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A review of the known MTA-cooperative PRMT5 inhibitors

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Protein arginine methyltransferase 5 (PRMT5), an epigenetic target with significant clinical potential, is closely associated with the occurrence and development of a range of tumours and has attracted considerable interest from the pharmaceutical industry and academic research communities. According to incomplete statistics, more than 10 PRMT5 inhibitors for cancer therapy have entered clinical trials in recent years. Among them, the second-generation PRMT5 inhibitors developed based on the synthetic lethal strategy demonstrate considerable clinical application value. This suggests that, following the precedent of poly ADP ribose polymerase (PARP), PRMT5 has the potential to become the next clinically applicable synthetic lethal target. However, due to the inherent dose-limiting toxicity of epigenetic target inhibitors, none of these PRMT5 inhibitors has been approved for marketing to date. In light of this, we have conducted a review of the design thoughts and the structure–activity relationship (SAR) of known methylthioadenosine (MTA)-cooperative PRMT5 inhibitors. Additionally, we have analysed the clinical safety of representative first- and second-generation PRMT5 inhibitors. This paper discusses the *in vivo* vulnerability of the aromatic amine moiety of the second-generation PRMT5 inhibitor based on its structure. It also considers the potential nitrosamine risk factors associated with the preparation process.

1 Introduction

Protein arginine methylation, as one of the post-translational modifications (PTMs), is catalysed by protein arginine methyltransferases (PRMTs).[1,2] And PRMTs are known to have nine members (PRMT1-9) in human cells. Although the fact that every member of the PRMT family can catalyse the methyl transfer reaction between the cofactor S-adenosylmethionine (SAM, as the methyl group's donor) and the protein arginine residues, the resulting forms of methylated arginine catalysed by the different members of the PRMT family are not consistent with one another.[3–5] Based on this, PRMTs may be further subdivided, including type I (PRMT1, 2, 3, 4, 6, 8), type II (PRMT5, 9), and type III (PRMT7).[6,7] Among these, PRMT5 is responsible for catalysing the formation of omega-NG, N'G-symmetric dimethylarginines (SDMA) and omega-NG-monomethyl arginine (MMA), which are involved in various physiological multifarious physiological and biochemical processes[8] such as cell cycle,[9,10] the splicing of DNA repair genes,[11] embryonic development and differentiation,[12–14] and so on. Moreover, there is evidence that the dysregulation of PRMT5 is associated with the development of several diseases, particularly malignant tumour pathogenesis and poor patient prognosis.[15–18]

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PRMT5, as a classical SAM-binding protein, has two orthosteric sites in its C-terminal structure, namely the SAM-binding site and the substrate-binding site. A passage way, designated the Arg tunnel, exists between the SAM pockets and the substrate pockets (as shown in the Fig. 1B, PDB: 4X61).[21,22] PRMT5, which has been identified as a potential target for antitumour therapy, has attracted increasing attention. Currently, according to incomplete statistics, there are more than 10 PRMT5 inhibitors entering clinical development worldwide, with the majority being in the clinical phase I. These small molecule inhibitors may be subdivided into the following several types according to their binding sites to PRMT5.

(I) The SAM mimetic. The structure of this inhibitor is characterised by the adenosine scaffold as its structural core (it is highlighted in red in Fig. 2). The binding of it to PRMT5 occurs in the SAM binding pocket and competes with SAM; because of all PRMTs use the cofactor SAM, as methyl group's donor, to catalyse the arginine methylation reaction and share a highly homologous SAM-dependent methyltransferase (MTase) domain which is the catalytic domain, how to make SAM competitive PRMT5 inhibitors have the target protein selectivity, which is one of the mayor challenges researchers face. In fact, scientists obtained a lot of SAM mimetics, including but not limited to SAH, sinefungin, dehydrosinefungin and DS-437 (structure not shown here) in the early stage of PRMT5 inhibitors development. Owing to the remarkable similarity in chemical structure between SAM and these mimetics, these compounds could occupy the SAM

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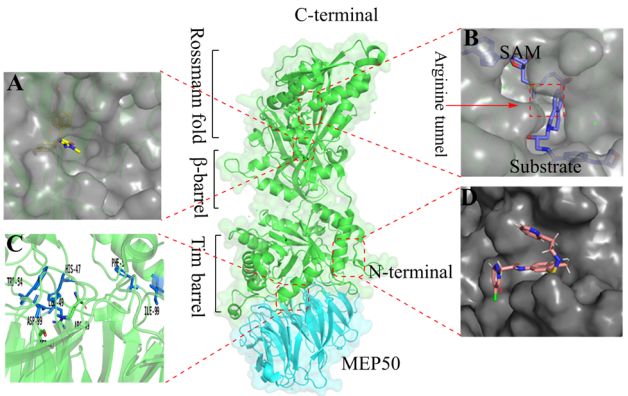

Fig. 1 Binding sites where PRMT5 can be used for the development of pharmaceutical agents. (A) Allosteric binding site (PDB: 6uxx).[19,20] (B) SAM binding site and substrate binding site. The substrate binding site and SAM-binding pocket are closely connected by the arginine tunnel (PDB: 4X61),[21,22] which can be exploited by certain inhibitors, such as JNJ-64619178, to block both the SAM-binding site and substrate binding pocket simultaneously.[23,24] (C) The PRMT5: MEP50 protein–protein surface interaction (PDB: 4gqb).[25] (D) The PBM groove (PDB: 6U0P). The substrate adaptor proteins (SAPs) contains a highly conserved linear peptide sequence, termed the PRMT5 binding motif (PBM). The PBM peptide and PRMT5 occurs within a shallow groove (the "PBM groove") of the TIM barrel[5,26–32]

binding pocket and show PRMT5 inhibitory activity to a certain degree, but lack of selectivity.[21,42–45] Interestingly, MTA, an endogenous ligand, is a selective SAM-competitive inhibitor of PRMT5, which shows more than 100-fold selectivity for PTMT5 over all other PRMTs (however, its potency is weak, $IC_{50} = 260$ nM).[46] In light of the findings of MTA research, non-covalently bound SAM competitive inhibitors, exemplified by **11** (LLy283),[33] have been developed. Concurrently, covalently bound SAM competitive inhibitors with **11** (LLy283) as the parent structure were also developed such as the hemiaminal compound. Although these SAM mimetics are similar in backbone structure, their preparation routes and methods are diverse. Among the various preparation methods, the strategy of using the adenosine scaffold structure as the starting material for the synthesis of the corresponding SAM mimetic has been the subject of extensive study and has a high success rate (see Fig. 2).[33–37]

(II) The dual SAM/substrate PRMT5 inhibitor. These inhibitors (such as JNJ-64619178,[47,48] see the structure in Fig. 2) could potentially prohibit binding of SAM and the substrate simultaneously by occupying the SAM-binding site, and then the quinoline moiety extending into the Arg tunnel, where it interacts with the key catalytic residues Glu444 of PTMT5. Moreover, the quinoline ring forms π–π stacking with Phe327. This unique binding pattern makes it possible to have a longer retention time at the target site, thereby reducing the required dose for clinical applications. However, this tight binding pattern, along with higher efficacy, may also result in more severe toxic side effects.[49–53] In terms of its chemical structure, this type of inhibitor is, in fact, a novel SAM analogue. Consequently, the most cost-effective preparation strategy is to use the adenosine scaffold structure as a starting material to synthesise the corresponding compounds. See Fig. 2.[23,24,38–41]

(III) The substrate competitive inhibitor. Its structure is characterised by the tetrahydroisoquinoline as its structural core (it is highlighted in red in Fig. 3). The binding site of this inhibitor with PRMT5 is in the substrate-binding pocket and competes with the substrate. Many studies have shown that the tetrahydroisoquinoline portion of this type of inhibitor forms a key cation–π interaction with the cofactor SAM, and at the same time, the aromatic ring of tetrahydroisoquinoline forms a potential π–π stacking interaction with Phe327. Phe327 is unique to PRMT5, and other PRMT enzymes at the same position are a Met. It is postulated that these factors contribute to the high selectivity and inhibitory activity of EPZ01566 against PRMT5.[61–63] The preparation of this class of inhibitors is to use (*R*)-2-(oxiran-2-ylmethyl)-1,2,3,4-tetrahydroisoquinoline as the starting material, and the corresponding compounds are made by aminolysis and acylation reactions (see Fig. 3).[54–60] And the PRMT5 degrader MS4322 with EPZ015666 as the target protein of interest (**POI**) ligand is also in preclinical studies.[64]

(IV) Allosteric inhibitors. when the PRMT5:MEP50 complex with the inhibitor binds in a new binding site (apart from the SAM-binding site and substrate-binding pocket), the backbone structure of protein is beginning to significant structural rearrangement and further influence both the SAM-binding site and substrate-binding pocket so that none of the SAM and substrate can bind with PRMT5 in their respective binding site, such as R-1a.[19,20]

Although all PRMT5 inhibitors described above have shown high selectivity and inhibitory activity against PRMT5, they lack specific selectivity for tumour cells and have a narrow therapeutic window in terms of their mechanism of action. Moreover, as mentioned previously, PRMT5 plays a crucial role in normal physiological processes; PRMT5 knockdown may result in embryonic lethality[42,65] and tissue-specific PRMT5 knockdown in the central nervous system,[66,67] skeletal muscle,[68] and haematopoietic system[69] all produce substantial toxicity. These toxicities may become dose-limiting in the clinical use of the drug, potentially affecting the progress of clinical studies of drugs targeting PRMT5. In clinical experiment, the majority of such PRMT5 inhibitors also exhibit significant tumour growth inhibitory effects. However, adverse events (AEs) associated with PRMT5 inhibitor therapy, including fatigue, anaemia, nausea, alopecia and dyspeptic events, are prevalent (for a list of representative PRMT5 inhibitors in clinical trials, please refer to Table 1). For example, 4 (PF-06939999) showed significant dose-limiting toxicities including thrombocytopenia, anaemia and neutropenia in a phase I dose-escalation study. Currently, clinical development has been terminated (NCT03854227).[70,71] Owing to the universality of these treatment-related AEs, the toxicity and side effects of these PRMT5 inhibitors, especially target associated haematotoxicity, need been further evaluated. Currently, the ability of these PRMT5 inhibitors to pass evaluation in further clinical trials depends on the assessment of their efficacy and safety; what is the benefit-risk ratio? Is the safety acceptable? Current clinical studies show that it is not certain. And, it may still be extremely valuable to explore other

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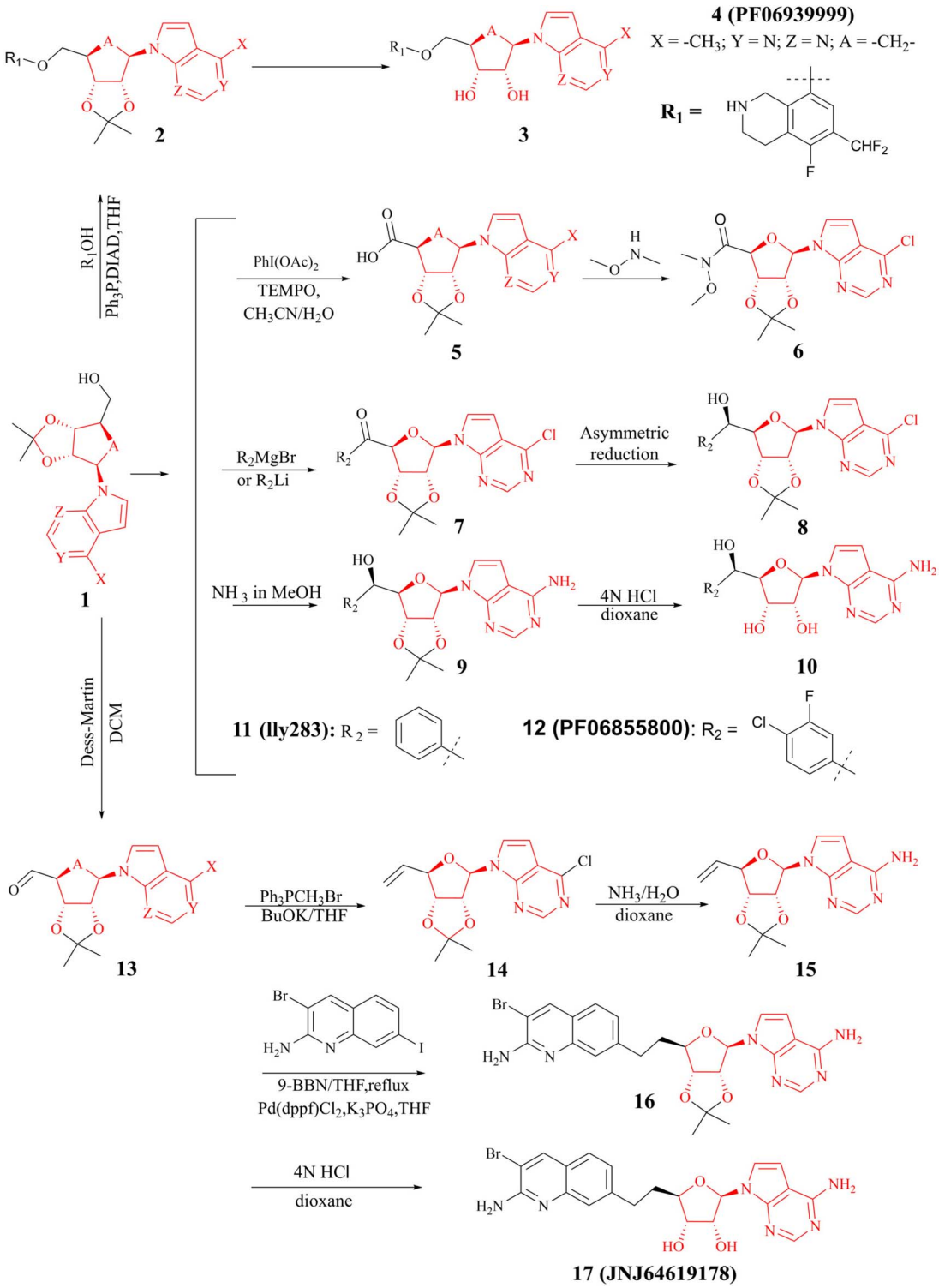

Fig. 2 Synthesis of the SAM mimetic using the adenosine scaffold as its structural core.[23,24,33–41] The adenosine scaffold is highlighted in red.

treatments with this class of drugs, such as combinations with other drugs. Hence, the study on PRMT5 inhibitors of new structures has been gradually focused on "synthetic lethality" to further higher selectivity and lower target associated toxicity.[72] In order to facilitate the distinction between the various types of PRMT5 inhibitors, the above mentioned inhibitors developed on the basis of the cofactor SAM-binding site and the substrate-binding site are also collectively referred to as the first-generation PRMT5 inhibitors.

Synthetic lethality was initially observed and defined in drosophila and yeast.[73] In a nutshell, if A and B genes constitute a synthetic lethal pair, mutations in either of these two genes

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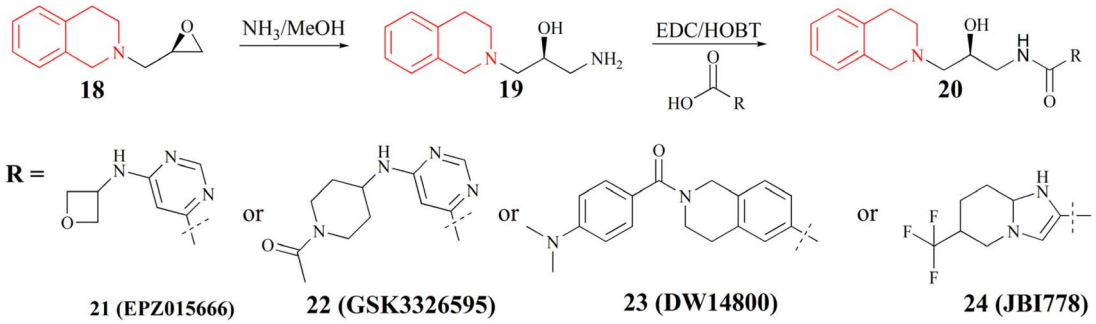

Fig. 3 Synthesis of compounds of formula 20.[54–60]

alone have a minimal impact on cells, but the simultaneous mutations of both genes will result in cell death.[72,74–76] This concept was further elucidated in 2005, when researchers found that pharmacological inhibition of the poly ADP-ribose polymerase (PARP) may result in the death of tumour cells bearing the BRCA mutation, whereas normal cells are minimally affected due to retaining the wild-type copy. (PARP and BRCA form a synthetic lethal pair here).[77,78] In 2014, the world's first synthetic lethal inhibitor olaparib, which are used in the treatment of BRCA-mutant ovarian cancer, was approved for marketing.[79]

It is widely known that tumour suppressor genes *CDKN2A* on chromosome 9p21 (*chr9p21*) are homozygously deleted in multiple solid tumours. The extent of this deletion varies considerably, often leading to neighbouring genes being deleted together. Homozygous deletion of the methylthioadenosine phosphorylase (*MTAP*) gene takes place frequently in multiple solid tumours owing to its proximity to the tumour suppressor gene *CDKN2A*.[80] It is noteworthy that the *MTAP* is in charge of methylthioadenosine (MTA) conversion in the methionine salvage pathway[81,82] (see Fig. 4). As an analogue of SAM, MTA fulfils the role of a natural SAM competitive inhibitor, showing more than 100-fold selectivity for PRMT5 over all other PRMT family members. In other words, *MTAP* loss results in the accumulation of intracellular MTA, which produces a frail PRMT5 state that is selectively sensitive to further inhibition by PRMT5 inhibitors.[42,43,46,83,84] Thus, both PRMT5 and MTAP constitute a synthetic lethal pair in this context. However, it is anticipated that the aforementioned clinical PRMT5 inhibitor will not exhibit selective inhibition of PRMT5 activity in MTAP-deleted tumours without impacting the normal cells due to the mechanism of action of these agents. Fortunately, MTA competes with SAM to generate the PRMT5-MEP50/MTA (PRMT5/MTA) complex in MTAP-deleted tumours. The researchers believe that a small molecule which could bind to and stabilise the PRMT5/MTA complex will selectively inhibit PRMT5 activity in MTAP-deleted tumour cells. Soon afterwards, several PRMT5/MTA complex inhibitors, including **28** (TNG908),[85,86] **27** (MRTX1719),[87,88] and **30** (AMG193)[89] were approved for clinical development successively.

In order to differentiate themselves from the aforementioned PRMT5 inhibitors, these MTA-cooperative synthetic lethal inhibitors have also been designated as second-generation PRMT5 inhibitor.

