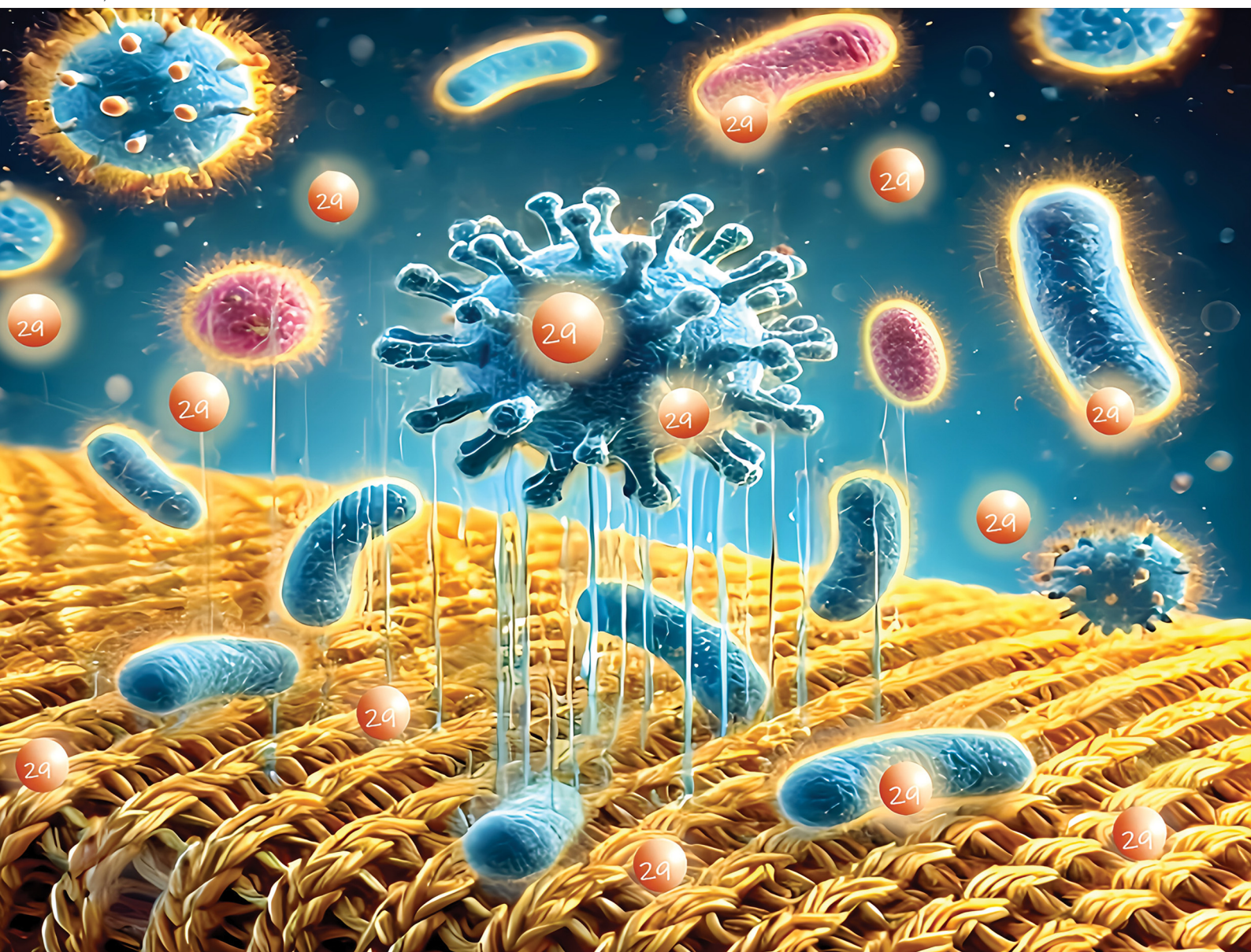


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PAPER

Chia-Ching Wu *et al.*
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6, 2507Industrially compatible manufacturing process of
wash-durable antimicrobial textiles using cuprous
oxide–polymer composites†Hung-Tung Chen,^a Ming-Cai Huang,^b Yi-Ying Chiang,^b Yong Chang^b and
Chia-Ching Wu  ^{★a}

Developing textiles with antibacterial and antiviral properties and excellent wash resistance can be a promising approach to combat multidrug-resistant bacteria, the influenza virus, and SARS-CoV-2, which are the major causes of nosocomial infections. Herein, an innovative method was developed to dope cuprous oxide nanoparticles (Cu₂O NPs) possessing antimicrobial properties into raw fiber materials, thus leading to the formation of Cu₂O NP-embedded masterbatches and then textiles. The textiles fabricated using the Cu₂O NP-embedded masterbatches exhibited excellent antimicrobial activity (>5.21; Japanese Industrial Standard L 1902:2015) against Gram-positive and Gram-negative bacteria and *Candida albicans*, and their antimicrobial activity slightly decreased by less than 5.9% after 50 washes. Thus, the Cu₂O NP-embedded textiles can effectively inhibit the growth of viruses and improve the efficacy of medical treatment. Our approach overcomes the drawback of the current techniques used for the post-processing of Cu₂O NP-related antimicrobial agents for textiles. In addition, Cu₂O NPs can be embedded into various raw materials used in the production of fabrics, such as polypropylene and polyethylene terephthalate, employing the developed technique. Furthermore, the developed approach can be readily commercialized.

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Introduction

Currently, textiles are widely used in various fields for multiple purposes. The textile industry has undergone significant advancements driven by technological innovation and increased environmental awareness. However, its primary goal remains to enhance human life. Consequently, textiles serve functions beyond protection. For instance, water-resistant jackets with high breathability can be both lightweight and warm.¹ Smart textiles integrate electronic components and sensing technologies to monitor physiological data.² Self-cleaning textiles, a type of functional material, can remove stains and odors without traditional washing.³ Moreover, textiles made from recycled polyester fibers, such as those extracted from plastic bottles, contribute to environmental sustainability.⁴

Since the outbreak of the COVID-19 pandemic, which triggered a global health crisis, national economies have been severely impacted, and public health systems have been strained. During this period, antimicrobial textiles garnered widespread attention and substantial research investment due to their ability to effectively inhibit bacterial and fungal growth.⁵

In healthcare settings, long-surviving pathogens can be transmitted through patients, healthcare personnel, and equipment, leading to hospital-acquired infections (HAIs). Highly contagious epidemic viruses, such as influenza, SARS-CoV, and SARS-CoV-2, can also cause hospital infections and community transmission. Thus, reducing the spread of viruses within healthcare facilities has become a critical focus of modern infection prevention strategies. In this case, antimicrobial agents are commonly applied to hospital textiles to mitigate infections. For example, treating hospital bed linens with antimicrobial agents can reduce bacterial colonization and curb the growth of pathogens such as methicillin-resistant *Staphylococcus aureus* (MRSA) and vancomycin-resistant *Enterococcus* (VRE), thereby decreasing HAIs.^{6–8} However, textiles require post-treatment with antimicrobial agents after each wash, making this method inconvenient and costly. As a result, extensive research has been conducted to develop

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antimicrobial textiles with durable properties. The use of antibacterial textiles can minimize microbial colonization, reducing the risk of infections from fabrics such as sheets, towels, and gowns. Incorporating copper oxide and copper composites into biocidal textiles for use in long-term care settings has been shown to significantly reduce HAIs, fever occurrences, antibiotic use, and medical resource consumption.^{8–10}

Key antimicrobial textile technologies include noble metal nanoparticle technology,¹¹ bio-based antimicrobial agents,¹² photocatalytic technology,¹³ antimicrobial coating technology,¹⁴ and smart antimicrobial textiles.¹⁵ Among them, noble metal nanoparticles have emerged as a highly effective and widely studied approach. Noble metals exert strong antimicrobial activity by disrupting microbial cell membranes, inhibiting enzymatic functions, and interfering with DNA replication, thereby exhibiting bactericidal effects.¹⁶

In noble metal nanoparticle technology, metals such as gold (Au), silver (Ag), and copper (Cu) are used to develop antimicrobial textiles.^{17–24} However, although silver nanoparticles have been extensively studied for their biocidal properties, few studies have explored the antimicrobial potential of Au and Cu. Gold nanoparticles have also attracted attention due to their unique physical and chemical properties. In this case, although Ag nanoparticles are more commonly used and typically exhibit stronger antimicrobial effects, gold nanoparticles possess notable antimicrobial activity. Additionally, Au nanoparticles have excellent biocompatibility and non-toxicity, making them suitable for antimicrobial textile applications.¹⁷ Copper, an essential trace element naturally present in plant and animal tissues, plays a crucial role in human metabolism. Moreover, Cu is significantly cheaper than Au and Ag, making it a cost-effective alternative for developing antimicrobial and antiviral textiles.

