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Oxygen self-supplying small size magnetic nanoenzymes for synergistic photodynamic and catalytic therapy of breast cancer†

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In recent years, tumor catalytic therapy based on nanozymes has attracted widespread attention. However, its application is limited by the tumor hypoxic microenvironment (TME). In this study, we developed oxygen-supplying magnetic bead nanozymes that integrate hemoglobin and encapsulate the photosensitizer curcumin, demonstrating reactive oxygen species (ROS)-induced synergistic breast cancer therapy. Fe₃O₄ magnetic bead-mediated catalytic dynamic therapy (CDT) generates hydroxyl radicals ([•]OH) through the Fenton reaction in the tumor microenvironment. The Hb-encapsulated Fe₃O₄ magnetic beads can be co-loaded with the photosensitizer/chemotherapeutic agent curcumin (cur), resulting in Fe₃O₄-Hb@cur. Under hypoxic conditions, oxygen molecules are released from Fe₃O₄-Hb@cur to overcome the TME hypoxia, resulting in comprehensive effects favoring anti-tumor responses. Upon near-infrared (NIR) irradiation, Fe₃O₄-Hb@cur activates the surrounding molecular oxygen to generate a certain amount of singlet oxygen (¹O₂), which is utilized for photodynamic therapy (PDT) in cancer treatment. Meanwhile, we validated that the O₂ carried by Hb significantly enhances the intracellular ROS level, intensifying the catalytic therapy mediated by Fe₃O₄ magnetic beads and inflicting lethal damage to cancer cells, effectively inhibiting tumor growth. Therefore, significant *in vivo* synergistic therapeutic effects can be achieved through catalytic–photodynamic combination therapy.

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1. Introduction

Nanocatalytic therapy is widely used in cancer treatment due to its enormous advantages such as high selectivity,¹ low toxicity,^{2,3} high efficiency,^{4–6} and tunability.⁷ Efficient tumor catalysts, such as hydrogen peroxide enzymes, metal nanoparticles, and porphyrin compounds, can induce the generation of reactive oxygen species (ROS), including singlet oxygen (¹O₂), hydroxyl radicals ([•]OH), and superoxide anions

([•]O²⁻).^{8,9} By harnessing the power of these natural ROS, it is possible to effectively induce oxidative damage to cancer cells and shrink tumors, which has been adopted and further explored for *in vivo* tumor therapy.¹⁰ Despite these significant advancements, there are still challenging factors that limit the potential efficacy of catalytic therapy.^{11,12} One of the major challenges is the inherently hypoxic tumor microenvironment (TME) within solid tumors, which significantly restricts the effectiveness of nanocatalytic therapy in clinical applications.^{13,14}

Chemodynamic therapy (CDT), which enhances intracellular ROS through the utilization of the Fenton reaction, has garnered significant attention in cancer treatment.^{15,16} Considering the overexpression of hydrogen peroxide (H₂O₂) and mild acidic conditions in the TME, extensive exploration has been conducted on advanced nanocatalysts to capture H₂O₂ as a substrate.^{17–19} However, achieving sufficient and satisfactory therapeutic efficacy remains challenging due to the limited endogenous oxygen (O₂) content.²⁰ Hemoglobin (Hb) is a composite protein composed of four subunits. Each Hb molecule consists of two α subunits and two β subunits, with each subunit containing a heme molecule capable of binding oxygen. The structure and function of hemoglobin enable it to

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encapsulate and transport oxygen.²¹ Using Hb to elevate the oxygen concentration within tumors, this combination therapy approach enhances the effectiveness of anti-tumor treatments.^{22,23} However, few researchers have explored the simultaneous elevation of O₂ levels and increased ROS within cancer cells to improve therapeutic outcomes. Therefore, there is an urgent need to develop novel combination therapies to enhance the efficacy of catalytic treatments. Curcumin (Cur), a diketone compound extracted from the rhizomes of turmeric, exhibits diverse pharmacological activities such as anti-inflammatory and anti-tumor effects.²⁴ Cur, being a natural photosensitizer, can mediate the generation of singlet oxygen in photodynamic therapy (PDT) to kill tumor cells. Additionally, as a chemotherapeutic agent, it synergizes with PDT to exert anti-tumor effects.^{25,26}

In this study, we constructed a composite magnetic nanosystem, utilizing a Hb-coated Fe₃O₄ based nanoenzyme and Cur (referred to as Fe₃O₄-Hb@cur), for enhancing ROS and designing catalytic reactions in cancer nanotherapy (Fig. 1). This strategy possesses the following characteristics: (1) Nanozymes exhibit inherent peroxidase-like activity, decomposing H₂O₂ into ·OH to induce tumor cell damage. (2) Oxygen

self-supply can be achieved through the encapsulation of Hb, and the increased O₂ can be activated by Cur to generate ¹O₂ for tumor cell death, thereby enhancing the catalytic performance of the nanozymes. Based on this therapeutic approach, we report the preparation of a small-sized magnetic nanozyme with intrinsic oxygen supply (Fe₃O₄-Hb@cur), which demonstrates synergistic effects on tumor damage through combined photodynamic and catalytic therapy. In summary, our work highlights the tremendous potential of modulating the TME using intelligent nanosystems to enhance the effectiveness of various treatment modalities, potentially inhibiting tumor growth and achieving comprehensive effects on cancer. This has significant implications in the field of nanocatalytic medicine.

2. Experimental section

2.1. Materials

Curcuminoids (Cur) and hemoglobin (Hb) were purchased from Macklin Biochemical Co., Ltd (Shanghai, China). 3,3',5,5'-Tetramethylbenzidine (TMB), 2',7'-dichlorofluorescein diacetate (DCFH-DA), 4',6-diamidino-2-phenylindole (DAPI) and 1,3-diphenylisobenzofuran (DPBF) were purchased from Aladdin Reagent Co., Ltd (Shanghai, China). Cy5.5 was purchased from Xi'an ruixi Biological Technology Co., Ltd. The cell counting kit was purchased from ZETA life. All reagents were of analytical grade and used without any purification.

2.2. Synthesis of Fe₃O₄

Monodisperse Fe₃O₄ nanoparticles with average diameters of around 15 nm were synthesized using a previously published protocol.²⁷ Afterwards, the product was precipitated with ethanol and redispersed in chloroform. The obtained Fe₃O₄ nanoparticles were then transferred into water by a polymer coating process, which was carried out by a published method.²⁸ Finally, Fe₃O₄ nanoparticles were stored in water at 4 °C for further use.

2.3. Synthesis of Fe₃O₄-Hb@cur

Under magnetic stirring, 2 mL of Fe₃O₄ (2 μM) was mixed with a 4 mL solution of Hb (dissolved in PBS at a concentration of 1 mg mL⁻¹) overnight. The mixture was then slowly added dropwise to 2 mL of cur (300 μM) and further blitzed overnight on a rotary mixer. The final product was concentrated using an ultrafiltration tube to 4 mL and stored at 4 °C.

