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Spectroelectrochemistry, the future of visualizing electrode processes by hyphenating electrochemistry with spectroscopic techniques

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The combination of electrochemistry and spectroscopy, known as spectroelectrochemistry (SEC), is an already established approach. By combining these two techniques, the relevance of the data obtained is greater than what it would be when using them independently. A number of review papers have been published on this subject, mostly written for experts in the field and focused on recent advances. In this review, written for both the novice in the field and the more experienced reader, the focus is not on the past but on the future. The scope is narrowed down to four techniques the authors claim to have the most potential for the future, namely: infrared spectroelectrochemistry (IR-SEC), Raman spectroelectrochemistry (Raman-SEC), nuclear magnetic resonance spectroelectrochemistry (NMR-SEC) and, perhaps slightly more controversial but certainly promising, electrochemistry mass-spectrometry (EC-MS).

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1 Introduction

1.1 Spectroelectrochemistry (SEC)

Spectroelectrochemistry is an established technique which hyphenates electrochemistry with spectroscopy. Electrochemistry

by itself is a technique that can be used in order to determine concentrations of known compounds or to obtain information concerning reaction kinetics. However, it is less suitable for elucidating unknown reaction intermediates or products.¹ By combining electrochemistry with an optical technique, more qualitative and quantitative information about the processes occurring at the electrodes can be obtained.

It is generally accepted in the SEC field that the work of Kuwana *et al.*² in 1964 is the first true SEC experiment. This

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early work on spectroelectrochemistry has resulted in a field containing a large variety of spectroscopic methods. Nowadays, a number of reviews exist concerning spectroelectrochemistry. For example the work by Dunsch from 2011, covering a wide range of multi-spectroelectrochemistry techniques.³ In 2013, Oberacher *et al.* published a paper on mass spectrometric methods in electrochemical cells.⁴ Wain and O'Connel wrote a paper in 2017 about surface-enhanced vibrational spectroelectrochemistry.⁵ Also in 2017, work by Tong on nuclear magnetic resonance spectroelectrochemistry focused on the challenges and prospects was published.⁶ The work by León and Mozo, published in 2018, describes in detail how to design a spectroelectrochemical cell.⁷ In 2018, Zhai *et al.* wrote a review in which they describe the recent advances in spectroelectrochemistry.⁸ Finally, in the work by Gazor-Ruiz *et al.* from 2019, the recent trends and challenges of spectroelectrochemistry are described.⁹

1.2 Visualizing the future of SEC

Most of the papers mentioned above are addressed towards experts in the fields, with a strong focus on recent advancements. This review tries to add to these existing review papers, firstly by focussing on the future of spectroelectrochemistry and secondly by writing a review paper in an accessible way for newcomers to the field. To be concise, the current review is focused on the four techniques that, in the opinion of the authors, have the biggest potential to undergo major improvements in the coming decades. The techniques covered in this review are infrared spectroelectrochemistry (IR-SEC), Raman spectroelectrochemistry (Raman-SEC), nuclear magnetic resonance spectroelectrochemistry (NMR-SEC) and, perhaps slightly more controversial but certainly promising, electrochemistry mass-spectrometry (EC-MS). These techniques will each be discussed in their own sections in the aforementioned

order. The basic principle of every technique is first explained, followed by the current state of the art in the field. To conclude, every chapter ends with a future perspective, based on the developments in the separate fields of the SEC technique and the substantiated opinion of the authors. The authors hope that this review will inform both the newcomers as well as the experts concerning the future of SEC. At best, we aim to give an overview of how the future of SEC may look like and at worst, to initiate a scientific discussion on the subject.

2 Infrared spectroelectrochemistry (IR-SEC)

2.1 Introduction IR-SEC

2.1.1 IR spectroscopy. In the most basic sense of the technique, infrared (IR) spectroscopy can be described as a technique where IR radiation is absorbed by molecules. The absorption of the infrared light occurs when the frequency of the absorbed radiation is equal to the vibrational frequency of the molecule. The resulting absorption spectrum provides information about the identity of the elements and structural composition of the molecule. Only vibrational modes showing a change in dipole moment are visible in the IR spectra. As a result molecules such as N₂ cannot be detected with this technique. IR spectroscopy operates over a wide spectral window between 2.5–25 μm (4000–400 cm^{-1}). The most commonly used IR technique is Fourier transform infrared spectroscopy (FTIR) (Fig. 1).

When considering IR spectroscopy, there are some key drawbacks associated with the technique. Most importantly, IR instruments often use a silicon carbide rod (such as a Globar), heated to 1000 °C or above, as a light source. Although these sources can cover a large spectral window, their power output is relatively low. This results in a relatively high detection limit



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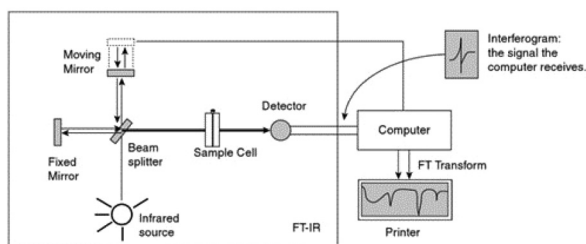


Fig. 1 Schematic diagram by Pavia *et al.*¹⁰ of an FTIR instrument. From Pavia/Lampman/Kriz/Vyvan. Introduction to Spectroscopy, 4E. © 2009 Brooks/Cole, a part of Cengage, Inc. Reproduced by permission. <http://www.cengage.com/permissions>.

compared to other analytical techniques. Another serious drawback is that water has strong absorbance bands in the IR region, which complicates measuring in aqueous solutions. In order to prevent the water bands from dominating the spectrum, Lambert–Beer law (eqn (1)) offers a practical solution.

$$A = \varepsilon \cdot c \cdot l \quad (1)$$

where A is the absorbance (arbitrary units), ε is the molar attenuation coefficient ($\text{m}^2 \text{mol}^{-1}$), c is the concentration (mol m^{-3}) and l is the path length (m). In order to reduce the absorbance effect of water, the path length is the only factor that can be changed when measuring in aqueous solutions. Meaning that in practice, the path length in FTIR of aqueous samples is kept in the order of 15 μm or lower.¹¹ This shorter path length also effects the analyte. The concentrations of analytes required for adequate signal strengths is therefore relatively high compared to other spectroscopic techniques, forming an obvious disadvantage of IR spectroscopy.

2.1.2 IR-SEC spectroscopy. As explained above, the requirement of a small cell volume is seen as a disadvantage, both due to the high sample concentration needed and difficulties in sample handling with such small volumes. However, it can be seen as an advantage when combining FTIR with EC. When the cell height is smaller than the diffusion length, total electrochemical conversion is more easily achieved.^{11,12}

Due to the requirement of such a small path length, an often used cell design in IR-SEC is the so called “thin-layer” configuration. This configuration is used as follows: the sample with electrolyte is applied on an electrode and a window of IR-transparent material (such as CaF_2) is pushed against the electrode. As a result the layer of liquid analyte is sandwiched between the electrode and the transparent IR window, with a thickness of several μm . A drawback of this method is that the exact thickness of the thin-layer is not as reproducible as with a fixed cell size, making the process of taking a background reference spectrum more difficult. The thickness of the layer can be calculated by measuring the absorbance of the bulk water vibration,¹³ potassium ferricyanide,¹⁴ or any other substance with a known concentration and molar attenuation coefficient and applying the Lambert–Beer law. This determination of the path length is not necessarily done using IR spectroscopy. De Lacey *et al.*¹⁵ used UV/Vis

spectroscopy of 8 mM cytochrome *c* to determine the path length of their thin-layer cell. Once the path length of both the sample and the background measurement is known, a background correction can be performed.

An alternative way to make a background correction when using the thin-layer configuration is by difference spectroscopy. In difference spectroscopy, a background reference measurement is performed at one potential and then subtracted from the measurement taken at a different potential. In this way, the contribution of the bulk solution is cancelled out and a spectrum is produced showing only the changes caused by the variation of the potential. There are, however, drawbacks to this technique, as adsorption and desorption of the analyte on the electrode changes the concentration of the measured analyte.¹⁶

2.1.3 Surface enhanced infrared absorption spectroscopy (SEIRAS). In order to increase the signal to noise ratio (S/N) of the measurements, researchers have been using surface enhanced infrared absorption spectroscopy (SEIRAS). After the successes in the 1970s in obtaining large enhancement factors in surface enhanced Raman spectroscopy (SERS), interest started to grow to apply similar concepts in infrared spectroscopy. The first SEIRAS experiments were reported in the 1980s by Hartstein *et al.*,¹⁷ although the term SEIRAS was only coined later.^{18–20} Since the 1990s early pioneering work in the field of SEIRAS was mostly done in the group of Osawa.^{21,22} Although not as powerful as in SERS, where local enhancement factors of up to 10^{10} have been reported,^{23,24} SEIRAS is still a valuable technique with enhancement factors of 10^1 – 10^3 being reported.²⁵ SEIRAS is performed on metallic surfaces, either in the form of roughened surfaces or arrays of nanostructures. A simplified explanation of SEIRAS is as follows: electromagnetic interactions between the IR light and the metallic structures can cause a phenomenon called plasmon resonance. Plasmon resonance amplifies the electromagnetic field, resulting in the enhancement. This enhancement only occurs close to the surface, and is negligible at distances bigger than 10 nm. SEIRAS is a complex phenomenon and the explanation above is simplified, excluding effects such as “chemical” enhancement. Therefore, the authors of this review recommend that, for an in-depth explanation of SEIRAS, the reader reads the following literature: Osawa,¹⁸ Aroca²⁶ and Neubrech.²⁷ In IR-SEC, SEIRAS is performed in combination with reflection spectroscopy and ATR spectroscopy, which will be discussed in the paragraphs below.

2.2 State of the art of IR-SEC

2.2.1 Transmission IR-SEC. In transmission IR-SEC, light emitted from the source is directed towards the sample. The sample, contained in an electrochemical cell, absorbs part of the IR radiation and the rest is transmitted through the cell towards the detector. The resulting difference between the emitted light and the detected light creates the IR spectrum. The electrochemical cell in transmission IR-SEC most frequently uses a mesh electrode configuration.^{11,15,28–31} This configuration was first reported by Murray *et al.*³² for the use



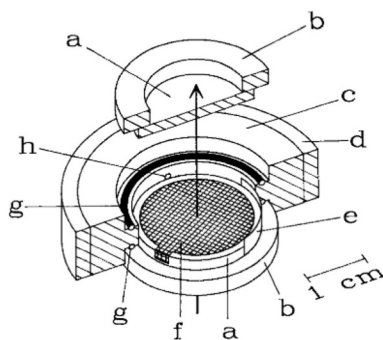


Fig. 2 Schematic representation of the IR-SEC cell. The arrow represents the propagation of the IR-light beam (a) IR-transparent CaF_2 window mounted onto a (b) Plexiglas ring, (c) Teflon body, (d) steel surround, (e) Pt counter electrode, (f) the mesh working electrode, (g) O-ring, (h) capillary connection to the reference electrode. Reprinted from the original work by Moss *et al.*,¹¹ Copyright 1990, with permission from John Wiley and Sons.

in UV/Vis-SEC and later adapted by Moss *et al.*¹¹ for the use in IR-SEC.

The design by Moss *et al.*¹¹ is shown in Fig. 2. As illustrated, a working electrode (WE) in the form of a Au mesh is sandwiched between two IR-transparent windows (CaF_2), creating a thin-layer configuration. A Teflon body keeps the cell together. The width of the cell and the WE is in the same order of magnitude as the width of the IR-light beam, allowing for the analysis of the entire cell. The design by Moss *et al.*¹¹ has been used by several researchers over the past few years with little change in the design.^{15,28–31,33} Most changes are small, such as changing the WE material into a platinum mesh instead of a Au mesh.^{30,31,33} One notable setup that does not use the above-mentioned mesh configuration is the work done in the group of Swain,^{34–36} where boron doped diamond (BDD) deposited on undoped silicon is used as transparent electrode material. This has as benefit over the mesh configuration that light is not blocked by the electrode. Among the large number of applications found in literature, some common applications for transmission IR-SEC are investigations of proteins, such as their oxidation, reduction and folding properties.^{11,15,28,29} Another application is the study of porphyrins.^{30,31,33}

2.2.2 Reflection IR-SEC

2.2.2.1 Basic principles reflection IR-SEC. The basic principles for IR reflection spectroscopy are as follows: the IR light is emitted by the light source. The IR light propagates through an IR-transparent window (often CaF_2) into the electrochemical cell. Next the light beam hits the electrode, an IR reflective material, and is reflected out of the cell, as shown in Fig. 3. The reflected beam passes back through the same window into the detector. In Fig. 3 a typical schematic representation of a reflectance cell by Alwis *et al.*^{14,37} is shown. The WE is on top of a CaF_2 window creating a thin-layer configuration. The reference electrode (RE) is separated from the cell by a Luggin capillary and the counter electrode (CE) is looped around the WE.



Fig. 3 Schematic representation of a reflection setup. Republished with permission of Journal of The Electrochemical Society, original by Alwis *et al.*,¹⁴ Copyright 2013; permission conveyed through Copyright Clearance Center, Inc.

Drawbacks of reflectance IR-SEC compared to transmittance IR-SEC mostly come from the somewhat more complex setup. When using a reflection setup it is necessary to introduce extra mirrors into the IR-spectrometer in order to focus the incident beam onto the electrode and focus the reflected beam onto the detector. Aligning these mirrors can be a time-consuming process, more labour intensive compared to transmission spectroscopy. The use of extra mirrors also introduces extra losses due to part of the light being absorbed instead of being reflected. An advantage of using reflection compared to transmission spectroscopy is that no light is blocked by the electrode material.

2.2.2.2 Fibre optic reflection IR-SEC. An alternative for the standard reflection setup is fibre optic reflectance spectroscopy. First developed by Shaw *et al.*³⁸ and used over the past decade primarily in, or in collaboration with, the group of Richter-Addo.^{38–41} The advantage of this specific configuration compared to reflection systems such as those by Alwis *et al.*^{14,37} is the ease of alignment of the source with the sample and electrode, allowing for smaller configuration of the setup. With the small thin-layer setup they claim better control over the applied potential than transmittance and ATR cells.³⁸ This particular configuration is mostly applied to the study of porphyrins,^{39–41} although it should be noted that it can also be used for applications where reflection IR is currently used.

