



Cite this: *Chem. Commun.*, 2016, 52, 6639

Received 16th February 2016,
Accepted 15th March 2016

DOI: 10.1039/c6cc01423b

www.rsc.org/chemcomm

Remote control over folding by light

Zhilin Yu† and Stefan Hecht*

Integrating stimulus-responsive components into macromolecular architectures is a versatile strategy to create smart materials that can be controlled by external stimuli and even adapt to their environment. Helical foldamers, which are omnipresent in Nature and display well-defined yet dynamic structures, serve as an ideal platform to integrate photoswitches to modulate their conformations by light. This feature article summarizes the development of photoswitchable foldamers, focussing on various design approaches that incorporate the photoswitches either at the side chains, as tethered loops, or directly in the main chain. Based on the emerging insight into the folding–switching relationship more advanced molecular designs should enable the development of photoresponsive foldamers with high sensitivity to control and power functional macromolecular and supramolecular systems.

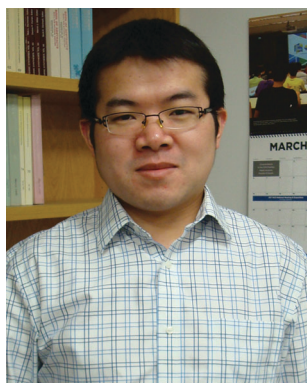
1. Introduction

Photoresponsive molecular systems, allowing for an external control of their structures and resulting properties by light, will be one of the key elements for creating “smart” materials with broad applications ranging from nanotechnology to pharmacology.^{1–3} Over the past two decades, photochromic moieties^{4–6} have been

incorporated into numerous materials that serve as molecular machines,^{7–14} remote-controlled catalysts,^{15–18} memory and storage devices,^{4,19,20} and optomechanical actuators,^{21,22} among others.^{23–29} However, the creation of real “smart” materials from these photoswitches requires maximization of the apparent functional responses to light at the macroscopic level. For this purpose – and alternatively to improve the intrinsic performance of the individual photoinduced events^{30–32} – incorporation of photoswitches into supramolecular³³ and macromolecular³⁴ systems allows for the potential amplification of the light signal by exploiting cooperative mechanisms.^{35–38} Considering the latter, covalent approach, dynamic helical macromolecules offer

Department of Chemistry and IRIS Adlershof, Humboldt-Universität zu Berlin, Brook-Taylor-Str. 2, 12489 Berlin, Germany. E-mail: sh@chemie.hu-berlin.de; Fax: +49-30-2093-6940; Tel: +49-30-2093-7365

† Present address: Department of Chemistry, Northwestern University, 2145 Sheridan Road, Evanston, IL 60208-3108, USA.



Zhilin Yu

Zhilin Yu received his BS degree from Tianjin University in 2006. Then he worked with Prof. Yu Liu at Nankai University on the complex of functional cyclodextrins and carbon materials, and obtained his MS degree in 2009. Following that, he continued his graduate study under the supervision of Prof. Stefan Hecht at the Humboldt-Universität zu Berlin, where his research involved the investigation of the structure–feature relationship of photo-

switchable foldamers composed of azobenzene units. He was awarded a PhD degree in 2013, and currently he is conducting his postdoctoral training with Prof. Samuel I. Stupp at Northwestern University.



Stefan Hecht

Stefan Hecht studied chemistry at Humboldt-Universität zu Berlin and obtained his PhD from the University of California at Berkeley in 2001, working under the guidance of Prof. Jean M. J. Fréchet. After establishing his own research group at Freie Universität Berlin and a subsequent position as a group leader at the Max-Planck-Institut für Kohlenforschung in Mülheim an der Ruhr, in 2006 he returned to his alma mater as the Chair of

Organic Chemistry and Functional Materials. His research interests range from synthetic macromolecular and supramolecular chemistry to surface science, with particular focus on utilizing photochromic molecules for remote controlling materials, devices, and processes. More information at: www.hechtlab.de.



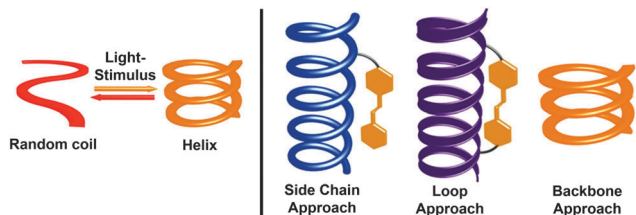


Fig. 1 Schematic illustration of a foldamer's (cooperative) helix-coil transition triggered by light (left). Conceptually different approaches to incorporate photoswitches, illustrated here for the most commonly used azobenzene, into the helical structures as side chains, tethered loops, or directly into the backbone (right).

the unique potential of cooperative folding³⁹ and, inspired by the importance of helices in nature, the past decade has witnessed the development of a variety of photoresponsive helical structures,^{38,40–46} which can undergo a light-induced (cooperative) folding and/or unfolding transition (Fig. 1, left).

In general, helical structures in foldamers are stabilized by a set of covalent local constraints and non-covalent intramolecular interactions.^{39,47–50} To successfully render foldamers photoswitchable one has to maximize the geometry change between the two different isomers of the photoswitch that should lead to selective stabilization or destabilization of the helical structure in one and/or the other switching form. Among the many established photoswitches,^{4–6} azobenzene offers the unique advantage that it features a remarkable change in molecular geometry (and dipole moment) based on the light-induced interconversion between its planar *E*-isomer and its non-planar *Z*-isomer.³¹ These dramatic structural changes occurring in the course of *E*-*Z* isomerization allow for facile variation of local constraints in the helical backbone but also influence π,π -stacking interactions and solvation, both of which contribute important enthalpic and entropic effects during helical folding. Alternatively, the light-induced conversion of charge-neutral spiropyrans to their zwitterionic merocyanine form⁵¹ has been used to locally enhance polarity and stabilize or destabilize helical structures.⁵² Thus far, three types of design approaches have been followed by integrating the photoswitches into the helical systems either (i) as side chains linked in a more or less flexible fashion to the main chain, or (ii) as tethered loops giving rise to constrained macrocyclic architectures, or (iii) directly into the helical backbone (Fig. 1, right).

In this feature article, we summarize the development of photoswitchable helical foldamers composed of various oligomeric/polymeric backbones and highlight their resulting light-induced (un)folding behavior in relation to the specific location of the photoswitches within the chain. Throughout we focus on the underlying design principle of the photoresponsive helices, following either side chain, loop, or backbone approaches, and the resulting relationship between local photoswitching and global foldamer conformation.

2. Side chain approach

In a significant body of work photoswitches have been attached to mostly polymeric foldamer backbones as their side chains.^{40,41,53,54}

Therefore, the local geometrical changes induced upon photo-switching do not affect the backbone conformation directly but rather alter the non-covalent interactions between the side chains, thereby indirectly leading to an overall conformational change. This design offers the advantage that minimizes the steric constraint for switching imposed by the backbone, thereby resulting in potentially lower energy barriers and hence more efficient photoswitching. However, the secondary effects of photoswitching on modulating side chain interactions are somewhat harder to predict, rendering a straightforward design more challenging. Note that in particular when using this design approach one should carefully distinguish between photomodulation of folding, *i.e.* influencing the helix-coil equilibrium by light, and photomodulation of (excess) helicity, *i.e.* influencing twist sense bias during folding by light. Experimentally, these two scenarios are sometimes not trivial to distinguish in particular when relying entirely on circular dichroism (CD) spectroscopy.⁵⁵

Early work on photoresponsive polymers reported light-induced structural changes of polyelectrolytes caused by non-covalently associated photoswitches.^{56,57} In these cases, the isomerization events modulate the interactions between photoswitches and polymers and thereby change the ionic repulsion along the hydrophobic polymer backbones, resulting in their extension or shrinkage. This strategy did inspire the development of covalent approaches to attach azobenzene and spiropyran units in the side chains of polymers.^{58,59} Due to the absence of defined (helical) conformations, researchers initially focused on monitoring viscosity changes of polymer solutions upon exposure to UV-light. The first example able to directly record light-induced conformational transition of photoresponsive polymers by CD spectroscopy reported the attachment of varying amounts of azobenzene units to the carboxyl side chains of poly(L-glutamic acid)s (Fig. 2).⁶⁰ In organic solvents, such as trifluoroethanol or trimethylphosphate, the modified polymeric backbones adopted an α -helical conformation as detected by CD spectroscopy.⁶¹ Exposure to 350 nm UV light led to *E* $\rightarrow$ *Z* photoisomerization of the azobenzenes yet the CD signal did not change indicating that the α -helix structure of the backbone was not disturbed. In contrast, when dissolving the polymer in water at pH = 7.6 assisted by the surfactant dodecylammonium chloride,⁶² the isomerization of azobenzene moieties gave rise to a reversible transition between random coil and α -helical conformations of the peptide backbone (Fig. 2). The observed solvent dependence of the photoswitchable folding behavior has been attributed to the change in the azobenzene's molecular dipole moment during photoisomerization, which is accompanied by a large change in the hydrophilicity of the side chain. Therefore, in the thermal resting state composed entirely of *E*-azobenzene side chains, their lipophilicity keeps the amphiphilic poly(L-glutamate) at the interface of the micelles where it adopts a random coil structure. Upon irradiation the metastable *Z*-isomer enriched peptide can adopt an α -helical conformation, which is able to expose the polar and hence more hydrophilic side chains to the aqueous environment dragging the peptide into the water phase. This behavior results in what we refer to as the photoinduced turn-on helix, *i.e.* light leads to helix formation while heat returns



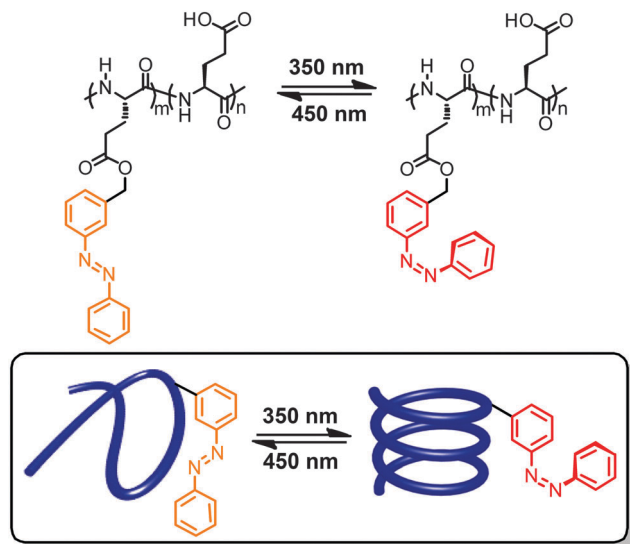


Fig. 2 Poly(L-glutamic acid) containing azobenzene units in the side chains undergoes a photoswitchable coil-helix transition in surfactant-water mixtures leading to turn-on helices.⁶²

the system to a random coil conformation due to the thermal instability of the *Z*-isomer.

