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3 **Development of Transmission Raman Spectroscopy towards the In Line,**
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5 **High Throughput and Non-Destructive Quantitative Analysis of**
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7 **Pharmaceutical Solid Oral Dose**
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32
33 **Abstract**
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35 Transmission Raman spectroscopy (TRS) is a recently introduced analytical technique to
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37 pharmaceutical analysis permitting volumetric sampling by non-destructive means. Here we
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39 demonstrate experimentally, for the first time, the enhanced speed of quantification of
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41 pharmaceutical tablets by an order of magnitude compared with conventional TRS. This is
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43 achieved using an enhancing element, “photon diode”, avoiding the loss of laser photons at
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45 laser coupling interface. The proof-of-concept experiments were performed on a complex
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47 mixture consisting of 5 components (3 APIs and 2 excipients) with nominal concentrations
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49 ranging between 0.4 and 89 %. Acquisition times as short as 0.01 s were reached with
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51 satisfactory quantification accuracy for all the sample components. Results suggest that even
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53 faster sampling speeds would be achievable for components with stronger Raman scattering
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55 cross sections or higher laser powers. This major improvement in speed of volumetric analysis
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3 enables high throughput deployment of TRS for *in line* quality control applications within the
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5 batch or continuous manufacturing process and facilitating non-destructive analysis of large
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7 fractions.
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10 11 12 **1. Introduction**

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14 The most widely used analytical method for the determination of assay, potency or content
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16 uniformity of pharmaceutical solid oral dose forms is High Performance Liquid
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18 Chromatography (HPLC). Although accurate and highly sensitive, this technique has several
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20 disadvantages: it is destructive, requires sample preparation and consumables, analysis times
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22 are typically longer than the batch manufacturing time of the product and consequently has
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24 limited throughput. HPLC is therefore unsuitable for routine analysis of large proportions of
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26 manufactured batches.
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32 The advantages of sampling larger proportions of the total batch size include: meeting
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34 regulator demand, reduction in batch rejects and higher confidence that the release product is
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36 of correct parameters. Analysis has to be with a non-destructive technique otherwise the cost of
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38 analysis would out way these benefits.
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43 Transmission Near Infrared (NIR) spectroscopy is a non-destructive technique that can, in
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45 some cases, quantify active ingredient in tablets and capsules with adequate accuracy. Analysis
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47 times are, typically, in the orders of tens of seconds. Recently a new potent volumetric
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49 analytical tool has emerged in this area – Transmission Raman Spectroscopy (TRS)^{1,2,3}. To
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51 date it has been used in several studies performed *at line* (i.e. off line, next to the production
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53 line and separately from it) demonstrating capability to predict tablets and capsule components
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55 in quantitative manner with required accuracy.^{4,5,6,7,8} The method was shown to provide
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3 volumetric sampling capability and inherently considerably higher sampling speeds than NIR.
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5 Typical TRS sampling times demonstrated range from seconds to tens of seconds per tablet or
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7 capsule when deployed in conventional form (although faster speeds are achievable with
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9 strong Raman scatterers or at high concentrations).
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14 In this paper, we demonstrate, for the first time, the capability of TRS to provide an order of
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16 magnitude enhanced sampling speeds in *quantitative* TRS analysis than achievable with
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18 conventional TRS technique. This is accomplished using a recently introduced signal
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20 enhancing ‘photon diode’ that is capable of increasing Raman light throughput at around ten-
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22 fold in a passive manner.⁹ Although the signal enhancement characteristics have been
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24 described the technique has not been demonstrated in quantitative analysis. As the signal-to-
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26 noise ratio (SNR) presents one of the limiting factors of conventional TRS methods in rapid
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28 quantitative analysis, the signal enhancement due to the photon diode, which also leads directly
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30 to SNR improvement, has a potential to translate itself directly into the improved precision of
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32 quantification. This hypothesis has been experimentally tested in the research presented here.
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39 In essence, the photon diode is a ‘unidirectional’ mirror permitting the transfer of photons from
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41 one side and acting as a reflector for photons impacting on it from the other side⁹. It is located
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43 in close proximity to sample placed directly over the laser illumination zone to prevent the loss
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45 of diffusely scattered photons from sample surface. As this loss can typically be substantial
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47 (>90 % of photons can escape by this mechanism from sample) its prevention leads to much
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49 higher coupling efficiency of laser photons into the sample.
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54 The photon diode itself is fabricated as a multilayer dielectric optical bandpass or edge filter
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56 and has identical properties. Its unidirectional property stems from the generic angular
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properties of dielectric filters for which the spectral profile shifts to shorter wavelengths as photons impact them at greater angles further away from normal incidence. The substantially increased coupling of laser radiation naturally leads to greater Raman signal and improved SNR of the detected Raman spectrum. Figure 1.