2.4. Characterization of Fe₃O₄-Hb@cur

The morphological features of Fe₃O₄ and Fe₃O₄-Hb were analyzed using Transmission Electron Microscopy (TEM, JEOL JEM 2100F from Japan Electron). UV-vis absorption spectra of Fe₃O₄, cur, Hb, and Fe₃O₄-Hb@cur were recorded using an ultraviolet-visible spectrophotometer (Lambda25, PerkinElmer, USA). A Dynamic Light Scattering (DLS) analyzer (Malvern Instruments, UK) was used to measure the particle size of Fe₃O₄-Hb@cur. Electron Paramagnetic Resonance (EPR,

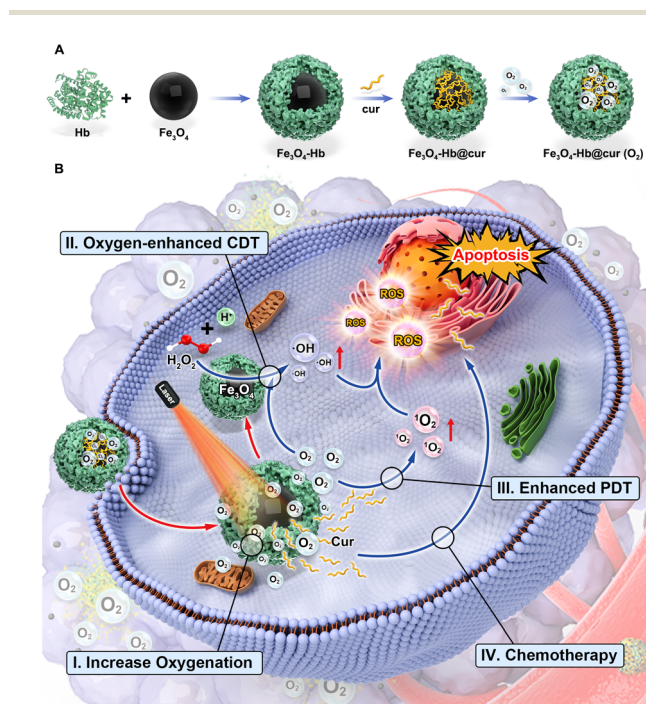


Fig. 1 A scheme of the preparation process (A) of the oxygen-supplying magnetic nanoparticle enzyme (Fe₃O₄-Hb@cur) and its mechanism for achieving photodynamic and catalytic therapy of tumor (B). In a hypoxic environment, oxygen molecules are released from Fe₃O₄-Hb@cur to overcome the oxygen-deficient TME. Hb carrying O₂ is co-loaded with the photosensitizer/chemotherapeutic agent cur, and upon NIR irradiation, it activates the surrounding molecular oxygen to generate a certain amount of ¹O₂ for PDT. Simultaneously, O₂-enhanced Fe₃O₄ mediates CDT by producing ·OH through the Fenton reaction. Therefore, Fe₃O₄-Hb@cur achieves significant *in vivo* synergistic therapeutic effects to accomplish the mechanism of catalytic-photodynamic therapy for breast cancer.

BRUKER EMXPLU) was used to measure the signals of reactive oxygen species. The EDS (Energy Dispersive Spectrometer) used for detecting the elements, composition information, and concentration of the samples was Oxford Xplore 30. *In vitro* magnetic resonance (MR) images using T_2 -weighted sequences were acquired using a 3.0 T clinical MR scanner (Erlangen, GER). The signal intensity in the region of interest (ROI) was measured for all T_2 -weighted MR images. To calculate the relaxation rates (r_2 , $1/T_2$), a range of concentrations of the $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ complex were measured. FL images were analyzed with the IVIS imaging system (CALIPER IVIS SPECTRUM). The collection parameters were $E_m = 700$ nm and $E_x = 640$ nm. Fluorescence spectral scanning was performed using an FL970 fluorescence spectrophotometer (Tianmei, China).

2.5. Hemolysis assay

0.5 mL of blood sample was added to 1 mL of PBS solution and centrifuged at 4500 rpm for 5 min, and the red blood cells were separated from the serum. Then, the red blood cells were washed 5 times with PBS and resuspended in 5 mL of PBS. Next, 0.3 mL of the above-mentioned red blood cells was mixed with 1.2 mL of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ at different concentrations (0–100 μM , prepared in PBS). The solution was gently mixed and allowed to stand at 25 °C for 4 h, followed by centrifugation to separate the red blood cells. The dissolved red blood cells could not be separated. The absorbance of the supernatant was then measured at a wavelength of 541 nm (mean \pm SD, $n = 3$). Red blood cells treated with PBS and de-ionized water were used as negative and positive control groups, respectively. The percentage of hemolysis was calculated using the equation:

$$\text{Hemolysis}(\%) = \frac{(\text{OD sample} - \text{OD negative control})}{(\text{OD positive control} - \text{OD negative control})} \times 100\%$$

2.6. H_2O_2 -dependent catalysis of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$

Briefly, 10 μL TMB (1 mM) and 150 μL acetic acid buffer solution (pH 5.5) were added to acetic acid buffer solution ($\text{H}_2\text{O}_2 + \text{Fe}_3\text{O}_4\text{@cur}$, $\text{H}_2\text{O}_2 + \text{Fe}_3\text{O}_4\text{-Hb@cur}$, and 150 μL H_2O_2 (1 mM)), respectively. Degradation of TMB was monitored with UV-Visible absorbance at 652 nm 1 h after the reaction. The degradation of TMB was monitored by observing the change in UV-Visible absorbance to evaluate the peroxidase activity of Fe_3O_4 . Specifically, different concentrations of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (3, 6, 9, 12, and 15 μM) were added to a mixture containing acetic acid buffer solution (pH 5.5) and H_2O_2 (1 mM), following the addition of 10 μL TMB (1 mM). The absorbance at 652 nm was immediately detected and the data were recorded continuously for 8 min.

2.7. Light-triggered ROS generation

Diphenylisobenzofuran (DPBF) was used in this experiment to trap the $^1\text{O}_2$ produced by $\text{Fe}_3\text{O}_4\text{@cur}$ under light conditions. We prepared $\text{Fe}_3\text{O}_4\text{@cur}$ (Fe_3O_4 (2 μM , 15 μL) + cur (300 μM , 150 μL) + 300 μL PBS) and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (Fe_3O_4 (2 μM , 15 μL)

+ cur (300 μM , 150 μL) + Hb (1 mg mL^{-1} , 50 μL) + 250 μL PBS). 100 μL of the above samples were taken and mixed with 10 μL DPBF (20 mM) at 25 °C. $\text{Fe}_3\text{O}_4\text{@cur}$ and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (300 μM as cur) and DPBF (20 mM) were irradiated with a 660 nm laser (0.5 W cm^{-2}) for 1, 2, 3, 4, or 5 min, respectively. Subsequently, the absorbance was recorded at a wavelength of 405 nm using a microplate reader.

