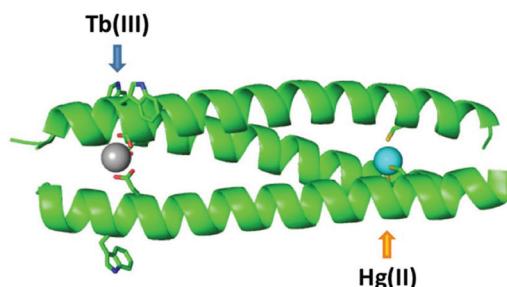




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## pH dependent binding in *de novo* hetero bimetallic coiled coils<sup>†</sup>

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**Fig. 1** Cartoon representation showing our designed hetero bimetallic coiled coil, featuring  $\text{LnAsp}_3$  and  $\text{HgCys}_3$  sites towards the N- and C-termini, respectively. Shown are the main chain atoms represented as helical ribbons (green), the Asp, Trp and Cys side chains in stick form (oxygen in red, nitrogen in blue and sulfur in orange), and the metal ions as spheres (lanthanide in grey and mercury in cyan).

the first metal inducing folding and promoting the formation of the second metal ion binding site.<sup>16</sup> A more recent example features a structural  $\text{HgCys}_3$  site<sup>17–19</sup> site and a catalytic  $\text{ZnHis}_3(\text{OH}/\text{OH}_2)$  site, inspired by the metalloenzyme carbonic anhydrase, within a three stranded coiled coil.<sup>1,20</sup>

The advantage of including two distinct mononuclear metal ion sites with different properties within a single assembly is clear, as it allows multi functionality and complexity, relating to interdependence, to be introduced. And so it remains to expand the number and type of such sites that can be combined in designs such as this, if complex multi-metalloc, multi-functional, systems are to be realised. Herein we explore the inclusion of a lanthanide binding site, of interest in biotechnology and protein design due to their appealing NMR and luminescent properties, into a heterobimetallic coiled coil, alongside the previously well characterized structural, but pH dependent,  $\text{HgCys}_3$  site,<sup>17</sup> see Fig. 1. Not only do we report the first example of a heterobimetallic coiled coil featuring a lanthanide binding site, we also report on the pH dependence of these lanthanide sites, and illustrate how pH can be used as to control and select for  $\text{Hg}(\text{II})$  and  $\text{Tb}(\text{III})$  binding, respectively.

## Results and discussion

### Design of a hetero bimetallic coiled coil

Given that the introduction of a metal binding site is a destabilizing modification to the coiled coil structure, the inclusion of two has to be carefully considered. Previously we reported a small family of mononuclear lanthanide binding sites within three stranded coiled coils, and found the location of the binding site to have a significant impact on coiled coil folding and stability. Terminal binding sites were significantly better tolerated than core sites, and the N-terminal site was better tolerated than the more buried C-terminal site. Furthermore, whereas the majority of binding sites required the substitution of two adjacent hydrophobic layers to accommodate the double AsnAsp binding site, when the binding site was introduced towards the N-terminus, the Asn was found to

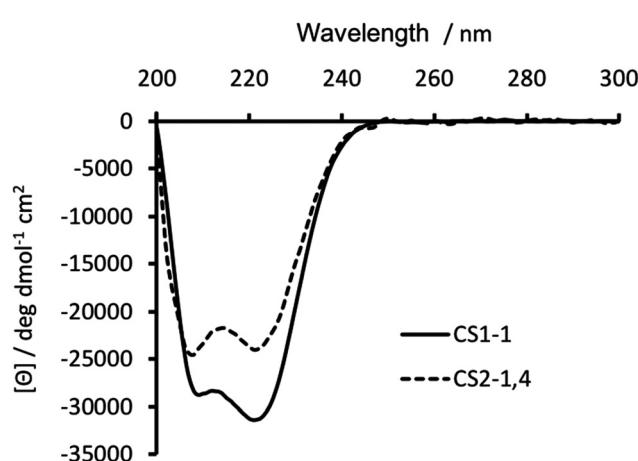
**Table 1** Peptide sequences used in this study