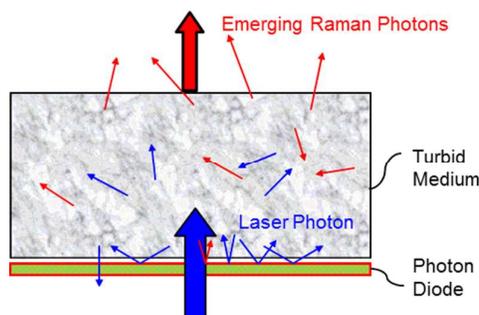


Figure 1: Schematic of a photon diode or ‘unidirectional mirror’. Tablet is placed directly on top of photon diode. Any laser photons which are re-radiated backwards from the tablet surface are reflected back into the sample. The net effect of increased laser photons into the sample results in an increase of emerging Raman photons.

An alternative enhancing concept to photon diode is a hemispherical mirror¹⁰ which too could be used to recycle diffusely reflected laser photons back into the sample to enhance Raman signal efficiency.

2. Experimental

The TRS spectra were collected using a commercial TRS instrument (TRS100, Cobalt Light Systems Ltd, Oxfordshire, UK). The system operates at 830 nm laser excitation wavelength.

The acquisition parameters employed a 4 mm laser illumination spot diameter and the power at sample was set to 200 mW. A Peltier cooled CCD detector was used to detect the Raman spectra (Andor Technology, DU420A-BR-DD, 1024x256 pixels, -75 °C). The acquisition time was varied systematically from 0.01 to 10 s. Multivariate modelling was performed using Solo (Eigenvector Research Inc., Wenatchee, WA, US).

The photon diode comprised of a 25 mm diameter dielectric bandpass filter (Iridian) centered at 830 nm with a bandwidth of 2.2 nm (FWHM) and transmission of >90% at the central wavelength.

Tablets were prepared from individual constituent powders weighed and mixed by hand, ground with a pestle and mortar and compressed using 8 mm flat surface tablet die in a bench top vice into two tablets per sample. The average mass of the tablets weighed ~200 mg with a range between 180 – 225 mg. The average thickness of the tablets measured 3.3 mm with a range 3.0 – 3.7 mm.

The basic mixture approximated commonly available over-the-counter cold & flu remedy tablets. It consisted of three active pharmaceutical ingredients (APIs): paracetamol (Sigma-Aldrich ~85 % w/w), caffeine (Sigma-Aldrich, ~4 % w/w), phenylephrine (Sigma-Aldrich ~1 % w/w); and two excipients: tablettose® [compressible lactose powder] (Meggler ~10 % w/w) and magnesium stearate (Sigma-Aldrich ~1% w/w).

The design of a calibration set followed a first-order (linear) 5 concentration level orthogonal design¹¹ uniformly covering the respective concentration ranges. Paracetamol, the bulk constituent, was used as the filler. Details of the individual 25 samples are shown in Table I.

Table I: Calibration DoE concentration ranges of components.

<i>Sample</i>	Phenylephrine %w/w	Caffeine %w/w	Mag Stearate %w/w	Tablettose %w/w	Paracetamol %w/w
1	1.22	5.19	1.29	10.03	82.27
2	1.18	1.36	0.46	8.19	88.81
3	0.51	1.50	2.28	9.55	86.15
4	0.39	9.07	0.95	7.49	82.10
5	1.94	3.26	1.88	13.43	79.48
6	0.96	8.42	1.28	7.98	81.36

