



Cite this: *Mater. Adv.*, 2024,
5, 1473

Received 7th October 2023,
Accepted 17th January 2024

DOI: 10.1039/d3ma00820g

rsc.li/materials-advances

Melanoma immunotherapy enabled by M2 macrophage targeted immunomodulatory cowpea mosaic virus†

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We have developed nanoparticle formulations targeting M2 macrophages for cancer immunotherapy by conjugating high-affinity binding peptides to cowpea mosaic virus as an immunostimulatory adjuvant. We confirmed the targeting of and uptake by M2 macrophages *in vitro* and the therapeutic efficacy of the nanoparticles against murine melanoma *in vivo*.

Main

Cancer immunotherapy is now the fourth pillar of cancer treatment following the approval of immune checkpoint blockade drugs targeting PD-1, PD-L1, and CTLA-4.^{1–3} However, its efficacy is hindered by primary resistance, acquired resistance, poor CD8⁺ T-cell infiltration, and irreversible T-cell exhaustion,^{4,5} which can be attributed to the immunosuppressive tumor microenvironment (TME).⁶ The TME is composed of tumor cells, tumor associated macrophages (TAMs), and various secreted immunosuppressive factors.⁷ Notably, M2-type pro-tumor TAMs (hereafter, referred to as M2 macrophages) play a pivotal role in the

immunosuppressive TME by hindering antigen presentation, promoting angiogenesis, tissue remodeling, and repair, suppressing the anti-tumor immune response, and promoting tumor progression and metastasis.^{7–9} M2 macrophages have therefore become a major target for the development of cancer immunotherapy.^{10,11}

Next-generation nanomedicines that target M2 macrophages can be based on synthetic or inorganic nanoparticles such as PLGA, chitosan, dextran, silica, gold, lipids and carbon.¹² One key mechanism is to reprogram pro-tumor M2 macrophages into anti-tumor M1 macrophages by incorporating Toll-like receptor (TLR) agonists, cytokines, antibodies, or RNAs.¹³ Many such formulations achieve the effective polarization of M2 macrophages, including β -cyclodextrin nanoparticles loaded with TLR7/8 agonists such as R848,¹⁴ poly(β -amino ester) nanoparticles loaded with IL-12¹⁵ or mRNA,¹⁰ and hyaluronic acid nanoparticles loaded with microRNA.¹⁶ We have developed nanoparticles based on the plant virus cowpea mosaic virus (CPMV), leveraging its natural immunostimulatory potential.^{17,18} While non-infectious toward mammals, the nucleoprotein assemblies are immunogenic and stimulate innate immune cells through multiple pathways: the highly organized and repetitive nature of the viral capsid assembly itself can be viewed as a pathogen-associated geometric pattern similar to pathogen-associated molecular patterns (PAMPs); further immunostimulation is provided through engagement with complement, presentation of T_h helper epitopes, and engagement with TLRs.^{19–21}

Here we describe the development of an intratumorally injected immunotherapy based on CPMV, which we modified to target M2 macrophages. CPMV has shown promising efficacy as an intra-tumoral agent for the treatment of murine melanoma, breast cancer, colorectal cancer, glioma, and ovarian cancer, as well as canine patients with oral melanoma and breast cancer.^{22–27} Mechanistic studies have shown that CPMV acts as a triple TLR agonist (TLRs 2, 4, and 7) to stimulate the innate immune system, thereby inducing an anti-tumor immune response.^{28,29} Intratumorally injected CPMV can

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† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d3ma00820g>

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reprogram the immunosuppressive TME into immunostimulatory by recruiting and activating N1-type neutrophils, natural killer (NK), dendritic cells, and converting M2 macrophages to M1 macrophages.²³ These activated and mature innate immune cells process tumor-associated antigens and neoantigens in the tumor to generate tumor-specific CD4⁺ and CD8⁺ effector and memory T cells.^{29–33} Therefore, to further improve efficacy by targeting and repolarization of M2 macrophages, we selected high-affinity peptides that target these cells^{34–38} and conjugated them to the surface of CPMV. We focused on two such linear peptides that bind strongly to the M2 macrophage mannose receptor, namely CSPGAK (CD206s, 561.7 Da) and CSPGAKVRC (CD206, 920.1 Da).³⁹ We evaluated CMPV particles conjugated with CD206s and CD206 for their efficacy in the treatment of murine melanoma through intra-tumoral administration.

