



Cite this: *Environ. Sci.: Water Res. Technol.*, 2020, 6, 779

Selective removal of natural organic matter during drinking water production changes the composition of disinfection by-products†

Anna Andersson, *^a Elin Lavonen,^{bc} Mourad Harir,^{de} Michael Gonsior,^f Norbert Hertkorn,^d Philippe Schmitt-Kopplin,^{de} Henrik Kylin ^{ag} and David Bastviken *^a

Disinfection by-products (DBPs) are potentially toxic compounds formed upon chemical disinfection of drinking water. Controlling the levels and characteristics of dissolved organic matter (DOM) as precursor material for DBPs is a major target to reduce DBP formation. A pilot-scale treatment including suspended ion exchange (SIX®), a ceramic microfilter (CeraMac®) with in-line coagulation and optional pre-ozonation followed by granular activated carbon (GAC) filtration was compared with a conventional full-scale treatment based on DOM removal and DBP formation using Fourier transform ion cyclotron resonance mass spectrometry (FT-ICR MS), rapid fractionation, liquid chromatography organic carbon detection (LC-OCD), adsorbable organic halogens (AOX) and trihalomethane (THM) analysis. The new treatment combination showed different selectivity for DOM removal, compared to the conventional, leading to changes in composition of the DBPs formed. SIX® and GAC had the largest impacts on reducing AOX and THM formation potentials but the high adsorptive capacity of GAC affected the diversity of detected DBPs most. Chlorination and chloramination of pilot treated water with doses normally used in Sweden produced low levels of AOX compared to the full-scale treatment, but FT-ICR MS revealed an abundance of brominated DBP species in contrast with the conventional treatment, which were dominated by chlorinated DBPs. This finding was largely linked to the high DOM removal by the pilot treatment, causing an increased Br⁻/C ratio and a higher formation of HOBr. Potential increases in Br-DBPs are important to consider in minimizing health risks associated with DBPs, because of the supposed higher toxicity of Br-DBPs compared to Cl-DBPs.

Received 18th October 2019,
Accepted 10th January 2020

DOI: 10.1039/c9ew00931k

rs.li/es-water

Water impact

Disinfection by-products (DBPs) are potentially toxic compounds formed during drinking water disinfection. Evaluating treatment processes for improved organic matter removal revealed shifts from Cl-DBPs towards Br-DBPs. Given the higher toxicity of Br-DBPs compared to Cl-DBPs, this work highlights that both qualitative and quantitative aspects should be taken into account for optimal water treatment.

1. Introduction

Disinfection by-products (DBPs) are unwanted and potentially toxic compounds that are formed during drinking water treatment when dissolved organic matter (DOM) reacts with primary and secondary chemical disinfectants, such as chlorine, chlorine dioxide or chloramine.¹ DBPs are of concern due to their potential health risk. Epidemiological studies have shown associations between exposure to chlorinated drinking water and increased risk for bladder cancer.² In addition, other health concerns have been reported, including miscarriages and congenital disabilities.³ Natural organic matter (NOM) together with anthropogenic compounds make up the complex mixture of organic compounds that interacts

^a Department of Thematic Studies – Environmental Change, Linköping University, SE-581 83 Linköping, Sweden. E-mail: anna.e.andersson@liu.se, david.bastviken@liu.se

^b Norrvatten, Kvalitet och Utveckling, SE-169 02 Solna, Sweden

^c Stockholm Vatten och Avfall, SE-168 67 Bromma, Sweden

^d Research Unit Analytical BioGeoChemistry, Helmholtz Centre Munich, Ingolstädter Landstraße 1, 85764 Neuherberg, Germany

^e Chair of Analytical Food Chemistry, Technische Universität München, 85354 Freising, Germany

^f Chesapeake Biological Laboratory, University of Maryland Center for Environmental Science, Solomons, Maryland 20688, USA

^g Research Unit: Environmental Sciences and Management, North-West University, Potchefstroom, South Africa

† Electronic supplementary information (ESI) available. See DOI: 10.1039/c9ew00931k



with the chemical disinfectants, giving rise to several hundreds of different DBP molecules. Disinfectant type, dosage and contact time also have major impacts on the formation of DBPs, as well as pH and temperature.⁴ Furthermore, presence of bromide or iodide in the source water can lead to the formation of bromine- or iodine-containing DBPs.^{5–7} Three different approaches are typically discussed as means of reducing human DBP exposure, *i.e.*, 1) changing the type of disinfectant or optimizing dosage, 2) removing reactive DOM species (precursors) prior to disinfection through upstream water treatment processes or 3) removing the DBPs after their formation through various end-user solutions applied before consumption, *e.g.*, filters based on activated carbon adsorption. The second approach, to improve DOM removal, is attractive for water treatment plants (WTPs) for other reasons than solely to minimize DBP exposure, *e.g.* to avoid saturation of activated carbon filters and bacterial regrowth in the distribution system.⁸ Increasing levels of DOM in source waters have already been observed in northern European lakes and climate change may result in even higher levels.⁹ Hence, some WTPs will need to modify their current treatment processes to maintain high drinking water quality.

To date, coagulation is the major DOM removal treatment for WTPs producing drinking water from surface water sources. Recently, a new treatment technique, suspended ion exchange (SIX®), became available which has the potential of removing DOM more efficiently than coagulation for certain water types.¹⁰ SIX® was developed to remove DOM upstream a ceramic membrane process to reduce fouling of the charged membrane surface and SIX® effectively removes the DOM fraction that is negatively charged at neutral pH.¹⁰ The full process evaluated in this study, combines SIX® with in-line coagulation, ceramic micro-filtration (CeraMac®), ozonation and GAC (granular activated carbon) filtration. In-line coagulation on the CeraMac® membrane removes particulate and high molecular weight DOM, which is not as efficiently removed by SIX®.¹¹ Ozonation in combination with GAC filtration offers additional potential for DOM removal, because ozone breaks down unsaturated organic molecules to oxygenated rather bioavailable compounds that can be consumed by microbial biofilms in the GAC filter downstream. The DOM species that are particularly relevant for the formation of DBPs constitute DOM-specific compositional and structural fractions that are highly reactive with the chlorine-based disinfectants. Non-target screening of the DOM pool, including assessment of formed DBPs, is currently only possible using ultrahigh resolution organic structural spectroscopy. Here, Fourier transform ion cyclotron resonance mass spectrometry (FT-ICR MS) specifically offers detection of thousands of molecular formulae, representing ionisable molecules directly out of mixtures and hence is capable of the discovery of treatment-specific changes in the DOM pool and associated DBP formation. Previous research based on detailed molecular-level analysis investigated the chlorine-DOM reaction^{12–15} and the specific DOM removal by drinking

water treatments, focused on coagulation,^{8,12,14,16,17} sand filtration¹⁶ and nano filtration.^{8,18} This study was undertaken to investigate the individual and combined potential of suspended ion exchange, ozonation, ceramic-microfiltration and GAC filtration to remove specific DOM components and influence DBP formation by using analytical tools with molecular resolution.

2. Methods

2.1 Evaluation approach

DOM removal was evaluated based on typical bulk characteristics, including dissolved organic carbon (DOC), absorbance at 254 nm and the specific absorbance at 254 nm (SUVA). To allow detailed molecular level analysis, Fourier transform ion cyclotron resonance mass spectrometry (FT-ICR MS) was used.¹⁹ To complement the detailed qualitative information provided by FT-ICR MS, DOM analysis based on fractionation techniques, specifically rapid fractionation²⁰ and size-exclusion chromatography using liquid chromatography organic carbon detection (LC-OCD),²¹ was included to provide further size- and composition-related information on DOM removal. Rapid fractionation and LC-OCD were performed according to Chow *et al.* 2004 (ref. 20) and Huber *et al.* 2011 (ref. 21), respectively. Additional information and results from this separate long-term evaluation of the pilot treatment process have been published in a Swedish report.²²

The effects of individual treatment steps, prior to chlorine-based disinfection on DBP formation, were assessed experimentally using the DBP formation potential (DBPFP) test. It is a common approach to determine the DBPFP from different DOM pools by adding chemical disinfectants in excess.²³ With the attempt to assess the mixture of formed DBPs, a non-target evaluation strategy was chosen, using adsorbable organic halogens (AOX) and FT-ICR MS for quantitative and qualitative DBP analysis, respectively. In addition, the commonly monitored trihalomethanes (THMs) were analysed.

2.2 Description of the pilot treatment

The pilot treatment processes evaluated were set up by Norrvatten and Stockholm Vatten och Avfall (SVOA), two drinking water producers that operate three WTPs in Stockholm, Sweden. All three WTPs use raw water from Lake Mälaren, together providing drinking water to about 2 million people in the Stockholm area. The background of this initiative was that Norrvatten and SVOA share several challenges to ensure high-quality drinking water in the future, where one is connected to the increasing and fluctuating levels of DOM in Lake Mälaren.²⁴ The pilot plant was set up at Lovö WTP in 2016. Lovö WTP currently uses coagulation with aluminium sulphate ($\text{Al}_2(\text{SO}_4)_3$), followed by sedimentation and rapid sand filtration, slow sand filtration, UV disinfection and dosing of monochloramine. Fig. 1 shows an overview of the treatment processes of the conventional full-scale plant at Lovö WTP and its connected pilot-