7	2.27	5.01	0.93	10.99	80.80
8	1.31	3.33	0.84	9.29	85.23
9	0.78	3.55	1.63	10.46	83.57
10	0.86	7.09	2.20	11.97	77.88
11	1.67	8.57	1.71	12.24	75.80
12	2.10	6.91	1.37	13.52	76.10
13	1.73	5.26	2.21	15.48	75.32
14	1.37	8.49	2.20	11.46	76.49
15	2.03	8.86	0.50	10.26	78.35
16	2.02	1.73	1.67	11.68	82.90
17	0.56	6.71	0.49	5.36	86.88
18	1.70	2.33	1.33	11.59	83.06
19	0.50	5.43	1.57	8.98	83.52
20	1.32	6.71	1.77	10.94	79.27
21	1.73	6.82	0.84	8.54	82.08
22	1.74	3.65	0.55	8.53	85.53
23	0.87	1.78	0.80	7.24	89.31
24	0.42	3.39	1.19	5.55	89.44
Average	1.3	5.2	1.3	9.8	82.4
Min	0.4	1.4	0.5	5.2	75.3
Max	2.3	9.1	2.3	15.5	89.4

A total of 768 spectra were collected for analysis by scanning all 48 tablets from either face, being placed on photon diode band pass filter or on a bare silica plate and scanned at each of the four acquisition parameters detailed in Table II. (24 Samples \times 2 tablets \times 2 Faces \times 2 Filter options \times 4 methods = 768 Spectra). Note that tablets were not moved between changing the acquisition parameters to minimise variation in tablet placement.

Table II: Acquisition Parameters for each scan time.

Method	Laser power (mW)	Sub-exposure time (s)	Number of accumulations	Total exposure time (s)
1	200	0.01	1000	10.00
2	200	0.01	100	1.00
3	200	0.01	10	0.10

4	200	0.01	1	0.01
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3. Results and Discussion

Prior to the quantification measurements we first characterised the enhancement effect of photon diode across the sample range by comparing the average spectra of the entire data set, and its influence on the signal-to-noise ratio (SNR) in TRS spectra.

Figure 2 and Table III illustrates the Raman intensity enhancement effect attained using a photon diode compared to a tablet placed on a bare silica plate acting as a reference for the 1 second total scan time data set. An overall Raman signal intensity enhancement of approximately 10 has been achieved.

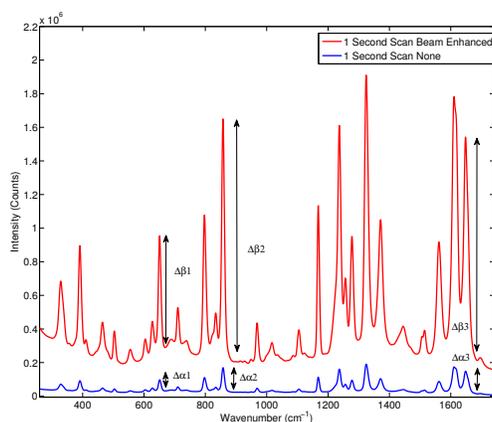


Figure 2 : Signal enhancement of the average spectra of the entire data set scanned for 1 second with (top trace) and without beam enhancer (bottom trace). Signal enhancement of 9.9 has been measured.

Table III: Beam Enhancement calculation (1 second scan time data).

Peak	Beam Enhanced $\Delta\beta$	None $\Delta\alpha$	$\Delta\beta/\Delta\alpha$
1	650901	64695	10.1

2	1417197	143841	9.9
3	1305761	133168	9.8
Average Enhancement			9.9

Figure 3 illustrates the improvement of SNR in TRS spectra upon the deployment of photon diode. The spectra are acquired under the same acquisition time (1 s). In the absence of any other source of noise the signal-to-noise ratio of TRS spectra is limited by the photon shot noise generated by the signal itself (i.e. Raman and any fluorescence background if present). The SNR is therefore theoretically expected to improve with the square root of enhancement factor, i.e. 3.3 times¹². From the statistical analysis of noise within the region 1730-1755 cm⁻¹ we estimate the measured improvement in signal to noise experimentally to be 5.6. A higher than theoretical value indicates an additional noise contribution originating possibly from the detection instrument (e.g. due to CCD detector read out noise). Similarly high SNR improvement factors with photon diode were estimated for the Raman spectra acquired at the other acquisition times.

