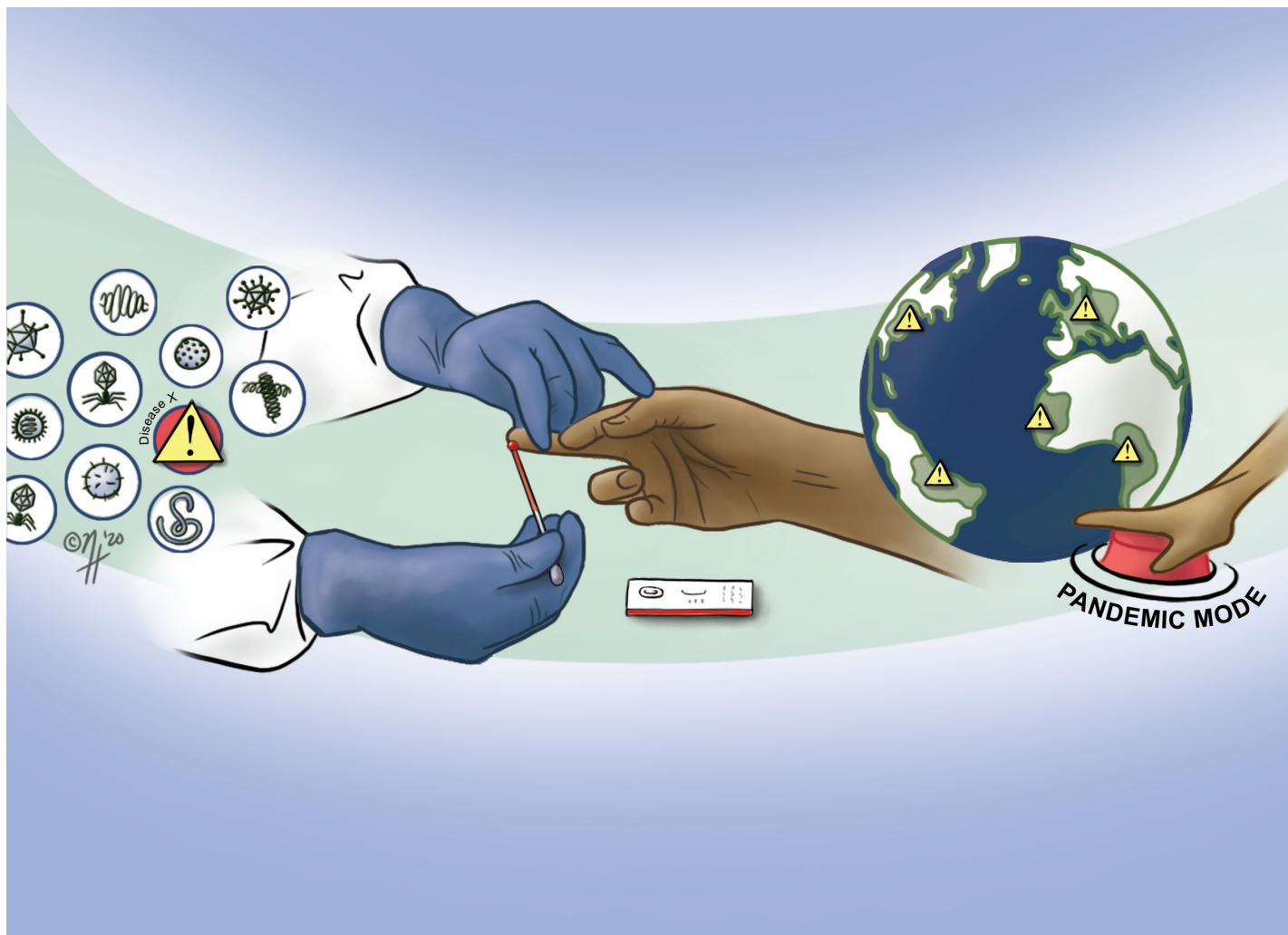




Showcasing research from Professor Winnie E. Svendsen's laboratory, Department of Bioengineering, Technical University of Denmark, Denmark.

Commercially available rapid diagnostic tests for the detection of high priority pathogens: status and challenges

This article describes the status and challenges of commercially available rapid diagnostic tests for detecting high priority pathogens listed by the World Health Organization as potential causes of present and future pandemics. The information these devices can collect is crucial for authorities to make effective decisions to contain the disease's advance. Image illustrated by Nicole Hampshire.

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Commercially available rapid diagnostic tests for the detection of high priority pathogens: status and challenges

Jaime Castillo-León, ^a Ramona Trebbien, ^b John J. Castillo ^c and Winnie E. Svendsen^a

The ongoing COVID-19 pandemic has shown the importance of having analytical devices that allow a simple, fast, and robust detection of pathogens which cause epidemics and pandemics. The information these devices can collect is crucial for health authorities to make effective decisions to contain the disease's advance. The World Health Organization published a list of primary pathogens that have raised concern as potential causes of future pandemics. Unfortunately, there are no rapid diagnostic tests commercially available and approved by the regulatory bodies to detect most of the pathogens listed by the WHO. This report describes these pathogens, the available detection methods, and highlights areas where more attention is needed to produce rapid diagnostic tests for future pandemic surveillance.

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1. Introduction

Disease outbreaks have occurred since the early days of human history in all regions of the world where humans have

migrated. One of the first reported disease outbreaks was the plague of Justinian in 541–542 AD. This pandemic affected the Byzantine Empire, killing as many as 50 million people.^{1,2} The cause of this pandemic was the bacterium *Yersinia pestis*, which is the same one that, centuries later, killed almost half of Europe's population during the years 1347–1351.³

The main reasons for disease outbreaks during human history are connected to the vast migration of humans throughout the planet and their interaction with new civiliza-

^aBioengineering Department, Technical University of Denmark, Ørsted's Plads, DK-2800 Kgs. Lyngby, Denmark. E-mail: jaic@dtu.dk

^bStatens Serum Institut, 5 Artillerivej, DK-2300 Copenhagen, Denmark

^cEscuela de Química, Universidad Industrial de Santander, Bucaramanga, Colombia



Jaime Castillo-León

Dr Jaime Castillo-León, Ph.D., is a chemist with experience in the manipulation, characterization, and integration of biological and semiconducting nanostructures into thin-films to fabricate miniaturized biosensing electrochemical and optical platforms and solar energy devices. His research has resulted in 47 scientific publications, four books edited, eight book chapters, and six patents.



Ramona Trebbien

Ramona Trebbien, DVM, Ph.D., Senior Scientist, Head of the National Influenza Center, Statens Serum Institut, Denmark. Dr Trebbien is part of the global and European virological networks, e.g., the Global Influenza Surveillance and Response System (GISRS) and the newly formed COVID-19 network of the World Health Organization and European Center for Disease Control.

Dr Trebbien has been working in the field of virology for nearly two decades and has vast experience in virus evolution and related methodology, e.g., genetic and phenotypic characterization. She has a background in investigations of the zoonotic and pandemic potential of influenza viruses.



tions and animals, resulting in the exchange of pathogens and a speeding up of diseases.⁴ Colonization expeditions, the foundation and expansion of cities, the opening of new and exotic trade routes, and lately, the phenomenon of globalization have contributed to the development and improvement of human life and increased risks of pandemics occurring.⁵

Simultaneously, scientific advances, an increase in the understanding of diseases, and improved life and hygienic conditions have been instrumental in preventing, controlling, and mitigating disease outbreaks. The development of new fabrication techniques, such as micro- and nanotechnology, and the improvement of sensing and diagnostic procedures during the last 100 years have been critical tools in the monitoring and preventing disease outbreaks.⁶ Having the possibility to quickly collect, identify, and analyze an increase in the number of infected people is an essential factor that allows authorities and health organizations to react early and take the necessary measurements to prevent disease spread. However, an unsatisfactory diagnostic capacity compromises outbreak detection and response, which results in the loss of lives.

Despite crucial advances in sensing and diagnostics, the need for more robust, sensitive, and useful devices for the rapid identification of infected individuals is still and will continue to be an urgent need.⁷ The latest report on the low efficiency of some of the existing rapid tests for detecting COVID-19 has raised questions about the quality of available rapid test devices.^{8,9}

Most of the more effective methods to identify pathogens responsible for disease outbreaks are based on molecular diagnostic assays that require the transportation of the samples to a lab, the use of specialized equipment and trained personnel. Moreover, since the test often takes more than 30 minutes, the more effective methods based on molecular assays are not ideal for analyzing thousands of samples.

The irruption of mobile phones and other devices, such as tablets, has opened a new spectrum of possibilities for com-

binning miniaturized biosensing analytical devices.¹⁰ A mobile phone provides an energy source, optical detection, analysis, storage, and data transmission on-site. Even though the combination of miniaturized analytical devices and mobile phones offers the perfect tools to be utilized for quick screening of infected individuals, the reality is that the combination of sensing platforms and mobile phone technology is still only available for research purposes.¹¹

This article highlights some of the deadly diseases that the WHO has identified as potential sources for future epidemics or pandemics.¹² Additionally, we present the status of existing commercially available rapid diagnostic tests to quickly identify infected individuals affected by these priority diseases identified by the WHO. Finally, we identify the prevailing challenges in developing more robust and effective rapid diagnostic test devices.

2. Rapid diagnostic test (RDT) – definition and characteristics

The WHO defines a rapid test as an assay that is “designed for use where a preliminary screening test result is required and is especially useful in resource-limited countries”.¹³ These tests are characterized as being:

- High-quality, easy-to-use tests for use in a resource-poor setting
- Quick and easy to perform and requiring little or no additional equipment
- Designed for use with a single or limited number of samples, making them more economical than, *e.g.*, ELISA in low-throughput laboratories
- Possible to store at room temperature for an extended period



John J. Castillo

John J. Castillo is currently an associate professor in analytical chemistry at the Universidad Industrial de Santander, Colombia. He obtained his Ph.D. in Chemistry from the Universidad Industrial de Santander in 2013. His main research interests are focused on the fabrication of novel amperometric biosensors based on nanomaterials integrated with plant peroxidases, and the development of gold and silver capped silicon nanopillar platforms for SERS detection.



Winnie E. Svendsen

Professor Winnie E. Svendsen has extensive experience in working with interdisciplinary research and innovation. She initially recognized the potential of combining novel micro and nanotechnology in biomedical research by detecting and manipulating DNA cancer biomarkers and later expanding this to lymphocytes, cancer cells, neuronal cells, bacteria, and brain tissue. She is the head of the Nano Bio Integrated Systems (NaBIS) group at DTU. NaBIS focuses on interdisciplinary research for solving the versatile challenges of integrating micro- and nanotechnology in biological analysis. Her research has resulted in 121 international journal publications, eight patents, and seven spin-off companies.



• Able to give same-day results, thus providing timely treatment interventions.

The acronym ASSURED, Affordable, Sensitive, Specific, User-friendly, Rapid, and Robust, was outlined by the WHO to summarize the set of criteria that the ideal RDT should have.

A lateral-flow assay works similarly to a chromatographic assay, where the components of a liquid sample are transported by capillary action inside a paper-based membrane. As shown in Fig. 2, an LFA has four main areas. The sample pad, where the sample is dropped; the conjugate pad, an area where the biorecognition elements, antibodies, are combined with labeled tags; test and control lines, where a color change indicates a positive or negative result; and the absorption pad, where excess sample and reagents are collected.

Two main types of immunoassays can be prepared using LFAs: sandwich and competitive assays. The sandwich assay is used to detect large analytes with multiple antigenic sites. In this format, labeled antibodies are immobilized on the conjugation pad, Fig. 2a. The label can be a metallic nanoparticle or a quantum dot. Once the sample is dropped, it is transported along the LFA membrane by capillary action. When the liquid sample passes through the conjugation pad, it rehydrates the labeled antibodies attached to the antigen epitopes. The labeled antibodies attached to the antigen continue traveling along the LFA membrane and reach the detection and control lines. If the antigen of interest is present, a colored line will appear at the detection lines. The color intensity at the test line is directly proportional to the analyte amount present in the sample. Regardless of the quantity of analyte in the sample, an anti-species antibody at the control line will bind the nanoparticle, yielding a solid control line signal, which demonstrates that the assay is functioning correctly.

When testing lower molecular weight analytes with a single antigenic site, the competitive lateral-flow assay is used, Fig. 2b. In this format, the test line typically contains the analyte antigen, and the conjugate pad contains the detection antibody-labeled conjugate. If the target antigen is present, the antigen will bind to the conjugate and prevent it from binding to the test line's antigen. If the antigen is not present, the con-

jugates will bind to the analyte at the test line, yielding a signal.

The signal intensity is inversely proportional to the amount of antigen present in the competitive format sample. As in the sandwich assay, the control line will bind the nanoparticle conjugate with or without the antigen, providing confidence that the assay is working correctly. In this case, only two types of antibodies are present in the LFA; a primary antibody coupled with labels and highly specific to the sample antigen, and a second species-specific anti-immunoglobulin antibody located at the control line.

Rapid diagnostic tests are point-of-need devices ideal for testing outside a dedicated laboratory space by personnel with minimal training. They are low-cost, require no or minimum energy, and can provide results within 5–30 minutes. RDTs are vital tools for rapidly detecting disease outbreaks, evaluating vaccine effectiveness, performing surveys of parasite prevalence, and allowing remote diagnosis in organized workforces entering endemic areas.

A key specification of RDTs is the ability to provide a rapid response to an analytical demand exactly where the demand is posed.¹⁴

RDTs simplify the analytical test by removing many of the steps required by traditional laboratory-based analysis, while still providing useful and reproducible information, as illustrated in Fig. 1. Additionally, RDTs also decrease the carbon footprint by reducing the amount of energy needed for analysis and skipping the transportation of samples to the lab to complete the analysis.^{15–17}

The technology behind most commercially available rapid diagnostic tests mainly focuses on lateral flow assays (LFA)¹⁸ and electrochemical methods.¹⁹ LFAs have shown their potential in applications such as pregnancy tests and the detection of illegal drugs. A lateral-flow-based test is ideal for the development of RDTs, mainly because of the advantages offered by using paper as the main component. Paper membranes are cheap to fabricate, easy to store, safe to dispose of, and can be used on-site anywhere and by anyone. Additionally, paper can be readily modified to create hydrophilic or hydrophobic areas

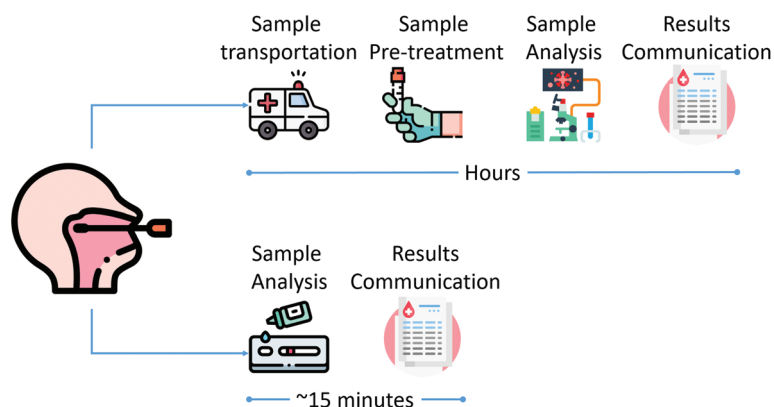


Fig. 1 Schematic workflow comparison between laboratory-based (top) and rapid diagnostic (bottom) testing.



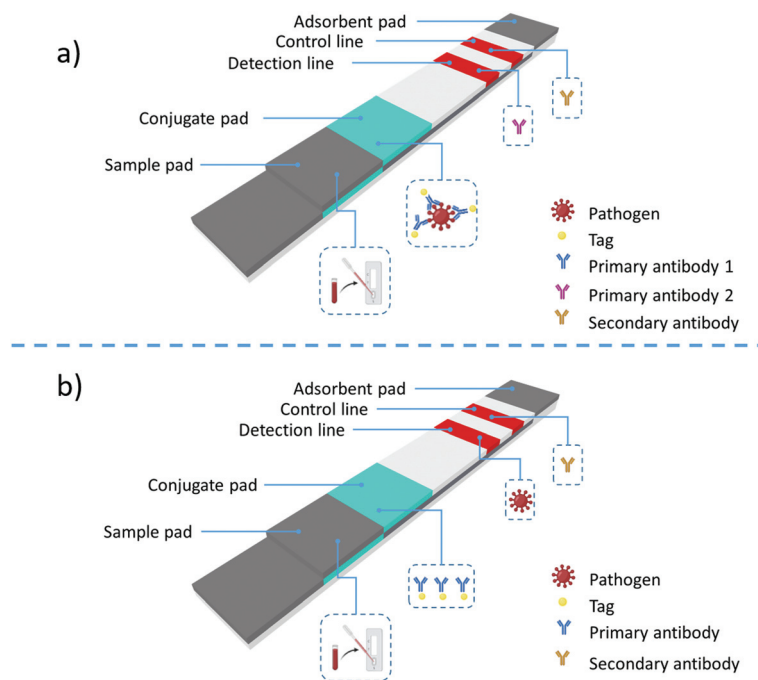


Fig. 2 Structure of a lateral-flow assay and the two main types of LFA formats: (a) sandwich format and (b) competitive format.

to control the reagents and sample flow. Paper-based sensing devices are light weight, compact, and easy to integrate into other technologies. Moreover, paper-based devices have been used in combination with mobile technology, enabling data management.²⁰

The majority of RDTs based on LFAs yield qualitative results, indicating the presence or not of a specific analyte by a change in color, giving a yes/no answer only. Lately, due to the integration of paper-based devices with mobile technology, it is possible to obtain a quantitative answer by relating the color intensity to the sample concentration.^{10,21,22} Additionally, extra features, such as storing and transmitting data, can be incorporated when combining LFAs with mobile technology.²³

Furthermore, LFA tests are disposable with no need for additional chemicals, and using LFAs brings about a massive reduction of the carbon footprint by avoiding transportation of samples to the lab and the use of specialized equipment during analysis.¹⁵ A recent study showed that the estimated CO₂ emissions of a single full-blood examination test done in a clinical lab were equivalent to driving a standard car for 770 m.^{16,17}

The success of electrochemical RDTs is centered around the amperometric glucose sensor's commercial success, an RDT used by millions of diabetic patients worldwide.²⁴ Electrochemistry has been suggested as a technique that could be used to develop RDTs to detect diseases in outbreaks.¹⁹ Electrochemical devices can be miniaturized, functionalized with different biosensing compounds (enzymes, antibodies, nanobodies), and offer the possibility of multiplexing. However, despite all of these advantages, the number of commercially available RDTs for detecting pathogens during disease outbreaks is low compared with lateral flow assays.