At around the same time, Ueno and coworkers reported the modification of poly(benzyl L-aspartate) by varying amounts of phenylazo units that were attached to the benzyl side chains at either their *meta*- or *para*-positions, leading to different orientations of the resulting azobenzenes with regard to the peptide backbone.^{63–65} Interestingly, in this case and using specific organic solvent mixtures the authors were able to show by a thorough analysis of CD spectra that photoisomerization of the azobenzene side chains causes reversible inversion of the helix screw sense.⁶⁶ When studying poly(benzyl L-aspartate) containing 49% *meta*-linked azobenzene side chains in a mixture of helix-stabilizing 1,2-dichloroethane and helix-denaturing trimethyl phosphate, 86% of the polymer chains adopt a left-handed helix while after irradiation with UV-light 100% of the polymer chains adopt a right-handed helix (Fig. 3). Even when reducing the azobenzene side chain content to 9.7% a modulation between 91% right-handed helix in the all-*E*-isomer and 74% in the *Z*-enriched sample could be observed. Furthermore, the fact that in the latter case only 50% *E* → *Z* photoconversion is necessary to cause the effect illustrates that overall only *ca.* 5% isomerization of the side chains is sufficient to cause a helix inversion of the majority of the sample.

In addition to azobenzene units, poly(L-glutamic acid) has also been functionalized by spiropyrans in the side chains.^{66–68} In hexafluoroisopropanol, the merocyanine form is thermodynamically more stable (so-called negative photochromism) and the polypeptide backbone carrying the merocyanine residues adopts a random coil conformation (Fig. 4).⁶⁶ However, upon irradiation with visible light the colorless spiropyran is formed and the polypeptide backbone adopts a right-handed α -helical conformation. In this particular case and under the specific conditions, the zwitterionic merocyanine has a strong tendency

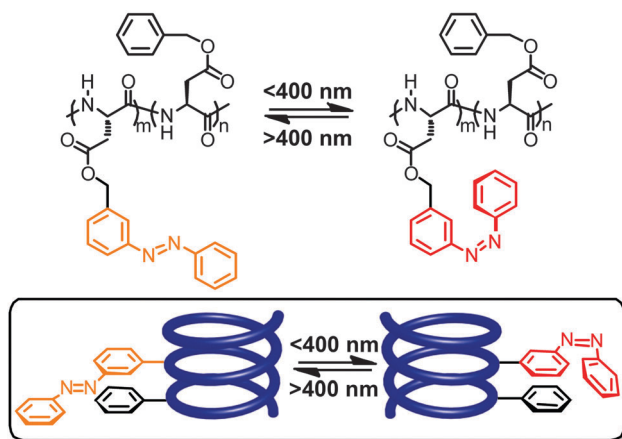


Fig. 3 Poly(L-aspartate) containing *meta*-linked azobenzene units undergoes a photoswitchable helicity inversion (change of helix screw sense) in the mixture of 1,2-dichloroethane and trimethyl phosphate.⁶⁴

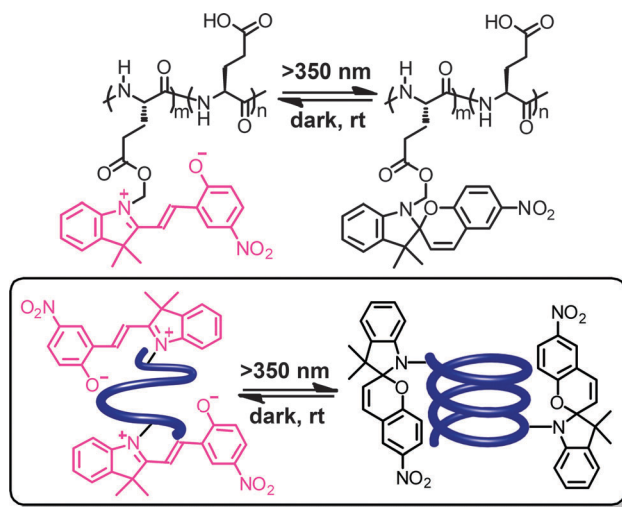


Fig. 4 Poly(L-glutamic acid) containing spiropyran units in their side chains undergoes a photoswitchable coil-helix transition in hexafluoroisopropanol leading to turn-on helices.⁶⁶

to dimerize⁶⁹ and therefore forces the polymer chain into a random coil structure. Visible light induces conversion to the spiropyran form and thereby weakens the side chain interactions such that the intrinsic helicogenicity of the backbone drives formation of the α -helix, giving rise to a turn-on helix once again.

Zentel and coworkers incorporated photoswitches in the side chains of abiotic foldamer backbones, such as polyisocyanates, to photomodulate their conformational behavior (Fig. 5).⁴⁰ Polyisocyanates composed of achiral monomers typically exist as a racemic mixture of right- and left-handed helices or isolated helical segments within the same polymer chain.^{70,71} The polyisocyanates decorated with chiral azobenzene side chains carrying one stereogenic center exhibited an intensity change in both ORD and CD spectra upon *E* → *Z* photoisomerization, indicating a change in the population of differently biased helical segments (center, Fig. 5).⁷² However, the *E* → *Z*



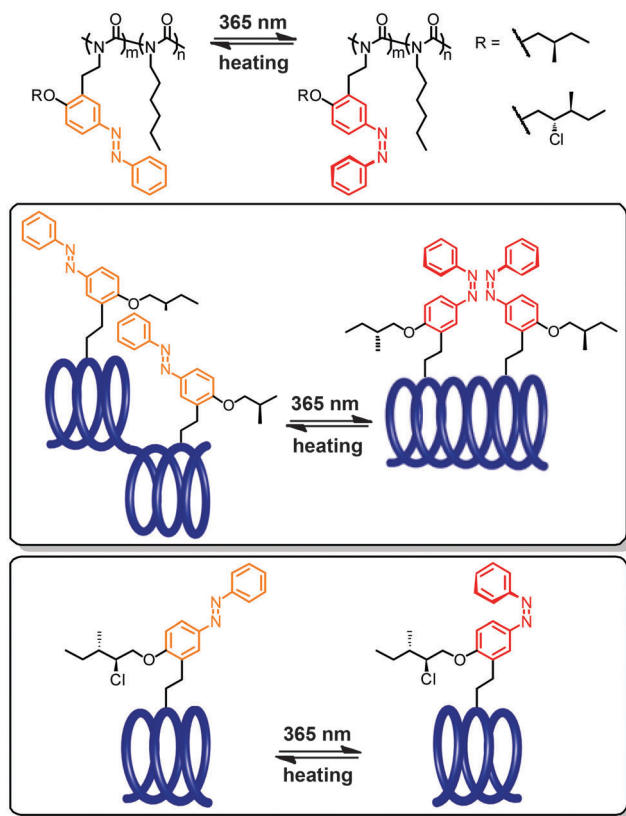


Fig. 5 Polyisocyanates containing chiral azobenzene units (top) exhibiting either a photoswitchable shift in their equilibrium between *P*- and *M*-helical segments (center)⁷² or helix inversion (bottom)⁷³ depending on the number of stereogenic centers in the azobenzene side chain.

photoisomerization of the azobenzene units with two chiral centers resulted in a more pronounced inversion of the Cotton effect (bottom, Fig. 5),⁷³ suggesting a helicity inversion of the polymers. The chiral amplification of the polyisocyanates was investigated involving a number of copolymers of various chiral azobenzene-containing isocyanates and over the course of their studies the authors observed a fine balance of various effects, including the distance of the stereocenters to the polymeric backbone, the solvent, and the concentration of photoswitches.^{74–76} By relating the *E*:*Z* ratio to the molar ellipticity in the CD spectra,⁷⁴ it could be shown that the *Z*-isomer exhibits a stronger induction power and rules the helix diastereoselection, in accordance with the majority rule.^{71,77}

Complementary to these aliphatic backbones, azobenzene moieties have also been attached to the side chains of aromatic foldamers, in particular poly(*meta*-phenylene ethynylene)s, prepared *via* Sonogashira–Hagihara polycondensation of 3,5-bis-(diethynyl)azobenzene and 1,3-diiodobenzene derived monomers (Fig. 6).⁷⁸ Intramolecular H-bonds between the amide and carbamate units allow this backbone to adopt stable helical conformations in a wide range of organic solvents.⁷⁹ Upon exposure to UV light, only subtle changes in the UV spectra were observed, indicating that just a small (7%) fraction of the azobenzene side chains underwent photoisomerization. This, presumably, is the reason why only limited degree of

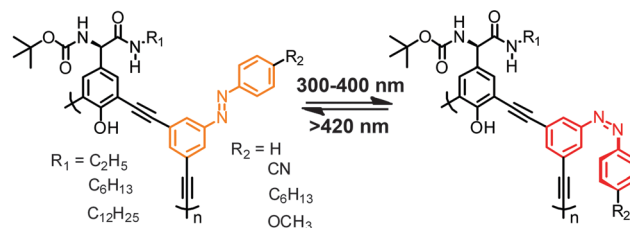


Fig. 6 Poly(*meta*-phenylene ethynylene) with appended phenylazo side chains folds into a helical structure in a variety of organic solvents due to H-bonding yet undergoes only little photoisomerization of the azobenzene moieties.⁷⁸

conformational response could be achieved as evident from the minimal changes in the corresponding CD spectra.