2.2.2.3 SNIFTIRS and PM-IRRAS. As mentioned in section 2.1.2, IR-SEC often operates with a thin-layer configuration. In reflection mode there are two different methods in order to perform background corrections. The first method, based on difference spectroscopy and most often reported in literature, is subtractive normalized interfacial Fourier transform infrared reflection spectroscopy (SNIFTIRS). Alternatively, polarization modulation infrared reflection absorption spectro-



scopy (PM-IRRAS) is used. SNIFTIRS is an IR-SEC reflection technique first introduced in the early 1980s by Pons.⁴² The authors would like to note that the use of the term SNIFTIRS seems to be used inconsistently throughout literature. Several papers use the acronym SNIFTIRS,^{14,37,42–44} while other papers, seemingly using SNIFTIRS, don't make use of the term.^{13,45–50} SNIFTIRS is a technique used for a number of different applications, such as the study of the oxidation of small organic molecules (such as ethanol and ethylene glycol)^{13,45,48,49} and metal ions.^{14,37,43} SNIFTIRS is also performed in combination with SEIRAS, for example for the studies of proteins.^{46,47}

PM-IRRAS, an alternative for SNIFTIRS, was first performed by Golden *et al.*⁵¹ in 1981. After this initial work, further notable improvements on the technique were done by Faguy *et al.*,^{52,53} the group of Corn^{54,55} and the group of Lipkowski.^{56–59} PM-IRRAS is most often used in cases where the potential modulation necessary for SNIFTIRS could damage the analyte of interest.¹⁶ Additional advantages of PM-IRRAS over SNIFTIRS, as proven by Faguy *et al.*,^{52,53} include that PM-IRRAS is very insensitive to atmospheric water and carbon species and has an increased signal to noise ratio.^{59,60} PM-IRRAS is especially suitable for measurements of samples adsorbed on metal surfaces, such as electrodes. Explained in a simplified manner, in PM-IRRAS the polarization of the reflected light is modulated and the reflection takes place under a grazing angle of incidence. Under s-polarization (perpendicular to the plain of incidence) there is no interaction between the light and electrode surface. However, when using p-polarization (parallel to the plain of incidence), constructive interference takes place at the electrode surface. By cycling the polarization of the light, a difference spectrum of the bulk (s-polarization) and at the electrode surface (p-polarization) can be obtained. PM-IRRAS has been reported in literature as, among others, a method for the study of self-assembled monolayers^{61,62} and lipid bilayers.^{56–59}

For a more in-depth description of SNIFTIRS and PM-IRRAS the authors recommend reading the work of Guidelli,¹⁶ Bard⁶³ and Alkire *et al.*⁵⁹

2.2.2.4 Electrode material and SEIRAS in reflection IR-SEC. Concerning SEIRAS in reflection IR-SEC, electrode materials used are Ni,^{14,37,43} Pt,^{13,38–41,50,64} Pd^{45,48,49} and Au.^{46,47,62,65,66} While only Liu *et al.* report on using the reflecting electrodes for surface enhancement,^{46,47} it is unlikely that the Au electrodes used by Hosseini *et al.*,⁶² Su *et al.*⁶⁵ and Villalba *et al.*⁶⁶ don't report any enhancement. For Pt, Pd and Ni, SEIRAS has been reported in the past,⁶⁷ but none of the papers above mentioned FTIR-SEC papers report this effect. This might be because it is not relevant for their application or that the SEIRAS effect is negligible.

2.2.3 Attenuated total reflection (ATR) IR-SEC

2.2.3.1 Basic principles of ATR-IR-SEC. Attenuated total reflection infrared spectroscopy (ATR-IR) operates under the following principle: IR light is coupled into an internal reflection element (IRE), as shown in Fig. 4. When the light hits the sample interface at an angle higher than the critical angle



Fig. 4 Internal reflection element. Reprinted and adapted from the original work by Schadle and Mizaikoff,⁶⁸ Copyright 2016, with permission from SAGE Publications.

(θ_{crit}), total internal reflection occurs, reflecting the light and creating an evanescent wave. This evanescent wave penetrates into the sample for several micrometres, with the electric field decaying at an exponential rate away from the surface. This has the advantage that the effective path length in ATR-IR is always in the order of several μm , and thus removes the need for a thin-layer configuration. In order to get a good estimation of the penetration depth in ATR-IR-SEC the following formula can be used:⁶⁸

$$de = \frac{\lambda}{2\pi \sqrt{\sin^2 \theta - \left(\frac{n_2}{n_1}\right)^2}} \quad (2)$$

In this equation de is defined as the penetration depth (m) at which the intensity of the wave is reduced to $1/e$ of its original value, or approximately 37%. Furthermore λ is the wavelength (m) used, n_1 is the refractive index of the IRE, n_2 the refractive index of the sample and θ is the angle of incidence ($^\circ$).⁶⁸ This formula can give a good estimation of the penetration depth and the effect of the IRE material on the penetration depth of the system. It can consequently be used to get an estimate of the path length through the sample.

2.2.3.2 IRE material ATR-IR-SEC. Penetration depth is one reason to choose for a certain ATR material, other reasons are availability, hardness and inertness. In Table 1, three different materials used for the IRE in ATR-IR-SEC are shown: diamond, ZnSe and Si. Diamond is more expensive than ZnSe or Si, but its large spectral window^{68,69} (2–25 μm), its chemical inertness and the high hardness are clear advantages. ZnSe is cheaper than diamond and has a similar spectral window^{68,69} (1–25 μm) but is relatively soft and can only be used between pH 5–9. Finally, and also cheaper than diamond, there is Si which is harder than ZnSe and somewhat more inert, but suffers from a reduced spectral window^{68,69} (2–7 μm). Surprisingly, Ge, a commonly used ATR material, does not seem to be used in ATR-IR-SEC. A possible reason for this is the fact that Ge is more conductive than diamond, ZnSe and Si and therefore could potentially cause interference with the simultaneous operation of the electrochemical cell.

Another variable in Table 1 is the configuration of the beam path through the IRE. As shown in the table, a single bounce or a multi bounce configuration can be used. Reasons to choose a certain configuration include the law of Lambert-



Table 1 Material and configuration used for IRE and electrode in ATR-IR-SEC. Setup I: Optically transparent electrode (Fig. 5), setup II: SEIRAS (Kretschmann configuration), setup III: spacer setup (Fig. 6)

IRE	Setup	Electrode material	Configuration	Author
Diamond	III	Carbon nanofibres	Single bounce	Dillard <i>et al.</i> ⁷⁷
Diamond	I	Boron doped diamond	Single bounce	Neubauer <i>et al.</i> ⁸²
Diamond	II	Boron doped diamond/Au nanoparticles	Single bounce	Izquierdo <i>et al.</i> ⁸¹
Si	III	Carbon paper	Multi bounce	López-Lorente <i>et al.</i> ³¹⁷
Si	III	Carbon paper	Multi bounce	Healy <i>et al.</i> ⁷⁶
Si	II	Au/Pd	Not mentioned	Paengnakorn <i>et al.</i> ⁷¹
Si	II	Au	Not mentioned	Yang ⁴⁸
Si	II	Au	Single bounce	Chequepán <i>et al.</i> ³¹⁸
Si	II	Cu/Au	Single bounce	Dunwell <i>et al.</i> ³¹⁹
Si	II	Au	Single bounce	Heyes <i>et al.</i> ⁷⁸
ZnSe	I or II	Sn/SnO _x	Single bounce	Ataka <i>et al.</i> ²¹
ZnSe	I	Nitrogen doped diamond like carbon	Multi bounce	Baruch <i>et al.</i> ⁷⁴
ZnSe	I	Au	Single bounce	Menegazzo <i>et al.</i> ⁸³
ZnSe	I	Boron doped silicon	Single bounce	Pfaffeneder-Kmen <i>et al.</i> ⁷⁹
ZnSe	II	Au	Single bounce	Purushothaman <i>et al.</i> ⁷⁰
ZnSe	II	Au	Multi bounce	Viinikanoja <i>et al.</i> ⁸⁰
				Zimmermann <i>et al.</i> ³²⁰

Beer (eqn (1)), since a multi bounce configuration, where the light interacts more often with the sample, effectively has a longer path length.

2.2.3.3 Electrode material and configuration in ATR IR-SEC. Also reported in Table 1 are the three different setups for the placement of the WE. The first configuration (denoted as I in Table 1), is the so called optically transparent electrode. This is either made out of a thick Au mesh deposited on the IRE, similar as those reported in transmission IR-SEC (2.2.1), or an optically transparent layer deposited on top of the ATR crystal. These optically transparent electrodes, such as boron doped diamond (BDD), nitrogen doped diamond like carbon (N-DLC) or boron doped silicon (BDSi), could potentially also be used as ATR material, but this does not seem to be applied in practice. A possible reason for this could be fabrication limitations. A schematic representation of an ATR-IR-SEC design using the optically transparent electrode configuration can be seen in Fig. 5.

The second configuration (denoted as II in Table 1), is similar to the first configuration in the sense that the electrode material is deposited on the top of the IRE, but instead of an optically transparent material or mesh, a thin layer of metal is deposited. This setup, known as the Kretschmann

configuration,^{72,73} is a method in ATR FTIR to perform SEIRAS experiments. The thin metal layer here acts as an electrode and as a surface for SEIRAS simultaneously. This thin-layer electrode has some drawbacks: if the layer is too thick, too much IR absorbance by the material occurs, if it is too thin, holes in the continuous layer might occur. Baruch *et al.*⁷⁴ deposited a thin layer of Sn/SnO_x as electrode material on top of an ATR crystal. They do not mention any SEIRAS effects, but one might argue that it is likely that SEIRAS does take place. The reasoning for this is that Sn has been reported to have surface enhanced properties in the infrared region,⁷⁵ making it debatable whether it should be in configuration I or II.

The final configuration (denoted as III in Table 1) uses a thin porous spacer between the IRE and the electrode. As one can see in Table 1 and in Fig. 6, a thin porous layer, either a Nafion membrane,⁷¹ carbon nanoparticles⁷⁶ or carbon fibres,⁷⁷ is placed on top of the IRE. A carbon paper electrode is placed in contact with this layer, covering the surface and functioning as a WE. This setup has as advantage that it does not need labour intensive deposition techniques.



Fig. 5 Schematic of an optically transparent electrode ATR IR-SEC setup, in a single bounce configuration. Reprinted with permission from Springer Nature: from the original work by Purushothaman *et al.*,⁷⁰ Copyright 2012.



Fig. 6 Schematic representation of the cell configuration using a spacer between the IRE device and the electrode. As shown, the Pt CE and calomel RE have more space than in the thin-layer configuration used for transmission and reflectance IR-SEC. In the inset one can see the IRE, the porous Nafion membrane containing the analyte and the carbon paper electrode on top. A multi-bounce configuration is used. Reprinted from the original work by – Paengnakorn *et al.*,⁷¹ Copyright 2017, published by The Royal Society of Chemistry.



A large number of applications regarding ATR-IR-SEC are reported in literature. Some examples are the study of metalloproteins,^{71,76} the study of CO₂ reduction without⁷⁴ and with the combination of SEIRAS,⁷⁸ the reduction of graphene oxide,^{79,80} technique development^{81–83} and practical applications such as observing lubricant degradation.⁷⁰

2.2.4 IR-SEC microscopy. In IR microscopy, the light from an IR light source is coupled into an IR microscope. Just as with conventional microscopy, the spatial resolution is limited by the diffraction limit of the light. Since IR light has a relatively large wavelength compared to visible light, the spatial resolution is, at its lowest, in the order of about 5 μm . Nowadays, infrared microscopes with focal plane arrays are available, allowing not only IR microscopy but also imaging the sample. The combination of IR microscopy and SEC has found some limited applications by using either a conventional source or synchrotron radiation.

2.2.4.1 Conventional source. The most notable work in using IR-SEC spectroscopy with a conventional source comes from the group of Sun,⁸⁴ who developed their own IR-SEC microscope in the late 1990s. They further developed this instrument in the last two decades for a variety of different applications regarding electrode interface reactions.^{85–87} Sun and Zhou also wrote a paper in which they further explain the use of FTIR microscopy for electrochemical applications.⁸⁸

2.2.4.2 Synchrotron source. Next to IR microscopy using conventional IR sources, instruments using synchrotron radiation have also been reported. The first experiments regarding the use of synchrotron radiation with electrochemistry were reported by Melendres *et al.*⁸⁹ in 1995, although the IR frequency used was in the far infrared. The first experiment that coupled mid infrared synchrotron radiation with electrochemistry was the work by the group of Burgess⁹⁰ in 2011 where they proved that, when using small aperture settings, synchrotron radiation has a better signal to noise ratio than conventional sources. Further notable work with synchrotron IR microscopy combined with electrochemistry was done by the group of Burgess on spatial mapping of methanol oxidation on a PtNi alloy,⁹¹ femtomole detection of ferrocyanide on ultra-microelectrodes⁹² and quantitative analysis of electrochemical diffusion layers.⁹³ Other notable work regarding synchrotron radiation in combination with electrochemistry was done by Ash *et al.*⁹⁴ on synchrotron based infrared microscopy to study biological redox processes under electrochemical control. In this paper they show that they can observe changes in the Flavin mononucleotide active site of a flavoenzyme with synchrotron radiation.

Although the power of synchrotron radiation is proven by the work done by researchers mentioned above, it has to be noted that the need of synchrotron radiation is a major drawback to using synchrotron based IR spectroscopy. Access to a synchrotron source is limited and will stay so in the foreseeable future.

2.3 Future perspective

The fields of IR and SEC are ever increasingly combined and advanced. Recently, a three-electrode electrochemical ATR cell

from Pike Technologies called the Jackfish SEC Cell has appeared on the market. The J1W version of this device uses commercially available disposable IRUBIS ATR-SEIRAS devices, making IR-SEC suitable for a broader audience.⁹⁵

It can be postulated that further improvements in the field of IR-SEC are linked to the advancements made in the fields of IR spectroscopy and electrochemistry. These advancements will take some time to be implemented into IR-SEC, but it is logical to assume that any advancement improving the separate fields, will eventually be implemented in IR-SEC.