Other techniques, such as optical sensing and bioFETS, have only been used in research and have yet to reach the market.^{25–28}

The way the high-priority pathogens can be detected using RDTs can be divided into two groups: direct detection using antigen-detecting and molecular assay RDTs and indirect detection using the body's immune response to the pathogen in the form of antibodies. Lateral-flow immunoassay (LFI) platforms dominate the antigen-detecting RDTs; however, lately, the incorporation of molecular-based techniques has also reached commercialization.

For the second group, antibody-detecting RDTs, LFIs are the primary platform used to develop this kind of RDT.

Table 1 displays a comparison between molecular-based detection methods and antigen- and antibody-detecting assays.

For the RDTs presented in this article, the test sensitivity is calculated as (true positives)/(true positives + false negatives) × 100.

The specificity is calculated as (true negatives)/(true negatives + false positives) × 100.

3. High-priority pathogens identified by the WHO as potential causes of future epidemics

It is a fact that the impact of globalization is accelerating the spread of contagious diseases.⁵ The increase in air travel and



Table 1 Advantages and challenges of detecting methods used to develop RDTs

Test type	Advantages	Challenges
Molecular diagnostic assays	• High sensitivity and specificity • Detects active infection	• Labor intensive • Requires specialized laboratory infrastructure and skilled personnel • Expensive • Requires sample transportation
Antigen-detecting	• Detects active infection • It can be used as a point of care • Easy to perform • Lower cost than molecular diagnostic assays • Rapid results (<30 min)	• Lower sensitivity and specificity than molecular diagnostic assays • Lower predictive value than for molecular-based tests • A negative result cannot be used to remove an individual from quarantine
Antibody-detecting	• Detects that an individual has been infected • It can be used as a point of care • Easy to perform • Lower cost than molecular diagnostic assays • Rapid results (<30 min)	• Provides mainly qualitative results • Lower sensitivity and specificity than molecular diagnostic assays • Positive results do not assure the presence of protective immunity • Interpretation of results depends on the timing of the disease, the epidemiology and prevalence within the setting, and the clinical morbidity of the individual

the exchange of goods positively impact the global economy and contribute to the rapid spread of infectious pathogens worldwide. We have experienced the long-transmission potential of global air travel during the last twenty years spreading emerging pathogens.⁴ The SARS coronavirus emerged in 2002 in China and quickly spread to more than 30 countries, resulting in more than 8000 cases and around 774 deaths worldwide.²⁹ In 2009, the H1N1 influenza pandemic originated in Mexico and spread worldwide, causing 284 000 deaths.³⁰ More recently, Ebola affected African countries and resulted in travel restrictions to and from affected areas. Currently, COVID-19 affects the whole world's population, resulting in an almost complete stop of air travel and paralyzing most goods exchange. When the COVID-19 pandemic hit, it became apparent that testing was the key to containing the spread. Massive and timely testing was implemented in countries like South Korea and New Zealand using a combination of molecular assay-based screening and serological-based rapid tests for the detection of antigenic proteins and antibodies. Mass testing limited the virus's spread by both symptomatic and asymptomatic viral carriers and significantly decreased the death toll in these countries.³¹

The rapid transmission of COVID-19 around the world and its significant impact on society and the economy have led to the accelerated development of new tests to detect infected individuals quickly. Recently, thanks to essential economic support from governmental and private institutions, research groups and companies have focused on developing more effective and rapid tests.^{32,33} Using the information provided by these tests, health authorities could implement measurements like declaring local quarantine or forcing the use of masks instead of a complete lockdown to stop the spread of COVID-19.

However, this is not the case for most of the WHO-distinguished diseases that pose the highest public health risk due to their epidemic potential. These diseases are:

- Crimean-Congo hemorrhagic fever
- Ebola virus disease and Marburg virus disease
- Middle East respiratory syndrome coronavirus (MERS-CoV) and severe acute respiratory syndrome (SARS)
- Nipah and henipaviral diseases
- Rift Valley fever
- Zika
- Coronavirus disease-2019 (COVID-19)

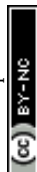
Next, we will describe each of these diseases, their impact areas, and the current rapid test status for their detection.

4. Crimean-Congo hemorrhagic fever (CCHF)

CCHF is a viral hemorrhagic fever with a high case fatality ratio caused by a virus transmitted to humans through a tick bite or direct contact with blood or tissue from infected ticks or viraemic vertebrates, wild animals, and livestock.³⁴ The virus can remain for up to one week in the bloodstream of infected animals, allowing the tick-animal-tick cycle to continue when another tick bites. Infected animals do not show any symptoms, which makes it challenging to identify them. Additionally, it allows the virus to maintain itself in nature.³⁵

It is considered the most geographically widespread tick-borne virus, with infection mortality in up to 30% of cases.³⁶ Regions of endemicity include parts of Africa, Asia, the Middle East, Russia, Eastern Europe, and Spain.

CCHF virus is a member of the family *Bunyaviridae*. It is an enveloped spherical particle of approximately 100 nm in diameter with a tripartite, single-stranded RNA genome of nega-



tive polarity. The three genome segments contain one open reading frame flanked by noncoding regions. It also contains four encoded structural proteins: the RNA-dependent RNA polymerase (L protein) expressed by the large (L) segment, the mature glycoproteins GN and GC are encoded by the medium (M) segment, and the nucleoprotein (N) by the small (S) segment.³⁴

Every year, 10 000 to 15 000 cases of CCHF are reported, mainly in endemic countries but also among travelers.

CCHF can be transmitted from human-to-human by direct contact with the body fluids of infected individuals. Human serological studies have revealed a mean seroprevalence of anti-CCHF antibodies of 4.7%. The seroprevalence of anti-CCHF antibodies is 7.5-fold higher among people with high-risk exposures. Increasing trends in CCHFV seroprevalence in both humans and animals, combined with evidence that people with frequent animal contact have more exposure to CCHFV, raise the concern that CCHFV could emerge as a zoonotic pathogen.³⁵

4.1 Symptoms

Patients who develop symptomatic CCHF progress through four stages: an incubation stage, a pre-hemorrhagic stage, a hemorrhagic stage, and a convalescent stage, as depicted in Fig. 3.³⁶

CCHF can be misdiagnosed with other viral hemorrhagic fevers, bacterial infections, viral infections, and parasitic infections such as malaria. Due to this, early laboratory confirmation of suspected cases is critical to prepare an adequate response.

CCHF symptoms could include myalgias, chills, fever, dizziness, headache, eye soreness, photophobia, sore throat, nausea, vomiting, and diarrhea.^{36,37}

Mortality occurs in approximately 30% of cases, often in the second week of illness.³⁷

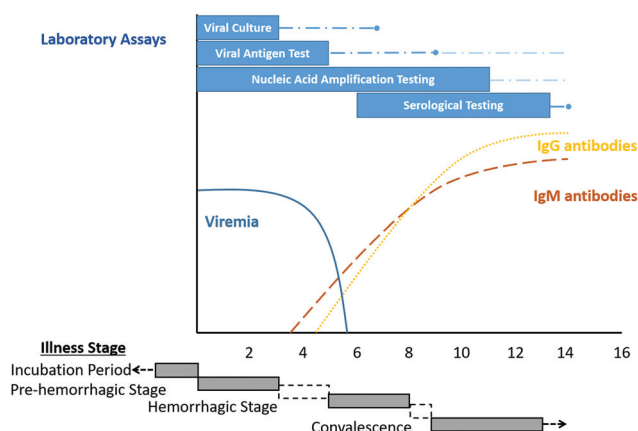


Fig. 3 Overview of diagnostic testing for acute illness due to the Crimean-Congo haemorrhagic fever virus in a non-fatal human infection. Reproduced from ref. 36 with permission from the American Society for Microbiology copyright 2020.

4.2 Diagnostic testing

There are currently two ways to diagnose CCHF in humans: by direct detection of the presence of CCHFV or by measuring the serological response consistent with acute infection by detecting the production of anti-CCHFV IgM or anti-CCHFV IgG antibodies.

The direct detection of CCHFV infection using nucleic acid amplification tests, viral antigen detection, and viral culture is most useful during the first week after symptom onset.^{37,38} Serological tests are most useful after the first week of illness, since IgM antibodies become detectable only after 7 to 9 days of symptom onset, except in some cases where IgM can be detected after four days of illness.³⁹ On the other hand, IgG antibodies become detectable simultaneously with IgM or one or two days later,³⁶ cf. Fig. 3.

4.3 Rapid test for the detection of CCHF

Currently, only one rapid test for the detection of CCHFV antibodies is commercially available. A lateral flow assay kit, CCHF Sero K-SeT, developed by CORIS BioConcept (Belgium), is used to detect IgM-specific antibodies in the patient's plasma, cf. Fig. 4. However, a study performed on patients in Iran showed very low sensitivity, 39.7%, and a specificity of 92.9%.⁴⁰ There is not a commercially available rapid test for the direct detection of the CCHF virus.



Fig. 4 CCHF Sero K-SeT IgM rapid test showing positive and negative results. (I) Positive result, (II) weakly positive result, and, (III) negative result. Reproduced from ref. 40 with permission from Elsevier copyright 2019.



5. Ebola virus (EV)

The Ebola virus is one of the most virulent pathogens to humans. The virus is endemic in African regions, where the first outbreaks took place in 1976 in a village near the Ebola River in the Democratic Republic of Congo.⁴¹ Since then, several outbreaks have been identified in 19 countries in Africa, Europe, Asia, and America.⁴² The largest recorded outbreak occurred in West African countries between December 2013 and 2016 and had a 40% fatality rate. Encroachment into forest areas due to population growth and direct interaction with wildlife contributed to the spread of the African region's Ebola virus.

Ebola virus is a single-stranded RNA virus belonging to the *Filoviridae* family along with the Marburg virus. Five virus species form the Ebola virus genus: *Reston ebolavirus*, *Tai Forest ebolavirus*, *Bundibugyo ebolavirus*, *Zaire ebolavirus*, and *Sudan ebolavirus*. The last two, *Zaire ebolavirus* and *Sudan ebolavirus*, are the most pathogenic and cause most outbreaks, with case fatality rates around 90%.⁴¹ The genome of the EV includes seven genes: the nucleoprotein, virion protein (VP) 35, VP40, glycoprotein, VP30, VP24, and RNA-dependent RNA polymerase (L)-5' trailer, cf. Fig. 5.

EV infection spreads by direct contact with bodily fluids from an infected animal or human, after which transmission occurs from human to human. EV has been identified in blood, feces, vomit, urine, semen, saliva, aqueous humor, vaginal fluid, breast milk, tears, and sweat.⁴³ Ingestion of undercooked bushmeat and contact while hunting or preparing a carcass have been identified as transmission methods to humans.⁴¹ The fruit bat has been singled out as the natural

host and is, in some cases, linked to direct transmission to humans.^{44,45}

5.1 Symptoms

EV commonly manifests with a hemorrhagic fever followed by an incubation period of 2–21 days. The incubation period is characterized by malaise, myalgia, chills, and fever. Other symptoms, like anorexia, nausea, vomiting, diarrhea, chest pain, cough, nasal discharge, prostration, conjunctival injection, headache, confusion, and coma, indicate multisystem involvement.⁴⁶ Additionally, by days 5–7 of the illness, erythema and desquamation can appear, which are used as differential diagnostic features.

5.2 Diagnostic testing

The primary detection assays used during EV outbreaks are ELISA, PCR, immunohistochemistry, virus isolation, and lateral-flow based assays.

ELISA and PCR techniques are sensitive methods that can detect EV in blood, serum, and tissue samples. They are lab-based techniques that require special equipment and trained personnel. They target viral nucleic acid, viral antigen, or virus-specific antibodies.⁴⁷ On the other hand, immunohistochemistry targets viral antigens in tissue samples such as the liver and skin. This is a qualitative imaging method that takes time to perform.

5.3 Rapid test for the detection of EV

Six RDTs for EV are available on the market, Table 2. These devices are based on lateral-flow and used during outbreaks, providing rapid information about the virus's presence in blood samples. OraQuick Ebola is a rapid antigen test approved by the U.S. Food & Drug Administration (the FDA).⁴⁸ This rapid antigen test is intended for use with whole blood obtained by venipuncture or finger stick. The sensitivity of this test on whole blood is 84%, with a specificity of 98%. Each test costs approximately USD 28.⁴⁹ After the addition of the sample, results are observed after 30 minutes.

The ReBOV antigen rapid test is a rapid chromatographic immunoassay intended for the qualitative detection of VP40 antigen from Zaire EV, Sudan EV, and Bundibugyo EV. This dipstick test has a sensitivity of 100%, with a specificity of 92%.⁵⁰ For this rapid test, whole blood from a finger stick is used (1 drop, 30 μ L), and results are observed after 15–25 min. The price of the ReBOV kit is USD 10.⁵¹

Brangel and co-workers have developed a serological point-of-care test for the detection of IgG antibodies against EV.²³ The novelty of this RDT device is the combination of the lateral-flow technique with mobile technology to obtain a quantitative result and the possibility to store or transmit data and tag the tested individuals geographically. Additionally, the device could simultaneously detect three related EV species: Sudan virus, Bundibugyo virus, and Zaire virus. Assay results are obtained within 15 minutes after the sample is added. The authors of this study reported a sensitivity and specificity of 100% compared with standard ELISA, and this work is an

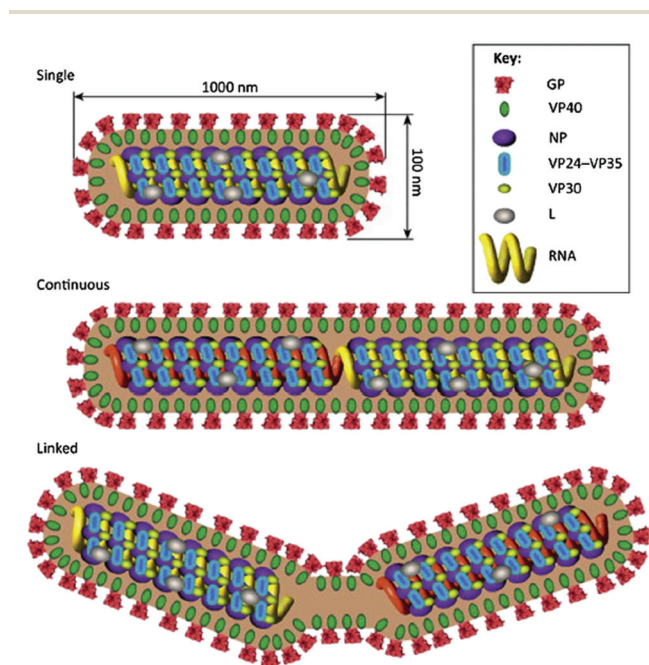


Fig. 5 Virion structure of the Ebola virus. Reproduced from ref. 41 with permission from Elsevier copyright 2016.