Peptide	Sequence (abcdefg)
CS1-1	Ac-G IAAIEWK <b>D</b> AAIEQK IAAIEQK IAAIEQK G-NH <sub>2</sub>
CS2-1,4	Ac-G IAAIEWK <b>D</b> AAIEQK IAAIEQK IAAIEQK CAAIEQK G-NH <sub>2</sub>
MB1-2	Ac-G IAAIEQK IAAN <u>E</u> WK <b>D</b> AAIEQK IAAIEQK IAAIEQK G-NH <sub>2</sub>

be non-essential. As such the least destabilizing site, which retains lanthanide binding, features a single Asp layer towards the N-terminus, CS1-1 and see Table 1.<sup>9</sup> Our hetero bimetallic coiled coil therefore features this lanthanide binding site, with an adjacent Trp residue which reports on  $\text{Tb}(\text{III})$  binding, and a second distinct metal binding site.

In order to achieve a high degree of metal selectivity for the two sites, we elected to generate a soft  $\text{Cys}_3$  metal binding site, to contrast with the hard lanthanide binding site generated by the  $\text{Asp}_3$  layer. The replacement of a core Ile with a Cys should be less destabilizing than the analogous mutation to Asp, but in an effort to minimize its impact, the Cys has again been introduced in a terminal rather than a core heptad, CS2-1,4 (and see Table 1, Fig. S1 and S2†). Though the binding of various soft metal ions has been reported to such a site,<sup>21,22</sup> we selected the well-studied and previously reported, structural  $\text{HgCys}$  site located in an “a” (the first position within a “abcdefg” heptad) rather than the “d” position (also located within the core of a three stranded coiled coil).

Substitution of Ile in position 30 with a Cys, was anticipated to be destabilizing, with previous reports<sup>18,19,23</sup> showing this to be the case. Circular dichroism (CD) spectroscopy was therefore used to establish the degree of peptide folding. Though spectra recorded for 30  $\mu\text{M}$  monomer solutions of both CS1-1 and CS2-1,4 in 5 mM HEPES buffer pH 7.0 displayed the characteristic double minima around 210 and 223 nm, consistent with an  $\alpha$ -helix, their intensities differed. CS1-1 was notably more folded ( $\Theta_{222} = -31\,320 \text{ deg dmol}^{-1} \text{ cm}^2 \text{ res}^{-1}$ , ca. 81% folded) than CS2-1,4 ( $\Theta_{222} = -23\,933 \text{ deg dmol}^{-1} \text{ cm}^2 \text{ res}^{-1}$ , ca. 62% folded), see Fig. 2.



**Fig. 2** CD spectra of 30  $\mu\text{M}$  CS1-1 (solid line) and CS2-1,4 (dashed line) peptide monomer, recorded at 293 K in 5 mM HEPES buffer pH 7.0.



## Retaining Tb(III) binding to Asp

Verification that Tb(III) binding is preserved in CS2-1,4, can be best obtained from luminescence spectroscopy, see Fig. 3A. Tb(III) emission is known to be sensitized on binding to the CS1-1 site, due to an adjacent Trp residue which is retained in the CS2-1,4 design. This sensitized Tb(III) emission is consistent with peptide binding, but more significantly, it is consistent with binding at the designed Asp<sub>3</sub> lanthanide binding site. This is because Trp sensitized Tb(III) emission is highly distance dependent, and therefore rules out the possibility of a binding site further away, and certainly rules out the Cys location (Trp – LnAsp<sub>3</sub> site *ca.* 12 Å *vs.* Trp – Cys<sub>3</sub> site *ca.* 32 Å).

## Retaining Hg(II) binding to Cys

Similarly, in order to verify that Hg(II) binding to the Cys site was retained, ultraviolet-visible (UV-vis) absorption spectra were recorded of solutions containing 30 μM CS2-1,4 peptide monomer in 10 mM HEPES buffer. On addition of 10 μM HgCl<sub>2</sub> at pH 7.0 we see a fairly featureless spectrum (see Fig. 4), but which is consistent with Hg(II) binding as HgCys<sub>2</sub> to Cys located in “a” sites at pH 7.0,<sup>17</sup> as would be the case here. However, as the pH is raised to 7.5 and ultimately 8.6, we observe the appearance of the characteristic signature for trigonal HgS<sub>3</sub>.<sup>17</sup> This pH dependence is consistent with the previously reported apparent pK<sub>a</sub> of 7.6, associated with coordination of trigonal HgS<sub>3</sub> to an “a” site.<sup>17</sup>