2.8. Cell culture

4T1 mouse mammary adenocarcinoma cells were procured from Beina Biotech Co., Ltd and maintained in culture medium (DMEM) supplemented with 10% (v/v) fetal bovine serum and a 1% penicillin-streptomycin mixture. The cells were cultured under standard conditions (5% CO_2 , 37 °C).

2.9. Cell uptake of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$

4T1 cells were inoculated in confocal culture dishes at a density of 1×10^5 cells per dish and cultured in 5% CO_2 at 37 °C or 24 h. In 300 μM $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ after replacing medium, the cells were subjected to further different incubation times (1 h, 5 h, 9 h). Subsequently, the culture medium was aspirated, and the cells were washed three times with PBS. Paraformaldehyde was then added for an 8-minute incubation period to fix the cells. After removing the fixative, DAPI was added and incubated for 20 min. The cell nuclei were stained with DAPI in the co-localization experiments. The cells were observed using a confocal laser scanning microscope (CLSM) with an excitation wavelength of 471 nm for cur and 405 nm for DAPI.

2.10. Cell viability

The viability of 4T1 cells treated with different concentrations of PBS (as control), Fe_3O_4 , $\text{Fe}_3\text{O}_4\text{@cur}$, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$, $\text{Fe}_3\text{O}_4\text{@cur}$ + laser, and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ + laser was determined. The 4T1 cells were cultured in 96-well plates at an initial density of 10^4 cells per well and with overnight incubation at 37 °C. After that, gradient concentrations of Fe_3O_4 , $\text{Fe}_3\text{O}_4\text{@cur}$ and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ were introduced and co-incubated with the cells for 24 h. A 100 μL medium solution containing 10 μL of CCK-8 was added to each well and incubated with the cells for 0.5 h. The optical density (OD) at 450 nm was measured using a microplate reader. Cell viability was calculated using the following formula:

$$\text{Cell viability}(\%) = \frac{(\text{OD}(\text{sample}) - \text{OD}(\text{blank group}))}{(\text{OD}(\text{control}) - \text{OD}(\text{blank group}))} \times 100.$$

2.11. Intracellular $\cdot\text{OH}$ detection

The intracellular $\cdot\text{OH}$ was measured *via* CLSM, and DCF served as a $\cdot\text{OH}$ indicator. The 4T1 cells were plated in 6-well plates and incubated at 37 °C overnight. Then, the cells were treated with different formulations (PBS (as a control), $\text{Fe}_3\text{O}_4\text{@cur}$, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$, $\text{Fe}_3\text{O}_4\text{@cur}$ + laser ($\text{Fe}_3\text{O}_4\text{@cur}$ (+)) and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ + laser ($\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (+)) (300 μM as cur)) for 12 h. Next, after washing the cells with PBS several times, the DCF solution (10 μM) was added and incubated

with the cells for 30 min. Finally, the cells were observed by CLSM.

2.12. Animal model

Female BALB/c mice aged 6 weeks and weighing 18–22 g were procured from Zhuhai Baishitong Biotechnology Co., Ltd. Subsequently, 4T1 cells which were suspended in 100 μ L of phosphate-buffered saline (PBS) at a concentration of 3×10^6 were inoculated into the subcutaneous site of the right hind limb of each BALB/c mouse. The tumor volume was estimated every other day according to the following formula: $(\text{length} \times \text{width}^2)/2$. All animal procedures were performed in accordance with the Guidelines for Care and Use of Laboratory Animals of Guangdong Medical University and approved by the Animal Ethics Committee of Guangdong Medical University (GDY2204043).

2.13. Monitoring the distribution of drugs in tumor-bearing mice

The distribution of nanoparticles was studied using the 4T1 mouse breast cancer model. Upon reaching a tumor volume of 150 mm³, mice bearing tumors were randomly assigned into groups ($n = 3$) and received intravenous injections of either free Cy5.5 (2 mg kg⁻¹) or Cy5.5@Fe₃O₄-Hb@cur (2 mg kg⁻¹ of free Cy5.5). At specified time points after injection (1, 2, 4, 8, 12, 24, and 48 h), fluorescence biodistribution was analyzed using the IVIS *in vivo* imaging system. The collection parameters were Em = 700 nm and Ex = 640 nm.

2.14. MRI evaluation

In order to evaluate the tumor accumulation of nanoparticles, 4T1 tumor-bearing mice were subjected to MRI imaging at different time intervals following intravenous administration of either PBS or Fe₃O₄-Hb@cur. The mice were intravenous injection with 100 μ L of PBS or Fe₃O₄-Hb@cur (containing 2 μ M Fe₃O₄). MRI intensity imaging and semi-quantitative analysis were conducted using a 3.0 T clinical MR scanner (Erlangen, GER) at two time points: immediately after injection and 3 hours post-injection.

2.15. *In vivo* synergistic photodynamic and catalytic antitumor therapy

A 4T1 xenograft tumor model was established as mentioned above for evaluation of antitumor efficacy. The mice were then randomly separated into five groups ($n = 6$ mice per group): saline (as a control group), Fe₃O₄, Fe₃O₄@cur, Fe₃O₄-Hb@cur, Fe₃O₄@cur + laser (Fe₃O₄@cur (+)) and Fe₃O₄-Hb@cur + laser (Fe₃O₄-Hb@cur (+)). Therapeutic measures were commenced when the tumors reached a volume of approximately 150 cubic millimeters. On day 0, mice received combined therapy *via* tail vein injection. The tumor size and body weight were documented for each mouse from day 0 to day 16, and the tumor volume V was computed as $V = W^2 \times L/2$ (W and L are the shortest and longest diameters of the tumor, respectively). After 16 days of treatment, all mice were euthanized, and the major organs and tumor tissues were collected for further analysis.

2.16. Statistical analysis

All statistical analyses were analyzed using GraphPad Prism version 9 software. Experimental data were collected three times or more. The results were expressed as means \pm standard deviation. Statistical analysis of the differences between two groups was conducted using a two-tailed Student's *t*-test. Difference between the two groups was considered statistically significant when the two-sided *P*-value was less than 0.05, where all significant values were indicated as follows: **P* < 0.05, ***P* < 0.01, and ****P* < 0.001.