**Table 2** Commercially available RDTs for the detection of high-priority pathogens identified by the WHO

Disease	Test name	Test type	Developer	Sample	Detected	Sensitivity	Specificity	Response time (min)	Approval	Link
CCHF	Sero K-Set	LFI	CORIS BioConcept	Plasma	IgM	98.5%	98.6%	15	—	http://www.corisbio.com
EV	Senova Ebola Kit	LFI	Senova	Whole blood, serum, plasma	Antigen	—	—	—	—	http://www.senova.de
EV	ReEBOV	LFI	Corgenix	Whole blood, serum, plasma	Antigen	100%	92%	15–25	—	http://www.corgenix.com
EV	SD Ebola Zaire	LFI	SD	Whole blood, serum, plasma	Antigen	84.5%	98.9%	—	The WHO EUAL	http://www.sdbiosensor.com
EV	OraQuick	LFI	OraSure Technologies	Whole blood	Antigen	84%	98%	30	The FDA EUAL, the WHO EUAL	http://www.orasure.com
EV	DPP Ebola Antigen System	LFI	Chembio	EDTA whole blood, plasma	Antigen	77.1%	91.7%	10	The FDA EUAL	https://chembio.com
EV	QuickNavi-Ebola	LFI	Denka Seiken	Serum, plasma	Antigen	85%	99.8%	—	—	http://www.denka.co.jp
LFV	ReLASV Pan-Lassa	LFI	Zalgen Labs	Whole blood, serum, plasma	Antigen	83.3%	92.8%	5–25	—	http://www.zalgen.com
LFV	ReLASV antigen rapid test	LFI	Zalgen Labs	Whole blood, serum, plasma	Antigen	76.9%	94.4%	5–25	—	http://www.zalgen.com
ZV	DPP Zika	LFI	Chembio	Whole blood, serum, plasma	IgG/IgM	85.8%	81.4%	15	The FDA EUAL	http://chembio.com
ZV	One Step Zika	LFI	Arttron Laboratories Inc	Whole blood	IgG/IgM	18%	84%	10	—	http://www.arttronlab.com
ZV	Accu-Tel Zika Virus IgG/IgM AB Cassette	LFI	AccuBioTech	Whole blood, serum, plasma	IgG/IgM	100%	100%	20	—	https://www.accubiotech.com
ZV	Zika IgG/IgM Combo Test Card	LFI	LumiQuick Diagnostics	Whole blood, serum, plasma	IgG/IgM	94.6%	96.5%	15–20	—	https://lumiquick.co
ZV	Zika NS1 Test Card	LFI	LumiQuick Diagnostics	Whole blood, serum, plasma	Antigen	—	—	—	—	https://lumiquick.co
ZV	Zika IgG/IgM & NS1 Duo Test	LFI	LumiQuick Diagnostics	Whole blood, serum, plasma	IgG/IgM and antigen	—	—	—	—	https://lumiquick.co
ZV	CDIA Zika IgG/IgM Rapid Test Kit	LFI	Creative Diagnostics	Whole blood, serum, plasma	IgG/IgM	—	—	15	—	https://www.cd-diatest.com
ZV	Zika Rapid Test	LFI	Maternova	Whole blood, serum, plasma	IgG/IgM	—	—	—	—	https://maternova.net
ZV	Zika IgG/IgM	LFI	BioPanda	Whole blood, serum, plasma	IgG/IgM	80%	94%	15	—	https://www.biopanda.co.uk
ZV	Zika NS1	LFI	BioPanda	Whole blood, serum, plasma	Antigen	99.9%	98.9%	15	—	https://www.biopanda.co.uk
ZV	Zika Combo	LFI	BioPanda	Whole blood, serum, plasma	IgG/IgM and antigen	94.4%	99.9%	15	—	https://www.biopanda.co.uk
ZV	Zika v IgG/IgM	LFI	AdvaGen	Whole blood, serum, plasma	IgG/IgM	—	—	—	—	http://www.advagen.com.br
ZV	Zika-v NS1 Ag	LFI	AdvaGen	Whole blood, serum, plasma	Antigen	—	—	—	—	http://www.advagen.com.br
ZV	Zika IgG/IgM	LFI	Ebram Produtos Laboratoriais	Whole blood, serum, plasma	IgG/IgM	99.9%	98.9%	20	—	http://ebram.com
ZV	ProDetect Zika Rapid Test	LFI	Medical Innovation Ventures	Whole blood, serum, plasma	IgG/IgM	—	—	10–20	—	http://mediven.com.my

Table 2 (Contd.)

Disease	Test name	Test type	Developer	Sample	Detected	Sensitivity	Specificity	Response time (min)	Approval	Link
ZV	Rapikit	LFI	Nectar Lifesciences Ltd	Whole blood, serum, plasma	Antigen	93.5%	99.4%	—	—	https://rapikit.com
SARS-CoV-2	SARS-CoV-2 IgG/IgM Rapid Test Kit	LFI	Abbexa	Whole blood, serum, plasma	IgG/IgM	96.7%	96.2%	10	—	https://www.abbexa.com
SARS-CoV-2	Panbio SARS-CoV-2 Ag Rapid Test Device	LFI	Abbot	Nasopharyngeal swab	Antigen	98.1%	99.8%	15	CE European Mark	https://www.globalpointofcare.abbott
SARS-CoV-2	Accu-Tell SARS-CoV-2 IgG/IgM Rapid Test Cassette	LFI	Accu Biotech	Whole blood, serum, plasma	IgG/IgM	—	—	10	CE European Mark	https://www.accubiotech.com
SARS-CoV-2	SARS-CoV-2 IgG/IgM LFI	LFI	AdvaGen Biotech	Whole blood, serum, plasma	IgG/IgM	93.6%	100%	15	—	http://www.advagen.com.br
SARS-CoV-2	RapCov Rapid SARS-CoV-2 Test	LFI	Advaite	Whole blood	IgG/IgM	90%	95.2%	15	EUA	https://rapcov.com/
SARS-CoV-2	Assure SARS-CoV-2 IgG/IgM Rapid Test Device	LFI	Assure Tech	Whole blood, serum, plasma	IgG/IgM	100%	98.8%	15	EUA	http://www.assuretech.com.cn
SARS-CoV-2	SARS-CoV-2 Test Kit	LFI	AccuQuick Tests	Whole blood, serum, plasma	IgG/IgM	99%	99%	10	CE, EUA	https://www.accuquicktestkits.com
SARS-CoV-2	SARS-CoV-2 IgM/IgG Rapid Test Kit	LFI	Aurora	Whole blood, serum, plasma	IgG/IgM	93.3%	95%	15	CE European Mark	https://www.aurorabiomed.com
SARS-CoV-2	Autobio Anti-SARS-CoV-2 Rapid Test	LFI	Autobio Diagnostics	Serum, plasma	IgG/IgM	93.3%	98.8%	15	EUA	http://www.autobio.com.cn
SARS-CoV-2	SARS-CoV-2 IgG/IgM Point of Care Rapid Test	LFI	Aytu Biosciences/Orient Gene Biotech	Whole blood, serum, plasma	IgG/IgM	100%	100%	10	EUA	https://aytubio.com
SARS-CoV-2	BD Veritor System for Rapid Detection of SARS-CoV-2	LFI	Becton, Dickinson, and Company (BD)	Nasal swabs	Antigen	84%	100%	15	EUA	https://www.bd.com
SARS-CoV-2	OnSite SARS-CoV-2 IgG/IgM Rapid Test	LFI	Bio Advanced Diagnostics	Whole blood, serum, plasma	IgG/IgM	96.7%	78.1%	10–15	—	https://www.bioadvanceddiag.com.br
SARS-CoV-2	2019-nCoV IgG/IgM Detection Kit (Colloidal Gold)	LFI	Biolidics	Whole blood, serum, plasma	IgG/IgM	96.7%	100%	10	CE European Mark	https://www.biolidics.com
SARS-CoV-2	SARS-CoV-2 IgG/IgM Rapid Test	LFI	BioMedomics	Whole blood, serum, plasma	IgG/IgM	99%	100%	10–15	—	https://www.biomedomics.com
SARS-CoV-2	Wantai SARS-CoV-2 Ab Rapid Test	LFI	Beijing Wantai Biological Pharmacy Enterprise Co., Ltd	Whole blood, serum, plasma	IgG/IgM	81.2%	99.5%	15	EUA	http://www.ystwt.cn
SARS-CoV-2	Sars-CoV-2 IgM/IgG Antibody test Kit	LFI	Biohit Healthcare (Hefei) Co., Ltd	Whole blood, serum, plasma	IgG/IgM	100%	99.1%	15	EUA	http://www.biohit.cn
SARS-CoV-2	Sars-CoV-2 IgG/IgM Kit	LFI	BioTime	Whole blood, serum, plasma	IgG/IgM	96.2%	98.7%	10	—	http://www.biotime.cn
SARS-CoV-2	qSars-CoV-2 IgG/IgM Rapid Test	LFI	Cellex, Inc.	Whole blood, serum, plasma	IgG/IgM	94.6%	99.5%	15	EUA	https://cellexcovid.com





Table 2 (Contd.)

Disease	Test name	Test type	Developer	Sample	Detected	Sensitivity	Specificity	Response time (min)	Approval	Link
SARS-CoV-2	OnSite SARS-CoV-2 IgG/IgM Rapid Test	LFI	CTK Biotech, Inc.	Whole blood, serum, plasma	IgG/IgM	97.1%	97.8%	15	—	https://ctkbiotech.com/
SARS-CoV-2	One-Step SARS-CoV-2 Test	LFI	Celer Biotechnologia	Whole blood, serum, plasma	IgG/IgM	—	—	15	—	https://celer.ind.br
SARS-CoV-2	SARS-CoV-2 Ag Respi-Strip	LFI (dipstick)	Coris Bioconcept	Nasal mucus swabs	Viral antigen	91.2%	99.4%	30	—	https://www.corisbio.com/
SARS-CoV-2	DPP SARS-CoV-2 IgG/IgM System	LFI	Chembio Diagnostics	Whole blood, serum, plasma	IgG/IgM	—	—	15	The FDA	http://chembio.com
SARS-CoV-2	Coronavirus Rapid Test	LFI	Diagnostica Industria e Comercio Ltd	Whole blood, serum, plasma	IgG/IgM	86.4%	99.6%	15	—	https://www.biocodiagnosticos.com.br
SARS-CoV-2	2019 nCoV IgG/IgM Rapid Test	LFI	Dynamiker	Whole blood, serum, plasma	IgG/IgM	>92%	—	10	—	http://en.dynamiker.com
SARS-CoV-2	Coronavirus IgG/IgM (SARS-CoV-2)	LFI	Ebram Produtos Laboratoriais	Whole blood, serum, plasma	IgG/IgM	93.1%	95.4%	15–20	—	http://ebram.com
SARS-CoV-2	SARS-CoV-2 IgG/IgM ECO test	LFI	Eco Diagnostica Ltd	Whole blood, serum, plasma	IgG/IgM	100%	97%	15	—	http://ecodiagnostica.com.br
SARS-CoV-2	SARS-CoV-2 Ag ECO Test	LFI	Eco Diagnostica Ltd	Nasopharyngeal or oropharyngeal swab	SARS-CoV-2 antigen	96.5%	>99.9%	2–15	—	http://ecodiagnostica.com.br
SARS-CoV-2	VivaDiag SARS-CoV-2 IgG/IgM Rapid Test	LFI	Everest Links Pre Ltd	Whole blood, serum, plasma	IgG/IgM	82.9%	100%	15	—	http://everestlinks.com
SARS-CoV-2	FIA SARS-CoV-2 IgM/IgG	LFI	GenBody	Whole blood, serum, plasma	IgG/IgM	91.7%	97.5%	10	—	http://www.genbody.co.kr
SARS-CoV-2	RightSign SARS-CoV-2 IgG/IgM Rapid Test Cassette	LFI	Hangzhou Biotest Biotech	Whole blood, serum, plasma	IgG/IgM	93.3%	100%	10	EUA	https://biotestsolutions.com
SARS-CoV-2	LYHER Novel Coronavirus (2019-nCoV) IgM/IgG Antibody Combo Test Kit	LFI	Hangzhou Laihe Biotech	Whole blood, serum, plasma	IgG/IgM	100%	98.8%	15	EUA	http://www.lyher.com
SARS-CoV-2	SARS-COV-2 IgG/IgM Rapid Test Cassette	LFI	Healgen Scientific	Whole blood, serum, plasma	IgG/IgM	96.7%	100%	10	EUA	https://www.healgen.com
SARS-CoV-2	2019-nCoV Ab Test (Colloidal Gold)	LFI	Innovita Biological Technology Co. Ltd	Whole blood, serum, plasma	IgG/IgM	93.3%	98.8%	15	—	http://www.innovita.com.cn
SARS-CoV-2	Anti SARS-CoV-2 IgG/IgM Rapid Test	LFI	Labtest Diagnostica S.A.	Whole blood, serum, plasma	IgG/IgM	85%	98.3%	15	—	https://labtest.com.br
SARS-CoV-2	StrongStep® SARS-CoV-2 IgG/IgM Combo Test	LFI	Liming Bio	Whole blood, serum, plasma	IgG/IgM	—	—	15	—	http://www.limingbio.com
SARS-CoV-2	Rapid Test SARS-CoV-2	LFI	Livzon Diagnostics	Whole blood, serum, plasma	IgG/IgM	90.6%	99.2%	15	—	https://livzon-covid-19test.com/
SARS-CoV-2	2019 Antibody (IgM/IgG) Combined Test Kit	LFI	Medical Systems Biotechnology	Whole blood, serum, plasma	IgG/IgM	—	—	5–10	—	http://en.nbmedicalsyste.com
SARS-CoV-2	Megna SARS-CoV-2 Antibody Test Kit	LFI	Megna Health	Whole blood, serum, plasma	IgG/IgM	100%	95%	15	EUA	https://www.megnahealth.com



Table 2 (Contd.)

Disease	Test name	Test type	Developer	Sample	Detected	Sensitivity	Specificity	Response time (min)	Approval	Link
SARS-CoV-2	SARS-CoV-2 rapid diagnostic test	LFI	Mologic	Whole blood, serum, plasma	IgA/IgG/IgM	—	—	—	CE European Mark	http://www.mologic.co.uk
SARS-CoV-2	2019-nCoV IgG/IgM Detection Kit (Colloidal Gold-Based)	LFI	Nanjing Vazyme Medical Technology Co. Ltd	Whole blood, serum, plasma	IgG/IgM	—	—	10	—	http://www.vazymebiotech.com
SARS-CoV-2	2019 COVID Antibody IGG IGM Test Cassette (Colloidal Gold)	LFI	Nantong Diagnose Biotechnology	Whole blood, serum, plasma	IgG/IgM	—	—	—	—	http://www.diagnosticsbio.com/en/
SARS-CoV-2	Rapid Test for SARS-CoV-2 IgM/IgG Antibody Detection Kit	LFI	Nirmidas Biotech, Inc.	Whole blood, serum, plasma	IgG/IgM	96.7%	97.5%	20	—	https://www.nirmidas.com
SARS-CoV-2	2019-nCoV IgG/IgM Rapid Test	LFI	Phamatech	Whole blood, serum, plasma	IgG/IgM	—	—	—	—	https://www.phamatech.com
SARS-CoV-2	SARS-CoV-2 IgG/IgM Rapid Test Cassette	LFI	Premier Biotech	Whole blood, serum, plasma	IgG/IgM	100%	93%	10	EUA	https://premierbiotech.com
SARS-CoV-2	Sofia SARS Antigen FIA	LFI AFI	Quidel	Direct nasal/nasopharyngeal swabs	Antigen	96.7%	100%	15	EUA	https://www.quidel.com
SARS-CoV-2	SARS-CoV-2 IgM/IgG Ab Rapid Test	LFI	Qingdao Hightop Biotech Co. Ltd	Whole blood, serum, plasma	IgG/IgM	93%	97.5%	—	—	http://www.hightopqd.com
SARS-CoV-2	Coronavirus (SARS-CoV-2) IgM/IgG Rapid Test Kit	LFI	Ray Biotech	Whole blood, serum, plasma	IgG/IgM	90.4%	98.3%	5–10	—	https://www.raybiotech.com
SARS-CoV-2	Salacor SARS-CoV-2 (Sars-CoV-2) IgG/IgM Rapid Test Kit	LFI	Salofa Oy	Whole blood, serum, plasma	IgG/IgM	94.9%	99.2%	—	EUA	https://www.salofa.com/en/
SARS-CoV-2	SARS-CoV-2 Antibody Test Strip	LFI (colloidal gold)	Sannuo Biotech (Sinocare)	Whole blood, serum, plasma	IgG/IgM	—	—	—	—	http://www.sinocare.com
SARS-CoV-2	STANDARD Q SARS-CoV-2 Ag	LFI	SD Biosensor	Nasopharyngeal swab	Viral antigen	87.9%	98.9%	15–30	—	http://sdbiosensor.com
SARS-CoV-2	STANDARD Q SARS-CoV-2 IgM/IgG Combo	LFI	SD Biosensor	Whole blood, serum, plasma	IgG/IgM	94.5%	69.1%	10–15	—	http://sdbiosensor.com
SARS-CoV-2	SARS-CoV-2 Rapid IgG/IgM combined Antibody assay Pre-screening test Kit	LFI	Sensing Self	Whole blood, serum, plasma	IgG/IgM	99.3%	100%	10	—	https://sensingself.me
SARS-CoV-2	Bioeasy 2019-nCoV IgG/IgM Ab GICA Rapid Test (Colloidal Gold)	LFI	Shenzhen Bioeasy Biotechnology Co. Ltd	Whole blood, serum, plasma	IgG/IgM	—	—	10–15	—	http://en.bioeasy.com.tr
SARS-CoV-2	Rapid Sars-CoV-2 antigen detection test	LFI	Sona Nanotech	Whole blood, serum, plasma	S1 domain of spike protein	84.6%	90%	15	—	https://sonanano.com

Table 2 (Contd.)

Disease	Test name	Test type	Developer	Sample	Detected	Sensitivity	Specificity	Response time (min)	Approval	Link
SARS-CoV-2	SGTI-flex SARS-CoV-2 IgM/IgG	LFI	Sugentech	Whole blood, serum, plasma	IgG/IgM	90.2%	100%	10–15	—	http://sugentech.com
SARS-CoV-2	Sars-CoV-2 IgM/IgG Antibody Rapid Test	LFI	Sure Biotech	Whole blood, serum, plasma	IgG/IgM	92.5%	99.2%	20	—	https://www.surebiotech.com
SARS-CoV-2	VivaDiag SARS-CoV-2 IgG/IgM Rapid Test	LFI	VivaChek Biotech	Whole blood, serum, plasma	IgG/IgM	—	—	15	—	https://www.vivachek.com
SARS-CoV-2	Sars-CoV-2 IgM/IgG Antibody Test Kit	LFI	Wuhan Easy Diagnosis Biomedicine Co., Ltd	Whole blood, serum, plasma	IgG/IgM	99.3%	96.2%	15	—	http://www.mdeasydiagnosis.com
SARS-CoV-2	SARS-CoV-2 IgG/IgM Rapid Test	LFI	Zhejiang Orient Gene Biotech Co. Ltd	Whole blood, serum, plasma	IgG/IgM	86.7%	>99%	15	—	http://www.orientgene.com
SARS-CoV-2	Diagnostic Kit for IgM/IgG Antibody to Coronavirus (SARS-CoV-2)	LFI	Zhuhai Livzon Diagnostics Inc.	Whole blood, serum, plasma	IgG/IgM	90.6%	99.2%	15	—	http://www.livzondiagnostics.com
SARS-CoV-2	Lucira COVID-19 All-in-one Test Kit	RT-LAMP	Lucira Health	Nasal swabs	SARS-CoV-2 RNA	98.4%	100%	30	The FDA EUA	https://www.lucirahealth.com

excellent example of the enormous potential of combining RDTs with mobile technology in disease outbreaks. Fig. 6 shows the developed lateral flow device and the smartphone application interface used to detect the intensity of the signal corresponding to the virus's presence and record the patient's details.