Recent studies have shown that different forms of Cu (*e.g.*, Cu(0), copper oxide, ionic copper, and copper-polymer composites) exhibit antimicrobial and antiviral properties, particularly at the nanoscale.²⁴ However, the underlying mechanisms for the antimicrobial and antiviral activity of Cu are not yet fully understood. It has been proposed that they involve the release of copper ions (Cu^{2+}), cell membrane damage, reactive oxygen species (ROS) generation, and bacterial metabolic disruption. A study demonstrated that coronaviruses survive on Cu surfaces for less than four hours, whereas they persist on plastic surfaces for over 72 hours.²⁵ Furthermore, Behzadinasab *et al.* reported that $\text{Cu}_2\text{O}/\text{PU}$ composite coatings significantly reduced the infectious titer of SARS-CoV-2, achieving a 99.9% reduction within one hour compared to the uncoated samples.²⁶ Consequently, Cu has regained attention due to its excellent antimicrobial characteristics. A review of clinical and animal studies confirmed that textiles containing copper oxide had no adverse effects on human skin.²⁷ Additionally, a 2009 report by the Environmental Protection Agency found no systemic toxicity or occupational risks associated with Cu exposure. The release of copper ions (Cu^{2+}) results in cell membrane damage catalyzed by reactive oxygen species (ROS), bacterial DNA degradation, metal-thiol interactions, and synergistic effects.^{28–40}

The functionalization of textiles with noble metals requires that their antimicrobial performance remains stable throughout their use, making it necessary to firmly embed them onto textiles. Various methods exist for incorporating noble metal nanoparticles into textiles, including surface coating (*e.g.*, dip-coating and spray-coating), *in situ* synthesis, and electrospinning. However, traditional coating techniques, such as spraying, result in Cu_2O nanoparticles being physically adsorbed onto textiles *via* van der Waals forces, leading to reduced antimicrobial effectiveness after washing. Similar results were observed when cotton fabrics were padded in salt noble metal solutions.⁴¹ A. Errokh *et al.* demonstrated that chemically immobilized Cu_2O nanoparticles could serve as sacrificial templates to anchor noble metals onto textiles.⁴² In the study by the Emama group, oxidizing agents such as hydrogen peroxide (H_2O_2) and sodium hydroxide (NaOH) were used to increase the content of carboxyl groups on cotton fabric surfaces. Although some CuO and TiO_2 were lost after multiple wash cycles, the antibacterial effect remained significant.^{43,44} Other techniques, such as grafting polymer brushes onto fabrics, enhanced the durability and maintained the long-term antibacterial efficacy of the fabrics.^{45,46} Additionally, cross-linkers improved the adhesion between noble metals and fabrics; however, their complex manufacturing processes limit their commercial feasibility.^{47,48} Researchers have also developed multifunctional antimicrobial textiles with properties such as dyeability, UV protection, water repellency, and photoluminescence, adding significant commercial value.^{49–56}

For noble metal-based antimicrobial textiles to achieve commercial viability, their antimicrobial efficacy must remain stable over repeated use. Unlike previous techniques that incorporate antimicrobial agents after fabric processing, this study focuses on embedding Cu_2O nanoparticles into masterbatches, specifically in their lower oxidation state. To the best of our knowledge, no previous research has explored the impregnation of Cu or its derivatives into masterbatches, followed by spinning and weaving them into textiles. We fabricated two types of masterbatches, Cu_2O NPs embedded in polypropylene (PP) and polyethylene terephthalate (PET). This approach prevented Cu_2O from detaching from the textile surface and inhibited its oxidation into CuO. Studies have shown that Cu_2O exhibits higher antimicrobial activity than CuO due to the greater release of Cu^+ ions.^{57–59} However, Cu_2O is prone to oxidation, which reduces its antimicrobial efficiency.

To validate our approach, we used Cu_2O NP-embedded PP masterbatches to produce melt-blown nonwoven fabric products. Our method aligns with conventional textile industry manufacturing techniques. The Cu_2O NP-PET melt-blown nonwoven fabrics fabricated using this technique exhibited excellent antimicrobial activity against both Gram-negative and Gram-positive bacteria, even after being stored for an extended period or undergoing 50 washing cycles. This durability is attributed to the antioxidant property of Cu_2O and its strong adhesion to PP. The high stability of Cu_2O NP-embedded PP for



antimicrobial melt-blown nonwoven fabrics has not been reported in previous Cu₂O-based research.

Experimental section

Materials and Cu₂O solution

Cu₂O powder was purchased from Taixing Smelting Plant (Cu₂O ≥ 98%, average diameter = 25 μm). Before use, the powder was dried at 150 °C overnight to remove absorbed moisture. The Cu₂O solution was prepared by dispersing 20% (by mass) of Cu₂O in water, using poly(propylene glycol) (purchased from EVONIK) as the dispersant. Polypropylene (PP) and polyethylene terephthalate (PET) powders, used as fiber raw materials, were obtained from Yuang Shen and David & Mike in Taiwan, respectively.

Synthesis of Cu₂O masterbatches

To ensure homogeneous mixing of the Cu₂O nanoparticles (NPs) with PP or PET powder, the Cu₂O solution was wet-milled using a NETZSCH LMZ10 system. This process refined the particles from 25 μm to the submicron scale. Subsequently, the refined Cu₂O powder was spray-dried at 150 °C. Next, a mixture of 12 wt% refined Cu₂O powder and 88 wt% PP or PET fiber raw material was prepared for masterbatch production. The mixture underwent compression, melting, extrusion, and pelletization using an extrusion-pelletizing machine from Kowin Recycling Machine, yielding Cu₂O NP-embedded masterbatches (Cu₂O NP-embedded PP or Cu₂O NP-embedded PET).

DLS and zeta potential measurements

The particle size distribution of the fine powders and the zeta potential of the Cu₂O NP-embedded PP masterbatches and nonwoven fabrics were analyzed using a Particulate Systems Nano Plus HD zeta/nanoparticle analyzer. In the dynamic light scattering (DLS) measurements, a 660-nm laser was directed at the prepared aqueous solution, and backscattered signals were collected using an avalanche photodiode detector at an angle of 165°. For the zeta potential measurements, the fabric samples were cut into 15 mm (*W*) × 35 mm (*L*) pieces. These samples were placed on a quartz crystal with an open side in the solid sample cell, where their zeta potential was measured. Subsequently, the quartz crystal was filled with reference polystyrene (PS) latex particles provided by Particulate Systems. A voltage was applied to the platinum electrodes, and the electroosmotic flow in the solid sample cell generated asymmetrical parabolic velocity profiles due to the charge differences on the upper and lower surfaces. These differences were analyzed using the Mori and Okamoto equation.⁶⁰ The apparent electrophoretic mobility at different cell positions was measured and converted into zeta potential using the Smoluchowski equation.⁶¹

Characterization

SEM images of the masterbatches and powder suspension were obtained using a Hitachi S-4800. The morphologies of Cu₂O

NP-PP and Cu₂O NP-PET meltblown nonwoven fabrics were characterized using a Thermo Scientific Phenom XL G2, which collects backscattered electrons and is equipped with an EDS system for elemental analysis. A TA instrument Q500 was used to perform TGA in a nitrogen atmosphere in the temperature range of 30 °C to 900 °C (heating rate: 10 °C min⁻¹). The crystallinity of the samples was examined using a Bruker D8 Discover diffractometer. XPS was performed using a Thermo Scientific K-Alpha X-ray photoelectron spectrometer with an Al Kα micro-focused monochromator as the radiation source. Before the measurement, a 6-nm surface of the PP masterbatch was sputtered with Ar ions to expose the embedded ingredients. The location of any color of textiles in the space was determined by its color coordinates; *L**, *a**, and *b**. The object was measured using the Datacolor® 500 family of benchtop spectrophotometers.