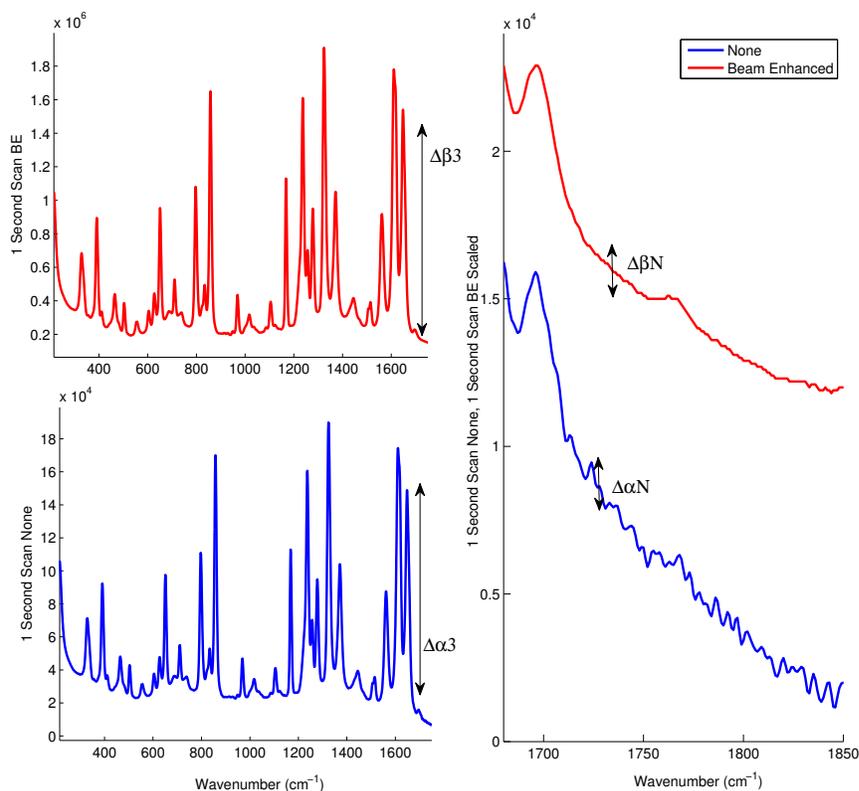


Figure 3: 1 s scan time. The Beam Enhanced spectra have been reduced and scaled to the same relative intensity as the 'None'. Signal-to-noise analysis of the TRS spectra with and without the photon diode was performed to illustrate the improvement of SNR accompanying the boost of the signal intensity with photon diode.

In the next series of experiments the sample set was subjected to TRS analysis performed with and without the photon diode. The tests were carried out separately at acquisition times of 10.00, 1.00, 0.10 and 0.01 s. Figure 4 shows the resulting TRS spectra of the average spectra for entire data set acquired at the different acquisition times, with and without photon diode. Visually the enhancement in signal to noise is uniform across each of the time points. Table IV summarises the enhancement factor and enhancement in SNR for each of the acquisition parameters. Statistically there is consistent enhancement in signal across the acquisition time points. The SNR improvement factor varied between 4.8 to 5.9.

