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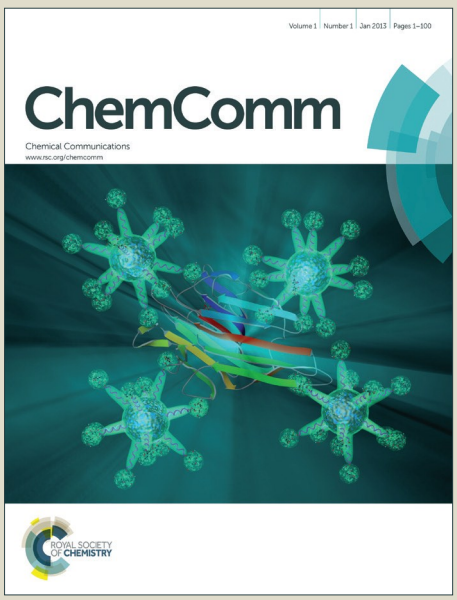

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Triggering autocatalytic reaction by host-guest interactions

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The acceleration of sequential reaction through electrostatic alteration of substrate basicity within a supramolecular host is demonstrated. In the presence of the host the reaction which is autocatalytic starts much sooner and exhibits substrate size selectivity.

Remote substrate activation is a distinctive trait of enzyme catalysis.[1] Being nested in enzyme pocket the substrate is exposed to electrostatic environment created by protein residues. This entails modifying the acid-base properties (pKa) of bound substrate, and in consequence, facilitates its hydrolysis. In some cases the magnitude of apparent pKa shift reaches even up to five units.[2] Similar (biomimetic) action has long been endeavored to reproduce in synthetic hosts which are reminiscent of enzyme cavities.[3-16] However, this proved very challenging as required the host which would meet several criteria, especially the capability of stabilizing the charged transition state. Raymond and co-workers have recently presented the first successful demonstration of electrostatic catalysis based on *ion-ion* interactions by using tetrahedral metal cluster which consists of hydrophobic cavity and negatively charged vertices.[17-24] It was found that the complex alters the pKa of encapsulated guest to the extent that enables acid hydrolysis at basic pH.[17, 21] Further research, in particular by Nau group, has revealed that the similar electrostatic action *via ion-dipole* interactions is also possible in cucurbit[n]uril (**CB**) hosts, which are a family of barrel-shaped macrocycles[25-38] composed of hydrophobic cavity and electronegative carbonyl-fringed rim. Like the metal cluster, **CB** host was shown to promote hydrolysis reactions via Coulombic stabilization of protonated substrate.[39-41] Most recently, the stabilization of positively charged species in transition state through *ion-π* interactions has also been suggested for resorcin[4]arene capsules.[42-46] Being intrigued by these results we pursued the studies to broad the scope of electrostatic catalysis towards multistep chemical transformations.

Herein we report sequential azo coupling-type reaction promoted by electrostatic **CB** host-guest interactions. Azo coupling is an important chemical process used in industry for production of azo dyes.[47] Apart textile-dyeing industry, azo compounds are also used as pH indicators and molecular photoswitches.[48, 49] The coupling reaction usually occurs between aromatic compound and diazonium salt which has to be generated *in situ* due to high instability. In this respect, triazene is a robust substitute of diazonium salt.[50] We found that triazene reacts with electron-rich arene in the presence of cucurbit[6]uril (**CB6**) affording azo dye. Since the background reaction is negligible at the timescale of catalytic event we may talk here about triggering the reaction.

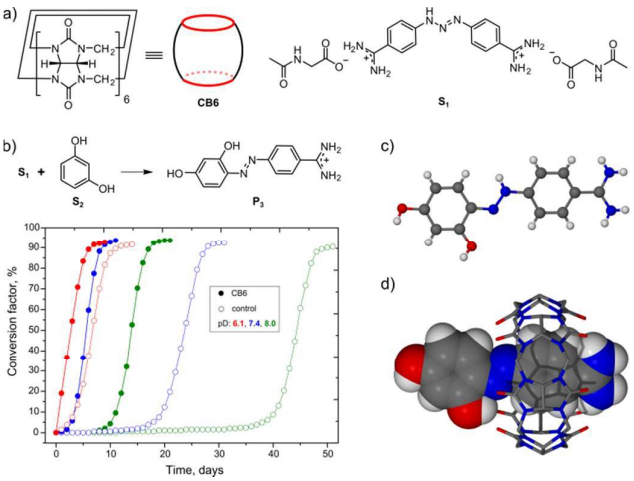

Fig. 1 a) Chemical structure of cucurbit[6]uril (**CB6**) and berenil (**S_1**); b) Reaction scheme and plots of the time dependencies of the conversion factor for azo coupling between **S_1** and resorcinol (**S_2**) in the presence and absence of **CB6** for different values of pD; c) X-ray structure of azo dye product **P_3**; d) X-ray structure of inclusion complex between **CB6** and **P_3**.