3. Loop approach

Rather than appending the photochromic units to the foldamer main chain on only one of their termini, tethering them at both of their ends to the backbone provides a macrocyclic loop architecture as a structural alternative to create photoswitchable foldamers.⁴⁶ Thus far peptides have been employed as backbones. Their versatile solid-phase synthesis allows for the positioning of amino acids carrying appropriate reactive groups at specific locations in the backbone, at which photoswitches with suitable geometry and size are subsequently tethered in a loop fashion. In principle, the geometry of one of the two isomers of the incorporated photoswitch – but not the other – is chosen to match the distance between the two reactive side chains in the targeted conformation of the peptide backbone. The photoinduced interconversion between these two isomers, associated with a significant local geometry change as in the case of the most commonly used azobenzenes, therefore reversibly makes and breaks the covalent local constraints favoring the specific backbone conformation, typically a peptide α -helix.

This loop approach has been pioneered by Woolley and coworkers, who tethered a variety of symmetrical azobenzene photoswitches *via* their *para*-positions to two cysteine residues within a peptide sequence (Fig. 7).⁸⁰ Molecular modelling suggested that upon attaching the azobenzene to cysteine residues in the *i* and *i* + 7 positions the peptide α -helix is only commensurate with the kinked *Z*-isomer, while the *E*-isomer should lead to unfolding and population of the random coil structure.⁸¹ Consequently, UV-induced *E* → *Z* isomerization promoted the formation of the α -helical conformation, while thermal or vis-light induced *Z* → *E* isomerization collapsed the helical conformation. Upon comparing the calculated distances between the two cysteine residues in various conformations, the authors could show that the α -helix content was improved from 2% in the *E*-isomer to 48% in the *Z*-isomer.⁸² On the other hand, tethering the peptide using cysteine residues in the more distant *i* and *i* + 11 positions renders the α -helix compatible with the extended *E*-azobenzene, which upon photoconversion to the shorter *Z*-azobenzene should consequently break the helical conformation. This nicely illustrates how either the



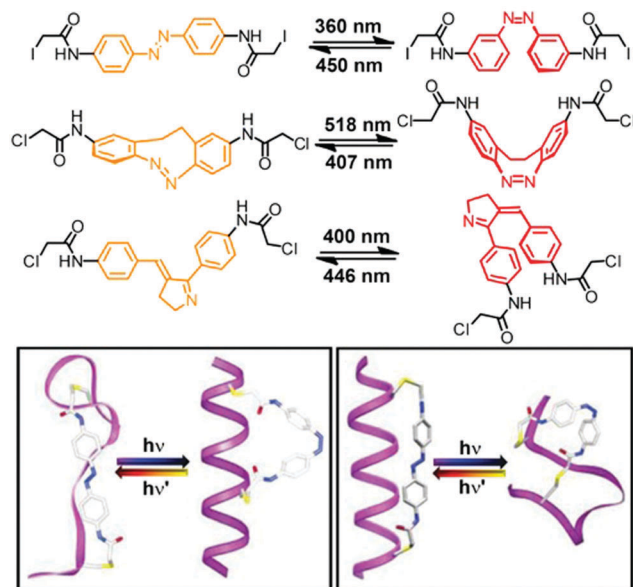


Fig. 7 Photoswitches that have been used as tethers to covalently link two side chain residues in peptide based foldamers in the loop approach (top). Turn-on (bottom left) and turn-off (bottom right) helices with the *E*- and *Z*-configured tethers connecting the *i* and *i* + 7 positions and the *i* and *i* + 11 positions of the peptide α -helix, respectively.⁸⁰

E-isomer or the *Z*-isomer of the azobenzene tether can be utilized to specifically stabilize the peptide's α -helix, resulting in a turn-off and turn-on foldamer, respectively. These two complementary approaches display some additional differences associated with the specific photoswitching behavior of azobenzene. In the turn-off design, the remaining *E*-isomer due to incomplete photoconversion leads to residual population of the α -helical conformation. In contrast, in the turn-on design helix formation can facilitate *E* $\rightarrow$ *Z* photoisomerization and thermally stabilize the *Z*-isomer while quantitative (photo)conversion of the *Z*-isomer back to the *E*-isomer leads to complete reversal of the process.

Rendering the loop approach more applicable, the group of Woolley developed and incorporated photoswitches, which do not require the use of UV-light and can be operated entirely with visible light.^{25,26} For example, they could trigger a reversible helix-coil transition by visible light and furthermore improve the extent of helical unfolding by using novel photochromic units (Fig. 7).^{83,84} On the one hand, a cyclic bis(*para*-acetamido)azobenzene derivative with separated $n \rightarrow \pi^*$ absorption bands for the two isomers⁸⁵ was tethered to the *i* and *i* + 11 positioned cysteine residues within the backbone.⁸³ As a result, the helix-coil transition could repetitively be switched by an alternating exposure to 407 nm and 518 nm light and thanks to the very efficient photoconversion between the *E*- and *Z*-isomers, the change in population of the different conformers was quite significant. On the other hand, to enhance the effect of the local photoswitching event on the conformational transition, *i.e.* improve the transduction efficiency, a non-symmetrical benzylidene-pyrroline chromophore, mimicking the Schiff base in rhodopsin and giving rise to a larger geometrical change upon

photoisomerization, has been linked to the peptide backbone.⁸⁴ Tethering the photoswitch between the *i* and *i* + 11 residues allowed the peptide backbone to adopt an α -helical conformation in the dark, while a random coil conformation was efficiently populated following *E* $\rightarrow$ *Z* photoisomerization of the benzylidene-pyrroline moiety. However, despite the high degree of photoconversion associated with a large structural change in the photochromic tether, the typical α -helix signature in the CD spectrum did not decrease to a large extent, which is probably due to the location of the photo-switch relative to the helix, leaving a larger part of the peptide and hence its conformation unaffected.

4. Backbone approach

The photoswitchable units can also be incorporated into the main chain^{86,87} of the helical structures to manipulate their folding behavior. This backbone approach is perhaps the most direct way to couple photoswitches with foldamers and both the photoswitching as well as the folding behavior are inextricably linked. Since most photoswitchable moieties consist of a more or less extended π -conjugated chromophore, their integration as monomer units is most readily accomplished using (hetero)aromatic foldamers based on either local constraints^{50,88} and/or solvophobic interactions.⁸⁹ Therefore, the geometrical changes during the course of the photoswitching event affect both the local conformational constraints as well as the typically intrastrand π , π -stacking interactions with non-nearest neighbors and of course *vice versa*. Following this approach thus far mostly azobenzene has been used due to its large geometrical changes, reflected in dramatically different angles and distances, which have also been exploited in the loop approach (see above), and in the inability of the twisted three dimensional *Z*-isomer to engage in stabilizing π , π -stacking contacts in strong contrast to the flat *E*-isomer. These azobenzene-containing foldamers have been connected by three different types of bridging units, *i.e.* carboxamide and triazole linkages, both providing some local conformational preferences,⁸⁸ as well as ethynylene (and also diethynylene) linkages, yielding three different classes of rigid photoswitchable foldamers, including oligoamides,⁵⁰ clickamers,⁹⁰ and oligo(*meta*-phenylene ethynylenes) (*OmpEs*)⁹¹ as covered below.

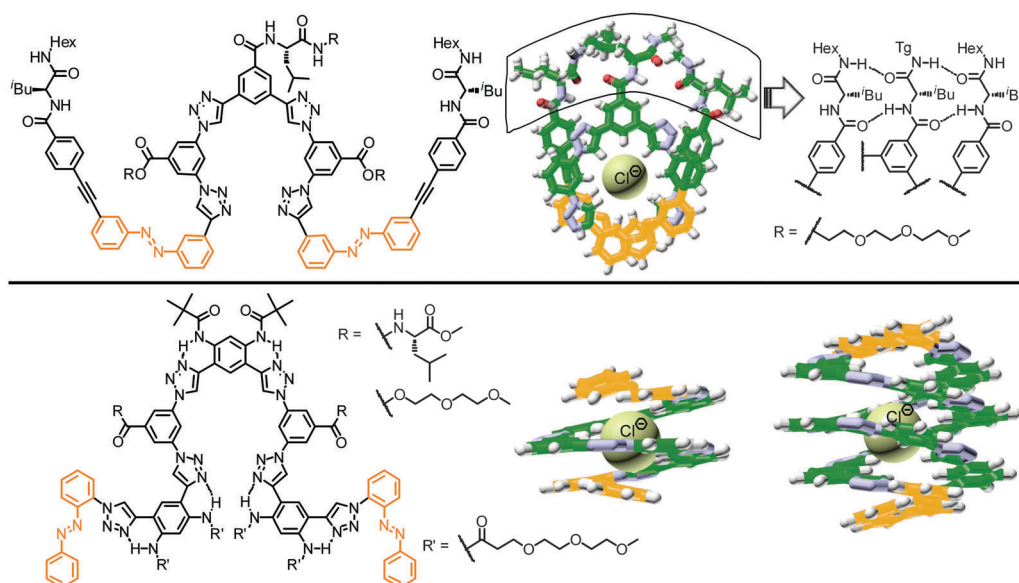
4.1 Photoresponsive oligoamides

The remarkable properties of the aromatic oligoamide foldamers,^{49,50} including their strong local conformational preferences, their high chemical stability, and their propensity to crystallize (and thereby enabling accurate structure determination by single crystal X-ray diffraction), inspire the design of their respective photoswitchable counterparts. Parquette and co-workers reported oligomeric backbones composed of alternating sequences of pyridine-2,6-dicarboxamides and *meta*-(phenylazo)azobenzene moieties (Fig. 8).⁴³ In its all-*E* configuration the foldamer adopts a helical structure with an approximately 3.4 Å helical pitch as predicted by molecular modelling and