In fact, this synthetic lethality also extends to the key points of the upstream and downstream molecular regulatory mechanisms of PRMT5, including the PRMT5 complex member located downstream (see Fig. 4).[26] It is known that a human PRMT5 is made up of a Rossmann fold which contains the catalytic site, a β-barrel that allows for the dimerisation of PRMT5, and a TIM barrel which houses the PBM groove and mediates interactions with different substrate adaptor proteins (SAPs).[5] Owing to the positive allosteric effect of the partner protein MEP50 (WDR77), the complex composed of PRMT5 and MEP50 usually possesses a significantly higher level of MTase activity than PRMT5 alone.[21,27] (Actually, a stable heterooctameric complex comprising four copies of PRMT5 and MEP50 is frequently observed within the cellular environment.)[21,28] Nevertheless, the identification and recruitment of substrates for the PRMT5/MEP50 complex depend on the involvement of SAPs. It is noteworthy that the research indicates that three SAPs (RioK1, pICln, and COPR5) share a highly conserved linear peptide sequence, which has been named as the PRMT5 binding motif (PBM). And the interaction between PBM and the PBM-binding groove is responsible for mediating the methylation of over 25 substrates, including nucleolin and RPS10.[29] Thus, the targeted inhibition of the protein–protein interactions (PPIs) represents another promising avenue for the development of MTA-cooperative synthetic lethal inhibitors (Fig. 4). Based on these results, the PBM-competitive inhibitor **31**(BRD0639),[31] the cyclic binding peptide inhibitor **32**,[30,32] and PRMT5:MEP50 PPI inhibitors **33**[25] were discovered successively.

However, the clinical status of anti-tumour treatment with second-generation inhibitors is under pressure and it will also face with a range of potential challenges. For instance, six different malignancies, including non-small cell lung cancer and melanoma, were treated with **27** (MRTX1719) in a dose-escalating manner, and the tumour burden was reduced by approximately 30–60%. However, it is important to note that these encouraging early clinical outcomes are limited by the small sample size and more work needs to be done to verify and confirm it. Furthermore, PRMT5 is essential for the survival of all cells, especially haematopoietic cells. Despite the treatment-related adverse events (AEs) in humans were not reported at the

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Table 1 Representative first-generation PRMT5 inhibitors in clinical trials[a]

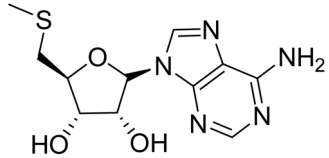
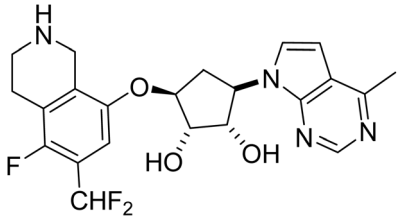
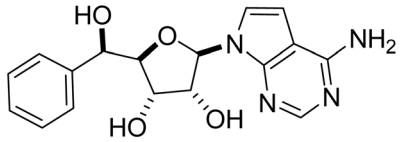
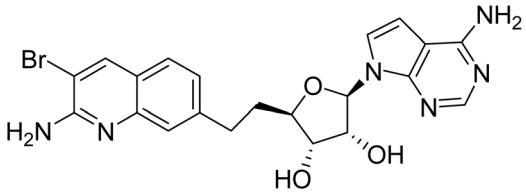
Comp.	Ligand–protein binding mechanism	Dose range	Toxic side effect	Safety conclusion	Phase	Ref.
 MTA		N/A	N/A	N/A	Preclinical	21
 **4** (PF0693999)	SAM-binding pocket. It forms a crucial interaction with Phe327	0.5–12 mg daily (QD or BID) dose	One patient (6 mg BID) had grade 4 treatment-related thrombocytopenia. Treatment-related adverse events occurred in 24 cases (86%)	6 mg QD was identified as the recommended monotherapy dose for expansion	Clinical termination	51–53
 **11** (Lly283)		N/A	N/A	N/A	Preclinical	33
Structure not disclosed SKL27969	N/A	N/A	N/A	N/A	Phase I terminated	24
 **17** (JNJ64619178)	SAM-binding pocket and reaches the substrate-binding pocket	2 mg QD and 3–4 mg 14 on/7 off	The dose-limiting toxicity was thrombocytopenia; 51 patients (91%) experienced a grade 3/4 trae with a treatment-related adverse event (TRAE) > 1 pt. Thirty patients (56%) experienced dose interruptions or reductions due to AEs	0.5 mg once daily was well tolerated 1.5 mg intermittent and 1.0 mg once daily dosing. Demonstrated manageable dose-dependent toxicity, however limited clinical benefit other than ACC	Phase I	19, 20, 65, and 66
	The substrate binding pocket and	12.5 mg to 600 mg once	In phase I of the clinical trial for patients with solid	A comprehensive assessment, AE is	NCT03614728 Terminated;	67–70

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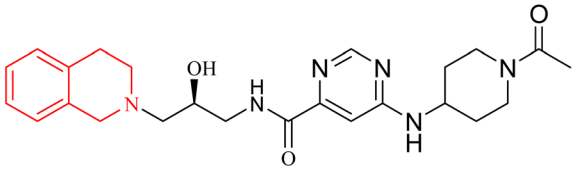
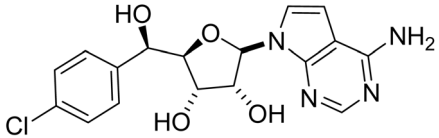
Table 1 (*Contd.*)

Comp.	Ligand–protein binding mechanism	Dose range	Toxic side effect	Safety conclusion	Phase	Ref.
 **22** (GSK3326595)	competes with the substrate	daily (QD) and 50 mg to 200 mg twice daily	tumour cancers, 89% of patients experienced treatment-related adverse events. Of these, 3/4 adverse events occurred	common but manageable	NCT04676516 completed; NCT02783300 completed	
 **25** (PRT543)	SAM-binding pocket. It forms a crucial interaction with Phe327	35 mg (n = 28) or 45 mg (n = 28)	Grade 3 treatment-related adverse events were experienced by 23% of patients, no grade 4/5 treatment-related adverse events were reported	The drug was observed to be tolerated in patients with R/M ACC, but its efficacy was limited	Phase I Completed	38 and 39
Structure not disclosed **26** (PRT811)		1.15–800 mg orally once daily 2.300 mg twice daily [BID]	One patient with GBM had grade 3 thrombocytopenia. 69% (n = 31) experienced adverse events. No deaths were reported	Well tolerated, with a good safety profile under either QD or BID regimens	Phase I Completed	23, 40 and 41

[a] N/A: Information not disclosed.

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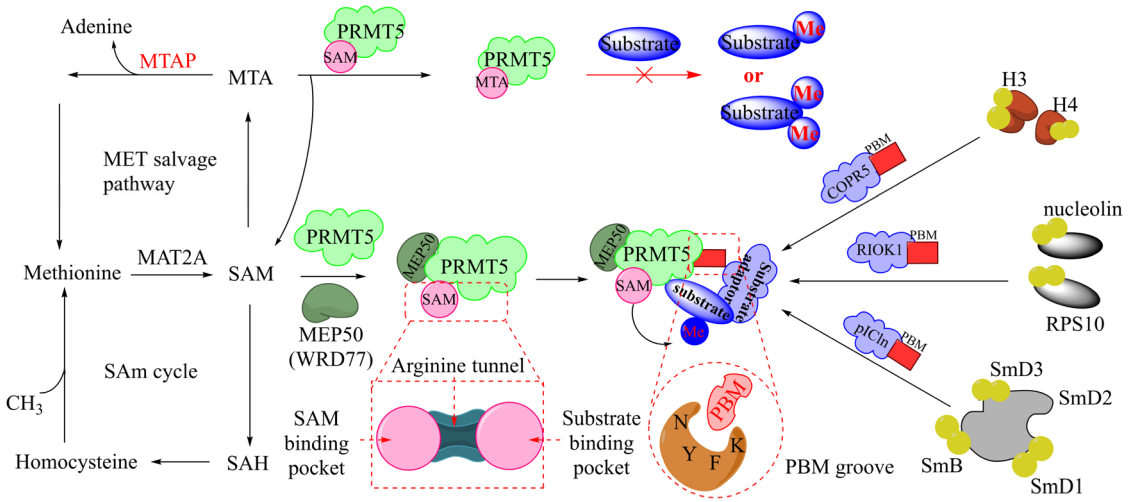

Fig. 4 The upstream and downstream molecular regulatory mechanisms of PRMT5. The substrate binding site and SAM-binding pocket are closely connected by the arginine tunnel, which can be exploited by certain inhibitors, such as **17** (JNJ-64619178), to block both the SAM-binding site and substrate binding pocket simultaneously. Furthermore, PRMT5's activity and substrate selectivity are regulated *via* the formation of multisubunit protein complexes. The SAPs, including Rio domain-containing protein (RioK1), cooperator protein of PRMT5 (COPR5), and chloride channel nucleotide-sensitive protein 1A (pICln), form the multisubunit complexes with PRMT5 by the interaction between the PBM and the PBM-binding groove.[5,21,26–30]

effective dose range of **27** (MRTX1719), there was a notable reduction in normal bone marrow SDMA levels (about 52% were inhibited) in comparison to the near-total loss of SDMA in MTAP-deficient tumours.[90] It has been reported that, *in vitro*, MTAP WT cells are capable of metabolising enriched MTA in MTAP del tumour cells. Furthermore, MTA is not enriched in human MTAP-deficient glioblastoma tissue.[91]

In addition, a series of studies have demonstrated that the PRMT5 enzyme is susceptible to further modulation by SAM levels in MTAP-null tumour cells, whereas its activity is either unaltered or less affected in normal tissues and in tumours where MTAP is retained. By inhibiting the upstream metabolic enzyme methionine adenosyltransferase II alpha (MAT2A) activity, SAM levels in MTAP-deleted cancers can be reduced, and selective inhibition of PRMT5 methylation activity is then possible (see Fig. 4). The MAT2A inhibitor **34** (AG-270) is currently being evaluated in clinical trials (NCT03435250).[92–96] However, we will not discuss it here because the drug target MAT2A is beyond the scope of this article.

It should be noted that caution should be exercised when interpreting SAR data across references since the data presented in this article are collected from different references. In addition, all the schematic diagrams and structures in this paper are summarised from the previous research results, and were created using the drawing software Kingdraw Professional 5.0. The crystal structure data were obtained from RCSB Protein Data Bank (https://www.rcsb.org/pages/about-us/index) (https://www.rcsb.org/). The crystal structure was created by PyMol

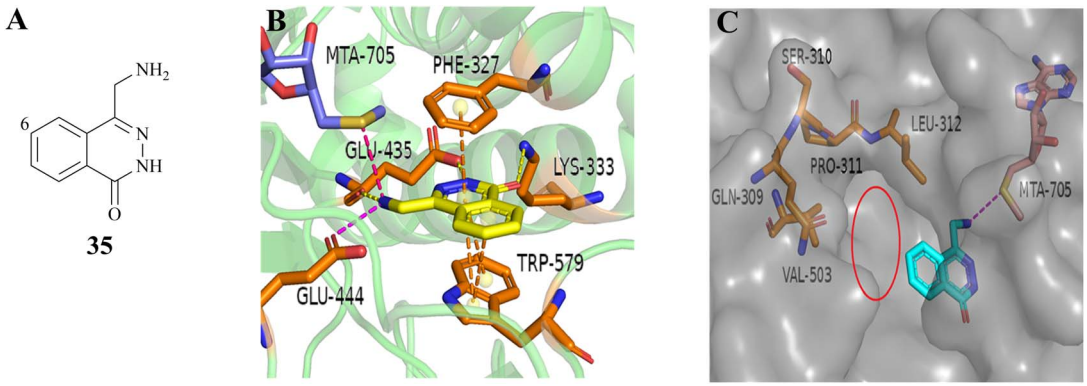

Fig. 5 (A) Chemical structure of **35**. (B) The co-crystal structure of fragment **35** with PRMT5/MTA complex (PDB ID: 7S0U). Fragment **35** forms a hydrogen bonding network with Glu435 and Lys333. Additionally, it is observed that there is an ionic interaction between the primary amine ($-NH_2$) of **35** and Glu444, a π–stack interaction between **35** and the side chains of Phe327 and Trp579, and a van der Waals interaction between **35** and the S-atom of MTA. (C) To investigate the pocket formed by five amino acid residues including the residues Leu312, the C6-position of **35** is the most appropriate position for fragment growth.[87]

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Table 2 Representative second-generation PRMT5 inhibitors in clinical trials[a]

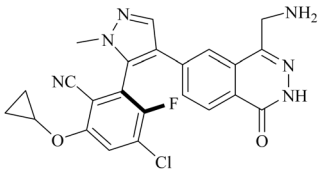
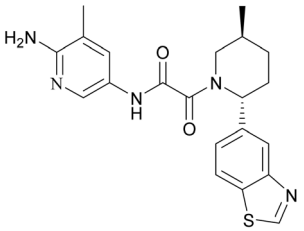
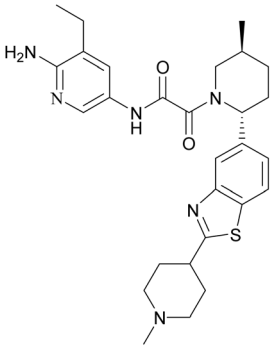
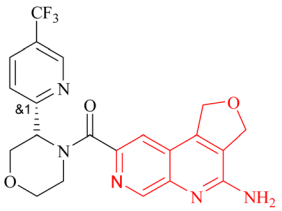
Comp.	Mechanism of action	Mechanisms of tumour cell selectivity	Dose range/ toxic side effect	Safety conclusion	Phase	Ref.
 **27** (MRTX1719)			50–800 mg orally once daily No dose-limiting adverse events associated with non-MTAP-selective PRMT5 inhibitors at dose levels of 400 mg q.d	MRTX1719 was well tolerated and no dose-limiting toxicity was observed at dose levels up to 400 mg q.d	Phase 1/2 recruiting	90, 91, 102, and 103
 **28** (TNG908)			N/A	N/A	Phase 1/2 recruiting	104
 **29** (TNG462)	Substrate competitiveness model. PRMT5/ MTA complex inhibitor	Synthetic lethal	N/A	N/A	Phase 1 recruiting	105
 **30** (AMG193)			40–1600 mg per day or 600 mg per day treatment-related adverse events were nausea (48.8% n = 80), fatigue (31.3%) and vomiting (30.0%). Dose-limiting toxicity was reported in 8 patients at a dose of 240 mg	The maximum tolerated dose was determined to be 1200 mg per day, and AMG 193 demonstrated a favourable safety profile without clinically significant myelosuppression	NCT06593522 phase 2 not yet recruiting NCT05975073 Phase 1/2 recruiting; NCT06360354 Recruiting NCT06333951 Recruiting NCT05094336 Recruiting	106–111

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Table 2 (*Contd.*)

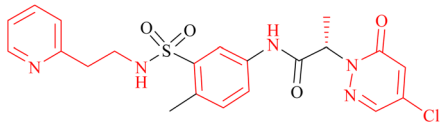
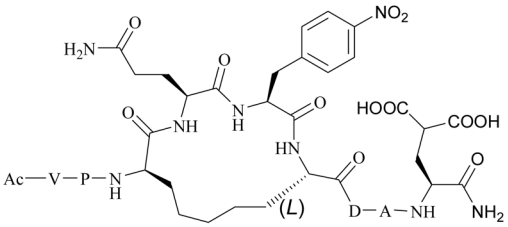
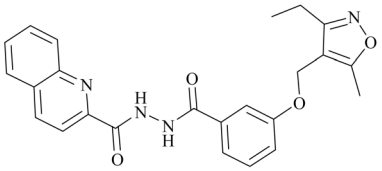
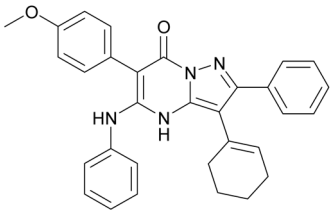
Comp.	Mechanism of action	Mechanisms of tumour cell selectivity	Dose range/ toxic side effect	Safety conclusion	Phase	Ref.
 **31** (BRD0639)			N/A	Poor *in vivo* stability and safety	Preclinical	31
 **32** (Cyclopeptide)	Occupy the PRMT5 PBM groove		N/A	Poor *in vivo* stability and safety	Preclinical	32
 **33** (PRMT5:MEP50 PPI inhibitors)	Impeding PRMT5 and MEP50 protein–protein surface reactions		N/A	Poor *in vivo* stability and safety	Preclinical	25
 **34** (AG-270)	MTA-Cooperative MAT2A inhibition		50 mg once daily (QD; $n = 3$), 100 mg QD ($n = 7$), 150 mg QD ($n = 6$), 200 mg QD ($n = 11$), 400 mg QD ($n = 6$), or 200 mg twice daily (BID; $n = 6$) Three patients developed grade 2 and 3 diffuse erythema during the second week of administration. Reversible but dose-limiting grade 3 and 4 liver enzyme elevations in two patients treated with 200 mg of BID	AG270 reduced plasma [SAM] and tumour SDMA levels at well-tolerated doses	Termination	92 and 93
Structure not disclosed SCR6290 (SCR6277)	N/A	Tumour-biased distribution. Intracellular enrichment concentration 50-fold greater	Daily doses of 10 to 160 mg Seventeen patients (81%) experienced treatment-related adverse events (TRAEs).	No dose-limiting toxicity (DLT) and grade 4 or 5 TRAE reported has a manageable safety profile	Phase I recruiting	112 and 113

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Table 2 (*Contd.*)

Comp.	Mechanism of action	Mechanisms of tumour cell selectivity	Dose range/ toxic side effect	Safety conclusion	Phase	Ref.
		than blood concentration	six patients experienced serious adverse events (SAEs) in which chronic colitis was associated with treatment			

[a] N/A: Information not disclosed.

2.1.0, a software for displaying the 3D structure of protein molecules.

2 MTA-cooperative PRMT5 inhibitors

It is well known that a lead compound with good druggability is critical to modern drug discovery. The identification of a lead compound in a timely and accurate manner has constituted a significant obstacle in the field of drug discovery. In recent years, a number of innovative drug screening techniques have emerged, including high-throughput screening (HTS), artificial intelligence-based drug molecule virtual screening (AIDD), fragment-based molecule library screening (FBDD), and structure-based molecule design (SBDD). These techniques offer potential solutions to this challenge. Among them, the concept of FBDD originated approximately 30 years ago. However, the rapid development of a wide range of experimental analytical techniques had made it an effective tool for modern drug discovery by the early 21st century. The implementation of FBDD is generally divided into three steps: the initial step is to identify a comprehensive and high-quality library of small molecules, or fragments. These small molecules should typically adhere to the Rule of Three (RO3) principle (molecular weight less than 300, no more than 3 hydrogen bond donors and acceptors, no more than 3 rotatable bonds, *c* Log *P* less than 3).[97–99] Step 2, determine an appropriate high-sensitivity screening method to identify suitable fragments from the library of fragments. Of these, the most critical of these is the experimental analysis technique used to identify the binding strength of the fragments to the target protein. Currently, the main experimental techniques used in practice include differential scanning fluorometry (DSF), microscale thermophoresis (MST), surface plasmon resonance (SPR), nuclear magnetic resonance (NMR), X-ray protein crystallography (X-ray), and so on.[99,100] Step 3, fragment growth and optimisation. The initial fragments obtained from FBDD screening are typically characterised by low activity. To enhance their target affinity and other properties, such as solubility, membrane permeability and bioavailability, these fragments must undergo structural extension and optimisation. In this process, SBDD is typically employed as the primary methodology to direct the expansion and enhancement of fragments.[101] SBDD represents a drug design method based on the concept of molecular recognition, which employs the three-dimensional structure of ligands and targets. In comparison to the conventional HTS approach, FBDD exhibits enhanced exploration efficiency, broader chemical space coverage, more favourable binding modes, and reduced input/output. These advantageous characteristics, coupled with the insights gleaned from the crystal structure of the protein, have led to the extensive utilisation of these innovative drug screening techniques, particularly the integration of FBDD and SBDD, in the development of MTA-cooperative PRMT5 inhibitors and structure–activity relationship (SAR) exploration, with notable success.