2.3.1 New sources for IR-SEC. One of the major drawbacks of IR spectroscopy is the low intensity of the light source. High power sources are available in the form of optical parametric amplifiers (OPA) and optical parametric oscillators (OPO) but these systems are expensive and labour intensive to both operate and construct. A more affordable, user friendly and compact option is a quantum cascade laser system (QCL). Some QCL instruments such as IRSweep's Iris-f1 are complete spectrometers, making IR-laser systems more available. In Fig. 7, a comparison between the output of a QCL by Daylight Solutions, a synchrotron and a Globar source is made by Weida and Yee.⁹⁶

The comparison as shown in Fig. 7 is made when passing the light through a 10 μm pinhole. Subsequent spot sizes of the three techniques are normalized. This comparison highlights the strength of QCLs in IR-microscopes, yet for normal spectroscopy this graph is skewed somewhat in favour of the QCL device. Nonetheless, the main message stays the same, being that a higher source power can be obtained with QCLs. Due to the higher power output of the QCLs, a shorter measurement time compared to regular IR instruments can be



Fig. 7 Comparison of Weida and Yee of three different IR sources of the signal through a 10 μm pinhole.⁹⁶ Reprinted and adapted from the original work by Weida and Yee,⁹⁶ Copyright 2011, with permission from SPIE and M. Weida.



achieved. A commercially available QCL system from IRSweep, using optical heterodyne detection, even further improves this time resolution. This IRSweep instrument can take a spectrum with a spectral width in the order of 100 cm^{-1} within $4\text{ }\mu\text{s}$. This allows for a more accurate understanding of the dynamic processes occurring at the electrode interface.

A drawback of QCLs is the shorter wavelength coverage compared with standard IR sources. The wavelength range of a single QCL is usually somewhere in the order of a micrometre. This can be compensated to some extent by having several lasers in a system. QCLs have already been used for several IR techniques,^{97,98} and therefore could be implemented in IR-SEC in the near future.

2.3.2 From IRE to (single mode) waveguides. When having access to laser based sources the use of waveguides instead of IREs also becomes more attractive. A simplified explanation of waveguides can be found in Fig. 8. In Fig. 8a and b an IRE is shown, while Fig. 8c displays a single mode waveguide. The figure illustrates that reducing the size of the IRE increases the amount of bounces the light makes through the device, effectively increasing the path length. In Fig. 8c the thickness of the device has been reduced to such an extent that it cannot be considered an IRE anymore, but a waveguide with a continuous evanescent field along the length of the device.⁶⁸ An added benefit of making the IRE smaller is that the losses due to the absorption of the IRE material are reduced, since the effective path length through the IRE decreases. While making the IRE smaller can improve the signal to noise ratio, it can also make it more difficult to couple light into the device. By using a source with a higher power and small beam size, such as the aforementioned QCLs, these losses might be mitigated. According to literature, a waveguide might lead to significantly increased sensitivity compared to an IRE device.^{68,69} Several

works on infrared waveguides^{99–106} have already been published and it is therefore not unthinkable that the first IR-SEC waveguide will be published in the near future.

2.3.3 Novel electrode material. An interesting material for IR-SEC to be used as a (true) transparent electrode in transmission mode or as waveguide material is BDD. BDD is optically transparent for infrared light and can be used as an electrode. BDD has already been used as transparent electrode material on top of an IRE⁸² and as an optical electrode in the transmission configuration.^{34–36} However, it might be possible to use the diamond layer as IRE/waveguide and electrode simultaneously. This would make it possible to measure as close to the electrode as physically possible. A drawback of using BDD as a waveguide are the extra losses the boron doping causes at wavenumbers of 1800 cm^{-1} and higher.^{34–36,107}

When the already small waveguide also functions as electrode, a device with a small footprint can be created. This way it can be easily combined with microfluidics in order to make lab-on-a-chip systems.

2.3.4 Microfluidics and lab-on-a-chip systems. The hyphenation of electrochemistry and microfluidics and lab-on-a-chip systems is already well established. For a review regarding this subject the authors recommend to read the paper by Odijk *et al.*¹⁰⁸

Although less established than electrochemistry, the combination of IR with microfluidics is also a developing field, for example: an ATR-IR device for the study of electric field-driven processes,¹⁰⁹ reaction monitoring¹¹⁰ and the analysis of chemical reaction intermediates.¹¹¹ A recent review written on spectroscopic microreactors for heterogeneous catalysis by Rizkin *et al.* also includes a section about IR.¹¹²

The combination of IR-SEC and microfluidics is not an established field yet, although some work has been done by Führer *et al.*¹¹³ It is expected that in the near future more work regarding IR-SEC is published.

2.3.5 Improving spatial resolution. As mentioned in section 2.2.4, the optical IR microscopy imaging is limited to a spatial resolution in the order of $5\text{ }\mu\text{m}$. Two different atomic force microscopy (AFM) based techniques can improve the spatial resolution of IR techniques, namely photothermal induced resonance (PTIR) and Nano-FTIR.

2.3.5.1 PTIR. PTIR is a technique that combines AFM with infrared spectroscopy. This hyphenated technique has the optical resolution of the AFM technique and the ability to obtain the chemical information usually obtained with IR. In PTIR, the probe tip of an AFM microscope is positioned close to the region of interest. Next, a tuneable IR laser is focused on the region close to the probe tip. If the sample has a vibrational frequency that corresponds to the wavelength emitted by the laser, absorption will take place. This absorption results in a thermal expansion of the sample, which then will be measured by the AFM cantilever.^{114,115} The resulting spatial resolutions for commercially available instruments are in the order of 20 nm , at least two orders of magnitude lower compared to optical microscopy.^{114,115} To our knowledge, this technique has not yet been combined with SEC. One possible

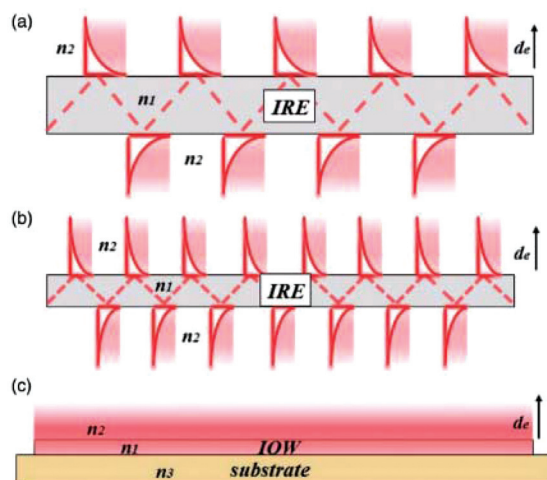


Fig. 8 Schematic representation of the evanescent wave/field with an ever decreasing IRE thickness. As one can see in (a) and (b), the smaller IRE has more reflections than the bigger IRE. When the size is small enough, as one can see in (c), a waveguide is created with a continuous evanescent field. Reprinted from the original work by Schadle and Mizaikoff,⁶⁸ Copyright 2016, with permission from SAGE Publications.



explanation could be that SEC introduces too much heat in the sample, which could interfere with the AFM measurement.

2.3.5.2 Nano-FTIR. In Nano-FTIR, FTIR and scattering-type scanning near-field optical microscopy (s-SNOM) are combined. IR light from a conventional IR laser source or a synchrotron source is focused on a conductive AFM probe tip. This probe tip acts as an antenna for the IR light and focuses and enhances it. The near field interaction between the probe tip and the sample influences how the light is being scattered. Recording the scattered light results in a spectrum of the sample. While imaging the surface, a map with special resolution as small as 10 nm can be obtained.^{114,116,117} One application of Nano-FTIR combined with SEC was reported by Lu *et al.*¹¹⁶ where they study the molecular structure of graphene-liquid interfaces, allowing new possibilities for the characterization of graphene-electrolyte interfaces.

2.3.6 Advanced SEIRAS structures. Finally, SEIRAS could be improved by replacing rough metal films with nanofabricated structured SEIRAS arrays or slits. Nanofabricated plasmonic nanoantennas have shown to be promising for the use in SEIRAS.¹¹⁸ These devices have been reported to have an improved SEIRAS effect, although they usually only operate at a small wavelength range. This however would make them a suitable to be used with QCLs, as they can be tuned to the same wavelength as the relatively narrow-band QCL laser.

3 Raman spectroelectrochemistry (Raman-SEC)

3.1 Introduction Raman-SEC

3.1.1 Raman spectroscopy. Just like IR spectroscopy, Raman spectroscopy is a technique used to get information from the vibrations of molecule. In short, Raman spectroscopy operates based on the Raman scattering effect. A light source, often a laser with a wavelength in the near infrared, visible or near ultraviolet, is directed at the sample. When the light interacts with the sample, photon scattering takes place. Most of the light is elastically scattered from the sample, also known as Rayleigh scattering, and has the same wavelength as the incoming light. A small percentage of the light is inelastically scattered, also known as Raman scattering, resulting in a shift in wavelength corresponding to the frequency of a molecular vibration. The shift can be to higher wavelengths (Stokes shift) or lower wavelengths (anti-Stokes shift).

Raman is often considered to be complementary to IR spectroscopy. The reason for this is that, in order to be visible in Raman spectroscopy, the vibration has to experience a change in polarizability. In other words, a change in how easy it is to distort the electron cloud around the molecule. As mentioned in the previous section, IR active molecules require a change in dipole moment in order to be visible in IR spectroscopy. Often vibrations that are polarizable do not have a dipole moment and *vice versa*, making the techniques complementary.



Fig. 9 Schematic representation of a Raman setup. Reprinted from the original work by Schmid and Dariz,¹¹⁹ Copyright 2019, with permission from Creative Commons. Licensed under CC BY 4.0.

A typical setup used in Raman spectroscopy is shown in Fig. 9, as reported by Schmid and Dariz.¹¹⁹ It consists of a light source, most often a laser, focused on the sample using a microscope objective. The scattered light is guided back into the instrument and a filter removes the Rayleigh scattered light. The Raman scattered light is guided onto the grating of a monochromator and eventually reaches the detector, where it records the resulting spectrum.

3.1.2 Surface enhanced Raman spectroscopy (SERS). Fleischmann *et al.*¹²⁰ made the first observation of SERS in 1974. Since then the work by the group of Van Duyne,^{121,122} the group of Moskovits,^{123–125} Otto¹²⁹ and Schlücker¹²⁶ further pioneered the field of SERS. In a simplified explanation, SERS is a technique that is used to enhance a Raman signal, similar to SEIRAS. This is primarily done through the electromagnetic interaction of the incident Raman light beam with the metal surfaces.^{19,24} This results in a phenomenon called plasmon resonance which amplifies the laser field.²⁴ This phenomenon only occurs very close to the surface of the metal substrate. Typically, the analyte is required to be relatively close to the substrate, not further away than 10 nm.^{19,24} The substrate, most often Ag or Au, should contain microstructures, either periodically nanofabricated or randomly orientated as encountered in roughened surfaces, to increase the intensity of the enhancement.

A second factor for enhancement that should be discussed is the “chemical” enhancement. Unlike the well-defined electrochemical enhancement, the term “chemical” enhance-



ment describes more of a collection of different phenomena.¹²⁷ Simplified, the chemical enhancement mostly describes phenomena due to the charge transfers between the analyte molecule and the metal substrate and other metal substrate/analyte molecule interactions. The chemical enhancement is significantly lower than the electrochemical enhancement, and much harder to theoretically explain and predict.¹²⁶

In practice, distinctions can be made between three different kinds of SERS substrate. The first are randomly orientated enhancement surfaces, such as roughened (electrode) surfaces and randomly orientated nanoparticles deposited or grown on a surface, which are relatively easy to fabricate. Second are the so called structured arrays, which can exist of a self-assembled array nanoparticles deposited on a substrate or nanofabricated arrays. These structured arrays can obtain higher enhancement factors than the roughened surfaces but are more difficult to fabricate. Finally, in order to improve measuring in liquid samples, metallic particles can be used in order to create SERS active colloidal solutions.²⁴ This last form, using colloidal particles, cannot be used in SERS-SEC, since it cannot function as or be in contact with the WE easily. With the right combination of SERS substrate material and configuration of the structures, enhancement factors of up to 10^{10} can be achieved, depending on the used definition of SERS enhancement.²⁴

One important note on working with nanoparticles is the need for proper cleaning of the particles in order to maximize the SERS effect. The recent work of Montiel *et al.* describes some of the methods that can be used for the cleaning of these particles.¹²⁸

SERS is a complex field with persistent misconceptions.¹²⁴ Therefore a complete review on the subject is beyond the scope of this study. The authors of this review recommend that the reader reads the work cited for a more in-depth explanation of SERS.^{24,67,120–126,129}

3.1.3 Raman-SEC. In Raman-SEC, a Raman instrument, often a Raman confocal microscope, is coupled to an electrochemical cell. Compared to FTIR SEC, a Raman setup is somewhat more versatile in the cell design. Since Raman is a scattering technique, it is not limited to transparent electrodes as is the case with transmission IR-SEC. Measurements usually take place at the electrode surface, so the laser is most often used in a configuration where it is focused on the electrode. Even compared to reflection and ATR IR-SEC, the setup of the cell can be considered more user friendly. The optical window of the cell can be made out of standard materials since a visible light source can be used. The thin-layer configuration is not necessary, as water does not have strong absorptions in typical wavelengths used for Raman spectroscopy. Moreover, confocal lenses are often used. Once access to a Raman instrument is available, combining it with SEC is therefore relatively straightforward.

3.1.4 SERS-SEC. As written above, the first report of the observation of the SERS phenomenon is considered to be made by Fleischmann *et al.*¹²⁰ In this experiment, SERS was observed by studying pyridine on a Ag electrode. This experi-

ment highlights the good synergy between SEC and Raman. The metallic electrodes used for the electrochemistry are naturally suited to perform SERS measurements, and adapting an electrode to be more SERS active usually does not negatively interfere with the electrode behaviour. For this reason, Raman-SEC measurements will experience the SERS effect to some extent, which can be either negligible or indeed the selling point of a certain configuration.

3.2 State of the art of Raman-SEC

3.2.1 Cell configuration and electrodes. There is a large variety of different materials for Raman electrodes. Reports range from standard metallic electrodes such as Au electrodes, to cells made by 3D printing,¹³⁰ screen-printed electrodes^{131–138} and even electrodes on fabric.¹³⁹

In the literature, several different cell configurations are described. Most commonly used is the stationary electrode configuration. Other configurations include the rotating cell, the linearly moving cell or the flow through cell. Two important reasons to choose a certain cell are the ease of use and the balance of fabrication *versus* potential photo-degradation. Photo-degradation of samples can be a severe problem in Raman spectroscopy. Constant illumination of a substrate can heat up the sample and eventually cause burning or evaporation of the sample or matrix. In the next section, we will discuss the different cells, their advantages and disadvantages, and how they deal with photo degradation.