3. Results and discussion

3.1. Construction and characterization of the Fe₃O₄-Hb@cur nanoplatform

In our study, homogeneous Fe₃O₄ nanoparticles with the average size of 15 nm were prepared by the thermal decomposition method, and after phase transfer into aqueous solution by polymer coating, a large number of carboxyl groups on their surface can then physically adsorb proteins (such as Hb) through electrostatic interactions. Small-sized magnetic nanozymes (Fe₃O₄-Hb) were further created by loading the Hb shells on the surface of Fe₃O₄ nanoparticles. The cavities of Hb can easily be loaded with many small molecule drugs (such as cur). Then, a photosensitizer/chemotherapeutic drug, cur, is loaded into the Fe₃O₄-Hb structure to obtain Fe₃O₄-Hb@cur. Therefore, the interaction during the formation of Fe₃O₄-Hb@cur is through physical absorption. Finally, oxygen is introduced into the solution to obtain Fe₃O₄-Hb@cur (O₂). Representative transmission electron microscopy (TEM) images show that the prepared nanozyme in this design have a single spherical morphology with an average diameter of about 15 nm (Fig. 2A). Notably, the successful encapsulation of Hb can be evidenced by the appearance of white halos on the surface of Fe₃O₄-Hb after Hb encapsulation of Fe₃O₄ (Fig. 2B). The dispersion of Fe₃O₄-Hb@cur was evaluated by dynamic light scattering (DLS). The DLS measurements indicate that Fe₃O₄-Hb@cur is homogeneous and well dispersed, with an average particle size of approximately 30 nm (Fig. S1†). Small-sized nanoparticles facilitate drug uptake and penetration within the tumor owing to elevated permeability and retention (EPR) effects.^{29,30} As shown in the UV-vis spectra, due to the interaction between the carrier during cur loading, a red shift occurs in the peak position. As shown in Fig. 2C, the UV absorption peak of Fe₃O₄-Hb@cur matches the peak shape of cur. This absorption red shift demonstrates the successful encapsulation of cur in Fe₃O₄-Hb@cur. Moreover, the spectra changed significantly after Hb encapsulation with a strong absorption at 410 nm, indicating the formation of the Hb layer. The Fe₃O₄-Hb@cur sample shows both Hb and Fe₃O₄ absorption peaks after cur and Hb-bound Fe₃O₄. The successful loading of Cur was indicated by the change in the zeta potential, which was -15 mV for Fe₃O₄, -0.15 mV for cur, and -5.5 mV for the synthesized Fe₃O₄-Hb (Fig. S2†). Subsequently, we tested the zeta potential of the final product,

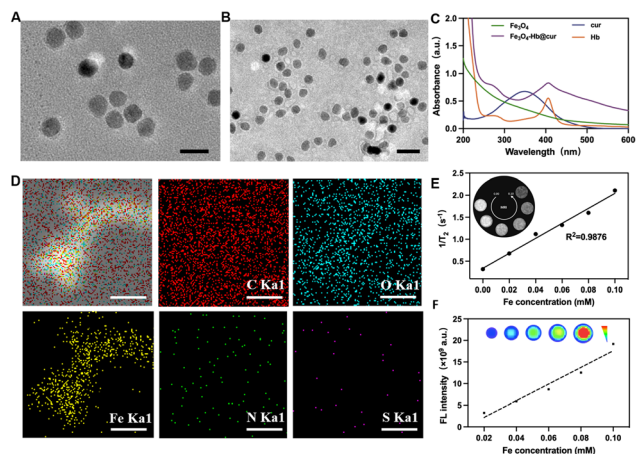


Fig. 2 Structural and compositional characterization of the $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ magnetic nanozyme. TEM images of (A) Fe_3O_4 , scale bar: 25 nm, and (B) $\text{Fe}_3\text{O}_4\text{-Hb}$, scale bar: 50 nm. (C) UV-vis spectra of Fe_3O_4 , cur, Hb, and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ NPs. (D) Dark-field TEM image and the corresponding elemental mapping of C, O, Fe, N, and S of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$. Scale bar: 30 nm. (E) Longitudinal relaxation rates ($1/T_2$) of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ solution treated with pH 7.4 as a function of Fe concentration. The following images show T_2 -weighted MR images of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ after treatment at a pH of 7.4. (F) FL intensity of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ solution at 673 nm as a function of Fe concentration. The inset shows the corresponding FL images of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ solution.

$\text{Fe}_3\text{O}_4\text{-Hb@cur}$, and the result was -1.5 mV. The loading of cur successfully neutralized the electronegativity of the final product. Moreover, we compared the fluorescence spectra of cur, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$, and $\text{Fe}_3\text{O}_4\text{-Hb}$ in the range of 450–700 nm under the excitation wavelength of cur (420 nm) (Fig. S3†). The results revealed that cur and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ exhibited identical fluorescence peak positions, providing evidence for the successful loading of cur onto $\text{Fe}_3\text{O}_4\text{-Hb@cur}$. However, $\text{Fe}_3\text{O}_4\text{-Hb}$ did not exhibit the same fluorescence peak. In addition, elemental mapping was performed to reveal the distribution of the elements C, N, O, S, and Fe in $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (Fig. 2D). $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ nanoenzymes contain elements C, O, Fe, N and S. The successful synthesis of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ was further demonstrated by the combination of UV-vis and zeta potential. The atomic percentages of the elements in the obtained $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ solution were as follows: C 86.41%, N 0.08%, O 13.24%, S 0.01%, and Fe 0.25%. The zeta potential, changes in DLS size, TEM imaging, and absorption peaks indicate the successful preparation of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$. Finally, we performed hemolysis experiments to demonstrate that $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ has good biocompatibility and safety (Fig. S4†).

Magnetic resonance imaging (MRI) and fluorescence intensity (FL) experiments were performed in order to reveal the bio-imaging capabilities of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$. As shown in Fig. 2E, MRI images of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ at different concentrations were acquired using a clinical magnetic resonance scanner at room temperature. The transverse relaxivity (r_2) measurement for iron concentration was found to be $17.04 \text{ mM}^{-1} \text{ s}^{-1}$, indicating a linear relationship. $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ exhibited a decrease in

signal intensity (shortening of T_2 relaxation time) in T_2 -weighted imaging with increasing concentration.^{31,32} The fluorescence properties of the nanoparticles were subsequently investigated by attaching Cy5.5 to $\text{Fe}_3\text{O}_4\text{-Hb@cur}$. We compared the fluorescence spectra of Cy5.5, $\text{Fe}_3\text{O}_4\text{-Hb@cur-Cy5.5}$, and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ in the range of 500–1000 nm at an excitation wavelength of Cy5.5 (673 nm). The results revealed that Cy5.5 and $\text{Fe}_3\text{O}_4\text{-Hb@cur-Cy5.5}$ exhibited identical fluorescence peak positions, providing evidence for the successful loading of Cy5.5 onto $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (Fig. S5†). Cy5.5 has a specific fluorescence emission upon excitation at 673 nm, which can be further used for *in vivo* imaging. The FL signal intensity in aqueous solution under 673 nm laser excitation was linearly correlated with the concentration of Fe (Fig. 2F). Overall, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ combined with Cy5.5 showed excellent MRI and FL imaging properties, which could be used as an ideal MRI contrast and fluorescence indicator for therapeutic diagnostic applications.