2.1 PRMT5/MTA complex inhibitors

As mentioned above, PRMT5 and MTAP have been supposed to be the next potential synthetic lethal pair and several PRMT5/MTA inhibitors are currently undergoing clinical trials. Among them, **27** (MRTX-1719), as one of the most clinical application value PRMT5/MTA inhibitors, has made the fastest progress (the clinical phase I/II: NCT05245500). The 4-(aminomethyl)phthalazin-1(2*H*)-one fragment (**35**, see Fig. 5A), as the starting point for the development of **27** (MRTX1719), was confirmed *via* a fragment-based lead discovery approach. Firstly, SPR was used as a screening platform because of its high sensitivity, ability to detect weak binding, *etc.* Matthew A. Marx *et al.* determined the K_D (K_D = 27 μM) for the binding of the PRMT5-MTA complex to EPZ015666 and used it as a positive control. In the initial phase of the screening process, approximately 1000 fragments were evaluated at a single concentration of 100 μM, leading to the identification of 17 fragmented compounds. In the subsequent phase, two additional fragment libraries, namely a diverse library comprising 1692 fragments and a focused library constructed of 194 specifically selected compounds, were screened at a concentration of 500 μM, resulting in the identification of a total of 188 hit points. Subsequently, the 205 fragments obtained from the screening

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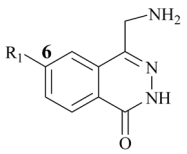
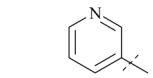
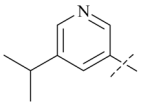
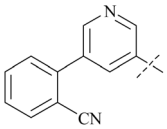
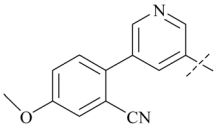
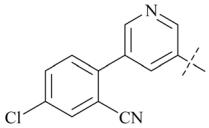
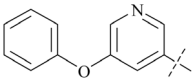
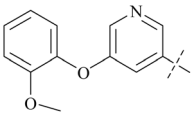
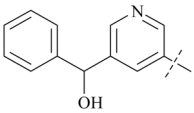
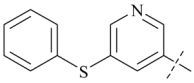
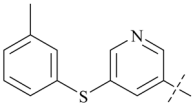
Table 3 SAR study of the C_6-position of fragment **35**



Comp.	Ex:#[a]	R_1	PRMT5 biochemical IC_{50} (nM)			Ref.
			MTA^+	MTA^-	MTA^+/MTA^-	
36	3		910	19 000	21	87
37	3-14		17	413	24	88
38	3-40		6	193	32	88
39	8-3		2.3	237	103	88
40	8-4		3.1	313	100.9	88
41	3-50		10	277	27.7	88
42	4-78		0.5	66	132	88
43	4-51		5.8	530	91.37	88
44	4-55		1.2	30	25	88
45	4-61		3	149	49.66	88

[a] Example numbers in the table are the same as those in the cited references.

were subjected to PRMT5-MTA K_D assay, and 100 of them exhibited saturation K_D values lower than 1 mM. From these, 24 fragments were selected for further characterisation with the objective of determining whether they preferentially bind to the PRMT5/MTA complex. The results demonstrated that the binding K_D of PRMT5-MTA ranged from 300 nM to 300 μM, while the selectivity for the PRMT5-SAM complex ranged from 0.2 to 35-fold, among the 24 fragments that were tested.

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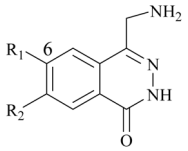
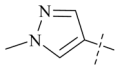
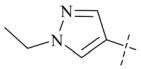
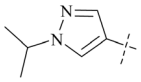
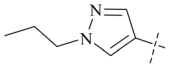
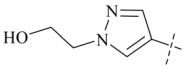
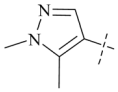
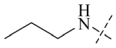
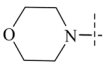
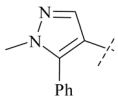
Table 4 Further SAR study about the C_6-position of fragment **35**



Comp.	Ex:#[a]	R_1	R_2	PRMT5 biochemical IC_{50} (μM) MTA$^+$	MTA$^-$	MTA$^+$/MTA$^-$	Ref.
46	9		H	0.30	5.8	19	87
47	10		H	0.51	10.6	21	87
48	11		H	0.67	11.4	17	87
49	12		H	0.32	7.65	24	87
50	13		H	0.71	11.2	16	87
51	14		H	0.05	1.29	26	87
52	6-2	H		>10^2	>10^2	—	88
53	5-1		H	72	>10^2	—	88
54	6-5		H	>10^2	>10^2	—	88
55	5-2		H	57	>10^2	—	88
56	4-48		H	1.6 (nM)	0.045	28	88

[a] Example numbers in the table are the same as those in the cited references.

Fragment 35, which exhibited a PRMT5-MTA K_D of 10 μM and a PRMT5-SAM K_D of 51 μM, was selected for further fragment growth and SAR exploration. SBDD is used as a guiding strategy.

In the co-crystal structure (PDB ID: 7S0U, Fig. 5B), it can be observed that the fragment **35** binds in the substrate-binding pocket of PRMT5 *via* a series of diverse and specific mutual effects. Further analysis of the co-crystal structure revealed the presence of a pocket constituted by five amino acid residues, including the residue Leu312, at the front of the C6-position of the **35** (see Fig. 5C). Molecular modelling using MOE suggested that it can accommodate a five or six-membered heterocyclic ring, in which the heteroatom with the lone-pair electron might make a beneficial H-bond interaction with Leu312.[87]

From this, a series of six-membered heterocyclic ring substituted compounds of **35** (see Table 3 for its representative compounds) were prepared. These compounds display high potency and selectivity in the PRMT5 biochemical assay, particularly compounds **39** (MTA$^+$ IC_{50} = 2.3 nM, MTA$^-$ IC_{50} = 237 nM, selectivity ratio MTA$^+$/MTA$^-$ = 103) and **40** (MTA$^+$ IC_{50} = 3.1 nM, MTA$^-$ IC_{50} = 313 nM, selectivity ratio MTA$^+$/MTA$^-$ = 100.9). It is regrettable that no further research data regarding the optimisation of this series of compounds was reported. In parallel, the five-membered heterocyclic ring substituted compounds of **35** were also prepared. Among them, those containing imidazole and N-substituted derivatives of imidazole exhibit a significant increase in potency and selectivity

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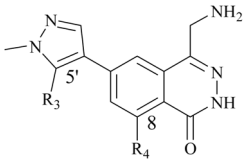
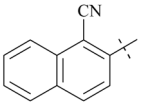
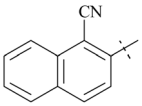
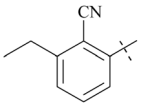
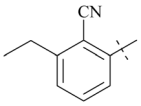
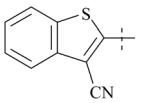
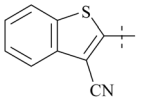
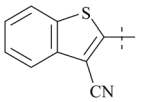
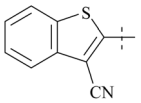
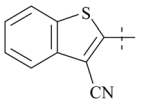
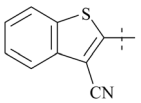
Table 5 The SAR of the 8-position (R_4) in the phthalazin-1(2*H*)-one scaffold **35**



Comp.	Ex: # [a]	R_3	R_4	SDMA IC_{50} HCT116 MTAP-del/MTAP-WT (nM)	Viability IC_{50} HCT116 MTAP-del/MTAP-WT (nM)/selectivity ratio	Ref.
61	10-1		–Cl	15/1123	36/2299/64	88
62	10-2		–F	6/2484	112/3378/30	88
63	10-3		–Cl	7/9215	128/>10^4/78	88
64	10-7		–CH_3	31/1000	103/8386/81	88
65	2		–H	5/>1000	58/5800/100	114
66	9		Cl–	17/>1000	52/734/14	114
67	10		–CH_3	17/>1000	30/1470/49	114
68	11		F_2HC–	25/759	37/2030/55	114
69	12		–OH	262/103	188/4350/23	114
70	**13**		–OCH_2CH_3	112/618	150/637/4	114

[a] Example numbers in the table are the same as those in the cited references.

compared to the **35**, especially compound **46** (see Table 4). The co-crystal structure of **46** with PRMT5/MTA (PDB 7S1Q, Fig. 6B)[87] indicates that the *N*-methylpyrazole-4-yl group occupies the aforementioned pocket (Fig. 5C) and a new H-bond is observed between the backbone N–H of Leu312 and the N_2 of *N*-methylpyrazole. Further investigation of the SAR shows that using any of the -ethyl, -propyl, -isopropyl, and -hydroxyethyl instead of H-atom of the pyrazole $N_{1'}$ is also tolerated (compounds **47–50**, see Table 3). Replacing the H-atom of the $C_{5'}$ position of the *N*-methylpyrazol-4-yl group with a methyl

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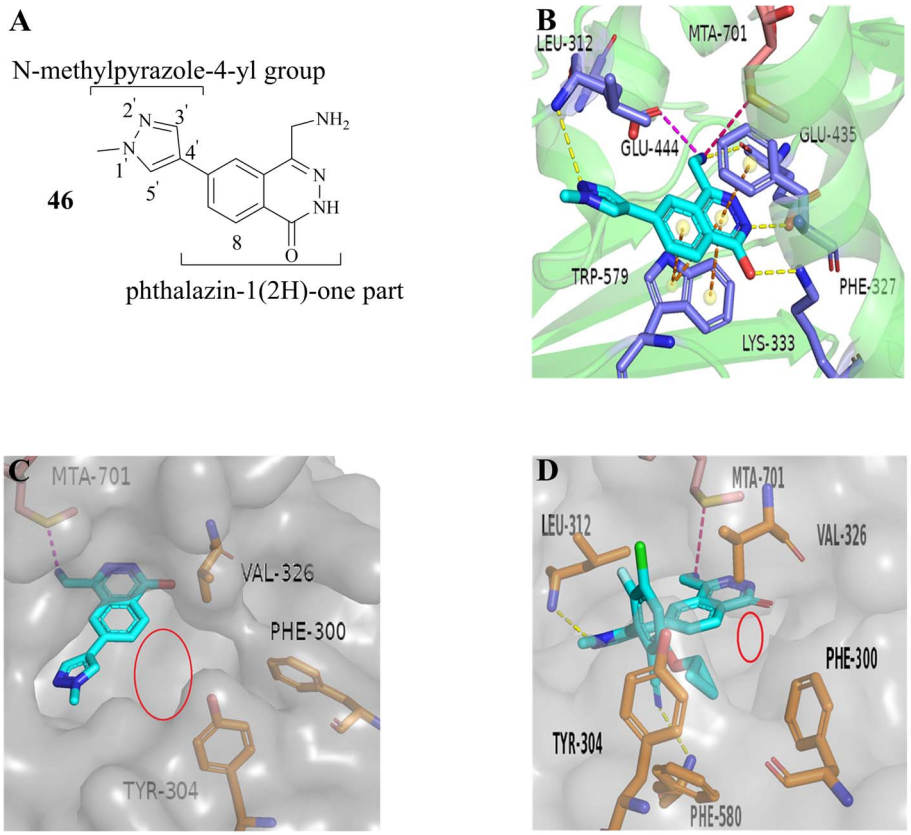

Fig. 6 (A) Structure of **46**. (B) The co-crystal structure of **46** with PRMT5/MTA (PDB 7S1Q). A new H-bond was observed between the backbone N–H of Leu312 and the N_2 of *N*-methylpyrazole. (C) The lipophilic pocket formed by the side chains of Phe300, Tyr304 and Val326 is observed and there is enough room to accommodate a lipophilic group. (D) The co-crystal structure of **27** (MRTX1719) with PRMT5/MTA (PDB 7S1S). Additionally, there is also a smaller cavity at the side of the 8-position in the phthalazin-1(2*H*)-one that could accommodate a group.[87]

group (compound **51**) generates a 10-fold boost in inhibition compared to **46**. Moreover, Phe300, Tyr304 and Val326 form the other lipophilic pocket and there is enough room to accommodate a lipophilic group (see Fig. 6C). Subsequently, compounds **56–60** were designed, synthesized, and tested (see Fig. 7 for structure and data).

The ^{1}H NMR spectra of compounds **57** and **58** indicate that they might be slowly interconverting rotational isomers. Then, atropisomers **27** (MRTX1719) and **(*P*)-27** (it is the enantiomer of **27** (MRTX1719), the structure is not shown here) are successfully separated by chira supercritical fluid chromatography (SFC). The co-crystal structure of **27** (MRTX1719) with PRMT5/MTA (PDB: 7S1S Fig. 6D) shows that the binding mode of compound **46** was maintained. The newly introduced 2-cyclopropoxy-4-chloro-5-fluoro-6-yl-benzo-nitrile substituent enters the lipophilic pocket and is oriented perpendicular to the *N*-methylpyrazole group. The nitrile plays a crucial role in improving the potency and selectivity of **27** (MRTX1719) due to the formation of a favourable H-bond between the nitrile and Phe580.

The F atom at the 5-position of the benzonitrile group points toward Leu312. However, using the Cl atom instead of F leads to an obvious loss of potency in the *MTAP*-del viability assay, and that is likely because the larger Cl-atom substituent creates a collision with the side chain of Leu312. **59** (M-31) was obtained by using the same SFC method.[87,115]

Based on the calculated torsion rotational energy, atropisomers are usually classified into three classes. The acquisition of **27** (MRTX1719) and **59** (M-31), which belong to class 3 (slow to no rotation), necessitates the use of chiral syntheses or kinetic racemic resolution (see Fig. 7). Significantly, the acquisition of compound **60**, as a class 1 compound (free rotation), does not need the chiral syntheses or kinetic racemic resolution. It exhibits high potency and selectivity (*MTAP*-del IC_{50} = 58 nM, *MTAP*-WT IC_{50} = 5800 nM, *MTAP*-WT/*MTAP*-del = 100 fold in the HCT116 viability). However, the human hepatocyte intrinsic clearance (Cl_{int} = 41 mL min^{-1} kg^{-1}) of **60** is higher than that of **27** (MRTX1719 Cl_{int} < 18 mL min^{-1} kg^{-1}). After considering all cases, including but not limited to the potency, selectivity, and PK profiling in preclinical species, **27** (MRTX1719) was selected for clinical trials.[87,88,114,115] In the meantime a scalable synthesis of this atropisomeric compound and an efficient isolation of the desired isomer have been established to support the clinical trials through further development of the racemic medicinal chemistry route.[116,117] Additionally, there is a smaller cavity at the side of the 8-position in the phthalazin-1(2*H*)-one **35**, which could accommodate a group (see Fig. 6C and D). And then, the SAR about the 8-

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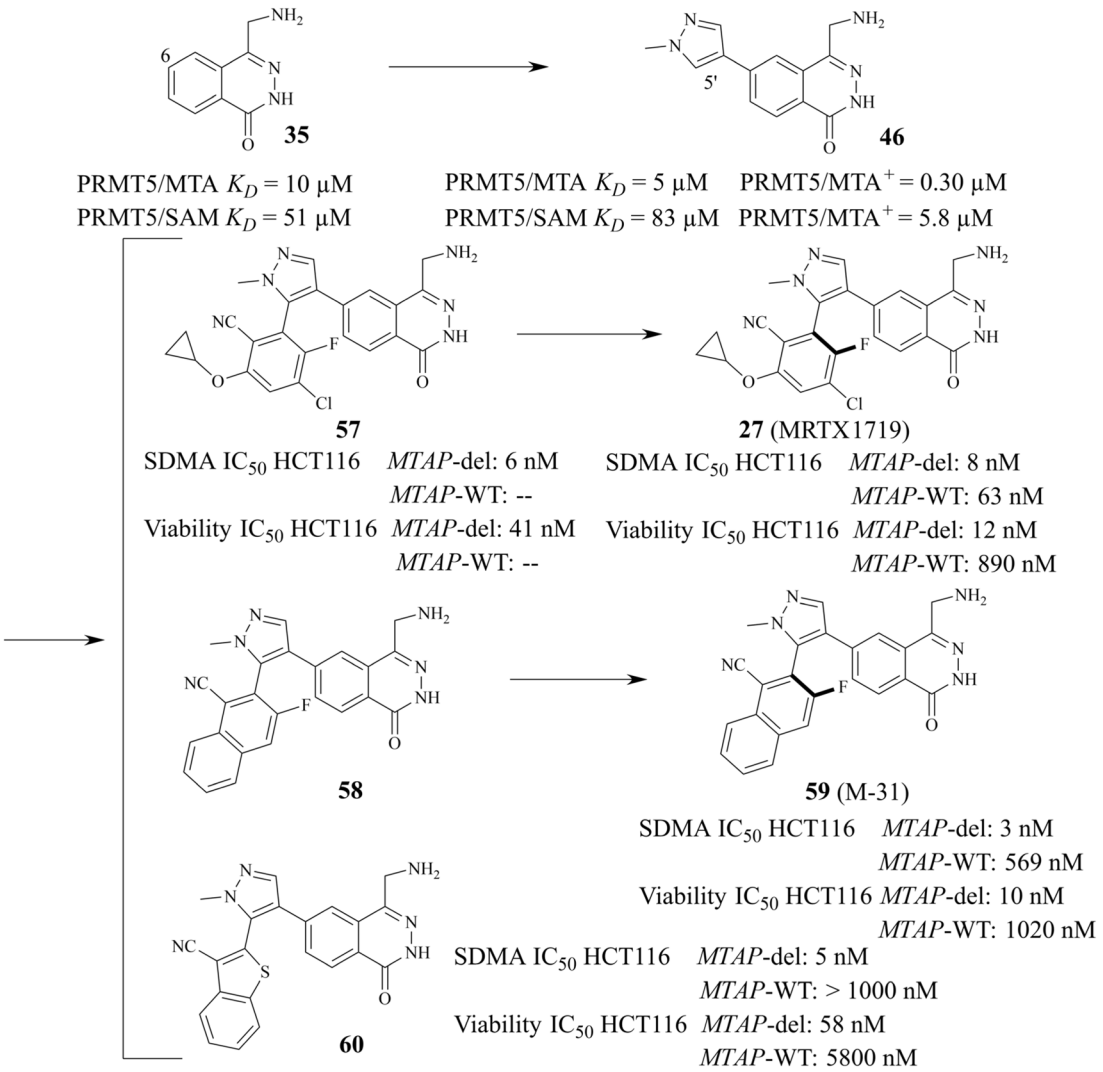

Fig. 7 The fragment **1** growth route of **27** (MRTX1719) and its analogues.

position of the phthalazin-1(2*H*)-one **35** was also investigated (the data and structure are given in Table 5). In general, a small substituent (such as compounds **61–68**) at the 8-position can show higher potency and selectivity than that of the larger substituents (such as compound **70**). For the compounds in Table 5, it can be observed that –CH_3-substituted compounds at this position consistently exhibit the highest potency in the HCT116 MTAP-del viability assay. Furthermore, an emerging

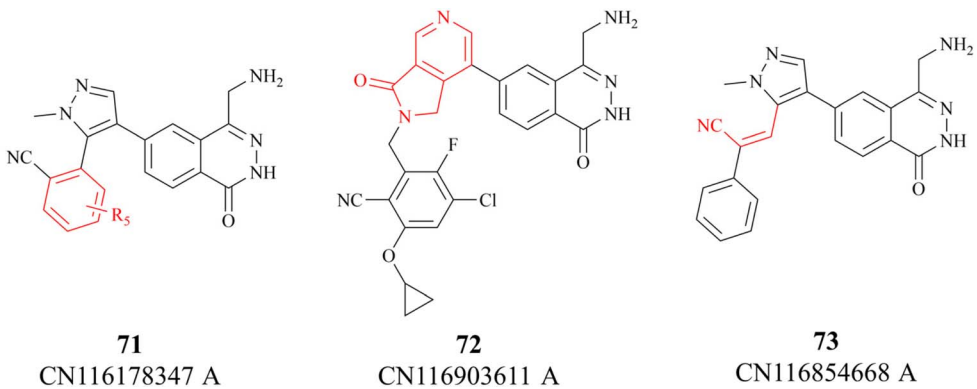

Fig. 8 **27** (MRTX1719) analogues.

