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Solid-state emitting twisted π-conjugate as AIE-active DSE-gen: *in vitro* anticancer properties against FaDu and 4T1 with biocompatibility and bioimaging†

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Dual-state emissive fluorogens (DSE-gens) are currently defining their importance as a transpiring tool in biological and biomedical applications. This work focuses on designing and synthesizing indole-anthracene-based solid-state emitting twisted π-conjugates using a metal-free protocol to achieve AIE-active DSE-gens, expanding their scope in biological applications. Special effort has been made to introduce proficient and photo/thermostable DSE-gens that inhibit cancer but not normal cells. Here, the lead DSE-gen initially detects cancer and normal cells by bioimaging; however, it could also confirm and distinguish cancer cells from normal cells by its abated fluorescence signal after killing cancer cells. In contrast, the fluorescence signals for a normal cell remain unscathed. Surprisingly, these molecules displayed decent anticancer properties against FaDu and 4T1 but not MCF-7 cell lines. From a series of newly designed indole-based molecules, we report one single 2,3,4-trimethoxybenzene-linked DSE-gen (the lead), exhibiting high ROS generation, less haemolysis, and less cytotoxicity than doxorubicin (DOX) for normal cells, crucial parameters for a biocompatible *in vitro* anticancer probe. Thus, we present a potentially applicable anticancer drug, offering a bioactive material with bioimaging efficacy and a way to detect dead cancer cells selectively. The primary mechanism behind the identified outcomes is deciphered with the support of experimental (steady-state and time-resolved fluorescence, biological assays, cellular uptake) and molecular docking studies.

1. Introduction

The dual-state emission (DSE) feature of aggregation-induced emission (AIE) active solid-state organic fluorogens (SSOF-gens) enables their effectiveness in biotechnology and biomedical applications due to the constant presence of emission in various states such as solutions, aggregates, and solids.[1–4] Besides, simple administration into cells and flexibility in structural modification are the main benefits of organic fluorescent molecules for their versatile application in biomedicine avoiding the ACQ effects that limit their ultimate use in image-guided treatment.[1,5] Hence, biologically non-toxic solid-state emitting AIE-active DSE-gens claim prominence in bioimaging, biosensing, and image-guided therapy systems (IGTs).[1,6,7] Moreover, small but efficient π-conjugated organic fluorogens are primarily desired in comparison to various inorganic-organic hybrid materials due to their easy synthesis/modification, high thermo/photostability, and cost-effectiveness.[8–13]

Recently, cancer cell detection by simple bio-imaging techniques using AIE-gens and DSE-gens has become a dependable, convenient, and cost-effective technique in cancer treatment.[7b,14–16] Notably, molecular imaging techniques can detect cancer mass based on the photophysical properties enabling cell visualization in living organisms at the cellular and molecular level without any disorder. Some earlier identified molecular probes for *in vitro* and *in vivo* bio-imaging have inadequate effectivity due to their photobleaching tendency, acute toxicity, and low-fluorescence efficacy.[17] Highly active commercial drugs are known to treat several types of cancers but with severe side effects.[18–21] Nevertheless, many admirable anticancer drugs are available, but with no bioimaging ability,[22,23] while numerous bioimaging agents are known without anticancer properties.[24] Hence, the discovery of an

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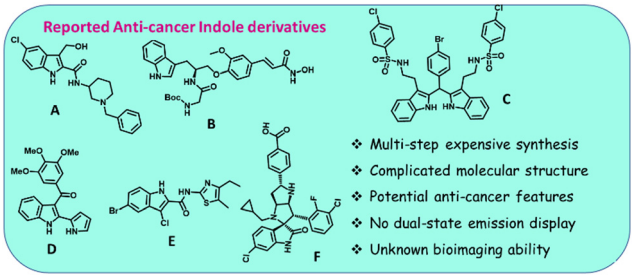

Fig. 1 Some known representative indole-based molecules (**A–F**).

anticancer agent with bioimaging ability is a looming challenge but a surging need of the time.[25]

To address this necessity, we herein focus on designing an indole-based fluorophore by incorporating anthracene and indole as the primary scaffold. Anthracene and indole are individually realized as effective anticancer agents (*vide infra*). A few essential indole-based molecules **A–F** and their pros and cons are presented in Fig. 1.[26–31]

Our thoughtful design makes the typical ACQ-fluorophore anthracene emissive in the aggregate and solid states. An additional conjugation was needed to enhance the emission intensity. The newly designed indole-anthracene-based dyes could be easily accessed economically and mainly established as efficient AIE-active DSE-gens. Here, a few aggressive and obdurate cell lines were chosen: (i) FaDu (one of the most aggressive hypopharyngeal carcinoma cell lines),[32–34] (ii) 4T1 (a highly destructive triple-negative mouse breast cancer cell line (TNBC), with a lack of treatment scope),[35–39] (iii) MCF-7 (a breast cancer cell line, but not triple negative),[40] and (iv) the non-cancerous human kidney embryonic cell line HEK-293 to investigate normal cell cytotoxicity and for comparison in bioimaging. Most approved marketed therapeutic drugs for breast cancer (sporadic for 4T1) are not emissive and have diverse side effects.[37] Among various substituents, the highly electron-rich 2,3,4-trimethoxybenzene (TMB) linked molecule only exhibits promising activity against the FaDu and 4T1 cell lines but is surprisingly not responsive to the MCF-7 cell line. The non-toxic nature towards the normal cell line HEK-293 was also verified. The pyridyl-linked emitters are active against FaDu and 4T1 but are cytotoxic to normal cell lines. Apart from inhibiting, the TMB-linked emitter could image both FaDu and 4T1 cell lines with green emission before cell death but the fluorescence signal was diminished 24 h after cell death. In addition, commendable ROS generation and less haemolysis exhibited by the lead molecule (**SB4**) enhance the scope of utilization. Thus, this report (Fig. 2) reports the discovery of new AIE(E)-active DSE-gens as cancer-inhibiting molecules with bioimaging ability. Relevant steady-state and time-resolved photophysical studies, biological assays, and docking studies mechanistically support the observed outcomes.

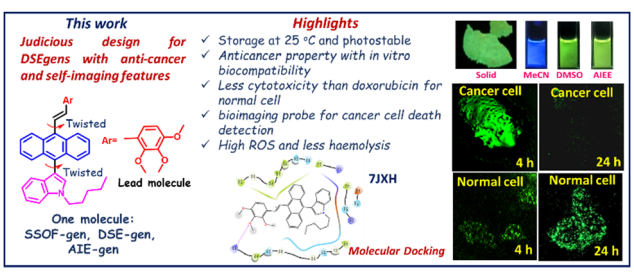

Fig. 2 Molecular design and highlights of this work.

2. Results and discussion

2.1. Molecular engineering

Considering the earlier identified design for organic solid-state emitters, specially DSEgens, we intended to obtain twisted organic molecules, having donor(D)–π–acceptor(A) and D–π–D combinations.[41–43] Thus, the flat indole (an established motif with anticancer properties) with a moderately long alkyl chain was tagged onto the flat anthracene ring. This was made possible through an easy electrophilic aromatic substitution reaction[44] using freshly prepared alkylated (*n*-hexyl) indole[45] as a nucleophile. These two cores, indole and anthracene, prefer to occupy different planes. Such a twisted structure would prohibit π–π stacking in aggregates, resulting in relaxation through a radiative channel and strong emission.

The designed D–π–A (**SB1-SB3**) and D–π–D combinations (**SB4-SB7**, Fig. 3) could be generated by Horner–Wadsworth–Emmons (HWE)-reactions of indole-anthracenyl phosphonates at room temperature through a metal/ligand-free protocol.[46]

The twisted structure and extended π-conjugation favorably compromise to emit with high efficiency in solution and solid states. Moreover, the indole and anthracene cores are individually potential anticancer agents.[47–51] Molecules with extended conjugation with the core consisting of pyridine, TMB, phenothiazine, and triphenylamine were conveniently created because these are well-known motifs for bioimaging and anticancer applications.[52–56] Unlike many previous reports, all synthesized molecules were intended to be non-charged, in order not to compromise with the fluorescence quantum yield,[57,58] despite a possible reduction in cellular

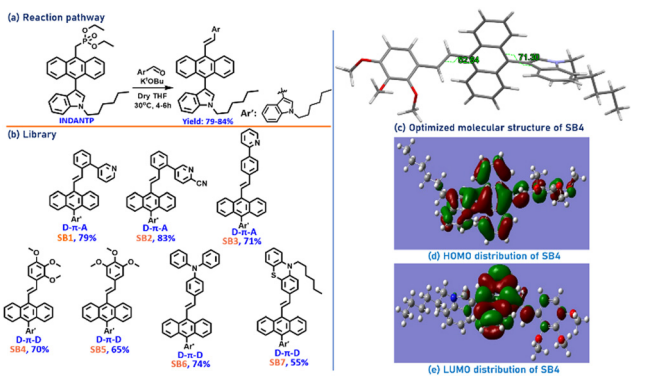

Fig. 3 (a) Synthetic route for the indole-anthracenyl analogs; (b) library of the synthesized molecules. (c) Optimized structure of one representative twisted π-conjugate **SB4** in the gas phase; (d) HOMO and (e) LUMO distributions of **SB4**.

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permeability.[59] Nevertheless, the long alkyl chain and many hydrophobic units in our synthesized probes assist in achieving commendable cellular uptake capacities (*vide infra*). Notably, these probes are significantly stable and storable at room temperature, in contrast to many other complex organic molecules due to chiral/spiro centers or complex functionalization.[60,61] Introduction of a conventional 'spacer' was also avoided to keep the fluorescence efficacy intact because it might disrupt the π-conjugation and introduce molecular motion, leading to quenched emission.[62]

Despite several attempts to generate a single crystal of these molecules, a suitable crystal for X-ray diffraction studies could not be generated. However, the viability of the molecular design and electronic effects on the lead molecule **SB4** were validated by determining its optimized molecular structure using density functional theory (DFT) calculations at the B3LYP/6-31G(d,p) level. The calculated gas-phase HOMO, LUMO, and optimized molecular structure are presented in Fig. 3c–e. The molecular structure is vastly twisted with a dihedral angle of 71.3° (anthracene/indole) and ~53° (anthracene-TMB). The HOMO is mainly distributed on the indole-anthracene part with a fine dispersion on the TMB ring, and the LUMO is mainly located at the core of anthracene. As the anthracene core contributes to both the HOMO and LUMO, generation of a charge transfer state is unexpected.