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In our study we employed berenil (**S_1**), a commercially available triazene which is an anti-infective drug (Fig. 1a). Resorcinol (**S_2**) served as aromatic partner in the reaction. In typical experiment, stoichiometric mixture of **S_1**, **S_2**, and **CB6** were suspended by agitation in heavy water and kept at room temperature. The experiment was carried out at near neutral pD without buffering. During the experiment, the pD of the suspension changed from 7.4 to 4.2 and azo dye product (**P_3**) precipitated as an orange solid. The process was monitored by ^{1}H NMR spectroscopy following the formation of 4-aminobenzamidine (**P_1**) which is another product of the reaction. According to NMR data, the reaction completed within two weeks, whereas only traces (<1 mol%) of **P_1** were detected in the control experiment performed without **CB6**. The azo dye product **P_3** was isolated from **CB6** precipitate by addition of calcium chloride followed by filtration and recrystallization from hydrochloric acid solution. X-ray diffraction of isolated orange crystals, along with NMR and MS analysis, confirmed the expected structure of azo dye (Fig. 1c). The azo product **P_3** was also crystalized as the inclusion complex with **CB6** upon the acidification of the crude reaction mixture (Fig. 1d).

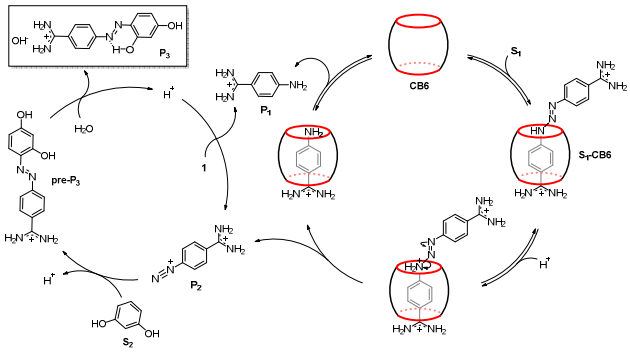

Fig. 2 Plausible mechanism of **CB6**-catalyzed sequential azo coupling-type reaction. The first step of the reaction is the acid hydrolysis of **S_1** promoted by electrostatic action of **CB6**. In the next step, the generated diazonium intermediary **P_2** reacts with **S_2** to yield azo compound **pre-P_3**. The compound **pre-P_3** is next transformed to final azo product **P_3** with a loss of proton which acidifies the reaction mixture accelerating the hydrolysis of **S_1**, and consequently the overall reaction.

To monitor the progress of the reaction, we calculated the quantity γ=[**P_1**]/([**S_2**]+[**P_1**])x100%. In the following, we refer to the quantity γ as the conversion factor. Time dependence of the conversion factor for the coupling reaction at different pD values is plotted in Fig. 1b. The coupling reaction is pH-sensitive exhibiting autocatalytic behavior in nearly neutral solutions. The involvement of **CB6** into the process is manifested by shorter induction period as well as a steeper curve during the acceleration phase when compared with the control experiment. The most spectacular effect of **CB6** action was observed in basic solution with the lag phase being reduced by one month.

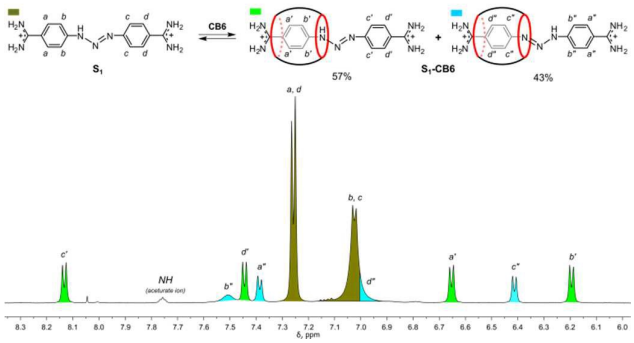

Fig. 3 Partial ^{1}H NMR spectrum of equimolar mixture of **S_1** and **CB6** in D_2O, 277 K. The tautomer ratio was calculated by the integration of the corresponding peaks.

