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A fluorescent probe strategy for the detection and discrimination of hydrogen peroxide and peroxyxynitrite in cells†

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Aryl boronate fluorescent probes allow the non-invasive study of dynamic cellular processes involving the reactive species, hydrogen peroxide (H₂O₂) and peroxyxynitrite (ONOO⁻). However, the ability of these probes to differentiate between these two species remains unclear. Here, we report a boronate-functionalised hemicyanine dye (HD-BPin) as a potential strategy to distinguish between H₂O₂ at 704 nm (red channel) and ONOO⁻ at 460 nm (blue channel) in solution and in cells. This work also highlights the choice of fluorophore before boronate functionalization can dictate the observed selectivity between these two species.

Hydrogen peroxide (H₂O₂) and peroxyxynitrite (ONOO⁻) are reactive oxygen (ROS) and reactive nitrogen species (RNS) found within living systems.¹ Both species have been identified as key signaling molecules that regulate a variety of cellular processes, ranging from cell growth, differentiation, migration, and programmed cell death.^{2–4} These chemical biomarkers also act as a source of oxidative and nitrosative stress, in which aberrant concentrations lead to the irreversible modification of important biomolecules such as proteins, enzyme, lipids, and DNA.^{2,5} The results of these pathological effects are linked to several diseases including cancer, cardiovascular disease, and neurodegenerative disease.^{6,7} For these reasons, extensive efforts have been devoted to developing methods that can provide detailed information on these species in biological settings.⁸ However, at present the specific identification of these individual RNS and ROS has proved challenging. Here, we report the fluorescent probe **HD-BPin** as a potential strategy for the detection of H₂O₂ (704 nm) and ONOO⁻ (460 nm) in aqueous solution and cells. The key feature of this strategy is that it takes advantage of differences in kinetic reactivity and emission wavelengths to signal the presence of each species.

Small-molecule fluorescent probes have emerged as powerful chemical tools that allow the non-invasive and real-time imaging of cellular processes and biological species.⁹ Pioneering work by the groups of Chang, Czarnik, Shabat, Lippard, and Tsien, among many others, exploited various aspects of molecular recognition and synthetic methodology to develop a suite of fluorescent probes with high level of specificity for various biological species, including ROS/RNS, biomolecules, and metal ions.^{9,10} However, as the field has grown, it has become apparent that these first- and second-generation systems built off various fluorescent scaffolds (*e.g.*, coumarin, xanthene, and BODIPY) have limitations.¹¹ For example, a first-generation H₂O₂-responsive fluorescent probe, peroxyresorufin-1 (PR1), has recently been reported to respond to ONOO⁻.^{12,13} This observed reactivity is due to the faster reaction kinetics of ONOO⁻ compared to H₂O₂ for aryl boronates.¹⁴ As a result, this functionality is now being increasingly used for the selective detection of ONOO⁻.^{14–16} However, with H₂O₂-selective aryl boronate probes still being reported,¹⁷ we rationalized that the choice of fluorescent scaffold may play a role in dictating selectivity between H₂O₂ and ONOO⁻ and therefore, we believed a strategy to distinguish these two species could be potentially identified. This study was undertaken in an effort to test this hypothesis.

Cyanine (Cy) and hemicyanine (HD)-based fluorophores have found extensive use in the design of fluorescent-based and photoacoustic-based chemical sensors.^{18–20} Many of these reported systems employ the boronate functionality to detect H₂O₂ *in vitro* and *in vivo*.^{21–23} This observed H₂O₂ selectivity is unexpected due to the greater reactivity of ONOO⁻ for aryl boronates.¹⁴ Previous studies suggest that ONOO⁻ reacts *via* the oxidative cleavage of the methine bridges (Fig. 1 – blue box) rather than the expected boronate oxidation (Fig. 1 – red box).^{24,25} We thus hypothesized boronate-based Cy- or HD-probes may provide the ability to discriminate between H₂O₂ and ONOO⁻. To test this hypothesis, we synthesized and evaluated two boronate-based fluorescent probes, **HD-BPin** and **Cy7-BPin** (Fig. 1) – see supporting information for synthetic procedures (ESI,† Scheme S1 and S2).

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Fig. 1 Chemical structures of **HD-BPin** and **Cy7-BPin**. Red box highlights H₂O₂ and ONOO⁻ reactive aryl boronate motif. Blue box highlights ONOO⁻ reactive methine bridge.

