Nanoscale



COMMUNICATION

View Article Online
View Journal | View Issue



Cite this: Nanoscale, 2024, 16, 11538

Received 25th March 2024, Accepted 28th May 2024 DOI: 10.1039/d4nr01314j

rsc.li/nanoscale

Near-infrared AlEgens for sulfatase imaging in breast cancer in vivo†

Aggregation-induced emission luminogens (AlEgens) enable highly sensitive and *in situ* visualization of sulfatase to benefit the early diagnosis of breast cancer (BC), but current sulfatase AlEgens always emit visible light (<650 nm). Herein, a near-infrared (NIR) AlEgen QMT-SFA is developed for sulfatase imaging *in vivo*. Hydrophilic QMT-SFA is cleaved by sulfatase to yield hydrophobic QMT-OH, which subsequently aggregates into nanoparticles to turn the AlE fluorescence "on", enabling sensitive sulfatase imaging in 4T1 cells and mouse models.

Breast cancer (BC) is the most frequently diagnosed malignant tumor in women worldwide, with high morbidity and mortality rates.^{1,2} In clinical practice, mammography plays an essential role in BC screening, but it has limitations in the early diagnosis of BC due to its low sensitivity;³ tissue biopsy can help diagnose BC but is commonly used until late cancer stages.4 Therefore, it is of clinical importance to establish advanced methodologies for the early diagnosis of BC. To this end, some molecular or nanoprobes have been recently developed for sensitive and in situ detection of BC biomarkers (e.g., matrix metalloproteinase, cathepsin, and sulfatase) at the molecular level, which benefits the identification of BC at early stages.5 Among these biomarkers, sulfatase is a highly conserved sulfate-hydrolyzing protease which is overexpressed upon the occurrence of BC.6,7 Thereby, sensitive imaging probes of sulfatase are highly desired for the early diagnosis of

Among the various imaging modalities, fluorescence imaging shows unique advantages in biomarker detection, such as non-invasiveness and superior temporal and spatial resolution.8-11 In recent years, a range of fluorescence probes have been devised for the sensitive imaging of sulfatase activity.12 Of note, these fluorescence probes with aggregationinduced emission (AIE) characteristics (i.e., AIEgens) have attracted high interest due to their small autofluorescence interference, low toxicity, and strong resistance to photobleaching. 13-16 However, current sulfatase AIEgens always emit visible light (<650 nm), which gives rise to nonnegligible autofluorescence interference and inferior tissue penetration, thus severely hindering their further biological application and clinical translation.¹⁷ In this regard, an AIEgen with a near-infrared (NIR) emission, which affords a higher signal-to-background ratio and deeper tissue penetration, 18,19 would benefit the sensitive imaging of sulfatase in vivo. However, so far, as we know, such sulfatase-activated AIEgens with NIR fluorescence emissions have not been reported.

Herein, a sulfatase-activatable NIR fluorescence probe (QMT-SFA) was fabricated, which contains an AIE luminophore (QMT)¹³ and a hydrophilic sulfatase-cleavable sulfate moiety (Fig. 1). As we hypothesized, QMT-SFA exhibited an initially weak fluorescence emission under physiological conditions because the hydrophilic QMT-SFA molecules were dispersive. When the sulfate group in QMT-SFA was cleaved by sulfatase inside BC cells, the obtained hydrophobic QMT-OH aggregated into nanoparticles, which emitted bright NIR fluorescence through the AIE mechanism. In this study, QMT-SFA has been successfully applied to visualize sulfatase activity *in vivo*.

[†]Electronic supplementary information (ESI) available. See DOI: https://doi.org/ 10.1039/d4nr01314j

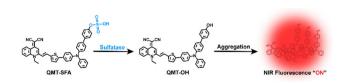


Fig. 1 Schematic illustration of the sulfatase-activatable near-infrared (NIR) aggregation-induced emission probe QMT-SFA.

^aState Key Laboratory of Digital Medical Engineering, School of Biological Science and Medical Engineering, Southeast University, Nanjing, Jiangsu 210096, China. E-mail: gliang@seu.edu.cn, xbsun@seu.edu.cn

^bState Key Laboratory of Analytical Chemistry for Life Science, School of Chemistry and Chemical Engineering, Nanjing University, Nanjing, Jiangsu 210023, China ^cHandan Norman Technology Co., Ltd, Guantao 057750, China