3.2.1.1 Stationary electrode. The stationary electrode, as shown in Fig. 10 is the most commonly used configuration. Since the laser is stationary with respect to the electrode, alternatives should be considered in order to reduce damage due to the laser beam. Depending on how susceptible the sample is to photo degradation, one should limit the laser power and the exposure time of the sample.

3.2.1.2 Linearly moving electrodes and cells. In order to reduce photo degradation, several setups have been designed to limit the time the Raman laser remains on a single spot on the substrate. One such a device operates by moving the entire cell and its electrode as can be seen in Fig. 11. The cell moves



Fig. 10 Example of a stationary electrochemical cell used for Raman experiments. Reprinted from the original work by Fleischmann *et al.*,¹²⁰ Copyright 1974, with permission from Elsevier.





Fig. 11 Schematic of a moving electrode cell. Ne is the Neon lamp and D is the rotating disk rotating around the axis represented by the black dot. Reprinted from the original work by Niaura *et al.*,¹⁴⁰ Copyright 1997, with permission from John Wiley and Sons.

together with the electrode in a linear path with respect to the Raman light source, reducing the time for which a single spot on the electrode is exposed to the Raman source. The linear moving electrode was first developed by Niaura *et al.*¹⁴⁰ and is mostly used by Mažeikienė *et al.* to study polyaniline,^{141–146} dyes^{147–150} and hexacyanoferrates.^{151,152}

3.2.1.3 Rotating electrodes and cells. Next to the moving electrode system, a rotating Raman system has been first developed by Kiefer and Bernstein¹⁵³ in 1971. One of the first rotating Raman-SEC setups was reported by Yamada *et al.*¹⁵⁴ in 1978, as shown in Fig. 12. An advantage from an electrochemical point of view for both the rotating electrode and the moving electrode is that there are no mass transport limitations due to the added convection. Some use of the rotating

electrode has been reported in the 70's and 80's,^{154–158} but, despite the potential of the rotating electrode, little mention of its use is reported in recent literature. An alternative to the rotating electrode configuration is the rotating cell configuration, in which instead of the electrode the entire cell is being rotated. This device was first used by Kavan *et al.*¹⁵⁹ in 2009. The WE is positioned at the centre of the motor axis while the sample rotates. Despite the potential for this setup, it is only mentioned once more in literature, for Fullerene C₆₀ in an aqueous medium.¹⁶⁰

3.2.1.4 Flow through cell. Another way to reduce the chance of photo-degradation is a flow through cell, which is easy to use for non-viscous liquids, but not applicable for solid samples. Most commonly, a flow through cell either operates by having a constant flow of fresh liquid flowing over the WE^{161,162} or by continuous mixing of the electrolyte solution.¹⁶³

3.2.1.5 Alternative designs. A special configuration that is worth mentioning is the setup by Ibañez *et al.*,¹⁶⁴ using a 4-electrode system in order to study the charge transfer at a polarisable liquid/liquid interface. Another interesting design is the Raman spectroscopy of a carbon super capacitor by Bonhomme *et al.*¹⁶⁵

3.2.2 Application Raman-SEC

3.2.2.1 Materials. Raman-SEC is used in a multitude of research fields and for different applications. An important research field for Raman-SEC is material science to investigate novel materials. One example is the research done on carbon nanotubes. Most of this extensive research comes from the groups of Dunsch and Kavan,^{166–177} although some work has been performed elsewhere.^{178–183} A related field is the investigation of graphene.^{162,163,184–190} Another popular application of Raman-SEC within material science is to get more insight on conductive polymers, studied by the group of Malinauskas^{141–146,191} and others.^{192–202} A number of applications are reported on the investigation of dyes,^{147–150,203,204} small organic molecules that can form monolayers on the electrode material^{205–209} and protein/cell studies.^{140,210–212}

3.2.2.2 Sensing applications and point of care. Research aimed towards real world applications is also present in Raman-SEC, although underrepresented and almost solely performed by the group of Brosseau. Some interesting applications include a portable SERS-SEC device for the detection of melanine in milk,¹³⁶ the detection of DNA biomarkers in synthetic urine,¹³⁷ the detection of 6-thiouric acid in synthetic urine,²¹³ the quantitative detection of uric acid in synthetic urine¹³⁸ and a point of care device based on SERS-SEC on a fabric substrate.¹³⁹

3.3 Future perspective

Raman-SEC is already a more mature technique than for example IR-SEC. This does not mean, however, that the technique has stopped to progress. The authors of this review are of opinion that most future development will take place on the SERS substrates and the development of new applications for Raman-SEC, especially towards real world applications.

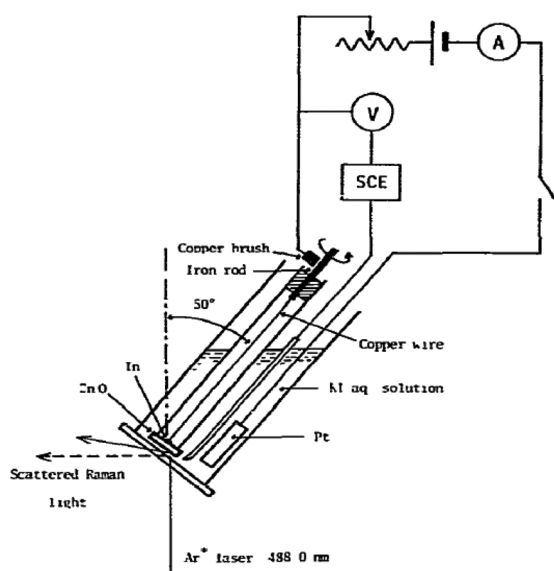


Fig. 12 Rotating Raman-SEC device. Reprinted from the original work by Yamada *et al.*,¹⁵⁴ Copyright 1978, with permission from Elsevier.



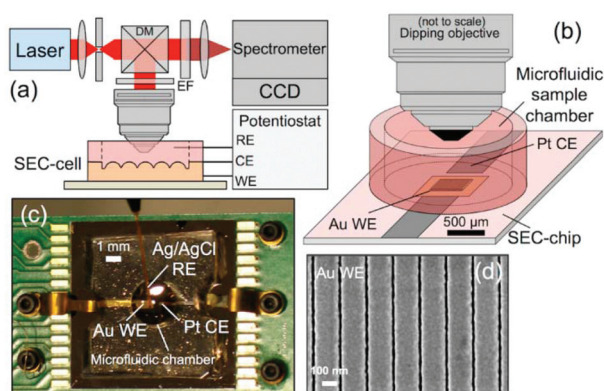


Fig. 13 *In situ* SERS-SEC system by Yuan *et al.*²¹⁴ (a) Schematic representation of the Raman SEC analysis system, (b) SEC-cell composed of a SEC-chip directly bonded to a small-volume microfluidic sample chamber with an optical interface to a microscope objective, (c) SEC-cell consisting of a Pt CE and nanostructured Au WE patterned on the Si SEC-chip and an external Ag/AgCl RE, and (d) representative SEM image of the Nanostructures Au WE. Reprinted with permission from Yuan *et al.*²¹⁴ Copyright 2015 American Chemical Society.

3.3.1 Nanofabricated SERS structures. Further developments towards structured nano arrays for SERS applications are expected. As is shown in Fig. 13 by Yuan *et al.*,²¹⁴ highly structured nano arrays consisting of thin Au lines can be used as SERS substrate, resulting in high enhancement factors while still functioning as electrode. New lithographic techniques make the wafer-scale fabrication of such substrates easier,²¹⁵ while novel methods to improve the adhesion of these Au structures on silicon²¹⁶ have also been developed in recent years.

Compared to, for instance, Au islands and randomly distributed particles, these nanostructured arrays can achieve a higher reliability, since the enhancement is higher and equal over the entire surface. This makes these structures more suitable for quantitative measurements compared to the randomly organized substrates.

3.3.2 Electrochemical-surface oxidation enhanced Raman scattering (EC-SOERS). EC-SOERS is a relatively new technique that was first introduced by Perales-Rondon *et al.*^{217,218} According to Perales-Rondon *et al.*, EC-SOERS cannot be easily explained using the classical models of SERS.^{217,218} In EC-SOERS, enhancement is only observed when the Ag WE is undergoing anodic oxidation. They postulate that during this process nanostructures are formed that are favourable for SERS enhancement. Garoz-Ruiz claims that this technique might be an alternative for molecules that are usually not very sensitive in SERS. Several questions need to be answered to determine the potential of this SOERS techniques for the future, such as elucidating the enhancement mechanism and determining suitable applications.

3.3.3 Improving time and spatial resolution

3.3.3.1 Time resolved Raman spectroscopy. Most commonly, Raman-SEC is performed using the so-called steady state method. In the steady state method, the electrode potential is

set to a desired value and given time to stabilize.^{219,220} After this stabilization period, a Raman spectrum is acquired. This steady state method has the benefit of being highly reproducible in obtaining data. However, information is lost when operating in this steady state configuration, such as information on reactive intermediates. Several experiments have been done in the 1990s regarding time resolved Raman spectroscopy.^{221–226} Van den Beld *et al.* have recently used an in-house developed graphene spectroelectrochemical cell to study the adsorption behaviour of simple redox couples in a more dynamic fashion, *i.e.* acquiring real-time Raman spectra while recording cyclic voltammograms at a scan rate of 25 mV s^{−1}.¹⁹⁰ Also recently, Zong *et al.*²²⁰ published on a transient SERS-SEC instrument that can acquire data at millisecond time resolution without the need to repetitively cycle the data acquisition. Further improvements in the field of time-resolved Raman-SEC might lead to a better understanding of reaction intermediates at the electrode surface.

3.3.3.2 Spatial resolution. One method in Raman spectroscopy that can be used in order to improve the spatial resolution is tip-enhanced Raman spectroscopy (TERS). First proposed by Wessel²²⁷ in 1985 and first independently developed by Stöckle *et al.*,²²⁸ Anderson,²²⁹ Hayazawa *et al.*²³⁰ and Pettinger²³¹ in 2000. In short, a metal or metal-coated tip is illuminated with a laser. The metal tip acts as a substrate for surface enhancement and can therefore be used for sensing purposes. In general, TERS achieves an enhancement factor of about 10³ to 10⁶, and has a spatial resolution between 10–80 nm,²³² much smaller than the diffraction limit of light. The combination of SEC and TERS is a relatively new field, first introduced in 2015 by Kurouski *et al.*²³³ and Zeng *et al.*²³⁴ As an emerging field, we expect that TERS-SEC will find more interest in the future.

3.3.4 Real world applications. As shown in paragraph 3.2.2, most of the research done with Raman-SEC is on fundamental research for material sciences. Recently, a paper was published on compact Raman-SEC for a portable point-of-care device,¹³⁶ as well as a paper on a low-cost portable device for the study of carbon nanotubes.¹⁸² These advancements might make Raman-SEC more accessible, possibly enabling a breakthrough towards more real-world applications.

3.3.5 Shell-isolated-enhanced-Raman-spectroscopy particles (SHINERS) and catalysis. Another new development related to SERS are so called SHINERS. SHINERS, developed by the group of Tian,²³⁵ are promising SERS particles coated with an inert layer for the use of catalyst monitoring. This both protects the typically Au nanoparticles from the high temperature and pressure conditions required for catalysis and at the same time prevents the Au to act as a catalyst, while the particles still exhibit the SERS effect.²³⁵ The group of Tian also performed notable work on combining SHINERS with electrochemistry. In the work Li *et al.*²³⁶ they combine SHINERS with electrochemical methods (EC-SHINERS) in order to monitor the pyridine absorption on single crystal Au surfaces *in situ*. The structured nanowires mentioned before (Fig. 13) could also be made into SHINERS-like structures by coating them



with Si. Although these SHINERS-like nanowires would no longer be suitable for use as electrodes, they could find applications in the field of *e.g.* elucidating photoelectrocatalytic reaction mechanisms.

4 Nuclear magnetic resonance spectroelectrochemistry (NMR-SEC)

4.1 Introduction NMR

Nuclear magnetic resonance (NMR) spectroscopy is a technique which is often used to obtain structural data about analytes. It typically operates within radio frequencies in the range of 60 to 1000 MHz (wavelengths of *ca.* 5 to 0.3 m). These low-energy waves can interact with nuclei that possess a magnetic spin, for example the isotopes ^1H and ^{13}C . For NMR spectroscopy, the different spin states of the nuclei get separated with a powerful magnetic field. The surrounding atoms and functional groups in a molecule influence how strongly the outside magnetic field affects the target nucleus locally. As a consequence, NMR spectroscopy is able to obtain comprehensive structural information of the molecule.

The sample can be manipulated and analysed with radio frequency radiation generated with a transmitter inside the spectrometer and the NMR probe around the sample. Nowadays, there are many different pulse patterns being used for sample excitation under varying experimental conditions.

For more detailed information, the authors refer the reader to a recent review paper by Colnago *et al.*²³⁷ and the references therein, explaining the fundamentals and challenges of NMR spectroscopy.

4.2 State of the art of NMR-SEC

NMR spectroscopy is a versatile tool to identify molecules non-destructively and accurately. It is most commonly used for the analysis of organic compounds, although there are also many applications which involve metals or metalorganic compounds. This versatility makes NMR spectroscopy a good coupling partner to EC, which can *e.g.* struggle to attribute signals correctly in the presence of side reactions. By combining the two techniques, however, mechanistic and kinetic information can be obtained *in situ* accurately and in a much more flexible way than *e.g.* with UV/Vis or IR spectroscopy, which can be limited by the functional groups present in the analyte.

However, because of the strong dependency of NMR measurements on the electromagnetic environment of a nucleus and the involvement of a strong outside magnetic field, NMR-SEC is not trivial. Combining EC with NMR leads to interference between the electrical and the magnetic fields, often caused by the electrode configuration, which can worsen the signal to noise ratio and signal resolutions in general. Additionally, instrumental failure upon simultaneous use of both systems has been reported, making real-time *in situ* measurements challenging.²³⁸ Pioneering work on NMR-SEC was performed by Wieckowski and associates. Many of their reported approaches were based on transferring the reaction

products from an EC-cell into an NMR tube and then analysing them with an NMR spectrometer in an offline manner.^{239–249} Other works included dedicated SEC-NMR cells^{250,251} or placing an EC-cell into an NMR spectrometer.²⁵²

A simple way to avoid possible interference between the two methods is to use an online technique which separates the electrochemical and the NMR-spectroscopic steps. However, this leads to much longer response times than is necessary to observe electrochemical intermediates and short-lived reaction products and is difficult to automate. To our knowledge, online EC-NMR methods are rarely being reported.