Evaluation of antimicrobial activity

The antimicrobial activity of the Cu₂O NP-embedded melt-blown nonwoven fabrics was assessed according to JIS L 1902:2015. The bacterial viability was evaluated using the pour plate method. The test organisms included Gram-negative bacteria (*Pseudomonas aeruginosa* ATCC 10145, *Escherichia coli* ATCC 8739, and *Klebsiella pneumoniae* ATCC 4352), Gram-positive bacteria (*Staphylococcus aureus* ATCC 6538P and methicillin-resistant *Staphylococcus aureus* (MRSA) ATCC 33591), and fungi (*Candida albicans* ATCC 10231). Briefly, 1 mL of bacterial culture grown in nutrient broth was spread onto an agar plate. The Cu₂O NP-embedded meltblown nonwoven fabrics were placed over the plates, while the control plates were left uncovered. The plates were incubated at 37 °C ± 1 °C for 18–24 h. The bacterial count (colony-forming units per milliliter, CFU per mL) was determined before and after incubation.

Laundrying

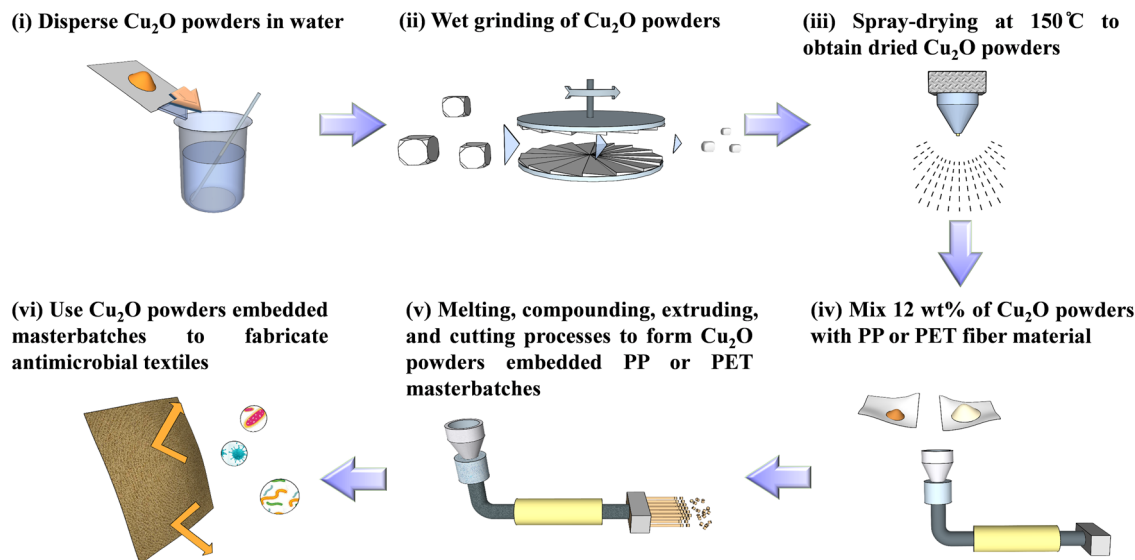
The antimicrobial activity of the Cu₂O NP-embedded melt-blown nonwoven fabrics after washing was evaluated following AATCC 135-2018, a standard established by the American Association of Textile Chemists and Colorists (AATCC). The fabrics were washed at 30 °C ± 3 °C and dried at temperatures below 60 °C for 50 cycles. After laundrying, antimicrobial activity tests were conducted to assess the retention of antibacterial properties.

Results and discussion

Synthesis of Cu₂O NP-embedded masterbatches and Cu₂O-embedded textiles

Scheme 1 illustrates the process for the preparation of Cu₂O NP-embedded masterbatches. Briefly, Cu₂O NP-embedded masterbatches were synthesized using a 20 wt% aqueous solution of Cu₂O NPs. The particles were wet-milled to obtain a fine powder suspension. The dynamic light scattering (DLS) analysis revealed that the hydrodynamic diameter of the fine powder





Scheme 1 Schematic of the preparation of Cu_2O NP-embedded masterbatches.

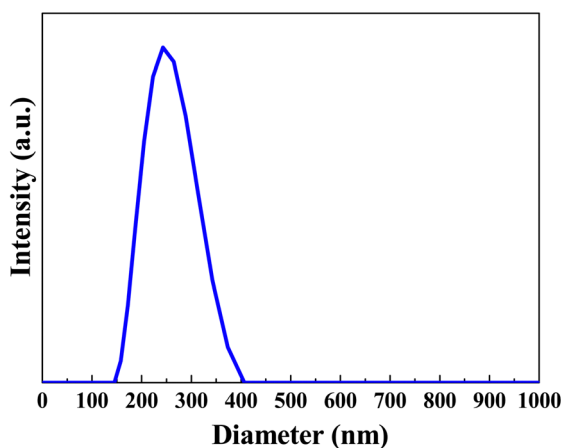


Fig. 1 Hydrodynamic diameter of Cu_2O NPs powders after the grinding process.

was 306 nm (Fig. 1). Subsequently, the suspension was spray-dried to yield dried powders, which were mixed with either PP or PET. Next, a continuous process involving compression, melting, extrusion, and cutting was employed to produce Cu_2O NP-embedded masterbatches (Fig. 2(a)). To fabricate yarns and textiles, 12 wt% Cu_2O NP-embedded (PP or PET) masterbatches were homogeneously blended with raw PP or PET masterbatches through melting and fusion processes. The thermogravimetric analysis (TGA) results for the Cu_2O NP-embedded PP masterbatches are presented in Fig. S1 (ESI[†]), demonstrating that the final degradation temperature and residual Cu_2O concentration were 467.7 °C and 11.33%, respectively. Fig. 2(b) shows a roll of the meltblown nonwoven fabric produced using the Cu_2O NP-embedded PP masterbatches (hereafter referred to as Cu_2O NP-PP meltblown nonwoven fabric), which can be utilized for fabricating face masks as one potential application (Fig. S2a, ESI[†]). Additionally, this

approach can be extended to produce various fiber-based materials and different types of Cu_2O NP-embedded textiles with antimicrobial properties. Fig. S2b and c (ESI[†]) illustrate the Cu_2O NP-embedded PET masterbatches and Cu_2O NP-embedded PET meltblown nonwoven fabric (hereafter referred to as Cu_2O NP-PET meltblown nonwoven fabric), respectively.

Morphology and crystallinity

Fig. 3(a), (b), and (d) present the scanning electron microscopy (SEM) images of the Cu_2O solution containing fine powders after grinding, Cu_2O NP-embedded PP masterbatches, and meltblown nonwoven fabric produced using Cu_2O NP-embedded PP masterbatches, respectively. As shown in Fig. 3(a), the average particle size of Cu_2O NPs after grinding was 306 nm. Fig. 3(b) shows the SEM image of the Cu_2O NP-embedded PP masterbatch, where the black regions correspond to PP, while the white and gray particles represent Cu_2O NPs. The Cu_2O NPs were observed to be uniformly distributed within the PP masterbatches. The chemical composition of the Cu_2O NP-embedded PP masterbatches was analyzed using energy-dispersive X-ray spectroscopy (EDS). Fig. 3(c) confirms the presence of Cu in the Cu_2O NP-embedded PP masterbatches. Fig. 3(d) illustrates the meltblown nonwoven fabric fabricated using the Cu_2O NP-embedded PP masterbatches. Additionally, Fig. S3 (ESI[†]) presents the elemental mapping of the meltblown nonwoven fabric produced from the Cu_2O NP-embedded PP masterbatches. As depicted in Fig. S3 (ESI[†]), carbon (C) exhibited the highest concentration in the meltblown nonwoven fabric, followed by oxygen (O), whereas the concentration of Cu was relatively low. This is because only 12 wt% of Cu_2O NPs was incorporated into the meltblown nonwoven fabric produced from the Cu_2O NP-embedded PP masterbatches. Fig. S4 (ESI[†]) further displays the meltblown nonwoven fabric fabricated using the Cu_2O NP-embedded PET masterbatches.