2.2. Photophysical studies in the solid state

We primarily focused on emission in solution, aggregates, and the solid-state for all these synthesized D–π–A (**SB1-SB3**) and D–π–D (**SB4-SB7**) molecules to make them suitable for biological applications. Notably, all these synthesized π-conjugates are SSOF-gens with green/greenish-yellow emission under 365 nm UV light except **SB7**, whose emission was orange (Fig. 4). The solid-state absorbance, emission, and other related parameters are tabulated elsewhere (see ESI† Fig. S1 and Table S1). In the solid-state UV-Vis studies, **SB1** shows maximum λ_{abs} at 426 nm with a bent phenyl-pyridinyl core while **SB2** has maximum λ_{abs} at 458 nm due to the suitably placed cyano group, improving π-conjugation in the ground state. In the emission profile, **SB1** displays emission at 510 nm with higher brightness (absolute quantum yield φ_f = 7.04%), but **SB2** is feebly emissive (φ_f = 1.29%) with λ_{em} at 505 nm. Another pyridinyl-conjugated dye **SB3** performed as an intense solid-state emitter (φ_f = 26.64%) with λ_{em} at 517 nm. Out of the two TMB-linked isomeric D–π–D systems, **SB4** with 2,3,4-TMB emits at $\lambda_{max.}$ = 534 nm (φ_f = 10.28%) with a 33 nm red-shift compared to its 3,4,5-TMB analog **SB5** ($\lambda_{max.}$ = 501 nm, φ_f = 7.00%), possibly due to the presence of –OMe at both the *ortho* and *para* position. The bulky and conformationally twisted triphenylamine provided intense emission for **SB6** ($\lambda_{max.}$ = 517 nm, φ_f = 20.11%). Still, the bowl-shaped phenothiazine moiety introduced a weak red emission in **SB7** ($\lambda_{max.}$ = 589 nm, φ_f = 2.66%).

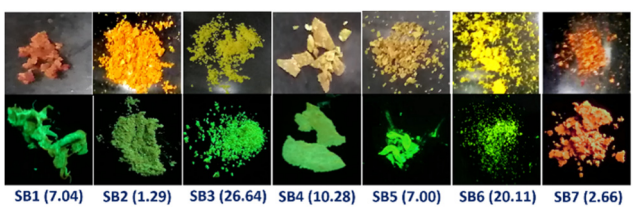

Fig. 4 The solid-state images of the molecules taken under sunlight (top row) and a 365 nm UV lamp (bottom row). The φ_f% values of these molecules are stated in parentheses.

2.3. Dual-state emission (DSE) feature studies

To examine the solution state emission property of the SSOF-gens, DMSO was chosen as a solvent due to its numerous applications in biology and medicine.[63] However, MeCN was also used as another polar solvent. Moreover, both DMSO and MeCN are water-miscible and, thus, suitable for AIE studies. All the synthesized SSOF-gens (**SB1-SB7**) were identified as dual-state emitters because weak/decent emissions were observed even in such polar solutions (Fig. 5a, b and Fig. S2, Table S2, ESI†) and the solid state (Fig. 4 and Fig. S1, Table S1, ESI†). The DSE-gen behaviors were well-supported by TD-DFT calculations performed with the representative D–π–A molecule **SB2** and D–π–D molecule **SB4**. The excited singlet state (S_1) achieves more planarity than the ground singlet S_0 state for **SB2** and **SB4** in both solvents, resulting in emissions in solutions (Fig. 5c, d and Fig. S3, S4, Table S3, ESI†). Compounds **SB2** and **SB4** both have a lower φ_f in DMSO than in MeCN, most possibly due to a relatively higher root mean square displacement (RMSD) (between S_0 and S_1) and weaker emission oscillator strength in DMSO.[64] The HOMO–LUMO energy gap in such polar solvents would be smaller to afford a weakly emissive solution. However,

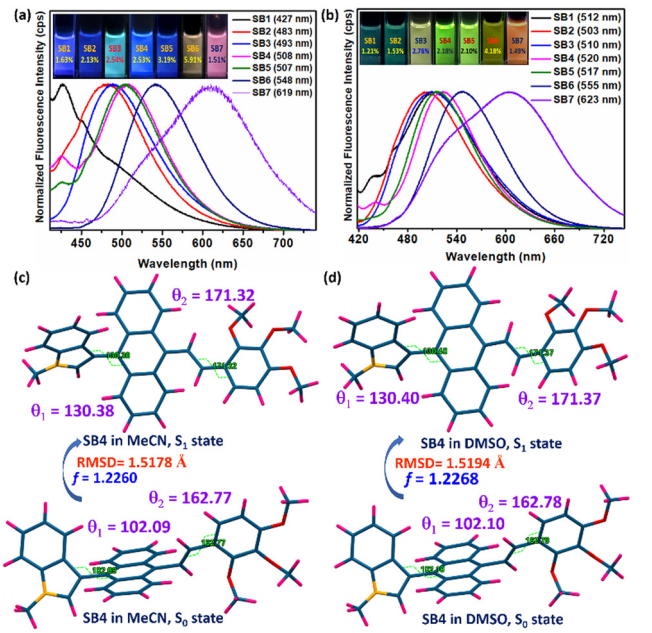

Fig. 5 Solution-state fluorescence with images (under a 365 nm UV lamp) of **SB1-SB7** in (a) MeCN and (b) DMSO with their emission λ_{max} and respective relative φ_f (%), [error: (±) 5%] [reference: 10–5 M quinine sulfate solution]; (c) TD-DFT optimized structures of **SB4** with selected torsion angles (θ_1 and θ_2). The DFT/TDDFT calculations were performed with *N*-methyl only as the long chain will not change its electronic properties (see 'Computational details' in the ESI†).

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Table 1 AIE/E-genic properties in a nutshell

SSOF-gen with AIE-property	$\lambda_{max.\ abs.}$ (nm) in agg. state	$\lambda_{max.\ em.}$ (nm) in agg. state	α-factor	*Z*-avg. (d nm) in agg. state	PDI	Life-time in agg. state (ps)
SB3	425	530	3.52 (8.94/2.54)	194.5	0.296	64
SB4	423	517	1.61 (4.07/2.53)	186.1	0.178	66
SB5	418	514	1.12 (3.57/3.19)	190.4	0.197	48
SB6	428	533	2.64 (10.52/3.91)	219.9	0.295	51
SB7	440	565	1.18 (2.25/1.91)	190.7	0.185	49

this energy gap was slightly larger for **SB4** than **SB2** (Fig. S5 and Table S3, ESI†), ensuring the slightly better φ_f for **SB4**.

2.4. Aggregation-induced emission (AIE) property of the DSE-gens

We aimed to verify the AIE activities of the synthesized DSE-gens and expected them to emit in the aggregated state as well. Notably, DSE-gens and SSOF-gens with AIE properties are more valuable in reporting biological information.[4b,4g,65,66] Initially, the AIE properties of all the DSE-gens were tested in an acetonitrile/water mixture (fraction of water in acetonitrile f_w: v/v%). All of the DSE-gens discussed here were not AIE-active. Especially, the admirable solid-state fluorophore **SB1** undergoes ACQ (Fig. S6, ESI†) with a gradual increment of water fraction (f_w), possibly due to the π-stacking of the phenyl/pyridine rings in the aggregated state.[67,68] Introduction of the cyano group in **SB2** might prevent π-stacking to a small extent, and the AIE-property of **SB2** was somewhat better but not promising (Fig. S7, ESI†). The AIE feature of **SB3** is more pleasing (Fig. S8, ESI†), with an 11 times fluorescence boost at f_w = 80%. The aggregate formation was also verified from the DLS (dynamic light scattering) studies (Table 1). A notable AIE feature (7 times increment) was noticed at f_w = 70% with **SB4**, containing three methoxy groups in an unsymmetrical fashion (Fig. 6a, b and Fig. S9a, ESI†). The indole with a long alkyl chain, twisted structure, and lack of symmetry may significantly restrict π-stacking and facilitate AIE. A simple viscofluorochromism study further supported this speculation. In MeOH, **SB4** is weakly emissive, and upon consecutive addition of glycerol into MeOH, the overall viscosity of the medium is gradually increased, which would help restrict the molecular motion, resulting in enhanced emission. To our delight, the emission efficacy of **SB4** was progressively enhanced with increasing viscosity up to a f_G of 70% (Fig. 6c, d and Fig. S9b, ESI†), indicating viscofluorochromism in **SB4**.

Another symmetrically different analog, **SB5**, was also identified as a good AIE-gen (Fig. S10, ESI†). The non-planar conformation of TPA in **SB6** and bowl-shaped-phenothiazine in **SB7** induce molecular twisting in the aggregate form, enabling destruction of the ACQ effect and the occurrence of AIE (Fig. S10, ESI†). An emission enhancement efficiency (I/I_0) comparison (Fig. 7) for all the synthesized molecules was also performed to gain a clear view and the AIE-genic properties are tabulated for clarity (Table 1). The photostability of **SB4** (Fig. 7) was investigated in its aggregated state (f_w of 70%). SEM images of the AIE-gens in their respective aggregated states were captured with different morphologies (Fig. S11, ESI†).