With **HD-BPin** and **Cy7-BPin** (Fig. 1) in hand, we first tested the response of each probe to H₂O₂ and ONOO⁻ in aqueous solution (PBS buffer, pH 7.2). As expected, the addition of H₂O₂ resulted in increases in the intensity of the near-infrared (NIR) fluorescence features (red emission) at 704 nm and 780 nm for **HD-BPin** and **Cy7-BPin**, respectively (Fig. 2A and ESI,† Fig. S1).^{7,8} These observations are consistent with previous reports (see ESI,† Schemes S3 and S4 for proposed reaction mechanisms).^{21,22,26} Changes in the absorption profiles of **HD-BPin** and **Cy7-BPin** were also seen (ESI,† Fig. S2 and S3). In contrast, the addition of ONOO⁻ (100 μM) to **Cy7-BPin** resulted in a dramatic change to its NIR absorption with a significant decrease at 750 nm and an appearance of a new absorption peak at ~525 nm. Unfortunately, no new fluorescent species were observed, which suggests degradation of the cyanine dye scaffold (ESI,† Fig. S4 and S5). Because of this, it was no longer a focus for the rest of the study. With the addition of ONOO⁻ (100 μM) to **HD-BPin**, an immediate color change from blue to colorless was observed (ESI,† Fig. S6). This color change was also accompanied by a rapid increase in blue fluorescence emission intensity at 460 nm ($\lambda_{\text{ex}} = 360$ nm, ESI,† Fig. S7), which is consistent with the ONOO⁻-mediated formation of a blue fluorescent xantheno dye^{24,27} – see ESI† for proposed reaction mechanism (ESI,† Scheme S5). Combined with previous reports on aryl boronate fluorescent probes (*e.g.*, PR1 and peroxyorange-1²⁸ (ESI,† Fig. S8 and S9)), this data shows the choice of fluorophore when designing a probe has potential to dictate the observed selectivity between H₂O₂ and ONOO⁻.

Next, we turned our attention to the evaluation of the dual-wavelength response of **HD-BPin** as a potential strategy for distinguishing H₂O₂ and ONOO⁻ in solution and in cells. Increasing H₂O₂ concentrations (0–450 μM) resulted in a dose-dependent increase at 704 nm with a calculated limit of detection (LOD) = 2.10 μM (Fig. 2A and ESI,† Fig. S10 and S11). Oxidation of the boronate functionality affording the red emissive species was confirmed by mass spectrometry (ESI,† Fig. S12). These responses required incubation times of >30 mins and an overall >20-fold change in red fluorescence emission intensity was observed (Fig. 2B and ESI,† Fig. S13). To our satisfaction, minimal changes in blue fluorescence emission intensity were observed, even at high H₂O₂ concentrations (*i.e.*, 200 and 400 μM) (ESI,† Fig. S14–S16).