To further advance their design, the same group prepared several new clickamers with stronger ion affinity utilizing interlocking and capping approaches.^{101,102} In the interlocking approach, the L-leucine residues forming β -sheet-type H-bonds were introduced to the central and terminal phenyl groups of the clickamer foldamer to initially stabilize the folded conformation as well as the foldamer-anion complexes by interlocking helical turns (Fig. 11, top).¹⁰¹ The formation of these interlocking H-bonds improved the chloride binding constant by almost one order of magnitude (from $1.26 \times 10^5 \text{ M}^{-1}$ to $9.7 \times 10^5 \text{ M}^{-1}$). Upon exposure to UV-light, the isomerization of the terminal azobenzene units significantly decreased the stability of the foldamer and hence led to unfolding, associated with a 84-fold reduced chloride affinity of the oligomer. In the alternative capping method, two clickamers composed of six triazole and five phenylene units were terminated with two *ortho*-linked azobenzene units (Fig. 11, bottom).¹⁰² These foldamers form single or double helical complexes with chloride ions in aqueous solution. In the latter double helical complex, a much larger percentage of the hydrophobic π -surface is buried ($\sim 80\%$ instead of $\sim 50\%$ in the single helix) and in addition the terminal *E*-configured azobenzene units shield the hydrogen-bond donors to chloride, thereby creating a solvent-excluding cavity capable of generating stronger $\text{CH}\cdots\text{Cl}^-$ H-bonds. While this work nicely illustrates the importance of creating specific



Chem. Commun., 2016, 52, 6639–6653 | 6645

This journal is © The Royal Society of Chemistry 2016

structure as revealed by standard solvent titration experiments.¹¹⁰ Unfortunately in this case, the azobenzene unit cannot be excited selectively due to the cross-conjugated *meta*-linkage and the photoisomerization of the helical foldamer was not possible due to the strong background absorption from the backbone.

To ensure selective excitation of the azobenzene core and hence access photoswitchability in the helically folded structure, the $\pi \rightarrow \pi^*$ absorption band of the azobenzene core was bathochromically shifted by introducing *para*-methoxy groups. As a consequence of the slightly weaker π, π -stacking contact of the more electron-rich 4,4'-dimethoxyazobenzene (DMAB) unit, the oligomer had to be elongated from a dodecamer to a tetradecamer (Fig. 13, bottom),⁴² which adopts a helical structure that is $1.7 \text{ kcal mol}^{-1}$ more stable than the coil structure, *i.e.* helix stabilization energy = $-1.7 \text{ kcal mol}^{-1}$, in acetonitrile.¹¹⁰ The formation of the helix was furthermore supported by the significant Cotton effect in CD spectra in aqueous acetonitrile solution due to the twist sense bias provided by the chiral side chains. As confirmed by both CD and UV/vis spectroscopy, exposure of the foldamer to UV-light induced formation of the *Z*-azobenzene core, which disrupts π, π -stacking interactions in the helix and consequently led to unfolding of the tetradecamer. This first prototype of a photoswitchable OmPE foldamer displayed approximately 40% unfolding in the PSS and sparked further efforts to design more efficient analogues.

Foldamers entirely composed of azobenzene. Maximizing the aspect ratio change of the photoswitchable foldamers induced by the $E \rightarrow Z$ photoisomerization of the incorporated azobenzene unit(s) requires a highly efficient conformational transition. As mentioned above our prototype of the azobenzene foldamers showed only a moderate light-induced change of the CD intensity, implying partial collapse of the foldamers. In order to increase the extent of the conformational change we sought to improve the efficiency of the photoinduced transition by enhancing the isomerization statistics. Hence, a series of foldamers entirely composed of azobenzene units with variable chain lengths, including oligomers **10**₅, **12**₆, and **14**₇ (Fig. 14), has been designed, synthesized, and characterized.⁴⁵ Solvent titration experiments revealed that in acetonitrile six azobenzene units are required for

the formation of a stable helix with two turns, assuming three azobenzene units per turn as suggested by molecular modelling and supported by ESR labelling studies.¹⁰⁶

For all three oligomers **10**₅, **12**₆, and **14**₇, the intensity of the Cotton effect in the CD spectra decreased dramatically or even quantitatively upon exposure to UV-light, indicating the occurrence of an efficient photoinduced unfolding transition following the $E \rightarrow Z$ photoisomerization (Fig. 14). The photoconversion was monitored by UV/vis spectroscopy and the content and distribution of *Z*-azobenzenes in the PSS was determined by NMR spectroscopy. Similarly to the findings of Parquette and coworkers,⁴³ the efficiency of the $E \rightarrow Z$ photoisomerization is dependent on the location of the azobenzene unit in the helix and the interior azobenzene units display a lower *Z*-content as compared to the terminal units. Importantly, the foldamer with a higher content of interior azobenzene units represents a lower rate constant of the photoinduced unfolding, illustrating that the untwisting process of these homogeneous backbones is predominately starting from the terminal positions. In contrast to Parquette's foldamers the helical refolding in this series of foldamers can readily be induced by visible light irradiation and resulting in full recovery of the helical structures over many cycles without any noticeable fatigue.

Foldamers with different relative orientations of azobenzenes.

It is well-known that the isomerization of photoswitches within natural or artificial systems strongly depends on their local surrounding.^{3,111,112} As illustrated for the above oligoazobenzene foldamer series, the particular microenvironment imparts a specific switching ability that presumably originates from different location dependent energy barriers due to specific intrastrand π, π -stacking interactions.^{43,45} To further optimize the photoisomerization and conformational transition of the oligoazobenzene foldamers, we investigated how specific locations of azobenzene units and their relative orientation as well as isomerization statistics affect the individual switching events and the global unfolding. As a result, the desire to vary the photochrome content while simultaneously positioning the individual azobenzene moieties precisely within the oligomeric backbone led us to design, synthesize, and characterize three different series of foldamers,

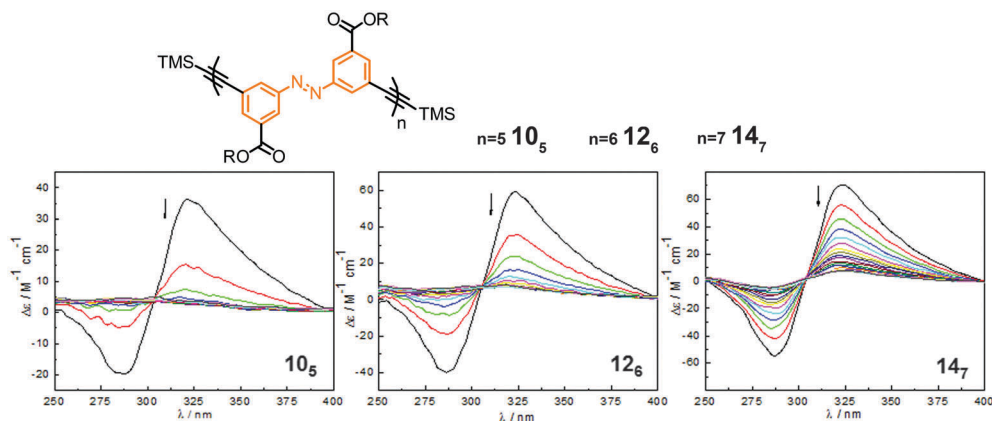


Fig. 14 Photoswitchable oligoazobenzene foldamers **10**₅, **12**₆, and **14**₇ (R = $-(\text{CH}_2\text{CH}_2\text{O})_3\text{CH}_3$ or $-(\text{CH}(\text{S-CH}_3)\text{CH}_2\text{O})(\text{CH}_2\text{CH}_2\text{O})_3\text{CH}_3$) and their corresponding CD spectra during the course of irradiation at $\lambda_{\text{irr}} = 358 \text{ nm}$ in CH_3CN at 25°C .⁴⁵

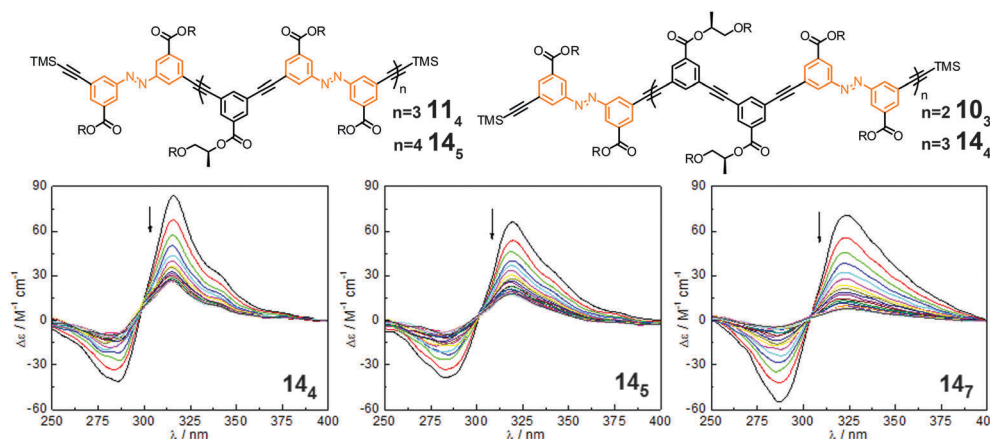


Fig. 15 Photoswitchable oligoazobenzene tetradecamers **14₄**, **14₅**, and **14₇** ($R = -(\text{CH}_2\text{CH}_2\text{O})_3\text{CH}_3$) and their corresponding CD spectra during the course of irradiation at $\lambda_{\text{irr}} = 358 \text{ nm}$ in CH_3CN at 25°C .¹⁰⁶

including oligomers **10₅**–**12₆**–**14₇** and **11₄**–**14₅** as well as **10₃**–**14₄** containing either 3 or 2 or 1.5 azobenzene units per turn (Fig. 15).¹⁰⁶

The variable relative orientations of azobenzenes within the helical structures led to three types of π, π -stacking interactions, *i.e.* azobenzene–azobenzene (azo–azo), phenylene–phenylene (Ph–Ph), and azobenzene–tolane (azo–tol). Calculated from the stabilization energies $\Delta G(\text{CH}_3\text{CN})$ of all the investigated foldamers determined by solvent titration experiments, the corresponding strengths of the three kinds of π, π -stacking interactions, *i.e.* azobenzene–azobenzene (azo–azo), phenylene–phenylene (Ph–Ph), and azobenzene–tolane (azo–tol), can be determined as $-0.86 \text{ kcal mol}^{-1}$, $-0.44 \text{ kcal mol}^{-1}$, and $-0.71 \text{ kcal mol}^{-1}$, respectively (Fig. 16). Based on the linear combination of these individual π, π -stacking interactions, the helix stabilization energy for various azobenzene-containing OmPE foldamers in acetonitrile can be estimated by:

$$\Delta G(\text{CH}_3\text{CN}, \text{kcal mol}^{-1}) = (-0.86)a + (-0.44)b + (-0.71)c$$

where a , b , and c stand for the number of the azo–azo, Ph–Ph, and azo–tol π, π -stacking interactions, respectively.