We were interested in measuring the nanosecond scale excited state lifetime of one of the molecules, **SB4** (as a lead; *vide infra*), in the solution, aggregate, and solid states. It was hard to measure the lifetime on a nanosecond scale due to its quick decay from the excited state. However, the decay profiles (ESI,† Fig. S12 and Table S4) were fitted better into bi or triexponential forms. The experiments were carefully repeated to confirm the lifetime in less than a nanosecond range. As observed in the emission intensity, the lifetime (ns) was enhanced in the aggregate/solid state, as previously reported for other relevant molecules.[69] The radiative (k_r) and nonradiative (k_{nr}) rate constants were measured (Table S4, ESI†) and the large nonradiative rate constant in solution explains the weak

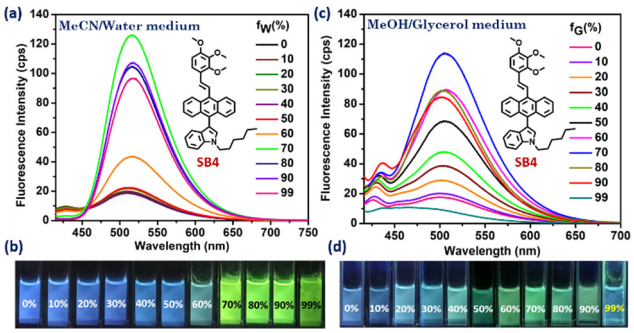

Fig. 6 AIE-property (a and b) and viscofluorochromism (c and d) of **SB4**; f_w is the fraction of water (v/v%) in MeCN and f_G is the fraction of glycerol (v/v%) in MeOH.

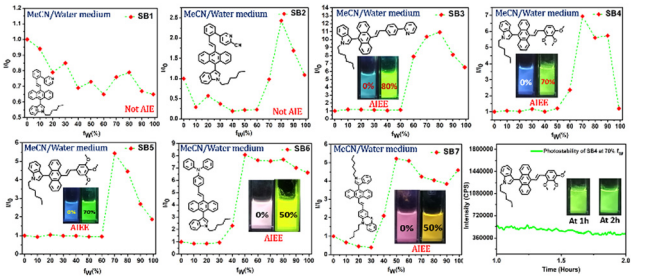

Fig. 7 Efficiency of AIE property of indole-anthracenyl derivatives and photostability of **SB4** at f_w (%) = 70. α-factor: φ_f in the aggregated state/φ_f in the solution state. DLS particle size is presented as '*Z*-avg.' in the 'nm' unit with the PDI value. Here, 'agg' stands for 'aggregated', 'avg.' stands for 'average', and 'PDI' is the polydispersity index.

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emission. The gradually improving k_r in the aggregate and solid state explains the emission enhancement.

2.5. Biological applications

2.5.1. Cytotoxicity of the synthesized compounds against different cell lines. The evaluation of *in vitro* cytotoxicity of all these DSE-gens was conducted against FaDu (a robust and aggressive cell line), the 4T1 triple-negative breast cancer cell line (TNBC), and the MCF-7 non-TNBC cancer cell line. After incubation with different concentrations of these dyes for 24 h, the cell viability was determined with the standard MTT assay. GI_{50} values for the individual cell lines were defined as the concentration of the compounds at 50% survival of cells. DOX was utilized as the standard drug. The anticancer activities of these dyes are tabulated below (Table 2).

From the cytotoxicity assay results using the cancer cell lines, only five active DSE-gens were selected to check their cytotoxicity against the non-cancerous human kidney embryonic cell line HEK-293. Notably, the DSE-gen **SB4** shows the best anticancer activity against both FaDu and 4T1 and is less cytotoxic than DOX for HEK-293 normal cells. Surprisingly, it was inactive against the comparative cell line MCF-7. The table also indicates good responses from **SB1** and **SB2** against FaDu and 4T1, but these compounds are acutely toxic towards HEK-293 and, thus, inappropriate for further use. The TPA-analog **SB6** also possesses favorable anti-cancer activity against FaDu but not 4T1.

2.5.2. Reactive oxygen species (ROS) generation and *in vitro* haemolysis study. Reactive oxygen species (ROS) is a phrase used to indicate several reactive molecules and free radicals derived from molecular oxygen. ROS dynamically influence the tumor microenvironment and are known to initiate cancer angiogenesis, metastasis, and survival at different concentrations.[70] At moderate concentrations, ROS activate the cancer cell survival signaling cascade. At high concentrations, ROS can cause cell apoptosis. Thus, this shows their potential as an effective anti-tumor modality and therapeutic target for treating malignancies.[70] To evaluate the ROS generation ability of the DSE-gens[7c] against a rigid and aggressive cell line like FaDu, we selected an oxidant-sensitive fluorescent dye named dichlorofluorescein diacetate (DCF-DA). Since DCF-DA has a high fluorescence quantum yield, the interference from the DSE-gens was minimized while evaluating the ROS generation ability. The efficiency of the lead molecule **SB4** was almost identical to that of the standard drug DOX (Fig. 8a).

Table 2 GI_{50} values of all the AIE-gens in different cancer cell lines

GI_{50} (μM) for each cell-line	Doxorubicin	SB1	SB2	SB3	SB4	SB5	SB6	SB7
FaDu	0.24–0.3	39.27	37.53	>100	24.94	>100	27.21	69.27
4T1	0.85–2.2	49.75	32.99	>100	20.76	82.11	>100	>100
MCF-7	1.07–2.57	88.7	>100	>100	>100	>100	>100	>100
HEK-293	3.3	35.63	33.72	—	>100	—	>100	>100

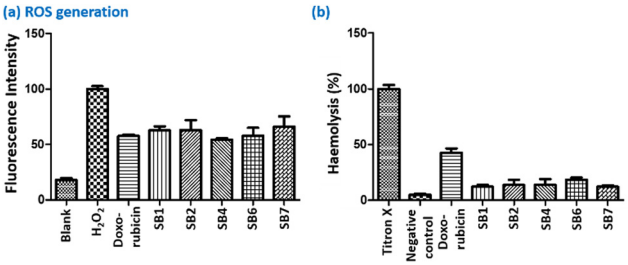

Fig. 8 (a) ROS and (b) haemolysis studies of the DSE-gens.

In addition, we treated red blood cells with these DSE-gens at their respective GI_{50} concentrations and then analyzed the treated cells by the haemolysis (rupture of red blood cells) assay. For the standard titron X, 100% haemolysis was considered, whereas DOX showed 40% haemolysis, a destructive impact. Interestingly, AIEgens **SB1**, **SB2**, **SB4**, **SB6**, and **SB7** exhibited an inferior haemolyzing effect compared to DOX (Fig. 8b). It is noteworthy that **SB1**, **SB2**, **SB4**, **SB6**, and **SB7** are less cytotoxic than DOX. Thus, the prime molecule **SB4** offers a decent anticancer property against FaDu and 4T1 with excellent ROS generation and a weaker haemolyzing effect in comparison with DOX.

2.5.3. Cellular uptake. The cellular uptakes of the DSE-gens **SB1**, **SB2**, **SB4**, **SB6**, and **SB7** were investigated by Fluorescence-Activated Cell Sorting (FACS) analysis against the FaDu cell line to understand the nature of the single-cell population. All the synthesized compounds displayed excellent cellular uptake (Fig. 9). Later cellular uptake of lead molecule **SB4** was investigated against other cell lines, 4T1, MCF-7, and HEK-293, and it displayed an excellent cellular uptake capacity for all. Hence, it is clear that the dyes can penetrate 4T1 and MCF-7 cells, and their specificity toward 4T1 is not due to any variance in cellular uptake capacity (Fig. 9).

2.5.4. Utilizing the anticancer agent SB4 in bioimaging of FaDu and 4T1 cell lines. As DMSO is a preferable solvent in a biological experiment regarding cell culture, the emission

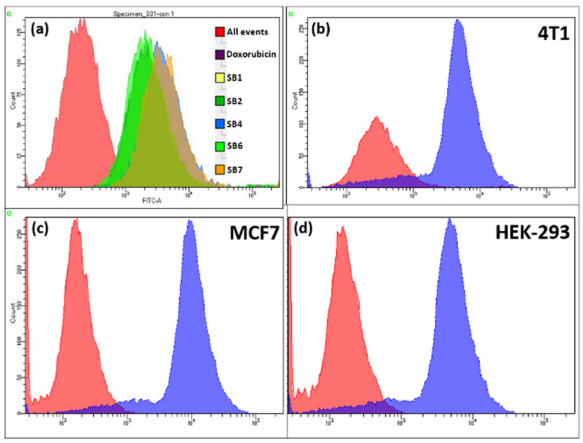

Fig. 9 (a) Cellular uptake capacity of DSE-gens into FaDu cells; cellular uptake capacity of lead DSE-gen **SB4** into (b) 4T1, (c) MCF-7, and (d) HEK-293 cells.

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properties of the biologically active molecules **SB1**, **SB2**, **SB4**, **SB6**, and **SB7** were also checked in a DMSO/water medium. To our satisfaction, the emission efficacies of these molecules were boosted in this new biological environment (Fig. S13, ESI†).

All the above results convincingly define **SB4** as a bioimaging agent able to visualize the cancer cells FaDu, 4T1, and HEK-293 before and after inhibition. After four hours of **SB4** treatment in both the FaDu and 4T1 cell lines at the GI_{50} concentration, the confocal experiment was performed, keeping the cells fixed, and the image was captured. Intense green fluorescence was noticed upon staining cells with **SB4** at its GI_{50} concentration. As the cytotoxicity assay against cancer cell lines was conducted at a 24 h time-point, confocal images were again captured 24 h after treatment using the same cell lines. Regarding the anticancer properties, **SB4** will inhibit the cells and will not provide any intense fluorescence signal to be observed by a confocal microscope at the 24 h time-point. But for HEK-293, a strong green fluorescence signal can be observed at 4 h and at 24 h because **SB4** does not inhibit HEK-293 normal cells. Thus, **SB4** can not only kill but also detect cancer cell death with its diminished fluorescence signal (Fig. 10).