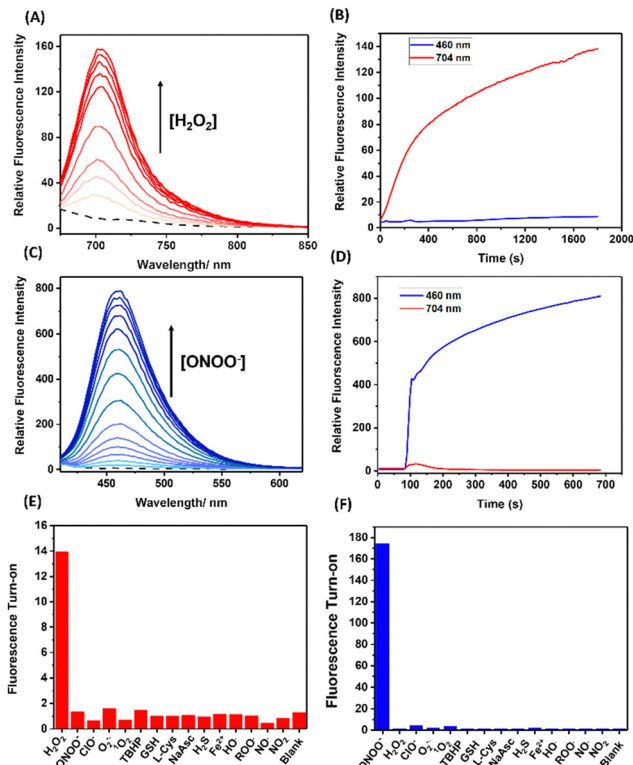


Fig. 2 (A) Fluorescent spectra of **HD-BPin** (10 μM) with increasing concentrations of H₂O₂ (0–450 μM, $\lambda_{\text{ex}} = 660$ nm, 30 min incubation). (B) Changes in relative fluorescence intensity of **HD-BPin** (10 μM) at 704 nm (Red) and at 460 nm (Blue) in the presence of H₂O₂ (200 μM), $\lambda_{\text{ex}} = 660$ nm (Red)/ $\lambda_{\text{ex}} = 360$ nm (Blue). (C) Fluorescence spectra of **HD-BPin** (10 μM) with increasing concentrations of ONOO⁻ (0–140 μM, $\lambda_{\text{ex}} = 660$ nm) (Red) and at 460 nm (Blue) in the presence of ONOO⁻ (100 μM), $\lambda_{\text{ex}} = 660$ nm (Red)/ $\lambda_{\text{ex}} = 360$ nm (Blue). (D) Changes in relative fluorescence intensity of **HD-BPin** (10 μM) at 704 nm (Red) and at 460 nm (Blue) in the presence of ONOO⁻ (100 μM), $\lambda_{\text{ex}} = 660$ nm (Red)/ $\lambda_{\text{ex}} = 360$ nm (Blue). (E) Reaction selectivity of **HD-BPin** (10 μM) with biologically relevant species (100 μM each species) at 704 nm ($\lambda_{\text{ex}} = 660$ nm). (F) Reaction selectivity of **HD-BPin** (10 μM) with biologically relevant species (100 μM each species) at $\lambda_{\text{ex}} = 360$ nm. All measurements were performed in PBS buffer solution pH = 7.20. Slit widths: 10 nm and 5 nm.

In contrast, the addition of ONOO⁻ (0–140 μM) resulted in a significant increase in fluorescence intensity at 460 nm with a calculated LOD = 0.28 μM (Fig. 2C and ESI,† Fig. S17). These ONOO⁻ induced responses required less than 30 s and an overall ~200-fold increase in blue fluorescence was observed (Fig. 2D). At low ONOO⁻ concentrations (0–15 μM), an initial increase in blue and red emission intensity was observed. However, when ONOO⁻ concentrations exceeded >10 μM, the fluorescence emission intensity at 704 nm decreased while the blue emission continued to increase in intensity (ESI,† Fig. S18 and S19). This observation suggests a stepwise deprotection mechanism, in which boronate oxidation is favored before the oxidation of the methine bridge (ESI,† Scheme S6). This potential stepwise mechanism is supported by the sigmoidal curve seen in titration experiments and mass spectroscopic analysis identifying the presence of both red and blue emissive species (ESI,† Fig. S21–S24). Although this increase in red emission was unexpected, this turn on response can be readily differentiated from



H₂O₂ due to the simultaneous increase in blue emission intensity at 460 nm. Together, these results demonstrate that **HD-BPin** can differentiate between H₂O₂ and ONOO⁻ responses in solution through differences in kinetics (ONOO⁻ (seconds) and H₂O₂ (minutes)) and differences in emission intensities at 704 nm and 460 nm. Before evaluating this strategy in cell studies, a selectivity assay was carried out against other ROS/RNS, reductants and thiols. As seen in Fig. 2E and F, excellent selectivity was observed for H₂O₂ at 704 nm and for ONOO⁻ at 460 nm (Fig. 2E and F).

Incubation of **HD-BPin** in A549 cells was shown not to impact cell viability (ESI,† Fig. S25). Subsequently, we evaluated the fluorescence response of **HD-BPin** in A549 cells *via* fixed cell imaging with the exogenous addition of H₂O₂ and SIN-1²⁹ (ONOO⁻ donor). SIN1 (500 μM) treatment led to an increase in blue fluorescence with its maximum intensity at 30 minutes (2.5-fold increase compared to time zero, Fig. 3A). No significant changes in red emission were observed over the measured time points (0–80 mins). Although a slight decrease in blue fluorescence emission can be observed after the optimal 30 min timepoint, the overall fluorescence emission intensity still differs in a statistically significant manner from untreated cells incubated with **HD-BPin** only. H₂O₂ treatment (100 μM) resulted in an expected time-dependent increase with an overall 8.6-fold increase in red fluorescence emission after 80 mins