The plausible mechanism of the reaction is depicted in Fig. 2. In the first step, the triazene molecule **S_1** is encapsulated by **CB** host. The encapsulation is facilitated by template effect of the positively charged amidinium terminus which interacts electrostatically with the electronegative **CB** rim. The formation of the host-guest species is clearly seen in NMR spectrum after mixing the reactants (Fig. S1). The proton resonances of aromatic ring placed inside **CB** cavity are shifted upfield, while signals of aromatic protons positioned outside the cavity move toward downfield region. Low-temperature NMR measurements of **S_1-CB6** mixture revealed that the observed shifts are attributed to the formation of two host-guest **CB6** complexes (**S_1-CB6**) comprising "frozen" tautomeric forms of thermodynamically stable *trans* isomer[51] of **S_1** (Fig. 3, S6-S8). The weak differentiation of the resonance structures of **S_1** upon the encapsulation indicates that either the NH group of triazene moiety in both tautomers is in a close proximity to carbonyl portals of **CB** rim or the hydrogen bonding is not a dominant force in the binding event that would be consistent with previous reports.[28] Hence it is not clear which tautomer is a reaction intermediate. The possible reaction scenario involving the major tautomer is shown in Fig. 2. The enhanced electron density at the **CB** rim increases the basicity of encapsulated substrate rendering triazene moiety susceptible to protonation and subsequent rupture with formation of 4-amidinophenyldiazonium (**P_2**). Despite high **CB6** loading, the cleavage reaction was triggered by catalytic amounts of **S_1-CB6** complex (ca. 15 mol%) formed at pre-equilibrium. Importantly, the employment of sub-stoichiometric amounts of **CB6** (20 mol%) did not affect the reaction rate significantly as the initial amount of the inclusion complex formed was about 5 mol% (Fig. S8). As the reaction progresses, the vast fraction of **S_1-CB6** complex is quickly consumed and not further replenished by virtue of competitive binding of **P_1** to **CB6**. In the next step, diazonium cation **P_2** reacts with **S_2** *via* electrophilic substitution to give azo compound **pre-P_3**. The substitution reaction is favored by alkaline pH, though remains feasible also in acidic conditions.[52] With the acidification of the reaction mixture, caused by the hydrolysis of **pre-P_3** to insoluble **P_3**, the hydrolysis of **S_1**, and consequently, the overall reaction

accelerates. Although **S_1-CB6** complex was not detected by NMR at this stage, the hydrolysis of **S_1**, as judged from the kinetic slopes, does take place inside the macrocycle cavity. The plot showing the pD of the reaction mixture as a function of time is shown in Fig. S10. The product **P_3** precipitates probably due to intramolecular hydrogen bonding between azo bond and nearby hydroxyl group.[52] To note, in strongly acidic conditions the hydrogen bond is vanished at the expense of protonation of azo dye bond (Fig. 1c).

To verify the proposed mechanism, we modeled the **CB6**-catalyzed sequential reaction by the following set of two rate equations: (i) $S_1 + D_2O + D^+ \xrightarrow{k_1} P_1 + P_2 + D^+$ and (ii) $S_2 + P_2 \xrightarrow{k_2} P_3 + D^+$. The first equation is the hydrolysis of **S_1**. The second reaction describes the azo coupling step. The acid specific hydrolysis rate constant, k_H, in equation (i) had the form $k_H = k_1^{n,c}[D^+]$, where the superscripts *n* and *c* refer, respectively, to the neutral (control) and **CB6**-catalyzed reaction. The constant term in k_1 was set equal to zero to account for the fact that in all experimental conditions the induction periods (the lag phases) were observed. The equations (i) and (ii) were solved numerically and the conversion factor, *γ*, was calculated as a function of time for pD = 8.0, 7.4, and 6.1. The rate constants k_1^n, k_1^c, and k_2 were determined by least-square fit to the experimental values of *γ* (see ESI for details). Numerical analysis of the rate equations showed the increase of both rate constants k_1^n and k_1^c with the decrease of pD. This tendency is in line with results reported recently for the hydrolysis of **S_1**.[53] As expected, we observed also substantial decrease of the rate constant k_2 with increasing acidity of the system. To quantify the catalytic effect of **CB6**, for each value of pD we calculated the acceleration factor defined as the ratio k_1^c/k_1^n. Based on the results of the numerical analysis of the rate constants, we obtained the acceleration factors 3.24, 5.47, and 5.12 for pD equal 8.0, 7.4, and 6.1, respectively.