Nanoscale

First, QMT-SFA was synthesized according to the procedure illustrated in Scheme S1,† and characterized by electrospray ionization mass spectrometry (ESI-MS, Fig. S1†). After synthesis, we initially explored the responsiveness of OMT-SFA toward sulfatase in phosphate buffered saline (PBS, 100 mM, pH 7.4, 1% DMSO). Briefly, QMT-SFA (10 µM) was incubated

with 40 U mL⁻¹ sulfatase at 37 °C for 1 h. The chemical evolution of **OMT-SFA** in response to sulfatase was traced by highperformance liquid chromatography (HPLC). As shown in Fig. 2a, 70% QMT-SFA (retention time: 11.6 min) was cleaved by sulfatase and converted to a new product, which shared the same retention time (18.0 min) as QMT-OH. ESI-MS results further certified that this compound (at 18.0 min) was QMT-OH (Fig. S2†). These results confirmed that QMT-SFA was successfully hydrolyzed by sulfatase to QMT-OH. Furthermore, molecular QMT-SFA could form into nanoparticles with an average diameter of ~25.5 nm (Fig. 2b, S3 and S4†). We further recorded the fluorescence spectra to verify whether this sulfatase-triggered nanoparticle formation could turn the NIR AIE fluorescence "on". As shown in Fig. 2c, under the above conditions, QMT-SFA exhibited a low fluo-

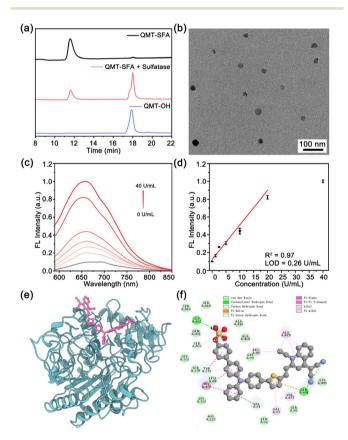


Fig. 2 (a) HPLC analyses of 10 μM QMT-SFA (black) and 10 μM QMT-SFA treated with 40 U mL $^{-1}$ sulfatase for 1 h (red), and 10 μ M QMT-OH (blue). (b) TEM image of 10 µM QMT-SFA treated with 40 U mL^{-1} sulfatase. (c) Fluorescence response of QMT-SFA (10 μ M) toward different levels of sulfatase. (d) The fitted relationship between the fluorescence intensity of QMT-SFA and sulfatase concentration. (e) Ribbon diagram of the binding interface between QMT-SFA and sulfatase. (f) 2D diagram of the interaction interface between QMT-SFA and sulfatase.

rescence signal at 660 nm, whereas the signals increased with the concentration (0-40 U mL⁻¹) of sulfatase, indicating that QMT-SFA was successfully activated by sulfatase to emit AIE fluorescence (Fig. 2c). Moreover, there was a linear relationship $(y = 0.04x + 0.12, R^2 = 0.97)$ between the fluorescence intensity and the concentration of sulfatase within the range of 0-20 U mL⁻¹ (Fig. 2d). The limit of detection (LOD) was calculated to be about 0.26 U mL⁻¹. These results indicated that QMT-SFA could be used as a sensitive probe for sulfatase detection in vitro.

Molecular docking calculations were performed to study the binding affinity between QMT-SFA and sulfatase. The protein structure of sulfatase (PDB ID: 1AUK) was chosen for docking analysis. 6,17 As shown in Fig. 2e and f, QMT-SFA forms strong hydrophobic interactions with seven amino acids (Val91, Tyr379, His405, Ala478, Pro8, Val93 and Val477) and hydrogen bonds with five amino acids (Glu481, Gly402, Val91, Tyr379 and Ser403) of sulfatase. Meanwhile, there is also a Coulomb interaction between GLU481 of sulfatase and QMT-SFA (Table S1†). Molecular docking data suggested that QMT-SFA has a high binding energy with sulfatase (-6.19 kcal mol⁻¹). The above results demonstrated the good binding ability between QMT-SFA and sulfatase, which permitted efficient enzymatic interactions.

We further investigated the potential of QMT-SFA in imaging intracellular sulfatase activity. Before cell imaging, the cytotoxicity of OMT-SFA was evaluated in 4T1 cells by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. As shown in Fig. S5,† after incubation with QMT-SFA at different concentrations (up to 320 µM), 4T1 or L02 cells retained high-level viability (>80%), indicating the low cytotoxicity of QMT-SFA. We then tested whether QMT-SFA could be used for sulfatase activity imaging in 4T1 cells. Timecourse fluorescence images of QMT-SFA-treated 4T1 cells indicated that the fluorescence of 4T1 cells rapidly increased with time and reached its maximum at 1 h (Fig. S6†), suggesting the fast activation of QMT-SFA in 4T1 cells. Then, we chose 1 h as the incubation time of QMT-SFA in the following cell experiments. As shown in Fig. 3, 4T1 cells incubated with 10 µM QMT-SFA showed bright AIE fluorescence, suggesting that QMT-SFA was activated by cellular sulfatase. In contrast, L02 (a normal cell that has undetectable sulfatase) showed negligible AIE fluorescence after incubation with 10 μM QMT-SFA for 1 h. Moreover, when the cells were pre-incubated with a 1 mM sulfatase inhibitor (hydroxylamine) for 0.5 h, no obvious red fluorescence was observed after QMT-SFA treatment, demonstrating that QMT-SFA was indeed activated by intracellular sulfatase. The above results indicated that QMT-SFA could be used for the sensitive imaging of intracellular sulfatase activity.