(Fig. 3B). Consistent with solution data, minimal changes in emission intensity were seen in the blue emission channel. Scavengers, ebselen (Ebs., ONOO⁻ scavenger) and catalase (Cat., H₂O₂ scavenger) confirmed the specificity of each observed signal (Fig. 3A and B). Similar trends were observed for **HD-BPin** when tested in H460 and HCT116 cell lines (ESI,† Fig. S26 and S27). Since 1–3% of SIN1 concentration is reported to form peroxyxynitrite,³⁰ we wanted to test the response of **HD-BPin** to various SIN1 and H₂O₂ concentrations. As shown in Fig. S28 (ESI,†), increasing SIN1 concentrations (0–1500 μM) resulted in an initial increase in both blue and red emission intensity followed by a further increase in blue emission with a concomitant decrease in emission in the red channel, reflecting the obtained cuvette data. Whereas, increasing H₂O₂ concentrations (0–300 μM, 80 min incubation) led to a dose-dependent increase in the red channel with minimal increases in blue emission (ESI,† Fig. S29). It is important to note at low H₂O₂ and ONOO⁻ concentrations differences between blue and red emission are less substantial and therefore one should take caution during cell analysis.

To demonstrate the potential utility of this strategy, **HD-BPin** was evaluated in A549 cells treated with known ROS inducers, which include cisplatin, menadione and antimycin A.^{31,32} Current literature reports suggest H₂O₂ contributes to cisplatin-mediated cell death.³³ Interestingly, as seen in Fig. S30 and S31 (ESI,†), the