3.2. H_2O_2 -dependent catalysis of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$

In the presence of acetic acid buffer solution (pH 5.5), Fe_3O_4 can exhibit peroxidase-like activity that catalyzes the degradation of H_2O_2 to generate highly toxic $\cdot\text{OH}$ for chemodynamic cancer therapy.^{33–35} 3,3',5,5'-Tetramethylbenzidine (TMB), oxidized by $\cdot\text{OH}$, changes the color of the mixture from colorless to blue-green with a typical absorbance peak at 652 nm. Therefore, it can be used to detect the generated $\cdot\text{OH}$. As shown in Fig. 3A, H_2O_2 in the absence of Fe_3O_4 based nanozymes cannot induce the Fenton reaction without obvious absorption at 652 nm. When H_2O_2 is mixed with $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ or $\text{Fe}_3\text{O}_4\text>@cur$, an evident absorption at 652 nm from TMB was detected and the color changes of the solution was recorded, indicating the peroxidase-like activity of Fe_3O_4 . The

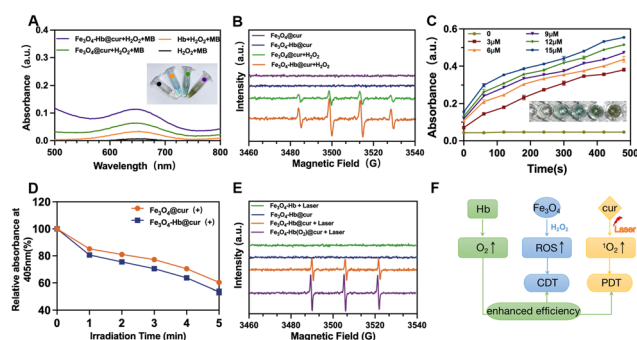


Fig. 3 Characterization of the enzyme-like activity of the $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ nanozyme. (A) UV-vis absorption spectra of TMB in different groups with pH 5.5. The inset shows the corresponding visual color changes. Generation of $^1\text{O}_2$ signals (B) and $\cdot\text{OH}$ signals (E) recorded by ESR spectra at different conditioned stimuli, respectively. (C) H_2O_2 -concentration dependence of UV-vis absorbance changes employing a TMB probe at 652 nm. (D) Changes in DPBF absorbance at 405 nm were observed after treating it with $\text{Fe}_3\text{O}_4\text>@cur$ and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ NPs under a 660 nm laser for different irradiation times. (F) Schematic representation of possible mechanisms of oxygen enhancement and tumor-expressed ROS under $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ conditions.

generated $\cdot\text{OH}$ can be used for chemokinetic therapy and combined with chemotherapy to achieve multi-mode cancer therapy. When Hb was loaded, the peak absorbance increased, indicating that O_2 carried on Hb enhanced the peroxidase activity of Fe_3O_4 and promoted the generation of $\cdot\text{OH}$. According to reports, the catalytic activity is associated with the generation capacity of $\cdot\text{OH}$.³⁶ Therefore, the detection of $\cdot\text{OH}$ production using electron spin resonance (ESR) is consistent. Compared to $\text{Fe}_3\text{O}_4@\text{cur}$ and $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ groups, the $\cdot\text{OH}$ signal was detected in both $\text{Fe}_3\text{O}_4@\text{cur}$ and $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ groups containing H_2O_2 . Furthermore, a more pronounced signal was detected in the $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ group containing H_2O_2 (Fig. 3B).

To systematically evaluate the catalytic performance of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$, we investigated the ability of the nanoenzyme to generate $\cdot\text{OH}$ under different pH conditions and found that our studied material only produced a significant amount of $\cdot\text{OH}$ at pH 5.5. In contrast, very little $\cdot\text{OH}$ was generated at pH 6.5 or pH 7.4. This indicates that a certain amount of H^+ is required for $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ to exhibit positive feedback, which is beneficial for the effective functioning of the nanoenzyme after entering tumor cells (Fig. S6†). Moreover, different concentrations (3, 6, 9, 12, and 15 μM) of H_2O_2 were added to $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ in the presence of TMB (1 mM). The absorbance was then monitored as a function of time and H_2O_2 concentration to observe the dependence (Fig. 3C). Therefore, $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ based enzyme can be used as an efficient-mimicking enzyme for cancer therapy.

3.3. O_2 -enhanced PDT effect *in vitro*

Oxygen is an essential element in the photodynamic therapy process, yet the hypoxic TME greatly reduces the efficacy of PDT for solid tumors. Therefore, an adequate supply of oxygen at the tumor site is very important for generating $^1\text{O}_2$ to treat tumors.^{37,38} In this study, the absorption of DPBF at 405 nm was irreversibly quenched by $^1\text{O}_2$, and thus was used as a probe for monitoring the production of $^1\text{O}_2$ by $\text{Fe}_3\text{O}_4@\text{cur}$ and $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ under 660 nm light irradiation.^{39,40} At various time points, the absorption values of the samples at 405 nm were continuously recorded using an UV-Vis spectrophotometer. As shown in Fig. 3D, at the fifth minute, there was a significant decrease in absorbance values for both groups. However, the group containing encapsulated O_2 in $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ exhibited a stronger signal. In particular, the $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ group showed a lower absorbance value, around 50%, indicating an increased generation of $^1\text{O}_2$. This observation suggests that the presence of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ (O_2) enhances the production of $^1\text{O}_2$. In summary, these results indicate that the release of O_2 is crucial for the antitumor effect of PDT. The loading of O_2 by Hb enhances the PDT efficacy of $\text{Fe}_3\text{O}_4@\text{cur}$, which has the potential to become a more efficient photocatalyst.

To further demonstrate the applicability of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ in PDT, the ESR technique was employed to investigate the generation of $^1\text{O}_2$ under near-infrared laser irradiation. As shown in Fig. 3E, in the absence of light and the photosensiti-

zer cur, no signal of $^1\text{O}_2$ was detected. Compared to the other groups, the $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ group carrying O_2 exhibited a stronger signal of $^1\text{O}_2$, indicating that O_2 effectively enhanced the PDT effect. This finding highlights the crucial role of O_2 in improving PDT within a hypoxic TME. These results emphasize the potential of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}(\text{O}_2)$ in enhancing the efficacy of PDT, particularly under hypoxic conditions, and provide promising prospects for cancer treatment. The aforementioned principle and mechanism are presented using a schematic diagram labeled in Fig. 3F.