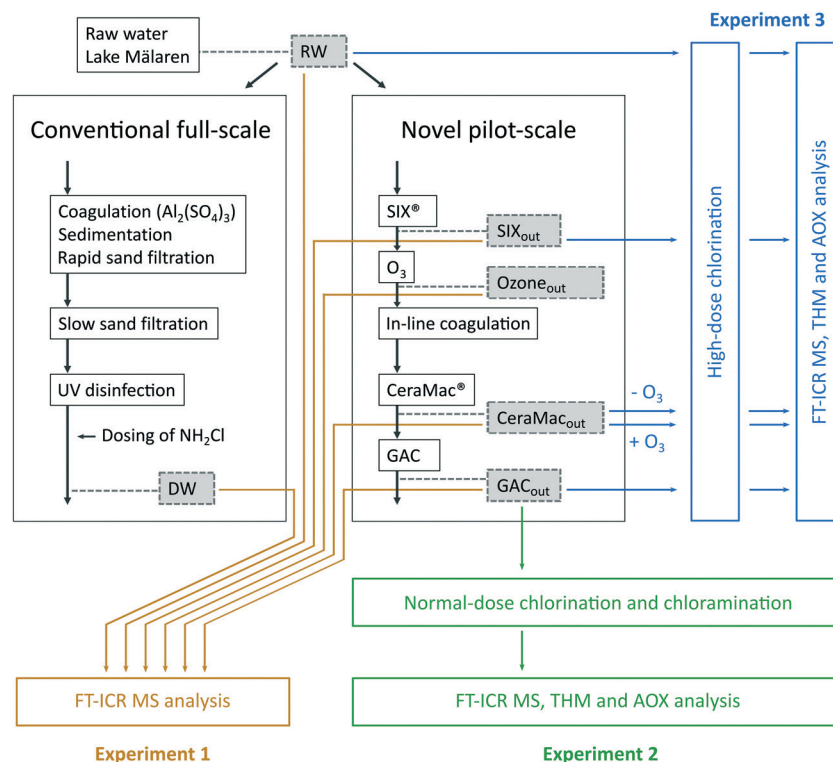


Fig. 1 Set up of the conventional full-scale and the novel pilot-scale treatments at Lovö WTP. Sampling points are highlighted in grey boxes and the experimental design and analysis methods are indicated in coloured boxes. The figure is modified from Lundqvist *et al.* 2019 (ref. 51) in which various bioassays were performed on the same extracts.

scale plant. The pilot plant included five treatment steps (Fig. 1) where the following doses and contact times were used during the sampling period:

1) Suspended ion exchange (SIX®) – a novel technology provided by PWNT Water Technology. In this case, an acrylic quaternary amine (strongly basic) was used as an anionic resin (Lewatit S5128, Lanxess, Germany). The dose and contact time were 15 mL L^{-1} and 30 minutes, respectively and the process was operated at $\text{pH} \approx 7.3\text{--}7.4$.

2) Ozonation, with estimated dose and contact time of $2.5\text{--}3.5 \text{ mg O}_3 \text{ per L}$ and $5\text{--}10$ minutes, respectively.

3) In-line coagulation using PIX-311 (FeCl_3) with the dose 4.5 mg Fe per L .

4) Direct filtration over a ceramic microfilter membrane (CeraMac®) provided by PWNT Water Technology, using a flux of $200 \text{ L m}^{-2} \text{ h}^{-1}$. This ceramic membrane has a pore size of $0.1 \mu\text{m}$ and is produced by Metawater (Japan).

5) GAC (granular activated carbon) (Norit 1240 W) filtration operated at $\text{pH} \approx 7.2\text{--}7.3$ with an empty bed contact time of 20 minutes.

At the time of sampling, the same SIX® resin had been in use and continuously regenerated for about 10 months. However, operational challenges associated with the CeraMac® membrane resulted in shorter operation of the GAC filter (in total about 30 days of continuous operation) at the time of sample collection. The performance of the GAC filter was therefore higher than would be expected for full-scale operation, where the activated carbon oftentimes is saturated and

used as a biological filter (BAC) as the material is not regularly regenerated or replaced.

2.3 Sampling and chlorination experiments

An overview of the three experiments and samples collected at Lovö WTP and the connected pilot plant are presented in Fig. 1. Briefly, experiment 1 focused on the analysis of DOM removal by the individual processes of the pilot treatment, experiment 2 focused on evaluating the formation of DBPs from the pilot treatment at typical disinfectant doses for Swedish WTPs, and experiment 3 aimed to analyse the effects of individual treatment processes on DBP formation potentials. For each sampling point (marked with dashed boxes in Fig. 1), three replicates of water samples (5 L each) were collected in amber glass bottles. The water samples were filtered immediately after sampling using pre-combusted ($450 \text{ }^\circ\text{C}$, 5 h) GF/F glass fibre filters ($0.7 \mu\text{m}$ porosity). The samples marked with yellow in Fig. 1 (experiment 1) were extracted and analysed according to the protocol described in 2.5.1 without further modifications, except for the sample collected after ozonation ($\text{Ozone}_{\text{out}}$) for which 2.5 mL L^{-1} $0.01 \text{ M Na}_2\text{S}_2\text{O}_3$ was added to quench excess ozone (1 mg L^{-1}) at this specific sampling point. In the other experiments, hypochlorite or pre-made monochloramine were added to quantify the potential DBP production. The hypochlorite and monochloramine were taken from Lovö WTP the same day. Immediately after addition, residual levels of hypochlorite and



monochloramine were measured as free chlorine (expressed as Cl_2) and total chlorine (expressed as Cl_2), respectively, using an eXact idip photometer. Free chlorine refers to the combined concentrations of chlorine in the forms of dissolved gas (Cl_2), hypochloric acid (HOCl) and hypochlorite (OCl^-) and total chlorine refers to the sum of free and combined chlorine levels, *i.e.*, including chloramines.

For normal-dose chlorination and chloramination (experiment 2), hypochlorite and monochloramine were added to water samples collected after the GAC filter (GAC_{out}), *i.e.*, to the finished water from the pilot treatment. A dose of 0.3–0.4 mg L^{-1} was used, representing total chlorine residuals typical for disinfected water at Swedish WTPs. The experiment included nine GAC_{out} samples in total, three replicates for control (no addition), three replicates for hypochlorite addition and three replicates for monochloramine addition. After dosing, samples were adjusted to pH ~ 8.5 using 1 M NaOH and were stored for 24 h at 15 °C before extraction and water collection for AOX and THM analysis. The finished water at the conventional full-scale treatment at Lovö WTP (after the addition of monochloramine) was used as a reference (sampling point DW, Fig. 1).

High-dose chlorination (experiment 3) was conducted using hypochlorite only. Hypochlorite was added in excess to a residual of $\sim 10 \text{ mg L}^{-1}$, measured as free chlorine, to water samples from each stage in the pilot treatment (RW, SIX_{out} , $\text{CeraMac}_{\text{out}} - \text{O}_3$, $\text{CeraMac}_{\text{out}} + \text{O}_3$, GAC_{out}). To investigate the effect of coagulant addition on the CeraMac® membrane alone (without ozonation), the sampling point after CeraMac® ($\text{CeraMac}_{\text{out}}$) was collected, both with the ozone turned off ($-\text{O}_3$) and on ($+\text{O}_3$). The ozone levels were monitored by on-line measurements and were also checked manually using the AccuVac MR method (HACH). Ozone residuals were not detected at $\text{CeraMac}_{\text{out}}$ and therefore, no addition of ozone quenching agent was needed. pH was ~ 8.5 (there was no need to adjust pH with NaOH this time) and the samples were kept in the dark at room temperature for 3 days of contact time. Hypochlorite was added to 15 samples (5 sampling points and 3 replicates). As control, one sample for each treatment stage without addition of hypochlorite was included. After 3 days, the free chlorine residuals were above 6 mg Cl_2 per L, for all samples, ensuring that all available, reactive DOM had reacted during the contact time.

2.4 Water analysis

2.4.1 DOC, absorbance and bromide. Dissolved organic carbon (DOC) was measured at the accredited lab of Norrvatten using the non-purgeable organic carbon (NPOC) method (Multi N/C 3100, Analytik Jena). The absorbance at 254 nm (UVA_{254}) was measured on site with a 4 cm cuvette using a HACH Lange DR6000. The specific UV absorbance (SUVA) at 254 nm was determined by dividing the absorbance at 254 nm by DOC and is reported in the unit $\text{L mg}^{-1} \text{m}^{-1}$.²⁵ Bromide was measured using HPLC-ICP-MS at an external accredited lab (method: EN ISO 5667-3:2004).

2.4.2 THMs. One replicate from the normal-dose chlorination and chloramination experiment (experiment 2) and two replicates from the high-dose chlorination experiment (experiment 3) were analysed for THMs. Samples were collected in amber glass bottles including quenching agent (sodium thio-sulphate, provided by Eurofins) and was sent to Eurofins, a Swedac accredited lab, for analysis (method reference LidMiljö.OA.01.16, based on HS-GC-MS).

2.4.3 AOX. Adsorbable organic halogens (AOX) is a method to quantitatively determine the amount of halogenated (chlorine, bromine or iodine) organic compounds that can be adsorbed to activated carbon. AOX was analysed according to ISO standard 9562:2004. Briefly, duplicate samples from the normal- and high-dose experiments (experiments 2 and 3) were quenched with 1 mg L^{-1} and 20 mg L^{-1} sodium thio-sulphate pentahydrate (normapur), respectively, and the samples were analysed in duplicates (two method replicates). For samples with concentrations below 100 $\mu\text{g Cl}_{\text{eq}}$ per L, 100 mL water were analysed. For samples subject to high-dose chlorination, 5–25 mL sample were diluted with Milli-Q to a final volume of 100 mL to not oversaturate the analyser cell (except for GAC_{out} where 100 mL could be used). The water samples were transferred to Erlenmeyer flasks, pH was adjusted to about pH 2 using concentrated HNO_3 , 5 mL acidic nitrate solution (0.02 M HNO_3 , 0.2 M KNO_3) and 50 mg (± 3 mg) activated carbon was added and the flasks were left on a shaking board (180 rpm, 60 minutes). The samples were then filtered through a polycarbonate filter (porosity 0.4 μm , diameter 25 mm, GE Healthcare Life Sciences) retaining the activated carbon with the adsorbed organic compounds. The filters were washed in two steps to remove remaining halide ions, first using an acid nitrate solution (1 mM HNO_3 , 10 mM KNO_3 , 2×10 mL) and then an acidified (using concentrated HNO_3) Milli-Q solution (pH 2, 2×10 mL). Finally, the adsorbed organic compounds were combusted at 1000 °C in O_2 atmosphere (ECS 3000, Thermo Fisher scientific) followed by halide detection through on-line micro coulometric titration. AOX was reported as μg chlorine equivalents per litre ($\mu\text{g Cl}_{\text{eq}}$ per L). However, AOX does not provide information about DBP speciation, *i.e.*, does not distinguish between Cl-, Br- or I-DBPs.