Hg(II) binding to Cys residues within the “a” site of a three stranded coiled coil has been extensively studied and found to be dependent on both the pH and the ratio of peptide monomer to Hg(II).<sup>17</sup> Consistent with these reports, the CS2-1,4 peptide, at pH 8.6 where Hg(II) is able to bind as trigonal HgS<sub>3</sub> to a Cys in an “a” site, does so up to the addition of a *ca.* 1:3 ratio of Hg(II) : CS2-1,4 monomer. On addition of further Hg(II) we observe the disappearance of the characteristic signal for HgCys<sub>3</sub>, and instead observe the featureless spectrum previously assigned to HgCys<sub>2</sub>, see Fig. S3,† again consistent with the literature.<sup>17</sup>

## pH dependence of Tb(III) binding in the presence of Hg(II)

The Tb(III) and Hg(II) binding data is consistent with selective binding to their respective Asp<sub>3</sub> and Cys<sub>3</sub> binding sites within

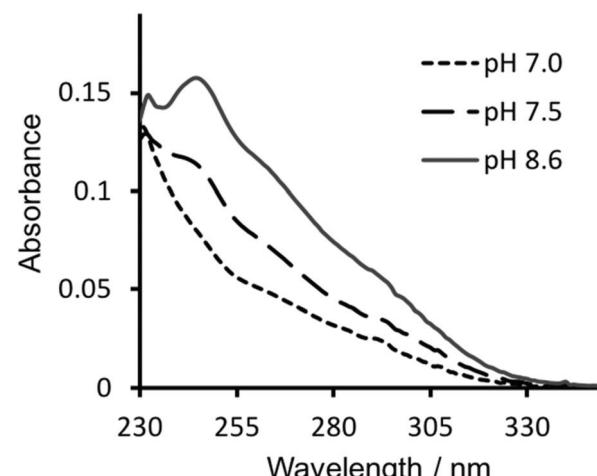


Fig. 4 UV-visible difference absorption spectra of 30 μM CS2-1,4 monomer upon the addition of 10 μM HgCl<sub>2</sub>, in 10 mM HEPES buffer at pH 7.0 (---), 7.5 (---) and 8.6 (solid line).

our two binding site peptide, CS2-1,4, and with similar chemistries to when they bind to these sites in isolation. However, with Tb(III) binding only studied at pH 7.0, but with Hg(II) binding being pH dependent with HgCys<sub>3</sub> forming above pH 8.6, it remained to be seen if Tb(III) could bind in the presence of Hg(II), and under the more basic conditions required for HgCys<sub>3</sub> formation.

Luminescence studies of 30 μM CS2-1,4 peptide monomer in the presence of both 10 μM TbCl<sub>3</sub> and 10 μM HgCl<sub>2</sub> at pH 6.8, still show enhanced emission, comparable to that recorded in the absence of Hg(II), consistent with similar Tb(III) binding when the Hg(II) is likely bound as HgCys<sub>2</sub> (see Fig. 3).<sup>24</sup> However, on raising the pH, the signal for the Tb(III) emission is significantly reduced, and lost entirely above pH 8, see Fig. 5. This could be due to reduced Tb(III) binding under these more basic conditions, due to competing formation of lanthanide hydroxide species. It could be a result of Hg(II) binding and distortion of the Asp<sub>3</sub> site on formation of HgCys<sub>3</sub>. Alternatively, as the Tb(III) CS1-1 type site is highly hydrated,<sup>9</sup> the lack of luminescence may be due to deprotonation