3.4. Cellular uptake of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$

The cellular uptake behavior of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ was studied by Confocal Laser Scanning Microscopy (CLSM). As shown in Fig. 4A, intense fluorescence of cur was observed in the cytoplasm, indicating the successful uptake of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ by 4T1 cells. With the prolongation of the incubation time, cur was significantly accumulated in the nucleus, which indicated that after the cellular uptake of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ magnetic nanoenzymes, the Fe_3O_4 nanocarriers were ruptured in acidic lysogeny and the intracellular cur was gradually released. Semi-quantitative analysis of CLSM images in Fig. 4B showed that the uptake of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ by 4T1 tumor cells increased with prolonged incubation times (1, 5, and 9 h).

3.5. Laser enhances cytotoxicity and cell apoptosis

Next, we investigated the synergistic photodynamic and catalytic effects of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ as a nanoenzyme on tumor photodynamic and catalysis *in vitro*. We firstly tested the cytotoxicity of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ in the dark. The release of cur and the generation of $\cdot\text{OH}$ could significantly enhance the synergistic photodynamic/catalytic treatment of cancer with the help of enhanced O_2 concentration. The synergistic anticancer effect of the drugs was studied using cell viability assay kit (CCK-8) analysis. When electronically coupled reagents are

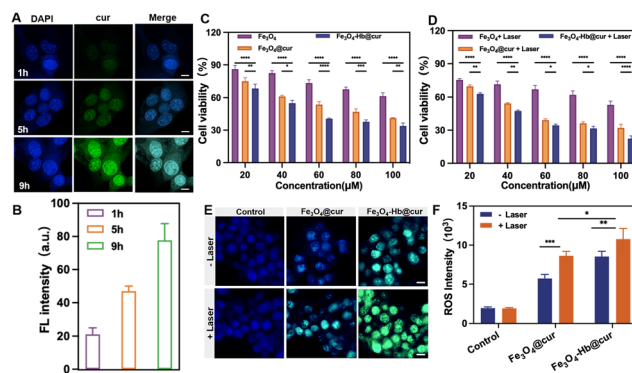


Fig. 4 Cellular uptake (A) and relative fluorescence intensity (B) of $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ after 1, 5, and 9 h of co-incubation with 4T1 cells. Scale bar: 8 nm. Cytotoxicity at different concentrations of Fe_3O_4 , $\text{Fe}_3\text{O}_4@\text{cur}$, and $\text{Fe}_3\text{O}_4\text{-Hb}@\text{cur}$ against 4T1 cells in the dark (C) and after 30 seconds of laser irradiation (D). Data are presented as means \pm SD ($n = 3$ per group). *** $P < 0.001$ and **** $P < 0.0001$. Detection of ROS levels (E) with the relative fluorescence intensity (F) after different treatments in DCFH-DA-stained 4T1 cells for 12 h. Scale bar: 10 nm.

present, WST-8 (2-(2-methoxy-4-nitrophenyl)-3-(4-nitrophenyl)-5-(2,4-dithiobenzyl)-2H-tetrazole monosodium salt) undergoes reduction by mitochondrial dehydrogenase, resulting in the formation of a water-soluble, orange-yellow product (formazan) that produces methane. The optical density (OD) of the formazan product can be measured at 450 nm using an enzyme marker. It can indirectly reflect the number of living cells. As shown in Fig. 4C, Fe₃O₄-Hb@cur showed stronger cell damage than Fe₃O₄@cur or free Fe₃O₄ at each concentration, which was attributed to the much stronger effect of synergistic photodynamic therapy/catalytic treatment than PDT or catalytic treatment alone. Furthermore, the synergistic killing effect of O₂ for both treatments was confirmed.

To evaluate the photodynamic conversion efficiency of cur, the cells were treated with a 660 nm laser (0.5 W cm⁻², 2 min). The relative survival was determined after 24 h using a colorimetric assay with tetramethyl azole salts. As shown in Fig. 4D, both types of irradiation with 660 nm excitation light exhibited enhanced drug cytotoxicity. Compared with the unenhanced light irradiation (Fe₃O₄-Hb@cur), the combined Fe₃O₄-Hb@cur under laser treatment showed the best killing effect on cancer cells at different concentrations (Fig. S7†). Therefore, the high antitumor effect of Fe₃O₄-Hb@cur in these treatments was mainly attributed to the enhanced catalytic therapy induced by the self-contained O₂ of the magnetic nanoenzymes as well as PDT.

3.6. Light-triggered ROS generation

Fe₃O₄@cur can react with local H₂O₂ to produce highly toxic ·OH in the acidic tumor microenvironment. In this study, 2',7'-dichlorofluorescein diacetate (DCFH-DA) was used to detect the intracellular ROS levels to confirm whether light irradiation increased intracellular ROS levels. Intracellular ROS can catalyze the generation of fluorescent 2,7-dichlorofluorescent diacetate (DCF) from non-fluorescent DCFH, allowing it to be used as a ROS probe to dynamically monitor ROS by fluorescent signals.^{41,42} As shown in Fig. 4E, little fluorescence was observed in 4T1 cells without any treatment, regardless of whether light was increased, indicating basal intracellular ROS levels. In contrast, ROS levels were significantly up-regulated after 4T1 cells co-cultured with Fe₃O₄@cur or Fe₃O₄-Hb@cur, which was shown as the intense green fluorescence in CLSM images (Fig. S8†). Thus, Fe₃O₄@cur contributed to the improvement of ROS production. Furthermore, a higher fluorescence intensity was detected in Fe₃O₄-Hb@cur treated cells, proving that O₂ in Fe₃O₄-Hb@cur was successfully involved in ¹O₂ generation. This further confirms that the PDT toxicity of Fe₃O₄@cur can be enhanced by the increase of O₂.

3.7. Biodistribution and tumor-targeting activity of Fe₃O₄-Hb@cur

The biological distribution and anticancer effect of Fe₃O₄-Hb@cur *in vivo* was further investigated in 4T1 Tumor-bearing Balb/c Mice. With its long excitation and emission wavelengths, Cy5.5 exhibits the capability of penetrating deep tissues, making it an ideal probe for the detection of tumor

accumulation. To visually observe the biological distribution of Fe₃O₄-Hb@cur *in vivo*, intravenous Fe₃O₄-Hb@cur was labeled with Cy5.5 (denoted as Fe₃O₄-Hb@cur-Cy5.5). Imaging and quantification of the whole body and major organs were recorded and presented using an IVIS spectrum small animal imaging system at different time intervals after intravenous injection (Fig. 5A and B). Due to the enhanced permeability and retention (EPR) effect of tumor, the material accumulates at the tumor site.^{43–45} As time progressed, the fluorescence intensity of the Fe₃O₄-Hb@cur-Cy5.5 group remained consistently stronger than that of the Cy5.5 group. Free Cy5.5 exhibited a shorter circulation half-life in mice, while the Fe₃O₄-Hb@cur-Cy5.5 group demonstrated a prolonged circulation time. At 12 and 48 h post-injection, the *in vivo* fluorescence intensity of the Fe₃O₄-Hb@cur-Cy5.5 group was 1.76-fold and 3.05-fold higher, respectively, compared to free Cy5.5. This indicates effective accumulation of Fe₃O₄-Hb@cur-Cy5.5 within the tumor (Fig. 5C). Moreover, *in vitro* imaging and quantitative analysis of major organs in Fig. 5D showed that there was still a strong fluorescence signal in Fe₃O₄-Hb@cur-