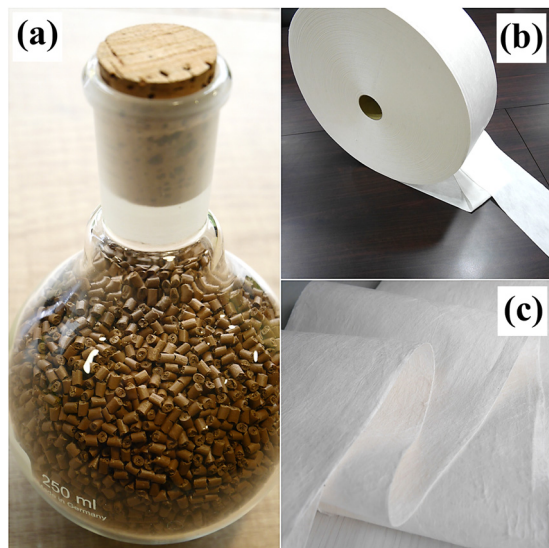


Fig. 2 Digital images of (a) Cu_2O NP-embedded PP masterbatches and (b) a roll of Cu_2O -PP meltblown nonwoven fabric. This fabric was used for the evaluation of antimicrobial activity in the present study. (c) Closer view of (b).

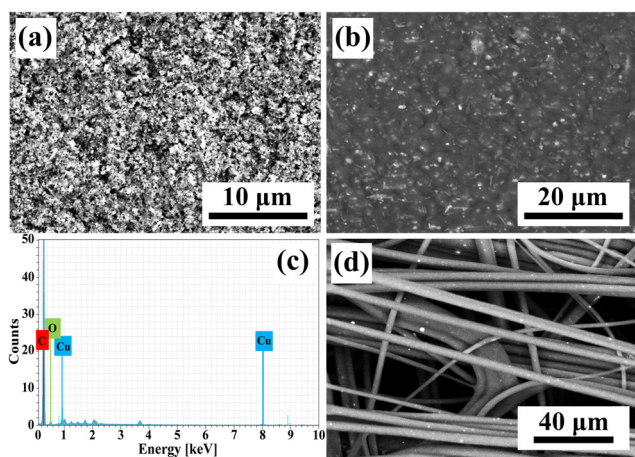


Fig. 3 SEM images of (a) fine powder suspension and (b) the Cu_2O NP-embedded PP masterbatch. (c) EDS spectrum of Cu_2O NP-PP meltblown nonwoven fabrics made from the Cu_2O NP-embedded PP masterbatch. (d) Cu_2O NP-PP meltblown nonwoven fabrics.

Fig. 4 shows the X-ray diffraction (XRD) patterns of the Cu_2O nanoparticles (NP) and Cu_2O NP-embedded polypropylene (PP) masterbatches. The diffraction peaks of Cu_2O NP at 2θ values of 36.5° , 42.7° , 47.1° , 61° , and 76.7° were assigned to the (111), (200), (211), (200), and (111) planes of Cu_2O , respectively (JCPDS no. 05-0667). No CuO phase, secondary phases, or unknown phases were observed, as shown in Fig. 4(a). Fig. 4(b) presents the XRD pattern of the Cu_2O NP-embedded PP masterbatches. The (131), (301), (060), and (220) planes of PP were observed at 2θ values of 21.1° , 21.8° , 25.4° , and 28.6° , respectively.⁶² The same diffraction peaks of Cu_2O NP at 2θ values of 36.5° , 42.7° , 47.1° , 61° , and 76.7° were assigned to the

(111), (200), (211), (200), and (111) planes, respectively. The very weak peaks at 36.1° and 38.9° were attributed to the (002) and (111) planes of CuO , respectively (JCPDS no. 48-1548). The XRD results suggest that slight oxidation of Cu_2O occurred during the fabrication of the masterbatches; however, this did not affect the antimicrobial properties of the textiles.

Chemical bonding state

The chemical composition of the Cu_2O NP-embedded polypropylene (PP) masterbatches was examined using X-ray photoelectron spectroscopy (XPS), as shown in Fig. 5. The survey spectrum in Fig. 5(a) revealed the presence of carbon (C), oxygen (O), and copper (Cu) elements, with the corresponding photoelectron peaks located at binding energies of 287 eV (C 1s), 531 eV (O 1s), 933.6 eV (Cu 2p), 76 eV (Cu 3p), and 122 eV (Cu 3s).

Fig. 5(b) shows the high-resolution scan of the Cu 2p core-level spectrum. The characteristic doublet peaks centered at 933.6 and 953.7 eV correspond to Cu $2p_{3/2}$ and Cu $2p_{1/2}$, respectively. Deconvolution of the Cu 2p electron binding energy revealed five peaks after Gaussian fitting at 932.7, 934.5, 947.6, 952.8, and 954.6 eV. The peaks at 932.7 and 952.8 eV were attributed to the Cu $2p_{3/2}$ and Cu $2p_{1/2}$ peaks of Cu^+ ions, confirming the presence of Cu_2O .⁶³ The weak peaks at 934.5 and 954.6 eV were assigned to the Cu $2p_{3/2}$ and Cu $2p_{1/2}$ peaks of Cu^{2+} ions, indicating the presence of CuO .^{64,65} The peak intensities for Cu_2O (932.7 and 952.8 eV) were higher than that for CuO (934.5 and 954.6 eV). Additionally, weak shake-up satellite bands were observed at 942–950 eV (with a peak at 947.6 eV), further suggesting the formation of Cu^{2+} ions.^{66–68} Shake-up bands occur when scattering photoelectrons interact with a valence electron, exciting it to a higher energy level. As a result, the kinetic energy of the core electron is slightly reduced, producing a satellite structure a few eV below the core-level position. The XPS results indicate that the Cu^+ ions have a filled electron configuration of $[\text{Ar}]3d^{10}$, while the Cu^{2+} ions have an unfilled $[\text{Ar}]3d^9$ configuration, with unpaired electrons in their d orbital. The photoelectrons collide with these d-electrons, exciting them to a higher energy level and reducing the kinetic energy of the 2p valence electrons. This interaction leads to the appearance of both a major XPS peak for Cu^{2+} ions and satellite peaks due to the interaction of the Cu 2p electron emission with the secondary excitation of outer valence electrons.⁶⁹ These XPS results demonstrate that Cu ions in the Cu_2O NP-embedded PP masterbatches exist in both the Cu^+ and Cu^{2+} chemical states. This Cu^+ and Cu^{2+} composition is likely due to the manufacturing process, which involves melting the PP fiber raw material at temperatures of around $250\text{--}260^\circ\text{C}$, leading to the slight oxidation of Cu_2O NPs.