The utility of **SB4** as a bioimaging probe for both cancer cell lines (Fadu and 4T1) was further investigated, and a comparative study was conducted with the standard drug DOX using DAPI'(4′,6-diamidino-2-phenylindole) as a typical stain.[71] The experimental outcomes indicate that DAPI and DOX could stain FaDu and 4T1 at both the time points of 4 h and 24 h. But, as stated earlier, **SB4** could stain cancer cells initially at the 4 h time-point, but a scathed fluorescence signal was noticed at the 24 h time-point for FaDu and 4T1 (Fig. 11 and 12). The used concentration of DAPI was 1 μg mL^{-1}, and DOX was used at its GI_{50} concentration for bioimaging (0.3 μM for FaDu and 2.2 μM for 4T1). DAPI is a well-known bis-intercalator with DNA and a minor-groove binder.[72] It can stain live and dead cells because the nucleus is not ruined instantly after the cell death. Interestingly, DOX also forms complexes by intercalation between DNA-base pairs,[73] and it can stain live and dead cells like DAPI.[74] These may be the reasons for recording fluorescence signals from both DAPI and DOX before and after the death of the cancer cells, respectively at 4 h and 24 h. However, the result is different for **SB4** than the standard drug DOX. The colocalization studies showed that **SB4** is not a nucleus-specific dye and stains the whole cell. Therefore, **SB4**- staining patterns remain intact for FaDu, 4T1, and HEK-293. As **SB4** is nontoxic against HEK-293, the cell is not ruptured, and hence **SB4** can possibly reside within the cell to provide a fluorescence signal even after 24 h and 4 h. Of note, aggregates are known to generate ROS.[7c] Likewise, the AIE-active DSE-gen **SB4** generates ROS and exhibits more affinity towards the HER2 protein, ensuring cancer cell inhibition. Meanwhile, cell membranes might be ruptured in dead cells and permit the dye to leak out of the cell. Hence, the dye **SB4** would tend to leak out of the cell, causing reduced fluorescence signals due to its lower concentration inside the cell. Nevertheless, the actual mechanism behind this observation needs further detailed investigation.

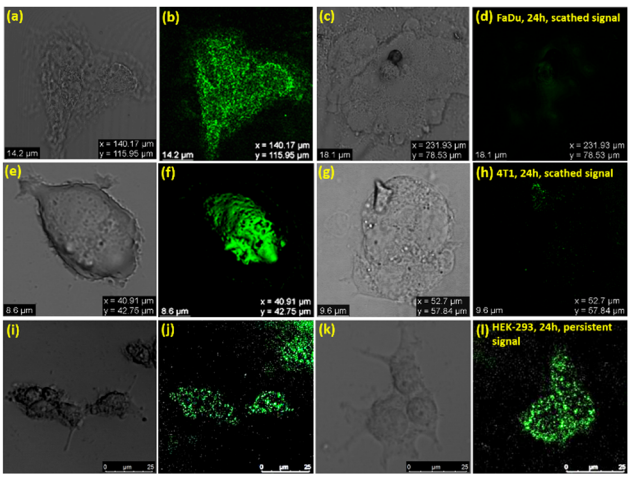

Fig. 10 Confocal microscopic images of **SB4**; For FaDu cells: at 4 h: (a) bright-field and (b) fluorescence images; after 24 h: (c) bright-field image and (d) diminished fluorescence; for 4T1 cells: at 4 h: (e) bright-field image and (f) fluorescence image; after 24 h: (g) bright-field image and (h) scathed fluorescence signal; for HEK-293 cells: at 4 h: (i) bright-field image and (j) fluorescence signal; after 24 h: (k) bright-field image and (l) persistent fluorescence.

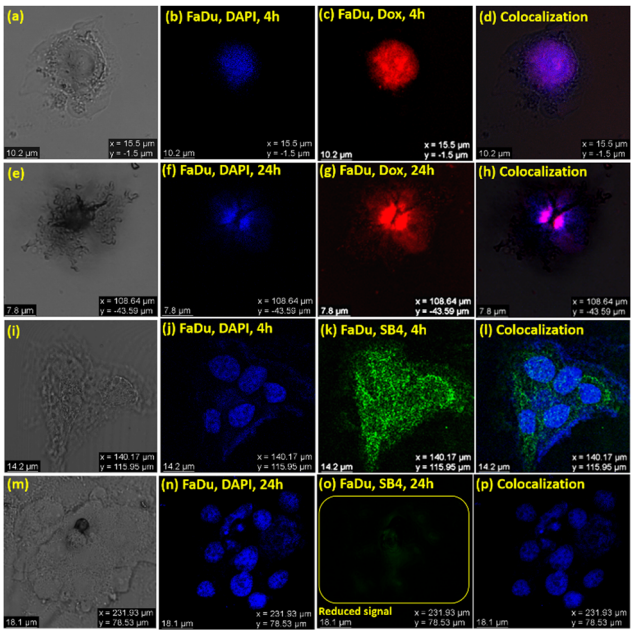

Fig. 11 Confocal microscopic images of FaDu cells with **DOX** and colocalization comparison with DAPI: At 4 h: (a) bright-field image, (b) fluorescence image (with **DAPI**), (c) fluorescence image (with **DOX**), and (d) merged images of a, b, and c to realize colocalization; after 24 h: (e) bright-field image, (f) fluorescence image (with **DAPI**), (g) persistent fluorescence image (with **DOX**), and (h) merged images of e, f, and g to recognize colocalization; confocal microscopic bioimages of FaDu cells with **SB4** and colocalization comparison with DAPI (other sets of cells, to distinguish anti-cancer features and imaging capabilities); at 4 h: (i) bright-field image, (j) fluorescence image (with **DAPI**), (k) fluorescence image (with **SB4**), and (l) merged imags of i, j, and k to realize colocalization; after 24 h: (m) bright-field image, (n) fluorescence image (with **DAPI**), (o) weakened fluorescent image (with **SB4**), and (p) merged images of m, n, and o to comprehend colocalization.

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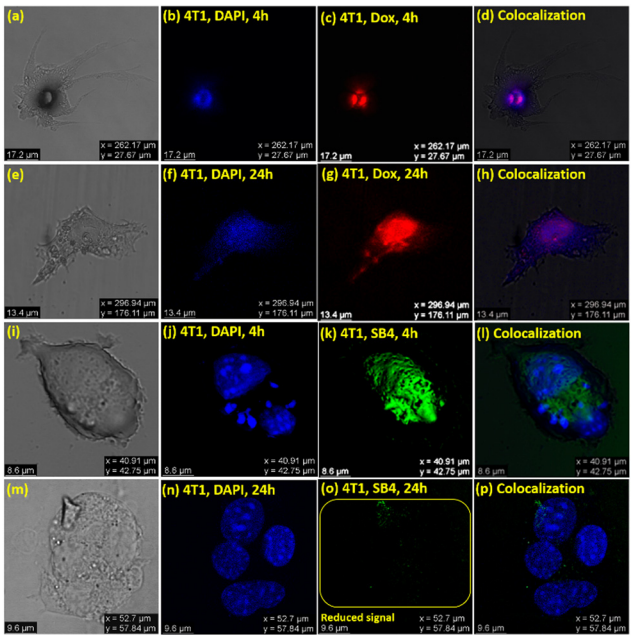

Fig. 12 Confocal microscopic images of 4T1 cells with **DOX** and colocalization comparison with DAPI: At 4 h: (a) bright-field image, (b) fluorescence image (with **DAPI**), (c) fluorescence image (with **DOX**), and (d) merged images of a, b, and c to realize colocalization; after 24 h: (e) bright-field image, (f) fluorescence image (with **DAPI**), (g) persistent fluorescence image (with **DOX**), and (h) merged images of e, f, and g to recognize colocalization; confocal microscopic bioimages of 4T1 cells with **SB4** and colocalization comparison with DAPI: at 4 h: (i) bright-field image, (j) fluorescence image (with **DAPI**), (k) fluorescence image (with **SB4**), and (l) merged images of i, j, and k to realize colocalization; after 24 h: (m) bright-field image, (n) fluorescence image (with **DAPI**), (o) weakened fluorescence image (with **SB4**), and (p) merged images of m, n, and o to comprehend colocalization.

Further, different confocal images (Fig. S14, ESI†) utilizing **DOX** and **SB4** were captured with colocalization comparison with DAPI to analyze and ultimately provide a quantitative contrast in a bar graph (Fig. 13) of mean fluorescence intensity using 'ImageJ' software.[75] The Fig. 13 illustrates a comparative view to recognize the unique quenching of the mean fluorescent intensity of **SB4** at a 24 h time-point for both Fadu and 4T1 cell lines.