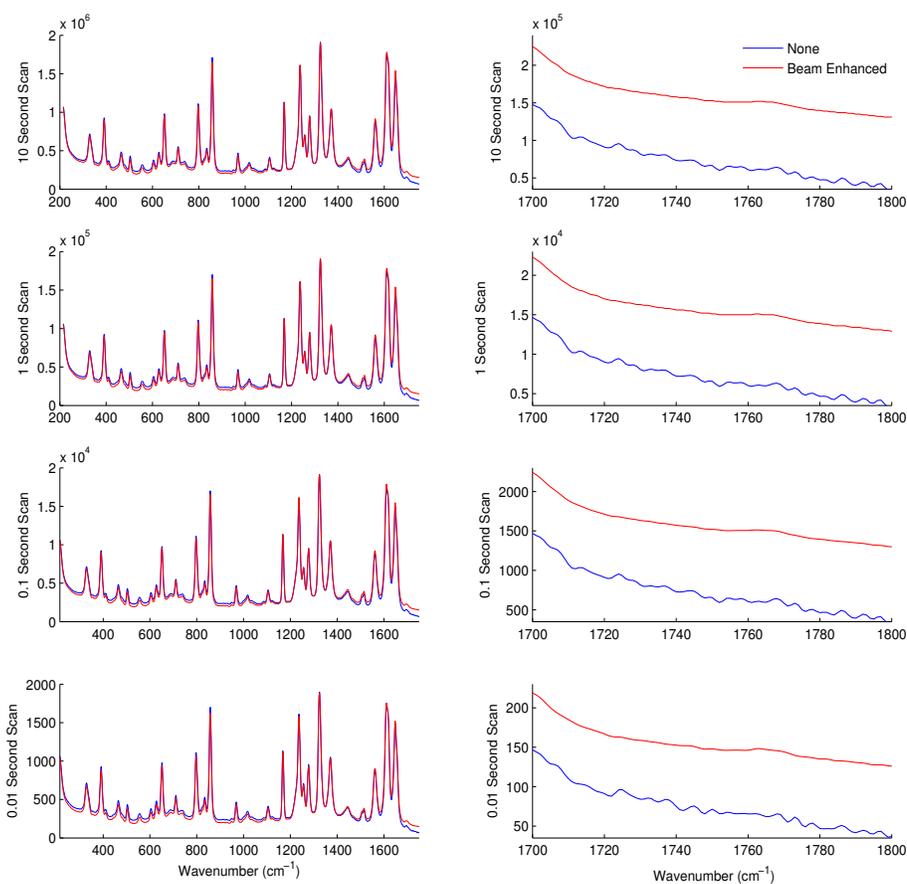


Figure 4: TRS spectra of the average spectra of the entire data set measured at four different acquisition times with and without photon diode. Beam enhanced spectra have been reduced in scale to the same relative intensity as 'None' to highlight enhancement in signal to noise ratios. Especially observable over the relatively blank regions 1700-1800 cm^{-1} .

Table IV: Signal to Noise Calculations for each of the acquisition parameters

	Signal Enhancement $\Delta\beta_3 \div \Delta\alpha_3$	Overall SNR Improvement $\text{SNR}_\beta \div \text{SNR}_\alpha$
10 Second Scan Time	10	4.8
1 Second Scan Time	10	5.6
0.1 Second Scan Time	10	5.9

0.01 Second Scan Time	10	4.9

Next we subjected the calibration data sets to PLS analysis and recovered RMSECV for each experimental configuration, acquisition parameters and beam enhancement. The results are depicted in Figure 5 and Figure 6. Further information can be found in the ESI.

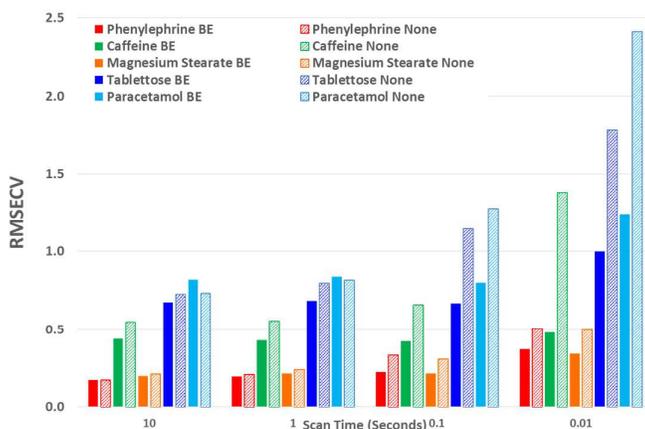


Figure 5: Graphical Summary of RMSECV Values.

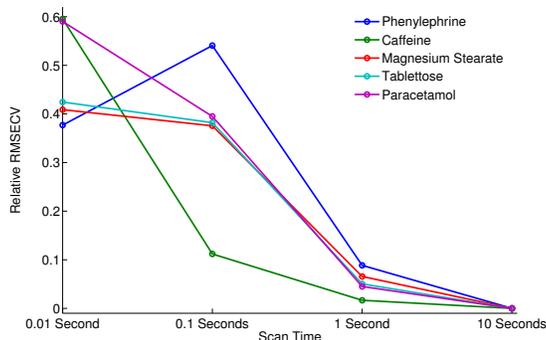


Figure 6: Relative improvement in RMSECV, scaled to the same Y axis for ease of comparison.