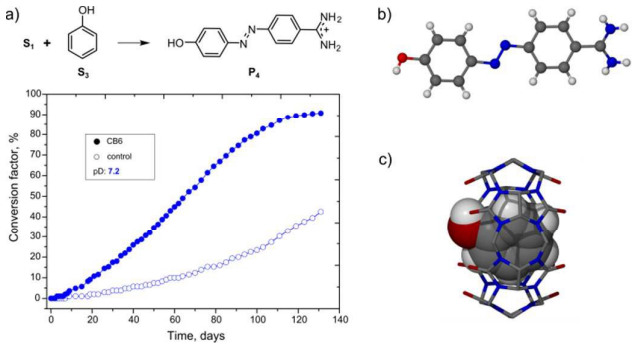

Fig. 4 a) Reaction scheme and plots of the time dependence of the conversion factor for azo coupling between **S_1** and phenol (**S_3**) in the presence and absence of **CB6**; b) X-ray structure of azo dye product **P_4**; c) X-ray structure of inclusion complex between **S_3** and **CB6**.

When **S_2** was substituted by phenol (**S_3**) the coupling reaction significantly slowed down (Fig. 4a). Since the resulting azo product (**P_4**, the X-ray structure is shown in Fig. 4b) lacks the intramolecular hydrogen bonding, the equilibrium does not shift towards its hydrolysis. As a result, the pD of the reaction mixture changed from 7.2 to 6.4, and no autocatalysis was observed. The slow reaction rate stems surprisingly also from competitive binding of **S_3** which displaces encapsulated **S_1** from **CB** cavity. Noteworthy, this kind of competition was not observed for **S_2** suggesting the size selectivity of **CB6** ring. During the first day of experiment, the content of **S_1-CB6** complex in the solution dropped below 0.5 mol%. The process is supposedly precipitation-driven considering the higher affinity of cationic guests toward **CB** host. The precipitation of phenol-**CB6** (**S_3-CB6**) complex can be delayed by addition of calcium chloride. In this case, the process of inclusion was observed through upfield shifts of phenolic NMR signals followed by slow crystallization of **S_3-CB6**, whose structure was further corroborated by single crystal X-ray analysis (Fig. 4c).

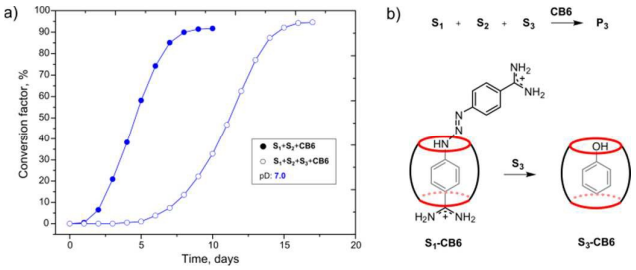

Fig. 5 a) Plots of the time dependence of the conversion factor for **CB6**-catalyzed azo coupling between **S_1** and **S_2** in the presence and absence of **S_3**; b) Schematic representation of competitive binding of **S_3** to **CB6** in the reaction between **S_1** and **S_2** in the presence of **S_3**.

To prove unequivocally the catalytic properties of **CB6** the coupling reaction between **S_1** and **S_2** was conducted with a stoichiometric admixture of **S_3**. As expected the reaction was greatly inhibited due to competitive binding of **S_3** for **CB** cavity that confirms nicely the action of **CB6** as a catalyst (Fig. 5). The reaction inhibition was also observed in the presence of 1,5-pentanediamine (cadaverine) which forms a strong complex with **CB6** (Fig. SX).

In summary, we designed a two-step azo coupling-type reaction induced by host-guest interactions. A peculiar feature of the reaction is that the first step is acid-catalyzed, whereas the next step prefers non-acidic conditions. Normally, if the reaction is performed in acidic medium, the fast hydrolysis step is followed by slow electrophilic substitution, and vice versa, the reaction is extremely sluggish in neutral and alkaline solutions. The use of **CB6** host as a catalyst facilitates the acid hydrolysis, and consequently overall reaction at basic pH through the electrostatic enhancement of substrate basicity. Accordingly, the reaction being autocatalytic begins much earlier, i.e., is triggered by **CB6**. The presented concept is of great importance for the development of chemical transformations involving discrete reaction steps occurring at different pH values. Furthermore, this strategy, given the substrate size selectivity, may be applicable for performing acid-catalyzed reactions on selected functional groups in the presence of other functionalities sensitive to acidic conditions.