CPMV was propagated in black eyed pea plants and purified as previously described.^{40,41} CPMV is 30 nm in diameter and consists of 60 copies of the small (S, 24 kDa) and large (L, 42 kDa) coat proteins encapsulating genomic single-stranded RNAs. Each particle also has 300 surface-exposed lysine residues suitable for bioconjugation *via* *N*-hydroxysuccinimide (NHS) chemistry.⁴² A two-step conjugation approach was used to attach the CD206s and CD206 peptides (Scheme 1). CD206s has a C-terminal cysteine residue allowing us to use a thiol-maleimide reaction in the second step for CD206s conjugation. First, CPMV was conjugated to the bispecific linker SM(PEG)₈ to introduce maleimide groups, and the subsequent cysteine and maleimide reaction allowed the formation of CPMV-CD206s particles. CD206 has cysteine residues at both the C-terminus and N-terminus, so we introduced a C-terminal azide group to ensure conjugation of CPMV to the CD206 C-terminus. CPMV was conjugated to the bispecific linker DBCO-PEG₄-NHS ester to introduce DBCO groups, and DBCO-azide click chemistry⁴³ then allowed the formation of CPMV-CD206 particles. To enable us to track particles in cell interaction studies, we also conjugated Cy5 to the surface using the same NHS chemistry with the SM(PEG)₈ and DBCO-PEG₄-NHS ester linkers. Accordingly, we produced five modified CPMV particles:

CPMV-CD206, CPMV-CD206s, CPMV-Cy5, CPMV-CD206-Cy5, and CPMV-CD206s-Cy5. Detailed conjugation protocols are provided in the ESI.†

Successful conjugation was confirmed by conducting a range of analytical tests, described in more detail in the ESI.† Native agarose gel electrophoresis confirmed the particles were intact because RNA bands detected under UV light co-migrated with protein bands detected by staining with Coomassie Brilliant Blue (Fig. 1a and b). Denaturing NuPAGE revealed two native bands (representing the S and L coat proteins at ~24 kDa and 42 kDa, respectively) – additional higher molecular weight bands were observed that corresponded to CD206(s)-modified CPMV. CD206s has a molecular weight of 561.1 Da and CD206 has a molecular weight of 920.1 Da. CPMV-CD206 resulted in two bands are around 25 kDa and 26 kDa which corresponds to S with one or two peptides conjugated. The CD206s formulation resulted in an additional band at ~24.5 kDa indicating that one peptide per S was conjugated (Fig. 1a and b). No additional bands were observed for the L protein, which is a phenomenon we often observe for modified CPMV and may indicate that the total number of peptides per CPMV is underestimated, *i.e.* the resolution of the NuPAGE is not sufficient to quantify the peptides on the L protein – or peptides were preferably conjugated to the S protein. Densitometry revealed that ~39 CD206 peptides and ~21 CD206s peptides were conjugated per CPMV particle (Fig. S1, ESI†). The elution profiles of the modified particles in size exclusion chromatography (SEC) experiments were similar to native CPMV, with a single absorbance peak synchronizing the 260 and 280 nm signals (representing RNA and proteins, respectively) thus indicating no broken particles in the column volume (Fig. 1c). However, CPMV-CD206 and CPMV-CD206s eluted at ~9 mL compared to 11 mL for native CPMV, reflecting the size increase due to peptide conjugation. The absorbance ratio at 260/280 nm was ~1.7 in all cases, indicating intact particles with packaged RNAs.⁴⁴ The size increase was confirmed by dynamic light scattering (Fig. 1d). Although the size distribution of CPMV-CD206 and CPMV-CD206s



Scheme 1 Overview of the two-step conjugation methods to produce CMPV particles displaying the peptides CD206s and CD206.