2.5.5. Molecular docking. Molecular docking is a methodology applied to study the molecular behavior of target protein binding. Indeed, it is a tool that is used extensively in drug discovery. We have performed an initial inspection with our molecules, although further investigation is needed to be carried out in the future. The existing literature reports that Epidermal Growth Factor Receptor (EGFR) is believed to play an essential role in oral, oropharyngeal, and triple-negative breast cancer (TNBC).[76,77] Human epidermal growth factor receptor 2 (HER2) is a member of the epidermal growth factor receptor family having tyrosine kinase activity. Amplification or overexpression of HER2 occurs in approximately 15–30% of breast cancers and HER2 testing and HER2-directed therapies are recommended only in breast and gastroesophageal cancers.[78,79]

Here, the Protein Preparation Wizard module, LigPrep module and Glide module from Schrodinger suites 2022-1 were employed to prepare the crystal structures of protein obtained from the PDB (PDB IDs: 7JXH, 3RCD and 3PP0 for HER2 protein and 5X2C and 4I22 for EGFR protein) for ligand preparation and for performing molecular docking, respectively. The crystal structures were chosen based on the availability of inhibitor ligands at the tyrosine kinase binding site and their resolutions. For HER2/ErbB2 protein, the resolution of structures was: 3PP0 with 2.25 Å, 7JXH with 3.27 Å, and 3RCD with 3.21 Å. For EGFR protein, the resolution of structures was: 5X2C with 2.05 Å and 4I22 with 1.71 Å. Post protein preparation, the co-crystal ligands and inhibitor ligands were prepared using the LigPrep module. The crystal structures were analyzed to find the active site of the kinase domain where the inhibitor binds. For all the proteins, a 20 × 20 × 20 Å size grid was generated at the ligand binding site of the protein before docking, and the ligand was docked to the active site of all

Table 3 Interactions and scores obtained from the docking study

PDB ID	Crystal ligand interacting residues and Glide docking score	Indole compound interacting residues and Glide docking score
3PP0	Lys753, Leu796, Met801, Asp863, Phe864 −14.784	Arg811 −5.931
7JXH	Ser783, Cys805, Asp808 −10.752	Lys736 −6.058
3RCD	Lys753, Leu796, Met801, Asp863 −11.277	Phe1004 −5.597
5X2C	Met793 −9.854	NA −6.502
4I22	Leu788, Met793 −10.065	NA −7.519

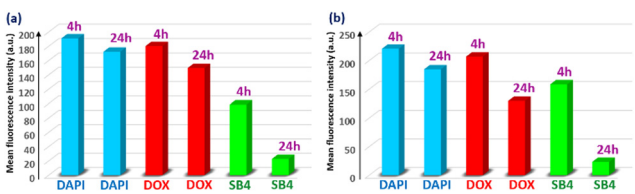

Fig. 13 Comparison of the mean fluorescence intensity in confocal images using 'ImageJ' for (a) FaDu and (b) 4T1.

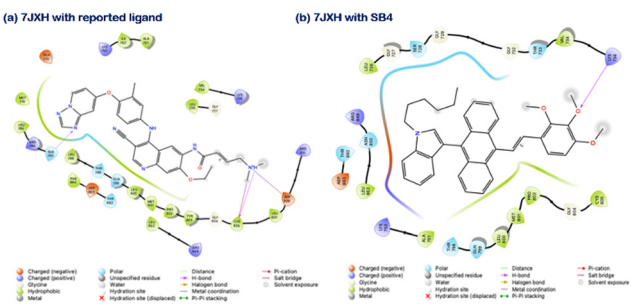

Fig. 14 2D view of interactions of (a) the reported ligand with the protein 7JXH and (b) **SB4** with the protein 7JXH.

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the proteins. Post-docking ligand protein interactions were analyzed. For the indole compound docked to the EGFR and HER2 proteins, the interacting amino acid residues are provided in Table 3. A comparison has been made with the interacting residues of the co-crystal ligands, suggesting the indole compound's affinity towards HER2 protein compared to EGFR protein (Fig. 14 and Fig. S15, ESI†). However, a further detailed investigation is highly needed for our future work.

3. Experimental

3.1. Materials and general conditions

All the syntheses were carried out in hot-air-oven-dried glassware. 1*H*-Indole, 1-bromohexane, 1-bromopentane, anthracene-9-carbaldehyde, diethyl phosphite, and piperazine were obtained from Alfa Aesar, Avra, and Merck and were utilized without further purification. Potassium hydroxide, potassium *tert*-butoxide, and methanesulfonic acid (MSA) were procured from Alfa Aesar and Sigma and were used as received. Dry DMSO, DCE, and THF were purchased from Finar. The rest of the chemicals were bought from Merck and Alfa Aesar and were used without any further purification. Spectroscopic grade solvents were used for UV-vis and fluorescence spectroscopy studies and procured from Sisco Research Laboratories (SRL). Column chromatography was performed by using silica gel (100–200 mesh), and the reactions were monitored *via* thin-layer chromatography on pre-coated silica gel 60 F254 plates (Merck & Co.) and observed using a 365 nm UV lamp. The NMR spectra were recorded at ambient temperature (*ca.* 20 °C) in $CDCl_3$ solution. The signal of TMS was considered as the reference one with the deuterated chloroform solvent peak as an internal standard (7.26 ppm for ^{1}H NMR, 77.0 ppm for ^{13}C NMR). Chemical shifts are reported here in ppm denoting s (singlet), d (doublet), dd (doublet of doublet), t (triplet), q (quintet), and m (multiplet). All the experiments were performed at room temperature (298 ± 2K). IR spectra (4000–400 cm^{-1}) were recorded using an FT/IR-4200 Jasco spectrometer (KBr pellets).

3.2. Steady-state absorption and fluorescence measurements

Solid-state absorption spectra were recorded using a JASCO-500 spectrophotometer, and the solution-state absorption spectra were recorded using a UV-vis-NIR spectrophotometer (Hitachi F7000, Japan). Solution-state emission spectra were obtained using an FP-6300 spectrometer (JASCO), using a 10 mm path-length quartz cuvette, while the solid-state emission spectra were recorded with a fluorimeter (Fluorolog, HORIBA). The emission spectra were recorded at the corresponding absorption wavelengths.

3.3. Absolute/relative quantum yield and lifetime decay measurement

For solid samples, the absolute quantum yield (φ_f) values were obtained by utilizing the calibrated integrating sphere method with a fluorimeter (Fluorolog, HORIBA). Absolute errors within ~± 2% are included. For the relative quantum yield (relative φ_f) in the solution state, the error is ±5%.

Time-resolved fluorescence measurements were performed using a time-correlated single-photon counting (TCSPC) unit (Horiba Deltaflex). The lasers used for all samples were 440 nm and 509 nm. All measurements were performed at room temperature. The decay fitting was completed, keeping the χ^2 value close to unity.

3.4. Molecular docking

As mentioned earlier, Schrodinger suites 2022-1 were employed to prepare the crystal structures of protein obtained from the PDB, for ligand preparation and performing molecular docking, respectively.

3.5. Cell-culture

The cell lines were purchased from NCCS, Pune, India and ATCC and maintained in DMEM supplemented with 10% FBS with 1% (v/v) penicillin-streptomycin. Cells were incubated at 37 °C in a humidified atmosphere comprising 05/95 CO_2/air. All culture supplies were procured from Thermo Fisher Scientific.

3.6. Cytotoxicity – MTT assay

All cells were seeded into 96 well plates at a density of 5×10^3 cells per well. After proper adhesion, cells were incubated with the compounds for 24 h. Cells treated with DMSO (less than 1%) were used as a control group. After 24 h, the supernatant from the plate was discarded and the cells were rinsed with 100 μL of sterile PBS. Freshly prepared 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) (MTT) (100 μL at 5 mg mL^{-1} in sterile PBS) was added. The 96 well plate was incubated at 37 °C for 4 h. The supernatant was removed after 4 h, and 100 μL per well DMSO was added to the 96 well plate. A multiplate reader was used to read absorbance at a wavelength of 570 nm. The anticancer activities of the synthesized indole-anthracene-based AIE-gens were evaluated against three cell lines: FaDu, which is a robust cell line, 4T1, a triple-negative breast cancer cell line, and MCF-7, which is a non-TNBC cell line, and one normal cell line: HEK 293, which is a human embryonic cell line using MTT assay. DOX is a clinical anticancer drug that was used as a positive control.

3.7. Cellular uptake (FACS)

Cells were used for quantitative analysis of the uptake of **SB4** using flow cytometry. Cell lines were seeded in a 12 well cell culture plate (1 × 106 cells per well) in DMEM (Dulbecco's Modified Eagle's Medium) containing 10% FBS with 1% antibiotics. After complete adhesion, cells were treated with different AIE-gens using their GI_{50} for 24 h. On the next day, cells were washed with sterile PBS followed by trypsinization of cells. Cells were collected in a 1.5 mL tube and centrifuged at 4 °C at 1000 rpm for 5 minutes. Cells were resuspended in 250 μL of sterile PBS (pH 7.4). A flow cytometer (BD FACS Aria™ III) was used for histogram generation of compound uptake.

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3.8. Reactive oxygen species (ROS) assay

FaDu cells were seeded in a 96 well plate using DMEM for 24 h. Later, cells were treated with the AIE-gens for 24 h. Cells were washed with PBS and incubated with a 10 μM 2′, 7′-Dichlorofluorescin diacetate (DCF-DA) solution for 30 min. The cells were washed 3 times with PBS and fluorescence was recorded using a Spectramax instrument at an excitation wavelength of 485 nm and an emission wavelength of 530 nm.

3.9. Haemolysis assay

Rat blood was collected and centrifuged at 2500 rpm for 10 minutes. Red blood cells (RBCs) were carefully separated from plasma and collected RBCs were washed with PBS and centrifuged at 3000 rpm for 10 minutes. RBCs were resuspended into PBS to obtain a working 5% v/v solution. Triton-X 100 (1%) was used as a positive control and the PBS treated sample was used as a negative control. Compounds were incubated at their GI_{50} concentration for 1 h and at 37 °C. Later, the RBC solution was centrifuged at 8000 rpm for 1 minute and the supernatant was collected for absorbance at 576 nm using a Spectramax™ multiplate reader (Molecular Devices, US). The formula used to calculate haemolysis (%) = $(A_s - A_0)/(A_{100} - A_0) \times 100$ where A_s is sample absorbance, A_0 is the negative control absorbance treated with PBS and A_{100} is (Absorbance of titron X-100).

3.10. Fixed cell confocal fluorescence imaging

Cell lines were seeded on a lysine coated sterile coverslip placed on a 12 well cell culture plate (1×10^5 cells per well) in DMEM containing 10% FBS with 1% antibiotics. Cells were allowed to adhere completely. After 24 h, cells were treated with different formulations using their GI_{50} for 4 h and 24 h. Later, cells were washed with sterile PBS and mounted on a glass slide. Cell imaging was done using a confocal microscope.