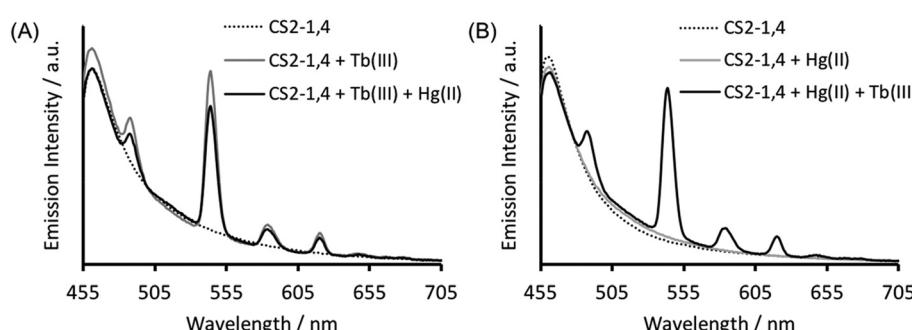
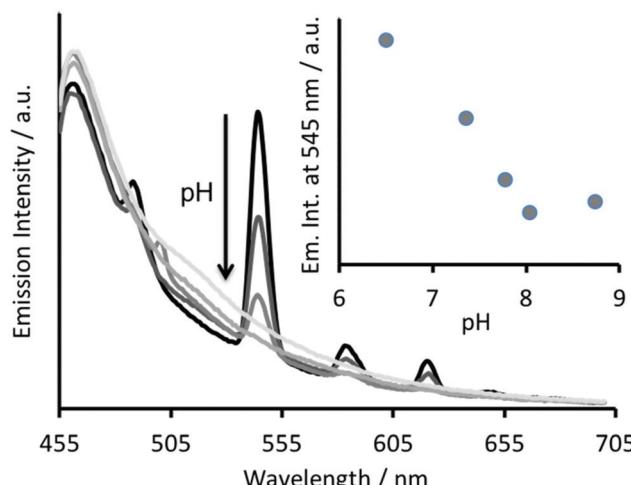


Fig. 3 Steady state luminescence spectra of 30 μM CS2-1,4 monomer (dashed line) in the presence of 10 μM TbCl<sub>3</sub> (grey line panel A), 10 μM HgCl<sub>2</sub> (grey line panel B) or both 10 μM TbCl<sub>3</sub> and 10 μM HgCl<sub>2</sub> (black line) in 5 mM HEPES buffer pH 6.8 on order of addition of (A) TbCl<sub>3</sub> or (B) HgCl<sub>2</sub>, first.  $\lambda_{\text{exc}} = 280$  nm.





**Fig. 5** Steady state luminescence spectra of 30  $\mu$ M CS2-1,4 monomer in the presence of 10  $\mu$ M  $\text{TbCl}_3$  and 10  $\mu$ M  $\text{HgCl}_2$ , at increasing pH.  $\lambda_{\text{exc}} = 280$  nm. Inset showing plot of emission intensity at 545 nm as a function of pH. The non-zero emission intensity above pH 8 is due to the contribution from the Trp residue.

tion of coordinated water and enhanced quenching by coordinated hydroxide.

In order to determine if the lack of  $\text{Tb}(\text{III})$  emission relates to  $\text{Hg}(\text{II})$  binding at the Cys site, luminescence spectra were recorded of 30  $\mu$ M CS2-1,4 monomer at pH 8.6 on addition of 10  $\mu$ M  $\text{TbCl}_3$ , but in the absence of  $\text{Hg}(\text{II})$ . However, this again showed a lack of emission at 545 nm, see Fig. S4.<sup>†</sup> Similar spectra were acquired in the presence of both 10  $\mu$ M  $\text{TbCl}_3$  and 10  $\mu$ M  $\text{HgCl}_3$ , and the order of addition altered, so as to rule out the formation of a kinetic product, but again no notable  $\text{Tb}(\text{III})$  emission was detected, see Fig. S4.<sup>†</sup> Therefore, we conclude that the lack of sensitized  $\text{Tb}(\text{III})$  emission is not due to binding of  $\text{Hg}(\text{II})$  at the Cys site, but rather is a feature of the  $\text{TbAsp}_3$  site at basic pH.

#### pH dependence of $\text{Tb}(\text{III})$ binding to single site peptides

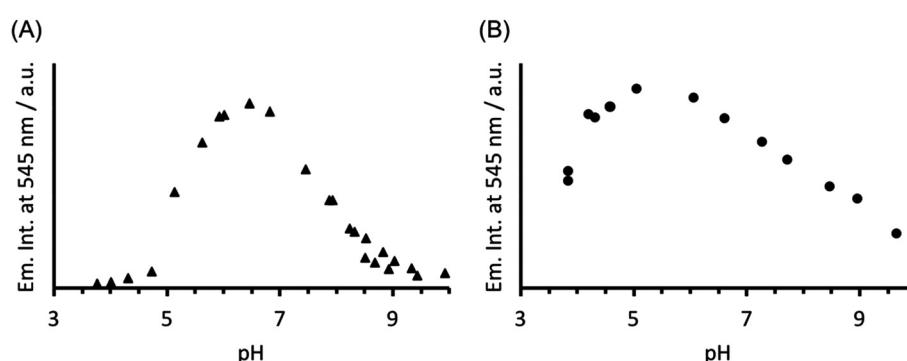
In an effort to study the pH dependence of  $\text{Tb}(\text{III})$  binding to the  $\text{Asp}_3$  site, we initially performed a pH titration on the