2.5 Detailed organic matter analysis

2.5.1 Solid phase extraction of DOM. Prior to extraction, pH was adjusted to pH 2.5 using 3 M HCl prepared by 32% hydrochloric acid (puriss p.a.) and ultrapure water (spectrophotometric grade). A vacuum manifold (Standard 24-port, 57 250-U, Sigma-A) connected to a peristaltic pump (Vantage 3000 C S 10, Svenska Pump AB) was used to drive the extraction and control the flow rate at 20 mL min^{-1} or below. Bond Elut PPL cartridges (modified styrene divinylbenzene polymer, 1 g sorbent in 6 mL cartridges, Agilent Technologies) were conditioned with methanol (10 mL, LC-MS Ultra Chromasolv®) followed by acidified ultrapure water (10 mL, pH 2.5, spectrophotometric grade) and the packing material



was kept wet at all times. Water samples were connected to the PPL cartridges *via* polytetrafluorethylene (PTFE) tubes. After the extraction, cartridges were washed with 0.1% formic acid water (10 mL, LC-MS Ultra Chromasolv®) to remove any remaining ions (*e.g.*, Cl⁻) that might interfere during FT-ICR MS analysis. Cartridges were dried ~15 seconds with air, eluted with methanol (10 mL, LC-MS Ultra Chromasolv®) and the extracts were collected in amber glass vials (22 mL) and stored in the freezer at -20 °C until FT-ICR MS analysis. To avoid interferences and incorrect interpretations upon FT-ICR MS analysis, no quenching agent was used to quench residual chlorine for the normal- and high-dose chlorination experiments (experiment 2 and 3) prior to extraction. However, water samples subject to high-dose chlorination (experiment 3), containing considerable chlorine residuals, were further acidified to pH 1.5 using 3 M HCl, which shifted the equilibrium towards gaseous chlorine, resulting in the removal of residual chlorine from these samples prior to extraction (free chlorine residuals below 1 mg L⁻¹). Further method discussions regarding sample preparation are provided in the ESI.†

2.5.2 FT-ICR MS analysis. A Bruker Solarix 12 Tesla FT-ICR MS, operating in negative electrospray ionization mode, was used for non-target analysis of DOM and DBPs. The methanol extracts were diluted based on water sample DOC to a concentration of 21 µg mL⁻¹. However, high-dose chlorination (experiment 3) greatly affected ionization selectivity of remaining molecules due to oxidation and degradation of DOM and the larger diversity of ions present after chlorination. Therefore, higher concentrations (150 µg mL⁻¹) were required for optimal detection for these samples. In these cases, extract concentrations were computed based on the dilution factor, extraction recovery and amount of DOC present in the sample before chlorination. Furthermore, extracts from samples collected after GAC filtration (GAC_{out}) were run at the highest concentration possible, based on the extract available. Analysis was performed with a spray current of -3.6 kV and a flow rate of 2 µL min⁻¹. For each mass spectrum, 300 scans were acquired over the mass range *m/z* 147.4 to 1000. Blanks were run every 20 samples and between samples from different experiments (no carry over was observed). Each mass spectrum was internally calibrated using a reference mass list of natural organic molecules often occurring in environmental samples (mass accuracy < 0.2 ppm error). For molecular formulae assignment, a software developed at Helmholtz Center Munich (Germany) was used and the search was limited to the following atoms: ¹²C₀₋₁₀₀, ¹H_{0-∞}, ¹⁶O₀₋₈₀, ¹⁴N₀₋₃, ³²S₀₋₂, ³⁵Cl₀₋₅ and ⁷⁹Br₀₋₅. Further method discussions regarding FT-ICR MS analysis are provided in the ESI.†

2.5.3 Filtration and verification. The data set of assigned molecular formulae was filtered based on intensity (total ion count (TIC) >3 000 000), mass error (<0.2 ppm) and various requirements to eliminate unrealistic formulae based on chemical prerequisites (number of C, H and O > 0, O/C ratio ≤1, H/C ratio ≤2.5 and double bond equivalences (DBE) ≥ 0). In addition, the so-called nitrogen rule was applied.²⁶ To identify chlorine and bromine containing DBPs, a filtration

approach was used where the ³⁷Cl or ⁸¹Br isotope variants of each chlorine (³⁵Cl) or bromine (⁷⁹Br) containing molecular formula were used to verify each DBP. This verification process has been further described elsewhere.²⁷ For the analysis of DOM removal, formulae containing carbon, hydrogen, oxygen, nitrogen (≤1) and sulfur (≤1) were considered but DBP assessment was limited to halogenated formulae with carbon, hydrogen and oxygen only. In the text, molecular formulae containing carbon, oxygen and hydrogen atoms are referred to as CHO formulae, without specifying the number of individual atoms. However, the number of chlorine and/or bromine atoms are specified, *e.g.*, CHOCl₂ refers to a molecular formula with carbon, oxygen, hydrogen and 2 chlorine atoms.

2.5.4 Data processing and analysis. Data analysis was performed using molecular formulae that were detected in all three replicates. To study DOM removal (experiment 1), changes in relative abundance of individual molecular formulae present before and after a treatment were compared between the three replicates. Calculations of average elemental compositions and indices were weighted against the average relative abundance of the three replicates. An exception from this routine were two samples from the high-dose chlorination study (experiment 3), CeraMac_{out} - O₃ and CeraMac_{out} + O₃, where common formulae and the relative intensities of two replicates were used. For normal and high-dose chlorination (experiment 2 and 3), verified DBPs present in the chlorinated or chloraminated samples were analysed unaltered, *i.e.*, halogenated CHO formulae present in the control were not subtracted. The reason was to obtain equivalent comparisons with the finished water at Lovö WTP (experiment 2) and because the dilution and ionization was very different between the chlorinated samples and the controls (experiment 3). For data analysis, indices, created to further characterize the organic molecules, were computed based on the elemental composition of individual formulae, including double bond equivalences (DBE), DBE normalized to the number of carbon atoms (DBE/C), the average carbon oxidation state (C_{OS}) and a modified aromaticity index (AI_{mod}), which have been explained in greater detail elsewhere.^{27,28}

3. Results and discussion

3.1 DOM removal by the pilot treatment (experiment 1)

3.1.1 Basic water characteristics. The pilot treatment had a large effect on DOM, both in terms of quantitative removal (*e.g.*, DOC) and on the relative abundance of DOM fractions with various structural characteristics (*e.g.*, SUVA). While the conventional treatment led to a DOC reduction from 6.9 to 3.9 mg L⁻¹, the pilot treatment resulted in a DOC concentration of 0.4 mg L⁻¹ in the GAC filtrate (Table 1). Primarily, the SIX® and GAC processes contributed to these changes. SIX® lowered DOC by 4.4 mg L⁻¹, whereas the conventional treatment lowered DOC by 3 mg L⁻¹. In-line coagulation and CeraMac® lowered DOC with 0.6 mg L⁻¹ and GAC resulted in a 1.6 mg L⁻¹ DOC decrease.



Table 1 Basic water characteristics including DOC, UVA₂₅₄, SUVA and AOX of water samples collected from the conventional full-scale and novel pilot-scale treatments (experiment 1). NA = not available

| | Sample collected | DOC (mg L ⁻¹) | UVA ₂₅₄ (cm ⁻¹) | SUVA (L mg ⁻¹ m ⁻¹) | AOX (µg Cl _{eq} per L) |
|-------------------------|------------------------|---------------------------|--|--|---------------------------------|
| Conventional full-scale | RW | 6.9 | 0.191 | 2.8 | 19 |
| | Lovö DW | 3.9 | 0.077 | 2.0 | 55 ^a |
| Novel pilot-scale | RW | 6.9 | 0.191 | 2.8 | 19 |
| | SIX _{out} | 2.5 | 0.048 | 1.9 ^b | 8.7 |
| | Ozone _{out} | 2.6 | 0.032 | 1.2 | NA |
| | CeraMac _{out} | 2.0 | 0.013 | 0.7 | NA |
| | GAC _{out} | 0.4 | 0.001 | 0.3 | <4 |

^a Lovö DW is chloraminated. ^b Long-term monitoring showed slightly higher SUVA values, reported in section 3.1.1.

Apart from lowering DOC, the pilot treatment also decreased UVA₂₅₄ and SUVA, which indicate that the treatment is effective in removing UV-absorbing compounds, *e.g.*, aromatic compounds. SUVA values after SIX® and the conventional treatment were similar at the time of sampling (Table 1). However, long-term monitoring of the pilot and conventional process show slightly different average SUVA values (2.4 ± 0.3 L mg⁻¹ m⁻¹ ($n = 98$) after SIX® and 1.8 L mg⁻¹ m⁻¹ after coagulation, average based on 2 years of monitoring), indicating that coagulation has an overall higher selectivity towards removing UV-absorbing compounds compared to SIX®. This is in line with previous findings.²⁹ No quantitative removal of DOM was observed upon ozonation, however, UVA₂₅₄ and SUVA decreased, highlighting changes in DOM characteristics. In-line coagulation over the

CeraMac® membrane (where the main effect on DOM is attributed to in-line coagulation²⁹) and GAC further decreased UVA₂₅₄ and SUVA, reflecting a selectivity towards UV-absorbing compounds also in these processes, resulting in almost blank absorbance at 254 nm after GAC. The level of AOX in Lake Mälaren was 19 µg Cl_{eq} per L (Table 1). That corresponds to the lower AOX range in a study based on 135 lakes in southern Sweden, where AOX varied between 11–185 µg Cl_{eq} per L and levels between 20–60 µg Cl_{eq} per L were most common.³⁰ AOX was decreased with approximately 50% during SIX® treatment (Table 1).