In this review paper, we instead focus on the liquid sample *in situ* EC-NMR methods reported in recent years and briefly mention their respective applications. For information on NMR applications in battery research, Hu *et al.* have recently published a review paper.²⁵⁴

4.2.1 Three-electrode cells as NMR tube insert. Because of the small confines imposed by standard NMR spectrometers, most of the recently reported *in situ* NMR-SEC approaches for liquid samples rely on integrating a homemade EC cell into a standard NMR test tube. An integral part of this kind of setup is the use of chokes to reduce the encountered interference between the NMR spectrometer and the potentiostat, which was specifically characterized in some recent publications.^{255,256}

4.2.1.1 Carbon fibre electrodes. Recent publications using carbon fibre WEs are based on a design originally published by Klod *et al.* in 2009.²⁵³ The design is shown in Fig. 14 and



Fig. 14 Schematic of a three-electrode NMR-SEC cell with carbon fibre WE and CE, partially placed in the NMR detection area. Reprinted with permission from Klod *et al.*²⁵³ Copyright 2009 American Chemical Society.



incorporated three electrodes. The WE and CE both consisted of carbon fibre filaments in sealed glass capillaries and the pseudo-RE was a PTFE-covered Ag wire with a AgCl tip. Only the active region of the WE was placed in the NMR detection area. The motivation to use carbon fibres was the higher possible reaction rate to obtain enough electrolysis product for ^1H and ^{13}C NMR measurements in lower amounts of time. Recent applications of this cell included the analysis of *p*-benzoquinone,^{253,257} 2-diphenylaminothiophene based compounds²⁵⁸ and a perfluoroalkylated fullerene.²⁵⁹ In the last case, ^{19}F NMR spectroscopy was performed as opposed to the more common ^1H and ^{13}C experiments. The measurements in all of the applications just mentioned were performed over large timeframes of up to 17 hours of electrolysis with periodic NMR spectroscopy sampling. Nunes *et al.* used a very similar configuration with the RE placed above the NMR detection area in a 10 mm NMR tube for the reduction of 9-chloroanthracene.²³⁸ They used a steady-state free precession (SSFP) pulse sequence to obtain a strong enough signal for ^{13}C experiments in the span of 11 minutes while halting the 150 minutes long electrolysis process during the measurement. Huang *et al.* used a different variation of the three-electrode design in which the WE carbon fibres just above the NMR detection area were loaded with a commercial electrocatalyst and a Pt gauze served as CE.²⁶⁰ They reported successful ^{13}C measurements of the electrocatalytic oxidation of ^{13}C -labeled ethanol by averaging 128 scans.

In 2013, Bussy *et al.* reported a slightly different three-electrode configuration in which carbon microfibres were again used as WE and CE.²⁶¹ The RE was replaced with a Pd wire. In this iteration, shown in Fig. 15, the electrodes were interfaced with the glass capillaries slightly differently and placed in different positions. The CE was still placed beneath the NMR detection area while the WE and the pseudo-RE were placed above it. Using this design, the oxidation of phenacetin was monitored periodically over the span of 6 hours.²⁶¹ Another application of this NMR-SEC cell was reported by Boisseau *et al.* when they performed ultrafast 2D COSY experiments on 9-chloroanthracene.²⁶² Periodic NMR samples were taken every 3 minutes in a chronoamperometric experiment over a time-frame of 80 minutes.

While the previously described cell designs focused on placing the WE close to or in the NMR detection area Ferreira Gomes *et al.* reported an NMR-SEC cell with three-electrode setup where all electrodes were deliberately placed above the NMR detection area. They used a carbon fibre WE, a platinized Pt CE, and a Ag/AgCl pseudo-RE.²⁶³ By moving the electrodes outside of the detection area, Ferreira Gomes *et al.* investigated the influence of the interference between the magnetic and the electric fields on mass transport within the cell. They reported that, by exploiting the resulting magnetoelectrolysis effect,^{264–266} which causes the solution to be stirred by a magnetohydrodynamic force, the electrochemical reaction rate for the electrochemical reduction of *p*-benzoquinone was greatly increased. The reaction was monitored *via* ^1H NMR spectroscopy.

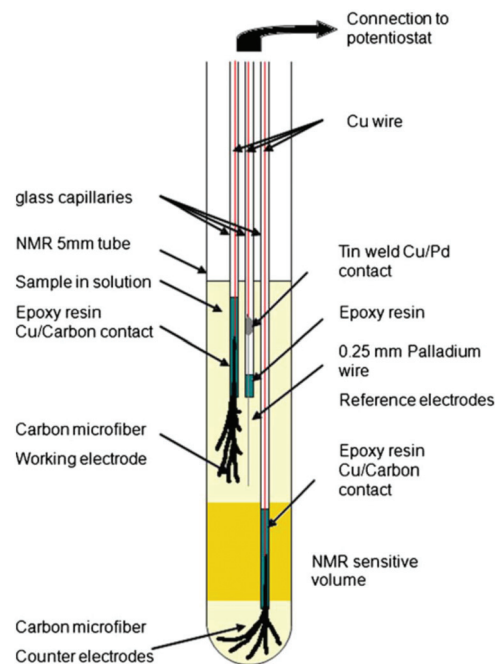


Fig. 15 Schematic of a three-electrode NMR-SEC cell with carbon microfibre WE and CE placed above and below the NMR detection area. Reprinted by permission from Springer Nature: from the original work by Bussy *et al.*,²⁶¹ Copyright 2012.

4.2.1.2 Thin film electrodes. While carbon microfibre approaches could aim to improve the electrochemical conversion inside the NMR-SEC cell, thin films with large surface areas hold the advantage of being near transparent to radio frequency radiation if they are thinner than their respective skin depth.²⁶⁷ This makes it possible to move the WE and possibly CE further into the NMR detection area to better monitor the electrochemical reaction close to the electrodes. One of the older approaches to introduce electrodes into an NMR tube based on a metal film was published by Zhang and Zwanziger in 2011.²⁵⁶ They used a thin Au film deposited on a 3 mm glass tube with small bored holes for ion exchange that served as WE and extended into the NMR detection area. A pseudo-RE was positioned inside of the WE glass tube and a curved platinized Pt-foil CE was placed between the NMR tube wall and the WE. With this metal film-based setup, Zhang and Zwanziger investigated the spectroelectrochemistry of *p*-benzoquinone, caffeic acid and 9-chloroanthracene in mechanistic approaches and with sampling times of up to two minutes. Cao *et al.* reported a similar NMR-SEC cell in which a Au coated glass tube sealed with a Nafion membrane to allow ion-exchange was used.²⁶⁸ Additionally, a Pt wire CE was placed inside the WE tube and a Pt foil pseudo-RE was placed between the NMR tube wall and the WE tube. Only the WE reached into the NMR detection area. They performed real-time ^1H NMR-SEC *in situ* measurements on quinone.

In more recent publications, Sorte *et al.* constructed an NMR-SEC cell with an interdigitated Au WE and CE (IGEs) in the NMR detection area and a Ag/AgCl pseudo-RE above





Fig. 16 Schematic and photograph of an NMR-SEC cell with interdigitated WE and CE placed in the NMR detection area. Reprinted from the original work by Sorte *et al.*,²⁶⁹ Copyright 2016, with permission from Elsevier.

it.^{269,270} The design is shown schematically and as a photograph in Fig. 16. The IGEs were printed on polyimide films and could be brought into close proximity inside a 5 mm NMR tube to promote better electrochemical performance. Sorte *et al.* successfully used this setup for *in situ* NMR-SEC measurements of the electrochemistry of ferrocene (^1H)²⁶⁹ and of the oxidation of methanol and ethanol (^{13}C).^{269,270}

Different modified tin oxide glass based coatings were explored by Zhang *et al.*, Wang *et al.* and Cao *et al.*^{271–274} Zhang *et al.* created a nano-polyaniline film (PAN) *via* electropolymerization on an indium tin oxide (ITO) electrode.²⁷¹ The schematic and a photograph are shown in Fig. 17. This PAN WE was then assembled with a Pt wire CE and a Ag pseudo-RE inside of a 5 mm NMR tube where only the WE extended into the NMR detection area. Using this simple NMR-SEC, they conducted real-time ^1H NMR-SEC experiments with hydroquinone. In 2019, Wang *et al.* used the same cell design to characterize the electrocatalytic performance of composite catalysts consisting of varying combinations of Pt, carbon, MoS_2 and graphene nanosheets electrodeposited on ITO-based WEs.^{272,273} They investigated the oxidation of ethanol with real time ^1H NMR-SEC measurements, among other techniques.

Lastly, Cao *et al.* used a slightly different substrate with a similar cell design.²⁷⁴ They employed two fluorinated tin oxide glass (FTO) slices positioned back to back to create a two sided WE and used a Pd wire as pseudo-RE. After synthesizing Pt nanocrystals on the WE surface, they characterized the electrocatalytic performance of the WE to oxidize short-chain alcohols with real-time ^1H NMR-SEC measurements.

4.2.1.3 Metal coil electrodes. A compromise between good NMR-SEC performance and a simple fabrication process was reported by da Silva *et al.* in 2019.²⁷⁵ They focused on developing a clutter-free NMR-SEC cell with simple materials and pro-



Fig. 17 Schematic and photograph of an ITO/PAN WE based three-electrode NMR-SEC cell. Reprinted from the original work by Zhang *et al.*,²⁷¹ Copyright 2018, with permission from Elsevier.

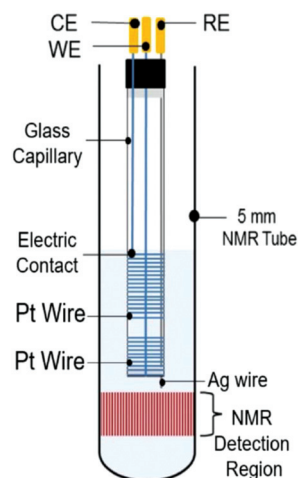


Fig. 18 Schematic of a coil-based three-electrode NMR-SEC cell. Reprinted from the original work by da Silva *et al.*,²⁷⁵ Copyright 2019, with permission from Elsevier.

vided an instructional video on how to assemble such a cell. The design schematic is shown in Fig. 18 and comprises two Pt wire coils around a glass capillary as WE and CE, as well as a Ag wire inside the capillary as pseudo-RE. All components were located above the NMR detection area. Da Silva *et al.* reported using the magnetoelectrolysis effect to stir the solution in a real-time ^1H NMR-SEC experiment on the oxidation of ascorbic acid, finding a doubled reaction rate in comparison to pure EC experiments.

4.2.2 Sealed pouch cell. Aside from the previously described approaches that focused on using standard NMR tubes as carriers, there have also been advances for different electrochemical reaction monitoring liquid *in situ* cells. One of



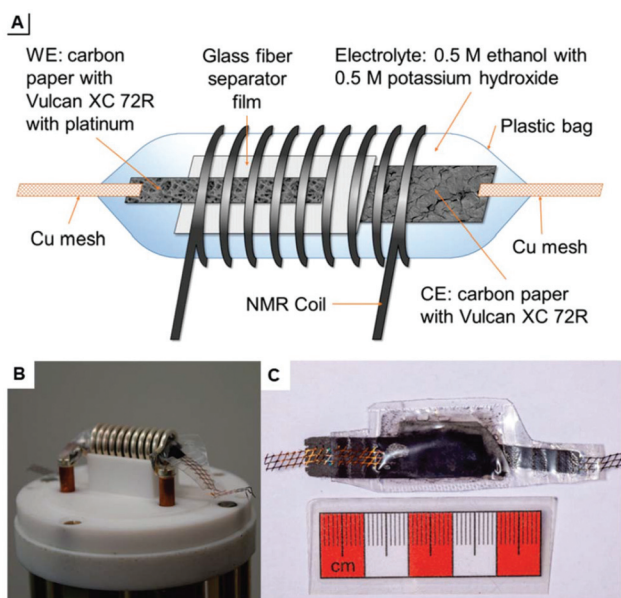


Fig. 19 (A) schematic of a plastic pouch NMR-SEC cell and (B and C) photographs thereof. Reprinted from the original work by Richter *et al.*,²⁷⁶ Copyright 2019, with permission from Creative Commons. Licensed under CC BY 3.0.

those was reported in 2019 by Richter *et al.*²⁷⁶ who assembled electrocatalyst-coated carbon paper electrodes separated by a glass fibre film into a sealed plastic pouch cell, based on a pouch design for capacitor characterization by Wang *et al.*²⁷⁷

The design schematic as well as photographs inside the solid state NMR spectrometer are shown in Fig. 19. With this cell and a solid state instrument, Richter *et al.* quantitatively monitored the oxidation of ethanol in real-time ^{13}C NMR-SEC *in situ* experiments with a time-resolution in the order of 10 minutes and during a total electrolysis time of 100 hours. The cell had to be punctured due to the evolution of CO_2 as reaction product.

4.3 Future perspective

The integration of the EC cell into a standard NMR tube is a simple and affordable approach. Since it is always done manually, however, it may be difficult to reproduce a cell exactly, even if an instructional video is provided.²⁷⁵ Creating a standardized electrochemical cell insert for NMR tubes could make NMR-SEC more accessible and allow for more comparable research between different research groups and experimental setups. On the other hand, creating more specialized equipment might lead to instrumental improvements in further avoiding negative interference between the two systems and making NMR-SEC methods more robust for real applications. An example of the influence of the instrumentation has very recently been given by Ferreira Gomes *et al.* in a review about the application of low magnetic field strengths for NMR-SEC.²⁷⁸

Especially ^{13}C NMR-SEC currently suffers from poor time-resolution. Underlying causes are low electrochemical reaction rates and the low natural occurrence of the isotope, often

leading to both very long electrolysis times of tens of hours and long measurement times of tens of minutes to achieve spectra of good enough quality.

There are general NMR spectroscopy advancements such as extra sample preparation steps or specific pulse sequences that can help to improve the sensitivity and reduce the timeframe of experiments. Specific pulse techniques like *e.g.* SSFP are already being investigated for applications in NMR-SEC,²³⁸ but, to our knowledge, sample preparation techniques are still not being investigated. One example is dynamic nuclear polarization (DNP),^{6,238,254} where a solidified sample can be polarized much more strongly in the solid state and then get quickly dissolved and analysed while retaining this hyperpolarization, leading to improved signal intensities.