To assess the stability of the Cu_2O NP-embedded PP masterbatch in air, it was exposed to air for 12 months, and subsequently examined using XPS (Fig. S5, ESI†). The peak intensity of $\text{Cu}^+/\text{Cu}^{2+}$ at 933.6 eV for the Cu_2O NP-embedded masterbatch exposed to air for 6 months was 1.17, which is similar to the peak intensity observed for the as-fabricated masterbatch that was not exposed to air. This result suggests



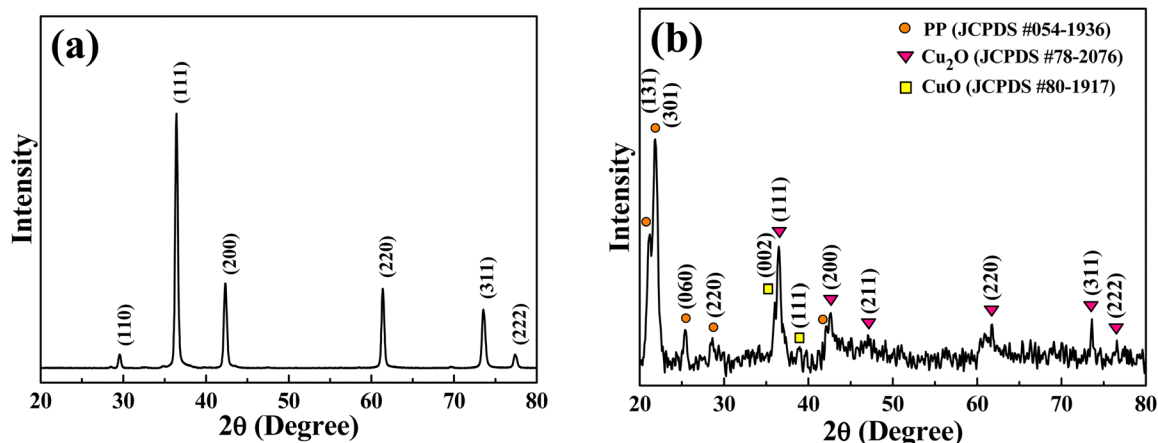


Fig. 4 X-ray diffraction patterns of (a) Cu_2O NP and (b) Cu_2O NP-embedded PP masterbatches.

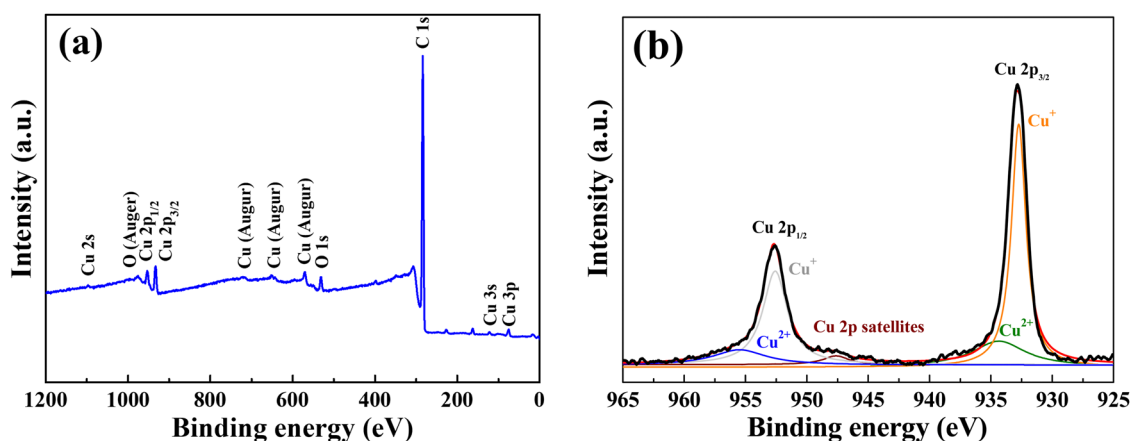


Fig. 5 XPS characterization of Cu_2O NP-embedded PP masterbatch. (a) Survey scan and (b) high-resolution Cu 2p spectra.

that the Cu_2O NPs in the PP polymer matrix were not further oxidized during the 6-month exposure to air. This stability can be attributed to the PP polymer, which isolates the Cu_2O NPs from moisture and oxygen in the air, preventing their further oxidation.

Color coordinates

To understand the effect of the incorporation of Cu_2O NPs on the dyeing performance of antimicrobial textiles, the samples were evaluated using the CIE $L^*a^*b^*$ (CIELAB) color space coordinates. The brightness value (L^*) of the pure PP textile was 94.31, with a^* and b^* values of 0.47 and 3.38, respectively. In the case of the Cu_2O NP-embedded PP textiles, their color coordinates were $L^* = 66.12$, $a^* = 1.92$, and $b^* = 18.2$, as shown in Table S1 (ESI[†]). Additionally, the K/S value of the pure PP textile was 0.143, while the value for the Cu_2O NP-embedded PP textile was 0.191.

Antimicrobial activity

The electrostatic interaction between positively charged metal ions and the negatively charged cell walls of bacteria can trigger

antimicrobial activity. It has been shown that the surface charge of silver (Ag) NPs significantly affects their antimicrobial properties. Positively charged Ag NPs exhibit a stronger antimicrobial effect than neutral or negatively charged Ag NPs. Thus, to investigate the surface charge of the fabrics prepared with Cu-embedded PP masterbatches, we measured the surface zeta potential of the meltblown nonwoven fabrics made with Cu_2O NP-embedded PP (Fig. S6, ESI[†]). The results revealed that the surface zeta potential of the meltblown nonwoven fabric was close to zero, indicating that it had a neutral surface, which resulted from the electrostatic treatment during the final spinning process.

To evaluate the antimicrobial activity of the meltblown nonwoven fabrics made with the Cu_2O NP-embedded PP masterbatches, tests were conducted according to the Japanese Industrial Standard (JIS) L 1902:2015. Five bacterial species were used to assess their antimicrobial activity, as shown in Fig. 6, including Gram-positive bacteria (*S. aureus* and MRSA), Gram-negative bacteria (*Escherichia coli*, *Pseudomonas aeruginosa*, and *Klebsiella pneumoniae*), and the fungus *Candida albicans*. The left column of Fig. 6 shows the bacterial count



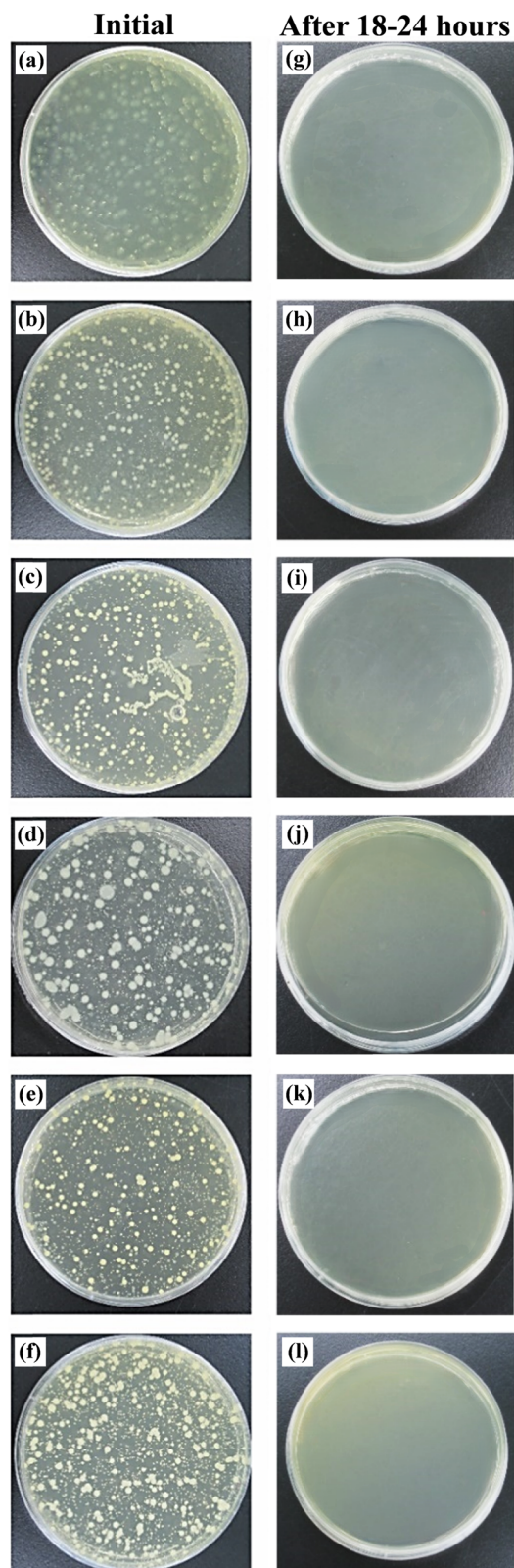


Fig. 6 Antimicrobial activity of Cu_2O NP-PP meltblown nonwoven fabrics against different species of microorganisms. The left column presents the pictures of the test just begun, and the right column depicts the results after 18–24 h. (a) and (g) *P. aeruginosa*. (b) and (h) *E. coli*. (c) and (i) Methicillin-resistant *Staphylococcus aureus* (MRSA). (d) and (j) *S. aureus*. (e) and (k) *K. pneumoniae*. (f) and (l) *C. albicans*.