control CS1-1 peptide, which lacks the  $\text{Cys}_3$  binding site, and monitored this by CD and luminescence spectroscopy. We also studied the pH dependence of  $\text{Tb}(\text{III})$  binding to MB1-2, a single site coiled coil, which in contrast to the highly hydrated  $\text{Tb}(\text{III})$  site generated on binding CS1-1, offers a coordinatively saturated  $\text{Tb}(\text{III})$  site. This would also allow us to address whether a different  $\text{Tb}(\text{III})$  binding site would be more attractive for use in designing hetero bimetallic coiled coils featuring a lanthanide binding site, as we have previously reported that the chemistry of  $\text{Tb}(\text{III})$  sites within these coiled coils, can be very different depending on ligand set presented and the linear location along the coiled coil.<sup>9</sup>

The CD titration of CS1-1 in the presence of  $\text{Tb}(\text{III})$  revealed very little change in folding as a function of pH, see Fig. S5A.<sup>†</sup> However, as the change in folding associated with  $\text{Tb}(\text{III})$  binding to CS1-1 is only modest,<sup>9</sup> this is not sufficient to address  $\text{Tb}(\text{III})$  de-complexation.

Performing the same pH CD titration for a buried  $\text{Tb}(\text{III})$  binding site, as featured in MB1-2, should show a notable change in folding on de-complexation.<sup>8,9</sup> Though the CD pH titration shows a steady decrease in MB1-2 folding as a function of pH, see Fig. S5B,<sup>†</sup> it is not consistent with full  $\text{Tb}(\text{III})$  de-complexation and complete conversion to the apo form, which is only  $21 \pm 3\%$  folded at pH 7.<sup>9</sup> These findings suggest that though  $\text{Tb}(\text{III})$  de-complexation may be occurring with increase in pH, it may not be complete in these single binding site control peptides. Therefore,  $\text{Tb}(\text{III})$  may remain bound to some extent in CS2-1,4 at pH 8.6.

The analogous luminescence pH titration of  $\text{Tb}(\text{CS1-1})_3$  revealed a loss in luminescence signal with increase in pH (see Fig. 6A), consistent with our previous observations with CS2-1,4. Again these findings are consistent with the Cys site, and its metallated  $\text{HgCys}_3$  derivative, not influencing the behaviour of the  $\text{TbAsp}_3$  site, and that the loss of luminescence signal relates to the chemistry of the  $\text{TbAsp}_3$  site in isolation. Intriguingly, the pH titration also revealed for the first time that  $\text{Tb}(\text{III})$  luminescence associated with binding to the  $\text{TbAsp}_3$  site, is not observed under acidic conditions, and is in fact optimal across a very narrow pH range between pH 6 and 7, the very pH conditions under which all such lanthanide



**Fig. 6** Plot of the change in emission intensity at 545 nm as a function of pH, for a solution containing 30  $\mu$ M (A) CS1-1 and (B) MB1-2 monomer in the presence of 10  $\mu$ M  $\text{TbCl}_3$ .  $\lambda_{\text{exc}} = 280$  nm.

binding sites have been studied to date. Lack of binding under acidic conditions may result from protonation of the Asp residues, making them unavailable for Tb(III) complexation, and for which the  $pK_a$  may be altered in the different peptide sequences due to the presence and absence of, for example, an adjacent Asn.

In order to establish if the lack of Tb(III) luminescence signal at basic pH, is in fact a result of enhanced quenching due to deprotonation of water bound to the TbAsp<sub>3</sub> site, an analogous luminescence pH titration was performed with Tb(MB1-2)<sub>3</sub>, in which the Tb(III) is coordinatively saturated and is not bound directly to any water molecules. A similar profile is observed, though with a much larger pH range for optimal Tb(III) binding (*ca.* pH 4–7), with a steady decrease in luminescence on increasing the pH (see Fig. 6B). However, as was the case in the analogous CD titrations, the decrease is less extreme than for CS1-1 and is only consistent with partial Tb(III) de-complexation at pH 8.6.