Considering the three tetradecamers **14₄**, **14₅**, and **14₇**, which only vary in the number and relative orientation of azobenzenes within the helix, we found that an increasing azobenzene content leads to both slower photoisomerization and unfolding, yet more complete denaturation according to the CD spectra upon UV-irradiation (Fig. 15). From this finding two competing effects become apparent: on the one hand, enhancing the number of azobenzene units is favorable as it increases the likelihood of light-induced formation of helix-breaking Z-isomers

(statistics); whereas on the other hand, avoiding strong π, π -stacking interactions, in particular between cofacial azobenzene units, enhances the efficiency of $E \rightarrow Z$ photoisomerization and subsequent unfolding (microenvironment). Therefore, both the chain length and azobenzene distribution throughout the backbone need to be optimized to maximize the photoresponse (efficiency) in these photoswitchable foldamers.

Foldamers with cooperative photoswitching events. Thus far we have been considering the effect of the local helix structure on the photoswitching ability of the embedded azobenzene units as a static effect. However, initial photoswitching events should influence the helical structure and change the environment, thereby changing the efficiency of a subsequent switching event. In such systems, multiple photoswitching events are coupled *via* the conformationally dynamic backbone, ideally in a cooperative fashion to further boost the sensitivity of the photoswitchable foldamers. In order to decipher the efficiency of individual photoswitching events within the foldamer backbone, *i.e.* to measure quantum yields for a specific azobenzene repeating unit, we have designed and synthesized, the three OmPE dodecamers **12_{2-int}**, **12_{2-core}**, and **12_{2-term}** with only two symmetrically placed azobenzene photochromes at either the core, the interior, or the termini of the helices, respectively (Fig. 17).¹⁰⁷ The limited content and symmetrical location of azobenzene units allow us to separate all three different possible isomers, *i.e.* E,E , E,Z , and Z,Z , and thereby quantify the photo-switching behavior. All three oligomers form stable helices in acetonitrile with helix stabilization energies associated with the location of azobenzene moieties, *i.e.* upon placing azobenzene units closer to the center, the helical structure becomes more stable.

Upon exposure to UV-light, the isomer content as well as the helicity were monitored and both were correlated for the three isomers (Fig. 17). Assuming that the photogenerated Z-isomer acts as a stopper for helix formation, which requires a critical chain length of approximately 10–12 phenylene units,¹¹³ the decays of the Cotton effect related to the loss of helicity in all the foldamers could quantitatively be correlated with the

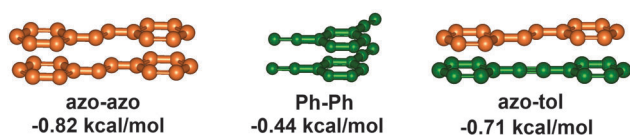


Fig. 16 Kinds of π, π -stacking interactions present in the folded backbones and their strengths.



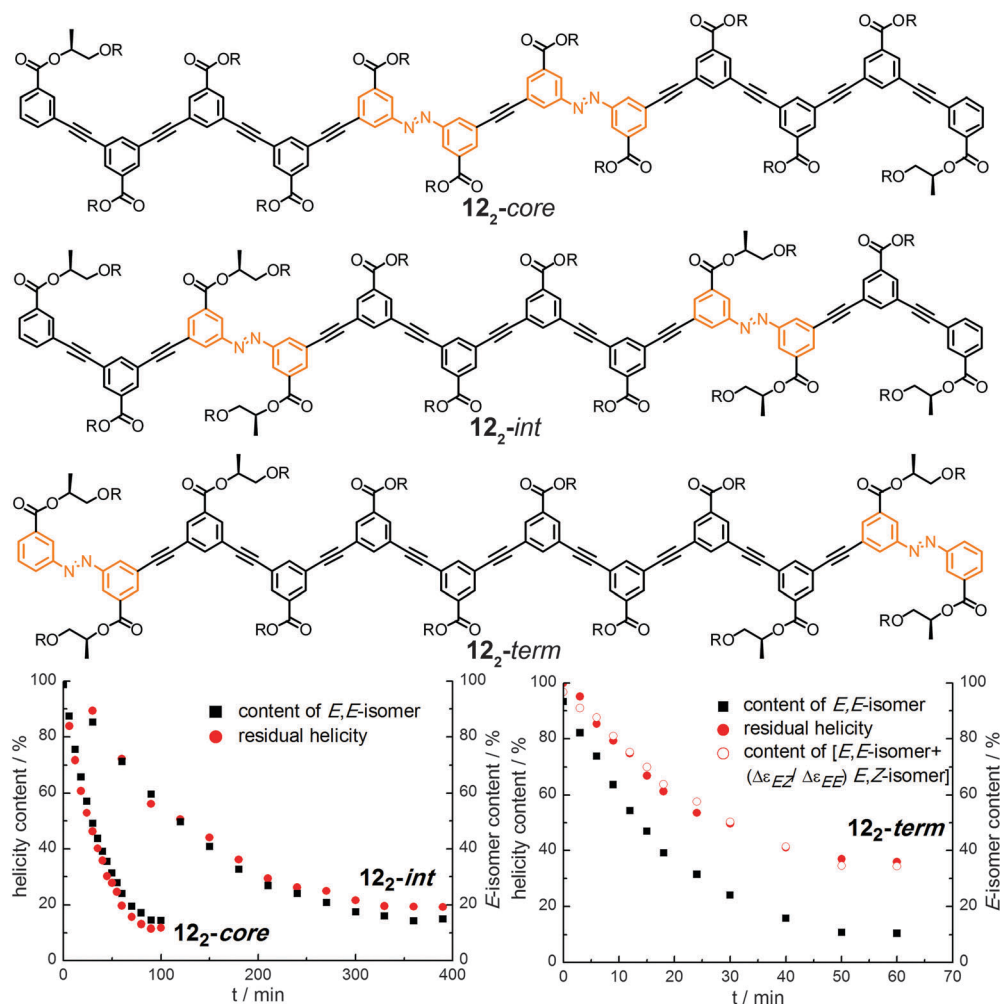


Fig. 17 Photoswitchable oligoazobenzene dodecamers **12₂-core**, **12₂-int**, and **12₂-term** ($R = -(CH_2CH_2O)_3CH_3$) and comparison of their CD intensities and isomer ratios during the course of irradiation at $\lambda_{irr} = 358$ nm in CH₃CN at 25 °C.¹⁰⁷

isomer contents, *i.e.* the *E,E*-isomer for foldamers **12₂-int** and **12₂-core** (Fig. 17, bottom left) or the *E,E*-isomer as well as the *E,Z*-isomer for foldamer **12₂-term** (Fig. 17, bottom right). The latter is explained by the fact that terminal isomerization yields a *E,Z*-configured foldamer, which can still adopt a (partially) helical structure giving rise to a Cotton effect. Importantly, this finding implies that there are no significant changes in the twist sense bias in these foldamers during photoswitching and furthermore no particular preference for untwisting either one of the *P*- or *M*-helices. Determination and analysis of the quantum yields for the individual *E* → *Z* photoisomerization events shows that photoswitching occurs more efficiently when the respective azobenzene moiety is located either at the helix terminus or in an already (partially) unfolded structure. This implies that once the initial photoswitching event in such structures has occurred, the subsequent photoisomerization event can be substantially facilitated and hence these bis-azobenzene systems exhibit (positive) cooperativity³⁶ with regard to its switching ability. Overall, the unfolding transition, which itself is cooperative, serves as a transduction mechanism to confer a cooperative behavior upon the embedded photoswitchable units

and thereby creates a photoresponsive macromolecular system with enhanced sensitivity.