(approximately 3×10^4 CFU per mL) immediately after inoculating the samples with the bacterial inoculum, while the right column shows the bacterial count after 18–24 h of incubation. Fewer than 20 colony-forming units per milliliter (CFU per mL) of Gram-positive bacteria, Gram-negative bacteria, and fungi were observed on the agar plates. The antibacterial activity (A) of the fabrics was calculated using the formula defined by the Japan Textile Evaluation Technology Council to evaluate the antimicrobial properties of textiles, as follows:

$$A = (\log C_t - \log C_0) - (\log T_t - \log T_0),$$

if $\log C_0 > \log T_0$, then $A = C_t - T_t$

where C_0 is the bacterial count observed immediately after inoculation on the standard fabric, C_t is the bacterial count observed after 18–24 h of incubation on the standard fabric, T_0 is the bacterial count noted immediately after inoculation on the fabrics made with Cu_2O NP-embedded PP masterbatches, and T_t is the bacterial count observed after 18–24 h of incubation on the fabrics made with Cu_2O NP-embedded PP masterbatches. A value greater than 3 indicates that the textile is highly effective against microorganisms. Fig. 7 shows a comparison of the antimicrobial activity of the standard fabric (control) with the meltblown nonwoven fabrics made with the Cu_2O NP-embedded PP masterbatches. The left column shows the results immediately after inoculation of the test bacteria, and the right column shows the results after 18–24 h of

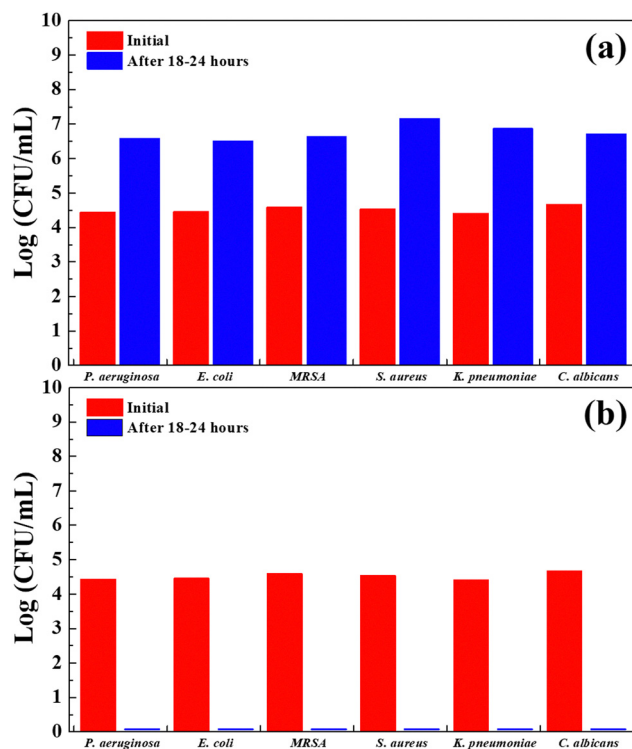


Fig. 7 Histograms representing the antimicrobial activity of Cu_2O NP-PP meltblown nonwoven fabrics against microorganisms. (a) Tests using the control. No Cu_2O NP-PP meltblown nonwoven fabric was placed in the agar plates in these tests. (b) Tests with the presence of Cu_2O NP-PP meltblown nonwoven fabrics.

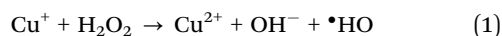


incubation. The results indicate that the meltblown nonwoven fabrics made with Cu₂O NP-embedded PP effectively inhibited bacterial growth.

The antimicrobial properties of Cu₂O NPs primarily rely on the release of Cu⁺ ions, which damage the cellular structure of microorganisms and disrupt their physiological functions. The antibacterial mechanism of Cu₂O NPs involves the release of free Cu ions, which catalyze the formation of reactive oxygen species (ROS) such as superoxide anions (O₂^{•−}), hydrogen peroxide (H₂O₂), hydroxyl radicals (•OH), and organic hydroperoxides in a Fenton-like reaction. These ROS damage cellular components and penetrate bacterial cells, leading to microbial cell death.^{29–32}

Given that Gram-positive bacteria (*S. aureus* and MRSA) lack a thick outer membrane that acts as a barrier to reactive oxygen species (ROS), the hydroxyl radicals (•OH) generated by Cu ions cause structural changes in phospholipids and disrupt the membrane integrity. Cu⁺ ions can pass through the cell membrane, leading to DNA degradation.

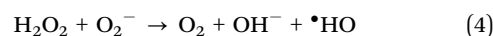
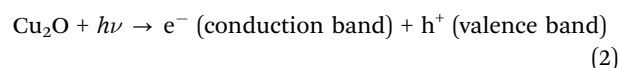
Fig. 8 presents a proposed model illustrating the antibacterial mechanisms of the meltblown nonwoven fabrics prepared using Cu₂O NP-embedded PP masterbatches when exposed to Gram-positive bacteria. Firstly, the Fenton reaction (eqn (1)) occurs, where Cu⁺ ions react with hydrogen peroxide (H₂O₂) to produce hydroxyl radicals, which degrade the peptidoglycan layer of the bacterial cell. In this process, Cu⁺ ions are released from the Cu₂O NP-embedded PP masterbatch.



Next, Cu⁺ ions enter the periplasmic space, separating the cytosol from the plasma membrane, which leads to membrane damage. This process follows one of the following pathways: (A) → (B), (A) → (B) → (C) → (b), or (A) → (B) → (C) → (c).

Following this, Cu⁺ ions penetrate the cell membrane, catalyzing ROS formation, which ultimately leads to DNA degradation. This pathway follows one of the following mechanisms: (a) → (b) or (a) → (c).

An additional minor antibacterial mechanism of Cu₂O NPs involves a photocatalytic reaction. Cu₂O has a direct bandgap of 2.1 eV. When visible light ($\lambda > 410$ nm) strikes the surface of Cu₂O NPs, an electron can jump from the valence band to the conduction band, as shown in eqn (2). Subsequently, the generated electrons are scavenged by molecular oxygen to form superoxide anions (O₂^{•−}), as shown in eqn (3). O₂^{•−} reacts with H₂O₂ to produce hydroxyl radicals (•HO), as shown in eqn (4). These hydroxyl radicals are the key oxidizing agents that damage bacterial cells.



Furthermore, the electrons generated in this reaction are paired, creating free radicals. These free radicals scavenge other electrons within the bacterial cells, pairing with them and causing damage to the cell wall, DNA, and proteins.

The mechanism by which Cu inhibits the growth of *E. coli* involves hydroxyl radicals induced by Cu ions, which drive the nonenzymatic peroxidation of unsaturated fatty acids in the bacterial cell membrane. This reaction initiates a chain process that leads to structural changes in phospholipids and disrupts the membrane integrity.⁶⁹ A similar mechanism is involved in the Cu-mediated inhibition of *P. aeruginosa* and *K. pneumoniae* growth. Fig. 9 illustrates how the Cu₂O NP-PP meltblown nonwoven fabrics inhibit the growth of Gram-negative bacteria.