Inspired by its excellent performance, we further employed QMT-SFA to monitor sulfatase in vivo. The 4T1 tumor-bearing nude mice were randomly divided into two groups. Briefly, mice in one group were merely intratumorally injected with 0.25 mg kg⁻¹ QMT-SFA (i.e., the "QMT-SFA" group), while those in the other group were pre-injected with hydroxylamine for 0.5 h, followed by the injection with QMT-SFA (i.e., the

Communication Nanoscale

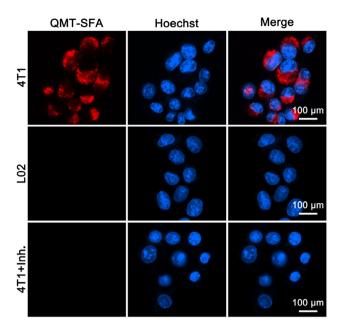


Fig. 3 Fluorescence images of 10 μM QMT-SFA-treated 4T1 cells, L02 cells, and hydroxylamine-pretreated 4T1 cells for 1 h.

"QMT-SFA + Inh." group). The fluorescence signals of the tumor regions were monitored by a small animal imaging system. As shown in Fig. 4, for the mice in the "QMT-SFA" group, the fluorescence within the tumor region was quickly turned "on" and reached the strongest brightness at 1 h postinjection, followed by a slow decrease within 16 h (Fig. S7†). In contrast, for the mice in the "QMT-SFA + Inh." group, a significantly low-level fluorescence profile was observed within the tumor region. The above results illustrated that QMT-SFA can be hydrolyzed by sulfatase to turn its NIR AIE fluorescence "on" in 4T1 tumor-bearing nude mice. In addition, the biodistribution of QMT-SFA in the above nude mice was further observed by ex vivo fluorescence imaging. As shown in Fig. S8,† fluorescence was undetectable in the major mouse organs (i.e., heart, liver, spleen, lungs, and kidney) from both

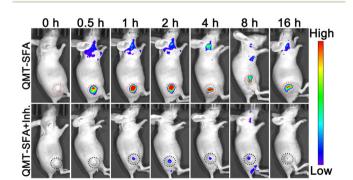


Fig. 4 Time-course fluorescence images of 4T1 tumor-bearing mice after the intratumoral injection of 0.25 mg \mbox{kg}^{-1} QMT-SFA. For the "QMT-SFA + Inh." group, tumors were pretreated with 0.02 mg kg⁻¹ inhibitor for 0.5 h before QMT-SFA injection. The dashed circles indicate tumor regions.

groups, as well as in the tumor from the "QMT-SFA + Inh." group. In contrast, the tumor from the "QMT-SFA" group showed a strong NIR fluorescence. These in vivo results demonstrated that QMT-SFA could be applied for sulfatase imaging in tumors of living animals.

Conclusions

In summary, we have successfully designed an NIR fluorescence probe QMT-SFA for the sensitive detection of sulfatase in vitro and imaging sulfatase activity in tumors of living mice. QMT-SFA could be cleaved by sulfatase to yield QMT-OH, which thereby aggregated into nanoparticles to turn the NIR AIE fluorescence "on" (centered at 660 nm). The probe exhibited high sensitivity toward sulfatase with an LOD of 0.26 U mL⁻¹ in vitro. Molecular docking calculation showed that **QMT-SFA** had a high binding affinity $(-6.19 \text{ kcal mol}^{-1})$ with the sulfatase docking score. In addition, QMT-SFA had been successfully employed for tracking sulfatase in 4T1 cells and 4T1 tumor-bearing nude mice. To the best of our knowledge, QMT-SFA was the first NIR AIEgen for sulfatase imaging in living cells and animals. Compared with current sulfatase AIEgens which emit visible light, QMT-SFA has higher potential in biological application owing to its improved tissue penetration and suppressed autofluorescence interference. We anticipate that this NIR AIEgen QMT-SFA could be employed for the early diagnosis of sulfatase-associated disease in clinic in near future.