6. Nipah virus (NiV)

Nipah virus is an emerging infectious disease that kills about 75% of those infected. Currently, there is no cure for this disease. NiV is a bat-borne pathogen; Pteropus fruit bats are considered the virus's natural reservoir.⁵² The bats are endemic to tropical and subtropical regions of Asia, East Africa, the Australian continent, and some oceanic islands.⁵³ There are no FDA-approved drugs to treat or prevent NiV.^{54,55}

NiV was discovered in 1998 during the first reported outbreak in the Sungai Nipah village (Malaysia), where locals became infected through contact with pigs, which are the intermediate host of the virus. Since then, recurring NiV outbreaks have been reported in Bangladesh, India, Ghana, Thailand, Madagascar, Cambodia, and Singapore.⁵⁶

NiV is an enveloped pleomorphic virus belonging to the genus *Henipavirus* of the family *Paramyxoviridae*. The virus structure consists of a non-segmented negative-sense single-stranded RNA, approximately 18.2 kb long, encoding six structural proteins: nucleocapsid (N), phosphoprotein (P), matrix protein (M), fusion protein (F), glycoprotein (G), and RNA polymerase (L), *cf.*, Fig. 7. The F and G proteins of NiV are necessary for attachment and entry into the host cell.^{53,57}

6.1 Symptoms

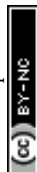
NiV infection causes febrile encephalitis or pneumonia at the early stage, making it difficult to distinguish from other febrile diseases. Other clinical features of NiV are headache, vomiting, behavioral changes, disorientation, and low levels of consciousness. Incubation periods have been reported to range from 4 to 14 days. Around 20% of encephalitis survivors sustained neurological dysfunction, including persistent seizures, fatigue, and behavioral abnormalities.⁵⁸

6.2 Diagnostic testing

The tests for detecting NiV include molecular assays, immunochemistry, histopathology, and serological tests.⁵⁹ Techniques such as PCR, next-generation sequencing, immunochemistry, and ELISA are among the most used for NiV detection. The type of samples collected for NiV detection in humans include throat and nasal swabs, urine, blood, and cerebrospinal fluid. All testing techniques are laboratory-based; there is no commercially available rapid test for the detection of NiV in humans or animals.⁶⁰

6.3 Rapid test for the detection of NiV

In 2014, a lateral-flow assay for the rapid detection of NiV in swine was reported by the Canadian Science Centre for



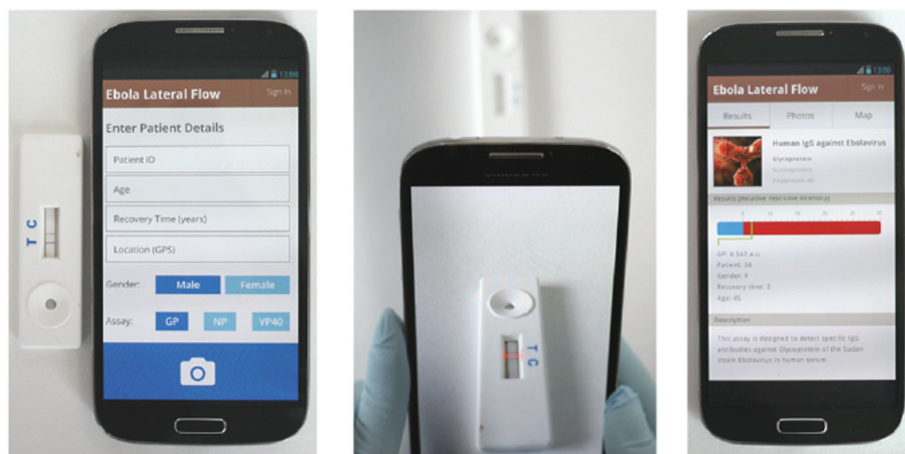


Fig. 6 Smartphone lateral-flow point-of-care test for Ebola virus IgG detection. Reproduced from ref. 23 with permission from ACS copyright 2018.

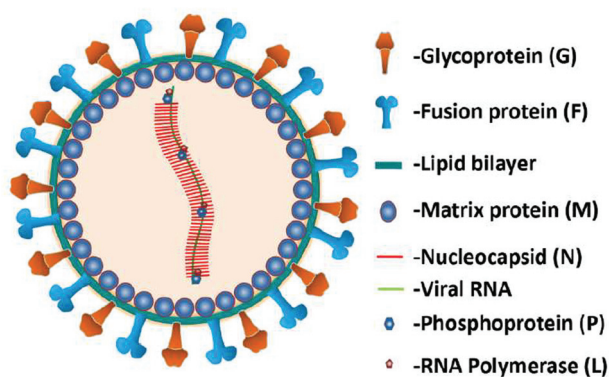


Fig. 7 Representation of the Nipah virus structure. Reproduced from ref. 53 with permission from MDPI copyright 2020.

Human and Animal Health.⁶¹ The developed rapid test was compared with PCR, and it was concluded that the lateral-flow assay was as sensitive as real-time PCR. Additionally, it was possible to detect viral protein as early as one-day post-inoculation in nasal swabs, and no cross-reaction with Hendra virus and other viruses, *e.g.*, Ebola virus, was observed. This RDT is, however, not commercially available.

7. Rift Valley fever (RVF)

Rift Valley fever is a viral zoonosis that affects animals, but can also infect humans. Approximately 1% of humans infected with RVF die from the disease.⁶² It primarily infects sheep, cattle, and goats and is maintained in nature by transovarial transmission in floodwater *Aedes* mosquitoes.⁶³ The virus was identified for the first time on a farm in the Rift Valley of Kenya in 1931. Since then, outbreaks have been reported in Niger, Mauritania, Madagascar, Sudan, Kenya, Somalia, Tanzania, Egypt, Yemen, Saudi Arabia, and South Africa.⁶⁴

The trade of animals between endemic areas and Mediterranean European countries actively contributes to viral

transmission, thus increasing the risk of contagion in the Mediterranean basin and Europe.⁶⁵ Moreover, the virus's ability to survive in a range of bioclimatic environments has raised concerns about its potential introduction into Europe and North American areas.⁶⁶

RVF is a mosquito-borne viral disease from the genera *Aedes* and *Culex*. These mosquito species are associated with freshly flooded temporary water bodies and are regarded as maintenance vectors in the *Aedes* species and as epidemic or amplifying vectors in the case of the *Culex* species.⁶⁷ Animals become infected through a bite from an infected mosquito. In the case of humans, they become infected *via* direct and aerosol exposure to infected animals or animal tissue and when bitten by infected mosquitoes. No human-to-human transmission of RVFV has been documented.

The RVFV genome is composed of three segments of single-stranded RNA: large (L), medium (M), and small (S). The viral RNA-dependent RNA polymerase (L protein) is encoded in the L segment. Four proteins are encoded in the M segment: the structural glycoproteins Gn and Gc, the non-structural protein Nsm, and a large 78 kDa glycoprotein (LGp). The structural glycoproteins mediate binding and entry to the target cell membrane *via* receptors. The nucleoprotein (N) and the non-structural NS protein, a significant determinant of virulence, are encoded in the S segment.⁶⁸

7.1 Symptoms

In humans, infection with RVFV is generally asymptomatic in 98% of cases. Only 2% of cases are severe, presenting three different complications: hemorrhagic fever, ocular disease (retinal vasculitis), and meningoencephalitis. Other symptoms corresponding to a mild infection form are flu-like fever, headache, joint pain, sensitivity to light, neck stiffness, vomiting, and appetite loss.⁶⁹

Individuals with clinical symptoms present short febrile illness and no long-term sequelae. During the initial phase of the disease, 1–6 days after infection, antibody levels rise while



viremia declines. Thrombosis and other vascular complications can manifest days to weeks after the initial infection.⁶⁵

7.2 Diagnostic testing

RVFV is detected using molecular assay techniques, namely, reverse transcriptase-polymerase chain reaction (RT-PCR) assay, loop-amplification mediated polymerase chain reaction (LAMP), ELISA, antigen detection tests, and virus isolation by cell culture. RT-PCR and LAMP are the primary assays used for detecting acute infections.^{70,71} For the detection of antibodies in several species, mainly ELISA platforms are used.^{72–74}

7.3 Rapid test for the detection of RVFV

Only one device for the rapid detection of RVFV has been reported.⁷⁵ The first-line lateral flow immunochromatographic strip test was developed to detect nucleoprotein (N) of the RVFV, cf. Fig. 8.

The fabricated device showed lower values of specificity and sensitivity compared with molecular-based techniques (PCR), but was, according to the authors of the study, adequate for the specific rapid initial detection of RVF outbreaks. This RDT is obtainable only for research purposes and is not commercially available.

8. Lassa fever virus (LFV)

Lassa fever is a zoonotic disease caused by the Lassa fever virus, a level IV pathogen. The virus' reservoir is a rodent of the genus *Mastomys*, known as a multimammate rat.⁷⁶ It is transmitted to humans by the ingestion of contaminated food or by contact with household items contaminated with rat urine and feces. Human-to-human transmission occurs through direct contact with blood, organs, or other body fluids from infected individuals. Rodent excretion aerosolized during cleaning activities could result in airborne transmission of the virus.⁷⁷

The first case of Lassa fever virus was detected in 1950 in Sierra Leone. Lassa virus belongs to the *Arenaviridae* family, which is responsible for causing viral hemorrhagic fever in African and South American countries.⁷⁸ The Lassa virus

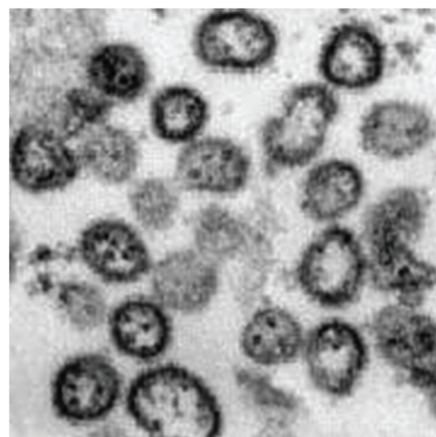


Fig. 9 Lassa virus electron micrograph reproduced from ref. 77 with permission from Elsevier copyright 2014.

measures between 70 and 150 nm in diameter, has a spherical shape, and belongs to the *Arenaviridae* family, cf. Fig. 9. The Lassa virus possesses a glycoprotein envelope with T-shaped spikes measuring 10 nm on its surface. The Lassa virus's genome is a single-stranded, bisegmented RNA consisting of a small (S) and a large (L) RNA fragment of 3.4 and 7 kilobases.⁷⁷

Lassa fever is endemic in West Africa, where a population of 58 million is at risk. However, imported cases have appeared in Germany, Sweden, the USA, Japan, the Netherlands, and the UK. Every year, approximately 100 000 to 300 000 cases of Lassa fever are reported in West Africa, with an estimated death toll of 5000 deaths per year.⁷⁹

8.1 Symptoms

LASV hinders the development of immune cells that are crucial to the immune response and that provide resistance to infection. The severity of LASV is correlated with high viremia levels, with peaks between 4 and 9 days after the onset of the disease. The initial diagnostic of LASV can be challenging, since the symptoms can mimic other endemic diseases, such as typhoid fever and malaria.⁸⁰

Around 30% of individuals infected with Lassa fever present visible bleeding, which could discriminate LASV from other febrile illnesses. Facial swelling, low blood pressure, hepatitis, fluid in the lung cavity, and petechiae or bruising indicate severe infection.⁸¹ Deaths due to LASV typically occur within 14 days of onset for 15–20% of severe cases. In the case of non-fatal cases, the patient's condition improves after 1–3 weeks. However, fatigue and renal damage may persist for several more weeks.⁸²

8.2 Diagnostic testing

The current LASV diagnostic test includes laboratory-based serological and nucleic acid amplification tests. However, the majority of commercial assays for PCR and serology are indicated for research use only. Additionally, endemic regions do

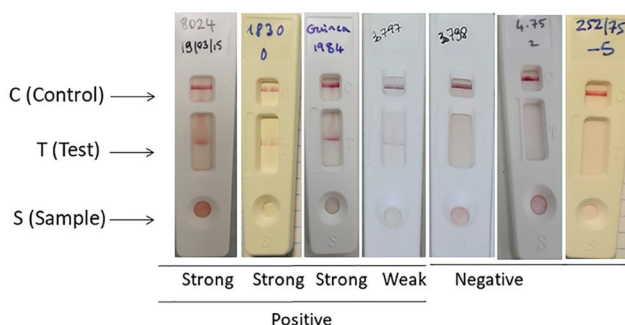


Fig. 8 RVF lateral flow test strips for the detection of RVF infection. Reproduced from ref. 75 with permission from PLOS copyright 2019.



not always have the financial resources and personnel to utilize these available diagnostics tests.⁸³

Immunofluorescent assay tests, western blotting, and ELISA are techniques used for the indirect detection of antibodies and antigens. LASV RNA can be detected using nucleic acid amplification, including techniques such as loop-mediated isothermal amplification (LAMP), strand displacement assays, and PCR. Virus isolation, PCR, and LASV antigen positivity are used to diagnose active infections.⁸⁴

8.3 Rapid test for the detection of LASV

Apart from the lab-based test, there are two commercially available kits for the rapid detection of LASV on the market: the ReLASV® Pan-Lassa Antigen Rapid Test and the ReLASV® Antigen Rapid Test.⁸⁵ However, they are labeled for research use only. These rapid detection tests are lateral flow-based dipsticks, *cf.*—Fig. 10. When applying a drop of whole blood, the

nucleoprotein antigen is detected with 91% sensitivity and 86% specificity for LASV lineage IV (Josiah strain). The performance of this test at temperatures between 18 and 30 °C is comparable to molecular tests such as RT-PCR and qPCR.^{86,87} However, data for the rapid test performance at high temperatures above 30 °C and high humidity are required to better indicate the test's performance in endemic areas. The test cost is around USD 30, with a turnaround time of <30 min.⁸⁵

9. Zika virus (ZV)

Zika virus is an enveloped, positive-sense, single-stranded RNA virus member of the *Flaviviridae* family that is transmitted to humans by mosquitoes from the *Aedes* species. It was discovered in 1947 in the Zika forest (Uganda).⁸⁸ Since then, several outbreaks of ZV have been reported in various countries in Africa, Asia, and South and Central America. One of the significant ZV outbreaks started in 2015 in Brazil and extended to most countries in the Caribbean, Central, and South American areas. Following this significant outbreak, ZV infection among travelers was reported in European countries, including the UK, Austria, Denmark, France, Finland, Italy, Ireland, Portugal, Germany, Sweden, the Netherlands, Switzerland, and Spain.⁸⁹

Before the large outbreaks in 2014 and 2015, research into ZK pathogenesis, detection, and treatment was neglected due to the mild symptoms and the high percentage of asymptomatic individuals.⁹⁰

The primary way of ZV transmission is through mosquito bites. However, additional transfer modes have been identified, such as materno-fetal by trans-placental transmission or during delivery when the mother is infected, sexual transmission, or through blood transfusion and organ transplantation,⁹¹ *cf.* Fig. 11.

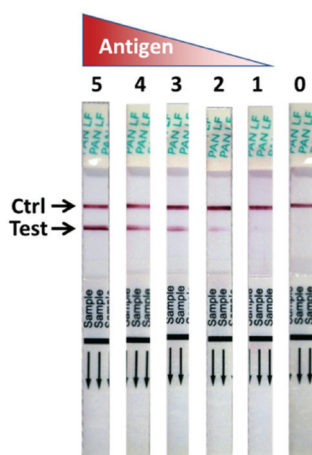


Fig. 10 Signal development of the ReLASV Pan-Lassa antigen rapid diagnostic test. Reproduced from ref. 87 with permission from Springer Nature copyright 2018.

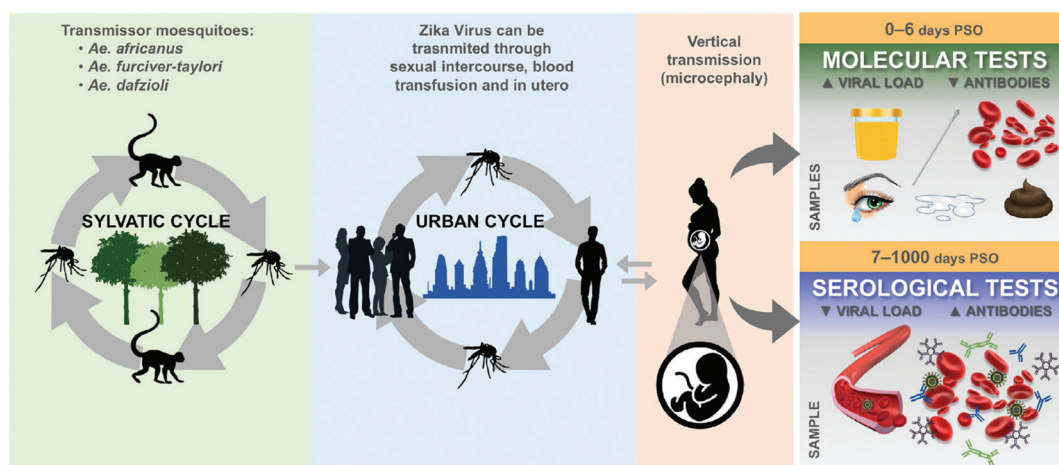


Fig. 11 The life cycle of the Zika virus and biological materials used to detect the disease. Reproduced from ref. 98 with permission from John Wiley & Sons copyright 2020.