Foldamers with a controllable unfolding pathway. The actual mechanism of unfolding is of importance for several processes, such as the binding of guests in molecular recognition involving helical hosts^{114–116} and the formation of double helices.^{117,118} Inspired by the observed location-dependent isomerization ability of the azobenzene units within the helix backbone, we sought to further enhance the localization of the isomerization events, thereby potentially controlling the unfolding pathway of the photoswitchable foldamers. For this purpose, a pair of tetradecamers **14_{3,2}** and **14_{4,1}** (Fig. 18) have been designed,¹⁰⁸ incorporating two different types of azobenzene units, *i.e.* the parent azobenzene and DMAB, which had been used as the core in our prototype.⁴² The excitation energy should be localized at the red-shifted DMAB units, either due to their direct (selective) excitation or indirect excitation followed by energy transfer from the parent azobenzenes. Therefore, the *E* → *Z* photoisomerization should be taking place primarily at the DMAB sites, which are localized in the terminal positions for foldamer **14_{3,2}** or in the core for foldamer **14_{4,1}**, and hence the unfolding

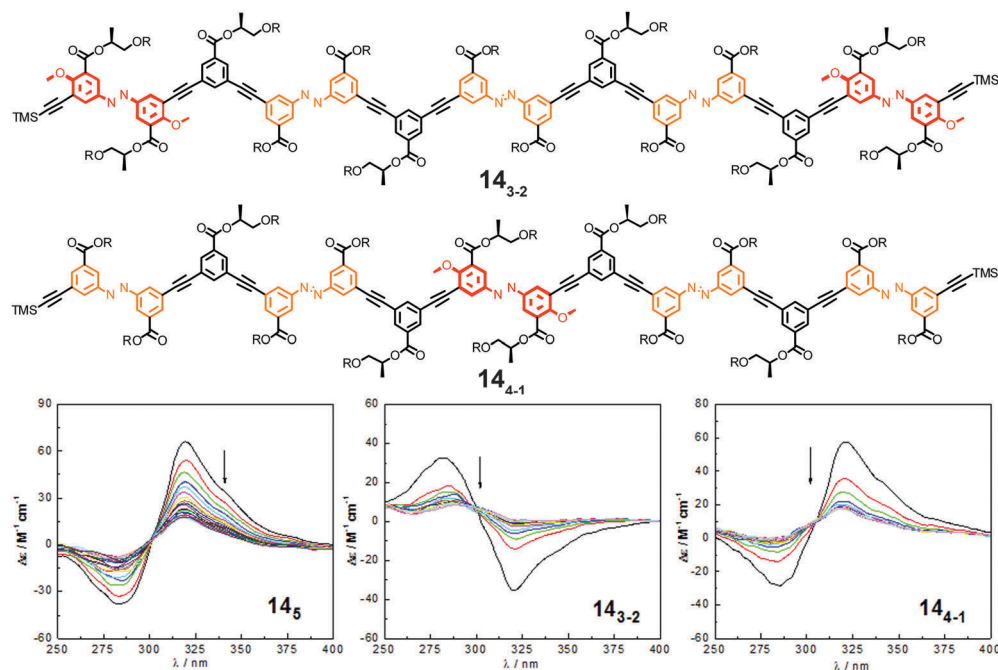


Fig. 18 Photoswitchable heterogeneous oligoazobenzene tetradecamers **14**₃₋₂ and **14**₄₋₁ ($R = -(\text{CH}_2\text{CH}_2\text{O})_3\text{CH}_3$) and the corresponding CD spectra (**14**₅ shown for comparison) during the course of irradiation at $\lambda_{\text{irr}} = 358 \text{ nm}$ in CH_3CN at 25°C .¹⁰⁸

pathway should be initiated either at the outside and progress inward in the case of foldamer **14**₃₋₂ or be initiated at the inside and progress outward in the case of foldamer **14**₄₋₁. The determined helix stability of the foldamers correlates well with the position of the DMAB units, which display weaker π,π -stacking interactions, within the backbone.

In comparison to the reference foldamer **14**₅ with a homogeneous backbone composed of parent azobenzene units only, the Z-content of photochromic units in the heterogeneous backbones in the PSS is increased specifically in the positions occupied by DMAB units, suggesting that indeed efficient intramolecular excitation energy transfer takes place in our foldamers exhibiting an energy gradient architecture.^{119–122} In contrast to the homogeneous oligomer **14**₅, in which untwisting the helical structure would predominantly arise from terminal isomerization events as described previously, the priority for isomerizing the DMAB units governs the unfolding process of foldamers **14**₃₋₂ and **14**₄₋₁. Specifically, initial terminal isomerization contributes mainly to the outside-in unfolding process of foldamer **14**₃₋₂, whereas initial interior isomerization dominates in the inside-out unfolding process of foldamer **14**₄₋₁ (Fig. 19).

Polymeric approach for various linkages. Despite the progress made in understanding photoswitchable foldamers and their (un)folding behavior, the synthesis and purification of discrete oligomers typically remain time-consuming with many transformations and low overall yields. This drawback clearly prohibits any reasonable scale-up and therefore precludes practical applications of such photoswitchable foldamers. As an alternative approach, polymerization can readily produce helically folding polymer chains at the cost of lower control over their length and sequence.^{78,122}

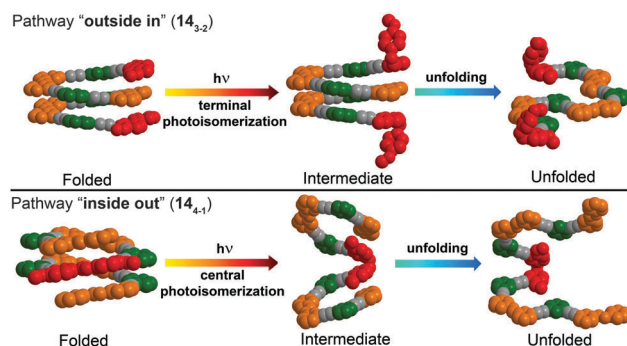


Fig. 19 Control over unfolding pathways in photoswitchable heterogeneous oligoazobenzene tetradecamers **14**₃₋₂ and **14**₄₋₁ exhibiting an energy gradient architecture leading to localization of the primary photoisomerization events at the DMAB positions (DMAB units are shown in red; parent azobenzene units in orange, meta-phenylene units in green, and ethynylene linkages in grey).¹⁰⁸

Along these lines, we designed and synthesized two types of poly(azobenzene)s using either ethynylene or butadiynylene linkages to connect azobenzene units in their meta-positions, leading to polymer **PAzoE** and **PAzoB**, respectively (Fig. 20).¹⁰⁹ While both polymers formed stable helices in a polar environment, their photoresponse was markedly different. While polymer **PAzoE** resembled the behavior of its oligomeric counterparts, incorporation of the extended diacetylene unit into polymer **PAzoB** considerably strengthened the π,π -stacking interactions among the helical structures, resulting in a pronounced aggregation tendency and suppressing photoisomerization in the folded state. This study demonstrates the importance of backbone connectivity to balance



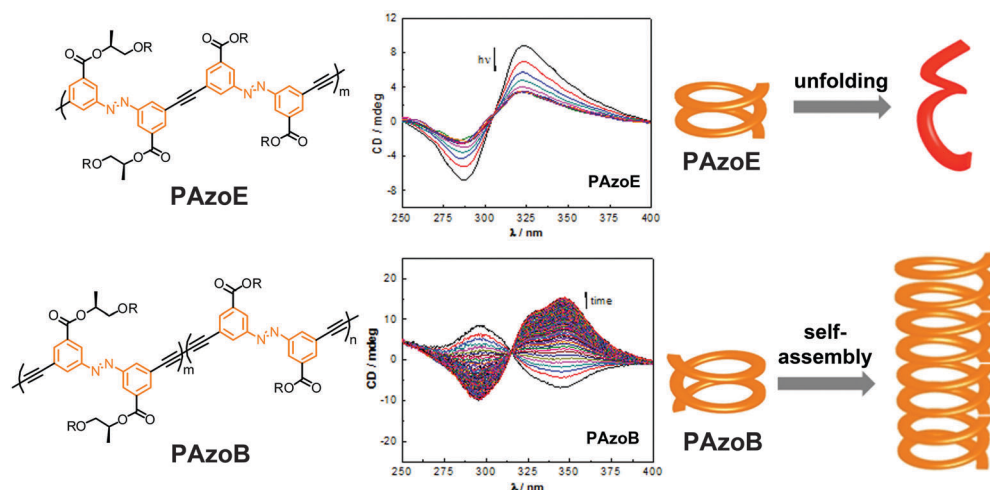


Fig. 20 Polyazobenzene foldamers containing ethynylene (PAzoE) and butadiynylene linkages (PAzoB) and CD spectra of PAzoE upon irradiation ($\lambda_{\text{irr}} = 358 \text{ nm}$) and of PAzoB over time in CH_3CN at 25°C , showing photoinduced unfolding and suppression of photoisomerization due to aggregation, respectively, as illustrated in the cartoon on the right.¹⁰⁹

intra- and intermolecular forces for the successful design of photoresponsive polymers.

5. Conclusion and perspectives

In this feature article, we detail the development of the photo-switchable foldamers with particular emphasis on the different design approaches for incorporating the photoswitches into the backbones. Depending on the manner of the attachment and the location of the incorporated photoswitches, the photo-induced interconversion between various (*meta*)stable isomers affects the local covalent constraints and/or the non-covalent interactions that govern the stabilization of the helix structure, thereby leading to conformational changes, in particular helix inversion and/or helix-coil transition. The coupling of the photoswitching event(s) and the (un)folding behavior is furthermore manifested in the profound effect of the folded architecture on the photoswitching ability and efficiency, leading to an overall “feedback”. Exploiting the strength of the oligomer approach to deduce clear structure-folding-switching relationships several design parameters, including the number, placement, relative orientation, and nature of photoswitches, can be optimized to maximize the conformational consequence of the photoswitching events and hence the sensitivity of photoresponse.

Considering the remarkable progress made over the past decade and the detailed understanding gained, we feel that it is time to exploit photoswitchable foldamers as a sophisticated tool to achieve remote control over the properties of various materials and device functions. As shown for photoswitchable clickamers, one of the promising applications of photoresponsive helices is their ability to act as dynamic hosts for the photo-controllable capture and release of guest molecules. Another main advantage is the significant change in the aspect ratio in these systems associated with their cooperative (un)folding that should aid the design of photoresponsive functional

macromolecular and supramolecular architectures, including sophisticated light-controlled catalysts^{15,17} based on photo-switchable foldamer scaffolds as well as molecular walkers¹⁰ for optomechanical systems^{12,3} based on an extension-shrinkage motion.³⁸ Beyond the molecular level, the collection and amplification of individual molecular photoswitching events still poses a major challenge.^{1,3,38} Therefore, it is important to organize the helical structures into unidirectional architectures spontaneously¹²⁴ or assisted by external force fields¹²⁵ and matrices.¹²⁶ In addition, non-covalent and covalent assembly of such functional helical architectures¹²⁷ will greatly enhance their availability and use. Combining all these aspects into new and more advanced molecular designs, we are absolutely confident that photoswitchable foldamers will prove invaluable to remote control and even power functional macromolecular and supramolecular systems operating at the mesoscopic and macroscopic level.