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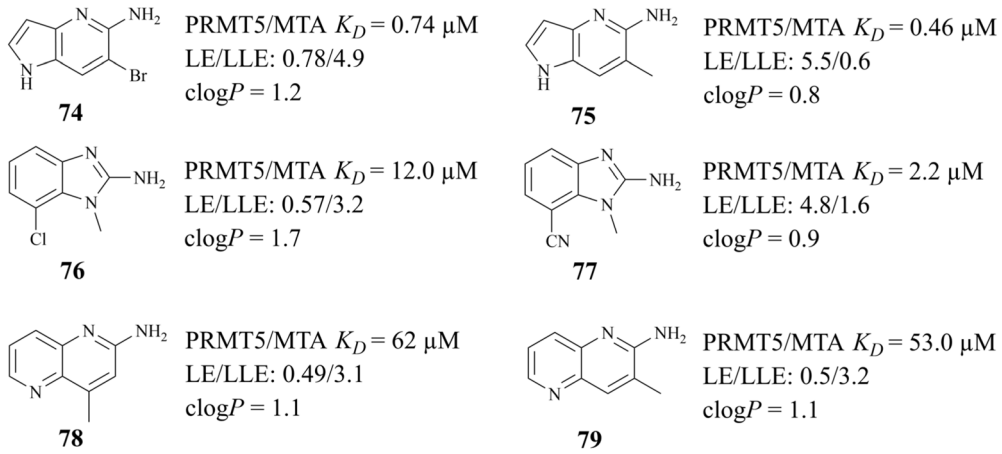

Fig. 9 Analogues of fragment **35**.

trend suggests that $-CH_3$ > –Cl > $-CHF_2$ > –F > –OH. However, their selectivity ratio (viability IC_{50} HCT116 MTAP-del/MTAP-WT) is markedly decreased compared to –H-substituted compounds (such as compound **65**). It is worth noting that the substituents of the 8-position in the **35** may have an effect on the human hepatocyte intrinsic clearance (Cl_{int}). For example, the Cl_{int} of compound **67** decreases to 27 mL min^{-1} kg^{-1} and the Cl_{int} of both (compounds **69** and **70**) is below 18 mL min^{-1} kg^{-1}, while compound **65** is in the high range (Cl_{int}: 41 mL min^{-1} kg^{-1}). The $-CHF_2$-substituted compound **68** (Cl_{int}: 44 mL min^{-1} kg^{-1}) remains similar to **65**. However, the Cl_{int} of –Cl-substituted compound (**66**) increases to 71 mL min^{-1} kg^{-1}.

In 2023, partial details of the SAR of the **27** (MRTX1719) are further revealed by three Chinese patents (see Fig. 8. These details are highlighted in red). Among them, patent CN116178347A discloses some new information about the substituents on the benzo nitrile. Moreover, compounds **72** and **73** have similar potency as **27**, possibly indicating a similar binding conformation for these three compounds (the data is not shown here).[118–120]

The success of **27** (MRTX1719) demonstrates that the development strategy combining the FBLD method with the co-crystal structure-based drug design (SBDD) is a highly feasible way for the discovery of PRMT5/MTA complex inhibitors. There are still several analogues of fragment **35** (fragments **74–79** shown in Fig. 9). Among them, fragments **74**, **76**, **78** and **79** were discovered together with the fragment **35** (PRMT5/MTA K_D = 10.2 μM, LE/LLE: 0.54/4.9, $c\log P$: 0.1) by using the FBLD approach. And fragments **75** and **77** were obtained by optimising the **74** and **76** respectively. Furthermore, each of these fragments might lead to the discovery of lead compounds just like **27** (MRTX1719). And interestingly enough, Matthew A. Marx *et al.* also describe another drug design strategy—chimeric design. Firstly, further analysis of the co-crystal structure of fragment **74** suggests that the 2-position of **74** is the most appropriate position for fragment growth. Subsequently, compound **80** was obtained by introducing the pyrimidine-4-carboxamide group of EPZ015666 (structure shown in red in Fig. 10) into this position. Compared to the original fragment **74** (PRMT5/MTA K_D = 740 nM), the addition of the pyrimidine-4-carboxamide substituent caused an 82-fold increase in binding K_D (PRMT5/MTA K_D = 9 nM), and the LLE increased from 4.6 to 7.4 (ΔLLE = +2.5). These results demonstrate that it is another economically feasible way to increase the potency of a fragment by a chimeric approach with known ligands.[121,122]

Another useful example in the development of the PRMT5/MTA complex inhibitors is the discovery of **28** (TNG908).[85,86] John P. Maxwell and colleagues initially employed a number of search strategies to identify potential lead compounds for drug discovery, including DEL and fragment screening. Among

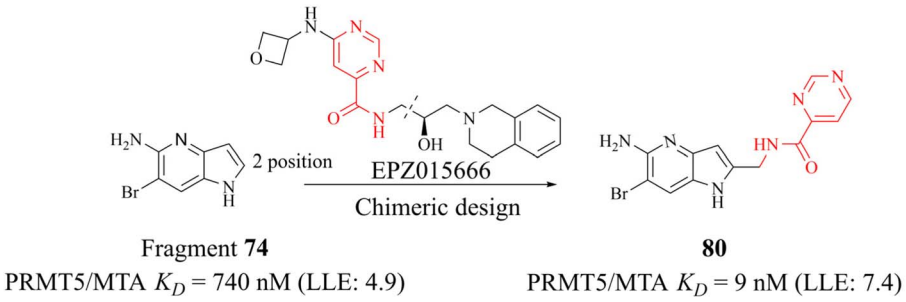

Fig. 10 The chimeric design enabled optimisation of fragment **74** to obtain **80**.

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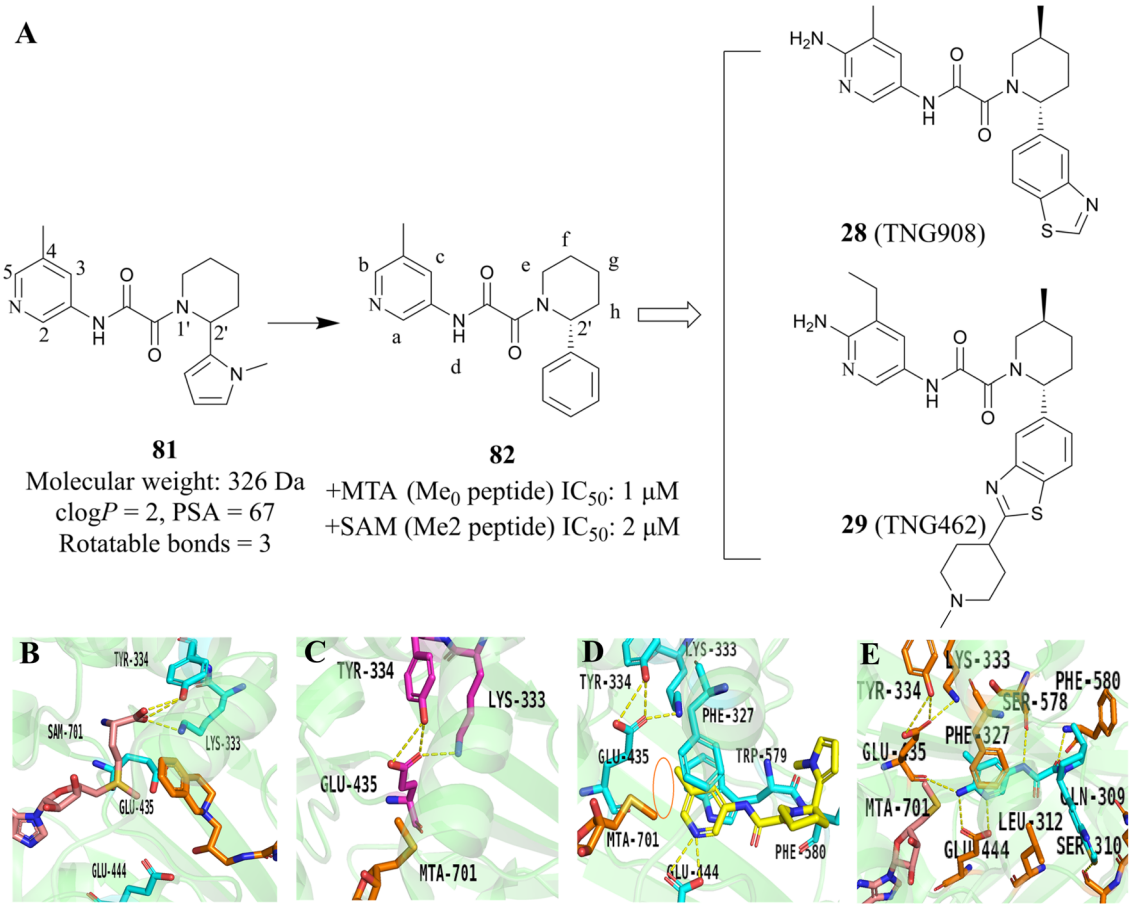

Fig. 11 The development strategy, which combined the HTS and SBDD methodologies, yielded the **28** (TNG908).[85,86] (A) The development route of **28** (TNG908) and **29** (TNG462). (B) X-ray cocrystal structure of PRMT5/SAM/substrate complex. Glu435 rotates toward substrate side chain (PDB 4X61).[21,22] (C) X-ray cocrystal structure of PRMT5/MTA complex. Glu435 rotates to fill space previously filled by SAM (PDB 8VEO).[21,22,85] (D) X-ray cocrystal structure of **82R** (yellow) bound to PRMT5/MTA (PDB 8VET).[85] (E) X-ray cocrystal structure of **28** (TNG908) bound to PRMT5/MTA (PDB 8VEY).[85]

these, a high-throughput screen of a 560 000-compound library at 20 μM using peptide substitution yielded a promising result. The fluorescence anisotropy (FA) method was employed to detect the displacement of a tamra-labelled histone H4 peptide from PRMT5 in the presence of 50 μM MTA. Subsequently, the compounds were subjected to further evaluation. Ultimately, 11 unique structural series, comprising 21 compounds, were obtained. Several series were eliminated due to an inability to demonstrate SAR traceability. Compound **81**, one of the three remaining structural series, was selected as the lead compound and SBDD was employed as a guide for further exploration of its SAR. (Structure and physicochemical properties of **81** are presented in Fig. 11A). Preliminary SAR studies of compound **81** indicate that the 2-position of the piperidine is tolerant to a variety of different substituents, including phenyl, cyclohexyl and pyridyl. However, it is intolerant to smaller substituents such as methyl. The substitution at this position exhibits a clear stereochemical preference, with the preferred stereochemistry being R. This information was used to obtain compound **82**.[85]

Subsequently, methyl groups were introduced at each position of **82** (positions a–h, as shown in Fig. 11A) in order to further investigate the SAR of **82**. The introduction of a trans methyl group at the f-position resulted in a 20-fold increase in potency while maintaining selectivity. During the SAR investigation of **82**, enantiopure isomers **82R** (the structure is not shown here) were obtained by chiral resolution of racemic **81** using SFC.

In order to obtain a compound that could cross the blood–brain barrier (BBB), the SBDD was employed to guide the further optimisation of compound **82**. The X-ray cocrystal structure of **82R** with the PRMT5/MTA complex reveals that the pyridine ring establishes van der Waals contacts with MTA and engages in π–stacking interactions with Phe327 and Trp579. Additionally, the N-atom on the pyridine ring forms a hydrogen bond with Glu444. It is noteworthy that the methyl group at the 5-position of the pyridine ring points to Lys333 and Glu435, thus locking Glu435 in the preferred spatial configuration for binding to MTA (see Fig. 11D). This prevents Glu435 from forming the rotamer necessary for binding to SAM, which helps to further explain the high selectivity of the PRMT5/MTA inhibitor (Fig. 11B and C). Furthermore, a small space near the 5- and 6-positions of the pyridine ring was observed to

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accommodate some small groups. The NH of oxamide forms a hydrogen bond with Ser578, and the carbonyl group of oxamide is also involved in the formation of the hydrogen bond (Fig. 11D).

Based on these understandings, the introduction of an amino group into the 6-position of the pyridine ring of **82** results in a 200-fold increase in potency compared to compound **82**, demonstrating a significant selectivity boost (HAP1 *MTAP*-null ICW 17 nM, HAP1 *MTAP*-null viability 420 nM, 14-fold viability selectivity). The substitution of the methyl group at the 5-position of the pyridine ring with the aminocarbonyl group leads to a 120-fold increase in potency relative to that of **82**, while exhibiting higher selectivity (30-fold viability selectivity). Further studies of the substituent at the 2′-position of the piperidine revealed that the introduction of a hydroxyl group in the *para*-position of the benzene ring may form a beneficial hydrogen-bonding interaction with the carbonyl of Ser310. However, it is observed that those compounds containing more hydrogen bond donors, such as NH and OH, display a tendency to have lower penetration and higher efflux than their matched compounds without additional hydrogen bond donors. Furthermore, they exhibit lower human liver microsomes stability. (The structure of these compounds is not shown here). Interestingly, an uncommon interaction of a C–S σ* orbital and the carbonyl of Ser310 is observed when the benzothiazole is used to replace the phenyl. This interaction demonstrates that it is possible to achieve a higher valence without the addition of an additional hydrogen bond donor. In addition, rational cyclisation studies from the pyridine onto the oxamide showed that the pyridine oxamide portion and the fragments depicted in Fig. 9 such as **74** and **75** are bioisosteric groups (it is highlighted in red in Fig. 12).[85,86] Nevertheless, although these cyclised compounds exhibit notable activity, they display reduced solubility and metabolic stability relative to their counterparts utilising the oxamide as a linker. By combining these SARs, **28** (TNG908),[86] a potent and selective, brain penetrant PRMT5/MTA inhibitor currently undergoing phase I/II clinical trials (NCT05275478), was discovered. Another structurally similar compound **29** (TNG462),[86] currently undergoing phase 1/2 clinical trials (NCT05732831), was also discovered.[123]

There are several other research teams developing or working in parallel on PRMT5/MTA inhibitors. Amgen Inc. has filed four patents in less than two years (2020–2022) on its PRMT5/MTA complex inhibitors, and the compounds covered by these patents possess a bicyclic (the representative compounds **83** and **84**) or tricyclic (the representative compounds **85–87**) quinolin-2-amine derivative scaffold, which almost all of these compounds exhibit similar biological activities as **27**.[124–128] The candidate compound **30** (AMG-193), as an MTA-cooperative PRMT5i, is in clinical trials (NCT06333951, NCT05094336).[89]

Interestingly, the quinolin-2-amine derivative scaffold is strikingly similar to that of the aforementioned fragment **79** (similar parts of their structures are highlighted in red in Fig. 13). Furthermore, the PRMT5 substrate binding site comprises the well-known double E ring (two acidic glutamate residues, Glu435 and Glu444), which facilitates the recognition of basic groups, including the protein arginine side chain.[28] And it can be postulated that the known PRMT5/MTA complex inhibitors may all have a key pharmacophore. And, the pharmacophores of these known PRMT5/MTA complex inhibitors should share some common features to ensure that specific binding reactions between the pharmacophore and the PRMT5/MTA complex can be formed. For instance, an appropriate aromatic planar structure may facilitate the formation of the π–stack interaction with the side chains of Phe327 and Trp579. And the basic substituents or basic atoms (such as primary amino groups and N-atoms on fragments **35** and **74–79**) on this aromatic planar structure may also be necessary for the formation of a stable specific interaction with Glu444 and MTA. Similarly, another basic or neutral substituent of appropriate size on this pharmacophore, such as the 5-methyl group on the pyridine ring of **28** (TNG908), may potentially improve target selectivity and potency.

2.2 PRMT5-Adaptor protein interaction inhibitors (PAPII)

As previously mentioned, the PBM-binding groove is located in the TIM barrel, and a small molecule which could occupy the PBM groove would affect the methylation of more than 25 PRMT5 substrates. In 2021, Ianari *et al.* disclosed a series of PBM-competitive inhibitors that the **31** (BRD0639) represents.[31] This research outcome remains one of the successful examples of the application of the SBDD methodologies. A competitive fluorescence polarisation (FP) analysis was employed to assess the capacity of compounds to displace the interaction between fluorophore-labelled RIOK1 PBM peptides and purified PRMT5:WDR77 heterodimers. And Ianari *et al.* conducted a screening of over 900K small molecule compounds with the objective of identifying inhibitors of PBM peptide interactions. Then, compound **58** (FP IC_{50} of 12 μM, solubility in PBS of 1.2

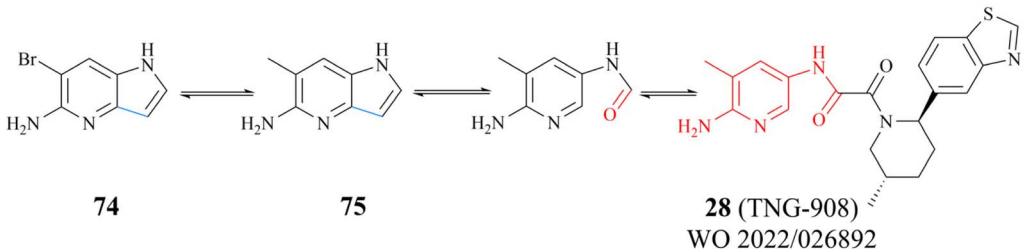

Fig. 12 The structural relationship between the 3-methylpyridin-2-amine pharmacophore and **74**.