### pH as an external trigger to control metal binding to a hetero bimetallic coiled coil

Taken together, the CD and luminescence pH titrations of CS1-1, MB1-2 and CS2-1,4, indicate that an increase in pH leads to Tb(III) de-complexation, and that not much Tb(III) remains bound to CS2-1,4 at pH 8.6. Future hetero bimetallic designs which were to feature a lanthanide binding site, might therefore benefit from the use of the double AsnAsp binding site, as featured in MB1-2, as it is more versatile with respect to pH. However, at neutral pH where Tb(III) is well bound, the Hg(II) can still bind to the Cys site in CS2-1,4 as HgCys<sub>2</sub>, but does require more basic conditions to bind fully as HgCys<sub>3</sub>. Therefore, in systems such as CS2-1,4, it is possible to use pH as an external trigger to control, in the case of the Hg(II) site, in what form it binds, and which metal site provides a spectroscopically measurable output. This offers a way to activate these outputs in more complex multi functional assemblies.

## Conclusions

Herein, and to date, two distinct mononuclear sites have been successfully engineered into a single coiled coil, by exploiting clear differences in metal binding preferences, such as HSAB theory. The ability to do so offers protein designers the opportunity to develop systems with clear distinct functional sites, which may operate in isolation or in a dependent fashion. Herein we report the first inclusion of a lanthanide binding site alongside a second distinct metal binding site, and illustrate that pH can be used as an external trigger to induce different binding at the two distinct metal binding sites. Through our efforts, we have for the first time reported on the pH dependence of these lanthanide binding sites, and describe the different chemistries associated with hydrated and coordinatively saturated lanthanide binding sites.

Though these findings further the metallo protein design field by contributing distinct and useful sites from which

metallo protein designers can select from, a real challenge remains to selectively bind, and distinguish between, two very similar metal ions but with different desirable properties, for which differences in HSAB or coordination chemistry, cannot be so easily exploited. Efforts in our laboratory are on-going to achieve this ultimate goal.

## Materials and methods

TbCl<sub>3</sub>·6H<sub>2</sub>O and HgCl<sub>2</sub> were purchased from Sigma Aldrich. Urea ( $\geq 99\%$  purity), xylene orange indicator, glacial acetic acid and 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES) were all purchased from Fisher Scientific Ltd. All Fmoc protected amino acids, dimethylformamide (DMF) and *N,N,N',N'*-tetramethyl-*O*-(1*H*-benzotriazol-1-yl)uronium hexafluorophosphate (HBTU) were purchased from Peptides Ltd, Leicester. The rink amide MBHA resin was obtained from AGTC Bioproducts Ltd, along with diisopropylethylamine (DIPEA) and trifluoroacetic acid (TFA). Gd(III) and Tb(III) standards were purchased from SCP Science, Quebec.

### Peptide synthesis and purification

Peptides were synthesized on a CEM Liberty Blue automated peptide synthesizer on rink amide MBHA resin (0.25 mmol scale, 0.65 mmol g<sup>-1</sup>), using standard Fmoc-amino acid solid-phase peptide synthesis protocols.<sup>25</sup> Peptides were purified as previously reported<sup>26</sup> and characterised by electrospray ionisation mass spectrometry recorded on a Waters Ltd. Synapt-G2-S, by direct injection with a mobile phase of 50% water: 50% acetonitrile and 0.05% formic acid, at a flow rate of 0.1 mL min<sup>-1</sup>. Data was processed with MassLynx. In the synthesis of CS2-1,4 the Fmoc-L-Cys(Trt)-OH was coupled twice and the temperature maintained below 50 °C.

### Sample preparation

Stock solutions of TbCl<sub>3</sub> and HgCl<sub>2</sub> (1 mM) were prepared in MilliQ water, and the Tb(III) concentrations determined spectroscopically using xylene orange indicator and Tb(III) standard solutions as reported by Fedeli *et al.*<sup>27</sup> Peptide concentrations were determined based on the tryptophan absorbance at 280 nm ( $\epsilon_{280} = 5690 \text{ M}^{-1} \text{ cm}^{-1}$ ) in 7 M aqueous solutions of urea, and in the case of CS2-1,4, verified by determining the available thiol concentration *via* the published Ellman's assay,<sup>28</sup> with experimental concentrations found to be within 10%.