3.1.2 FT-ICR MS, rapid fractionation and LC-OCD. CHO-Type molecules were most abundant (58–77%) and the average nitrogen and sulphur content was between 0.1–0.2% at all treatment stages (Table 2). However, CHNOS-type

Table 2 Counts of *m/z* ions as computed from negative electrospray 12T FT-ICR mass spectra for singly charged ions reported in neutral form (the mass of a proton added). Only molecular compositions present in three replicates were included and the relative abundance was computed based on the average relative intensity of the three replicates. For definition of column heads, see Fig. 1. For SIX_{out} and Ozone_{out} the percentage of DOM compositions deviates from 100 due to rounding

| Experiment 1 | | | | | | |
|---|-------|--------------------|----------------------|------------------------|--------------------|---------|
| | RW | SIX _{out} | Ozone _{out} | CeraMac _{out} | GAC _{out} | Lovö DW |
| Number of DOM compositions | | | | | | |
| Total | 2731 | 2856 | 2841 | 2398 | 1841 | 2371 |
| CHO | 1823 | 1914 | 1649 | 1403 | 1426 | 1398 |
| CHNO | 528 | 570 | 563 | 594 | 334 | 525 |
| CHOS | 375 | 362 | 609 | 386 | 81 | 444 |
| CHNOS | 5 | 10 | 20 | 15 | 0 | 4 |
| Percentage of DOM compositions | | | | | | |
| CHO [%] | 66.8 | 67.0 | 58.0 | 58.5 | 77.5 | 59.0 |
| CHNO [%] | 19.3 | 20.0 | 19.8 | 24.8 | 18.1 | 22.1 |
| CHOS [%] | 13.7 | 12.7 | 21.4 | 16.1 | 4.4 | 18.7 |
| CHNOS [%] | 0.2 | 0.4 | 0.7 | 0.6 | 0.0 | 0.2 |
| Characteristics of DOM compositions | | | | | | |
| Average H [%] | 42.8 | 45.3 | 45.3 | 45.4 | 43.9 | 44.5 |
| Average C [%] | 37.2 | 36.8 | 36.1 | 35.8 | 35.9 | 36.8 |
| Average O [%] | 19.9 | 17.7 | 18.3 | 18.6 | 20.1 | 18.4 |
| Average N [%] | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 |
| Average S [%] | 0.1 | 0.1 | 0.2 | 0.1 | 0.1 | 0.1 |
| Computed average H/C ratio | 1.15 | 1.23 | 1.25 | 1.27 | 1.22 | 1.21 |
| Computed average O/C ratio | 0.53 | 0.48 | 0.51 | 0.52 | 0.56 | 0.50 |
| Computed average N/C ratio | 0.003 | 0.003 | 0.003 | 0.003 | 0.002 | 0.003 |
| Computed average S/C ratio | 0.003 | 0.003 | 0.005 | 0.003 | 0.002 | 0.004 |
| Average carbon oxidation state (C _{os}) | -0.07 | -0.25 | -0.23 | -0.22 | -0.10 | -0.20 |
| Average DBE | 9.17 | 8.60 | 8.03 | 7.65 | 8.59 | 8.58 |
| Average DBE/C | 0.48 | 0.44 | 0.43 | 0.42 | 0.44 | 0.45 |
| Average Al _{mod} | 0.28 | 0.26 | 0.23 | 0.21 | 0.22 | 0.26 |
| Mass weighted average [Da] | 419.0 | 414.5 | 405.6 | 393.7 | 434.4 | 407.3 |



molecular compositions were rarely detected. Weighted elemental compositions and indices (DBE, DBE/C, AI_{mod} and C_{OS}) for CHO, CHNO, CHOS and CHNOS molecular compositions, calculated for each group individually, are presented in Table S1.† Fig. 2 and S5† show the molecular formulae that decreased in relative abundance by more than 50% after each process.

DOM compositions detected after SIX® showed higher hydrogen and lower oxygen content compared to the raw water, indicating a selective removal of unsaturated oxygen rich compounds (Table 2). Oxygenated molecules of a wide mass range (250–650 m/z) (Fig. 2) and a high average carbon oxidation state (Fig. S5†) decreased in relative abundance after SIX®, mainly because molecules with oxygen containing functional groups are likely to be deprotonated and charged and therefore easily removed with ion exchange. A separate rapid fractionation experiment (Fig. 3) further showed that SIX® removed large portions of the very and slightly hydrophobic acids (VHA, SHA), and all the hydrophilic charged (CHA) molecules.

Ozonation selectively transformed DOM in the mass range of 600–750 Da (Fig. S1 and S3†), which resulted in a decrease in average mass (Table 2). Molecular compositions that decreased in relative abundance upon ozonation had low H/C and O/C ratios (Fig. 2), indicating that ozone preferably reacted with aromatic compounds and/or highly unsaturated molecules (compounds with multiple double bonds). This is in line with previous findings where electrophilic addition of ozone to unsaturated bonds has been identified as the main reaction pathway between ozone and DOM, leading to the formation of aldehydes, ketones and carboxylic acids.^{31,32} Interestingly, the average carbon oxidation state was rather unchanged after ozonation (Table 2). This is explained by the formation of oxygen-rich end products, like short chained aldehydes and carboxylic acids and finally CO_2 and H_2O , not captured with FT-ICR MS.^{32,33} Hence, the oxidation of freshwater DOM leads to a defunctionalisation of the remaining, extractable DOM.

In-line coagulation and CeraMac® (where in-line coagulation, rather than CeraMac® has the major effect on DOM) further removed high molecular weight DOM (Fig. S1 and S3,† Table 2) including very and slightly hydrophobic acids (Fig. 3), unsaturated, oxygenated CHO-molecules (van Krevelen diagram, Fig. 2 and DBE, AI_{mod} and C_{OS} plots, Fig. S5†) and rather saturated CHOS-molecules (van Krevelen diagram, Fig. 2 and DBE and AI_{mod} plots Fig. S5†). The pronounced removal of CHOS-molecules observed by in-coagulation and CeraMac® might be linked to the higher abundance of CHOS-molecules detected before this treatment (Table 2), as the number of detected CHOS formulae were higher after ozonation compared to the other treatment stages.

The effect of GAC on DOM compositions requires careful evaluation. Before sample collection, the GAC filter had been operated a limited amount of time (approximately 30 days of continuous operation), which means that the GAC filter still had a substantial adsorptive capacity, which is in contrast to

many full-scale WTPs in Sweden, where activated carbon filters are often operated as biological filters only, *i.e.*, as BAC filters. We therefore expect that the GAC filter in this pilot study removed higher DOC levels than would be economically feasible in a full-scale BAC application. FT-ICR MS indicate that GAC, at a stage where removal was dominated by adsorption, was selective in removing molecules of mid to high H/C ratios (1–1.7) (Fig. 2 and S3†). Also, less oxygenated compounds were selectively removed while the average mass increased after GAC (Fig. S5,† Table 2), suggesting a selective removal of low molecular weight compounds of low oxidation state. This is in line with previous studies where small, neutral hydrophobic molecules were preferentially removed after GAC operated through adsorption.^{31,34} The fractionation experiment showed that the major components found after GAC constituted neutral hydrophilic compounds (Fig. 3). Consequently, while we expect to capture a majority of DOM upon extraction with Bond Elut PPL and subsequent FT-ICR MS analysis, the largest fraction of DOM remaining after GAC, may not have been efficiently captured. However, overall amounts of DOC remaining after GAC were low.

The pilot treatment was compared with finished water from the full-scale conventional treatment at Lovö, where coagulation has the highest impact on DOM abundance and composition (Table 2; Fig. 2 and S3–S5†).¹⁶ The selectivity of the conventional treatment partly resembled that of SIX® (Fig. 2), which likely can be linked to the removal of humic substances of both treatments.²⁹ However, the conventional process selectively removed aromatic DOM (Fig. 2) of higher molecular weight (Table 2), when compared to SIX®, which is consistent with previous findings.²⁹ Previous studies based on FT-ICR MS showed that coagulation was selective to compounds of H/C and O/C ratios of 0.4–1.4 and 0.4–0.9, respectively, which were attributed to polyphenol-like DOM, resembling our findings.^{12,17}

Analysis of DOM fractions using LC-OCD were performed to further compare the selectivity between SIX® and coagulation. In general, LC-OCD showed that SIX® is less selective compared to coagulation (Fig. 4). Coagulation removed a part of the early eluting fraction constituting larger molecules, described as biopolymers in the method defined by Huber *et al.* 2011,²¹ which were not affected by SIX® (Fig. 4). Furthermore, coagulation showed higher selectivity to remove larger humic substances (shorter retention times), while SIX® removed medium-sized humic substances (longer retention times) more effectively compared to coagulation. Finally, SIX® affected the removal of building blocks and low molecular weight humic substances to a higher extent than coagulation (Fig. 4). These differences in selectivity are in agreement with previous work,^{10,29} but variations in selectivity have been observed for different raw waters depending on DOM characteristics.²⁹

The differences in selectivity can be linked to the interactions and forces involved in these two processes, respectively. Ion exchange primarily removes DOM based on charge, while