3.11. Synthetic routes

Synthesis of diethyl ((10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)methyl)phosphonate. Diethyl (anthracen-9-yl(hydroxy)-methyl)phosphonate (1.45 mmol, 1 equiv., 0.5 g) was put into one 100 mL round bottom flask and subjected to vacuum drying for 10 min. Next, under N_2, dichloroethane (15 mL) was poured into it and allowed to stir for 5 min in an ice bath. Later, methanesulphonic acid (MSA) (5.05 mmol, 3.48 equiv., 0.33 mL) was added under N_2 and allowed to stir for another 15 min. Next, the ice bath was taken away and the reaction mixture was brought back to room temperature and the solution turned maroon. Then, freshly prepared alkylated indole (2.59 mmol, 1.79 equiv., 0.52 g) was added slowly under N_2 and allowed to stir for 15 min until it changed from marron to green and finally to blue. The alkylation of indole was performed following a reported procedure.[45] Next, the mixture was heated under reflux for 3 h at 90 °C and then cooled down to room temperature. The completion of the reaction can be monitored by observing the formation of a cyan color fluorescent product spot at R_f = 0.17 while eluting the TLC (Thin Layer Chromatography) plate into hexane/EtOAc medium (40% EtOAc into hexane (v/v). The reaction mixture was subjected to work up with 30 mL of an EtOAc and brine water mixture of 1 : 2 EtOAc : H_2O. The maroon color organic layer was separated and made into a slurry with silica gel (60–120 mesh) to perform column chromatography (30% EtOAc into hexane (v/v)), which yielded (50%, 0.38 g) a gummy cyan fluorescent compound. IR (KBr, cm^{-1}): 3057, 2930, 2854, 2355, 1586, 1462, 1390, 1254, 1161, 1057, 1021, 964, 740. M.p.: 79–81 °C. NMR spectroscopy: ^{1}H NMR (400 MHz, $CDCl_3$, 25 °C, δ): 8.39 (d, J = 8.92 Hz, 2H), 7.92 (d, J = 8.76 Hz, 2H), 7.54–7.45 (m, 3H), 7.29–7.21 (m, 4H), 7.05–6.98 (m, 2H), 4.33–4.25 (m, 4H, PCH_2 doublet signals are merged), 4.01–3.85 (m, 4H), 2.00–1.93 (m, 2H), 1.42–1.26 (m, 6H), 1.12 (t, J = 7.04 Hz, 6H), 0.88 (t, J = 6.48 Hz, 3H). ^{13}C NMR (100 MHz, $CDCl_3$, 25 °C, δ): 136.1, 131.7, 130.8, 130.5, 129.8, 128.9, 128.3, 125.7, 125.1, 124.7, 123.3, 121.8, 120.5, 119.5, 111.9, 109.7, 62.3 (d, J= 6.84 Hz), 46.6, 31.5, 30.3, 27.3 (d, J = 140 Hz), 26.8, 22.6, 16.3 (d, J= 5.98 Hz), 14.0. ^{31}P NMR (162 MHz, $CDCl_3$, 25 °C, δ): 26.08. Mass spectrometry: HRMS (ESI) m/z: calcd for $C_{33}H_{38}NO_3P$ 527.2589, found: 528.2445 $[M + H]^+$.

Synthesis of 2-(pyridin-3-yl)benzaldehyde. 2-bromobenzaldehyde (1.08 mmol, 1 equiv., 0.2 g) and pyridin-3-ylboronic acid (2.16 mmol, 2 equiv., 0.27 g) were added into a two-neck round bottom flask and subjected to vacuum drying for 5 min. Later, under N_2, 15 mL of dioxane/water (9 : 1, (v/v)) was poured into it and stirred for 5 min again. Next, $Pd(PPh_3)_4$ (0.09 mmol, 0.08 equiv., 0.09 g), K_2CO_3 (1.63 mmol, 1.5 equiv., 0.22 g), and Tetrabutylammonium bromide (0.09 mmol, 0.08 equiv., 0.03 g) were added into the reaction solution, stirred for 15 min and the N_2 environment was maintained. Then, the reaction mixture was refluxed at 90 °C for 18 h. The progress of the reaction was monitored by eluting TLC into hexane/EtOAc (20% EtOAc in hexane (v/v)) medium to observe an intense product spot formed at R_f = 0.32. The reaction mixture was worked up with 30 mL of an EtOAc and brine water mixture of 1 : 2 EtOAc : H_2O. The organic layer was separated, passed through a Na_2SO_4 layer, and made into a slurry with silica gel (60–120 mesh) to perform column chromatography (12% EtOAc into hexane (v/v)), which yielded (81%, 0.16 g) a solid whitish compound. IR (KBr, cm^{-1}): 3055, 2898, 2768, 1677, 1599, 1469, 1252, 1191, 836. M.p.: 60–62 °C. NMR spectroscopy: ^{1}H NMR (400 MHz, $CDCl_3$, 25 °C, δ): 9.98 (s, 1H), 8.72–8.67 (m, 2H), 8.09–8.05 (t, J = 8.33 Hz, 1H), 7.75–7.56 (m, 3H), 7.46–7.29 (m, 2H). ^{13}C NMR (100 MHz, $CDCl_3$, 25 °C, δ): 191.3, 150.1, 149.3, 141.8, 137.2, 133.9, 133.8, 133.7, 131.0, 128.7, 128.5, 123.2. Mass spectrometry: LCMS (ESI) m/z: calcd for $C_{12}H_9NO$ 184.0684, found: 184.1500 $[M]^+$.

Synthesis of 5-(2-formyl phenyl)picolinonitrile. 5-bromopicolinonitrile (1.09 mmol, 1 equiv. 0.2 g) and (2-formyl phenyl)boronic acid (2.19 mmol, 2 equiv., 0.33 g) were put in a two-neck round bottom flask and subjected to vacuum drying for 5 min. Later, under N_2, 15 mL of dioxane/water (9 : 1, (v/v)) was poured into it and stirred for 5 min again. Next, $Pd(PPh_3)_4$ (0.09 mmol, 0.08 equiv., 0.1 g), K_2CO_3 (1.64 mmol, 1.5 equiv., 0.23 g), and Tetrabutylammonium bromide (0.09 mmol, 0.08 equiv., 0.03 g) were added into the reaction solution, stirred for

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15 min and the N_2 environment was maintained. Then, the reaction mixture was refluxed at 90 °C for 18 h. The progress of the reaction was monitored by eluting TLC into hexane/EtOAc (20% EtOAc into hexane (v/v)) medium to observe an intense product spot formed at R_f = 0.36. The reaction mixture was worked up with 30 mL of an EtOAc and brine water mixture of 1 : 2 EtOAc : H_2O. The organic layer was separated, passed through a Na_2SO_4 layer, and made into a slurry with silica gel (60–120 mesh) to perform column chromatography (12% EtOAc into hexane (v/v)), which yielded (71%, 0.16 g) a solid whitish compound. IR (KBr, cm^{-1}): 3040, 2925, 2782, 2232, 1688, 1595, 1462, 1260, 1197, 992. M.p.: 120–122 °C. NMR spectroscopy: ^{1}H NMR (400 MHz, $CDCl_3$, 25 °C, δ): 9.98 (s, 1H), 8.75–8.74 (m, 1H), 8.08–8.06 (d, J = 7.68 Hz, 1H), 7.87–7.65 (m, 4H), 7.42–7.39 (d, J = 7.58 Hz). ^{13}C NMR (100 MHz, $CDCl_3$, 25 °C, δ): 190.6, 151.3, 139.6, 137.8, 134.1, 133.9, 133.7, 133.2, 131.1, 130.7, 129.8, 127.8, 117.0. Mass spectrometry: LCMS (ESI) m/z: calcd for $C_{13}H_8N_2O$ 208.0637, found: 209.2000.$[M]^+$.

Synthesis of (*E*)-1-hexyl-3-(10-(2-(pyridin-3-yl)styryl)anthracen-9-yl)-1*H*-indole (SB1). Diethyl ((10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)methyl)phosphonate (0.37 mmol, 1 equiv., 0.2 g), 2-(pyridin-3-yl)benzaldehyde (0.82 mmol, 2.2 equiv., 0.15 g) and K^{t}OBu base (0.93 mmol, 2.5 equiv., 0.1 g) were put in a two-neck round bottom flask and subjected to vacuum drying for 5 min. Later, N_2 was purged throughout the flask and it was sealed with a silicon septum. On one end, a N_2-containing balloon was placed, and at the other neck, dry THF (15 mL) was injected into the flask that was set to stir for 4 h. The progress of the reaction was monitored by eluting TLC into hexane/EtOAc (20% EtOAc into hexane (v/v)) medium to observe an intense product spot formed at R_f = 0.35. The reaction mixture was worked up with 30 mL of an EtOAc and brine water mixture of 1 : 2 EtOAc : H_2O. The organic layer was separated, passed through a Na_2SO_4 layer, and made into a slurry with silica gel (60–120 mesh) to perform column chromatography (12% EtOAc into hexane (v/v)), which yielded (79%, 0.16 g) a maroon sticky solid compound under room light but a green fluorescent compound under a UV-365 nm bulb. IR (KBr, cm^{-1}): 3452, 3178, 2966, 2852, 1728, 1597, 1453, 1429, 1380, 1328, 1190, 1119, 1062, 1015. NMR spectroscopy: ^{1}H NMR (400 MHz, $CDCl_3$, 25 °C, δ): 8.73 (s, 1H), 8.53–8.51 (dd, J = 1.6 Hz, 1.6 Hz 1H), 8.35 (d, J = 8.76 Hz, 2H), 8.15–8.13 (d, J = 7.56 Hz, 1H), 7.99–7.89 (m, 3H), 7.79–7.76 (m, 1H), 7.61–7.39 (m, 6H), 7.30–7.25 (m, 5H), 7.11–6.89 (m, 3H), 4.29 (t, J = 7.04 Hz, 2H), 2.02–1.95 (quint, J =7.36 Hz, 2H), 1.39–1.28 (m, 6H), 0.89 (t, J = 7.02 Hz, 3H). ^{13}C NMR (100 MHz, $CDCl_3$, 25 °C, δ): 150.2, 148.3, 137.4, 137.1, 136.5, 136.1, 136.0, 135.4, 132.3, 131.5, 130.5, 130.3, 129.7, 129.5, 128.8, 128.6, 128.1, 127.9, 127.7, 126.5, 125.8, 125.2, 124.8, 123.0, 121.8, 120.6, 119.5, 111.8, 109.6, 46.6, 31.4, 29.7, 26.8, 22.6, 14.0. Mass spectrometry: HRMS (ESI) m/z: calcd for $C_{41}H_{36}N_2$ 556.2878, found: 557.2743 $[M + H]^+$.