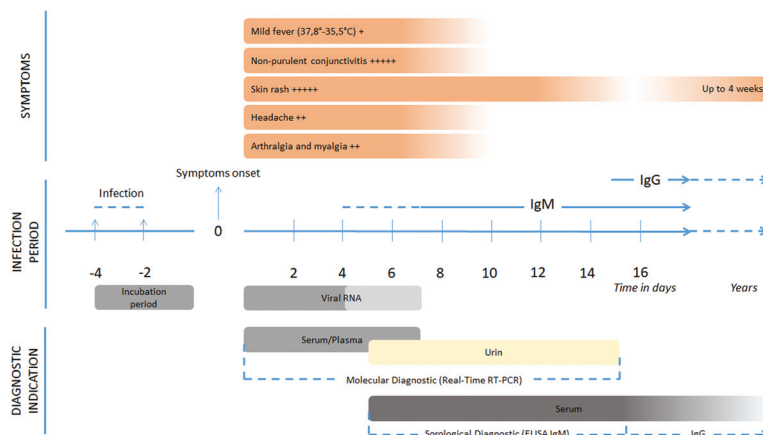


Fig. 12 Chronology of viral replication, relationship with clinical and biological symptoms, and the possibility of the detection of Zika virus by laboratory techniques. Reproduced from ref. 98 with permission from John Wiley & Sons copyright 2020.

9.1 Symptoms

Due to serologic cross-reactivity to other flaviviruses, such as the dengue virus, ZV diagnosis has been challenging.⁸⁸ The incubation period for ZV ranges between 4 and 12 days after infection, followed by a period of up to sixteen days, during which symptoms such as mild fever, skin rash, non-purulent conjunctivitis, headache, arthralgia, and myalgia may appear, *cf.* Fig. 12.

If a woman is infected during pregnancy, the fetus has a high risk of suffering congenital central nervous malformations and microcephaly. In some cases, fetal losses have been reported during Zika outbreaks.⁹²

9.2 Diagnostic testing

The irruption of ZK in South and North America and Europe has accelerated the development of assays to detect infected individuals.^{89,93} For instance, after the 2015 outbreak, the FDA granted emergency use authorization to 14 molecular and five serological assays to detect ZV.⁹³

One of the primary molecular assays used for ZV detection is real-time reverse transcriptase PCR (rRT-PCR), which targets the ZV E gene in serum, whole blood, urine, cerebrospinal fluid, and amniotic fluid.⁹⁴ Another molecular assay used for ZV detection is transcription-mediated amplification (TMA). This assay singles out the ZV NS1 and NS4/NS5 genes in serum and urine.⁹⁵

ELISA has been used for the colorimetric detection of ZV and its differentiation from the dengue virus. This qualitative test detects the presence of ZV IgM antibodies in human sera with a sensitivity of >90%, a specificity of >96%, and a turn-around time of four hours.⁹⁶

9.3 Rapid test for the detection of ZV

Several RDTs for ZV detection are currently on the market.^{97,98} They are mainly immunochromatographic assays for the detection of IgG, IgM antibodies, or NS1 protein.⁹⁹ Chembio Diagnostic Systems developed a lateral flow assay, DPP® Zika



Fig. 13 DPP Zika IgM system developed by Chembio for Zika virus detection (image from <http://www.chembio.com>).

IgM System, read by an automated reader,¹⁰⁰ *cf.* Fig. 13. This RDT is the only immunochromatographic assay that is FDA Emergency Use Authorization (EUA) approved. According to the manufacturer, the RDT can be used to detect IgM and IgG antibodies to ZV in whole blood, serum, or plasma, and the results are displayed after 15 minutes. The RDT requires only 10 µL of sample to provide a semi-quantitative answer when combined with the reader. Each kit's price is around USD 35/card, while the reader's price is around USD 754.

Apart from the DPP Zika RDT, there are other immunochromatographic rapid assays available on the market, some of which are listed below:

- Artron One Step Zika Virus Test Kit, Artron Laboratories Inc (Canada),¹⁰¹
- Accu-Tell Zika Virus IgG/IgM AB Cassette, AccuBioTech (China),¹⁰²
- Zika NS1 Test Card, Zika IgG/IgM & NS1 Duo Test, LumiQuick Diagnostics Inc. (USA),¹⁰³



Apart from these commercially available RDTs for ZV detection, other devices based on electrochemistry and lateral flow have been reported in the literature, but are only for research purposes.^{99,109–112}

MERS-CoV is a lineage C beta coronavirus that was first identified in June 2012 in Saudi Arabia. Since then, several outbreaks have been reported in 27 countries in Asia, Africa, and Europe, with mortality of around 34.4%.^{113–115} MERS-CoV's genotype is closely related to bat coronaviruses, such as BtCoV-HKU4 and BtCoV-HKU5, but its evolutionary pathway is still not fully understood. MERS-CoV exists as enveloped extracellular particles with protruding spike (S) proteins.¹¹⁶ MERS-CoV targets the cellular receptor using its spike glycoprotein (S), a putative receptor-binding domain.¹¹⁷ The MERS-CoV infection results in profound apoptosis of infected human bronchial epithelial Calu-3 cells within 24 hours.¹¹⁸

Several studies link MERS-CoV to dromedary camels.¹¹³ Additional studies have indicated potential cross-infection between dromedary camels and humans due to close contact. Compared with other coronaviruses, MERS-CoV has a lower basic reproduction number (R_0); this is the average number of infections caused by one infected individual in a fully susceptible population. MERS-CoV R_0 is around 1.0 throughout various clusters and outbreaks.¹¹⁹ Transmission between humans occurs through close and unprotected human contact and nosocomial transmission, mainly at healthcare locations and households.^{71,120–122}

The incubation period for MERS is approximately five days. Individuals infected with MERS-CoV will present chills, fever, shortness of breath, cough, generalized myalgia, nausea, vomiting, and diarrhea. These symptoms are similar to other respiratory diseases, such as SARS. Due to the absence of specific laboratory abnormalities or clinical data to differentiate MERS-CoV pneumonia from pneumonia caused by other viruses, real-time PCR is used to identify the virus from respiratory secretions.¹¹⁷

Older age (>60 years), as well as a medical history of chronic lung disease, diabetes, cancer, or renal disease, are considered risk factors that could aggravate the condition of MERS patients, resulting in respiratory failure and the need

for mechanical ventilation and support in intensive care units.¹²³

As mentioned before, the primary method to identify and differentiate MERS-CoV from other respiratory syndromes is real-time PCR using samples from the lower respiratory tract, namely, sputum, tracheal aspirates, or bronchoalveolar lavage.¹²⁴

ELISA, indirect immunofluorescence, and neutralization tests are serological tests used for MERS-CoV detection.^{125–127} However, these tests require specialized equipment and trained personnel.

There are no commercially available RDTs for the detection of MERS-CoV in humans. Chen and co-workers developed an immunochromatographic RDT for MERS-CoV detection in human samples for research purposes. Their device showed high specificity, but moderate sensitivity, 81%, with a detection limit that was 25–100 times less sensitive than ELISA.¹²⁸

Coronavirus disease-2019 (COVID-19) is a respiratory disease caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). This disease was first reported in December 2019 in Wuhan (China), where it caused an outbreak of atypical pneumonia.¹²⁹ Four months later, on March 11, 2020, a global pandemic was declared by the WHO.¹³⁰ As of March 16, 2021, COVID-19 had infected more than 120 million individuals worldwide with a death toll of approximately 2.6 million deaths reported to the WHO.^{131,132} Additionally, the COVID-19 outbreak has caused a global economic crisis due to the almost complete paralysis of all primary industries, activities, travel, and goods exchange.

SARS-CoV-2 is an enveloped positive-sense RNA virus. Its genome comprises around 30 kilobases. It belongs to the same *Coronaviridae* family as SARS-CoV and the Middle East respiratory syndrome virus (MERS). SARS-CoV-2 enters into target cells by using the spike (S) protein of coronaviruses. Entry depends on binding of the surface unit, S1, to a cellular receptor, facilitating viral attachment to target cells,¹³³ cf. Fig. 14. After membrane fusion and endocytosis, the viral

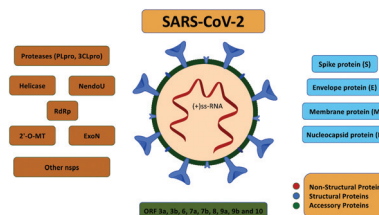


Fig. 14 SARS-CoV-2 encoded proteins reproduced from ref. 134 with permission from ACS copyright 2020.

nucleocapsid releases the genome payload into the infected cell's cytoplasm. Inside the infected cell, the virus forms numerous double-membrane vesicles that protect the viral genome, thus allowing its replication. After this process, the viral genomes are translated into viral polyproteins, which are then cleaved into viral proteins. Immediately after this, viral particles and virions assemble in the endoplasmic reticulum/Golgi compartment and are transported to the cell surface from which they are released *via* exocytosis and they continue to infect other cells.¹³⁴

It is unclear which animal was the intermediate host that transmitted the SARS-CoV-2 virus to humans, but bats and pangolin are among the main suspects. SARS-CoV-2 is highly contagious and can be transmitted from human-to-human by contact with saliva, tissue from infected individuals, and contaminated surfaces.

11.1 Symptoms

In most individuals infected with SARS-CoV-2, the disease manifests as a respiratory tract infection, with many different symptoms that are common to other infections, such as influenza. Due to this, a laboratory test is required to distinguish SARS-CoV-2 from other respiratory diseases. Among the reported symptoms are a cough, difficulty breathing, fever, sore throat, nasal congestion, headache, nausea or vomiting, muscle and joint pain, diarrhea, and a loss of taste and smell.^{135,136} Since SARS-CoV-2 is not fully understood, the list of symptoms is continuously being updated.¹¹

The majority of infected patients present mild symptoms and can recover at home without professional medical care. However, people with a medical history of diabetes, heart or lung disease, obesity, and older adults seem to be at higher risk of developing more severe complications. The symptoms may appear 2–14 days after exposure to the virus. The mortality rate for SARS-CoV-2 is around 1%.¹³⁷

11.2 Diagnostic testing

Due to the current emergency with SARS-CoV-2, pharmaceutical companies and research labs worldwide are focusing on developing quick and effective tests.^{138,139} Commercially available tests for the detection of SARS-CoV-2 can be classified into two major groups: molecular assays and immunological and serological assays.

Molecular assays focus on detecting viral RNA to identify infected individuals during the acute phase of infection. Around 90% of the available molecular assays use PCR or reverse transcription-PCR. The remaining 10% are based on isothermal amplification techniques, hybridization technologies, or CRISPR.^{140,141} One of the main advantages of these assays is the high sample throughput and, in some cases, an analysis time below 30 minutes.¹⁴² However, the tests are based on specialized equipment, skilled personnel, and high complexity labs.

Immunological and serological assays focus on detecting antibodies or antigenic proteins produced by individuals exposed to the virus. Immunological and serological RDTs are

usually paper-based lateral-flow assays, providing the advantage of transporting samples and reagents by capillary action, making the device more straightforward, and maintaining a low-cost per device. These tests provide a qualitative answer within 10–20 minutes. Currently, there are more than 200 commercially available rapid lateral flow immunoassays (LFIs) for the detection of SARS-CoV-2, detecting primarily antibody response.¹⁴³ From these 200 commercially available RDTs, only nine have received EUA by the US FDA.¹⁴⁴

Tests based on the detection of the virus antigen are, due to low sensitivity, not recommended by the WHO.¹⁴⁵ The sensitivity of antigen-based RDTs for respiratory diseases similar to SARS-CoV-2 varies from 34% to 80%. This variation in sensitivity means that more than half of infected individuals could be overlooked by such tests.¹⁴⁶

Additionally, the efficacy of rapid LFIs for SARS-CoV-2 has been questioned in the media during the last months.^{8,9} The incorrect results are attributed to the lack of specificity and sensitivity of several LFIs on the market due to the use of antibodies that are not specific enough or low-quality materials for their fabrication. These reports highlight the need to develop more robust and efficient RDTs controlled and validated by governmental regulator bodies before becoming available on the market.

Recently, a molecular-based RDT for COVID-19 self-testing at home got EUA approval by the FDA.¹⁴⁷ This RT-LAMP test kit developed by Lucira Health has a cost of approximately 50 USD and can detect SARS-CoV-2 RNA in nasal swabs in 30 minutes.¹⁴⁸ The user gets a positive or negative answer indicated by illumination of a light in the test unit display shown in Fig. 15.

12. Existing challenges to develop robust rapid diagnostic tests for use during disease outbreaks

The current SARS-CoV-2 pandemic that affects the whole world has exposed the lack of available RDTs to detect high-priority



Fig. 15 Lucira COVID-19 All-In-One test kit developed by Lucira Health for self-testing at home. It is the first molecular-based RDT approved by the FDA under EUA (image from <http://www.lucirahealth.com>).



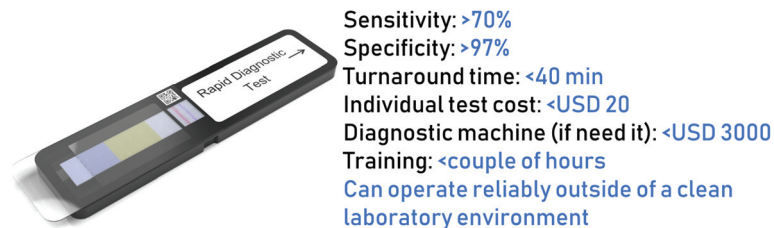


Fig. 16 Specifications for the ideal RDT for application during disease outbreak, according to criteria listed by the WHO.

pathogens during disease outbreaks. The number of available RDTs correlates with the geographic extension of disease outbreaks and their impact on developed countries' populations. Currently, there are no commercially available RDTs for the detection of LFV, RVF, and NV. Outbreaks connected to these viruses have affected areas located in undeveloped countries, with only a few cases reported in Europe or North America. Meanwhile, the number of commercially available RDTs for detecting EV, ZV, and COVID-19 is much higher. These outbreaks have affected not only undeveloped countries, but have also led to a large number of cases in Europe and North America, in the case of EV and ZV, and throughout the whole world, in the case of COVID-19.

Looking over the RDTs listed in Table 2, it is clear that most of these devices are based on a lateral-flow immunoassay platform. Lateral-flow immunoassay platforms are simple, low-cost, and provide a rapid response. Two types of immunoassays can be implemented in a lateral-flow platform. The first type is known as a sandwich assay, in which an antigen can be detected by the combination of an antibody pair, where the first antibody labeled with a colloidal gold nanoparticle binds to one antigen epitope, and a second capture antibody, immobilized on the lateral flow strip, binds to a second epitope on the same antigen. In this assay type, the antigen's concentration will be proportional to the signal intensity at the test line.

The second type of test is known as a competitive assay, and it is used for the detection of analytes using an antigen. As opposed to the sandwich assay detection, this test line's intensity is inversely proportional to the sample concentration in the competitive assay.

The fabrication of most LFIs involves combining several materials, *e.g.*, paper membranes, Au nanoparticles, antibodies, and antigens. Despite the overwhelming number of scientific reports and discussion of how the integration of LFIs with novel materials with intrinsic properties, such as carbon nanotubes,^{149,150} III-V nanostructures,^{151,152} and graphene,^{153–155} will disrupt the diagnostic market, only a few materials and techniques have made the transition from the lab bench to commercialization for disease detection during outbreaks. More efficient interactions between researchers in fields like immunology, biochemistry, materials science, diagnostics, and regulating bodies are required to accelerate the development of robust RDTs that fulfill the criteria listed by

the WHO for the ideal rapid detection device during disease outbreaks,¹³ Fig. 16.

One of the challenges when a new pandemic or outbreak occurs is the lack of time to develop new robust assays to detect the new pathogen. Ideally, it should be easy and quick to change RDT technology to accommodate new pathogens, to be used instantly in an occurring outbreak. However, development often takes time, especially when it is an entirely new pathogen where no preexisting data are present.

During an outbreak, health authorities must collect data from testing, patient details, geography, *etc.* This is to estimate the magnitude of outbreaks, transmission routes, exposure, underlying diseases of importance for the development of the disease, risk behaviors, *etc.* Epidemiological analyses and data collection are essential for implementing the preventive measures needed during an epidemic outbreak. Results of RDTs should ideally be reported in a way that the health authorities can use data. This could be done by implementing electronic reporting in the RDT technology, *e.g.*, by mobile phone data transfer.

Next, we discuss the areas that need further improvement to achieve more efficient RDTs for disease outbreaks.