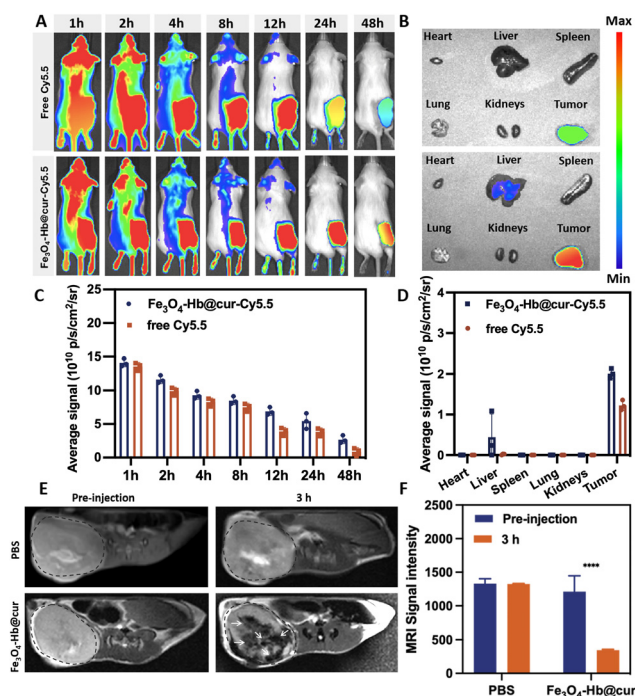


Fig. 5 *In vivo* imaging study of Fe₃O₄-Hb@cur. (A and B) *In vivo* fluorescence images of 4T1 tumor-bearing mice dissected organs, and tumors after intravenous injected with free Cy5.5 or Fe₃O₄-Hb@cur-Cy5.5 for different times. (C) Quantitative mean fluorescence intensity analysis of tumor tissues at different time intervals (*n* = 5 per group). ****P* < 0.001 and *****P* < 0.0001. (D) Fluorescence intensities of the organs and tumors collected at different times. (E) Over a period of time following the intravenous administration of Fe₃O₄-Hb@cur nanoparticles, *in vivo* magnetic resonance (MR) imaging was performed in the tumor region to monitor changes. (F) T₂-Weighted magnetic resonance imaging of mouse tumors at 0 h and 3 h after tail vein injection of Fe₃O₄-Hb@cur nanoparticles.

Cy5.5 in the tumor and liver 48 h after injection, further confirming the long cycle and EPR effect of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$.

Furthermore, MRI was employed to evaluate the accumulation of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$. To evaluate the accumulation of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ *in vivo*, T_2 -weighted MRI was performed by utilizing 4T1 breast cancer xenografts after intravenous administration of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$. Fig. 5E shows the observed MRI signal, which increased over time. Furthermore, further analysis was conducted on the statistically semi-quantitative T_2 signal intensity. The signal intensity of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ was the lowest at 3 h, while the signal intensity of the PBS group showed no difference at 3 h, consistent with the imaging results (Fig. 5F). This further confirms the strong tumor accumulation ability of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ through prolonged circulation and the EPR effect. Therefore, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ nanoparticles can effectively target tumor sites.

3.8. *In vivo* synergistic photodynamic and catalytic antitumor effects of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$

The combined therapeutic effect of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$, which exhibits effective tumor targeting and PDT effects, was evaluated by inoculating 4T1 cells in BALB/c mice. As shown in Fig. 6A, the mice were randomly divided into 5 groups (6 mice per group), and each group was injected with 100 μL saline (as a control), Fe_3O_4 , $\text{Fe}_3\text{O}_4\text{@cur}$, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$, $\text{Fe}_3\text{O}_4\text{@cur}$ + laser ($\text{Fe}_3\text{O}_4\text{@cur}$ (+)) and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ + laser ($\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (+)) through the caudal vein (an equivalent dose of 25 mg kg^{-1} cur or 0.75 mg kg^{-1} Fe_3O_4). When the tumor volume reached approximately 150 mm^3 , the treatments were initiated. Irradiation was scheduled at 12 and 24 h after drug injection based on the results of biodistribution of nanoparticles. Subsequently, the tumors were exposed to a 660 nm laser with a power density of 0.5 W cm^{-2} for a duration of 5 minutes. Fig. 6B–D illustrates that only the $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (+) group exhibited a significant inhibition of tumor volume and tumor load, following the completion of the treatments, whereas the other four groups displayed only slight tumor growth suppression. In contrast, the $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ treatment group observed a strong inhibitory effect on the tumor, which was completely suppressed within 16 days of tumor growth. The therapeutic effect was clearly observed in post-treatment mouse tumor photographs (Fig. 6B). It is worth noting that the $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ nanoparticle group also demonstrated a moderate level of antitumor activity, further confirming the important role of O_2 carried by Hb in enhanced catalytic therapy and PDT. Additionally, $\text{Fe}_3\text{O}_4\text{@cur}$ also exhibited certain therapeutic efficacy, which may be attributed to the ability of $\text{Fe}_3\text{O}_4\text{@cur}$ to effectively accumulate tumors and administer chemotherapy. Overall, there were few significant fluctuations in the body weights of the mice after the various treatments, suggesting that Fe_3O_4 nanoparticles can have negligible side effects in the tumor treatment at the doses used (an equivalent dose of 25 mg kg^{-1} cur or 0.75 mg kg^{-1} Fe_3O_4) (Fig. S9†).

At the end of the treatment, the mice were euthanized, and tumor tissues were dissected and stained for histological analysis (Fig. 6E). Hematoxylin and eosin (H&E) staining of the tumor tissues revealed typical characteristics: enlarged and darkly stained cell nuclei with minimal cytoplasm. Compared to the saline group, the treatment group with $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (+) showed severe cell necrosis in the tumor tissue, while other groups exhibited varying degrees of tumor necrosis induction. Additionally, the TUNEL assay and Ki67 immunohistochemistry were performed to further support the above findings, indicating that $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ had the strongest positive reaction and the most significant therapeutic effect. Therefore, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ demonstrated effective tumor targeting and synergistic anti-tumor activity.