Synthesis of (*E*)-5-(2-(2-(10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)vinyl)phenyl)picolinonitrile (SB2). Diethyl ((10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)methyl)phosphonate (0.37 mmol, 1 equiv., 0.2 g), 5-(2-formylphenyl)picolinonitrile (0.82 mmol, 2.2 equiv., 0.17 g) and K^{t}OBu base (0.93 mmol, 2.5 equiv., 0.1 g) were put in a two-neck round bottom flask and subjected to vacuum drying for 5 min. Later, N_2 was purged throughout the flask and it was sealed with a silicon septum. On one end, a N_2-containing balloon was placed; at the other neck, dry THF (15 mL) was injected into the flask and it was stirred for 4 h. The progress of the reaction was monitored by eluting TLC into hexane/EtOAc (20% EtOAc into hexane (v/v)) medium to observe an intense green fluorescent product spot formed at R_f = 0.27. The reaction mixture was worked up with 30 mL of an EtOAc and brine water mixture of 1 : 2 EtOAc : H_2O. The organic layer was separated, passed through a Na_2SO_4 layer, and made into a slurry with silica gel (60–120 mesh) to perform column chromatography (12% EtOAc into hexane (v/v)), which yielded (83%, 0.18 g) a solid yellowish compound under room light but a green fluorescent compound under a UV-365 nm bulb. IR (KBr, cm^{-1}): 3436, 3123, 2924, 2848, 1690, 1569, 1464, 1419, 1364, 1021, 963. M.p.: 168–170 °C. NMR spectroscopy: ^{1}H NMR (400 MHz, $CDCl_3$, 25 °C, δ): 8.67 (s, 1H), 8.33 (d, J = 8.76 Hz), 8.21–8.15 (m, 2H), 8.0–7.97 (m, 3H), 7.82–7.60 (m, 2H), 7.53–7.40 (m, 5H), 7.30–7.27 (m, 2H), 7.24 (s, 1H), 7.10–6.86 (m, 3H), 5.51 (s, 1H), 4.30 (t, J = 7.04 Hz, 2H), 1.98 (quint, J = 7.36 Hz, 2H), 1.45–1.26 (m, 6H), 0.89 (t, J = 7.1 Hz, 3H). ^{13}C NMR (100 MHz, $CDCl_3$, 25 °C, δ): 148.9, 148.1, 139.5, 138.3, 136.5, 136.2, 136.1, 135.1, 131.9, 131.5, 130.5, 130.4, 129.7, 129.6, 129.5, 129.3, 129.1, 128.9, 128.3, 128.2, 127.9, 126.7, 125.7, 125.3, 124.8, 121.9, 121.8, 120.6, 119.5, 109.6, 46.6, 31.4, 30.2, 26.8, 22.6, 13.9. Mass spectrometry: HRMS (ESI) m/z: calcd for $C_{42}H_{35}N_3$ 581.2831, found: 582.2720 $[M + H]^+$.

Synthesis of (*E*)-1-hexyl-3-(10-(4-(pyridin-2-yl)styryl)anthracen-9-yl)-1*H*-indole (SB3). Diethyl ((10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)methyl)phosphonate (0.37 mmol, 1 equiv., 0.2 g), 4-(pyridin-2-yl)benzaldehyde (0.82 mmol, 2.2 equiv., 0.15 g) and K^{t}OBu base (0.93 mmol, 2.5 equiv., 0.1 g) were put in a two-neck round bottom flask and subjected to vacuum drying for 5 min. Later, N_2 was purged throughout the flask and it was sealed with a silicon septum. At one end, a N_2-containing balloon was placed, and at the other neck, dry THF (15 mL) was injected into the flask that was set to stir for 5 h. The progress of the reaction was monitored by eluting TLC into hexane/EtOAc (12% EtOAc into hexane (v/v)) medium to observe an intense green fluorescent product spot formed at R_f = 0.29. The reaction mixture was worked up with 30 mL of an EtOAc and brine water mixture of 1 : 2 EtOAc : H_2O. The organic layer was separated, passed through a Na_2SO_4 layer, and made into a slurry with silica gel (60–120 mesh) to perform column chromatography (10% EtOAc into hexane (v/v)), which yielded (71%, 0.15 g) a solid yellowish compound under room light and a bright but slightly yellowish-green fluorescent compound under a UV-365 nm bulb. IR (KBr, cm^{-1}): 3445, 3048, 2944, 2849, 1580, 1467, 1431, 1345, 1322, 1233, 1147, 1017, 958. M.p.: 154–156 °C. NMR spectroscopy: ^{1}H NMR (400 MHz, $CDCl_3$, 25 °C, δ): 8.74 (s, 1H), 8.46 (d, J = 8.76 Hz, 2H), 8.13–7.93 (m, 5H), 7.82–7.75 (m, 4H), 7.53–7.43 (m, 3H), 7.32–7.23 (m, 5H), 7.14–7.01 (m, 3H), 4.30 (t, J = 7.04 Hz, 2H), 1.99 (quint, J = 7.32 Hz, 2H), 1.46–1.23 (m, 6H), 0.90 (t, J = 7.04 Hz, 3H). ^{13}C NMR

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(100 MHz, $CDCl_3$, 25 °C, δ): 156.9, 149.8, 138.8, 138.1, 136.9, 136.1, 132.4, 131.6, 130.2, 129.8, 129.7, 128.9, 127.9, 127.3, 127.2, 127.0, 126.2, 126.1, 125.3, 124.9, 122.2, 121.8, 120.6, 120.5, 119.5, 111.9, 109.6, 46.6, 31.5, 30.3, 26.8, 22.6, 14.0. Mass spectrometry: HRMS (ESI) *m*/*z*: calcd for $C_{41}H_{36}N_2$ 556.2878, found: 557.2674 $[M + H]^+$.

Synthesis of (*E*)-1-hexyl-3-(10-(2,3,4-trimethoxystyryl)anthracen-9-yl)-1*H*-indole (SB4). Diethyl ((10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)methyl)phosphonate (0.37 mmol, 1 equiv., 0.2 g), 2,3,4-trimethoxybenzaldehyde (0.82 mmol, 2.2 equiv., 0.16 g) and K^{t}OBu base (0.93 mmol, 2.5 equiv., 0.1 g) were put in a two-neck round bottom flask and subjected to vacuum drying for 5 min. Later, N_2 was purged throughout the flask and it was sealed with a silicon septum. On one end, a N_2-containing balloon was placed, and at the other neck, dry THF (15 mL) was injected into the flask and set to stir for 5 h. The progress of the reaction was monitored by eluting TLC into hexane/EtOAc (12% EtOAc into hexane (v/v)) medium to observe an intense green fluorescent product spot formed at R_f = 0.33. The reaction mixture was worked up with 30 mL of an EtOAc and brine water mixture of 1 : 2 EtOAc : H_2O. The organic layer was separated, passed through a Na_2SO_4 layer, and made into a slurry with silica gel (60–120 mesh) to perform column chromatography (10% EtOAc into hexane (v/v)), which yielded (75%, 0.16 g) a solid yellowish compound under room light but a green fluorescent compound under a UV-365 nm bulb. IR (KBr, cm^{-1}): 3447, 3401, 2959, 2934, 2854, 1596, 1465, 1378, 1324, 1287, 1091. M.p.: 125–127 °C. NMR spectroscopy: ^{1}H NMR (400 MHz, $CDCl_3$, 25 °C, δ): 8.47 (d, J = 8.72 Hz, 2H), 7.94–7.90 (m, 3H), 7.62–7.42 (m, 4H), 7.31–7.00 (m, 7H), 6.84 (d, J = 8.76 Hz, 1H), 4.31 (t, J = 7.06 Hz, 2H), 3.92 (s, 6H), 3.88 (s, 3H), 1.99 (quint, J = 7.36 Hz, 2H), 1.45–1.28 (m, 6H), 0.90 (t, J = 7.34 Hz, 3H). ^{13}C NMR (100 MHz, $CDCl_3$, 25 °C, δ): 153.6, 151.9, 142.7, 136.1, 133.3, 131.9, 131.6, 129.8, 129.7, 128.9, 127.8, 126.3, 125.0, 124.8, 124.7, 121.7, 121.1, 120.6, 119.4, 112.0, 109.6, 107.9, 61.4, 61.0, 56.2, 46.6, 31.5, 29.7, 26.8, 22.6, 14.0. Mass spectrometry: HRMS (ESI) *m*/*z*: calcd for $C_{39}H_{39}NO_3$ 569.2930, found: 570.2792 $[M + H]^+$.