12.1 Sensitivity

The majority of available RDTs for virus detection are based on the immobilization of antibodies against the specific virus on a surface acting as a transducer. The efficiency of binding between the antibody and the antigen will determine the assay's ultimate sensitivity. Three factors can influence this binding efficiency: the specificity of the antibody, the antibody's orientation, and the attachment of the antibody to the surface. These parameters influence the performance of RDTs based on LFIs, since they are critically dependent on the concentration of the analyte in a biological fluid. False-negative results from analyte concentrations below the LFI limit of detection are significant concerns.¹⁵⁶

These are critical factors that need to be considered when developing RDTs. Making available the structure of the virus or pathogen to be detected facilitates the development of antibodies. Recently, nanobodies have appeared as a promising novel class of therapeutic proteins based on single-domain antibody fragments containing the unique structural and functional properties of naturally occurring heavy-chain-only antibodies. These small structures are arousing increasing atten-



tion in immunosensing due to their higher binding specificity, greater thermal stability, and a lower tendency for aggregation than conventional antibodies.^{157,158}

Once a specific antibody or nanobody against a particular target is developed, its attachment and correct orientation must ensure efficient binding with the target pathogen. Several methods are available with the aim of obtaining a covalent attachment between the capturing molecule and the surface, acting as a transducer.^{159,160} The available immobilization methods can be classified as covalent and non-covalent, depending on the coupling chemistry used. Both methods aim to assure the sensing molecule's attachment to the surface without affecting its binding activity and specificity.

Antibodies and nanobodies can adopt different orientations when deposited on a surface. This orientation will affect the binding capacity of these glycoproteins, since their binding sites could entirely or partially face the substrate, thus affecting the immunoassay's performance.¹⁶¹

During the current COVID-19 pandemic, the high number of false negatives when using LFIs for antigen detection has accelerated the development of a rapid molecular diagnostic test with a shorter turnaround time and higher sensitivity.^{162,163}

Current efforts are focus on combining the high sensitivity of molecular diagnostic assays with the simplicity of lateral flow platforms. An example of this is the AmpliVue platform developed by Quidel.¹⁶⁴ This platform combines a rapid nucleic acid amplification that does not require a thermal cycler with the advantages of a lateral-flow-based cassette, avoiding the need for an electronic reader. The AmpliVue platforms are used to detect *Bordetella pertussis*, *Clostridium difficile*, group B streptococcus, Herpes simplex virus type 1 (HSV-1), and Herpes simplex virus type 2 (HSV-2). The turnaround time of the AmpliVue platform for the detection of HSV-1 is around one hour.

Another attempt to combine lateral flow with genetic analysis was recently developed by Wang and co-workers.¹⁶⁵ In their work, the CRISPR (clustered regularly interspaced short palindromic repeats)/Cas system was integrated into a lateral flow assay platform prepared using a nitrocellulose membrane. Using this hybrid platform, it was possible to identify the African swine fever virus in a non-laboratory environment from swine serum samples within 40 minutes.

Using a similar approach, Broughton and co-workers reported developing a rapid (<60 min) CRISPR-Cas12-based lateral flow assay for the detection of SARS-CoV-2 from respiratory swab RNA extracts.¹⁶⁶ The platform was validated using contrived reference samples and clinical samples from patients with COVID-19 infection and patients with other viral respiratory infections, obtaining 95% positive predictive agreement and 100% negative predictive agreement. This qualitative assay does not require bulky instrumentation and is currently waiting for EUA approval from the US FDA. Other examples of combining CRISPR with analytically portable and simple sensing platforms have recently been reviewed.¹⁶⁷

12.2 Quantitative answer

As shown in Table 2, most commercially available RDTs are based on the immunochromatographic technique using a paper-based membrane to immobilize the sensing chemistry and transport samples and reagents. The response of these devices is colorimetric, *i.e.*, there is a color change and an increase in the color intensity proportional to the sample concentration. Even though a couple of devices provide the option of transforming the change in color into a concentration value, most of the devices offer only a qualitative answer. This "yes" or "no" response might be insufficient or not sufficiently clear if the limit of detection is not clearly stated, *i.e.*, at what concentration the color change will occur. For these situations, a quantitative detection method will provide a more precise and robust answer. A natural combination to provide a simple answer to this challenge could be to use the cameras on mobile phones with lateral-flow assay devices.^{168,169}

Additionally, the latest advances in the development of apps and algorithms offer the possibility of incorporating this software in any mobile phone to obtain a concentration value from a picture of the RDT reaction area.¹⁷⁰ Moreover, the mobile phone will also add the option to detect and report additional data to health authorities, which is crucial from a surveillance point of view. These additional data are geographic location, temperature conditions, and time of the analysis, just to mention a few. Several publications reporting on the status of the combination of LFA and mobile phones are available,^{10,171–173} as well as the FDA-approved apps for the analysis of urine and wounds.¹⁷⁴ Using these types of apps for the quantitative detection of RDTs during disease outbreaks will provide the health authorities with more complete and faster information to facilitate the implementation of measurements aiming to stop the spread of the disease.

12.3 Alternatives to lateral flow and colorimetric detection

As mentioned previously, paper-based devices offer several advantages in terms of cost and simplicity for the development of RDTs. Additionally, their fabrication is simple and could be integrated with, for instance, conductive materials (gold, graphene) for the manufacture of more advanced electrochemical LF devices.^{171,175} These systems have been reported in the literature to develop point-of-care biomedical devices.¹⁷⁶ Despite the high number of existing articles, the amount of commercially available devices is still meager.

Since the irruption of electrochemical biosensors, these devices have been put forward as an ideal option for developing POC systems due to their presenting the possibility of miniaturization, integration with sensing molecules (enzymes, antibodies, aptamers, cells), sensitivity and specificity, and simplicity.¹⁷⁷ Still, after more than 20 years of research, only a handful of electrochemical biomedical sensors are commercially available and successful, of which the main one is the glucose biosensor based on an enzymatic reaction.^{178,179} Despite all the mentioned advantages offered by electrochemical sensors, none of the existing RDTs listed in Table 2



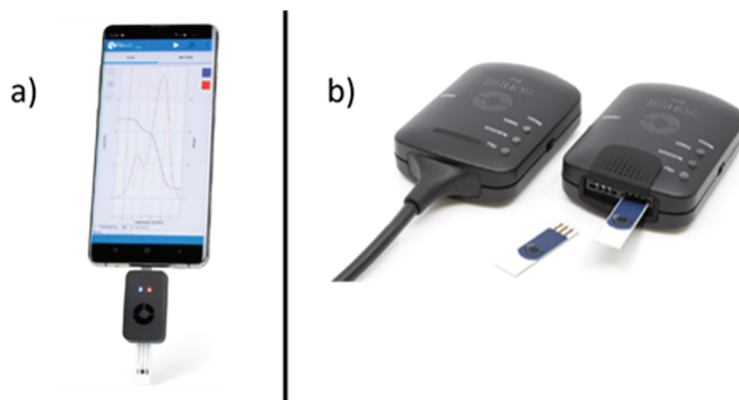


Fig. 17 (a) SenSit-smart miniaturized potentiostat connected to a mobile phone, and (b) Sensit-bt miniaturized wireless potentiostat developed by PalmSens (images from <http://www.palmsens.com>).

use electrochemistry as their detection technique. One of the reasons for this is that electrochemical sensors utilized in combination with immunochemistry are mainly based on impedance. This method requires the presence of a redox couple in a solution for the quantification of antigen concentration. Typically, a solution of ferro-/ferricyanide is used as the source for the redox couple that will be employed to quantify the amount of sample bound to the antibody.¹⁸⁰ This means that, in practice, the user will need to add the sample, wait for the binding between the antigen and the antibody to occur, and then add the redox solution to quantify the presence of the analyte. This extra step makes this method complex and not user-friendly, limiting its use as a point-of-care device.¹⁸¹ Recently, the use of thiols containing a tethered redox molecule was suggested as an alternative to immunoassays based on electrochemical impedance.^{182,183} The obtained results for the detection of the dengue virus showed that this technique had great potential for facilitating the development of RDTs for biomedical applications.^{184,185}

Recently, a hybrid approach combining electrochemistry detection with the trans-cleavage property of CRISPR, E-CRISPR, was presented by a multidisciplinary research team from Case Western Reserve University.¹⁸⁶ Using the E-CRISPR platform, viral nucleic acids, such as papillomavirus 16 (HPV-16) and parvovirus B19 (PB-19), were detected with picomolar sensitivity within one hour. This is another excellent example of nucleic acid assays combined with miniaturized electrochemical platforms and used outside the lab.

Another recent development that could facilitate the use of electrochemical sensors in the development of RDTs is the miniaturization of a potentiostat that can be integrated into smartphones,¹⁸⁷ cf. Fig. 17a. The smartphone potentiostat allows the evaluation of electrochemical sensors based on amperometric, voltammetric, pulsed techniques, and electrochemical impedance, of which the results can be stored and evaluated using a corresponding app on a mobile phone. PalmSens has developed a miniaturized handheld wireless potentiostat that uses the USB port of a smartphone or tablet

to communicate with and send data from an electrochemical sensor,¹⁸⁸ cf. Fig. 17b. This miniaturized potentiostat enables electrochemical analysis on-site with lower consumption of energy and with data transmission.

13. Conclusions and perspectives

The current SARS-CoV-2 pandemic and previous SARS and MERS pandemics demonstrated the crucial role of rapid testing devices as an essential tool for health authorities to obtain rapid and precise information that could be used to adopt quick decisions aiming to stop the spread of disease. Simultaneously, it exposed the high dependence on lateral-flow devices and colorimetric assays as the primary technique to develop rapid test devices, as defined in the introduction. Even though more precise molecular-based assays are developing rapidly and more user-friendly sensing platforms are emerging, these methods are not yet at the level of simplicity, low-cost and rapid turnaround of lateral-flow-based assay rapid test devices. However, recently a molecular-based RDT device for SARS-CoV-2 self-testing at home on nasal swabs received approval from the FDA. This RT-PCR self-testing device can detect SARS-CoV-2 in 30 minutes with 94% positive percent agreement and 98% negative percent agreement.^{148,189} This molecular-based RDT could be the first step towards developing more precise and straightforward RDTs that could hopefully be applied to detect other pathogens causing pandemics.

As shown in Table 2, due to the worldwide extension of the current SARS-CoV-2 pandemic, the development of new RDTs for SARS-CoV-2 received a tremendous boost from government and private organizations. This vast support resulted in the fast development of hundreds of new RDT devices that effectively provided rapid data to decrease the virus's spread, as demonstrated in New Zealand and South Korea. However, the situation is not the same for most of the pathogens identified by the WHO as potential causes of future pandemics.



As discussed, it is necessary to accelerate the integration of more robust techniques with the lateral-flow assay platform's simplicity. Techniques such as electrochemistry, optical, nanoplasmonics, and molecular-based assays have already demonstrated promising results in sensitivity, selectivity, and reproducibility. The irruption of new micro- and nano-fabrication techniques would definitively accelerate integrating these highly sensitive assays with low-cost and simple platforms. It is vital to maintain the simplicity, low-cost, and robustness to facilitate the distribution and use of these new platforms worldwide.

Quick turnaround RDTs should be effectively combined with the advantages of existing mobile technology. Effective communication of data for disease surveillance will be achieved by standardizing the recording and reporting of results. Existing mobile technology provides the possibilities of capturing, recording, and transmitting data using a single device in resource-limited locations. The development of applications that standardize the way data extracted from RDTs are recorded and reported will contribute to a rapid exchange of information between health organizations worldwide. Moreover, the use of the global positioning system integrated into mobile phones will link geographical location data (time, temperature, humidity) to the assay results, allowing the status of disease spread to be precisely mapped and strategies adopted to contain the advance of a particular disease.

Research into RDTs should be supported and conducted to develop these new and smarter technologies. The research should also include the critical perspective that a given technology should have the ability to shift focus if new pathogens occur swiftly. This is essential for immediate implementation and response in an outbreak situation.

Another critical factor that should be highlighted is more effective coordination between government agencies in charge of evaluating and approving new RDTs used in disease surveillance. This collaboration will decrease the release of commercially available RDTs that do not provide complete and reliable information, which delays practical measurements to prevent the disease from spreading.

All the actions suggested in this review should be implemented as soon as possible through coordinated efforts involving research groups, health authorities, and funding bodies. Climate change and closer interaction between humans and animals in previously unexplored environments contribute to the emergence of new diseases, resulting in epidemics and pandemics due to increased human mobility and goods trade.

Author contributions

Jaime Castillo-León: conceptualization, investigation, writing original draft, writing & review. Ramona Trebbien: conceptualization, investigation, writing original draft, writing & review. John J. Castillo: investigation, writing original draft, writing &

review. Winnie E. Svendsen: conceptualization, investigation, writing original draft, writing & review.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 D. M. Wagner, J. Klunk, M. Harbeck, A. Devault, N. Waglechner, J. W. Sahl, J. Enk, D. N. Birdsell, M. Kuch, C. Lumibao, D. Poinar, T. Pearson, M. Fourment, B. Golding, J. M. Riehm, D. J. D. Earn, S. DeWitte, J. M. Rouillard, G. Grupe, I. Wiechmann, J. B. Bliska, P. S. Keim, H. C. Scholz, E. C. Holmes and H. Poinar, *Lancet Infect. Dis.*, 2014, **14**, 319–326.
- 2 G. Vogel, *Science*, 2014, **345**, 1549–1550.
- 3 S. Haensch, R. Bianucci, M. Signoli, M. Rajerison, M. Schultz, S. Kacki, M. Vermunt, D. A. Weston, D. Hurst, M. Achtman, E. Carniel and B. Bramanti, *PLoS Pathog.*, 2010, **6**, e1001134.
- 4 S. A. Truelove, L. Mier-y-Teran-Romero, P. Gastanaduy, A. Taylor Walker, A. Berro, J. Lessler and M. Johansson, *medRxiv*, 2020, 10.1101/2020.05.08.20095414.
- 5 I. of M. (US) F. on M. Threats, S. Knobler, A. Mahmoud, S. Lemon and L. Pray, *The Impact of Globalization on Infectious Disease Emergence and Control*, National Academies Press, 2006.
- 6 J. Castillo-León, in *Lab-on-a-Chip Devices and Micro-Total Analysis Systems*, Springer International Publishing, 2015, pp. 1–15.
- 7 M. D. Perkins, C. Dye, M. Balasegaram, C. Bréchet, J. V. Mombouli, J. A. Røttingen, M. Tanner and C. C. Boehme, *Lancet*, 2017, **390**, 2211–2214.
- 8 Are We Failing the Coronavirus-Antibody Test? | The New Yorker, <https://www.newyorker.com/news/daily-comment/are-we-failing-the-coronavirus-antibody-test>, (accessed 14 July 2020).
- 9 Mayo Clinic doctors find many COVID-19 antibody tests fail their quality standards: ABC News exclusive - ABC News, <https://abcnews.go.com/US/mayo-clinic-doctors-find-covid-19-antibodytests/story?id=70803740>, (accessed 14 July 2020).
- 10 A. W. Martinez, S. T. Phillips, E. Carrilho, S. W. Thomas, H. Sindi and G. M. Whitesides, *Anal. Chem.*, 2008, **80**, 3699–3707.
- 11 D. A. Drew, L. H. Nguyen, C. J. Steves, C. Menni, M. Freydin, T. Varsavsky, C. H. Sudre, M. J. Cardoso,