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Notes and references

1. A. Warshel, P. K. Sharma, M. Kato, Y. Xiang, H. Liu and M. H. M. Olsson, *Chem. Rev.*, 2006, **106**, 3210-3235.
2. F. H. Westheimer, *Tetrahedron*, 1995, **51**, 3-20.
3. M. Raynal, P. Ballester, A. Vidal-Ferran and P. W. N. M. van Leeuwen, *Chem. Soc. Rev.*, 2014, **43**, 1734-1787.
4. J. Meeuwissen and J. N. H. Reek, *Nat. Chem.*, 2010, **2**, 615-621.
5. M. Yoshizawa, J. K. Klosterman and M. Fujita, *Angew. Chem. Int. Ed.*, 2009, **48**, 3418-3438.
6. S. H. A. M. Leenders, R. Gramage-Doria, B. de Bruin and J. N. H. Reek, *Chem. Soc. Rev.*, 2015, **44**, 433-448.
7. T. S. Koblenz, J. Wassenaar and J. N. H. Reek, *Chem. Soc. Rev.*, 2008, **37**, 247-262.
8. C. J. Walter and J. K. M. Sanders, *Angew. Chem. Int. Ed.*, 1995, **34**, 217-219.
9. J. Kang and J. Rebek, *Nature*, 1997, **385**, 50-52.
10. M. Yoshizawa, M. Tamura and M. Fujita, *Science*, 2006, **312**, 251-254.
11. S. R. Shenoy, F. R. Pinacho Crisóstomo, T. Iwasawa and J. Rebek, *J. Am. Chem. Soc.*, 2008, **130**, 5658-5659.
12. P. Thordarson, E. J. A. Bijsterveld, A. E. Rowan and R. J. M. Nolte, *Nature*, 2003, **424**, 915-918.
13. E. Anslyn and R. Breslow, *J. Am. Chem. Soc.*, 1989, **111**, 5972-5973.
14. F. Ortega-Caballero, C. Rousseau, B. Christensen, T. E. Petersen and M. Bols, *J. Am. Chem. Soc.*, 2005, **127**, 3238-3239.
15. M. J. Wiester, P. A. Ulmann and C. A. Mirkin, *Angew. Chem. Int. Ed.*, 2011, **50**, 114-137.
16. A. G. Salles, S. Zarra, R. M. Turner and J. R. Nitschke, *J. Am. Chem. Soc.*, 2013, **135**, 19143-19146.
17. M. D. Pluth, R. G. Bergman and K. N. Raymond, *Science*, 2007, **316**, 85-88.
18. W. M. Hart-Cooper, K. N. Clary, F. D. Toste, R. G. Bergman and K. N. Raymond, *J. Am. Chem. Soc.*, 2012, **134**, 17873-17876.
19. C. J. Hastings, M. P. Backlund, R. G. Bergman and K. N. Raymond, *Angew. Chem. Int. Ed.*, 2011, **50**, 10570-10573.
20. C. J. Hastings, M. D. Pluth, R. G. Bergman and K. N. Raymond, *J. Am. Chem. Soc.*, 2010, **132**, 6938-6940.
21. M. D. Pluth, R. G. Bergman and K. N. Raymond, *J. Am. Chem. Soc.*, 2008, **130**, 11423-11429.
22. C. Zhao, F. D. Toste, K. N. Raymond and R. G. Bergman, *J. Am. Chem. Soc.*, 2014, **136**, 14409-14412.
23. C. Zhao, Q.-F. Sun, W. M. Hart-Cooper, A. G. DiPasquale, F. D. Toste, R. G. Bergman and K. N. Raymond, *J. Am. Chem. Soc.*, 2013, **135**, 18802-18805.
24. Z. J. Wang, K. N. Clary, R. G. Bergman, K. N. Raymond and F. D. Toste, *Nat. Chem.*, 2013, **5**, 100-103.
25. J. W. Lee, S. Samal, N. Selvapalam, H.-J. Kim and K. Kim, *Acc. Chem. Res.*, 2003, **36**, 621-630.
26. J. Lagona, P. Mukhopadhyay, S. Chakrabarti and L. Isaacs, *Angew. Chem. Int. Ed.*, 2005, **44**, 4844-4870.