Synthesis of (*E*)-1-hexyl-3-(10-(3,4,5-trimethoxystyryl)anthracen-9-yl)-1*H*-indole (SB5). Diethyl ((10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)methyl)phosphonate (0.37 mmol, 1 equiv., 0.2 g), 3,4,5-trimethoxybenzaldehyde (0.82 mmol, 2.2 equiv., 0.16 g) and K^{t}OBu base (0.93 mmol, 2.5 equiv., 0.1 g) were put in a two-neck round bottom flask and subjected to vacuum drying for 5 min. Later, N_2 was purged throughout the flask and it was sealed with a silicon septum. On one end, a N_2-containing balloon was placed, and at the other neck, dry THF (15 mL) was injected into the flask that was set to stir for 5 h. The progress of the reaction was monitored by eluting TLC into hexane/EtOAc (12% EtOAc into hexane (v/v)) medium to observe an intense green fluorescent product spot formed at R_f = 0.24. The reaction mixture was worked up with 30 mL of an EtOAc and brine water mixture of 1 : 2 EtOAc : H_2O. The organic layer was separated, passed through a Na_2SO_4 layer, and made into a slurry with silica gel (60–120 mesh) to perform column chromatography (10% EtOAc into hexane (v/v)), which yielded (77%, 0.17 g) a solid yellowish compound under room light but a green fluorescent compound under a UV-365 nm bulb. IR (KBr, cm^{-1}): 3417, 3064, 2955, 2930, 2359, 1580, 1504, 1461, 1417, 1319, 1231, 1128, 1013. M.p.: 108–110 °C. NMR spectroscopy: ^{1}H NMR (400 MHz, $CDCl_3$, 25 °C, δ): 8.43 (d, J = 8.76 Hz, 2H), 7.9–7.89 (m, 3H), 7.54–7.43 (m, 3H), 7.32–7.28 (m, 4H), 7.13–6.91 (m, 5H), 4.32 (t, J = 7.04 Hz, 2H), 3.99 (s, 6H), 3.94 (s, 3H), 2.00 (quint, J = 7.38 Hz, 2H), 1.45–1.24 (m, 6H), 0.91 (t, J = 7.1 Hz, 3H). ^{13}C NMR (100 MHz, $CDCl_3$, 25 °C, δ): 153.6, 138.3, 137.3, 136.1, 134.3, 133.2, 132.3, 131.6, 130.1, 129.8, 129.7, 128.9, 127.9, 126.1, 125.2, 124.8, 121.8, 120.6, 119.5, 111.9, 109.6, 103.7, 61.1, 56.3, 46.6, 31.5, 30.3, 26.8, 22.6, 14.0. Mass spectrometry: HRMS (ESI) *m*/*z*: calcd for $C_{39}H_{39}NO_3$ 569.2930, found: 570.2744 $[M + H]^+$.

Synthesis of (*E*)-4-(2-(10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)vinyl)-*N*,*N*-diphenylaniline (SB6). Diethyl ((10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)methyl)phosphonate (0.37 mmol, 1 equiv., 0.2 g), 4-(diphenylamino)benzaldehyde (0.82 mmol, 2.2 equiv., 0.22 g) and K^{t}OBu base (0.93 mmol, 2.5 equiv., 0.1 g) were put in a two-neck round bottom flask and subjected to vacuum drying for 5 min. Later, N_2 was purged throughout the flask and it was sealed with a silicon septum. On one end, a N_2-containing balloon was placed, and at the other neck, dry THF (15 mL) was injected into the flask that was set to stir for 5 h. The progress of the reaction was monitored by eluting TLC into hexane/EtOAc (8% EtOAc into hexane (v/v)) medium to observe an intense green fluorescent product spot formed at R_f = 0.38. The reaction mixture was worked up with 30 mL of an EtOAc and brine water mixture of 1 : 2 EtOAc : H_2O. The organic layer was separated, passed through a Na_2SO_4 layer, and made into a slurry with silica gel (60–120 mesh) to perform column chromatography (5% EtOAc into hexane (v/v)), which yielded (84%, 0.20 g) a solid greenish compound under room light and a bright green fluorescent compound under a UV-365 nm bulb. IR (KBr, cm^{-1}): 3029, 2928, 2860, 2351, 2323, 1588, 1505, 1494, 1326, 1280, 1173. M.p.: 190–192 °C. NMR spectroscopy: ^{1}H NMR (400 MHz, $CDCl_3$, 25 °C, δ): 8.46 (d, J = 8.72 Hz, 2H), 7.93–7.87 (m, 3H), 7.60–7.42 (m, 5H), 7.32–7.28 (m, 8H), 7.19–7.16 (m, 6H), 7.13–6.93 (m, 5H), 4.31 (t, J = 7.04 Hz, 2H), 1.99 (quint, J = 7.34 Hz, 2H), 1.49–1.26 (m, 6H), 0.90 (t, J = 7.12 Hz, 3H). ^{13}C NMR (100 MHz, $CDCl_3$, 25 °C, δ): 147.7, 147.6, 139.9, 136.8, 136.1, 132.8, 131.7, 131.6, 129.8, 129.7, 129.6, 129.3, 128.9, 127.9, 127.5, 126.2, 125.0, 124.8, 124.5, 123.8, 123.6, 123.1, 121.7, 120.6, 119.5, 113.1, 113.0, 112.9, 111.9, 109.6, 46.6, 31.5, 30.3, 26.8, 22.6, 14.0. Mass spectrometry: HRMS (ESI) *m*/*z*: calcd for $C_{48}H_{42}N_2$ 646.3348, found: 647.3156 $[M + H]^+$.

Synthesis of (*E*)-3-(2-(10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)vinyl)-10-pentyl-10*H*-phenothiazine (SB7). Diethyl ((10-(1-hexyl-1*H*-indol-3-yl)anthracen-9-yl)methyl)phosphonate (0.37 mmol, 1 equiv., 0.2 g), 10-pentyl-10*H*-phenothiazine-3-carbaldehyde (0.82 mmol, 2.2 equiv., 0.25 g) and K^{t}OBu base (0.93 mmol, 2.5 equiv., 0.1 g) were put in a two-neck round bottom flask and subjected to vacuum drying for 5 min. Later, N_2 was purged throughout the flask and it was sealed with a silicon septum. On one end, a N_2-containing balloon was placed, and at the other neck,

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dry THF (15 mL) was injected into the flask that was set to stir for 4 h. The progress of the reaction was monitored by eluting TLC into hexane/EtOAc (8% EtOAc into hexane (v/v)) medium to observe an intense green fluorescent product spot formed at R_f = 0.37. The reaction mixture was worked up with 30 mL of an EtOAc and brine water mixture of 1 : 2 EtOAc : H_2O. The organic layer was separated, passed through a Na_2SO_4 layer, and made into a slurry with silica gel (60–120 mesh) to perform column chromatography (5% EtOAc into hexane (v/v)), which yielded (63%, 0.19 g) a solid orange compound (it becomes solid after 2–3 days though initially may remain as gummy) under room light but an orange-red fluorescent compound under a UV-365 nm bulb. IR (KBr, cm^{-1}): 3441, 3061, 2953, 2927, 2856, 1573, 1463, 1334, 1247. M.p.: 119–121 °C. NMR spectroscopy: ^{1}H NMR (400 MHz, $CDCl_3$, 25 °C, δ): 8.41 (d, J = 8.76 Hz, 2H), 7.93–7.83 (m, 3H), 7.64–7.37 (m, 5H), 7.31–7.23 (m, 4H), 7.17–7.00 (m, 4H), 6.96–6.85 (m, 4H), 4.29 (t, J = 7 Hz, 2H), 3.87 (quint, J = 6.28 Hz, 2H), 1.99 (quint, J = 7.12 Hz, 2H), 1.86 (quint, J = 7.14 Hz, 2H), 1.45–1.26 (m, 10H), 0.94–0.86 (m, 6H). ^{13}C NMR (100 MHz, $CDCl_3$, 25 °C, δ): 145.1, 144.9, 136.1, 132.7, 132.0, 131.6, 129.9, 129.8, 129.7, 128.9, 127.9, 127.5, 127.3, 126.2, 125.9, 125.4, 125.1, 124.8, 124.4, 123.6, 122.5, 121.8, 120.6, 119.5, 115.9, 115.5, 115.4, 114.8, 111.9, 109.6, 47.9, 47.6, 46.6, 31.5, 30.3, 29.2, 26.8, 26.7, 22.7, 22.5, 14.1. Mass spectrometry: HRMS (ESI) m/z: calcd for $C_{47}H_{46}N_2S$ 670.3382, found: 671.3252 $[M + H]^+$.

Conclusions

In conclusion, this work introduces a new class of *N*-alkyl indole-linked anthracenyl twisted π-conjugates as efficient DSE-gens. All these emitters were tested in anticancer studies, especially with the aggressive and robust FaDu and TNBC 4T1 cell lines. Among all these molecules, the 2,3,4-TMB substituted probe SB4 exhibited potential and was a convincing AIE-active DSE-gen that not only inhibited both FaDu and 4T1 (not MCF-7 and HEK-293) at 24 h but could also detect cell death by its diminished fluorescence signal. The HEK-293 normal cells and cancer cells could be easily distinguished by comparing the intensity of intact fluorescence signals. The noncytotoxic nature, high ROS generation, and less haemolysis make SB4 a commendable biocompatible drug with imaging capability, even in comparison to DOX. The intrinsically anticancer indole and anthracene play a crucial role in providing anticancer features with their affinity to HER2 protein, as suggested by the primary docking studies. The hydrophobicity of the lead probe may have assisted in penetrating the cell membrane (validated by FACS) to form a closer interaction and provide an image of the cells with DSE properties. Hence, a new class of easily accessible indole-anthracenyl π-conjugate compounds offers a promising solution to kill cancer cells selectively and detect the damaged cells through diminished fluorescence signals. This research showcases the prospects of such newly designed DSE-gens in challenging cancer research. Finding a detailed mechanistic path to decipher the selectivity of 4T1 and modification of the probe to improve the sensitivity and specificity are currently underway in our laboratory.

Author contributions

SB: conceptualization of the whole work, planning, molecular structure designing, synthesis and characterization, steady-state and time-resolved photophysical studies, analysis of the TD-DFT optimized structures, assistance in confocal microscopy, confocal image quantification, collaborative discussions, writing the original manuscript; PS: cell culture, cytotoxicity, ROS, haemolysis, FACS, confocal microscopy and suggestions in manuscript editing; PC: molecular docking and analysis, manuscript editing and assistance in confocal image analysis; OK: time to time discussion in the biological part and manuscript revision; MC: overall supervision, scientific guidance, analysis through theoretical and experimental support, a detailed plan on manuscript presentation, revision, continuous discussions and monitoring of the work.

Conflicts of interest

There are no conflicts to declare.

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