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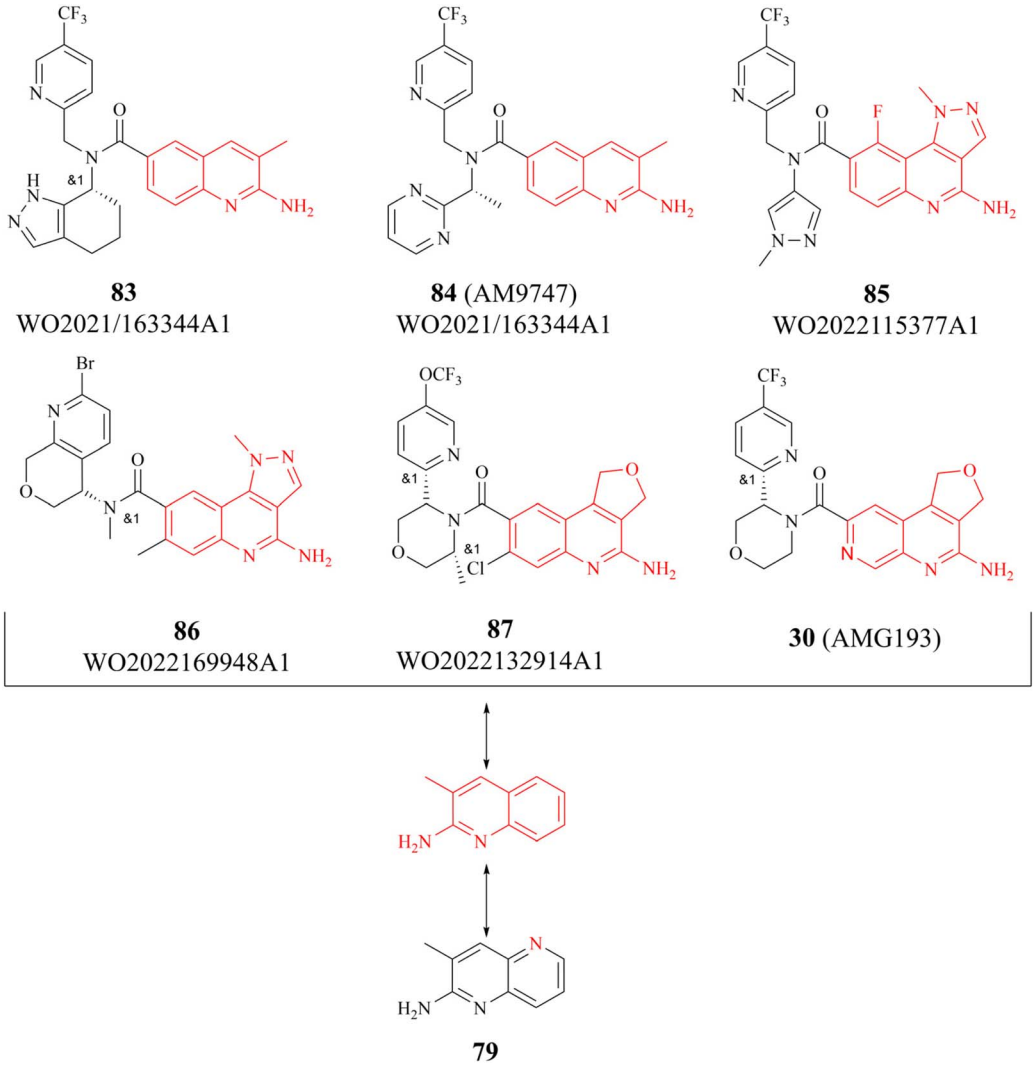

Fig. 13 The heterocycles as PRMT5/MTA complex inhibitors.

μM) was finally identified as the lead compound. The subsequent SAR exploration process was guided by SBDD. This process can be further divided into two phases. Initially, solubility and potency were the primary objectives, whereas in the subsequent phase, a balance between potency and glutathione (GSH) electrophilic reactivity was the key focus. The co-crystal structure of the lead compound **88** (Fig. 14A) with the PRMT5 complex shows that the PBM groove is occupied, and that cys278 makes a covalent bond with the lead compound **88** *via* a nucleophilic substitution reaction. In the co-crystal structure, a four-ring π–π stacking interaction (Tyr286-pyridine-arylcore-Phe243) is observed. Moreover, multiple hydrogen bonding interactions are also observed between the sulfonamide and Asn239, the sulfonamide and Lys241, the acid amides and Ser279, and the pyridazinone and Gln282 (see Fig. 14B). Further SAR studies have revealed that the replacement of the pyridyl group in the R_1-portion with a six- or seven-membered nitrogen heterocyclic ring results in a loss of potency (the structure is not shown here). The pyridine isomers (compounds **89** and **90**) exhibit a similar potency and stability to that of **88** (see Fig. 14C). With regard to the modification of R_2, aromatic six-membered rings are tolerated such as the pyridyl ring (compound **91**). Furthermore, the introduction of a methyl group at the b- or c-position is tolerated (compounds **93** and **94**), while the introduction of a methyl group at the a-position results in a significant loss of potency (compounds **92**). However, these modifications at the a-, b-, and c-positions may cause a reduction in solubility. And the introduction of a methyl group at the d-position is markedly less active (the structure and data are not shown here). A methyl group at the e-position is tolerated, with a very strong stereochemistry preference (the preferred stereochemistry is S such as compound **95**). Meanwhile, the introduction of a methyl group at this position markedly enhances the stability of the compounds due to the steric hindrance (as exemplified by compounds **95** and **96**). The electrophilic reactivity of R_3 showed a negative correlation with the selectivity of the compounds. To achieve an acceptable balance between activity and reactivity, this part was further

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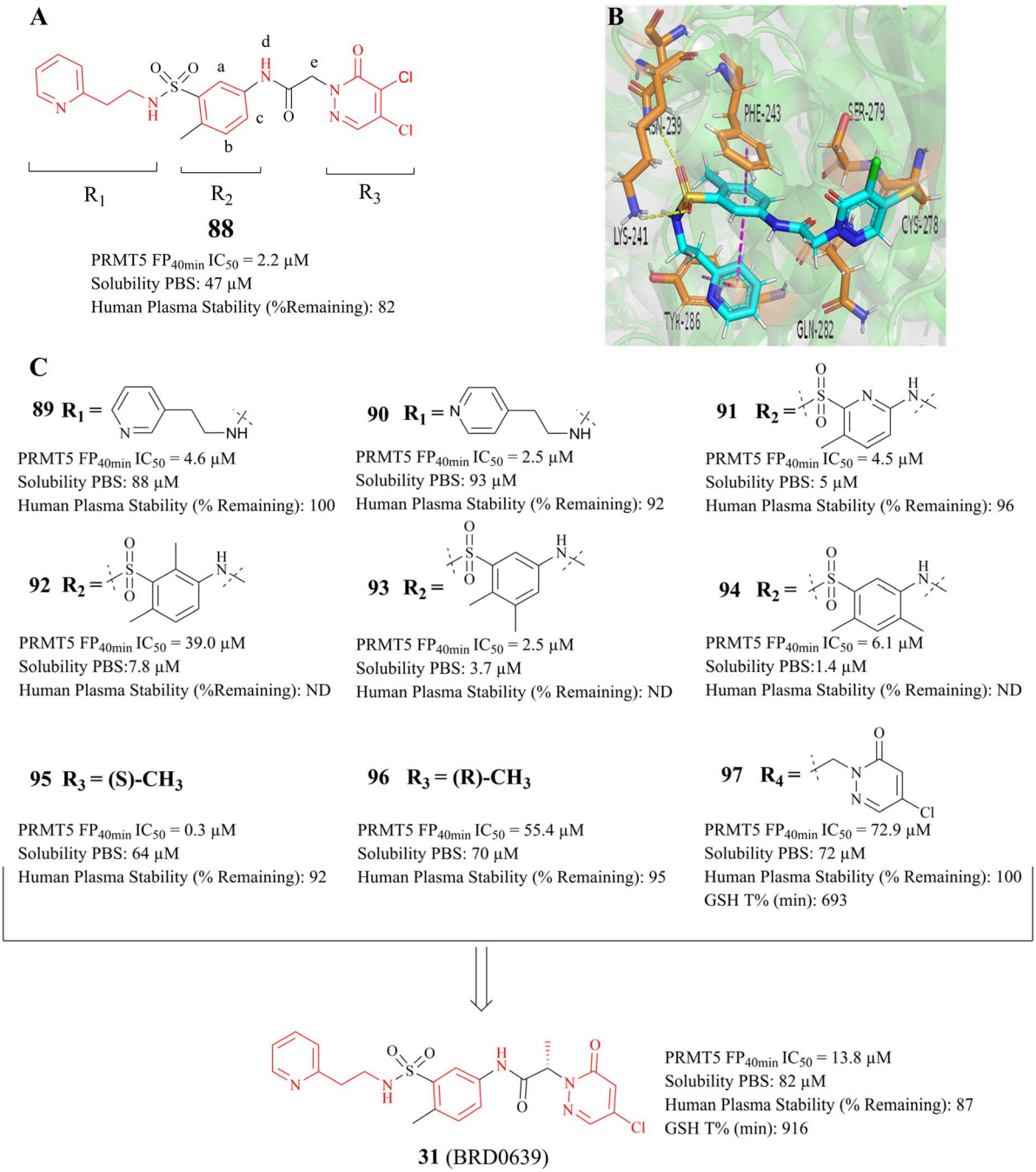

Fig. 14 (A) Lead compound **88**. (B) The cocrystal structure of compound **88** bound to the PRMT5 complex (PDB: 7M05).[31] (C) Further SAR exploration of **88** results in a PBM-competitive inhibitor **31** (BRD0639). The FP is a competition fluorescence polarization assay.

optimised. And the electrophilic reactivity of the compounds towards glutathione was used as an evaluation index of the reactivity of the compounds. As a result, the monochlorine-substituted compound **97** has been found to exhibit an optimal balance. From this, **31** (BRD0639) was obtained.[31] (Fig. 14C).

In 2022, Krzyzanowski and colleagues developed another type of PRMT5–adaptor protein interaction inhibitor (PAPII). The research group initially conducted a screening and identification process, whereby the RioK1-derived PBM sequence was identified as a promising candidate. It is well-known that the short amino acid chains are more easily hydrolysed by the proteolytic enzymes than those of the long chains in the physiological state. Thus, the peptide stapling technique is employed to improve the stability of the short chain. According to the co-crystal structure of RioK1-derived PBM bound to the PBM groove, Gly14 and Asp17 on the RioK1-derived PBM sequence (Fig. 15) are identified as the most ideal tether position for structural optimisation. And then these two residues may be linked through a suitable flexible chain to form

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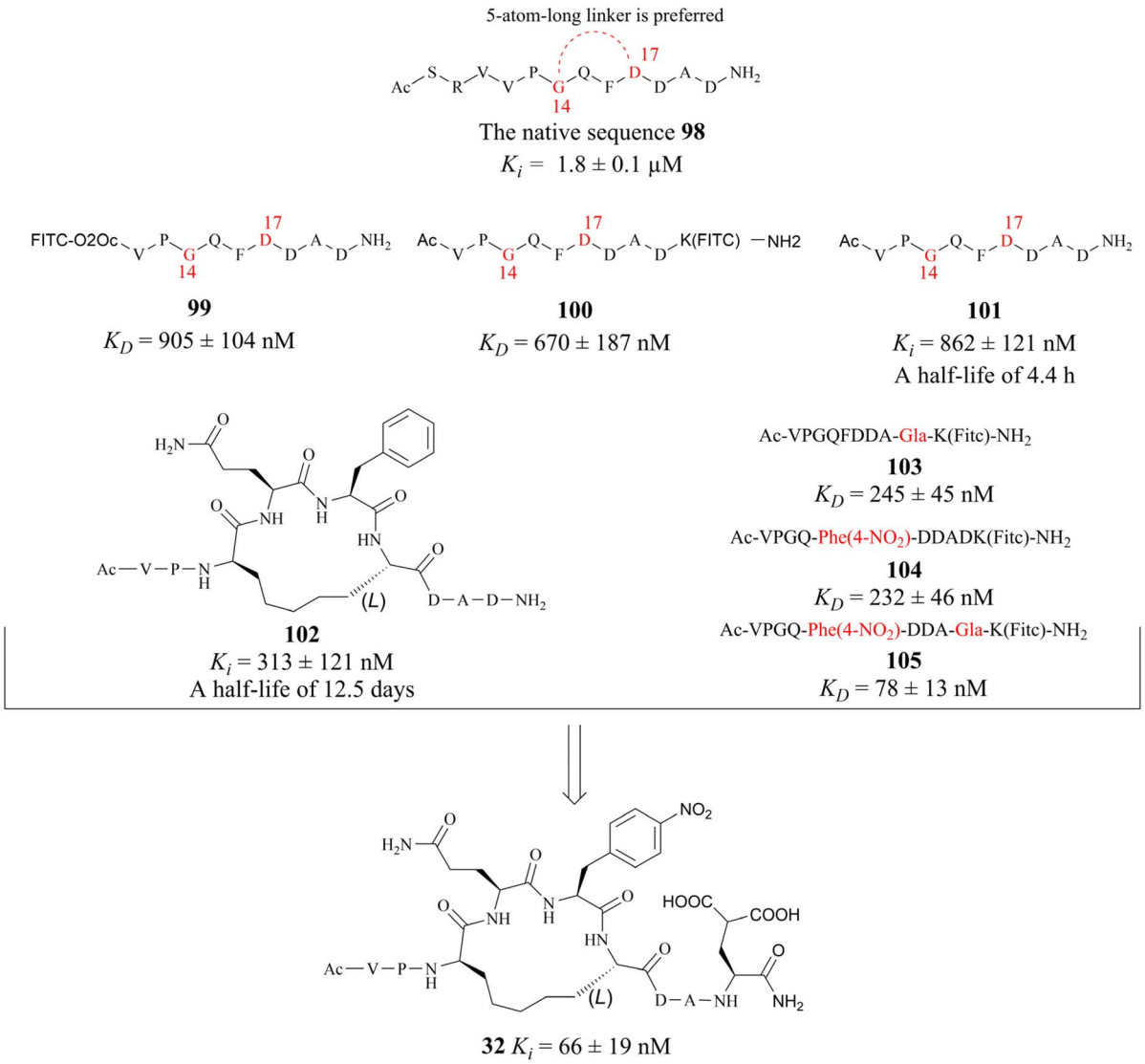

Fig. 15 Macrocyclization of the RioK1-derived PBM sequence results in a PAPII **32**.

a macrocycle, which would contain the key residues Gln15 and Phe16. This could maintain the spatial conformation of the short-peptide and effectively improve its enzymatic stability. Further studies have shown that this N-terminal SRV motif is not necessary for the affinity and that the N-terminal VP motif is essential for maintaining affinity (such as compounds **99–101**). Owing to a potential steric effect of the attached group, the N-terminal fluorescein isothiocyanate (FITC) label (such as compound **99**) may result in a reduced apparent protein binding affinity. The C-terminal FITC label (compound **100**) does not interfere with binding to the PBM groove. Cyclopeptides with 5-atom-long linker are preferred such as compound **102**. Then a 4-atom-long linker with a double bond is also tolerated, but only one of the isomers of the unsaturated linker will show a strong affinity for the target (the structure is not shown here). Replacing Asp20 with Gla and using Phe(4-NO_2) instead of Phe16 are both beneficial in improving affinity (compounds **103–105**). The combination of these modifications in the compound resulted in the cyclopeptide **32**. Cyclopeptide **32** (IC_{50}: 654 ± 476 nM) shows a comparable level of potency to that of the PBM-competitive inhibitor **31** (BRD0639) (IC_{50}: 568 ± 284 nM) in a competitive assay. Moreover, cyclopeptide **32** does not interfere with the interaction between PRMT5 and MEP50. However, the physicochemical properties of cyclopeptide **32** reveal that its cell permeability may be poor and further optimisation is required.[30,32] (Fig. 15).

In addition, the Hu group disclosed a series of novel PRMT5:MEP50 PPI inhibitors in 2022, represented by compounds **106** and **33** (see Fig. 16). Although both **106** and **33** have almost identical chemical structures, differing only in a substituent group on the isoxazole ring (shown in red in Fig. 16), the potency of compound **33** (IC_{50}: 430 nM) is approximately 4-times greater than that of **106** in the LNCaP cell growth inhibition assay. Nevertheless, these inhibitors, which are intended to be used as anti-cancer drugs in clinical trials, will still face significant challenges. For example, the **33** may

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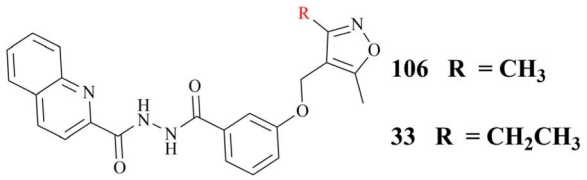

Fig. 16 Structures of compounds **106** and **33**.

have low solubility and its hydrazide linker does not appear to be beneficial for toxicity and stabilisation *in vivo*. Therefore, more work may need to be done to optimise this. Interestingly, the docking model of **106** into the TIM barrel shows that its isoxazole moiety is solvent-exposed, suggesting that it may serve as a suitable handle for structural modification.[25]

3 Other PRMT5 inhibitors based on the allosteric, covalent, and PROTAC modes of action

3.1 Allosteric inhibitors

In 2020, a compound **108** (R-1a) (it was originally used as the BACE inhibitor) was screened and identified by Palte *et al.* as a potent allosteric PRMT5 inhibitor (EC_{50}: **108** (R-1a) = 16 nM).[19,20] In the co-crystal structure (Fig. 17B, PDB: 6UXX), the adamantane moiety of **108** (R-1a) occupies a hydrophobic cage formed by five amino acid residues (Leu436, Leu437, Phe519, Phe555, and Tyr468). The iminohydantoin core forms two hydrogen bonds with Glu444. The methoxyphenyl group is surrounded by aromatic residues Phe519, Phe584, and Phe602, and makes an edge-to-face stacking interaction with Tyr613. Moreover, this group may occupy the position of Phe440 residue, thereby forcing it to move towards the SAM binding pocket. This unique binding mode deeply impacts both the SAM-binding site pocket and substrate-binding site so that none of the SAM and substrate can bind with PRMT5 in their respective binding sites. Surprisingly, the co-crystal structure of the PRMT5/MEP50 complex with **108** (PDB: 6UXX) shows that there is a narrow channel between the allosteric binding site and the substrate binding site, which appears to provide an opportunity for further structural optimisation.[19] In addition, the use of other hydrophobic groups (such as a propylcyclohexane group) instead of the adamantyl moiety seems to be ok, but further screening studies on hydrophobic groups are necessary.