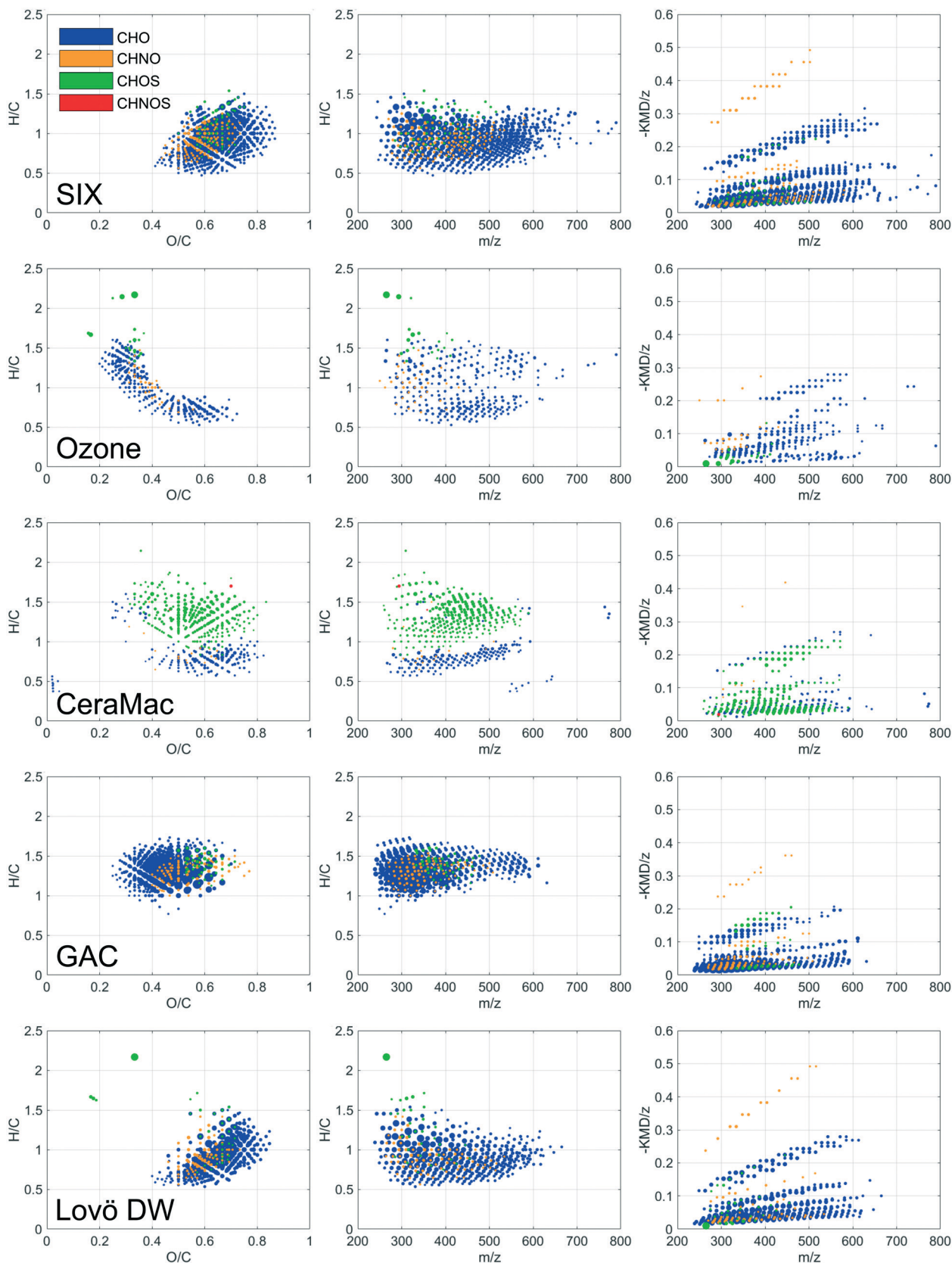


Fig. 2 Van Krevelen diagrams (left panel), mass edited H/C ratios (middle panel) and modified Kendrick mass plots (right panel) of molecular compositions that decreased in relative abundance by more than 50% after each pilot treatment process. Comparisons were made for compositions detected in three replicates with final water from the conventional treatment at Lovö used as a reference (Lovö DW).



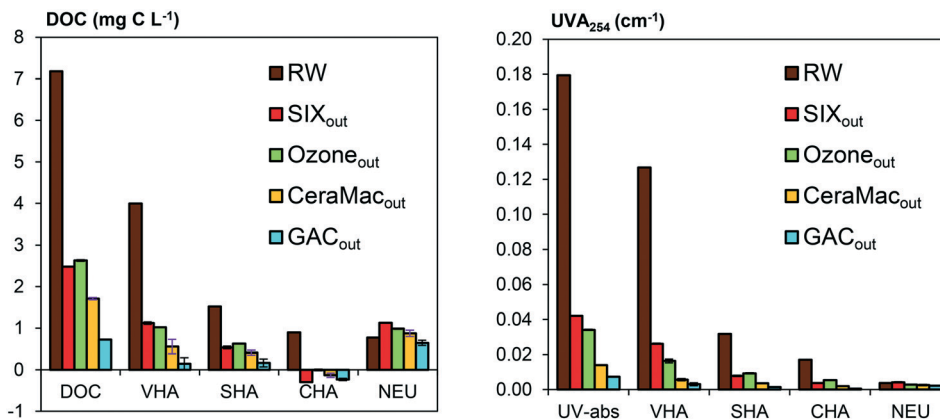


Fig. 3 Distribution of very hydrophobic acids (VHA), slightly hydrophobic acids (SHA), hydrophilic charged (CHA) and hydrophilic neutral (NEU) compounds at different stages of the pilot treatment analyzed by rapid fractionation.

the coagulation process also depends on the hydrophobicity of DOM and specific structural characteristics, such as the number and positions of carboxyl and hydroxyl groups, affecting the complexation reaction with the coagulant salt to form larger aggregates.^{35,36} Concerning size, a large molecular size leads to decreased charge density, which affect the removal efficiency of ion exchange.³⁷ The size of DOM also affects the propensity of a molecule to enter the ion exchange resin pores, *e.g.*, the gel type resins (the type used in this study) typically has smaller pores compared to macroporous resins.^{38,39} For coagulation, large DOM molecules are instead favoured for removal, partly because molecules of high molecular weight likely have higher hydrophobicity and abundance of carboxylic groups, leading to increased aggregation through hydrophobic interactions and electrostatic forces, respectively.^{35,40} Furthermore, high-molecular weight compounds can adsorb to multiple other particles, linking them together and thereby increase aggregation during coagulation.⁴⁰

3.2 Normal-dose chlorination and chloramination (experiment 2)

3.2.1 THM and AOX formation. The level of AOX following chlorination and chloramination of water collected after the pilot treatment (GAC_{out}), was 21 and 6.4 $\mu\text{g Cl}_{\text{eq}}$ per L respectively, which can be compared with 55 $\mu\text{g Cl}_{\text{eq}}$ per L for the conventional full-scale treatment at Lovö WTP, where monochloramine is dosed prior to distribution (Fig. S6[†]). The only measurable levels of THMs were found after chlorination of pilot treated water, but at total levels of 5.5 $\mu\text{g L}^{-1}$ (Table 3), well below regulatory limits.⁴¹ It is known that chloramine produces less amounts of THMs compared to chlorine, because of its less oxidizing power to produce so-called end-DBPs, such as THMs.^{42,43} Importantly, the pilot treated water could be both chlorinated and chloraminated with limited AOX formation and without exceeding regulatory limits of THMs.

3.2.2 DBP formation detected by FT-ICR MS. Fig. 5 shows the detected verified DBPs from the normal-dose experiment that were present in all three replicates, visualized using van Krevelen diagrams, H/C to mass-to-charge (m/z) and modified Kendrick mass plots. Plots based on DBE, AI_{mod} and C_{OS} are shown in Fig. S7[†] and a table with computed weighted averages is presented in Table S2[†]. The finished water from Lovö WTP (sampling point DW, experiment 1) showed a diversity of CHOCl , CHOCl_2 and CHOBr DBP compositions, with O/C, H/C and m/z distributions similar to those observed in other WTPs, subject to DBP assessments using FT-ICR MS.^{12,14,17,27} In these previous studies, CHOCl_2 type DBPs have primarily been detected in waters subjected to chlorination, but in this experiment no such DBP species were found after chlorination of the pilot treated water, indicating that the precursors had a more important role than disinfectant type for DBP speciation in this case. The Kendrick mass plots (Fig. 5) showed that a group of DBPs with high-KMD/ z , indicative of high oxygenation, was detected after chlorination and chloramination of pilot treated water, but not in Lovö drinking water, also showing that the pilot treatment changed the diversity of DBPs formed. Particularly, the pilot treatment shifted DBP

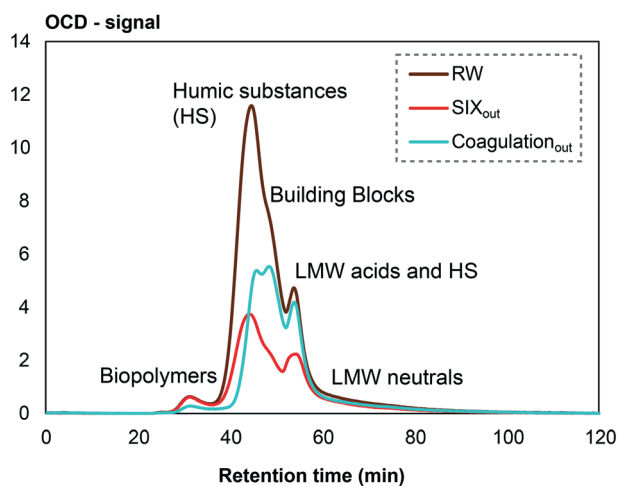


Fig. 4 Selectivity of pilot-scale SIX[®] compared to full-scale conventional coagulation and rapid sand filtration analyzed by LC-OCD. LMW = low molecular weight.



Table 3 THM concentrations after normal-dose chlorination (HOCl) and chloramination (NH₂Cl) of finished water (GAC_{out}) of the pilot treatment (experiment 2; note that only water after GAC filtration were studied). Finished water from the conventional full-scale treatment at Lovö (Lovö DW), where chloramine is used, was included as a reference

| Experiment 2 | Pilot treatment | | Pilot treatment | | Lovö DW | |
|----------------------|-----------------------------|--|-----------------------------|--|-----------------------------|--|
| | HOCl | | NH ₂ Cl | | NH ₂ Cl | |
| | Conc. (µg L ⁻¹) | | Conc. (µg L ⁻¹) | | Conc. (µg L ⁻¹) | |
| Trichloromethane | <1.0 | | <1.0 | | <1.0 | |
| Bromodichloromethane | <1.0 | | <1.0 | | <1.0 | |
| Dibromochloromethane | 1.6 | | <1.0 | | <1.0 | |
| Tribromomethane | 2.8 | | <1.0 | | <1.0 | |
| Total THMs | 5.5 | | <4.0 | | <4.0 | |

formation towards brominated species. DBP components in samples collected after GAC with subsequent chlorination and chloramination were dominated by CHOBr compositions, which composed 73% and 93% of all formulae respectively (Fig. 5). In addition, the CHOBr DBPs formed af-

ter the pilot treatment were distributed across a wider chemical space, *e.g.*, at higher O/C ratios and at higher molecular weights, compared to the CHOBr DBPs detected in Lovö drinking water. Few Cl-DBPs were detected in the pilot treated water after chloramine addition, compared to



Fig. 5 Van Krevelen diagrams (left panel), mass edited H/C ratios (middle panel) and modified Kendrick mass plots (right panel) of the verified chlorine and bromine containing molecular formulae (neutral form) present after normal-dose chlorination (HOCl) and chloramination (NH₂Cl) of finished water from the pilot process (GAC_{out}) using final treated water from Lovö DW (Lovö DW) as a reference (experiment 2). Presented DBPs are limited to formulae detected in all three replicates.



after chlorine addition (Fig. 5). This is likely due to the different reactivities of chloramine and chlorine with DOM.⁴⁴

The diversity of available precursors was greatly affected by the pilot treatment (as described in section 3.1), and the removal of large quantities of DOM affected the Br⁻/DOC ratio, which also explains the impact of the pilot treatment on DBP speciation. SIX® removed 34 ± 10% bromide (from 56 to 37 µg L⁻¹) through sorption to the ion exchange resin but the even higher decrease of DOC (63 ± 3%, computed based on average DOC removal during the entire test period) still resulted in a net increase of the Br⁻/DOC ratio from 0.008 to 0.014 ± 0.003 mg Br⁻/mg DOC. There was no significant effect on bromide levels after ozonation, coagulation or GAC filtration (demonstrated by separate experiments performed at Norrvatten). This is why we expect that the bromide level remained rather constant after SIX®, resulting in a final Br⁻/DOC ratio of about 0.07 mg Br⁻/mg DOC after GAC. Of the different processes, GAC likely had the greatest impact on the Br⁻/DOC ratio, due to the decreased DOC and unchanged Br⁻ levels and therefore likely contributed most to the observed shift towards higher relative abundances of Br-DBPs. According to the experiments described above, the conventional full-scale treatment was not expected to have a significant effect on bromide.