27. B. C. Pemberton, R. Raghunathan, S. Volla and J. Sivaguru, *Chem. Eur. J.*, 2012, **18**, 12178-12190.
28. K. I. Assaf and W. M. Nau, *Chem. Soc. Rev.*, 2015, **44**, 394-418.
29. A. L. Koner, C. Márquez, M. H. Dickman and W. M. Nau, *Angew. Chem. Int. Ed.*, 2011, **50**, 545-548.
30. X. Lu and E. Masson, *Org. Lett.*, 2010, **12**, 2310-2313.
31. H. Cong, Z. Tao, S. F. Xue and Q. J. Zhu, *Curr. Org. Chem.*, 2011, **15**, 86-95.
32. L. Zheng, S. Sonzini, M. Ambarwati, E. Rosta, O. A. Scherman and A. Herrmann, *Angew. Chem. Int. Ed.*, 2015, **54**, 13007-13011.
33. W. L. Mock, T. A. Irra, J. P. Wepsiec and M. Adhya, *J. Org. Chem.*, 1989, **54**, 5302-5308.
34. S. Y. Jon, Y. H. Ko, S. H. Park, H.-J. Kim and K. Kim, *Chem. Commun.*, 2001, **37**, 1938-1939.
35. B. C. Pemberton, N. Barooah, D. K. Srivatsava and J. Sivaguru, *Chem. Commun.*, 2010, **46**, 225-227.
36. B. C. Pemberton, R. K. Singh, A. C. Johnson, S. Jockusch, J. P. Da Silva, A. Ugrinov, N. J. Turro, D. K. Srivastava and J. Sivaguru, *Chem. Commun.*, 2011, **47**, 6323-6325.
37. C. Yang, T. Mori, Y. Origane, Y. H. Ko, N. Selvapalam, K. Kim and Y. Inoue, *J. Am. Chem. Soc.*, 2008, **130**, 8574-8575.
38. S. J. Barrow, S. Kasera, M. J. Rowland, J. del Barrio and O. A. Scherman, *Chem. Rev.*, 2015, **115**, 12320-12406.
39. N. i. Saleh, A. L. Koner and W. M. Nau, *Angew. Chem. Int. Ed.*, 2008, **47**, 5398-5401.
40. C. Klöck, R. N. Dsouza and W. M. Nau, *Org. Lett.*, 2009, **11**, 2595-2598.
41. N. Basilio, L. García-Río, J. A. Moreira and M. Pessêgo, *J. Org. Chem.*, 2010, **75**, 848-855.
42. A. Cavarzan, A. Scarso, P. Sgarbossa, G. Strukul and J. N. H. Reek, *J. Am. Chem. Soc.*, 2011, **133**, 2848-2851.
43. G. Bianchini, G. L. Sorella, N. Canever, A. Scarso and G. Strukul, *Chem. Commun.*, 2013, **49**, 5322-5324.
44. Q. Zhang and K. Tiefenbacher, *J. Am. Chem. Soc.*, 2013, **135**, 16213-16219.
45. L. Catti and K. Tiefenbacher, *Chem. Commun.*, 2015, **51**, 892-894.
46. Q. Zhang and K. Tiefenbacher, *Nat. Chem.*, 2015, **7**, 197-202.
47. K. Hunger, P. Mischke and W. Rieper, in *Ullmann's Encyclopedia of Industrial Chemistry*, Wiley-VCH Verlag GmbH & Co. KGaA, 2000.
48. A. A. Beharry and G. A. Woolley, *Chem. Soc. Rev.*, 2011, **40**, 4422-4437.
49. R. Klajn, *Pure Appl. Chem.*, 2010, **82**, 2247-2279.
50. S. Bräse, *Acc. Chem. Res.*, 2004, **37**, 805-816.
51. M. Barra and N. Chen, *J. Org. Chem.*, 2000, **65**, 5739-5744.
52. O. Macháčková, V. Štěrba and K. Valter, *Collect. Czech. Chem. Commun.*, 1972, **37**, 1851-1860.
53. M. Campbell, R. J. Prankerd, A. S. Davie and W. N. Charman, *J. Pharm. Pharmacol.*, 2004, **56**, 1327-1332.