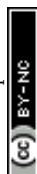
- S. Ourselin, J. Wolf, T. D. Spector and A. T. Chan, *Science*, 2020, **368**, 1362–1367.
- 12 Prioritizing diseases for research and development in emergency contexts, <https://www.who.int/activities/prioritizing-diseases-for-research-and-development-in-emergency-contexts>, (accessed 25 August 2020).
 - 13 WHO | Simple/Rapid tests, https://www.who.int/diagnostics_laboratory/faq/simple_rapid_tests/en/, (accessed 6 July 2020).
 - 14 L. Anfossi, C. Giovannoli and C. Baggiani, in *Rapid Test - Advances in Design, Format and Diagnostic Applications*, InTech, 2018.
 - 15 J. B. Lopez, D. Jackson, A. Gammie and T. Badrick, *Clin. Biochem. Rev.*, 2017, **38**, 3–11.
 - 16 S. McAlister, A. L. Barratt, K. J. Bell and F. McGain, *Med. J. Aust.*, 2020, **212**, 377–382.
 - 17 K. Ni, Y. Hu, X. Ye, H. S. AlZubi, P. Goddard and M. Alkahtani, *Energies*, 2018, **11**, 3105.
 - 18 T. Ozer, C. McMahon and C. S. Henry, *Annu. Rev. Anal. Chem.*, 2020, **13**, 85–109.
 - 19 T. Ozer, B. J. Geiss and C. S. Henry, *J. Electrochem. Soc.*, 2020, **167**, 037523.
 - 20 X. Ding, M. G. Mauk, K. Yin, K. Kadimisetty and C. Liu, *Anal. Chem.*, 2019, **91**, 655–672.
 - 21 K. H. Foyals, S. E. Seo, M. J. Kim, O. S. Kwon and J. W. Chong, *Sensors*, 2019, **19**, 4812.
 - 22 M. M. Gong and D. Sinton, *Chem. Rev.*, 2017, **117**, 8447–8480.
 - 23 P. Brangel, A. Sobarzo, C. Parolo, B. S. Miller, P. D. Howes, S. Gelkop, J. J. Lutwama, J. M. Dye, R. A. McKendry, L. Lobel and M. M. Stevens, *ACS Nano*, 2018, **12**, 63–73.
 - 24 J. D. Newman and A. P. F. Turner, *Biosens. Bioelectron.*, 2005, **20**, 2435–2453.
 - 25 Y. Chen, R. Ren, H. Pu, X. Guo, J. Chang, G. Zhou, S. Mao, M. Kron and J. Chen, *Sci. Rep.*, 2017, **7**, 10974.
 - 26 M. Soler, M. C. Estévez, M. Cardenosa-Rubio, A. Astua and L. M. Lechuga, *ACS Sens.*, 2020, **5**, 2663–2678.
 - 27 W. E. Svendsen, M. Jørgensen, L. Andresen, K. B. Andersen, M. B. B. Larsen, S. Skov and M. Dimaki, in *Procedia Engineering*, Elsevier, 2011, vol. 25, pp. 288–291.
 - 28 A. Zulfiqar, F. Patou, A. Pfreundt, C. Papakonstantinopoulos, W. E. Svendsen and M. Dimaki, *Sens. Bio-Sensing Res.*, 2017, **13**, 88–95.
 - 29 M. Park, R. S. Thwaites and P. J. M. Openshaw, *Eur. J. Immunol.*, 2020, **50**, 308–311.
 - 30 2009 H1N1 Pandemic (H1N1pdm09 virus) | Pandemic Influenza (Flu) | CDC, <https://www.cdc.gov/flu/pandemic-resources/2009-h1n1-pandemic.html>, (accessed 25 August 2020).
 - 31 M. G. Baker, N. Wilson and A. Anglemeyer, *N. Engl. J. Med.*, 2020, e56.
 - 32 Rapid COVID-19 testing breaks free from the lab - C&EN Digital Magazine, <https://cendigitalmagazine.acs.org/2020/08/14/rapid-covid-19-testing-breaks-free-from-the-lab-3/content.html>, (accessed 18 August 2020).
 - 33 B. J. Tromberg, T. A. Schwetz, E. J. Pérez-Stable, R. J. Hodes, R. P. Woychik, R. A. Bright, R. L. Fleurence and F. S. Collins, *Rapid Scaling Up of Covid-19 Diagnostic Testing in the United States-The NIH RADx Initiative*, 2020.
 - 34 R. Flick, in *Crimean-Congo Hemorrhagic Fever: A Global Perspective*, Springer Netherlands, 2007, pp. 35–44.
 - 35 H. Nasirian, *Acta Trop.*, 2019, **196**, 102–120.
 - 36 V. N. Raabe, *J. Clin. Microbiol.*, 2020, **58**, e01580-19.
 - 37 M. Zivcec, F. E. M. Scholte, C. F. Spiropoulou, J. R. Spengler and É. Bergeron, *Viruses*, 2016, **8**, 106.
 - 38 L. T. Mazzola and C. Kelly-Cirino, *BMJ Glob. Health*, 2019, **4**, 1114.
 - 39 W. Wu, S. Zhang, J. Qu, Q. Zhang, C. Li, J. Li, C. Jin, M. Liang and D. Li, *Virus Res.*, 2014, **187**, 84–90.
 - 40 V. Baniasadi, M. H. Pouriayeali, T. Jalali, M. Fazlalipour, K. Azadmanesh and M. Salehi-Vaziri, *J. Virol. Methods*, 2019, **265**, 49–52.
 - 41 R. Reece, M. A. Smit and T. P. Flanagan, in *Encyclopedia of Immunobiology*, Elsevier Inc., 2016, vol. 4, pp. 355–362.
 - 42 Outbreaks | Ebola (Ebola Virus Disease) | CDC, <https://www.cdc.gov/vhf/ebola/outbreaks/index-2018.html>, (accessed 6 October 2020).
 - 43 E. Rose, in *Life-Threatening Rashes*, Springer International Publishing, Cham, 2018, pp. 291–300.
 - 44 E. M. Leroy, A. Epelboin, V. Mondonge, X. Pourrut, J. P. Gonzalez, J. J. Muyembe-Tamfum and P. Formenty, *Vector-Borne Zoonotic Dis.*, 2009, **9**, 723–728.
 - 45 E. M. Leroy, B. Kumulungui, X. Pourrut, P. Rouquet, A. Hassanin, P. Yaba, A. Délicat, J. T. Paweska, J. P. Gonzalez and R. Swanepoel, *Nature*, 2005, **438**, 575–576.
 - 46 H. Feldmann and T. W. Geisbert, *Lancet*, 2011, **377**, 849–862.
 - 47 S. Su, G. Wong, X. Qiu, G. Kobinger, Y. Bi and J. Zhou, *Lancet Infect. Dis.*, 2016, **16**, 294–295.
 - 48 First FDA-approved vaccine for the prevention of Ebola virus disease, marking a critical milestone in public health preparedness and response | FDA, <https://www.fda.gov/news-events/press-announcements/first-fda-approved-vaccine-prevention-ebola-virus-disease-marking-critical-milestone-public-health>, (accessed 7 July 2020).
 - 49 OraQuick Ebola Rapid Antigen Test, kit/25, <https://supply.unicef.org/s0003883.html>, (accessed 7 July 2020).
 - 50 M. J. Broadhurst, J. D. Kelly, A. Miller, A. Semper, D. Bailey, E. Groppelli, A. Simpson, T. Brooks, S. Hula, W. Nyoni, A. B. Sankoh, S. Kanu, A. Jalloh, Q. Ton, N. Sarchet, P. George, M. D. Perkins, B. Wonderly, M. Murray and N. R. Pollock, *Lancet*, 2015, **386**, 867–874.
 - 51 ReBOV Antigen Rapid Test Kit/50, <https://supply.unicef.org/s0003750.html>, (accessed 7 July 2020).
 - 52 M. Letko, S. N. Seifert, K. J. Olival, R. K. Plowright and V. J. Munster, *Nat. Rev. Microbiol.*, 2020, **18**, 461–471.
 - 53 V. Soman Pillai, G. Krishna and M. Valiya Veettil, *Viruses*, 2020, **12**, 465.
 - 54 R. Ojha, A. Pareek, R. K. Pandey, D. Prusty and V. K. Prajapati, *ACS Omega*, 2019, **4**, 13069–13079.



- 55 R. K. McLean and S. P. Graham, *Front. Vet. Sci.*, 2019, **6**, 16.
- 56 S. P. Luby, *Antiviral Res.*, 2013, **100**, 38–43.
- 57 A. Tamin, B. H. Harcourt, T. G. Ksiazek, P. E. Rollin, W. J. Bellini and P. A. Rota, *Virology*, 2002, **296**, 190–200.
- 58 J. J. Sejvar, J. Hossain, S. K. Saha, E. S. Gurley, S. Banu, J. D. Hamadani, M. A. Faiz, F. M. Siddiqui, Q. D. Mohammad, A. H. Mollah, R. Uddin, R. Alam, R. Rahman, C. T. Tan, W. Bellini, P. Rota, R. F. Breiman and S. P. Luby, *Ann. Neurol.*, 2007, **62**, 235–242.
- 59 L. T. Mazzola and C. Kelly-Cirino, *BMJ Glob. Health*, 2019, **4**, e001118.
- 60 Nipah virus, <https://www.who.int/news-room/fact-sheets/detail/nipah-virus>, (accessed 7 October 2020).
- 61 M. Mingyi and L. Li, *A novel pen A novel pen-side strip test side strip test for the detection of for the detection of Nipah Nipah virus in swine virus in swine*, http://cahln-rectlsa.com/wp-content/uploads/2014/11/mingyi_li.pdf.
- 62 Rift Valley Fever, <https://www.cdc.gov/vhf/rvf/RVF-FactSheet.pdf>, (accessed 9 October 2020).
- 63 *Modern Infectious Disease Epidemiology - Concepts, Methods, Mathematical Models, and Public Health*, ed. A. Krämer, Springer, 2010.
- 64 R. B. Mandell and R. Flick, *Hum. Vaccin.*, 2010, **6**, 597–601.
- 65 V. Chevalier, *Clin. Microbiol. Infect.*, 2013, **19**, 705–708.
- 66 B. Bazanow, D. Stygar, E. Romuk, B. Skrzep-Poloczek, J. Pacoń, Ł. Gadzała, M. Welz and J. Paweńska, *J. Vector Borne Dis.*, 2018, **55**, 324.
- 67 J. Bingham and P. Jansen van Vuren, *Microbiol. Aust.*, 2020, **41**, 28.
- 68 J. Gonzalez-Valdivieso, B. Borrego, A. Girotti, S. Moreno, A. Brun, J. F. Bermejo-Martin and F. J. Arias, *Mol. Pharm.*, 2020, **17**, 1608–1620.
- 69 K. L. Mansfield, A. C. Banyard, L. McElhinney, N. Johnson, D. L. Horton, L. M. Hernández-Triana and A. R. Fooks, *Vaccine*, 2015, **33**, 5520–5531.
- 70 C. A. Le Roux, T. Kubo, A. A. Grobbelaar, P. J. Van Vuren, J. Weyer, L. H. Nel, R. Swanepoel, K. Morita and J. T. Paweska, *J. Clin. Microbiol.*, 2009, **47**, 645–651.
- 71 C. Drosten, S. Götting, S. Schilling, M. Asper, M. Panning, H. Schmitz and S. Günther, *J. Clin. Microbiol.*, 2002, **40**, 2323–2330.
- 72 P. Jansen van Vuren and J. T. Paweska, *J. Virol. Methods*, 2009, **157**, 15–24.
- 73 J. T. Paweska, F. J. Burt, F. Anthony, S. J. Smith, A. A. Grobbelaar, J. E. Croft, T. G. Ksiazek and R. Swanepoel, *J. Virol. Methods*, 2003, **113**, 103–112.
- 74 B. Faburay, W. C. Wilson, A. Secka, B. Drolet, D. Scott McVey and J. A. Richt, *J. Clin. Microbiol.*, 2019, **57**, e01058-19.
- 75 C. Cêtre-Sossah, A. Pédarrieu, M. Juremalm, P. Jansen Van Vuren, A. Brun, A. B. Ould EL Mamy, J.-M. Héraud, C. Filippone, J.-P. Ravalohery, H. Chaabihi, E. Albina, L. Dommergues, J. Paweska and E. Cardinale, *PLoS Negl. Trop. Dis.*, 2019, **13**, e0007700.
- 76 N. E. Yun and D. H. Walker, *Viruses*, 2012, **4**, 2031–2048.
- 77 O. Ogbu, in *Encyclopedia of Food Safety*, Elsevier, 2014, vol. 2, pp. 208–213.
- 78 D. S. Grant, H. Khan, J. Schieffelin and D. G. Bausch, in *Emerging Infectious Diseases: Clinical Case Studies*, Elsevier Inc., 2014, pp. 37–59.
- 79 E. L. Hamblion, P. Raftery, A. Wendland, E. Dweh, G. S. Williams, R. N. C. George, L. Soro, V. Katawera, P. Clement, A. N. Gasasira, E. Musa and T. K. Nagbe, *Int. J. Infect. Dis.*, 2018, **66**, 65–73.
- 80 D. M. Emperador, S. A. Yimer, L. T. Mazzola, G. Norheim and C. Kelly-Cirino, *BMJ Glob. Health*, 2019, **4**, e001119.
- 81 O. Azeez-Akande, *African J. Clin. Exp. Microbiol.*, 2016, **17**, 282.
- 82 V. Raabe and J. Koehler, *J. Clin. Microbiol.*, 2017, **55**, 1629–1637.
- 83 A. N. Happi, C. T. Happi and R. J. Schoepp, *Curr. Opin. Virol.*, 2019, **37**, 132–138.
- 84 L. T. Mazzola and C. Kelly-Cirino, *BMJ Glob. Health*, 2019, **4**, 1116.
- 85 Lassa Diagnostic Kits – Zolgen Labs, <https://www.zolgen.com/product-category/vhf-diagnostic-kits/lassa-diagnostic-kits/>, (accessed 7 October 2020).
- 86 M. L. Boisen, E. Uyigwe, J. Aiyepada, K. J. Siddle, L. Oestereich, D. K. S. Nelson, D. J. Bush, M. M. Rowland, M. L. Heinrich, P. Eromon, A. T. Kayode, I. Odia, D. I. Adomeh, E. B. Muoebonam, P. Akhilomen, G. Okonofua, B. Osiemi, O. Omoregie, M. Airende, J. Agbukor, S. Ehikhametalar, C. O. Aire, S. Duraffour, M. Pahlmann, W. Böhm, K. G. Barnes, S. Mehta, M. Momoh, J. D. Sandi, A. Goba, O. A. Folarin, E. Ogbaini-Emovan, D. A. Asogun, E. A. Tobin, G. O. Akpede, S. A. Okogbenin, P. O. Okokhere, D. S. Grant, J. S. Schieffelin, P. C. Sabeti, S. Günther, C. T. Happi, L. M. Branco and R. F. Garry, *Sci. Rep.*, 2020, **10**, 1–14.
- 87 M. L. Boisen, J. N. Hartnett, J. G. Shaffer, A. Goba, M. Momoh, J. D. Sandi, M. Fullah, D. K. S. Nelson, D. J. Bush, M. M. Rowland, M. L. Heinrich, A. P. Koval, R. W. Cross, K. G. Barnes, A. E. Lachenauer, A. E. Lin, M. Nekoui, D. Kotliar, S. M. Winnicki, K. J. Siddle, M. Gbakie, M. Fonnies, V. J. Koroma, L. Kanneh, P. C. Kulakosky, K. M. Hastie, R. B. Wilson, K. G. Andersen, O. O. Folarin, C. T. Happi, P. C. Sabeti, T. W. Geisbert, E. O. Saphire, S. H. Khan, D. S. Grant, J. S. Schieffelin, L. M. Branco and R. F. Garry, *Sci. Rep.*, 2018, **8**, 17.
- 88 J. N. Maslow and C. C. Roberts, in *Methods in Molecular Biology*, Humana Press Inc., 2020, vol. 2142, pp. 1–8.
- 89 J. J. Waggoner and B. A. Pinsky, *J. Clin. Microbiol.*, 2016, **54**, 860–867.
- 90 H. Song, J. Qi, J. Haywood, Y. Shi and G. F. Gao, DOI: 10.1038/nsmb.3213.
- 91 J. M. Mansuy, M. Dutertre, C. Mengelle, C. Fourcade, B. Marchou, P. Delobel, J. Izopet and G. Martin-Blondel, *Lancet Infect. Dis.*, 2016, **16**, 405.
- 92 M. H. Collins and J. J. Waggoner, *ACS Infect. Dis.*, 2019, **5**, 1055–1069.