3.3 High-dose chlorination (experiment 3)

3.3.1 THM and AOX formation potentials. Of the four major processes of the pilot treatment, THM formation potential (THMFP) and AOX formation potential (AOXFP) were primarily lowered through SIX® and GAC filtration (Fig. 6). The reduction by SIX® was 66% and 54% for THMFP and AOXFP, respectively. After GAC, THMFP and AOXFP had been lowered by 97% and 96% compared to the raw water, respectively, resulting in a low DBP formation potential in the finished water. Both THMFP and AOXFP were strongly correlated with DOC, UVA₂₅₄ and SUVA (R^2 values of 0.84 to 0.99; Fig. S8 and S9†), which re-

lates to the general dependency of organic matter substrates. The relationship with DOC was the strongest (R^2 0.98–0.99) because the treatment processes affecting UVA₂₅₄ and SUVA, but not DOC, *e.g.*, ozonation, did not show a clear effect on THMFP and AOXFP (Fig. 6). We expect that the effect of ozonation on DBP formation potentials would have been more pronounced if UVA₂₅₄ and SUVA were higher at the point of ozonation, because the efficiency of ozonation to destroy DBP precursors has been related to SUVA.⁴⁵ This means that ozonation, due to its reactivity, has a larger impact on DOM modification and consequently the formation of DBPs when SUVA is high.

The patterns of THMFP and AOXFP largely corresponded, but SIX® decreased THMFP to a higher extent, compared to AOXFP (Fig. S10†). This indicates that SIX® more efficiently removed precursors associated with the formation of THMs, *i.e.*, hydrophilic low molecular weight compounds^{46,47} – the selective SIX®-induced removal of medium-sized humic substances, building blocks and low molecular weight humic substances was specifically shown in the LC-OCD fractionation experiment (Fig. 4). Previous research has found that coagulation has a substantially higher impact on reducing the formation potential of haloacetic acids (HAA) compared to SIX®, which can be related to the more efficient removal of aromatic precursors by coagulation, while the effect on THMFP was rather similar.²⁹ Thus, the strength of correlations between AOX and specific DBP species will depend on the DOM characteristics of individual waters and the disinfectant type employed, which both affect DBP speciation.^{42,48}

The formation potential per mg carbon (AOXFP/C or THMFP/C), show the relative DOM reactivity, and was primarily reduced through GAC filtration (Fig. 6), demonstrating a selective removal of THM and AOX precursors by this treatment under prevailing conditions. Changes in THMFP/C and AOXFP/C between processes were slightly different, *e.g.*, THMFP/C decreased while AOXFP/C increased after SIX®, further emphasizing that SIX® selectively removes THM reactive precursors.

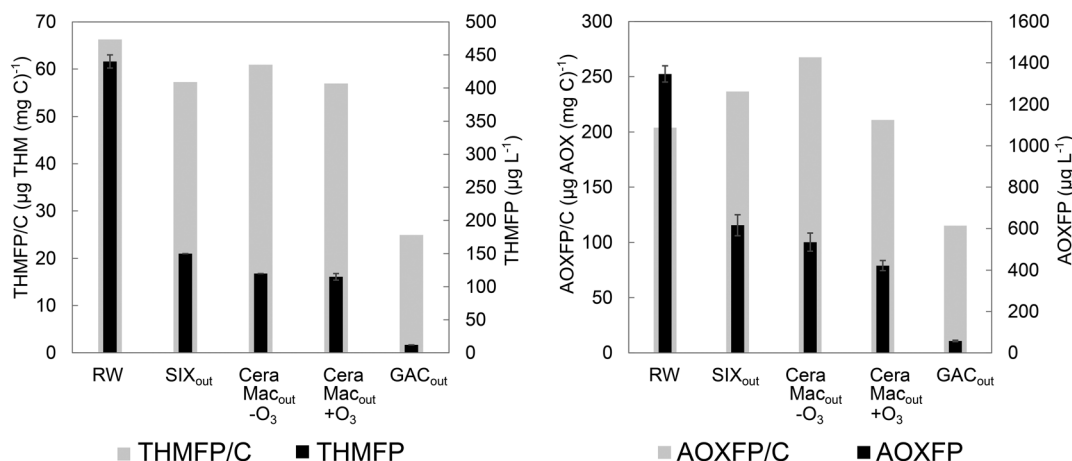


Fig. 6 THM formation potential (THMFP), AOX formation potential (AOXFP), THM formation potential per mg carbon (THMFP/C) and AOX formation potential per mg carbon (AOXFP/C) upon chlorination (~10 mg L⁻¹ chlorine residual) at different stages of the pilot treatment (RW, SIX_{out}, CeraMac_{out} - O₃, CeraMac_{out} + O₃, GAC_{out}) (experiment 3).



The relative formation of bromine containing DBP species (Br-DBPs) increased along the treatment stages and especially after GAC (Fig. 7). However, the high chlorine residual drives the formation towards Cl-DBPs and therefore this pattern was even more distinct when chlorination was performed at a normal dose (experiment 2), when tribromomethane and dibromochloromethane were the only THM species detected after GAC (Table 3). In addition to the increased Br⁻/DOC ratio (discussed in 3.2.2), this shift might be explained by UVA₂₅₄ and SUVA reductions caused by the pilot treatments. Bromine substitution has been found to decrease with increasing SUVA for a range of DBP classes.⁴⁹ This is because chlorine substitution reactivity is associated with aromatic, hydrophobic DOM (which is related to SUVA), while bromine substitution reactivity is associated with carboxylate-rich hydrophilic DOM.⁴⁵

3.3.2 DBP formation potentials detected by FT-ICR MS.

High-dose chlorination (experiment 3) induced the formation of many DBPs (>1000 verified DBP compositions for chlorinated raw water) including CHOCl, CHOCl₂, CHOCl₃, CHOCl₄ and CHOCl₅ compositions. Overall, DBP species with multiple chlorine atoms were distributed towards higher O/C and lower H/C ratios, had higher aromaticity (AI_{mod}) and fewer number of carbon atoms. This trend could be followed from CHOCl to CHOCl₄ formulae (Fig. 8 and S11†), showing that multiple chlorine substitution is linked to certain precursor characteristics, especially unsaturated, and highly oxygenated DOM compositions. As for THMs, the formation of brominated species was limited due to the high chlorine residuals.

At a first glance, the distribution of DBPs formed upon chlorination of raw water and after SIX® appeared overall comparable as shown in van Krevelen diagrams (Fig. 8). However, the computed weighted average elemental ratios and indices revealed higher H/C ratios and lower O/C ratios as well as lower DBE and AI_{mod} among DBPs formed after SIX® (Table S3†). This matches with the observation that SIX® selectively removes DOM compositions of high O/C and low H/C ratios (Fig. 2). However, the weighted average *m/z* did not change notably (Table S3†). In-line coagulation and CeraMac® limited the *m/z* distribution of DBPs formed (Fig. 8) and the average *m/z* was shifted from 358 to 340 Da

(Table S3†), further demonstrating that in-line coagulation preferentially removes high molecular weight precursors.

No changes in DBE, AI_{mod} or average mass of verified DBPs after ozonation were observed (Table S3†). GAC filtration showed the largest impact on the formation of FT-ICR MS amenable DBPs. After GAC, the detected DBPs appeared in a more restricted chemical space in the van Krevelen diagram (DBPs with O/C below 0.4 or H/C below 1.0 were rarely found) and less DBPs with three, four and five chlorines were formed, in favor of mixed chlorine–bromine DBPs (Fig. 8).

By comparing the individual DBP formulae formed after chlorination at different treatment stages (RW, SIX_{out}, CeraMac_{out} - O₃, CeraMac_{out} + O₃, GAC_{out}, Fig. 1) in a Venn diagram (Fig. 9), it is possible to distinguish between unique DBPs, *i.e.*, DBP formulae formed after a single treatment, and shared DBPs, *i.e.*, DBP formulae formed after more than one treatment stage. 173 DBPs were detected following chlorination at all stages, which means that the precursors producing these DBPs remained throughout the pilot treatment. This group of DBPs comprised one and two chlorine containing DBPs with H/C ratios above one, in a mass range of 270–400 Da. The largest group of common DBPs (266 formulae) was shared between all treatment stages except GAC, further emphasizing that GAC had the biggest impact on DBP diversity of the individual treatment processes. DBPs detected after chlorination of raw water only were dominated by unsaturated compounds, underlining that SIX® treatment removed this group of precursors.

3.4 Outlook and summary

The connection between selective precursor removal and DBP formation is not straightforward.⁵⁰ Bond *et al.* (2010) showed that DOM constituents of similar physiochemical properties (that were equally removed during treatment) could have completely different DBP formation potential.⁵⁰ Therefore, DBP formation potential tests remain important to evaluate effective removal of DBP precursors. In the present study, patterns of selective DOM removal by the treatments and subsequent DBP formation could be linked *via* ultrahigh-resolution mass spectrometry, but the different patterns of



Fig. 7 Percentage of different THM species formed during high-dose chlorination after each stage of the pilot treatment (RW, SIX_{out}, CeraMac_{out} - O₃, CeraMac_{out} + O₃, GAC_{out}), based on the average of two replicates (experiment 3). Tribromomethane was not detected.