A different approach to increase the reaction yield and reduce the time necessary for ^{13}C measurements could rely on the previously introduced NMR-SEC exclusive magnetoelectrolysis effect.^{263–266} Many cell designs rely on diffusion to move the reaction products into the NMR detection area. By investigating the exact influence of cell and electrode geometries on the stirring induced by the magnetoelectrolysis effect and optimizing them, the sensitivity limitations imposed by electrochemical reaction rates and product diffusion could be greatly alleviated. One approach to do this in the three-electrode cell type could for example focus on micromachining electrodes. By designing them in such a way that they induce defined non-uniformities in the magnetic field, the magnetoelectrolysis effect could be further investigated on a more fundamental level. Additionally, the possibilities of high-surface area electrodes to improve the conversion speed should be further investigated, since the larger currents can lead to more product but also to complications in the NMR-SEC setups.

To conclude, there are many promising areas for NMR-SEC developments to take place in the near future as it still is very much a developing technique with great potential.

5 Mass spectroelectrochemistry (EC-MS)

5.1 Introduction EC-MS

Mass spectrometry (MS) is a powerful and versatile technique to identify and quantify various kinds of analytes almost universally. The general MS process consists of the ionization of the analyte followed by the mass selection of the ions for their mass to charge ratio m/z and finally, the detection of the ions in a universal ion detector. There are many ionization and mass selector techniques with their respective advantages and drawbacks, varying for example in the ionization success for different kinds of molecules and the mass resolution that is achievable. Additionally, tandem mass spectrometry is the intentional fragmentation of the ions, for example *via* collision with an inert gas. The fragments can then be mass selected and detected or fragmented again for more complex experiments. More detailed information about mass spectrometry can be found in the literature.^{279,280}



While MS is not generally regarded as a spectroscopic method,²⁸¹ its universal capabilities to generate qualitative as well as quantitative data make it an excellent method to pair with EC. However, three precautions need to be kept in mind: firstly, the analyte needs to be successfully ionized. Secondly, MS requires a high-vacuum environment for the mass selector. And thirdly, MS generally uses a destructive detection method, making the analyte unavailable for follow-up procedures. Because of these precautions, *in situ* MS measurements are not possible in the same way as with, for example, the previously described IR spectroscopy. This also means that almost no surface-related data can be obtained. However, MS is an excellent method to quantify and identify reaction products and intermediates in online approaches. The qualitative information about reaction products and their structure gained complements the electrochemical information that can be obtained, especially for more complex reactions.

In the following sections, we will present often used as well as novel and innovative experimental setups used for EC-MS measurements as well as discuss future prospects. We focus on coupling mechanisms for liquid samples and not on the electrochemical and mass spectrometric instrumentation.

5.2 State of the art of EC-MS

Liquid EC-MS can only be realized as an online method. Since the lifetime of intermediates and unstable products is limited by definition, the method needs to have a low response time between these two steps to be able to detect them. In the following sections, we present a selection of recently reported online EC-MS methods to achieve low response times and successful intermediate detection. Methods with coupled chromatography steps will be excluded since they strictly raise the response time between product generation and detection. In research questions, where no intermediates need to be detected, online EC inductively coupled plasma (ICP) MS can be used, since it destroys species information. In 2019, Kasian *et al.*²⁸² reviewed EC-ICP-MS as a technique for catalyst research in an online configuration with reported response times 30–40 s.^{283,284}

5.2.1 Coupled electrospray ionization (ESI) techniques. A very useful ionization technique for liquid samples is electrospray ionization (ESI). For this technique, the analyte solution is introduced into a capillary-like device which is electrically connected to the MS inlet interface. By applying a high potential of several kV, a Taylor cone can be created that emits charged droplets towards the MS inlet. These charged droplets disintegrate into smaller and smaller droplets to finally form ionized molecules.²⁸⁵ The high potential necessary for this process can be dangerous to the equipment and the user. Therefore, special care has to be taken to decouple the EC reaction from the ionization. In combined systems, the ESI potential can be applied to the EC system as a floating potential.²⁸⁶

5.2.1.1 Commercial thin-layer EC-MS flow cells. As has been mentioned previously,^{11,12} thin-layer cells and microfluidic devices can hold several advantages over bigger cell geometries for better electrochemical performance. Over the last decade,

there have been many applications of commercial thin-layer flow cells for EC-MS, often but not always coupled to liquid chromatography (LC). In a review from 2014, Faber *et al.* covered the principal designs and flow charts of these setups,²⁸⁸ focusing on applications in metabolism studies. In a more recent publication, Zabel and Weber reported the use of a thin-layer flow cell setup with a response time of ~25 s for the study of sulphuric amino acid and glutathione oxidation.²⁸⁹ As using thin-layer flow cells is a very established and commercialized approach, the authors do not go into further detail here.

5.2.1.2 Microfluidic EC-MS flow cells. As described previously, microfluidic EC methods can hold many advantages over methods with larger reactors. Since EC-MS methods are online techniques, a microfluidic EC cell generally does not need to be specifically designed for coupling to MS.

In 2012, Odijk *et al.*²⁹⁰ reported an EC chip coupled to a microchip-ESI, which was previously developed by Qiao *et al.*²⁹¹ They investigated the oxidation of mitoxantrone as well as the influence of frit channels on the EC performance of the chip. As a special precaution to separate the electrical circuits, a grounded transfer tubing with high resistance was used to transfer the EC products to the microchip-ESI. Using the same chip design as Odijk *et al.*, van den Brink *et al.* integrated a commercially available ESI emitter to couple the chip to an MS system²⁸⁷ (Fig. 20). Van den Brink had less issues with circuit separation, as the ESI needle itself was grounded while the orifice of the MS was set to high voltage. Using this latter setup, response times of ~4.5 s were achieved.

5.2.1.3 EC-MS droplet in front of MS-interface. A coupling method with a very simple configuration was employed by Yu *et al.* in 2018,²⁹² as shown in Fig. 21. They used a droplet-approach in which a microscopy glass cover slide served as a carrier. This setup was developed from a previously reported pure MS approach with a glass carrier from Jiang *et al.*²⁹³



Fig. 20 Photographs of a microfluidic EC cell integrated with a commercially available ESI needle in an EC-MS setup. Reprinted with permission from van den Brink *et al.*²⁸⁷ Copyright 2015 American Chemical Society.



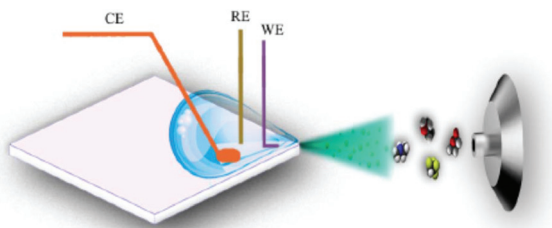


Fig. 21 D Schematic of the droplet-based EC-MS approach. Reprinted with permission from Yu *et al.*²⁹² Copyright 2018 American Chemical Society.



Fig. 22 Schematic of the thin-layer double-barrel capillary setup. Reprinted from the original work by Qiu *et al.*,²⁹⁴ Copyright 2016, with permission from Creative Commons. Licensed under CC BY 3.0.

The glass carrier was operated with a sample volume of 80 μL forming a droplet in one corner. In that corner, there was also a three-electrode setup consisting of a Pt WE and CE as well as a Ag/AgCl RE. In addition to the electrochemical reaction, the WE and CE were also used to form the electrospray together with the MS inlet. The authors do not specify a response time between the reaction and subsequent detection, but demonstrate real-time behaviour of the setup system to obtain mechanistic EC reaction data.

As a droplet-based approach, this setup is very small and simple to use. A disadvantage is the non-continuity of the experiment. The droplet is gradually desaturated of the analyte and saturated with reaction products over the course of the experiment which limits the versatility of this method. Additionally, the electrospray removes more and more liquid, slightly changing the entire system over time. The exact positioning and amount of the liquid in the droplet is also difficult to exactly reproduce. As a consequence, this setup is well suited for qualitative mechanistic studies but not for quantitative investigations.

5.2.1.4 Thin liquid layer double-barrel capillary. Qiu *et al.*²⁹⁴ and later Guo *et al.*²⁹⁵ reported a homemade capillary device where the EC reaction takes place inside a thin liquid layer formed just outside the tip of a hydrophilic double-barrel capillary. One of the two barrels is capped with previously *in situ* generated carbon while the other barrel contains the analyte solution as well as a AgCl/Ag wire. Primary ions for the formation of an electrospray are generated with a piezoelectric pistol at the beginning of the capillary.

By positioning the EC reaction at the tip outside of the capillary at a distance of 2 mm to the MS inlet, the response time is reduced greatly. While the authors don't give an exact value, they succeeded in detecting a diamine intermediate formed by the oxidation of uric acid with a reported half-life of about 23 ms (ref. 296) which makes this method an excellent online EC-MS technique with a very low response time. Later, Guo *et al.* applied this technique to detect intermediates in organometallic Ru- and Ir-based chemiluminescence experiments.

Qiu *et al.*²⁹⁴ have created a powerful online EC-MS method with a small footprint, as shown in Fig. 22. Their setup needs careful preparation to form the carbon electrode, but shows an

excellent response time. The ionization process is unusual, however, and probably the most variable part between different experiments. In addition to this drawback, there is no RE, further complicating the use of this setup. With careful calibration, however, this method can serve as a great tool to investigate short-lived reaction products and intermediates.

5.2.2 Differential electrochemical mass spectrometry (DEMS). A different approach to ionize quickly transport reaction products is the differential electrochemical mass spectrometry (DEMS) method, where volatile compounds inside a liquid evaporate through a membrane or porous frit into the MS. This technique is inherently restricted by the vapor pressure of the analyte and was first mentioned as a dedicated technique by Wolter and Heitbaum in 1984.^{298,299} The main component of DEMS is a non-wetting porous membrane that allows volatile compounds to evaporate into a two-step vacuum as a function of their formation rate, hence the name "differential".³⁰⁰

5.2.2.1 Thin-layer DEMS cells. Modern DEMS systems, as described by Abd-El-Latif *et al.*,³⁰⁰ are generally built in a thin-layer configuration, where an electrode is placed on one side and a porous membrane on the other side. The electrochemical reaction products have to cross from the electrode to the other side of the cell to pass the membrane, causing response times of up to 2 s.

One example of a thin-layer cell still in use at present with modifications was published by Jusys *et al.* in 1999, and is depicted in Fig. 23.²⁹⁷ It was originally designed for a combination of DEMS and an electrochemical quartz crystal microbalance (EQCM) in two separate chambers. The membrane used in this instance is a porous Teflon membrane, placed on top of a stainless steel frit that leads to an MS inlet. In 2007, Heinen *et al.* slightly changed the cell to include a Pt thin-film layer on a Si prism instead of an EQCM³⁰² to allow a combined EC-IR-MS method. These two designs have been used in multiple recent publications with applications ranging from CO oxidation and desorption on nanocrystals,³⁰³ ethylene glycol oxidation,^{304,305} methanol oxidation³⁰⁶ to the oxygen evolution reaction.³⁰⁷ The reported response time of the setups lies in the range of 1–2 s.^{297,304}

A simpler one chamber thin-layer cell was reported by Trimarco *et al.* in 2018.³⁰¹ Here, they used a previously reported microfabricated Si membrane with supporting SiO_2





Fig. 23 DEMS cell; A: exploded sideview; B: top view. Image republished and modified with permission of Journal of The Electrochemical Society, from the original work by Jusys *et al.*²⁹⁷ Copyright 1999, permission conveyed through Copyright Clearance Center, Inc.

pillars and an integrated sampling chamber to separate the liquid phase from the gaseous phase.³⁰⁸ The thin-layer cell had a simple one chamber geometry with a WE directly above the membrane chip, allowing for low response times. A CE and RE were positioned in slanted side channels. The membrane chip itself was mounted onto a vacuum system to enable DEMS, shown schematically in Fig. 24. Using this setup, Roy *et al.* investigated the oxidation evolution reaction catalysed by nanoparticle catalysts.³⁰⁹ Additionally, Winiwarter *et al.* investigated the mechanisms of partial propene oxidation.³¹⁰



Fig. 24 Membrane DEMS setup for thin-layer cell; (a) membrane chip photograph; (b) scanning electron microscope image of the membrane chip; (c) schematic of the membrane chip; (d) schematic of the setup. Image republished and modified with permission from Trimarco *et al.*³⁰¹ Copyright 2018 American Chemical Society.



Fig. 25 Membrane fabrication scheme. Reprinted from the original work by Trimarco *et al.*,³⁰⁸ Copyright 2015, with the permission of AIP Publishing.

5.2.2.2 Microfluidic DEMS. In a similar fashion to the ESI method, a microfluidic approach for DEMS has recently been reported. In 2015, Trimarco *et al.* reported a “sniffer chip” assembly that uses the previously mentioned microfabricated membrane.³⁰⁸ The microfabrication process is illustrated in Fig. 25. The Si and SiO₂ based membrane was incorporated into a microfluidic chip with integrated electrodes a small total internal volume of *ca.* 31 nL and a low response time about 0.5 s. Trimarco *et al.* made proof of principle measurements of hydrogen and oxygen evolution reactions in an aqueous solvent. An envisioned application is to use the chip as a dip-in probe that immediately saturates with the gasses in the solution to provide a quick qualitative readout of the sample.

5.2.3 Microfluidic EC-SIMS (secondary ion mass spectrometry). A different ionization approach based on a microfluidic chip was recently reported by Liu *et al.*,³¹¹ Wang *et al.*³¹² and Yu *et al.*³¹³ They were using microfluidic EC cells with a silicon nitride (SiN) membrane on the backside of a thin Au WE, as shown in Fig. 26. In SIMS, a primary ion beam is used to ionize the analyte molecules as secondary ions. In this device, the primary ion beam was used to drill a hole through the SiN membrane and the WE, exposing the liquid to the vacuum of the mass spectrometer in a $\sim 2 \mu\text{m}$ wide spot. For sequential mechanistic measurements, new holes could be drilled to capture the current state close to the electrode in the instant the hole is created. All of the publications cited above focused on imaging the electrode/electrolyte interface, by using KI^{311,313} or ascorbic acid³¹² as analytes with a response

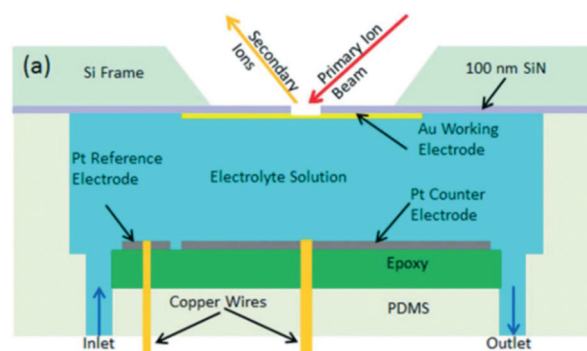


Fig. 26 Schematic of an EC-SIMS cell. Reprinted from the original work by Liu *et al.*,³¹¹ Copyright 2014 – Published by The Royal Society of Chemistry.