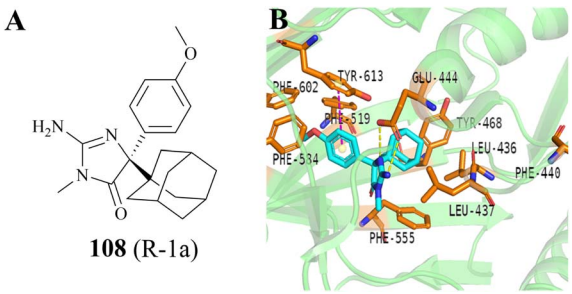

Fig. 17 (A) The structure of **108**. (B) The co-crystal structure of the PRMT5/MEP50 complex with **108** (PDB: 6UXX).[19]

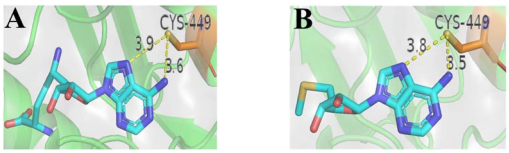

Fig. 18 (A) The space length between C449 and C_6–NH_2 (and N_7 atom) of the adenine ring of **SAM** is 3.9 (and 3.6) Å (PDB: 4GQB).[25] (B) The space length between C449 and C_6–NH_2 (and N_7 atom) of the adenine ring of **MTA** is 3.8 (and 3.5) Å (PDB: 5FA5).[46] Note: the standard length of a carbon–carbon bond is approximately 1.54 Å.[133]

3.2 Covalent inhibitors

In general, the binding of SAM competitive inhibitors to PRMT5 occurs by non-covalent means;[33,49,129,130] However, the potency of these reversible SAM competitive inhibitors is likely to be affected by SAM levels in tumour cells.[131,132] Interestingly, in the SAM binding pocket, there is a unique non-catalytic cysteine (C449), whereas other PRMTs contain a serine residue at the same position. The co-crystal structure shows that the space length between C449 and C_6–NH_2 (and N_7 atom) of the adenine ring of MTA (or SAM) could accommodate a covalent warhead (see Fig. 18). Moreover, C449 exists in part as a thiolate ion under physiological conditions. These findings provide a foundation for the development of irreversible SAM competitive inhibitors. Based on this information, two acrylamide derivatives of MTA (see Fig. 19A compounds **109** and **110**) were designed and synthesized in 2017; However, they exhibited a poor inhibitory activity (IC_{50}: 5.01 μM and 2.38 μM, respectively) in a biochemical methylation assay.[134] And then, Luengo *et al.* developed another series of PRMT5 covalent inhibitors using **LLY-283** as the parent compound in 2019 (compounds **111–115** see Fig. 19A)[34] Both compounds **112** and **113** show valuable enzymatic inhibitory potency (comparable IC_{50}: 11 nM and 19.5 nM, respectively) in the PRMT5/MEP50 biochemical assay. Compound **112** is initially converted to **113** under the assay conditions (which closely resemble physiological conditions, with a pH of 8.0). Subsequently, a plausible trans double bond (see Fig. 19B) is formed between the S atom of Cys449 and the adenine ring C_6–NH_2; However, the covalent bond between Cys449 and the adenine ring of compounds **111**, **114**, and **115** has not been formed. This is probably due to the fact that these compounds lack the chemical reactivity with Cys449 under assay (pH: 8.0) conditions or the corresponding aldehyde does not match the active site or the potential affinity of these compounds for the SAM binding pocket is reduced. Additionally, the co-crystal structure of PRMT5/MEP50 complex with compound **113** (PDB: 6K1S) shows that, apart from the plausible trans double bond interaction, all other key binding interactions between **113** and PRMT5/MEP50 complex are similar to those of the parent compound **11** (LLY-283) (PDB: 6CKC). It should be noted that the aldehyde is very easy to be metabolized

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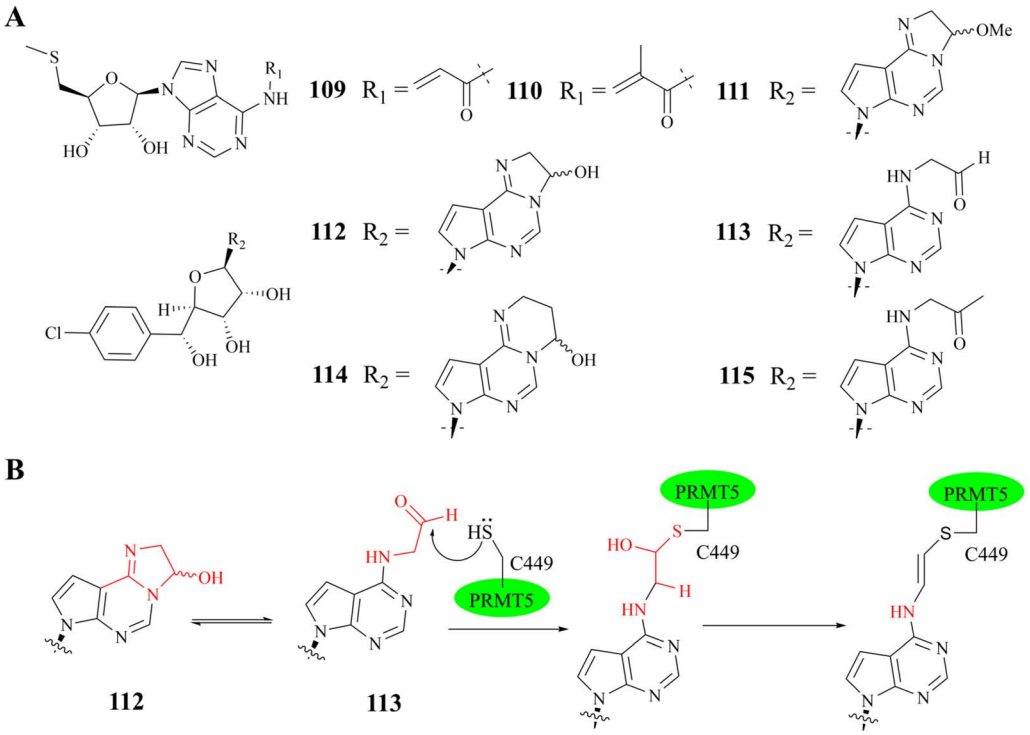

Fig. 19 (A) PRMT5 covalent inhibitors. (B) Proposed mechanism of vinyl-thio ether formation.

by enzymes *in vivo*, and further structural optimisation will be necessary.[34]

3.3 The PRMT5 degraders based on PROTACs

Proteolysis targeting chimeras (PROTACs) have the ability to degrade a specific target protein by hijacking the E3 ubiquitin ligase machinery of the ubiquitin-proteasome system (UPS).[135,136] In 2020, a selective substrate-competitive inhibitor EPZ015666 was selected as the target protein of interest (POI) ligand because Shen, *et al.* observed that the oxetane moiety of EPZ015666 is solvent-exposed in the co-crystal structure (PDB: 4X61 Fig. 20B). Subsequently, the first PRMT5 degrader **123** (MS4322) and two control compounds **122** (MS4370) and **121** (MS4369) were designed and prepared.[64] As shown in Fig. 20C, **123** (MS4322) is composed of three parts: EPZ015666, (*S*, *R*, *S*)-AHPC-Me (VHL-2), and the linker. A preliminary study of the SAR indicates that the PRMT5 degraders with a relatively longer polyethylene glycol (PEG) linker have possessed stronger potency than other PRMT5 degraders which contain a short PEG linker or an all-carbon linker. For example, the PRMT5 protein level in a multitude of cell lines is clearly reduced by compound **120** at 1–5 μM (with greater efficacy observed at 5 μM), whereas none of compounds **116–119** seem to exhibit the efficacy of PRMT5 protein degradation at this concentration range. And although **123** (MS4322), **122** (MS4370), **121** (MS4369), and compound **116** share a common PEG linker, their efficacy is obviously distinct; and when is administered to MCF-7 cell lines, PRMT5 levels are significantly reduced in a time-, dose-, VHL-, and proteasome-dependent manner ($DC_{50} = 1.1 \pm 0.6$ μM and $D_{max} = 74 \pm 10\%$); in contrast, EPZ015666 (alone), **122** (MS4370) which contains the diastereoisomer of **VHL-2**, and **121** (MS4369) could not reduce PRMT5 levels in MCF7 cells at 5 μM. Compound **116** displays a weaker PRMT5 degradation activity due to the fact that **VHL-2** is more effective than **VHL-1** at the degrading of protein; Furthermore, **121** (MS4369) exhibits poor PRMT5 inhibition activity compared to EPZ015666 (IC_{50}: 30 ± 3 nM), and it is probably because the 1, 2, 3, 4-tetrahydroisoquinoline moiety plays an important role in maintaining and enhancing the PRMT5 substrate-competitive inhibitors activity. These results suggest that the efficacy of the degrader **123** (MS4322) may be influenced by multiple factors, including the POI ligand, linker, and the E3 ligase ligand.[64]

4 The potential nitrosamine risk factors associated with the preparation process

Since the initial detection of *N*-nitrosodimethylamine (NDMA) in valsartan in 2018, the quality control and regulation of genotoxic impurities in the *N*-nitrosamine (NA) class has become a pivotal concern for pharmaceutical manufacturers and regulatory bodies. This is because they were subsequently identified in the active pharmaceutical ingredients (APIs) of other drugs, including sartan, pioglitazone, ranitidine and metformin. In consequence, drugs such as varenicline, propranolol, quinupril and orphenadrine were also recalled due to the presence of the respective nitrosamine drug-substance-related impurity (NDSRI). It is well established that the properties of amines,

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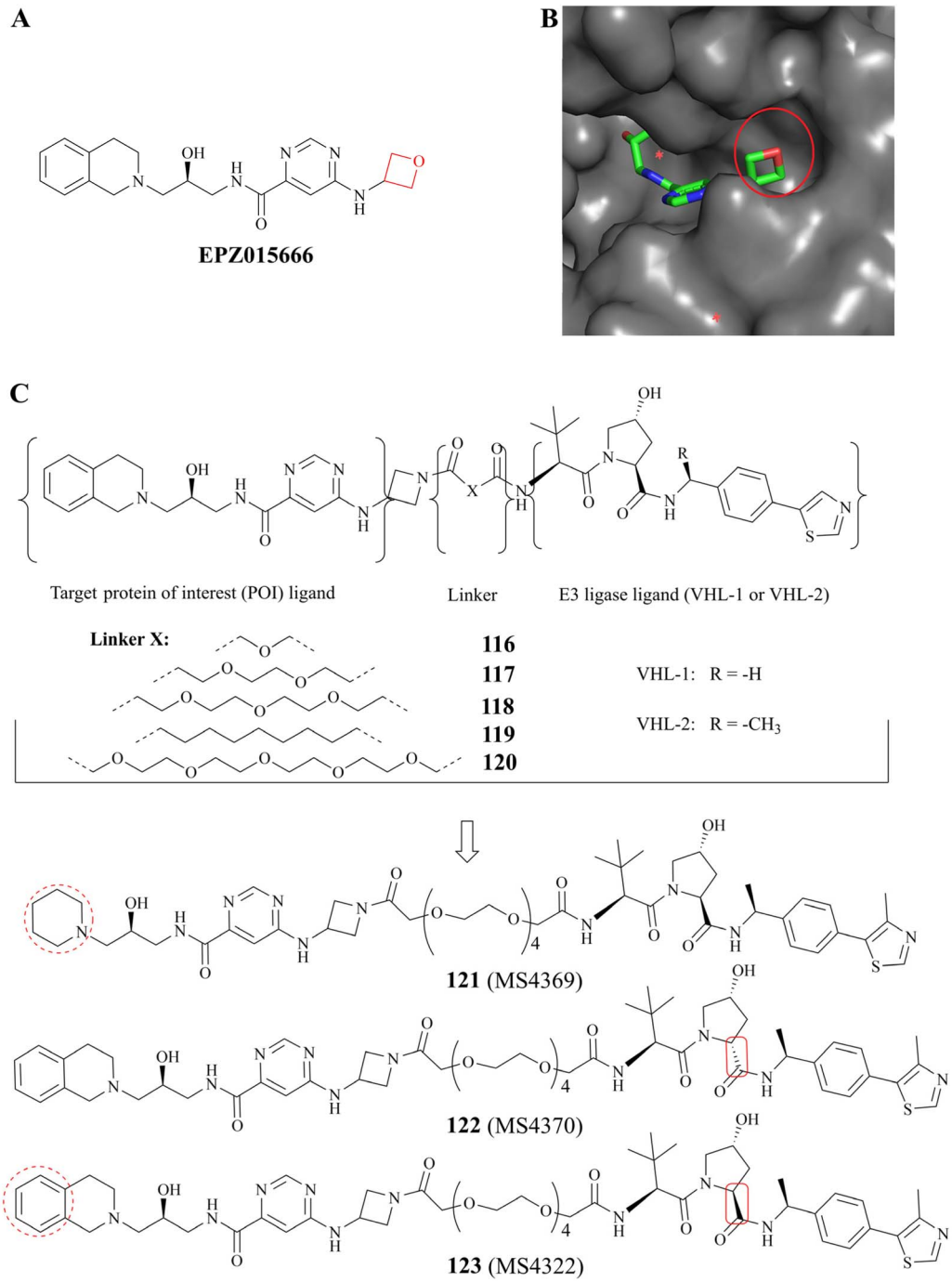

Fig. 20 (A) EPZ015666 (the oxetane moiety is shown in red); (B) the co-crystal structure of PRMT5/MEP50 complex with EPZ015666 (PDB: 4X61). It can be observed that the oxetane moiety of EPZ015666 is solvent-exposed. (C) A brief study of the SAR of **123** (MS4322).

particularly those of secondary amines, are frequently necessary for effective binding to protein biological targets. Furthermore, amines typically exhibit a higher volume of distribution *in vivo*, which facilitates the enhancement of numerous pharmacokinetic parameters. However, recent studies have demonstrated that approximately 40% of active pharmaceutical ingredients (APIs) and 30% of API impurities are potential precursors of nitrosamine (NA) formation due to the presence of susceptible amine groups. If only the more reactive secondary amines are considered, there is still a potential risk for 13–15% of APIs.[137–140] It is noteworthy that even in solid drug products (DPs), nitrites can form NDSRIs from levels of 1 part per million due to factors such as storage environment. Furthermore, the default Acceptable Intake (AI) for *N*-nitrosamines (NAs) is only 18 ng per day.

The origin of nitrosamines in pharmaceuticals is generally considered to be multifactorial. Primarily, the simultaneous use of secondary amines and nitrite reagents in the synthesis routes

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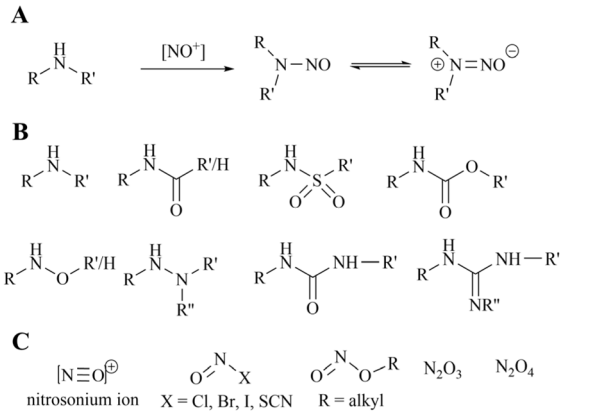

Fig. 21 (A) Mechanism of formation of nitrosamines from secondary amines, (B) the various secondary amine structures that have the potential to form nitrosamines. (C) Common nitrosating reagents.

of active pharmaceutical ingredients (APIs) (see Fig. 21) can be necessary for the formation of nitrosamines in the general assessment process. Secondly, the source of nitrosamines in pharmaceuticals can also be attributed to the use of pharmaceutical excipients or packaging materials. The formation of nitrosamines is less likely to occur with the use of tertiary amines due to the additional dealkylation step involved. In standard conditions, secondary amines are 1000 times more reactive than tertiary amines. Additionally, the generation of different cross-nitrosamine compounds may be influenced by the presence of different alkyl substituents. Furthermore, compared to secondary and tertiary amines, primary amines are challenging to produce stable nitrosamines. Typically, primary amines undergo diazotisation and subsequent amine removal to achieve this stability.[141–143] Therefore, primary amines are the least likely to result in the formation of nitrosamines. Consequently, this study focuses on secondary amines in the synthesis process.

4.1 MRTX1719

The early preparative route to **27** (MRTX1719) focused on the cross-coupling of two compounds (**124** and **125**) to give racemic **126**. Subsequently, the pure enantiomer **27** (MRTX1719) was obtained through a separation and deprotection process utilising chiral preparative chromatography.[87] However, as the clinical trials progressed, the chiral preparative chromatographic separation to obtain the highly pure (*M*)-enantiomers was no longer adequate for the split. In 2023, Michal Achmatowicz *et al.* reported an economically viable method for the production of high-purity (*M*)-isomers on a multi-kilogram scale. Firstly, the conventional chiral resolution method, which utilises Boc-*d*-phenylalanine (BDP), has replaced the enantiomeric chromatographic separation that was previously required by the original route. The efficiency of the chiral resolution process was significantly enhanced, enabling the conversion of the racemate to 98.4% d.e. MRTX1719-BDP salt **127** ((*M*)-4-BDP) in 75% separation yield on a current multi-kilogram scale.[116] The preparation is shown in Fig. 22.