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3 From the analysis and consideration of the statistical outputs of the PLS models, R^2 , RMSECV
4 values and visualised fit and trending of these values (Figure 5 to Figure 6), we have found
5 consistently improved results were obtained with the photon diode permitting good
6 quantification performance to be attained down to a level of 0.01 s acquisition times. Overall
7 the use of photon diode alone clearly lowered the require acquisition times down
8 approximately by an order of magnitude for achieving the same level of quantification
9 accuracy as without the photon diode. This permitted 0.01s acquisition times per tablet to be
10 reached.
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22 Due to the sample set containing different constituents at varying concentrations we were able
23 to observe the improvement of the quantification of individual constituents at different
24 concentration levels as a function of scan time and beam enhancer. It would appear that the
25 improvement of the quantification of individual constituents due to the engagement of photon
26 diode vary differently, for instance it depends on the inherent intensity and Raman scattering
27 properties of the constituent and the relative proportion or concentration present in the sample.
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38 For example, caffeine gives an intense Raman signal and at a nominal concentration of 5%, the
39 use of the photon diode gives improved model statistics across all scan times. Conversely,
40 Paracetamol, although an excellent Raman scatterer, at 85% nominal concentration the beam
41 enhancer does not improve model statistics at longer scan times, conversely model statistics
42 slightly worsen. It could be suggested that under the experimental conditions, with these
43 particular samples we have reached the optimal model statistics for paracetamol, and no further
44 improvement can be made by scanning the samples for longer, or using beam enhancers. This
45 is presumably due to some other effect independent of SNR within Raman spectra, which
46 clamps the achievable RMSECV values at such longer acquisition times. We speculate that
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3 these clamping effects could for example be due to remaining heterogeneity of tablets not
4 removed fully by the TRS method that becomes a dominant factor below certain level of SNR
5 within spectra or due to residual errors in weighing and blending individual tablet components.
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11 Considering relative improvement in RMSECV values, caffeine relative improvement at 0.01 s
12 scan times is steep, suggesting that even faster scan times and relative improvements in
13 RMSECV can be achieved using the beam enhancer technology.
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19 Overall it would seem that beam enhancers have the greatest and most noticeable improvement
20 at short acquisition times. This is expected as SNR due to photon shot noise within spectra
21 becomes the limiting factor for achievable quantification in this regime in contrast with longer
22 acquisition times where other effects such as tablet heterogeneity or sample preparation
23 accuracy may be the limiting factors for reaching lower RMSECV values. The relative
24 enhancement is also dependent on the inherent nature of the constituent being analysed.
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36 **5. Conclusions**

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38 The study demonstrated the feasibility of dramatically shortening the acquisition times (by an
39 order of magnitude) by using a photon diode signal enhancing element. Acquisition times as
40 short as 0.01 s per tablet were sufficient to achieve a good degree of quantification with API at
41 concentrations ranging from 0.4 to 89%. Still higher sampling speeds would be achievable for
42 higher API concentrations or API with stronger Raman scattering cross sections than those
43 used here. The method paves the way for the next generation of TRS analytical tools
44 permitting the use of this technology *in line* in manufacturing environment for fast non-
45 destructive analysis of large portions of manufactured products than possible to date.
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References

1. B. Schrader and G. Bergmann, *Zeitschrift fur Analytische Chemie Fresenius*, 1967, **225**, 230-247.
2. P. Matousek and A.W. Parker, *Appl. Spectrosc.*, 2006, **60**, 1353-1357.
3. P. Matousek and A.W. Parker, *J. Raman Spectrosc.*, 2007, **38**, 563-567.
4. J. Johansson, A. Sparen, O. Svensson, S. Folestad and M. Claybourn, *Appl. Spectrosc.*, 2007, **61**, 1211-1218.
5. C. Eliasson, N.A. Macleod, L.C. Jayes, F.C. Clarke, S.V. Hammond, M.R. Smith and P. Matousek, *J. Pharm. Biomed. Anal.*, 2008, **47**, 221-229.
6. M.D. Hargreaves, N.A. Macleod, M.R. Smith, D. Andrews, S.V. Hammond and P. Matousek, *J. Pharm. Biomed. Anal.*, 2010, **54**, 463-468.
7. A. Aina, M.D. Hargreaves, P. Matousek and J.C. Burley, *Analyst*, 2010, **135**, 2328-2333.
8. M. Fransson, J. Johansson, A. Sparen and O. Svensson, *J. Chemometrics*, 2010, **24**, 674-680.
9. P. Matousek, *Appl. Spectrosc.*, 2007, **61**, 845-854.
10. M.J. Pelletier, *Appl. Spectrosc.*, 2013, **67**, 829-840.
11. Brereton R, *Analyst*, 1997, **122**, 1521-1529.
12. P. Matousek, M. Towrie and A.W. Parker, *J. Raman Spectrosc.*, 2002, **33**, 238-242.