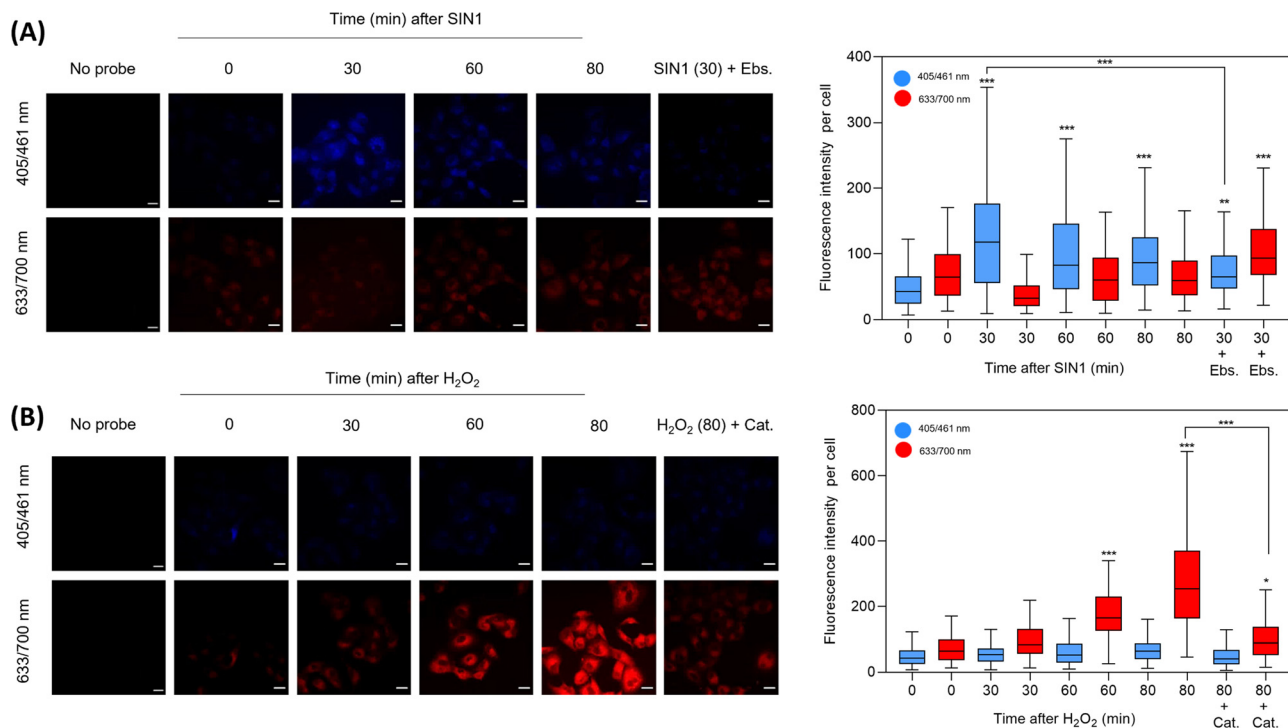


Fig. 3 Fluorescence images of A549 cells incubated with **HD-BPin** (10 μM) followed by addition of SIN-1 or H₂O₂. A549 cells were pre-treated with **HD-BPin** (10 μM) and then treated with either (A) SIN1 (500 μM) or (B) H₂O₂ (100 μM) and fixed at the indicated times. For scavenging experiments, cells were pre-treated for 4.5 hours with either Ebselen (10 μM, Ebs.) to scavenge ONOO⁻ or Catalase (1000 U mL⁻¹, Cat.) to scavenge H₂O₂. Representative images of time points taken at 405/461 nm to visualise changes in blue emission and 633/700 nm to visualise changes in red emission. Scale bar represents 20 μm. Images taken at 63x magnification. (Quantification data is fluorescence intensity per cell, black line indicates mean. For each condition 100 cells were imaged and quantified. Outliers were excluded using Tukeys box plot. Error bars represent SD. Significance tested *via* One way ANOVA. * $p < 0.05$ ** $p < 0.01$ and *** $p < 0.001$. $n = 3$.)



obtained images suggest an increase in ONOO⁻ production as time progressed in A459 cells treated with cisplatin (15 μM). Exogenous addition of Cat. (H₂O₂) and Ebs. (ONOO⁻) resulted in decreases in the respect red and blue emission suggesting presence of both H₂O₂ and ONOO⁻. PO1 was used as a comparison (ESI,† Fig. S32). A549 cells treated with menadione and antimycin A showed a dose-dependent increase in red emission with little changes seen in the blue emission channel (ESI,† Fig. S33 and S34). This observation suggests a sole increase in intracellular H₂O₂. These image profiles shows this present strategy has the potential to provide greater molecular insight and overcome the limitations of current commercially available aryl boronate fluorescent probes (e.g., PO1).

In summary, this report demonstrates the fluorescent probe HD-BPIn as a potential strategy to distinguish between H₂O₂ and ONOO⁻ in solution and in cells. Differences in kinetics and resultant emission intensities successfully enables the ability to distinguish between a H₂O₂ induced response and ONOO⁻ induced response. However, at low ONOO⁻ and H₂O₂ concentrations, the user should take caution when evaluating differences in both red and blue channels in cells. We anticipate this report to be a useful guide for the development of future ROS/RNS fluorescent probes when using the aryl boronate motif.

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Conflicts of interest

There are no conflicts of interests to declare.