### Circular dichroism (CD) spectroscopy

CD spectra for solutions containing 30  $\mu$ M monomer in the absence or presence of 10  $\mu$ M metal (TbCl<sub>3</sub> or HgCl<sub>2</sub>), in 5 mM HEPES buffer pH 7 or 8.6, were recorded in a 1 mm pathlength quartz cuvettes on a Jasco J-810 spectropolarimeter. The observed ellipticity in millidegrees was converted into molar ellipticity, ( $\Theta$ ), and is reported in units of deg dmol<sup>-1</sup> cm<sup>2</sup>. The percentage folding, %<sub>Folded</sub>, was calculated (eqn (1)) based on the theoretical maximum ellipticity, [ $\Theta$ ]<sub>max</sub>, determined from



( $-42\,500 \times (1 - (3/n))$ ), where  $n$  is the number of residues in the sequence, and  $[\Theta]_{\text{coil}}$  is the ellipticity of a random coil, based on reports by Scholtz *et al.*<sup>29</sup>

$$\%_{\text{Folded}} = \frac{[\Theta]_{222\text{ nm}} - [\Theta]_{\text{coil}}}{[\Theta]_{\text{max}} - [\Theta]_{\text{coil}}} \times 10. \quad (1)$$

### UV-visible spectroscopy

UV-visible spectra for solutions containing 30  $\mu\text{M}$  CS2-1,4 monomer, 10  $\mu\text{M}$   $\text{HgCl}_2$  in 10 mM HEPES buffer pH 7.0, 7.5 or 8.6 were recorded in a 1 cm pathlength quartz cuvettes on a Cary50 spectrometer.

### Luminescence

Emission spectra were recorded in a 1 cm pathlength quartz cuvette using an Edinburgh Instruments Fluorescence FLS920 system with a 450 W Xenon arc lamp and a Hamamatsu R928 photomultiplier tube. The emission monochromator was fitted with two interchangeable gratings blazed at 500 nm and 1200 nm and the data was collected using F900 spectrometer analysis software. Aliquots of a 1 mM stock solution of  $\text{TbCl}_3$  or  $\text{HgCl}_2$  were titrated into 30  $\mu\text{M}$  peptide monomer solutions in 5 mM HEPES buffer pH 7 or 8.6, and the emission profile recorded after 20 min equilibration. Solutions were excited at 280 nm and the emission was scanned from 475–750 nm using a 455 nm long pass filter. Spectra were corrected for instrument response (grating/PMT) in all cases. The integration of the  $\text{Tb}^{(III)}$  emission at 545 nm was measured between 535–555 nm.

### Conflicts of interest

The authors declare no conflicts of interest.