Author contributions

Lingling Xu: investigation, formal analysis, methodology, validation, visualization, and writing - original draft. Yu Deng: investigation and validation. Hang Gao: validation and project administration. Yuchen Yao: validation. Xiaoyang Liu: visualization and formal analysis. Wenjun Zhan: supervision and project administration. Gaolin Liang: funding acquisition, project administration, supervision, and writing - review & editing. Xianbao Sun: conceptualization, funding acquisition, project administration, and writing - review & editing.

Ethical statement

All animal experiments were approved and performed according to the guidelines of the Animal Care and Use Committee of the Southeast University Laboratory Animal Center (No: 20240306009).

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (Grants 22204019 and 22234002), the Full-time Talents Program of Hebei Province (Grant 2023HBQZYCXY027), and the China Postdoctoral Science Foundation funded project (Grant 2023M730577).

References

Nanoscale

- 1 S. Loibl, P. Poortmans, M. Morrow, C. Denkert and G. Curigliano, *Lancet*, 2021, **397**, 1750–1769.
- 2 A. N. Giaquinto, H. Sung, K. D. Miller, J. L. Kramer, L. A. Newman, A. Minihan, A. Jemal and R. L. Siegel, *CA Cancer J. Clin.*, 2022, 72, 524–541.
- 3 D. Barba, A. León-Sosa, P. Lugo, D. Suquillo, F. Torres, F. Surre, L. Trojman and A. Caicedo, *Crit. Rev. Oncol. Hematol.*, 2021, **157**, 103174.
- 4 I. H. Kunkler, L. J. Williams, W. J. L. Jack, D. A. Cameron and J. M. Dixon, *N. Engl. J. Med.*, 2023, **388**, 585–594.
- 5 D. Yue, M. Wang, F. Deng, W. Yin, H. Zhao, X. Zhao and Z. Xu, *Chin. Chem. Lett.*, 2018, 29, 648–656.
- 6 S. R. Hanson, M. D. Best and C.-H. Wong, Angew. Chem., Int. Ed., 2004, 43, 5736–5763.
- 7 R. Mashima and M. Nakanishi, *Int. J. Mol. Sci.*, 2022, 23, 8153.

- 8 L. Xu, N. Liu, W. Zhan, Y. Deng, Z. Chen, X. Liu, G. Gao, Q. Chen, Z. Liu and G. Liang, ACS Nano, 2022, 16, 19328– 19334.
- 9 X. Wu, R. Wang, N. Kwon, H. Ma and J. Yoon, *Chem. Soc. Rev.*, 2022, **51**, 450–463.
- 10 S. He, P. Cheng and K. Pu, *Nat. Biomed. Eng.*, 2023, 7, 281–297
- 11 J. Li, H. Gao, R. Liu, C. Chen, S. Zeng, Q. Liu and D. Ding, *Sci. China: Chem.*, 2020, **63**, 1428–1434.
- 12 M. H. Xiang, Z. Y. Jiang, W. L. Zhao, E. Zhang, L. Xia, R. M. Kong, Y. Zhao, W. Kong, X. Liu, F. Qu and W. Tan, ACS Sens., 2023, 8, 2021–2029.
- 13 L. Xu, H. Gao, W. Zhan, Y. Deng, X. Liu, Q. Jiang, X. Sun, J.-J. Xu and G. Liang, J. Am. Chem. Soc., 2023, 145, 27748– 27756.
- 14 Z. Zhu, Q. Wang, X. Chen, Q. Wang, C. Yan, X. Zhao, W. Zhao and W. H. Zhu, *Adv. Mater.*, 2022, 34, e2107444.
- 15 S. Jia, Z. Gao, Z. Wu, H. Gao, H. Wang, H. Ou and D. Ding, CCS Chem., 2022, 4, 501–514.
- 16 J. Qi, H. Ou, Q. Liu and D. Ding, Aggregate, 2021, 2, 95-113.
- 17 Y. Xu, M. Cui, W. Zhang, T. Liu, X. Ren, Y. Gu, C. Ran, J. Yang and P. Wang, *Chem. Eng. J.*, 2022, **428**, 132514.
- 18 L. Xu, H. Gao, Y. Deng, X. Liu, W. Zhan, X. Sun, J. J. Xu and G. Liang, *Biosens. Bioelectron.*, 2024, 255, 116207.
- 19 H. Shen, F. Sun, X. Zhu, J. Zhang, X. Ou, J. Zhang, C. Xu, H. H. Y. Sung, I. D. Williams, S. Chen, R. T. K. Kwok, J. W. Y. Lam, J. Sun, F. Zhang and B. Z. Tang, *J. Am. Chem. Soc.*, 2022, 144, 15391–15402.