Acknowledgements

Our own contributions to the field have been generously supported by the German Research Foundation (DFG *via* SFB 765) and the European Research Council (*via* ERC-2012-STG_308117 “Light4Function”).

References

- 1 M.-M. Russew and S. Hecht, *Adv. Mater.*, 2010, **22**, 3348–3360.
- 2 W. A. Velema, W. Szymanski and B. L. Feringa, *J. Am. Chem. Soc.*, 2014, **136**, 2178–2191.
- 3 J. M. Abendroth, O. S. Bushuyev, P. S. Weiss and C. J. Barrett, *ACS Nano*, 2015, **9**, 7746–7768.
- 4 M. Irie, *Chem. Rev.*, 2000, **100**, 1683–1890.
- 5 H. B.-L. Heinz Dürr, *Photochromism: Molecules and Systems*, Elsevier, Amsterdam, 2003.
- 6 W. R. Browne and B. L. Feringa, in *Molecular Switches*, Wiley-VCH Verlag GmbH & Co. KGaA, 2011, pp. 121–179.
- 7 W. R. Browne and B. L. Feringa, *Nat. Nanotechnol.*, 2006, **1**, 25–35.



- 8 E. R. Kay, D. A. Leigh and F. Zerbetto, *Angew. Chem., Int. Ed.*, 2007, **46**, 72–191.
- 9 S. Saha and J. F. Stoddart, *Chem. Soc. Rev.*, 2007, **36**, 77–92.
- 10 M. von Delius and D. A. Leigh, *Chem. Soc. Rev.*, 2011, **40**, 3656–3676.
- 11 V. Serreli, C.-F. Lee, E. R. Kay and D. A. Leigh, *Nature*, 2007, **445**, 523–527.
- 12 M. Yamada, M. Kondo, J. Mamiya, Y. Yu, M. Kinoshita, C. J. Barrett and T. Ikeda, *Angew. Chem., Int. Ed.*, 2008, **47**, 4986–4988.
- 13 Q. Li, G. Fuks, E. Moulin, M. Maaloum, M. Rawiso, I. Kulic, J. T. Foy and N. Giuseppone, *Nat. Nanotechnol.*, 2015, **10**, 161–165.
- 14 G. Ragazzon, M. Baroncini, S. Silvi, M. Venturi and A. Credi, *Nat. Nanotechnol.*, 2015, **10**, 70–75.
- 15 R. S. Stoll and S. Hecht, *Angew. Chem., Int. Ed.*, 2010, **49**, 5054–5075.
- 16 J. Wang and B. L. Feringa, *Science*, 2011, **331**, 1429–1432.
- 17 R. Göstl, A. Senf and S. Hecht, *Chem. Soc. Rev.*, 2014, **43**, 1982–1996.
- 18 V. Blanco, D. A. Leigh and V. Marcos, *Chem. Soc. Rev.*, 2015, **44**, 5341–5370.
- 19 F. K. Bruder, R. Hagen, T. Rolle, M. S. Weiser and T. Facke, *Angew. Chem., Int. Ed.*, 2011, **50**, 4552–4573.
- 20 T. J. Kucharski, N. Ferralis, A. M. Kolpak, J. O. Zheng, D. G. Nocera and J. C. Grossman, *Nat. Chem.*, 2014, **6**, 441–447.
- 21 H. Yu and T. Ikeda, *Adv. Mater.*, 2011, **23**, 2149–2180.
- 22 N. Hosono, T. Kajitani, T. Fukushima, K. Ito, S. Sasaki, M. Takata and T. Aida, *Science*, 2010, **330**, 808–811.
- 23 S. Hecht, *Small*, 2005, **1**, 26–29.
- 24 G. Mayer and A. Heckel, *Angew. Chem., Int. Ed.*, 2006, **45**, 4900–4921.
- 25 C. Raimondo, N. Crivillers, F. Reinders, F. Sander, M. Mayor and P. Samori, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 12375–12380.
- 26 P. Theato, B. S. Sumerlin, R. K. O'Reilly and I. I. T. H. Epps, *Chem. Soc. Rev.*, 2013, **42**, 7055–7056 and entire special issue.
- 27 W. Szymański, J. M. Beierle, H. A. V. Kistemaker, W. A. Velema and B. L. Feringa, *Chem. Rev.*, 2013, **113**, 6114–6178.
- 28 J. Broichhagen, J. A. Frank and D. Trauner, *Acc. Chem. Res.*, 2015, **48**, 1947–1960.
- 29 P. K. Kundu, D. Samanta, R. Leizrowice, B. Margulis, H. Zhao, M. Börner, T. Udayabhaskararao, D. Manna and R. Klajn, *Nat. Chem.*, 2015, **7**, 646–652.
- 30 A. A. Beharry and G. A. Woolley, *Chem. Soc. Rev.*, 2011, **40**, 4422–4437.
- 31 H. M. D. Bandara and S. C. Burdette, *Chem. Soc. Rev.*, 2012, **41**, 1809–1825.
- 32 D. Bléger and S. Hecht, *Angew. Chem., Int. Ed.*, 2015, **54**, 11338–11349.
- 33 S. Yagai, T. Karatsu and A. Kitamura, *Chem. – Eur. J.*, 2005, **11**, 4054–4063.
- 34 M. Irie, *Adv. Polym. Sci.*, 1990, **94**, 27–67.
- 35 D. Zhao and J. S. Moore, *Org. Biomol. Chem.*, 2003, **1**, 3471–3491.
- 36 C. A. Hunter and H. L. Anderson, *Angew. Chem., Int. Ed.*, 2009, **48**, 7488–7499.
- 37 T. F. De Greef, M. M. Smulders, M. Wolffs, A. P. Schenning, R. P. Sijbesma and E. W. Meijer, *Chem. Rev.*, 2009, **109**, 5687–5754.
- 38 D. Blegler, Z. Yu and S. Hecht, *Chem. Commun.*, 2011, **47**, 12260–12266.
- 39 D. J. Hill, M. J. Mio, R. B. Prince, T. S. Hughes and J. S. Moore, *Chem. Rev.*, 2001, **101**, 3893–4012.
- 40 S. Mayer and R. Zentel, *Prog. Polym. Sci.*, 2001, **26**, 1973–2013.
- 41 A. Natansohn and P. Rochon, *Chem. Rev.*, 2002, **102**, 4139–4176.
- 42 A. Khan, C. Kaiser and S. Hecht, *Angew. Chem., Int. Ed.*, 2006, **45**, 1878–1881.
- 43 C. Tie, J. C. Gallucci and J. R. Parquette, *J. Am. Chem. Soc.*, 2006, **128**, 1162–1171.
- 44 Y. Hua and A. H. Flood, *J. Am. Chem. Soc.*, 2010, **132**, 12838–12840.
- 45 Z. Yu and S. Hecht, *Angew. Chem., Int. Ed.*, 2011, **50**, 1640–1643.
- 46 G. A. Woolley, *Acc. Chem. Res.*, 2005, **38**, 486–493.
- 47 S. Hecht and I. Huc, *Foldamers: Structure, Properties, and Applications*, Wiley-VCH, Weinheim, 2007.
- 48 E. Yashima, K. Maeda, H. Iida, Y. Furusho and K. Nagai, *Chem. Rev.*, 2009, **109**, 6102–6211.
- 49 G. Guichard and I. Huc, *Chem. Commun.*, 2011, **47**, 5933–5941.
- 50 D.-W. Zhang, X. Zhao, J.-L. Hou and Z.-T. Li, *Chem. Rev.*, 2012, **112**, 5271–5316.
- 51 R. Klajn, *Chem. Soc. Rev.*, 2014, **43**, 148–184.
- 52 A. Koçer, M. Walko, W. Meijberg and B. L. Feringa, *Science*, 2005, **309**, 755–758.
- 53 O. Pieroni, A. Fissi, N. Angelini and F. Lenci, *Acc. Chem. Res.*, 2001, **34**, 9–17.
- 54 B. L. Feringa, R. A. van Delden, N. Koumura and E. M. Geertsema, *Chem. Rev.*, 2000, **100**, 1789–1816.
- 55 J. E. G. David and A. Lightner, *Organic Conformational Analysis and Stereochemistry from Circular Dichroism Spectroscopy*, Wiley-VCH, Weinheim, 2000.
- 56 R. Lovrien, *Proc. Natl. Acad. Sci. U. S. A.*, 1967, **57**, 236–242.
- 57 G. v. d. Veen, R. Hogue and W. Prins, *Photochem. Photobiol.*, 1974, **19**, 197–204.
- 58 M. Irie, A. Menju and K. Hayashi, *Macromolecules*, 1979, **12**, 1176–1180.
- 59 L. Matějka and K. Dušek, *Makromol. Chem.*, 1981, **182**, 3223–3236.
- 60 O. Pieroni, J. L. Houben, A. Fissi, P. Costantino and F. Ciardelli, *J. Am. Chem. Soc.*, 1980, **102**, 5913–5915.
- 61 J. L. Houben, A. Fissi, D. Bacciola, N. Rosato, O. Pieroni and F. Ciardelli, *Int. J. Biol. Macromol.*, 1983, **5**, 94–100.
- 62 O. Pieroni, D. Fabbri, A. Fissi and F. Ciardelli, *Macromol. Rapid Commun.*, 1988, **9**, 637–640.
- 63 A. Ueno, J.-i. Anzai, T. Osa and Y. Kadoma, *Bull. Chem. Soc. Jpn.*, 1979, **52**, 549–554.
- 64 A. Ueno, K. Takahashi, J. Anzai and T. Osa, *J. Am. Chem. Soc.*, 1981, **103**, 6410–6415.
- 65 A. Ueno, K. Adachi, J. Nakamura and T. Osa, *J. Polym. Sci., Part A: Polym. Chem.*, 1990, **28**, 1161–1170.
- 66 F. Ciardelli, D. Fabbri, O. Pieroni and A. Fissi, *J. Am. Chem. Soc.*, 1989, **111**, 3470–3472.
- 67 T. M. Cooper, K. A. Obermeier, L. V. Natarajan and R. L. Crane, *Photochem. Photobiol.*, 1992, **55**, 1–7.
- 68 A. Fissi, O. Pieroni, F. Ciardelli, D. Fabbri, C. Ruggeri and K. Umezawa, *Biopolymers*, 1993, **33**, 1505–1517.
- 69 F. Würthner, T. E. Kaiser and C. R. Saha-Möller, *Angew. Chem., Int. Ed.*, 2011, **50**, 3376–3410.
- 70 S. Lifson, M. M. Green, C. Andreola and N. C. Peterson, *J. Am. Chem. Soc.*, 1989, **111**, 8850–8858.
- 71 M. M. Green, J.-W. Park, T. Sato, A. Teramoto, S. Lifson, R. L. B. Selinger and J. V. Selinger, *Angew. Chem., Int. Ed.*, 1999, **38**, 3138–3154.
- 72 M. Mueller and R. Zentel, *Macromolecules*, 1994, **27**, 4404–4406.
- 73 G. Maxein and R. Zentel, *Macromolecules*, 1995, **28**, 8438–8440.
- 74 M. Müller and R. Zentel, *Macromolecules*, 1996, **29**, 1609–1617.
- 75 S. Mayer, G. Maxein and R. Zentel, *Macromolecules*, 1998, **31**, 8522–8525.
- 76 S. Mayer and R. Zentel, *Macromol. Chem. Phys.*, 1998, **199**, 1675–1682.
- 77 M. M. Green, B. A. Garetz, B. Munoz, H. Chang, S. Hoke and R. G. Cooks, *J. Am. Chem. Soc.*, 1995, **117**, 4181–4182.
- 78 H. Sogawa, M. Shiotsuki, H. Matsuoka and F. Sanda, *Macromolecules*, 2011, **44**, 3338–3345.
- 79 M. Banno, T. Yamaguchi, K. Nagai, C. Kaiser, S. Hecht and E. Yashima, *J. Am. Chem. Soc.*, 2012, **134**, 8718–8728.
- 80 J. R. Kumita, O. S. Smart and G. A. Woolley, *Proc. Natl. Acad. Sci. U. S. A.*, 2000, **97**, 3803–3808.
- 81 D. C. Burns, D. G. Flint, J. R. Kumita, H. J. Feldman, L. Serrano, Z. Zhang, O. S. Smart and G. A. Woolley, *Biochemistry*, 2004, **43**, 15329–15338.
- 82 D. G. Flint, J. R. Kumita, O. S. Smart and G. A. Woolley, *Chem. Biol.*, 2002, **9**, 391–397.
- 83 S. Samanta, C. Qin, A. J. Lough and G. A. Woolley, *Angew. Chem., Int. Ed.*, 2012, **51**, 6452–6455.
- 84 M. Blanco-Lomas, S. Samanta, P. J. Campos, G. A. Woolley and D. Sampedro, *J. Am. Chem. Soc.*, 2012, **134**, 6960–6963.
- 85 R. Siewertsen, H. Neumann, B. Buchheim-Stehn, R. Herges, C. Näther, F. Renth and F. Temps, *J. Am. Chem. Soc.*, 2009, **131**, 15594–15595.
- 86 M. Irie and W. Schnabel, *Macromolecules*, 1981, **14**, 1246–1249.
- 87 M. Irie, Y. Hirano, S. Hashimoto and K. Hayashi, *Macromolecules*, 1981, **14**, 262–267.
- 88 I. Huc and L. Cuccia, *Foldamers*, Wiley-VCH Verlag GmbH & Co. KGaA, 2007, pp. 1–33.
- 89 Y. Zhao and J. S. Moore, *Foldamers*, Wiley-VCH Verlag GmbH & Co. KGaA, 2007, pp. 75–108.