Table 2 Overview of the described EC–MS techniques and their special characteristics

Ionization method/ technique	Response time	Advantages	Drawbacks	Ref.
ESI, thin-layer	~25 s	Standardized, exchangeable electrode	Long transfer times, special care in gasket handling	289
ESI, microfluidic	~4.5 s	Optimized design, very low analyte amounts	Limited flexibility, difficult fabrication	287 and 290
ESI, droplet	fast enough to detect intermediates	Very small footprint, simple fabrication	Static experiments, poor reproducibility	292
ESI, double-barrel capillary	≤23 ms	Extremely fast, small footprint	Complicated fabrication and operation	294 and 295
DEMS, thin-layer	1–2 s	Simple operation, measures formation rate	Long transfer times, restricted to volatile analytes	297, 301–307, 309 and 310
DEMS, microfluidic	~0.5 s	Simple and fast operation, very small footprint	Restricted to volatile analytes, difficult fabrication	308
SIMS, microfluidic	<100 ms	Very fast and sensitive, can probe electrode/electrolyte interfaces	Small probed area, destroys the WE	311–313

time of under 100 ms and imaging integration times of 1–2 minutes. Wang *et al.* also succeeded in detecting ascorbic acid oxidation intermediates.

5.3 Future perspective

We have shown several established and innovative approaches to couple electrochemical devices to mass spectrometers. Table 2 shows the different techniques and semi-quantitative statements about their reported response times, advantages and restrictions.

Compared to other spectroelectrochemical methods, however, EC-MS is not being researched to the same degree. As stated previously, the most important part in coupling these techniques is the ionization technique.

The already established microfluidic ESI methods described previously mostly use commercially available nano-ESI needles. A simple way to improve on the methods could lie in the monolithic fabrication of the device. By integrating the needle into the chip design, it could be simpler to setup, avoiding damage to the nano-ESI needle and possible leaks. More importantly, it reduces dead volume. Additionally, the influence of the needle geometry on the ionization success could be investigated more systematically. This has been demonstrated in a relatively straightforward manner by the group of Girault,^{314,315} using a single carbon paste electrode. To the best of our knowledge it has not been demonstrated with a three-electrode microchip yet.

Additionally, there have been ESI setups where the capillary itself served as an electrochemical reactor in the past.^{286,316} This approach promises the lowest possible response times and simplest setups and should be investigated further even in present times.

The microfluidic possibilities of using EC-DEMS chips to monitor EC reactions are still only emerging. A spin-off company, SpectroInlets, has created a dedicated system to use their membrane chip technology. It is to be expected that microfluidic DEMS will be a topic of future research.

Lastly, as a general note, there are limitations to the use of ESI and DEMS. While DEMS is strongly restricted to volatile

analytes, ESI can struggle with the ionization of certain classes of compounds and ion suppression effects.²⁷⁹ It is therefore also important to investigate other already established ionization methods as a coupling partner to EC. We presented an example that employed SIMS to obtain impressive information about electrode/electrolyte interfaces and could also take short snapshots of intermediates. There are, however, more established and likely also suitable techniques like *e.g.* atmospheric pressure chemical ionization (APCI), atmospheric pressure photoionization (APPI), atmospheric pressure plasma jet (APPI) or ICP ionization, the latter having been already mentioned in section 5.2 for methods where species information is not important. These methods mostly use external gasses to nebulize and/or ionize the analytes and thus require additional instrumentation outside the EC cell. They can, however, offer better ionization efficiencies or better versatility than other ionization methods that require less instrumentation because they use different mechanisms to create the ions necessary for MS.

6 Concluding remarks

6.1 IR-SEC

We expect that the advent of QCL lasers will change the future of IR-SEC, solving the inherent problems of low intensity light sources. The QCL lasers will make laser IR spectroscopy more accessible, allowing for faster analysis times. The higher source power will alleviate the need for using the thin-layer configurations and at the same time push the field of ATR IR-SEC towards the use of waveguides. This, together with advancements in nanofabrication will create the possibility of constructing complete IR-SEC lab on chip systems.

By making use of IR-transparent conductive waveguides, both the waveguide and the electrode can be combined into one structure. The footprint of the device could be made small enough to be installed as sensing systems in organs on chip, as wearable sensors or in miniaturized high throughput screening platforms.



6.2 Raman-SEC

Unlike IR-SEC, Raman-SEC is already a mature technique. However, especially the field of SERS-SEC can be further developed by combining it with new advancements in the field of nanofabrication. These advancements will possibly lead to developments where SERS enhancement structures can give more reliable and uniform enhancements and at the same time increase the area of the enhancement structures and simultaneously decrease the cost.

At the moment Raman-SEC is mostly used for a large number of applications in material sciences, but as of yet does not have a large number of real world applications, some notable exceptions are for instance the work by the group of Brosseau.^{136–139} When the advancements of SERS-SEC as highlighted above take place, the technique will likely become more attractive for real world applications.

6.3 NMR-SEC

NMR spectroscopy is a very versatile technique that still has much potential to grow and mature. As a consequence, NMR-SEC is also still in its infancy and will benefit from general NMR improvements. There are, however, certain NMR-SEC specific recommendations that can be made.

Because of the unique restrictions caused by the interference between NMR and EC systems, dedicated NMR-SEC instruments should be investigated. Also, standardizing or making the often used three-electrode cell type commercially available would be beneficial to make research from different groups both more comparable and reliable.

NMR-SEC can already be used for real-time ¹H measurements, but especially real-time ¹³C measuring methods still need to be improved. Research approaches could be based on sample preparation techniques or the exploitation of NMR-SEC specific effects like magnetoelectrolysis to reduce the time needed for an NMR-SEC experiment.

6.4 EC-MS

Although research towards EC-MS is not being performed to the same degree as the SEC techniques covered in this review, clear paths for future advancements can be discussed. Any improvements on EC-MS regarding the response time between reactions at the electrodes and the ionization of the sample would make the technique more attractive. Directly combining the ionization mechanism, *e.g.* ESI, with a microfluidic system *via* new microfabrication techniques could result in monolithic devices with very short response times. Additional ionization techniques that are used for MS but could also be of interest for EC-MS are SIMS, APCI, APPI, APPJ and ICP, as partly already mentioned previously. These other ionization techniques could help to increase the ionization efficiency of certain analytes while also mostly not suffering from the same coupling problems of EC with MS as is the case with ESI. Some of them have their own application restrictions, however, making a careful selection all the more important.

6.5 Hyphenation

Hyphenation of the (already hyphenated) SEC techniques, also known as multi-spectroelectrochemistry, is quite common in literature. In 2011 Dunsch wrote a review paper on this very topic.³

As the field of SEC advances, so will the number of hyphenation of different techniques. Already in 2011 Dunsch predicted a greater variety in triple multi-spectroelectrochemistry. In order to get more information from a system, we expect that more and more techniques will be combined into multi-spectroelectrochemistry in the future.

Author contributions

Authors J. J. A. Lozeman and P. Führer contributed equally to this review paper. J. J. A. Lozeman was responsible for IR-SEC and Raman-SEC, P. Führer had the responsibility for NMR-SEC and EC-MS. M. Odijk and W. Olthuis supervised and managed the review and edited the final version of the paper.

Conflicts of interest

There are no conflicts to declare.

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References

- W. Kaim and J. Fiedler, *Chem. Soc. Rev.*, 2009, **38**, 3373–3382.
- T. Kuwana, R. K. Darlington and D. W. Leedy, *Anal. Chem.*, 1964, **36**, 2023–2025.
- L. Dunsch, *J. Solid State Electrochem.*, 2011, **15**, 1631–1646.
- H. Oberacher, F. Pitterl, R. Erb and S. Plattner, *Mass Spectrom. Rev.*, 2015, **34**, 64–92.
- A. J. Wain and M. A. O'connell, *Adv. Phys.: X*, 2017, **2**, 188–209.
- Y. Y. J. Tong, *Curr. Opin. Electrochem.*, 2017, **4**, 60–68.
- L. León and J. D. Mozo, *TrAC, Trends Anal. Chem.*, 2018, **102**, 147–169.
- Y. Zhai, Z. Zhu, S. Zhou, C. Zhu and S. Dong, *Nanoscale*, 2018, **10**, 3089–3111.



- 9 J. Garoz-Ruiz, J. V. Perales-Rondon, A. Heras and A. Colina, *Electroanalysis*, 2019, 1254–1278.
- 10 D. L. Pavia, G. M. Lampman, G. S. Kriz and J. A. Vyvyan, *Introduction to Spectroscopy*, Cengage Learning, 2008.
- 11 D. Moss, E. Nabadryk, J. Breton and W. Mantele, *Eur. J. Biochem.*, 1990, **187**, 565–572.
- 12 F. T. G. van den Brink, W. Olthuis, A. van den Berg and M. Odijk, *TrAC, Trends Anal. Chem.*, 2015, **70**, 40–49.
- 13 P. A. Christensen and S. W. M. Jones, *J. Phys. Chem. C*, 2014, **118**, 29760–29769.
- 14 K. H. K. L. Alwis, M. R. Mucalo and B. Ingham, *J. Electrochem. Soc.*, 2013, **160**, 803–812.
- 15 A. L. De Lacey, E. C. Hatchikian, A. Volbeda, M. Frey, J. C. Fontecilla-Camps and V. M. Fernandez, *J. Am. Chem. Soc.*, 1997, **119**, 7181–7189.
- 16 R. Guidelli, *Bioelectrochemistry of Biomembranes and Biomimetic Membranes*, 2016.
- 17 A. Hartstein, J. R. Kirtley and J. C. Tsang, *Phys. Rev. Lett.*, 1980, **45**, 201–204.
- 18 M. Osawa, in *Near-Field Optics and Surface Plasmon Polaritons*, ed. S. Kawata, Springer Berlin Heidelberg, Berlin, Heidelberg, 2001, pp. 163–187.
- 19 R. Aroca, *Surface-Enhanced Vibrational Spectroscopy*, John Wiley & Sons, Ltd, Chichester, UK, 2006.
- 20 Y. Hu, Á. I. López-Lorente and B. Mizaikoff, *ACS Photonics*, 2019, **6**, 2182–2197.
- 21 K. I. Ataka, T. Yotsuyanagi and M. Osawa, *J. Phys. Chem.*, 1996, **100**, 10664–10672.
- 22 M. Osawa, K. Ataka, K. Yoshii and T. Yotsuyanagi, *J. Electron Spectrosc. Relat. Phenom.*, 1993, **64–65**, 371–379.
- 23 H. Im, K. C. Bantz, N. C. Lindquist, C. L. Haynes and S. H. Oh, *Nano Lett.*, 2010, **10**, 2231–2236.
- 24 E. C. Le Ru and P. G. Etchegoin, *Principles of Surface-Enhanced Raman Spectroscopy*, Elsevier, 2009.
- 25 K. Ataka and J. Heberle, *Anal. Bioanal. Chem.*, 2007, **388**, 47–54.
- 26 R. Aroca, *Surface-Enhanced Vibrational Spectroscopy*, Wiley, 2006.
- 27 F. Neubrech, C. Huck, K. Weber, A. Pucci and H. Giessen, *Chem. Rev.*, 2017, **117**, 5110–5145.
- 28 C. Di Bari, N. Mano, S. Shleev, M. Pita and A. L. De Lacey, *J. Biol. Inorg. Chem.*, 2017, **22**, 1179–1186.
- 29 Y. Kato and T. Noguchi, *Biochemistry*, 2014, **53**, 4914–4923.
- 30 F. Tutunea and M. D. Ryan, *J. Electroanal. Chem.*, 2012, **670**, 16–22.
- 31 F. Tutunea, A. Atifi and M. D. Ryan, *J. Electroanal. Chem.*, 2015, **744**, 17–24.
- 32 R. W. Murray, W. R. Heineman and G. W. O'Dom, *Anal. Chem.*, 1967, **39**, 1666–1668.
- 33 Z. Wei and M. D. Ryan, *Inorg. Chem.*, 2010, **49**, 6948–6954.
- 34 Y. Dai, D. A. Proshlyakov, J. K. Zak and G. M. Swain, *Anal. Chem.*, 2007, **79**, 7526–7533.
- 35 J. K. Zak, J. E. Butler and G. M. Swain, *Anal. Chem.*, 2001, **73**, 908–914.
- 36 J. Stotter, J. Zak, Z. Behler, Y. Show and G. M. Swain, *Anal. Chem.*, 2002, **74**, 5924–5930.
- 37 K. H. K. L. Alwis, M. R. Mucalo and J. R. Lane, *RSC Adv.*, 2015, **5**, 31815–31825.
- 38 M. J. Shaw, R. L. Henson, S. E. Houk, J. W. Westhoff, M. W. Jones and G. B. Richter-Addo, *J. Electroanal. Chem.*, 2002, **534**, 47–53.
- 39 N. Xu, J. Lilly, D. R. Powell and G. B. Richter-Addo, *Organometallics*, 2012, **31**, 827–834.
- 40 M. A. El-Attar, N. Xu, D. Awasabisah, D. R. Powell and G. B. Richter-Addo, *Polyhedron*, 2012, **40**, 105–109.
- 41 D. Awasabisah, N. Xu, K. P. S. Gautam, D. R. Powell, M. J. Shaw and G. B. Richter-Addo, *Eur. J. Inorg. Chem.*, 2016, **2016**, 509–518.
- 42 S. Pons, *J. Electroanal. Chem.*, 1983, **150**, 495–504.
- 43 D. Aurbach and O. Chusid, *J. Electrochem. Soc.*, 1993, **140**, L155–L157.
- 44 M. R. Mucalo and Q. Li, *J. Colloid Interface Sci.*, 2004, **269**, 370–380.
- 45 X. Fang, L. Wang, P. K. Shen, G. Cui and C. Bianchini, *J. Power Sources*, 2010, **195**, 1375–1378.
- 46 L. Liu, L. Zeng, L. Wu and X. Jiang, *Anal. Chem.*, 2017, **89**, 2724–2730.
- 47 L. Liu, L. Zeng, L. Wu and X. Jiang, *J. Phys. Chem. C*, 2015, **119**, 3990–3999.
- 48 Y. Y. Yang, J. Ren, H. X. Zhang, Z. Y. Zhou, S. G. Sun and W. Bin Cai, *Langmuir*, 2013, **29**, 1709–1716.
- 49 L. Wang, H. Meng, P. K. Shen, C. Bianchini, F. Vizza and Z. Wei, *Phys. Chem. Chem. Phys.*, 2011, **13**, 2667–2673.
- 50 Q. Zhang, W. L. Song, A. M. Showkot Hossain, Z. Di Liu, G. J. Hu, Y. P. Tian, J. Y. Wu, B. K. Jin, H. P. Zhou, J. X. Yang and S. Y. Zhang, *Dalton Trans.*, 2011, **40**, 3510–3516.
- 51 W. G. Golden, D. S. Dunn and J. Overend, *J. Catal.*, 1981, **71**, 395–404.
- 52 P. W. Faguy and W. N. Richmond, *J. Electroanal. Chem.*, 1996, **410**, 109–113.
- 53 P. W. Faguy, W. N. Richmond, R. S. Jackson, S. C. Weibel, G. Ball and J. H. Payer, *Appl. Spectrosc.*, 1998, **52**, 557–564.
- 54 M. J. Green, B. J. Barner and R. M. Corn, *Rev. Sci. Instrum.*, 1991, **62**, 1426–1430.
- 55 B. J. Barner, M. J. Green, E. I. Sáez and R. M. Corn, *Anal. Chem.*, 1991, **63**, 55–60.
- 56 V. Zamylnny, I. Zawisza and J. Lipkowski, *Langmuir*, 2003, **19**, 132–145.
- 57 I. Zawisza, X. Bin and J. Lipkowski, *Langmuir*, 2007, **23**, 5180–5194.
- 58 N. Garcia-Araez, C. L. Brosseau, P. Rodriguez and J. Lipkowski, *Langmuir*, 2006, **22**, 10365–10371.
- 59 R. C. Alkire, D. M. Kolb, J. Lipkowski and P. N. Ross, *Diffraction and Spectroscopic Methods in Electrochemistry*, Wiley, 2009.
- 60 E. A. Monyoncho, V. Zamylnny, T. K. Woo and E. A. Baranova, *Analyst*, 2018, **143**, 2563–2573.
- 61 T. Wang, J. Bai, X. Jiang and G. U. Nienhaus, *ACS Nano*, 2012, **6**, 1251–1259.
- 62 P. Hosseini, G. Wittstock and I. Brand, *J. Electroanal. Chem.*, 2018, **812**, 199–206.