### Acknowledgements

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### References

- 1 M. L. Zastrow, A. F. A. Peacock, J. A. Stuckey and V. L. Pecoraro, *Nat. Chem.*, 2012, **4**, 118.
- 2 A. J. Reig, M. M. Pires, R. A. Snyder, Y. Wu, H. Jo, D. W. Kulp, S. E. Butch, J. R. Calhoun, T. Szyperski, T. G. Szyperski, E. I. Solomon and W. F. DeGrado, *Nat. Chem.*, 2012, **4**, 900.
- 3 A. Roy, D. J. Sommer, R. A. Schmitz, C. L. Brown, D. Gust, A. Astashkin and G. Ghirlanda, *J. Am. Chem. Soc.*, 2014, **136**, 17343.
- 4 R. L. Koder, J. L. R. Anderson, L. A. Solomon, K. S. Reddy, C. C. Moser and P. L. Dutton, *Nature*, 2009, **458**, 305.
- 5 W. D. Kohn, C. M. Kay and R. S. Hodges, *J. Pept. Res.*, 1998, **51**, 9.
- 6 W. D. Kohn, C. M. Kay, B. D. Sykes and R. S. Hodges, *J. Am. Chem. Soc.*, 1998, **120**, 1124.
- 7 A. Kashiwada, K. Ishida and K. Matsuda, *Bull. Chem. Soc. Jpn.*, 2007, **80**, 2203.
- 8 M. R. Berwick, D. J. Lewis, Z. Pikramenou, A. W. Jones, H. J. Cooper, J. Wilkie, M. M. Britton and A. F. A. Peacock, *J. Am. Chem. Soc.*, 2014, **136**, 1166.
- 9 M. R. Berwick, L. N. Slope, C. Smith, S. M. King, S. L. Newton, R. Gillis, G. Adams, A. Rowe, S. Harding, M. M. Britton and A. F. A. Peacock, *Chem. Sci.*, 2016, **7**, 2207.
- 10 O. A. Kharenko and M. Y. Ogawa, *J. Inorg. Biochem.*, 2004, **98**, 1971.
- 11 M. Matzapetakis and V. L. Pecoraro, *J. Am. Chem. Soc.*, 2005, **127**, 18229.
- 12 O. Iranzo, S. Chakraborty, L. Hemmingsen and V. L. Pecoraro, *J. Am. Chem. Soc.*, 2011, **133**, 239.
- 13 O. Iranzo, C. Cabello and V. L. Pecoraro, *Angew. Chem., Int. Ed.*, 2007, **46**, 6688.
- 14 A. F. A. Peacock, L. Hemmingsen and V. L. Pecoraro, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 16566.
- 15 M. R. Ghadiri and M. A. Case, *Angew. Chem., Int. Ed.*, 1993, **32**, 1594.
- 16 T. Tanaka, T. Mizuno, S. Fukui, H. Hiroaki, J. Oku, K. Kanaori, K. Tajima and M. Shirakawa, *J. Am. Chem. Soc.*, 2004, **126**, 14023.
- 17 V. L. Pecoraro, A. F. A. Peacock, O. Iranzo and M. Luczkowski, Understanding the Biological Chemistry of  $\text{Hg}^{(II)}$  Using a De NovoDesign Strategy, in *Bioinorganic Chemistry Cellular Systems and Synthetic Models, ACS Symposium Series 1012*, ed. E. Long and M. J. Baldwin, American Chemical Society, Washington, 2009, ch. 12, pp. 183–197.
- 18 G. R. Dieckmann, D. K. McRorie, D. L. Tierney, L. M. Utschig, C. P. Singer, T. V. O'Halloran, J. E. Penner-Hahn, W. F. DeGrado and V. L. Pecoraro, *J. Am. Chem. Soc.*, 1997, **119**, 6195.
- 19 G. R. Dieckmann, D. K. McRorie, J. D. Lear, K. A. Sharp, W. F. DeGrado and V. L. Pecoraro, *J. Mol. Biol.*, 1998, **280**, 897.
- 20 M. L. Zastrow and V. L. Pecoraro, *J. Am. Chem. Soc.*, 2013, **135**, 5895.



21 D. S. Touw, J. A. Stuckey, C. E. Nordman and V. L. Pecoraro, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 11969.

22 M. Matzapetakis, D. Ghosh, T.-C. Weng, J. E. Penner-Hahn and V. L. Pecoraro, *J. Biol. Inorg. Chem.*, 2006, **11**, 876.

23 N. E. Zhou, C. M. Kay and R. S. Hodges, *Biochemistry*, 1993, **32**, 3178.

24 The small reduction following Hg(II) addition to Tb(CS<sub>2-1,4</sub>)<sub>3</sub> may be due to small structural changes to the TbAsp<sub>3</sub>(OH<sub>2</sub>)<sub>3</sub> site, or the emission quenching effect of the Hg(II).

25 W. C. Chan and P. D. White, *Fmoc Solid Phase Peptide Synthesis: A Practical Approach*, Oxford University, New York, 1st edn, 2000.

26 A. F. A. Peacock, G. A. Bullen, L. A. Gethings, J. P. Williams, F. H. Kriel and J. Coates, *J. Inorg. Biochem.*, 2012, **117**, 298.

27 A. Barge, G. Cravotto, E. Gianolio and F. Fedeli, *Contrast Media Mol. Imaging*, 2006, **1**, 184.

28 M. Mantle, G. Stewart, G. Zayas and M. King, *Biochem. J.*, 1990, **266**, 597.

29 J. K. Myers, C. N. Pace and J. M. Scholtz, *Proc. Natl. Acad. Sci. U. S. A.*, 1997, **94**, 2833.