3.9. Biosafety evaluation

To investigate the *in vivo* toxicity of the nanomedicine, on the 16th day of treatment, the mice were killed by cervical dislocation, and tumors were isolated and weighed. Comprehensive analysis of blood, physiological, and biochemical parameters of the mice was conducted. H&E staining was performed on major organ tissue samples (Fig. 7A). No significant organ damage or inflammatory injury was observed in any of the treatment groups compared with the physiological saline group, indicating that photodynamic and catalytic synergistic therapies had no significant impact on the organ function of mice. To assess liver and kidney damage, blood levels of AST, ALT, BUN and CRE were measured and analyzed. No significant differences occurred and remained within the

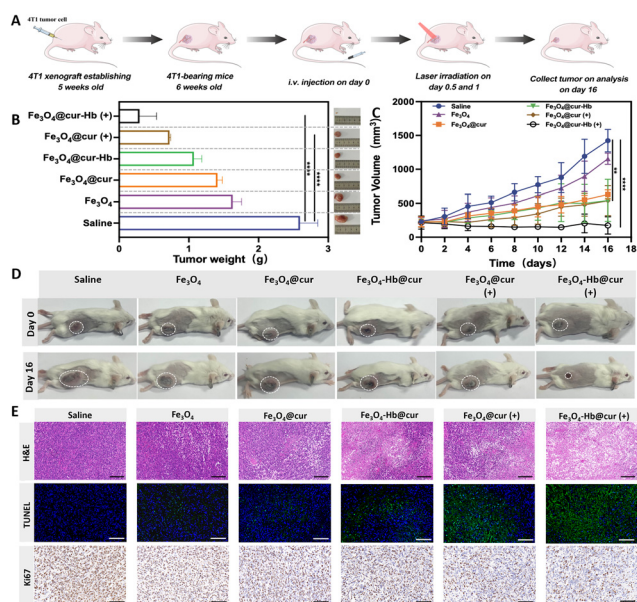


Fig. 6 (A) Schematic illustration showing the time line of the efficacy study. (B) Tumor weights in each group at the end of 16 days of treatment. (C) Tumor volume curves of 4T1 tumor-bearing mice with different treatments at 16 days. Obvious inhibition occurred in the $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (+) group ($***P < 0.001$ and $****P < 0.0001$). (D) Representative images of the 4T1-bearing mice at day 0 and day 16 after different treatments. (E) Tumor histological images of H&E staining and Ki67 staining of tumors after different treatments. Scale bar: 100 μm . TUNEL staining for apoptosis in tumors extracted from mice with different treatments (green, TUNEL; blue, and DAPI). Scale bar: 50 μm .

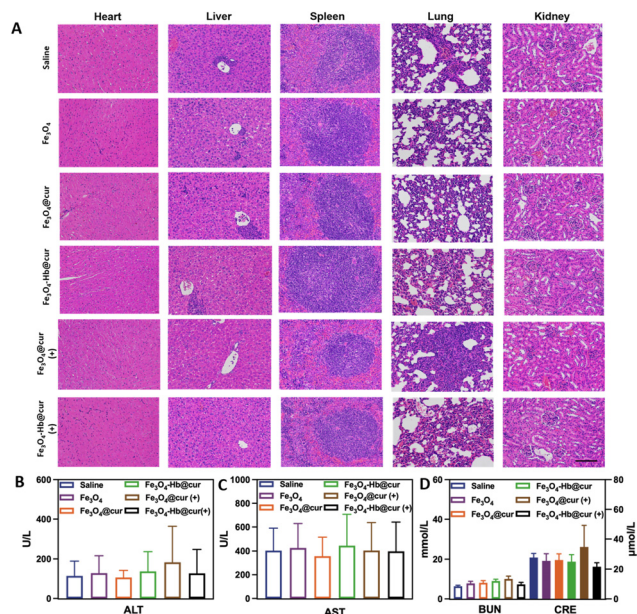


Fig. 7 *In vivo* biosafety of $\text{Fe}_3\text{O}_4\text{-Hb@cur}$. (A) H&E-stained images of major organs (heart, liver, spleen, lungs, and kidneys) harvested from 4T1 tumor-bearing mice. Scale bar, 100 nm. Each mouse was treated with a single dose of saline (as a control), Fe_3O_4 , $\text{Fe}_3\text{O}_4\text{@cur}$, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$, $\text{Fe}_3\text{O}_4\text{@cur}$ + laser ($\text{Fe}_3\text{O}_4\text{@cur}$ (+)) and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ + laser ($\text{Fe}_3\text{O}_4\text{-Hb@cur}$ (+)) through the caudal vein (an equivalent dose of 25 mg kg^{-1} cur or 0.75 mg kg^{-1} Fe_3O_4) through i.v. administration. Mice were sacrificed 16 days after drug administration. (B–D) The serology panel for biomarkers of body function and hepatic function of mice in the control and various Fe containing groups (an equivalent dose of 25 mg kg^{-1} cur or 0.75 mg kg^{-1} Fe_3O_4) after treatment for 16 days. $n = 6$ mice per group. AST: aspartate aminotransferase; ALT: alanine transaminase; BUN: blood urea nitrogen; and CRE: creatinine.

normal physiological range with regard to all these biomarkers (Fig. 7B–D). Therefore, all the measured parameters and pathological assessments indicated that Fe_3O_4 , $\text{Fe}_3\text{O}_4\text{@cur}$, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$, $\text{Fe}_3\text{O}_4\text{@cur}$ + laser and $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ + laser (at an equivalent dose of 25 mg kg^{-1} cur or 0.75 mg kg^{-1} Fe_3O_4) did not exhibit significant toxicity.

4. Conclusions

In summary, we reported a unique type of O_2 enhanced multi-functional nano-sonosensitizer platform using Fe_3O_4 and cur, encapsulated by Hb ($\text{Fe}_3\text{O}_4\text{-Hb@cur}$) successfully. Fe_3O_4 can catalyze the conversion of H_2O_2 to $\cdot\text{OH}$ through the Fenton reaction, while Hb can self-supply O_2 , overcoming the hypoxia barrier of tumor cells. Working as a photosensitizer, cur was effectively stimulated by low-intensity laser irradiation at 660 nm and this excited the surrounding O_2 to generate a certain amount of $^1\text{O}_2$ for tumor inhibition. Therefore, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ exhibited a synergistic CDT/PDT anticancer ability *in vitro* and *in vivo*. Additionally, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$ remarkably enhanced T_2 -weighted MR imaging. Overall, $\text{Fe}_3\text{O}_4\text{-Hb@cur}$

with MR imaging-guided synergistic cancer therapy has the potential for bio-application in the bio-nanomedicine field.

Author contributions

X.Y.C. directed and conceived this project with input from T.Y. and Q.Z. synthesized the materials, performed most of the experimental work, and wrote the draft of the manuscript. T.T. X., R.D. and D.Z. performed the experimental data analysis. G. Q.C. and W.C.Z. performed the theoretical study. H.P. and J.J. H. designed the project. All the authors discussed the results and contributed to the writing and revision of the manuscript.

Conflicts of interest

There are no conflicts to declare.

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