Nucleophilic reactions between compounds containing secondary amines and nitrosating reagents have the potential to

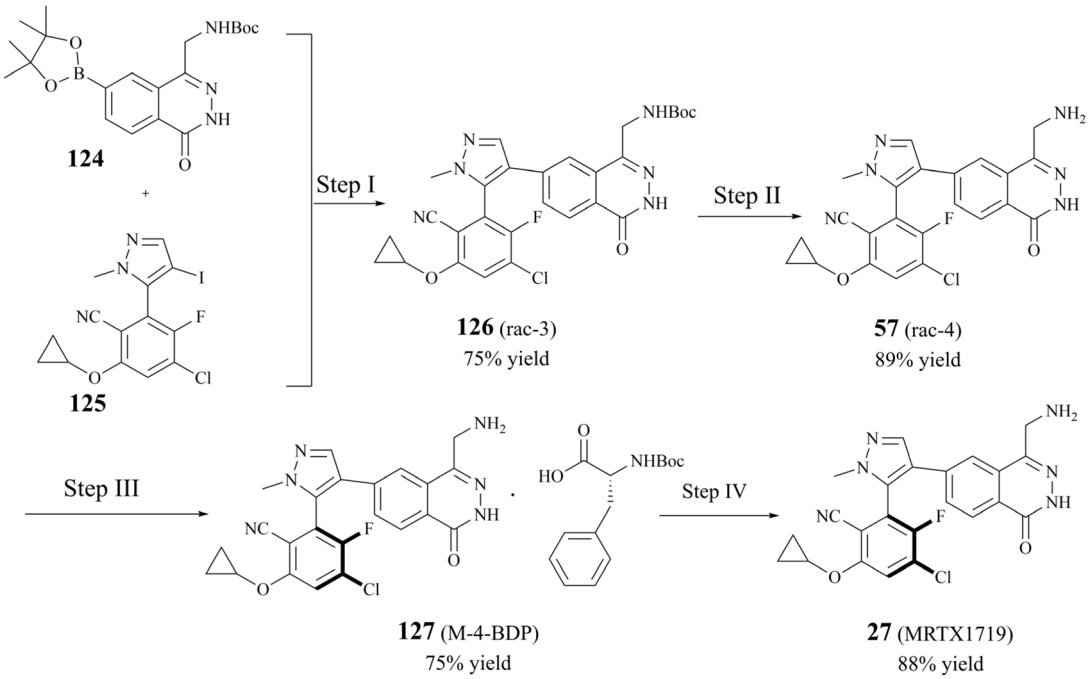

Fig. 22 Current multikilogram GMP synthesis of **27** (MRTX1719). (Step I) 3.0 mol% Ad_2nBuP-Pd-G3,3.0 eq., Cs_2CO_3, 12 vol. toluene, 4 vol. H_2O, 55–60 °C, 18 h. (Step II) 4 M HCl/EtOAc (5 vol.), 20 vol. MeOH, 15–25 °C, 18 h; 7 M NH_3/MeOH (1 vol.),10 vol. MeOH, 10–15 °C, 20 h. (Step III) 1.2 eq. Boc-*D*-Phe, 15 vol. EtOH/H_2O 98 : 2, crystallizer: 20–25 °C, 16 h, racemizer: 160 °C, t_R 2–4 min; 7 vol. EtOH/H_2O 85 : 15, 15–25 °C, 16 h. (Step IV) 4.5 vol. THF, 1.5 vol. H_2O, 20–25 °C; 39 vol. H_2O, 1 vol. conc. aq. NH_3, 20–25 °C, 6 h; 9 vol. H_2O, 1 vol. IPA, 20–25 °C, 6 h.

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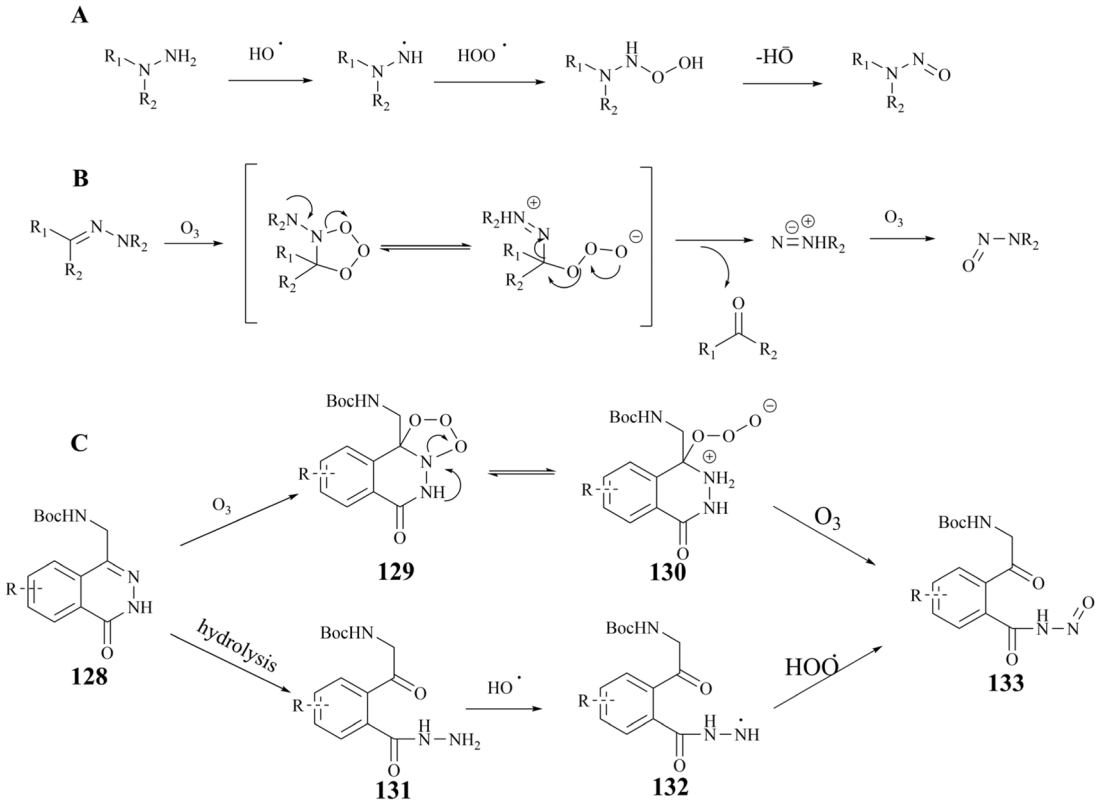

Fig. 23 (A) The reaction mechanism of the oxidation of hydrazine. (B) The reaction mechanism of the ozonolysis of hydrazine (C) compound **133** possible production routes.

produce nitrosamine impurities, as illustrated in Fig. 21. It is noteworthy that the secondary amines referred to herein encompass a range of compounds, including fatty secondary amines, amides, carbamates, hydroxylamines, hydrazines, hydrazones, ureas, and guanidines. In Fig. 22, there seems to be a structural rationale for the formation of nitrosamine-like impurities such as the presence of several intermediates with Boc-protected amide structures, including compound **124**, **126** (rac 3), and **BDP**. Compound **124**, which can be hydrolysed to boric acid under acidic conditions. Alkyl nitrites have been reported to generate nitrosamines from arylboronic acids in the presence of a copper catalyst, such as the Chan–Lam reaction. However, we carefully reviewed the conditions of the preparative process for step I–IV in the route, and did not find any obvious nitrosation reagents, and the Boc-protected amide structures exhibited a generally relatively low reactivity. It is worth noting that nitrosamines can be produced by two specific pathways, the oxidation of hydrazine and the ozonolysis of hydrazone.[143] Some studies have shown that hydrazine can be oxidised in the presence of air or metal catalysts to form nitrosamines (the reaction mechanism is shown in Fig. 23A. In the presence of ozone, hydrazones can be oxidised to form nitrosamines and the reaction mechanism is shown in Fig. 23B.

4.2 AMG193

As previously stated, Amgen Inc. has patented a series of inhibitors targeting the PRMT5/MTA complex. Possibly based on synthetic convenience considerations, all of the inhibitors in this series have an aromatic pharmacodynamic core connected to another molecular group *via* an amide bond. Thus, the patented compounds in this series have similar preparation routes. We would like to briefly summarise them here, taking the preparation route of AMG193 as a representative. As shown in Fig. 24, the pharmacophore 4-amino-1,3-dihydrofuro [3,4-*c*][1,7]naphthyridine-8-carboxylic acid (Compound **136**) and (*S*)-3-(5-(trifluoromethyl)pyridin-2-yl)morpholine (compound **138**) *via* a condensation reaction to produce **30** (AMG193) in the presence of the peptide condensation reagent 2-(7-azabenzotriazol-1-yl)-*N*,*N*,*N*′,*N*′-tetramethyluronium hexafluoro phosphate (HATU). Compound **136** was synthesised using methyl 5-amino-4-bromo-2-pyridinecarboxylate (**133**) as the initial starting material in a three-step process involving Suzuki coupling reaction, cyclisation and hydrolysis. The production of compound **138** was accomplished through a chiral splitting approach utilising the SFC method. Notably, this same chiral splitting strategy can also be employed after the synthesis of **30** (AMG193) racemates.[124–127]

In additional, the morpholine moiety in compound **137** and **138** represents a cyclic secondary amine warning structure. However, of greater significance is the peptide condenser HATU. In the presence of HATU, the carboxylic acid anions within the condensation reaction system initially attack HATU, resulting in the generation of an unstable *O*-acyl (tetramethyl) isourea salt intermediate through an addition–elimination

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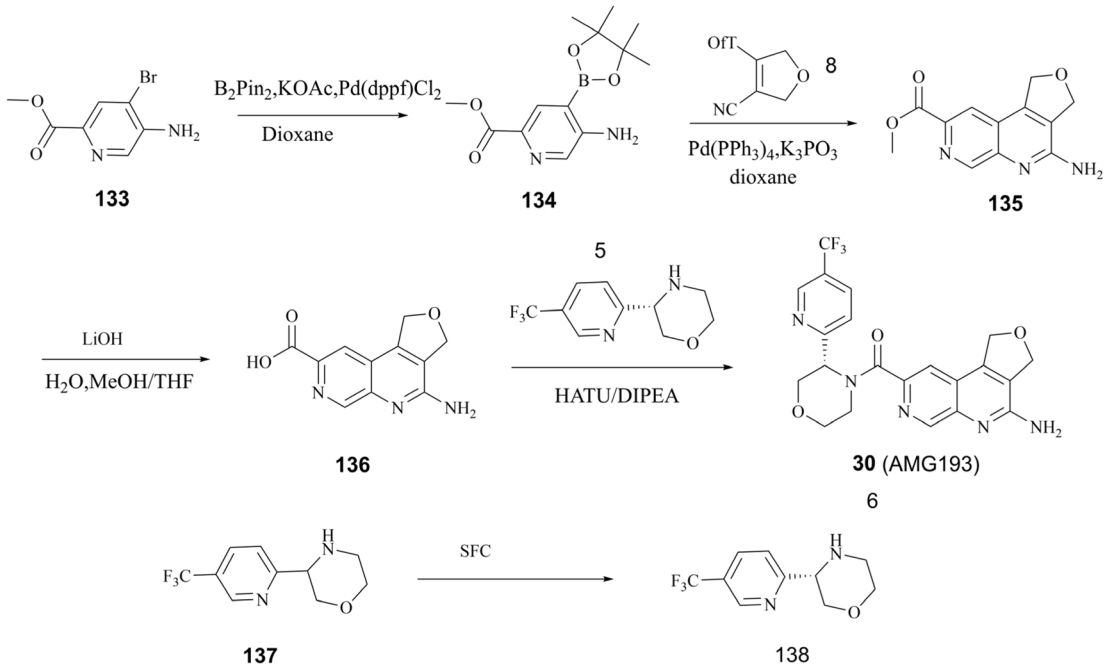

Fig. 24 Preparation of **30** (AMG193).

process. Subsequently, the OAt negative ion attacks the isourea salt, resulting in the formation of the HOAt active ester and tetramethylurea. The amine and HOAt reactive ester then undergo a reaction, forming the amide and HOAt. Therefore, tetramethylurea has the potential to serve as a source of dimethylamino group (see Fig. 25).

4.3 TNG908

The process route for the preparation of **28** (TNG908), as illustrated in Fig. 26, comprises two principal sections. In one phase of the process, pyridin-2(1*H*)-one (compound **139**) was used as the initial reactant, which was reduced by 10% Pd/C in order to obtain racemic compound **140**. Subsequently, compound **140** was protected by Boc, and in the presence of 1,1,1-trifluoro-*N*-phenyl-*N*(trifluoromethylsulfonyl)methanesulfonamide (1.25 eq., compound A), the triflate compound **141** was generated at low temperature (between −78 and −25 °C). The compound **143** was prepared by Suzuki coupling of boronates or boronic acids with Boc-protected triflate **142**. Subsequently, the protection group of compound **143** was removed by treatment with trifluoroacetic acid (TFA), compound **145** is gain by a further reduction of the double bond. The other part was the preparation of compound 12 using 3-methyl-5-nitropyridine-2-amine (compound **146**) as starting material. The condensation of compounds **145** and **150** was conducted in the presence of the peptide condensation reagent HATU. The Boc protecting group was then removed, and **28** (TNG908) was obtained by chiral splitting using SFC or chiral HPLC. Given that **29** (TNG462) and other derivatives possess the same fundamental backbone structure as **28** (TNG908), they can be prepared in a similar manner, and the associated details are therefore not repeated herein.[85,86]

From the perspective of compound preparation, the process route for **28** (TNG908) preparation in the initial exploratory phase of SAR incorporates the preparation of **28** (TNG908) and its derivatives, which is a highly practical process preparation route. Nevertheless, more rigorous quality control is necessary for clinical applications and GMP production environments. Firstly, with regard to the potential for the formation of nitrosamine impurities, triethylamine (TEA) and dimethylformamide (DMF) are frequently employed as reagents in the synthesis process depicted in Fig. 26. It is generally accepted

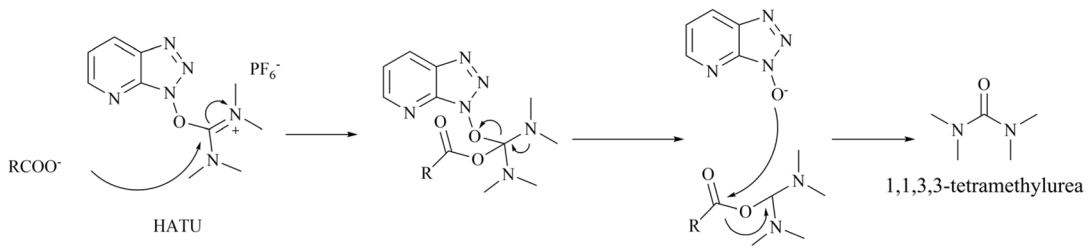

Fig. 25 Reaction mechanism of HATU to generate 1,1,3,3-tetramethylurea.[144]

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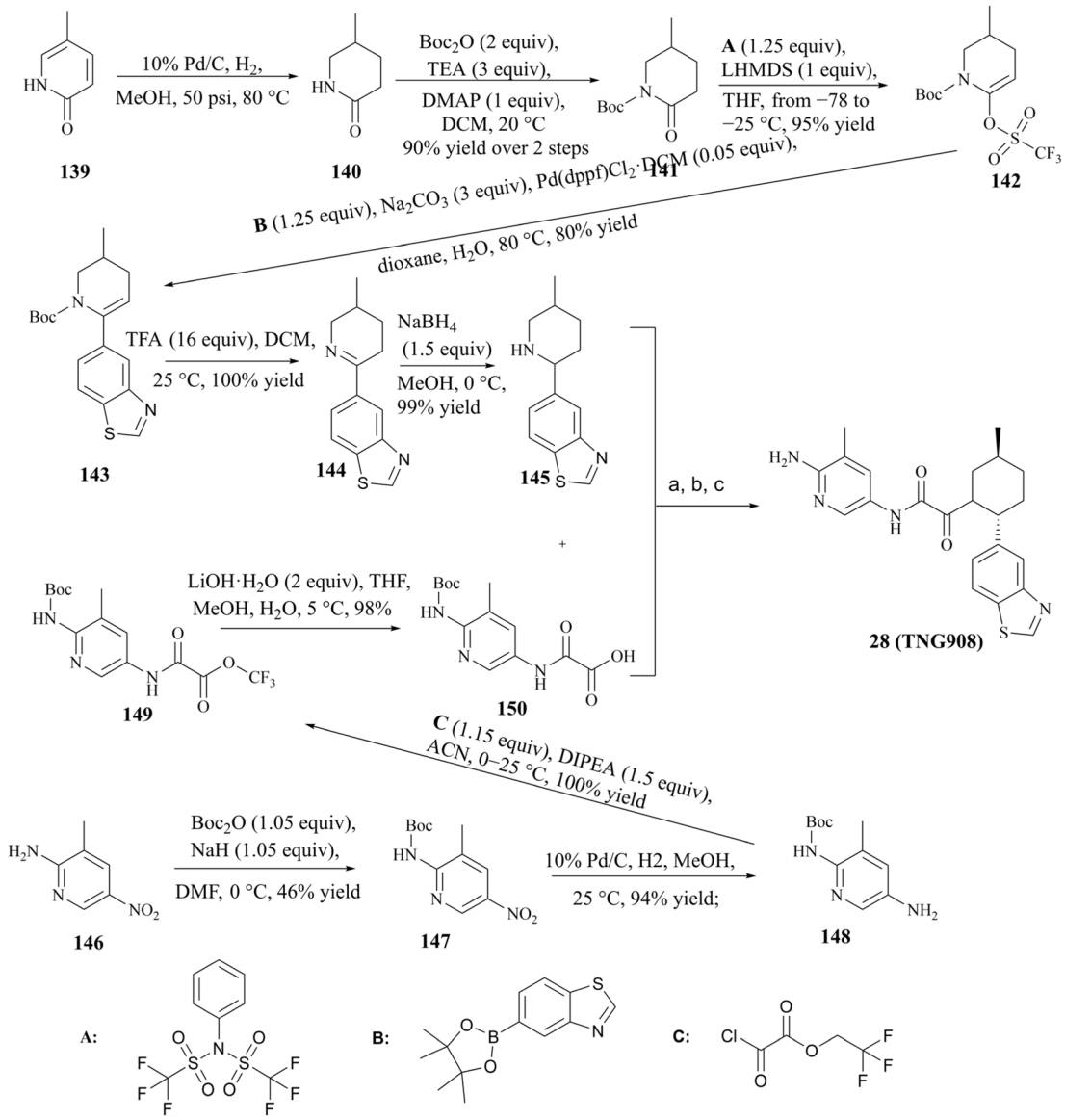

Fig. 26 28 (TNG908) preparation route. (a) HATU (1 equiv.), TEA (6 equiv.), DMF, 25 °C; (b) 4 M HCl in dioxane, 25 °C, 59% yield over 2 steps for **28** (TNG908); (c) chiral HPLC separation of enantiomers, 48% yield for **28** (TNG908).

that these two reagents are the source of additional secondary amines in the synthesis process. TEA may contain diethylamine, while DMF may contain dimethylamine. It is hypothesised that 1,1,1-trifluoro-*N*-phenyl-*N*(trifluoromethylsulfonyl) methanesulfonamide (compound A) may be a potential source of sulfonamide-type secondary amines. Furthermore, intermediates **140**, **144**, and **145** are cyclic secondary amine alert structures. In the preparative process, the nitro group of intermediate **147** was reduced by hydrogenation with 10% Pd/C, resulting in the production of an amine. This reaction is characterised by low impurities and high yield. However, the reduction reaction does in fact undergo an intermediate process of nitroso compounds, namely *N*-hydroxyhydroxylamine (see Fig. 27), which could potentially act as a source of nitroso groups if the process is not operated correctly or the reaction is not monitored with sufficient accuracy. Furthermore, nitroso compounds and *N*-hydroxyhydroxylamine can undergo a reaction to form oxoazo compounds, which can then, upon further reduction, yield intermediates such as azoic compound, 1,2-dihydrocarbonyldihydrazine, and so on. These intermediates require longer hydrogenation-catalysed reductions to be fully reduced to amine groups.[145]

It should be noted that the nitrosamine risk factors discussed above are based on the chemical structures described in the current literature, which indicate the potential for the generation of nitrosamine-like impurities under certain conditions. These chemical structures must meet sufficient conditions, such as the introduction of nitrosating compounds or potentially nitrosating reagents in the process route due to unknown factors, to generate possible nitrosamine impurities.