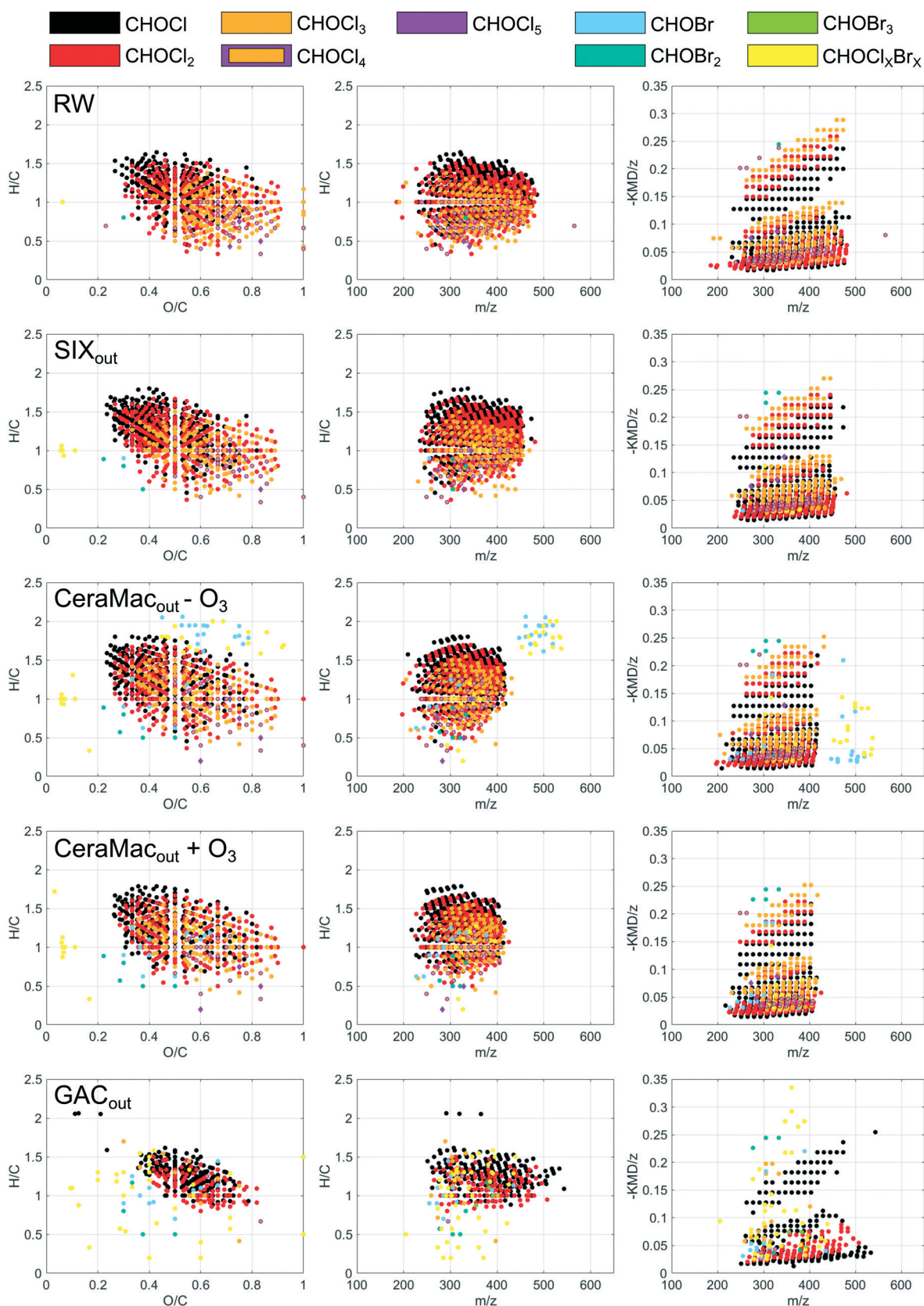


Fig. 8 Van Krevelen diagrams (left panel), mass edited H/C ratios (middle panel) and modified Kendrick mass plots (right panel) of the verified chlorine and bromine containing molecular formulae (neutral form) present after high-dose chlorination (experiment 3) at different stages of the pilot treatment (RW, SIX_{out}, CeraMac_{out} - O₃, CeraMac_{out} + O₃, GAC_{out}).





Fig. 9 Venn diagram showing the number of unique and shared verified DBPs formed upon high-dose chlorination at different stages of the pilot treatment (RW, SIX_{out}, CeraMac_{out} - O₃, CeraMac_{out} + O₃, GAC_{out}) (experiment 3).

THM and AOX formation highlight that the selective DOM removal affect the formation of various DBP groups differently.

The challenge remains in distinguishing the most relevant precursor groups to remove. Therefore, the extracts chemically analysed in the present study were also analysed with various bioassays to measure the toxic effects from the mixture of DBPs formed.⁵¹ The DBPs were activating an oxidative stress response system (the Nrf2 pathway), which is an early cell response that can cause, *e.g.*, genotoxicity.⁵² The results from the high-dose chlorination experiment showed that SIX® reduced the Nrf2 induced activity of formed DBPs by ~70%, which is similar to the decreases in THM formation potentials. However, the treatments that solely reduced UVA₂₅₄, *e.g.*, ozonation, affected Nrf2 induced activity more than AOX and THM formation potentials.⁵¹ The drastic decrease in THM and AOX formation after GAC due to the high DOC removal did not resonate with a similar drop in Nrf2 induced activity from DBPs formed. We suggest that this observation can be explained by the shift towards higher relative abundances of more toxic Br-DBPs after GAC, observed through both THM and non-target FT-ICR MS analysis in this study. The following statements summarize our findings:

- Quantity matters 1: overall, the largest attenuation of DBP formation potential was associated with decreased DOC concentration, which was also reflected by UVA₂₅₄ and SUVA reductions.

- Quantity matters 2: SIX® and GAC were the treatments that had the highest effect on decreasing THM and AOX formation potentials, but the formation potential per mg DOC was remarkably similar after all treatment steps except GAC (with a relatively new filter).

- Quality matters 1: different treatments showed distinct selectivity in DOM removal, regarding *e.g.* molecular weight and hydrophilic/hydrophobic properties, which together with the Br⁻/DOC ratio, affected the types of DBPs formed.

- Quality matters 2: genotoxic responses are not necessarily associated with the quantitative sum of DBP formation and toxic effects can be associated with qualitative rather than quantitative properties of DBPs.

4. Conclusions

This study implies that qualitative aspects of DBP formation, *i.e.*, which DBPs that are formed, can be highly affected by increased DOM removal during drinking water treatment. Because DBP speciation influences toxicological responses, qualitative aspects of DBP formation need to be considered in water quality assessments. Furthermore, the interplay between quantitative and qualitative aspects are important, *e.g.*, for experimental design and subsequent evaluations. At low doses of reactive chlorine, formation of Br-DBPs is induced to the extent possible given the Br⁻ presence and the excess and characteristics of DOC, whereas at high chlorine concentrations, chemical equilibria favour formation of Cl-DBPs. Consequently, the dosage of chlorine will highly influence which types of DBPs that are formed. To avoid formation of Br-DBPs, the balance between raw water Br⁻ levels, Br⁻ removal through treatment and DOM levels and characteristics need to be considered. In conclusion, apart from removing DOM from water, it is also important to consider how the DOM removal influence which types of DBPs that are formed in order to reduce health effects associated with DBPs. Further studies combining quantitative and detailed qualitative aspects of DBP formation appear needed to reach the optimal solution for each water treatment.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

This project was funded by the Swedish Research Council for Sustainable Development, FORMAS (grant no. 2013-1077), Svenskt Vatten Utveckling (project no. 16-104), Norrvatten, and Stockholm Vatten och Avfall. This is contribution of the University of Maryland Center for Environmental Science No, 5737 (Ref. No. [UMCES] CBL 2020-025). We thank Kristina Dahlberg for input on experimental design and practical assistance during sample collection and Ulf Eriksson for helpful input on process related issues. We also thank the staff at Lovö WTP for very accommodating support during the days of sample collections, experiments and extractions, and Susanne Karlsson, Nguyen Than Duc, Mårten Dario, and Ingrid Sundgren for valuable help and advice. Finally, gratitude goes to Elias Andersson who offered major practical assistance.

References

- 1 S. D. Richardson, M. J. Plewa, E. D. Wagner, R. Schoeny and D. M. DeMarini, Occurrence, genotoxicity, and carcinogenicity of regulated and emerging disinfection by-