Fig. 1 Characterization of modified CPMV particles. (a) and (b) Comparison of CPMV to (a) CPMV-CD206 and (b) CPMV-CD206s by agarose gel electrophoresis and NuPAGE. (c) Analysis of CPMV, CPMV-CD206, and CPMV-CD206s by size exclusion chromatography. (d) Analysis of CPMV, CPMV-CD206, and CPMV-CD206s by dynamic light scattering. (e) Transmission electron microscopy images of CPMV, CPMV-CD206, and CPMV-CD206s.

was similar to native CPMV, the diameter increased from 38.2 nm to 62.1 nm for CPMV-CD206 and to 46.6 nm for CPMV-CD206s. This was concordant with the SEC data, again confirming that peptide conjugation increased the particle size. Specifically for the size increase for CPMV-CD206, it was possible that there is some degree of interparticle crosslinking – which may be possible through disulfide formation between the N-terminal cystines of conjugated CD206 peptides. As demonstrated by transmission electron microscopy (Fig. 1e), all formulations showed intact particles and most CPMV and CPMV-CD206s remained monodisperse, however, more CPMV-CD206s were associated together possibly due to the interparticle crosslinking. The Cy5-labeled particles were characterized in the same manner. The Cy5 signal

co-migrated with the CPMV proteins in agarose gel electrophoresis and NuPAGE experiments (Fig. S2a and b, ESI†) and the absorbance peak at 647 nm corresponded to the 260 and 280 nm peaks in SEC experiments (Fig. S2c, ESI†). UV-Vis spectrophotometry confirmed that ~32 Cy5 molecules were conjugated per CPMV particle, ~17 per CPMV-CD206-Cy5 particle, and ~15 per CPMV-CD206s-Cy5 particle (Fig. S2d, ESI†).

Having confirmed the integrity of the conjugated particles, we next evaluated their ability to target M2 macrophages *in vitro*. We prepared M1 and M2 macrophages by polarizing murine RAW264.7 cells (M0) *in vitro* using lipopolysaccharide (LPS) or interleukins IL-4 and IL-13, respectively.⁴⁵ First, we confirmed the identity of the polarized M1 and M2



macrophages by assessment of nitric oxide synthase (iNOS) expression which is characteristic for the M1 phenotype and Arginase 1 which is characteristic for the M2 phenotype.⁴⁶ The M1/M2 polarization was confirmed (Fig. 2a and b); we also observed that only M2 macrophages expressed CD206 on the cell surface post IL-4 and IL-13 simulation (Fig. 2c). M0, M1 and M2 macrophages were then incubated with the same quantity of CPMV-Cy5, CPMV-CD206-Cy5 or CPMV-CD206s-Cy5 particles in 12-well plates. After 24 h, the cells were harvested and analyzed by flow cytometry. CPMV-CD206-Cy5

and CPMV-CD206s-Cy5 were able to target M2 macrophages more efficiently than M0 and M1 macrophages as confirmed by the right-shift in Cy5 fluorescence (Fig. 2d and e). The mean fluorescence intensities (MFIs) confirmed that both peptides targeted M2 macrophages preferentially, increasing the Cy5 signal (Fig. 2g and h). CPMV-CD206s-Cy5 was taken up more efficiently than CPMV-CD206-Cy5 (Fig. 2f), as confirmed by comparing the MFIs (Fig. 2i), even though more CD206 peptides were conjugated per CPMV particle. Overall, *in vitro* analysis confirmed that the conjugation of



Fig. 2 Flow cytometry. (a) iNOS, (b) arginase 1, and (c) CD206 expression within or on the surface of M0, M1, and M2 macrophages. (d) and (e) Analysis of M0, M1 and M2 macrophages incubated with (d) CPMV-CD206-Cy5 and (e) CPMV-CD206s-Cy5 particles. (f) Analysis of M2 macrophages incubated with CPMV-Cy5, CPMV-CD206-Cy5 and CPMV-CD206s-Cy5 particles. (g)–(i) Mean fluorescence intensities (MFIs) of each sample in (d) and (e) plotted as bar graphs. Statistical significance was determined by ordinary one-way ANOVA (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$, ns = not significant).



Biosciences Inc. The other authors declare no potential conflicts of interest.

Acknowledgements

This work was supported in part by the NIH (R01-CA253615 and R01-CA224605) to N. F. S. and the Shaughnessy Family Fund for Nano-ImmunoEngineering (nanoIE) at UCSD. We thank the Nano3 core facility at UCSD for transmission electron microscopy imaging. Nano3 is the San Diego Nanotechnology Infrastructure (SDNI) of the University of California, San Diego, a member of the National Nanotechnology Coordinated Infrastructure (NNCI), which is supported by the National Science Foundation (Grant ECCS-1542148).

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