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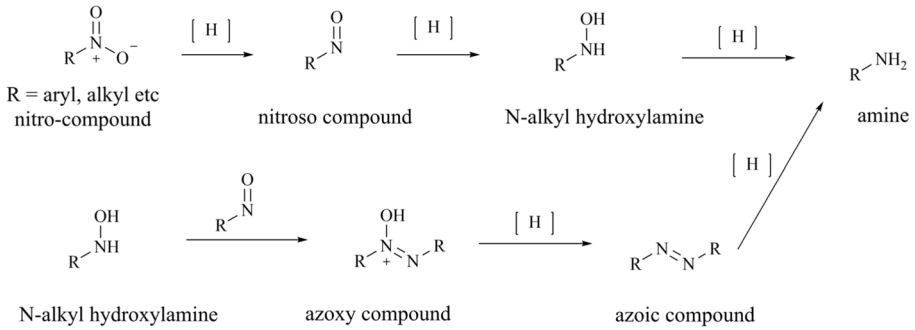

Fig. 27 Nitrohydrogenation reduction mechanism.

While various types of secondary amine structures are generated in our current process routes for PRMT5/MTA inhibitor preparation, in all mature processes, there are no significant nitrosation conditions. However, rigorous process operation and quality control standards remain necessary considerations based on safety concerns.

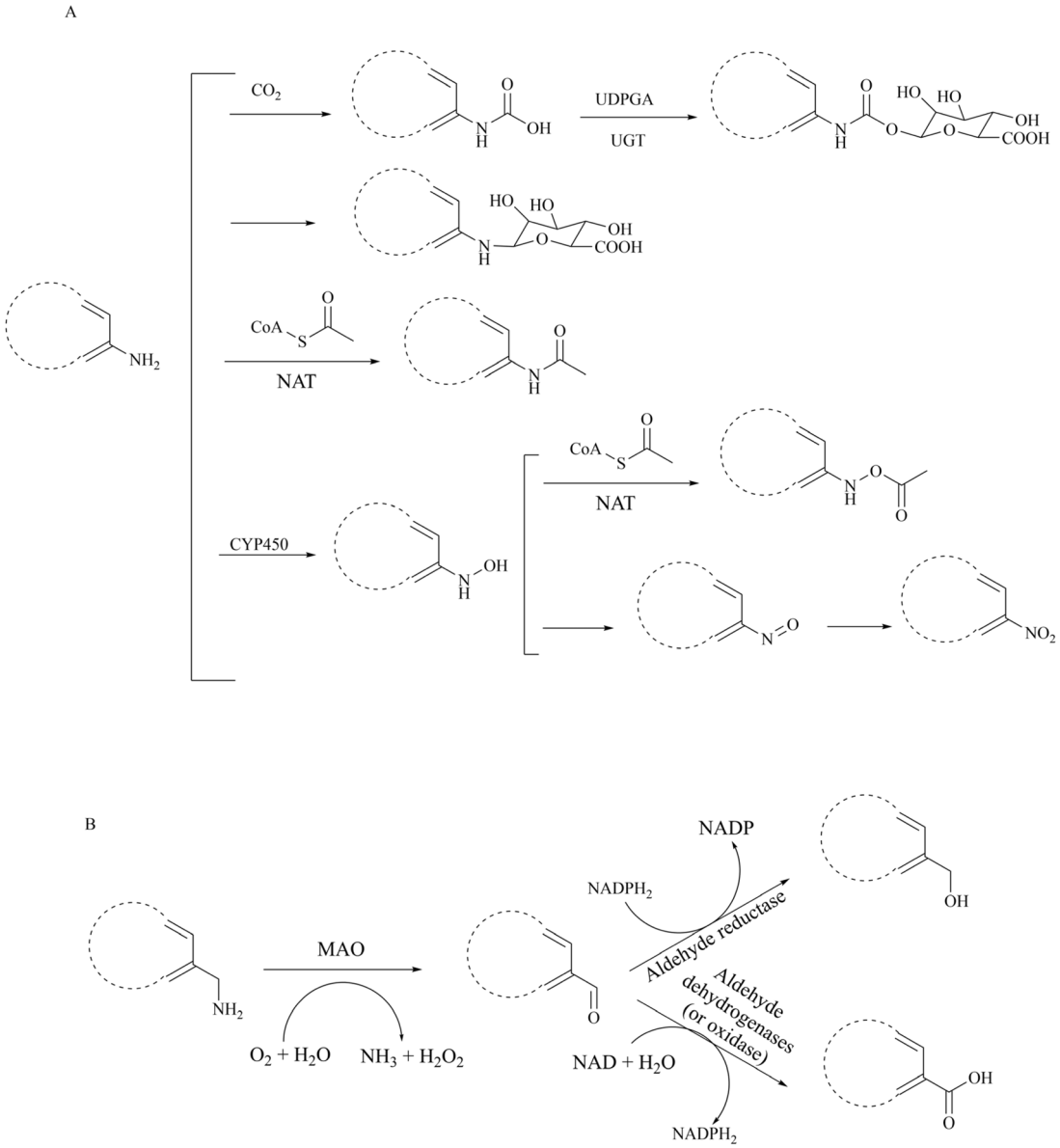

Fig. 28 Major metabolic pathways of primary amines *in vivo*. (A) Major metabolic pathways of aryl amines *in vivo*.[148–152] (B) Major metabolic pathways of benzyl amines *in vivo*. The vulnerability of arylamine *in vivo*.[153]

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5 The vulnerability of arylamine *in vivo*

As previously stated, all PRMT5/MTA complex inhibitors possess a vital pharmacophoric functional group, namely the primary amine moiety (see Fig. 9). The primary amine can form H-bond interactions with the key catalytic residues Glu444 or Glu435. Additionally, it forms a specific interaction with the MTA bound in the protein complex, which serves to stabilise the PRMT5/MTA complex.

Furthermore, the primary amine group is a basic group for the salt formation of these inhibitors, which facilitates the improvement of many pharmacokinetic parameters. These finds suggest that the primary amine moiety is of primary importance with regard to the efficacy, protein complex selectivity, and MTAP-deficient tumour cell selectivity of PRMT5/MTA complex inhibitors. However, the greater the importance, the greater the fragility. Small changes in primary amines inevitably result in changes in the potency and safety profile of PRMT5/MTA complex inhibitors. In the initial stages of drug discovery, this vulnerability posed significant organic synthesis challenges for structural extensions and the SAR exploration of fragments. Some promising fragments were ultimately excluded after a comprehensive evaluation.[101,146,147] *In vivo*, exogenous drugs containing amine groups are usually metabolised by the cytochrome P450 (CYP) family of enzymes. However, arylamines can also be catalyzed by NAT enzymes to produce acetylated metabolites or conjugated with glucuronic acid to produce water-soluble products. Generally, arylamines are metabolised by cytochrome P450 (CYP) enzymes without deamination, but rather to hydroxylated metabolites (see Fig. 28A). However, many of the hydroxylated metabolites formed by exogenous arylamines are carcinogenic, such as 2-amino-1-methyl-6-phenylimidazo[4,5-*b*]pyridine (PhIP).[148–152] On the basis of this information, it is reasonable to speculate that *in vivo*, arylamine groups may represent a significant metabolic reaction site for drugs bearing an arylamine moiety, and that these metabolic reactions are readily occurring. However, the arylamine moiety is of critical importance for the efficacy and safety of PRMT5/MTA complex inhibitors *in vivo*, and changes in its structure would lead to changes in its inhibitory potency and pharmacological mechanism of action, including cellular selectivity. Moreover, these inhibitors are PRMT5 substrate competitive. Hypothetically, these inhibitors such as **30** (AMG193) and **28** (TNG908) have preferential metabolism of primary amines over other metabolic sites, and there is a potential for an indirect dose-related toxicity in the case of long-term administration. It is noteworthy that the primary amine of **27** (MRTX1719) is a benzylamine group, which may offer an alternative metabolic possibility (*e.g.*, Fig. 28B). Benzylamine molecules can be catalytically deaminated by MAO to produce aromatic aldehydes.[153] However, aromatic formaldehyde is reactive *in vivo* and is generally further oxidised or reduced to the corresponding aromatic formic acid or aromatic methanol. These aromatic formic acids or aromatic methanols are generally more water-soluble than their parent drug. This may help to explain the lack of dose-limiting toxicity observed *in vivo* for MRTX1719 at a dose level of 400 mg q.d. However, it must be acknowledged that a clear picture of the dose-limiting toxicity of MRTX1719 *in vivo* remains elusive. Regrettably, no authoritative studies on these aspects could be identified in public databases.

6 Conclusions and perspectives

In this paper, we have conducted a review of the design thoughts and the structure–activity relationship (SAR) of known methylthioadenosine (MTA)-cooperative PRMT5 inhibitors. We expect to elucidate the chemical-structural basis of the protein- and cell-selectivity of these inhibitors through such a comprehensive review. Additionally, we have analysed the clinical safety of representative first- and second-generation PRMT5 inhibitors. In the binding pocket, first-generation inhibitors achieve high selectivity for PRMT5 by forming a potential π–π stacking or cation–π interaction with Phe327, a specific amino acid residue of PRMT5. And the molecular structure of SAM-competitive inhibitors commonly frequently comprises an aromatic ring, which facilitates the formation of π–π stacking interaction with Phe327. However, given the critical function of PRMT5 in human cells, it is expressed in almost all cells, especially in the haematopoietic system. *In vivo*, this selectivity for the enzyme does not reflect the high selectivity for tissue cells, resulting in a narrow therapeutic window. Consequently, these inhibitors typically exhibit dose-limiting toxicity in clinical trials. In contrast to the first-generation PRMT5 inhibitors, the second-generation inhibitors, which were developed based on a synthetic lethality strategy, exploit the elevated levels of MTA in MTAP-deficient tumour cells, thereby achieving enhanced cellular selectivity and representing a novel therapeutic option. In the clinical phase I trial, the second-generation inhibitors demonstrated higher tolerated doses, lower dose-limiting toxicity, and higher clinical value, particularly the PRMT5/MTA complex inhibitors. The PRMT5/MTA complex inhibitors occupy the PRMT5 substrate-binding pocket and form a specific interaction with MTA bound in the SAM pocket, thereby stabilising stabilises the PRMT5/MTA complex. Moreover, the substrate-binding pocket is predominantly composed of acidic amino acids, which are more prone to interaction with basic groups. The core pharmacophore of these inhibitors is typically a nitrogen-containing aromatic ring with a spatially directed substituted amine group. Owing to the binding mechanism of these small molecule inhibitors to the PRMT5/MTA complex, minor alterations to primary amines will inevitably result in modifications to the potency and safety profile of PRMT5/MTA complex inhibitors. Consequently, the vulnerability of its aromatic amine moiety is discussed. It can be reasonably assumed that the aromatic amine moiety is, *in vivo*, a readily metabolisable group. A further consideration is that PRMT5/MTA complex inhibitors are substrate competitive, which may prove to be a disadvantage to their use in clinical applications.

A number of second-generation inhibitors have been approved for clinical trials (Table 2 lists several representative second-generation inhibitors). Compared to the first-generation PRMT5 inhibitors, the majority of the second-generation inhibitors demonstrated satisfactory dose tolerance due to their rational pharmacological mechanism of action. For

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example, maximum tolerated dose of **27** (AMG193) is determined to be 1200 mg per day,[111] and no dose-limiting toxicity was observed in patients treated with **30** (MRTX1719) at a dose of 400 mg once daily.[103] The distinctive physicochemical characteristics of TNG908, which permit penetration of the blood–brain barrier, have facilitated promising advances in the treatment of gliomas. These results suggest that, following the precedent of poly ADP ribose polymerase (PARP), PRMT5 has the potential to become the next clinically applicable synthetic lethal target. In light of these issues, the primary challenges currently facing this field of research can be summarised as follows: firstly, it is essential to confirm the efficacy of these MTA-cooperative PRMT5 chemical inhibitors *in vivo* against a range of tumour types at doses that can be tolerated by patients. Secondly, it is crucial to identify any other biomarkers associated with these inhibitors. Thirdly, the mechanisms of resistance and metabolism of these inhibitors should be elucidated. It is notable that SCR6290 has been demonstrated to exhibit tumour cell selectivity through a tumour-biased distribution profile, characterised by an intracellular enrichment concentration that is 50-fold greater than that observed in the blood.[112,113] This may represent a promising avenue for reducing dose-limiting toxicity of PRMT5 inhibitors. However, the exact mechanism leading to the presence of a biased tumour distribution of this inhibitor *in vivo* has not been disclosed.

Following comprehensive research and discussion of the design strategies and SARs of various PRMT5 inhibitors, we found that the dose-limiting toxicity of this class of compounds that this may a common problem in the drug delivery field regarding selective and effective delivery of targeted inhibitors and the development of an optimal drug delivery system has the potential to significantly address this problem and enhance the efficacy. It is well known that, in general, tumour blood vessels are much more permeable than normal blood vessels. Furthermore, the compromised lymphatic blood flow pathways allow for the preferential accumulation of certain substances in tumour tissues, where they remain for extended periods, which is known as the enhanced permeability and retention (EPR) effect.[154] A nanodrug delivery strategy may be defined as the utilisation of either EPR effect-mediated passive targeting or ligand-mediated active targeting, with the objective of delivering nanodrugs to tumour tissues subsequent to systemic delivery.[155] This technology allows for the precise delivery of drugs into tumours, either by targeting tumour cells directly or by targeting APC (including macrophages and DCs). In 2020, Chenggang Zhu *et al.* reported the development of a novel injectable *in situ*-forming implantation system. This system consisted of *n*-butyl-2-cyanoacrylate (NBCA), ethyl oleate, and a sol–gel phase transition, and was used for the delivery of protein arginine methyltransferase 1 (PRMT1) inhibitor TC-E-5003.[156] In 2023, the team of Camino de Juan Romero reported the utilisation of small extracellular vesicles (EVs) isolated from diverse cell lines and encapsulated with temozolomide and EPZ015666 *via* direct incubation for the treatment of pancreatic cancer and glioblastoma (GBM).[157,158] Nanomedicines have been developed based on active targeting nanotechnology. These include ligand-implanted nanomedicines that facilitate specific tumour delivery and enhance therapeutic efficacy. These include trastuzumab (Tra) mediated targeting of NSCLC (HER-2 positive) and substance P peptide mediated targeting of gliomas. Furthermore, Jordan J. Green and colleagues devised a bioreducible nanoparticle for the systemic delivery of siRNA into patient-derived glioblastoma cells in an *in situ* mouse tumour model.[159] It is regrettable that, although the targeted delivery strategy may prove an effective means of enhancing the clinical value of PRMT5 inhibitors, research in this area is still in its infancy. No studies on targeted delivery systems applicable to PRMT5 inhibitors have been identified in the relevant journals or research communities.

Allostery is an inherent property of epigenetic enzymes in which topographically distinct binding sites are functionally coupled and transmissible.[160] Meanwhile, due to the binding site of allosteric inhibitors being located outside of the orthosteric sites, allosteric inhibitors always show non-competitiveness with endogenous cofactors and substrates in orthosteric sites. Thus, they possess remarkable potency and safety in lower dose levels compared to orthosteric inhibitors.[161] Interestingly, Palte *et al.* also noted that, although the cocrystal structure of the known allosteric inhibitor with PRMT5 shows that the eleven-amino acid loop (Glu435-Leu445) prevents the binding of SAM (PDB ID: 6UXY), the different conformations of this loop can be stabilized depending on the presence of allosteric inhibitors. And this suggests that the configurations in which both an allosterically binding molecule and SAM are bound seem possible.[19] So, is there another possible configuration in which both an allosteric binding molecule and MTA are bound?

In addition, the MTA-cooperative PRMT5 inhibitors provide an opportunity to study the MTA-cooperative PRMT5 degraders. In general, a suitable POI ligand from the vast array of highly potent and selective inhibitors can obviously enhance the selectivity of the degraders, which may exhibit more targeted degradation selectivity than the parent inhibitor, particularly against structurally highly homologous proteins.[136] A POI ligand (only EPZ015666 has been screened) is not sufficient to elucidate the profile of the PRMT5 degraders and we would like to see more extensive research in this area.

Abbreviations

AE	adverse events
ADMA	omega-NG, NG-asymmetric dimethylarginines
AIDD	artificial intelligence-based drug molecule virtual screening
APIs	active pharmaceutical ingredients
BDP	Boc-*d*-phenylalanine
Cl_{int}	human hepatocyte intrinsic clearance
COPR5	cooperator protein of PRMT5
DNMTs	DNA methyltransferases
DSF	scanning fluorometry
FBDD	fragment-based molecule library screening
FBLD	fragment-based lead discovery
FP	fluorescence polarisation
GBM	glioblastoma
GSH	glutathione

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HDACs	histone deacetylases
HMTs	histone methyltransferases
HTS	high-throughput screening
MEP50	methylosome protein 50
MET	methionine
MMA	omega-NG-monomethylarginines
MTA	methylthioadenosine
MTAP	methylthioadenosine phosphorylase gene
MST	microscale thermophoresis
NA	*N*-nitrosamine
NDSRI	nitrosamine drug-substance-related impurity
NDMA	*N*-nitrosodimethylamine
NMEs	new molecular entities
NMR	nuclear magnetic resonance
PARP	poly ADP ribose polymerase
PBM	PRMT5-binding motif
PPIs	protein–protein interactions
pICln	chloride channel nucleotide sensitive protein 1A
PRMTs	histone arginine methyltransferases
PTMs	post-translational modifications
PROTACs	proteolysis targeting chimeras
RioK1	Rio domain-containing protein
TKIs	tyrosine kinase covalent inhibitors
SAM	S-5-adenosyl-L-methionine
SAH	S-adenosyl homocysteine (SAH)
SAR	structure–activity relationship
SBDD	structure-based molecule design
SDMA	omega-NG, N'G-symmetric dimethylarginines
SFC	chiral supercritical fluid chromatography
SPR	surface plasmon resonance
TRAE	treatment-related adverse event
UPS	ubiquitin–proteasome system
VHL	Von Hippel-Lindau E3 ubiquitin ligase ligand
WD	tryptophan-aspartate
X-ray	X-ray protein crystallography

Data availability

No primary research results. Software or code have been included and no new data were generated or analysed as part of this review.

Author contributions

All authors have equal sharing in all preparation steps for the review in its current final form.

Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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