- products in drinking water: a review and roadmap for research, *Mutat. Res.*, 2007, **636**(1), 178–242.
- 2 C. M. Villanueva, S. Cordier, L. Font-Ribera, L. A. Salas and P. Levallois, Overview of Disinfection By-products and Associated Health Effects, *Curr. Environ. Health Rep.*, 2015, **2**(1), 107–115.
 - 3 F. Bove, Y. Shim and P. Zeitz, Drinking water contaminants and adverse pregnancy outcomes: A review, *Environ. Health Perspect.*, 2002, **110**(SUPPL. 1), 61–74.
 - 4 L. Liang and P. C. Singer, Factors influencing the formation and relative distribution of haloacetic acids and trihalomethanes in drinking water, *Environ. Sci. Technol.*, 2003, **37**(13), 2920–2928.
 - 5 C. Postigo, S. D. Richardson and D. Barceló, Formation of iodo-trihalomethanes, iodo-haloacetic acids, and haloacetaldehydes during chlorination and chloramination of iodine containing waters in laboratory controlled reactions, *J. Environ. Sci.*, 2017, **58**, 127–134.
 - 6 M. B. Heeb, J. Criquet, S. G. Zimmermann-Steffens and U. Von Gunten, Oxidative treatment of bromide-containing waters: Formation of bromine and its reactions with inorganic and organic compounds - A critical review, *Water Res.*, 2014, **48**(1), 15–42.
 - 7 J. Tan, S. Allard, Y. Gruchlik, S. McDonald, C. A. Joll and A. Heitz, Impact of bromide on halogen incorporation into organic moieties in chlorinated drinking water treatment and distribution systems, *Sci. Total Environ.*, 2016, **541**, 1572–1580.
 - 8 S. J. Köhler, E. Lavonen, A. Keucken, P. Schmitt-Kopplin, T. Spanjer and K. Persson, Upgrading coagulation with hollow-fibre nanofiltration for improved organic matter removal during surface water treatment, *Water Res.*, 2016, **89**, 232–240.
 - 9 C. D. Evans, D. T. Monteith and D. M. Cooper, Long-term increases in surface water dissolved organic carbon: Observations, possible causes and environmental impacts, *Environ. Pollut.*, 2005, **137**(1), 55–71.
 - 10 G. Galjaard, E. Koreman, D. Metcalfe, G. Moore and P. Ericsson, Nom-removal by the six®-process, *Water Pract. Tech.*, 2018, **13**(3), 524–541.
 - 11 C. Rockey, D. C. Metcalfe, G. Galjaard, H. Shorney-Darby and J. Zheng, Ceramic microfiltration; a novel and compact process for the treatment of surface water, *Water Sci. Technol.: Water Supply*, 2018, **18**(6), 2035–2043.
 - 12 M. Gonsior, P. Schmitt-Kopplin, H. Stavklint, S. D. Richardson, N. Hertkorn and D. Bastviken, Changes in Dissolved Organic Matter during the Treatment Processes of a Drinking Water Plant in Sweden and Formation of Previously Unknown Disinfection Byproducts, *Environ. Sci. Technol.*, 2014, **48**(21), 12714–12722.
 - 13 B. D. Harris, T. A. Brown, J. L. McGehee, D. Houserova, B. A. Jackson and B. C. Buchel, *et al.* Characterization of Disinfection By-Products from Chromatographically Isolated NOM through High-Resolution Mass Spectrometry, *Environ. Sci. Technol.*, 2015, **49**(24), 14239–14248.
 - 14 E. E. Lavonen, M. Gonsior, L. J. Tranvik, P. Schmitt-Kopplin and S. J. Köhler, Selective chlorination of natural organic matter: Identification of previously unknown disinfection byproducts, *Environ. Sci. Technol.*, 2013, **47**(5), 2264–2271.
 - 15 H. Zhang, Y. Zhang, Q. Shi, J. Hu, M. Chu and J. Yu, *et al.* Study on transformation of natural organic matter in source water during chlorination and its chlorinated products using ultrahigh resolution mass spectrometry, *Environ. Sci. Technol.*, 2012, **46**(8), 4396–4402.
 - 16 E. E. Lavonen, D. N. Kothawala, L. J. Tranvik, M. Gonsior, P. Schmitt-Kopplin and S. J. Köhler, Tracking changes in the optical properties and molecular composition of dissolved organic matter during drinking water production, *Water Res.*, 2015, **85**, 286–294.
 - 17 H. Zhang, Y. Zhang, Q. Shi, S. Ren, J. Yu and F. Ji, *et al.* Characterization of low molecular weight dissolved natural organic matter along the treatment trait of a waterworks using Fourier transform ion cyclotron resonance mass spectrometry, *Water Res.*, 2012, **46**(16), 5197–5204.
 - 18 N. Cortés-Francisco, M. Harir, M. Lucio, G. Ribera, X. Martínez-Lladó and M. Rovira, *et al.* High-field FT-ICR mass spectrometry and NMR spectroscopy to characterize DOM removal through a nanofiltration pilot plant, *Water Res.*, 2014, **67**, 154–165.
 - 19 N. Hertkorn, M. Frommberger, M. Witt, B. P. Koch, P. Schmitt-Kopplin and E. M. Perdue, Natural organic matter and the event horizon of mass spectrometry, *Anal. Chem.*, 2008, **80**(23), 8908–8919.
 - 20 C. W. K. Chow, R. Fabris and M. Drikas, A rapid fractionation technique to characterise natural organic matter for the optimisation of water treatment processes, *J. Water Supply: Res. Technol.-AQUA*, 2004, **53**(2), 85–92.
 - 21 S. A. Huber, A. Balz, M. Abert and W. Pronk, Characterisation of aquatic humic and non-humic matter with size-exclusion chromatography - organic carbon detection - organic nitrogen detection (LC-OCD-OND), *Water Res.*, 2011, **45**(2), 879–885.
 - 22 E. Lavonen, I. Bodlund, K. Dahlberg, U. Eriksson, A. Andersson and S. Bertilsson, *et al.*, *Dricksvattenberedning med nya reningstekniker - en pilotstudie*, 2018, Contract No.: 2018–07.
 - 23 B. Chen and P. Westerhoff, Predicting disinfection by-product formation potential in water, *Water Res.*, 2010, **44**(13), 3755–3762.
 - 24 S. J. Köhler, D. Kothawala, M. N. Futter, O. Liungman and L. Tranvik, In-Lake Processes Offset Increased Terrestrial Inputs of Dissolved Organic Carbon and Color to Lakes, *PLoS One*, 2013, **8**(8), e70598.
 - 25 J. L. Weishaar, G. R. Aiken, B. A. Bergamaschi, M. S. Fram, R. Fujii and K. Mopper, Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of dissolved organic carbon, *Environ. Sci. Technol.*, 2003, **37**(20), 4702–4708.
 - 26 K. Mopper, A. Stubbins, J. D. Ritchie, H. M. Bialk and P. G. Hatcher, Advanced instrumental approaches for characterization of marine dissolved organic matter: Extraction techniques, mass spectrometry, and nuclear magnetic resonance spectroscopy, *Chem. Rev.*, 2007, **107**(2), 419–442.



- 27 A. Andersson, M. Harir, M. Gonsior, N. Hertkorn, P. Schmitt-Kopplin and H. Kylin, *et al.* Waterworks-specific composition of drinking water disinfection by-products, *Environ. Sci.: Water Res. Technol.*, 2019, **5**(5), 861–872.
- 28 B. P. Koch and T. Dittmar, From mass to structure: An aromaticity index for high-resolution mass data of natural organic matter, *Rapid Commun. Mass Spectrom.*, 2006, **20**(5), 926–932.
- 29 D. Metcalfe, C. Rockey, B. Jefferson, S. Judd and P. Jarvis, Removal of disinfection by-product precursors by coagulation and an innovative suspended ion exchange process, *Water Res.*, 2015, **87**, 20–28.
- 30 G. Asplund and A. Grimvall, Organohalogens in nature, *Environ. Sci. Technol.*, 1991, **25**(8), 1346–1350.
- 31 T. Bond, E. H. Goslan, S. A. Parsons and B. Jefferson, Treatment of disinfection by-product precursors, *Environ. Technol.*, 2011, **32**(1), 1–25.
- 32 U. Von Gunten, Ozonation of drinking water: Part I. Oxidation kinetics and product formation, *Water Res.*, 2003, **37**(7), 1443–1467.
- 33 X. Zhong, C. Cui and S. Yu, Exploring the pathways of aromatic carboxylic acids in ozone solutions, *RSC Adv.*, 2017, **7**(55), 34339–34347.
- 34 J. E. Kilduff, T. Karanfil, Y. P. Chin and W. J. Weber Jr., Adsorption of natural organic polyelectrolytes by activated carbon: A size-exclusion chromatography study, *Environ. Sci. Technol.*, 1996, **30**(4), 1336–1343.
- 35 D. Wang, Y. Zhao, J. Xie, C. W. K. Chow and J. Van Leeuwen, Characterizing DOM and removal by enhanced coagulation: A survey with typical Chinese source waters, *Sep. Purif. Technol.*, 2013, **110**, 188–195.
- 36 J. Song, X. Jin, X. C. Wang and P. Jin, Preferential binding properties of carboxyl and hydroxyl groups with aluminium salts for humic acid removal, *Chemosphere*, 2019, **234**, 478–487.
- 37 J. P. Croué, D. Violleau, C. Bodaire and B. Legube, Removal of hydrophobic and hydrophilic constituents by anion exchange resin, *Water Sci. Technol.*, 1999, 207–214.
- 38 H. Humbert, H. Gallard, V. Jacquemet and J. P. Croué, Combination of coagulation and ion exchange for the reduction of UF fouling properties of a high DOC content surface water, *Water Res.*, 2007, **41**(17), 3803–3811.
- 39 D. L. Separations, *DOWEX Ion Exchange Resins - Fundamentals of Ion Exchange*, 2000.
- 40 J. K. Edzwald, *Water quality & treatment. a handbook on drinking water*, McGraw-Hill, 6th edn, 2012.
- 41 Livsmedelsverket. Livsmedelsverkets föreskrifter om dricksvatten, SLVFS, 2001, 30 2015 [Available from: <http://www.livsmedelsverket.se/globalassets/om-oss/lagstiftning/dricksvatten—naturl-mineralv—kallv/slvfs-2001-30-kons-2015-3-webb.pdf>].
- 42 X. Zhang, S. Echigo, R. A. Minear and M. J. Plewa, Characterization and comparison of disinfection by-products of four major disinfectants, *ACS Symp. Ser.*, 2000, 299–314.
- 43 C. M. M. Bougeard, E. H. Goslan, B. Jefferson and S. A. Parsons, Comparison of the disinfection by-product formation potential of treated waters exposed to chlorine and monochloramine, *Water Res.*, 2010, **44**(3), 729–740.
- 44 X. Zhu and X. Zhang, Modeling the formation of TOCl, TOBr and TOI during chlor(am)ination of drinking water, *Water Res.*, 2016, **96**, 166–176.
- 45 G. Hua and D. A. Reckhow, Effect of pre-ozonation on the formation and speciation of DBPs, *Water Res.*, 2013, **47**(13), 4322–4330.
- 46 G. Hua and D. A. Reckhow, Characterization of disinfection byproduct precursors based on hydrophobicity and molecular size, *Environ. Sci. Technol.*, 2007, **41**(9), 3309–3315.
- 47 J. P. Croué, D. Violleau and L. Labouyrie, Disinfection by-product formation potentials of hydrophobic and hydrophilic natural organic matter fractions: A comparison between a low- and a high-humic water, *ACS Symp. Ser.*, 2000, 139–153.
- 48 N. Ates, S. S. Kaplan, E. Sahinkaya, M. Kitis, F. B. Dilek and U. Yetis, Occurrence of disinfection by-products in low DOC surface waters in Turkey, *J. Hazard. Mater.*, 2007, **142**(1–2), 526–534.
- 49 M. S. Ersan, C. Liu, G. Amy and T. Karanfil, The interplay between natural organic matter and bromide on bromine substitution, *Sci. Total Environ.*, 2019, **646**, 1172–1181.
- 50 T. Bond, E. H. Goslan, S. A. Parsons and B. Jefferson, Disinfection by-product formation of natural organic matter surrogates and treatment by coagulation, MIEX® and nanofiltration, *Water Res.*, 2010, **44**(5), 1645–1653.
- 51 J. Lundqvist, A. Andersson, A. Johannisson, E. Lavonen, G. Mandava and H. Kylin, *et al.* Innovative drinking water treatment techniques reduce the disinfection-induced oxidative stress and genotoxic activity, *Water Res.*, 2019, **155**, 182–192.
- 52 B. I. Escher, M. Dutt, E. Maylin, J. Y. M. Tang, S. Toze and C. R. Wolf, *et al.* Water quality assessment using the AREc32 reporter gene assay indicative of the oxidative stress response pathway, *J. Environ. Monit.*, 2012, **14**(11), 2877–2885.