- 90 R. M. Meudtner and S. Hecht, *Angew. Chem., Int. Ed.*, 2008, **47**, 4926–4930.
- 91 J. Moore and C. Ray, *Adv. Polym. Sci.*, 2005, **177**, 91–149.
- 92 E. D. King, P. Tao, T. T. Sanan, C. M. Hadad and J. R. Parquette, *Org. Lett.*, 2008, **10**, 1671–1674.
- 93 C. J. Gabriel and J. R. Parquette, *J. Am. Chem. Soc.*, 2006, **128**, 13708–13709.
- 94 D. M. Bassani, J.-M. Lehn, G. Baum and D. Fenske, *Angew. Chem., Int. Ed.*, 1997, **36**, 1845–1847.
- 95 D. Zornik, R. M. Meudtner, T. El Malah, C. M. Thiele and S. Hecht, *Chem. – Eur. J.*, 2011, **17**, 1473–1484.
- 96 H. Juwarker, J. M. Lenhardt, D. M. Pham and S. L. Craig, *Angew. Chem., Int. Ed.*, 2008, **47**, 3740–3743.
- 97 Y. Li and A. H. Flood, *J. Am. Chem. Soc.*, 2008, **130**, 12111–12122.
- 98 Y. Li and A. H. Flood, *Angew. Chem., Int. Ed.*, 2008, **47**, 2649–2652.
- 99 R. M. Meudtner, M. Ostermeier, R. Goddard, C. Limberg and S. Hecht, *Chem. – Eur. J.*, 2007, **13**, 9834–9840.
- 100 Y. Hua and A. H. Flood, *Chem. Soc. Rev.*, 2010, **39**, 1262–1271.
- 101 S. Lee, Y. Hua and A. H. Flood, *J. Org. Chem.*, 2014, **79**, 8383–8396.
- 102 Y. Hua, Y. Liu, C.-H. Chen and A. H. Flood, *J. Am. Chem. Soc.*, 2013, **135**, 14401–14412.
- 103 Y. Wang, F. Bie and H. Jiang, *Org. Lett.*, 2010, **12**, 3630–3633.
- 104 S. Hecht and A. Khan, *Angew. Chem., Int. Ed.*, 2003, **115**, 6203–6206.
- 105 A. Khan and S. Hecht, *Chem. – Eur. J.*, 2006, **12**, 4764–4774.
- 106 Z. Yu, S. Weidner, T. Risse and S. Hecht, *Chem. Sci.*, 2013, **4**, 4156–4167.
- 107 Z. Yu and S. Hecht, *Chem. – Eur. J.*, 2012, **18**, 10519–10524.
- 108 Z. Yu and S. Hecht, *Angew. Chem., Int. Ed.*, 2013, **52**, 13740–13744.
- 109 Z. Yu and S. Hecht, *J. Polym. Sci., Part A: Polym. Chem.*, 2015, **53**, 313–318.
- 110 R. B. Prince, J. G. Saven, P. G. Wolynes and J. S. Moore, *J. Am. Chem. Soc.*, 1999, **121**, 3114–3121.
- 111 S. Hecht and J. M. J. Fréchet, *Angew. Chem., Int. Ed.*, 2001, **40**, 74–91.
- 112 P. D. Kiser, M. Golczak and K. Palczewski, *Chem. Rev.*, 2014, **114**, 194–232.
- 113 M. T. Stone, J. M. Heemstra and J. S. Moore, *Acc. Chem. Res.*, 2006, **39**, 11–20.
- 114 A. Tanatani, M. J. Mio and J. S. Moore, *J. Am. Chem. Soc.*, 2001, **123**, 1792–1793.
- 115 J.-L. Hou, X.-B. Shao, G.-J. Chen, Y.-X. Zhou, X.-K. Jiang and Z.-T. Li, *J. Am. Chem. Soc.*, 2004, **126**, 12386–12394.
- 116 M. Inouye, M. Waki and H. Abe, *J. Am. Chem. Soc.*, 2004, **126**, 2022–2027.
- 117 V. Berl, I. Huc, R. G. Khoury, M. J. Krische and J.-M. Lehn, *Nature*, 2000, **407**, 720–723.
- 118 Y. Tanaka, H. Katagiri, Y. Furusho and E. Yashima, *Angew. Chem., Int. Ed.*, 2005, **44**, 3867–3870.
- 119 C. Devadoss, P. Bharathi and J. S. Moore, *J. Am. Chem. Soc.*, 1996, **118**, 9635–9644.
- 120 T. Weil, E. Reuther and K. Müllen, *Angew. Chem., Int. Ed.*, 2002, **41**, 1900–1904.
- 121 W. R. Dichtel, S. Hecht and J. M. J. Fréchet, *Org. Lett.*, 2005, **7**, 4451–4454.
- 122 H. Sogawa, M. Shiotsuki and F. Sanda, *Macromolecules*, 2013, **46**, 4378–4387.
- 123 T. Hugel, N. B. Holland, A. Cattani, L. Moroder, M. Seitz and H. E. Gaub, *Science*, 2002, **296**, 1103–1106.
- 124 T. Aida, E. W. Meijer and S. I. Stupp, *Science*, 2012, **335**, 813–817.
- 125 G. Singh, H. Chan, A. Baskin, E. Gelman, N. Repnin, P. Král and R. Klajn, *Science*, 2014, **345**, 1149–1153.
- 126 H. Cölfen and S. Mann, *Angew. Chem., Int. Ed.*, 2003, **42**, 2350–2365.
- 127 Z. Yu, F. Tantakitti, T. Yu, L. C. Palmer, G. C. Schatz and S. I. Stupp, *Science*, 2016, **351**, 497–502.