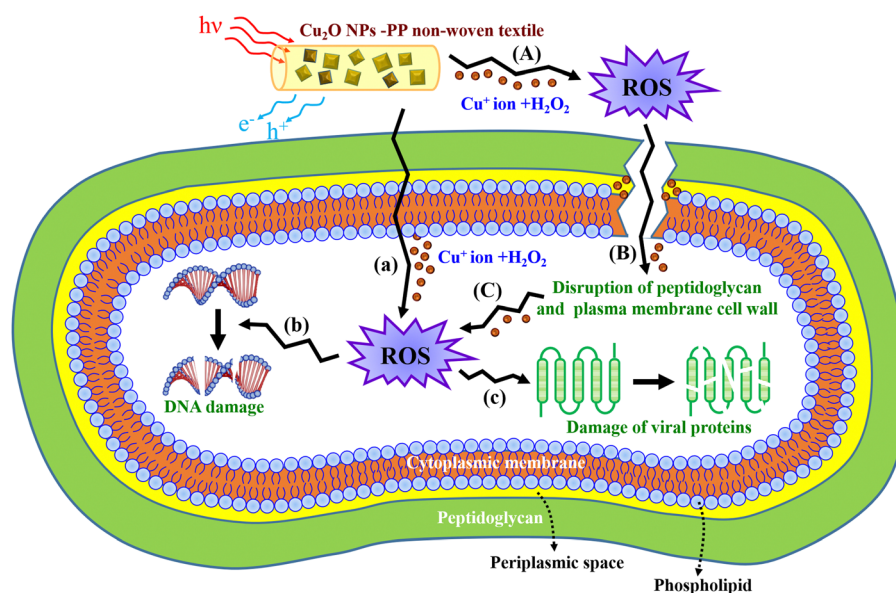


Fig. 8 Schematic of the probable antimicrobial mechanism of Cu₂O NP-PP meltblown nonwoven fabrics against Gram-positive bacteria.



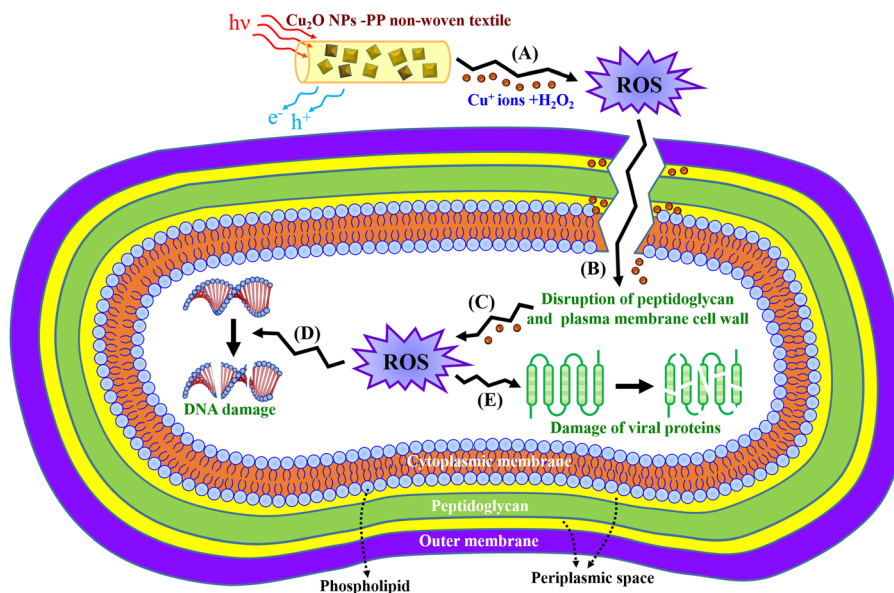


Fig. 9 Schematic of the probable antimicrobial mechanism of Cu_2O NP-PP meltblown nonwoven fabrics against Gram-negative bacteria.

Gram-negative bacteria have a three-layered cell envelope, consisting of an outer membrane, peptidoglycan, plasma membrane, and periplasmic space. Cu^+ ions react with hydrogen peroxide (H_2O_2) to produce hydroxyl radicals, which break down the peptidoglycan layer. Following this, Cu^+ ions enter the periplasmic space, separating the cytosol from the plasma membrane, which leads to membrane damage. Some Cu^+ ions penetrate the cell and catalyze the formation of reactive oxygen species (ROS), which damage DNA and proteins. The pathway for this mechanism can be described as follows: (A) \rightarrow (B), (A) \rightarrow (B) \rightarrow (C) \rightarrow (D), or (A) \rightarrow (B) \rightarrow (C) \rightarrow (E).

The antibacterial effect of Cu_2O NPs also involves a photocatalytic reaction. Compared to Gram-positive bacteria, Gram-negative bacteria exhibit slight resistance to ROS, which may influence the antibacterial mechanisms in these bacteria. Therefore, the specific mechanism(s) responsible for the antibacterial effects of Cu may vary between Gram-positive and Gram-negative bacteria. However, it remains unclear whether a single mechanism or a combination of mechanisms is responsible for the overall antibacterial effect.

C. albicans, a dimorphic fungus, can cause several infections, ranging from mild, superficial cutaneous infections to severe invasive candidiasis, which may lead to disability and mortality in immunocompromised patients. As shown in Fig. 6(f) and (i), we observed circular, white cells of *C. albicans*. White cells are epigenetically distinct from opaque cells, and although they are less efficient at mating, they have been shown to cause more virulent systemic infections.^{70,71}

The mechanism by which Cu_2O NP-PP meltblown nonwoven fabrics inhibit the growth of *C. albicans* is as follows: the nonwoven textile prepared using the Cu_2O NP-embedded PP masterbatches releases Cu^+ ions, which trigger the production of significant amounts of reactive oxygen species (ROS) in *C. albicans* through Fenton-like reactions. In the presence of H_2O_2 ,

Cu^+ ions oxidize various cellular substrates, causing cellular damage in a typical Fenton-like reaction, similar to the behavior of iron.⁷² Moreover, the level of membrane damage induced by Cu^+ ions was significantly higher than that caused by Cu^{2+} ions. This can be attributed to the fact that Cu_2O NP-PP meltblown nonwoven fabrics continue to release Cu^+ ions over time after contact with *C. albicans*. The high positive surface charge of Cu^+ ions may contribute to membrane damage. Given that *C. albicans* cells are electronegative, their cell walls can effectively bind with positively charged inorganic ions, potentially facilitating electrostatic interactions with Cu^+ ions. This interaction may play a significant role in the cellular damage of *C. albicans*.⁷³

The antibacterial effect of noble metals on Gram-positive and Gram-negative bacteria varies due to differences in their cell morphology. A recent study showed that Cu_2O NPs are more effective against Gram-positive bacteria than Gram-negative bacteria, which can be attributed to the differences in their cell wall structures. Gram-negative bacteria have a three-layered cell envelope, with a thick outer membrane that acts as a barrier to reactive oxygen species (ROS) generated through the Fenton reaction. Ren *et al.* observed that Cu^+ ions exhibited stronger antibacterial activity against *S. aureus* compared to *E. coli*. They attributed this result to the higher isoelectric point of the membranes of Gram-positive bacteria at the same pH, which creates a more negatively charged surface. This enhanced negative charge facilitates stronger interaction with metallic ions, thereby more effectively inhibiting *S. aureus*.^{74,75}

Table 1 summarizes the antibacterial activity of the fabrics prepared using Cu_2O NP-embedded PP masterbatches against the selected bacteria and fungus. The Cu_2O NP-PP meltblown nonwoven fabric exhibited an antibacterial activity value of 5.21 against *Escherichia coli* after 18–24 h of contact, as shown in



Table 1 Antimicrobial activities of Cu₂O NP-PP meltblown nonwoven fabrics at the contact times of 18–24 h

Bacteria species	Antimicrobial activity (As-fabricated)	Antimicrobial activity (One year later)
<i>P. aeruginosa</i> (ATCC 10145)	5.24	5.19
<i>E. coli</i> (ATCC 8739)	5.21	5.15
MRSA (ATCC 33591)	5.33	5.27
<i>K. pneumonia</i> (ATCC 4352)	5.57	5.51
<i>S. aureus</i> (ATCC 6538P)	5.86	5.79
<i>C. albicans</i> (ATCC 10231)	5.38	5.32