References

- 1 E. A. Veal, A. M. Day and B. A. Morgan, *Mol. Cell*, 2007, **26**, 1–14.
- 2 H. Sies, *Redox Biol.*, 2017, **11**, 613–619.
- 3 P. Pacher, J. S. Beckman and L. Liaudet, *Physiol. Rev.*, 2007, **87**, 315–424.
- 4 C. Szabó, H. Ischiropoulos and R. Radi, *Nat. Rev. Drug Discovery*, 2007, **6**, 662–680.
- 5 S. Bartesaghi and R. Radi, *Redox Biol.*, 2018, **14**, 618–625.
- 6 M. P. Lisanti, U. E. Martinez-Outschoorn, Z. Lin, S. Pavlides, D. Whitaker-Menezes, R. G. Pestell, A. Howell and F. Sotgia, *Cell Cycle*, 2011, **10**, 2440–2449.
- 7 E. Gochman, J. Mahajna and A. Z. Reznick, *Anticancer Res.*, 2011, **31**, 1607–1617.
- 8 Y. Zhang, M. Dai and Z. Yuan, *Anal. Methods*, 2018, **10**, 4625–4638.
- 9 D. Wu, A. C. Sedgwick, T. Gunnlaugsson, E. U. Akkaya, J. Yoon and T. D. James, *Chem. Soc. Rev.*, 2017, **46**, 7105–7123.
- 10 K. J. Brummer, S. W. M. Crossley and C. J. Chang, *Angew. Chem., Int. Ed.*, 2020, **59**, 13734–13762.
- 11 J. Chan, S. C. Dodani and C. J. Chang, *Nat. Chem.*, 2012, **4**, 973–984.
- 12 M. Weber, A. B. Mackenzie, S. D. Bull and T. D. James, *Anal. Chem.*, 2018, **90**, 10621–10627.
- 13 E. W. Miller, A. E. Albers, A. Pralle, E. Y. Isacoff and C. J. Chang, *J. Am. Chem. Soc.*, 2005, **127**, 16652–16659.
- 14 A. Sikora, J. Zielonka, M. Lopez, J. Joseph and B. Kalyanaraman, *Free Radical Biol. Med.*, 2009, **47**, 1401–1407.
- 15 N. Rios, L. Piacenza, M. Trujillo, A. Martinez, V. Demicheli, C. Prolo, M. N. Alvarez, G. V. Lopez and R. Radi, *Free Radical Biol. Med.*, 2016, **101**, 284–295.
- 16 J. Zielonka, A. Sikora, J. Joseph and B. Kalyanaraman, *J. Biol. Chem.*, 2010, **285**, 14210–14216.
- 17 J. Hou, M. Qian, H. Zhao, Y. Li, Y. Liao, G. Han, Z. Xu, F. Wang, Y. Song and Y. Liu, *Anal. Chim. Acta*, 2018, **1024**, 169–176.
- 18 S. H. Gardner, C. J. Brady, C. Keeton, A. K. Yadav, S. C. Mallojjala, M. Y. Lucero, S. Su, Z. Yu, J. S. Hirschi, L. M. Mirica and J. Chan, *Angew. Chem., Int. Ed.*, 2021, **60**, 18860–18866.
- 19 Y. Tan, L. Zhang, K. H. Man, R. Peltier, G. C. Chen, H. T. Zhang, L. Y. Zhou, F. Wang, D. Ho, S. Q. Yao, Y. Hu and H. Y. Sun, *ACS Appl. Mater. Interfaces*, 2017, **9**, 6796–6803.
- 20 J. Yin, Y. Kwon, D. Kim, D. Lee, G. Kim, Y. Hu, J. H. Ryu and J. Yoon, *J. Am. Chem. Soc.*, 2014, **136**, 5351–5358.
- 21 J. Weber, L. Bollepalli, A. M. Belenguer, M. D. Antonio, N. De Mitri, J. Joseph, S. Balasubramanian, C. A. Hunter and S. E. Bohndiek, *Cancer Res.*, 2019, **79**, 5407–5417.
- 22 J. Zhang, L. Shi, Z. Li, D. Li, X. Tian and C. Zhang, *Analyst*, 2019, **144**, 3643–3648.
- 23 Y. Hao, Z. Li, N. Ding, X. Tang and C. Zhang, *Spectrochim. Acta, Part A*, 2022, **268**, 120642.
- 24 D. Y. Zhou, O. Y. Juan, Y. Li, W. L. Jiang, T. Yang, Z. M. Yi and C. Y. Li, *Dyes Pigm.*, 2019, **161**, 288–295.
- 25 T. Hou, K. Zhang, X. Kang, X. Guo, L. Du, X. Chen, L. Yu, J. Yue, H. Ge, Y. Liu, A. M. Asiri, K. A. Alamry, H. Yu and S. Wang, *Talanta*, 2019, **196**, 345–351.
- 26 A. Sagi, R. Weinstein, N. Karton and D. Shabat, *J. Am. Chem. Soc.*, 2008, **130**, 5434.
- 27 L. L. Wu, J. H. Liu, X. Tian, R. R. Groleau, S. D. Bull, P. Li, B. Tang and T. D. James, *Chem. Sci.*, 2021, **12**, 3921–3928.
- 28 B. C. Dickinson, C. Huynh and C. J. Chang, *J. Am. Chem. Soc.*, 2010, **132**, 5906–5915.
- 29 I. M. Bonilla, A. Sridhar, Y. Nishijima, S. Gyorke, A. J. Cardounel and C. A. Carnes, *J. Cardiovasc. Pharmacol.*, 2013, **61**, 401–407.
- 30 F. J. Martin-Romero, Y. Gutierrez-Martin, F. Henao and C. Gutierrez-Merino, *J. Fluoresc.*, 2004, **14**, 17–23.
- 31 G. Loor, J. Kondapalli, J. M. Schriever, N. S. Chandel, T. L. Vanden Hoek and P. T. Schumacker, *Free Radical Biol. Med.*, 2010, **49**, 1925–1936.
- 32 W. H. Park, Y. W. Han, S. H. Kim and S. Z. Kim, *J. Cell. Biochem.*, 2007, **102**, 98–109.
- 33 Y. I. Chirino and J. Pedraza-Chaverri, *Exp. Toxicol. Pathol.*, 2009, **61**, 223–242.