- 93 E. S. Theel and D. Jane Hata, *J. Clin. Microbiol.*, DOI: 10.1128/JCM.01972-17.
- 94 *Zika Virus RNA Qualitative Real-Time RT-PCR For use under an Emergency Use Authorization only Rx Only Instructions For Use*, 2017, <https://testdirectory.questdiagnostics.com/test/test-detail/93870/zika-virus-rna-qualitative-real-time-rt-pcr?cc=MASTER>.
- 95 P. Ren, D. A. Ortiz, A. C. B. Terzian, T. E. Colombo, M. L. Nogueira, N. Vasilakis and M. J. Loeffelholz, *J. Clin. Microbiol.*, 2017, **55**, 2198–2203.
- 96 ZIKV Detect™ IgM 2.0 Capture ELISA Kit (USA) – InBios International, Inc., <https://inbios.com/zikv-detecttm-igm-capture-elisa-kit-usa/>, (accessed 7 October 2020).
- 97 A. Goncalves, R. W. Peeling, M. C. Chu, D. J. Gubler, A. M. De Silva, E. Harris, M. Murtagh, A. Chua, W. Rodriguez, C. Kelly, A. Wilder-Smith and A. Wilder-Smith, *J. Infect. Dis.*, 2018, **1060**, 1060–1068.
- 98 F. A. Jorge, M. V. Thomazella, D. Castro Moreira, L. D. G. Lopes, J. J. V. Teixeira and D. A. Bertolini, *Rev. Med. Virol.*, 2020, **30**, e2105.
- 99 I. Bosch, H. De Puig, M. Hiley, M. Carré-Camps, F. Perdomo-Celis, C. F. Narváez, D. M. Salgado, D. Senthooor, M. O. Grady, E. Phillips, A. Durbin, D. Fandos, H. Miyazaki, C. W. Yen, M. Gélvez-Ramírez, R. V. Warke, L. S. Ribeiro, M. M. Teixeira, R. P. Almeida, J. E. Muñoz-Medina, J. E. Ludert, M. L. Nogueira, T. E. Colombo, A. C. B. Terzian, P. T. Bozza, A. S. Calheiros, Y. R. Vieira, G. Barbosa-Lima, A. Vizzoni, J. Cerbino-Neto, F. A. Bozza, T. M. L. Souza, M. R. O. Trugilho, A. M. B. De Filippis, P. C. De Sequeira, E. T. A. Marques, T. Magalhaes, F. J. Díaz, B. N. Restrepo, K. Marín, S. Mattar, D. Olson, E. J. Asturias, M. Lucera, M. Singla, G. R. Medigeshi, N. De Bosch, J. Tam, J. Gómez-Márquez, C. Clavet, L. Villar, K. Hamad-Schifferli and L. Gehrke, *Sci. Transl. Med.*, 2017, **9**, 29.
- 100 Chembio | DPP® Zika IgM System, <http://chembio.com/dpp-zika-igm-system/>, (accessed 10 July 2020).
- 101 Artron | Zika Test Kit, <http://www.artronlab.com/products/new/zika-test.html>, (accessed 8 October 2020).
- 102 ZIKA Rapid Test, <https://www.accubiotech.com/product-list-zika-rapid-test.html>, (accessed 8 October 2020).
- 103 Infectious Diseases - Lumiquick Diagnostics, Inc | Lumiquick Diagnostics, Inc., <https://lumiquick.co/infectious-diseases>, (accessed 8 October 2020).
- 104 CDIA™ Zika IgG/IgM Rapid Test Kit - Creative Diagnostics, https://www.cd-diatest.com/cdia-sup-tm-sup-zika-igg-igm-rapid-test-kit_p570.html, (accessed 8 October 2020).
- 105 Zika Virus Rapid Test Kit - Maternova Inc., <https://maternova.net/products/zika-rapid-test>, (accessed 8 October 2020).
- 106 Zika Rapid Test Kits - Biopanda Reagents, https://www.biopanda.co.uk/php/products/rapid/infectious_diseases/zika.php, (accessed 8 October 2020).
- 107 ProDetect® Zika Rapid Test | Mediven - Diagnostics Solutions - COVID-19 Test Kits, <http://mediven.com.my/index.php/catalog/prodetect-zika-rapid-test/>, (accessed 8 October 2020).
- 108 Zika IgG/IgM virus Antigen Rapid Test - Rapikit, <https://rapikit.com/products/zika-iggigm-virus-antigen-rapid-test/>, (accessed 8 October 2020).
- 109 A. Priye, S. W. Bird, Y. K. Light, C. S. Ball, O. A. Negrete and R. J. Meagher, DOI: 10.1038/srep44778.
- 110 A. Kaushik, A. Yndart, S. Kumar, R. D. Jayant, A. Vashist, A. N. Brown, C. Z. Li and M. Nair, *Sci. Rep.*, 2018, **8**, 1–5.
- 111 A. Kaushik, S. Tiwari, R. D. Jayant, A. Vashist, R. Nikkhah-Moshaie, N. El-Hage and M. Nair, *Trends Biotechnol.*, 2017, **35**, 308–317.
- 112 A. Kaushik, S. Tiwari, R. Dev Jayant, A. Marty and M. Nair, *Biosens. Bioelectron.*, 2016, **75**, 254–272.
- 113 H. A. Mohd, J. A. Al-Tawfiq and Z. A. Memish, *Virol. J.*, 2016, **13**, 87.
- 114 A. S. Omrani, J. A. Al-Tawfiq and Z. A. Memish, *Pathog. Glob. Health*, 2015, **109**, 354–362.
- 115 N. Petrosillo, G. Viceconte, O. Ergonul, G. Ippolito and E. Petersen, *Clin. Microbiol. Infect.*, 2020, **26**, 729.
- 116 E. Qing, M. P. Hantak, G. G. Galpalli and T. Gallagher, in *Methods in Molecular Biology*, Humana Press Inc., 2020, vol. 2099, pp. 9–20.
- 117 J. A. Al-Tawfiq and Z. A. Memish, in *Emerging Infectious Diseases: Clinical Case Studies*, Elsevier Inc., 2014, pp. 185–190.
- 118 X. Tao, T. E. Hill, C. Morimoto, C. J. Peters, T. G. Ksiazek and C.-T. K. Tseng, *J. Virol.*, 2013, **87**, 9953–9958.
- 119 I. M. Mackay and K. E. Arden, *Virol. J.*, 2015, **12**, 222.
- 120 A. Assiri, A. McGeer, T. M. Perl, C. S. Price, A. A. Al Rabeeah, D. A. T. Cummings, Z. N. Alabdullatif, M. Assad, A. Almulhim, H. Makhdoom, H. Madani, R. Alhakeem, J. A. Al-Tawfiq, M. Cotten, S. J. Watson, P. Kellam, A. I. Zumla and Z. A. Memish, *N. Engl. J. Med.*, 2013, **369**, 407–416.
- 121 Z. A. Memish and J. A. Al-Tawfiq, *Am. J. Infect. Control*, 2014, **42**, 1258–1260.
- 122 Z. A. Memish, A. M. Assiri and J. A. Al-Tawfiq, *Int. J. Infect. Dis.*, 2014, **29**, 307–308.
- 123 C. D. Kelly-Cirino, J. Nkengasong, H. Kettler, I. Tongio, F. Gay-Andrieu, C. Escadafal, P. Piot, R. W. Peeling, R. Gadde and C. Boehme, *BMJ Glob. Health*, 2019, **4**, 1179.
- 124 S. Al Johani and A. H. Hajeer, *J. Infect. Public Health*, 2016, **9**, 216–219.
- 125 Y. Chen, K.-H. Chan, Y. Kang, H. Chen, H. K. Luk, R. W. Poon, J. F. Chan, K.-Y. Yuen, N. Xia, S. K. Lau and P. C. Woo, *Emerg. Microbes Infect.*, 2015, **4**, 26.
- 126 J. F. W. Chan, S. Sridhar, C. C. Y. Yip, S. K. P. Lau and P. C. Y. Woo, *J. Microbiol.*, 2017, **55**, 172–182.
- 127 R. Perera, P. Wang, M. Gomaa, R. El-Shesheny, A. Kandeil, O. Bagato, L. Siu, M. Shehata, A. Kayed, Y. Moatasim, M. Li, L. Poon, Y. Guan, R. Webby, M. Ali, J. Peiris and G. Kayali, *Eurosurveillance*, 2013, **18**, 20574.
- 128 Y. Chen, K. H. Chan, C. Hong, Y. Kang, S. Ge, H. Chen, E. Y. M. Wong, S. Joseph, N. G. Patteril, U. Wernery, N. Xia, S. K. P. Lau and P. C. Y. Woo, *J. Infect.*, 2016, **73**, 82–84.
- 129 F. Wu, S. Zhao, B. Yu, Y. M. Chen, W. Wang, Z. G. Song, Y. Hu, Z. W. Tao, J. H. Tian, Y. Y. Pei, M. L. Yuan,



- Y. L. Zhang, F. H. Dai, Y. Liu, Q. M. Wang, J. J. Zheng, L. Xu, E. C. Holmes and Y. Z. Zhang, *Nature*, 2020, **579**, 265–269.
- 130 Archived: WHO Timeline - COVID-19, <https://www.who.int/news-room/detail/27-04-2020-who-timeline--covid-19>, (accessed 7 October 2020).
- 131 WHO Coronavirus Disease (COVID-19) Dashboard | WHO Coronavirus Disease (COVID-19) Dashboard, https://covid19.who.int/?gclid=CjwKCAjwr7X4BRA4EiwAUXjbt46IjGAWI4MuFY7U11TK1j9x3RoBIXX_x8I7sRflgVp91yqcdtfaZRoC-CAQAvD_BwE, (accessed 7 October 2020).
- 132 COVID-19 Map - Johns Hopkins Coronavirus Resource Center, <https://coronavirus.jhu.edu/map.html>, (accessed 7 October 2020).
- 133 M. Hoffmann, H. Kleine-Weber, S. Schroeder, M. A. Mü, C. Drosten and S. Pö, *Cell*, 2020, **181**, 271–280.
- 134 E. E. A. Osman, P. L. Toogood and N. Neamati, *ACS Infect. Dis.*, 2020, **6**, 1548–1552.
- 135 C. Menni, A. M. Valdes, M. B. Freidin, C. H. Sudre, L. H. Nguyen, D. A. Drew, S. Ganesh, T. Varsavsky, M. J. Cardoso, J. S. El-Sayed Moustafa, A. Visconti, P. Hysi, R. C. E. Bowyer, M. Mangino, M. Falchi, J. Wolf, S. Ourselin, A. T. Chan, C. J. Steves and T. D. Spector, *Nat. Med.*, 2020, **26**, 1037–1040.
- 136 C. Menni, C. H. Sudre, C. J. Steves, S. Ourselin and T. D. Spector, *Lancet*, 2020, **395**, e107–e108.
- 137 Y. Roussel, A. Giraud-Gatineau, M.-T. Jimeno, J.-M. Rolain, C. Zandotti, P. Colson and D. Raoult, *Int. J. Antimicrob. Agents*, 2020, **55**, 105947.
- 138 B. Udugama, P. Kadhiresan, H. N. Kozlowski, A. Malekjahani, M. Osborne, V. Y. C. Li, H. Chen, S. Mubareka, J. B. Gubbay and W. C. W. Chan, *ACS Nano*, 2020, **14**, 3822–3835.
- 139 L. J. Carter, L. V. Garner, J. W. Smoot, Y. Li, Q. Zhou, C. J. Saveson, J. M. Sasso, A. C. Gregg, D. J. Soares, T. R. Beskid, S. R. Jervey and C. Liu, *ACS Cent. Sci.*, 2020, **6**, 591–605.
- 140 M. Mahmoudi, DOI: 10.1021/acs.molpharmaceut.0c00371.
- 141 D. Administration, *FDA Combating COVID-19 With Medical Devices*, 2020.
- 142 Home - Credo Diagnostics Biomedical Pte. Ltd, <https://www.credodxbiomed.com/en/>, (accessed 7 October 2020).
- 143 SARS-CoV-2 diagnostic pipeline - FIND, https://www.finddx.org/covid-19/pipeline/?avance=Commercialized&type=Rapid+diagnostic+tests&test_target=all&status=all§ion=show-all&action=default, (accessed 7 October 2020).
- 144 In Vitro Diagnostics EUAs | FDA, <https://www.fda.gov/medical-devices/coronavirus-disease-2019-covid-19-emergency-use-authorizations-medical-devices/vitro-diagnostics-euas#isft1>, (accessed 7 October 2020).
- 145 Advice on the use of point-of-care immunodiagnostic tests for COVID-19, <https://www.who.int/news-room/commentaries/detail/advice-on-the-use-of-point-of-care-immunodiagnostic-tests-for-covid-19>, (accessed 7 October 2020).
- 146 A. H. L. Bruning, M. M. G. Leeflang, J. M. B. W. Vos, R. Spijker, M. D. De Jong, K. C. Wolthers and D. Pajkrt, *Clin. Infect. Dis.*, 2017, **1026**, 1026–1058.
- 147 Coronavirus (COVID-19) Update: FDA Authorizes First COVID-19 Test for Self-Testing at Home | FDA, <https://www.fda.gov/news-events/press-announcements/coronavirus-covid-19-update-fda-authorizes-first-covid-19-test-self-testing-home>, (accessed 20 November 2020).
- 148 COVID-19 At-home Test - Lucira Health, <https://www.lucirahealth.com/>, (accessed 20 November 2020).
- 149 Y. Huang, Y. Wen, K. Baryeh, S. Takalkar, M. Lund, X. Zhang and G. Liu, *Microchim. Acta*, 2017, **184**, 4287–4294.
- 150 A. Abera and J. W. Choi, *Anal. Methods*, 2010, **2**, 1819–1822.
- 151 C. Gui, K. Wang, C. Li, X. Dai and D. Cui, *Nanoscale Res. Lett.*, 2014, **9**, 1–8.
- 152 Y. J. Xie, Y. Yang, W. J. Kong, S. H. Yang and M. H. Yang, *Chin. J. Anal. Chem.*, 2015, **43**, 618–628.
- 153 V. Shirshahi, S. N. Tabatabaei, S. Hatamie and R. Saber, *J. Pharm. Biomed. Anal.*, 2019, **164**, 104–111.
- 154 E. Morales-Narváez, T. Naghdi, E. Zor and A. Merkoçi, *Anal. Chem.*, 2015, **87**, 8573–8577.
- 155 R. Álvarez-Díduk, J. Orozco and A. Merkoçi, *Sci. Rep.*, 2017, **7**, 1–9.
- 156 T. R. Kozel and A. R. Burnham-Marusch, *J. Clin. Microbiol.*, 2017, **55**, 2313–2320.
- 157 L. Huang, S. Muyldermans and D. Saerens, *Expert Rev. Mol. Diagn.*, 2010, **10**, 777–785.
- 158 Y. Wang, Z. Fan, L. Shao, X. Kong, X. Hou, D. Tian, Y. Sun, Y. Xiao and L. Yu, *Int. J. Nanomed.*, 2016, **11**, 3287–3303.
- 159 A. K. Trilling, J. Beekwilder and H. Zuillhof, *Analyst*, 2013, **138**, 1619–1627.
- 160 N. G. Welch, J. A. Scoble, B. W. Muir and P. J. Pigram, *Biointerphases*, 2017, **12**, 02D301.
- 161 J. Hu, M. Gao, Z. Wang, Y. Chen, Z. Song and H. Xu, *Appl. Nanosci.*, 2020, **1**, 3.
- 162 J. Van Elslande, E. Houben, M. Depypere, A. Brackenier, S. Desmet, E. Andr, M. Van Ranst, K. Lagrou and P. Vermeersch, *Clin. Microbiol. Infect.*, 2020, **26**, 1082.
- 163 S. J. C. Pallett, M. Rayment, A. Patel, S. A. M. Fitzgerald-Smith, S. J. Denny, E. Charani, A. L. Mai, K. C. Gilmour, J. Hatcher, C. Scott, P. Randell, N. Mughal, R. Jones, L. S. P. Moore and G. W. Davies, *Lancet Respir. Med.*, 2020, **8**, 885–894.
- 164 AmpliVue | Quidel, <https://www.quidel.com/molecular-diagnostics/amplivue-products>, (accessed 23 September 2020).
- 165 X. Wang, E. Xiong, T. Tian, M. Cheng, W. Lin, H. Wang, G. Zhang, J. Sun and X. Zhou, *ACS Nano*, 2020, **14**, 2497–2508.
- 166 J. P. Broughton, X. Deng, G. Yu, C. L. Fasching, V. Servellita, J. Singh, X. Miao, J. A. Streithorst, A. Granados, A. Sotomayor-Gonzalez, K. Zorn, A. Gopez, E. Hsu, W. Gu, S. Miller, C. Y. Pan, H. Guevara, D. A. Wadford, J. S. Chen and C. Y. Chiu, *Nat. Biotechnol.*, 2020, **38**, 870–874.
- 167 J. E. van Dongen, J. T. W. Berendsen, R. D. M. Steenbergen, R. M. F. Wolthuis, J. C. T. Eijkel and L. I. Segerink, *Biosens. Bioelectron.*, 2020, **166**, 112445.



- 168 A. Carrio, C. Sampedro, J. Sanchez-Lopez, M. Pimienta and P. Campoy, *Sensors*, 2015, **15**, 29569–29593.
- 169 C. Ruppert, N. Phogat, S. Laufer, M. Kohl and H. P. Deigner, *Microchim. Acta*, 2019, **186**, 1–9.
- 170 L. Pezzarossa, S. I. Preus, W. E. Svendsen and J. Madsen, *A Computer Vision Algorithm for the Digitalization of Colorimetric Lateral Flow Assay Readouts*, Institute of Electrical and Electronics Engineers (IEEE), 2020, pp. 1–6.
- 171 C. Carrell, A. Kava, M. Nguyen, R. Menger, Z. Munshi, Z. Call, M. Nussbaum and C. Henry, *Microelectron. Eng.*, 2019, **206**, 45–54.
- 172 D. C. Christodouleas, B. Kaur and P. Chorti, *ACS Cent. Sci.*, 2018, **4**, 1600–1616.
- 173 W. Xiao, C. Huang, F. Xu, J. Yan, H. Bian, Q. Fu, K. Xie, L. Wang and Y. Tang, *Sens. Actuators, B*, 2018, **266**, 63–70.
- 174 Healthy.io | Turning the smartphone into a medical device, <https://healthy.io/>, (accessed 4 August 2020).
- 175 G. Ruiz-Vega, M. Kitsara, M. A. Pellitero, E. Baldrich and F. J. del Campo, *ChemElectroChem*, 2017, **4**, 880–889.
- 176 P. D. Sinawang, V. Rai, R. E. Ionescu and R. S. Marks, *Biosens. Bioelectron.*, 2016, **77**, 400–408.
- 177 J. Wang, in *Biosensors and Bioelectronics*, Elsevier, 2006, vol. 21, pp. 1887–1892.
- 178 Y. Dai and C. C. Liu, *Angew. Chem.*, 2019, **131**, 12483–12496.
- 179 E. T. S. G. da Silva, D. E. P. Souto, J. T. C. Barragan, J. de F. Giarola, A. C. M. de Moraes and L. T. Kubota, *ChemElectroChem*, 2017, **4**, 778–794.
- 180 X. Chen, Y. Wang, J. Zhou, W. Yan, X. Li and J. J. Zhu, *Anal. Chem.*, 2008, **80**, 2133–2140.
- 181 T. Bertok, L. Lorencova, E. Chocholova, E. Jane, A. Vikartovska, P. Kasak and J. Tkac, *ChemElectroChem*, 2019, **6**, 989–1003.
- 182 R. M. B. Oliveira, F. C. B. Fernandes and P. R. Bueno, *Electrochim. Acta*, 2019, **306**, 175–184.
- 183 P. R. Bueno, in *Nanobiosensors for Personalized and Onsite Biomedical Diagnosis*, Institution of Engineering and Technology, 2016, pp. 293–316.
- 184 J. Cecchetto, A. Santos, A. Mondini, E. M. Cilli and P. R. Bueno, *Biosens. Bioelectron.*, 2020, **151**, 111972.
- 185 J. Cecchetto, F. C. B. Fernandes, R. Lopes and P. R. Bueno, *Biosens. Bioelectron.*, 2017, **87**, 949–956.
- 186 Y. Dai, R. A. Somoza, L. Wang, J. F. Welter, Y. Li, A. I. Caplan and C. C. Liu, *Angew. Chem., Int. Ed.*, 2019, **58**, 17399–17405.
- 187 Sensit Smart - PalmSens, <https://www.palmsens.com/product/sensit-smart/>, (accessed 4 August 2020).
- 188 Sensit BT - PalmSens, <https://www.palmsens.com/product/sensit-bt/>, (accessed 4 August 2020).
- 189 R. Francis, M. Le Bideau, P. Jardot, C. Grimaldier, D. Raoult, J. Yaacoub, B. Khalil and B. La Scola, *Clin. Microbiol. Infect.*, 2021, **27**, 128e1–128e7.