Table 1 and Table S2 (ESI[†]). Additionally, all the activity values were in the range of 5–6, indicating that the Cu₂O NP-PP meltblown nonwoven fabrics demonstrated excellent antimicrobial activity against all the selected microorganisms. Our results showed no significant difference in the effectiveness of the Cu₂O NP-PP meltblown nonwoven fabrics against Gram-positive and Gram-negative bacteria after 18–24 h of contact. The similar effectiveness against both types of bacteria can be attributed to the relatively long contact time used in our experiments compared to previous studies. To further test our hypothesis, we examined the antibacterial activity of the Cu₂O NP-PP meltblown nonwoven fabrics against *S. aureus*, *E. coli*, and *P. aeruginosa* with contact times of 1, 2, and 4 h. As shown in Fig. S7 (ESI[†]), the Cu₂O NP-PP meltblown nonwoven fabrics more effectively inhibited *S. aureus* than *E. coli* and *P. aeruginosa* when the contact time was under 2 h. Significant antimicrobial activity was observed against *S. aureus*, but not against the other bacteria within the first 2 h. This could be because the cell membrane of Gram-positive bacteria is thinner than that of Gram-negative bacteria, allowing Cu⁺ ions to penetrate more easily without causing immediate damage. Subsequently, Cu⁺ ions catalyze the formation of ROS, which damages the DNA of the cell. The antimicrobial mechanism follows path (a) → (b) or (a) → (c), as shown in Fig. 8. However, Cu⁺ ions require more time to penetrate and damage the cell membrane of Gram-negative bacteria, which have a more complex, three-layered cell envelope. Our results suggest that Gram-negative bacteria exhibit stronger resistance to Cu₂O NPs during shorter contact times. Furthermore, we tested the antibacterial activity of the Cu₂O NP-PP melt-blown nonwoven fabric after 12 months of storage. The results showed that the antibacterial activity slightly decreased compared to the freshly prepared Cu₂O NP-PP melt-blown nonwoven fabric. However, the antibacterial activity remained above 5, indicating that the fabric still exhibited good antibacterial properties after one year. The main reason for this result is that we embedded the Cu₂O NPs into the PP polymer, which protected the Cu₂O NPs from environmental factors and prevented their oxidation. The detail parameters are shown in Table S3 (ESI[†]). These results are generally consistent with the XPS result, as shown in Fig. S5 (ESI[†]).

The relatively longer time required for effective antimicrobial activity against Gram-negative bacteria, compared to Gram-positive bacteria can be attributed to several factors. In addition

to the thicker cell membrane of Gram-negative bacteria, the surface of the Cu₂O NP-PP meltblown nonwoven fabrics was found to be electrostatically neutral, as shown by the zeta potential measurements. As a result, the nonwoven fabric prepared with the Cu₂O NP-embedded PP masterbatches did not exhibit immediate antimicrobial activity upon direct contact with bacteria. However, after a certain period of contact, the fabric could release copper or cuprous ions. The interaction of these positively charged ions with the negatively charged bacterial surface could inhibit bacterial growth. The primary antimicrobial mechanism for long contact times with Gram-negative bacteria follows path (A) → (B) → (C) → (D), or (A) → (B) → (C) → (E). The antimicrobial mechanism for Gram-positive bacteria is similar but occurs along paths (A) → (B) → (C) → (b) and (A) → (B) → (C) → (c). Thus, to enhance the antimicrobial activity against Gram-negative bacteria, the concentration of Cu₂O NPs in the fabric should be increased in future applications.

Washing resistance is crucial for the commercialization of antimicrobial textiles using noble metals as antimicrobial agents. However, the coating processes commonly used in the textile industry, such as spraying and dipping, weakly bond Cu to fabrics. Additionally, these post-processing methods generate a large amount of wastewater, which is harmful to the environment. Furthermore, another approach, which involves using bridging agents to improve the Cu-fabric affinity, may not be universally applicable to all types of fibers. In contrast, our approach, where Cu₂O NPs are directly embedded into masterbatches, provides satisfactory washing resistance for antimicrobial textiles. This method can be applied not only to PP but also PET, a fiber widely used in functional textiles. The results showed that the antimicrobial activity against *E. coli* decreased by only 3.8% after 50 washes. Moreover, the antimicrobial activity against all the tested bacteria remained above 94%, as shown in Table 2 and Table S4 (ESI[†]). These results for *E. coli* are more favorable than the reductions observed in previous studies, such as a 5% reduction after 50 washes for AgNP@HTCS antibacterial fabrics, 25% reduction after 10 washes for surfactants stabilizing CuO-NPs on fiber surfaces, 2% reduction after 10 washes for improving the adhesion properties of CuO-NPs on cotton fibers using a non-toxic, biocompatible starch material (with a 7.7% reduction in *S. aureus*), and 26.4% reduction after 25 washes for Cu₂O fabrics.^{76–79}

Table 2 Washing resistance of antimicrobial activities of Cu₂O NP-PET meltblown nonwoven fabrics

Bacteria species	Antimicrobial activity		
	Before washes	After washes	Reduction (%)
<i>P. aeruginosa</i> (ATCC 10145)	5.77	5.43	94.1
<i>E. coli</i> (ATCC 8739)	5.77	5.55	96.2
MRSA (ATCC 33591)	5.83	5.59	95.9
<i>K. pneumonia</i> (ATCC 4352)	5.86	5.55	94.7
<i>S. aureus</i> (ATCC 6538P)	5.62	5.42	96.4



The antimicrobial activity was categorized as very effective according to JIS L 1902:2015. Thus, our results demonstrate that our approach can solve the problem of antimicrobial durability in textiles. The COVID-19 pandemic has caused widespread morbidity and mortality globally, disrupting economies and lifestyles. The SARS-CoV-2 virus, which causes COVID-19, can remain viable on some solid surfaces for up to a week.^{80,81} Therefore, mechanisms for reducing the viability of SARS-CoV-2 on surfaces are urgently needed. Our study, along with previous research, suggests that Cu₂O-based polymer nanocomposites can reduce the viability of bacteria and viruses.⁸² Consequently, the Cu₂O NP-embedded PP and PET masterbatches developed in this study can be applied for the production of face masks, surgical masks, and high-efficiency particulate air (HEPA) filters used in hospitals. These Cu₂O NP-embedded masks and filters can reduce the viability of viruses and improve the efficacy of medical treatments.

Conclusions

This study described an innovative method of doping Cu₂O NPs into masterbatches to manufacture Cu₂O NP-embedded textiles that showed excellent antimicrobial activity. When the contact time was in the range of 18–24 h, the antimicrobial activity of the Cu₂O NP-embedded PP textiles showed their effectiveness in inhibiting Gram-positive and Gram-negative bacteria. The textiles made from these masterbatches could effectively eliminate copper-green *Pseudomonas*, *Escherichia coli*, MRSA, *Klebsiella pneumoniae*, *Staphylococcus aureus*, and *Candida albicans*, even after being stored for a long period or washed 50 times, demonstrating long-lasting antibacterial activity. In terms of application, we demonstrated that the Cu₂O NP-embedded PP masterbatches can be used to manufacture antibacterial masks and similar products. We also successfully embedded Cu₂O NP into PET, which can potentially be used in medical textiles in the future, such as surgical gowns, hospital bed sheets, and air filters, to reduce hospital-acquired infections.

Abbreviations

PET	Polyethylene terephthalate
HAIs	Healthcare-associated infections
MRSA	Methicillin-resistant <i>Staphylococcus aureus</i>
VRE	Vancomycin-resistant <i>Enterococcus</i>
ROS	Reactive oxygen species
Cu NPs	Copper nanoparticles
PP	Polypropylene
Cu ₂ O	Cuprous oxide
DLS	Dynamic light scattering
CuO	Copper oxide
Cu-PP	Cu-embedded PP
SEM	Scanning electron microscopy
EDS	Energy dispersive spectroscopy
TGA	Thermogravimetric analysis
XRD	X-Ray diffraction

XPS	X-Ray photoelectron spectroscopy
CFU per mL	Colony forming unit per milliliter
JIS	Japanese Industrial Standard
JTETC	Japan Textile Evaluation Technology Council
PS	Polystyrene
AATCC	American Association of Textile Chemists and Colorists
HEPA	High-efficiency particulate air filter

Author contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

Data availability

Data are not available for sharing. The raw/processed data required to reproduce the above findings cannot be shared at this time as the data also form part of an ongoing study.

Conflicts of interest

The authors declare no competing financial interest.

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