- 63 A. J. Bard, *Electroanalytical Chemistry: A Series of Advances*, Taylor & Francis, 1986.
- 64 G. Neophytides, L. Quaroni, F. N. Büchi, A. Orfanidi, S. G. Neophytides and T. J. Schmidt, *Electrochem. Commun.*, 2013, **34**, 200–203.
- 65 P. Su, V. Prabhakaran, G. E. Johnson and J. Laskin, *Anal. Chem.*, 2018, **90**, 10935–10942.
- 66 M. Villalba, L. P. M. De Leo and E. J. Calvo, *ChemElectroChem*, 2014, **1**, 195–199.
- 67 R. Aroca, D. Ross and C. Domingo, *Appl. Spectrosc.*, 2004, **58**, 324A–338A.
- 68 T. Schadle and B. Mizaikoff, *Appl. Spectrosc.*, 2016, **70**, 1625–1638.
- 69 M. Sieger and B. Mizaikoff, *Anal. Chem.*, 2016, **88**, 5562–5573.
- 70 B. K. Purushothaman, M. Pelsozy, P. W. Morrison, V. F. Lvovich and H. B. Martin, *J. Appl. Electrochem.*, 2012, **42**, 111–120.
- 71 P. Paengnakorn, P. A. Ash, S. Shaw, K. Danyal, T. Chen, D. R. Dean, L. C. Seefeldt and K. A. Vincent, *Chem. Sci.*, 2017, **8**, 1500–1505.
- 72 A. Hatta, Y. Suzuki, T. Wadayama and W. Suëtaka, *Appl. Surf. Sci.*, 1991, **48–49**, 222–226.
- 73 E. Kretschmann, *Z. Phys.*, 1971, **241**, 313–324.
- 74 M. F. Baruch, J. E. Pander, J. L. White and A. B. Bocarsly, *ACS Catal.*, 2015, **5**, 3148–3156.
- 75 R. Aroca and B. Price, *J. Phys. Chem. B*, 1997, **101**, 6537–6540.
- 76 A. J. Healy, P. A. Ash, O. Lenz and K. A. Vincent, *Phys. Chem. Chem. Phys.*, 2013, **15**, 7055–7059.
- 77 C. Dillard, A. Singh and V. Kalra, *J. Phys. Chem. C*, 2018, **122**, 18195–18203.
- 78 J. Heyes, M. Dunwell and B. Xu, *J. Phys. Chem. C*, 2016, **120**, 17334–17341.
- 79 M. Pfaffeneder-Kmen, I. F. Casas, A. Naghilou, G. Trettenhahn and W. Kautek, *Electrochim. Acta*, 2017, **255**, 160–167.
- 80 A. Viinikanoja, J. Kauppila, P. Damlin, M. Suominen and C. Kvarnström, *Phys. Chem. Chem. Phys.*, 2015, **17**, 12115–12123.
- 81 J. Izquierdo, B. Mizaikoff and C. Kranz, *Phys. Status Solidi A*, 2016, **213**, 2056–2062.
- 82 D. Neubauer, J. Scharpf, A. Pasquarelli, B. Mizaikoff and C. Kranz, *Analyst*, 2013, **138**, 6746–6752.
- 83 N. Menegazzo, M. Kahn, R. Berghauser, W. Waldhauser and B. Mizaikoff, *Analyst*, 2011, **136**, 1831–1839.
- 84 S. Sun, S. Hong, S. Chen, G. Lu, H. Dai and X. Xiao, *Sci. China, Ser. B: Chem.*, 1999, **42**, 261–267.
- 85 J. T. Li, S. R. Chen, F. S. Ke, G. Z. Wei, L. Huang and S. G. Sun, *J. Electroanal. Chem.*, 2010, **649**, 171–176.
- 86 J. T. Li, Q. S. Chen and S. G. Sun, *Electrochim. Acta*, 2007, **52**, 5725–5732.
- 87 H. Gong, S. G. Sun, Y. J. Chen and S. P. Chen, *J. Phys. Chem. B*, 2004, **108**, 11575–11584.
- 88 S.-G. Sun and Z.-Y. Zhou, in *In-situ Spectroscopic Studies of Adsorption at the Electrode and Electrocatalysis*, ed. S.-G. Sun, P. A. Christensen and A. Wieckowski, Elsevier Science B.V., Amsterdam, 2007, pp. 139–178.
- 89 C. A. Melendres, B. Beden and G. A. Bowmaker, *J. Electroanal. Chem.*, 1995, **383**, 191–193.
- 90 S. M. Rosendahl, F. Borondics, T. E. May, T. M. Pedersen and I. J. Burgess, *Anal. Chem.*, 2011, **83**, 3632–3639.
- 91 K. Tu, M. J. Lardner, T. A. Morhart, S. M. Rosendahl, S. Creighton and I. J. Burgess, *J. Phys. Chem. C*, 2016, **120**, 23640–23647.
- 92 S. M. Rosendahl, F. Borondics, T. E. May and I. J. Burgess, *Anal. Chem.*, 2013, **85**, 8722–8727.
- 93 M. J. Lardner, K. Tu, B. C. Barlow, S. M. Rosendahl, F. Borondics and I. J. Burgess, *J. Electroanal. Chem.*, 2017, **800**, 184–189.
- 94 P. A. Ash, H. A. Reeve, J. Quinson, R. Hidalgo, T. Zhu, I. J. McPherson, M. W. Chung, A. J. Healy, S. Nayak, T. H. Lonsdale, K. Wehbe, C. S. Kelley, M. D. Frogley, G. Cinque and K. A. Vincent, *Anal. Chem.*, 2016, **88**, 6666–6671.
- 95 J. A. Sigrist, E. S. Lins, T. A. Morhart, J. L. Briggs and I. J. Burgess, *Appl. Spectrosc.*, 2019, **73**, 1394–1402.
- 96 M. J. Weida and B. Yee, *Imaging Manip. Anal. Biomol. Cells Tissues IX*, 2011, **7902**, 79021C.
- 97 J. M. Ramirez, V. Vakarin, C. Gilles, J. Frigerio, A. Ballabio, P. Chaisakul, X. Le Roux, C. Alonso-Ramos, G. Maisons, L. Vivien, M. Carras, G. Isella and D. Marris-Morini, *Opt. Lett.*, 2017, **42**, 105.
- 98 P. Wägli, Y. C. Chang, A. Homsy, L. Hvozdar, H. P. Herzig and N. F. De Rooij, *Anal. Chem.*, 2013, **85**, 7558–7565.
- 99 P. G.-Q. Lo, A. A. Bettiol, A. E.-J. Lim, K.-W. Ang and U. Younis, *IEEE Photonics Technol. Lett.*, 2016, **28**, 2447–2450.
- 100 C. B. E. Gawith, A. Z. Khokhar, C. Littlejohns, G. S. Murugan, J. S. Penades, V. Mittal, M. Nedeljkovic, J. S. Wilkinson, G. Z. Mashanovich and L. G. Carpenter, *Opt. Express*, 2017, **25**, 27431.
- 101 N. Singh, A. Casas-Bedoya, D. D. Hudson, A. Read, E. Mägi and B. J. Eggleton, *Opt. Lett.*, 2016, **41**, 5776.
- 102 V. Mittal, J. S. Wilkinson and G. S. Murugan, *High-contrast GeTe₄ waveguides for mid-infrared biomedical sensing applications*, ed. J. E. Broquin and G. Nunzi Conti, International Society for Optics and Photonics, 2014, p. 89881A.
- 103 Z. Cheng, X. Chen, C. Y. Wong, K. Xu and H. K. Tsang, *IEEE Photonics J.*, 2012, **4**, 1510–1519.
- 104 Y. C. Chang, P. Wägli, V. Paeder, A. Homsy, L. Hvozdar, P. Van Der Wal, J. Di Francesco, N. F. De Rooij and H. Peter Herzig, *Lab Chip*, 2012, **12**, 3020–3023.
- 105 M. Malmström, M. Karlsson, P. Forsberg, Y. Cai, F. Nikolajeff and F. Laurell, *Opt. Mater. Express*, 2016, **6**, 1286.
- 106 A. Osman, A. Z. Khokhar, M. Nedeljkovic, Y. Wu, G. Z. Mashanovich, K. Debnath, Z. Qu and J. Soler Penades, *Opt. Lett.*, 2018, **43**, 5997.
- 107 Y. Tzeng, *Diamond Films Technol.*, 1991, **1**, 31.
- 108 M. Odijk and A. van den Berg, *Annu. Rev. Anal. Chem.*, 2018, **11**, 421–440.



- 109 A. Susarrey-arce, R. M. Tiggelaar, M. Morassutto, J. Geerlings and R. G. P. Sanders, *Sens. Actuators, B*, 2015, **220**, 13–21.
- 110 R. Hersig-Marx, K. T. Queeney, R. J. Jackman, M. A. Schmidt and K. F. Jensen, *Anal. Chem.*, 2004, **76**, 6476–6483.
- 111 E. Karabudak, B. L. Mojet, S. Schlautmann, G. Mul and H. J. G. E. Gardeniers, *Anal. Chem.*, 2012, **84**, 3132–3137.
- 112 B. A. Rizkin, F. G. Popovic and R. L. Hartman, *J. Vac. Sci. Technol., A*, 2019, **37**, 050801.
- 113 P. Führer, J. J. A. Lozeman, H. L. de Boer, J. G. Bomer, W. Olthuis and M. Odijk, 22nd Int. Conf. Miniaturized Syst. Chem. Life Sci. μ TAS, 2018, vol. **2018**, pp. 2127–2130.
- 114 E. Gross, *Nano Res.*, 2019, **12**, 2200–2210.
- 115 A. Dazzi and C. B. Prater, *Chem. Rev.*, 2017, **117**, 5146–5173.
- 116 Y. H. Lu, J. M. Larson, A. Baskin, X. Zhao, P. D. Ashby, D. Prendergast, H. A. Bechtel, R. Kostecki and M. Salmeron, *Nano Lett.*, 2019, **19**, 5388–5393.
- 117 F. Huth, A. Govyadinov, S. Amarie, W. Nuansing, F. Keilmann and R. Hillenbrand, *Nano Lett.*, 2012, **12**, 3973–3978.
- 118 R. Adato, S. Aksu and H. Altug, *Biochem. Pharmacol.*, 2015, **18**, 436–446.
- 119 T. Schmid and P. Dariz, *Heritage*, 2019, **2**, 1662–1683.
- 120 M. Fleischmann, P. J. Hendra and A. J. McQuillan, *Chem. Phys. Lett.*, 1974, **26**, 163–166.
- 121 D. L. Jeanmaire and R. P. Van Duyne, *J. Electroanal. Chem.*, 1977, **84**, 1.
- 122 P. L. Stiles, J. A. Dieringer, N. C. Shah and R. P. Van Duyne, *Annu. Rev. Anal. Chem.*, 2008, **1**, 601–626.
- 123 M. Moskovits, *Rev. Mod. Phys.*, 1985, **57**, 783–826.
- 124 M. Moskovits, *Phys. Chem. Chem. Phys.*, 2013, **15**, 5301–5311.
- 125 M. Moskovits, *J. Raman Spectrosc.*, 2005, **36**, 485–496.
- 126 S. Schlücker, *Angew. Chem., Int. Ed.*, 2014, **53**, 4756–4795.
- 127 J. Kim, Y. Jang, N. J. Kim, H. Kim, G. C. Yi, Y. Shin, M. H. Kim and S. Yoon, *Front. Chem.*, 2019, **7**, 1–7.
- 128 M. A. Montiel, F. J. Vidal-Iglesias, V. Montiel and J. Solla-Gullón, *Curr. Opin. Electrochem.*, 2017, **1**, 34–39.
- 129 A. Otto, *J. Raman Spectrosc.*, 1991, **22**, 743–752.
- 130 M. F. dos Santos, V. Katic, P. L. dos Santos, B. M. Pires, A. L. B. Formiga and J. A. Bonacin, *Anal. Chem.*, 2019, **91**, 10386–10389.
- 131 D. Martín-Yerga, A. Pérez-Junquera, D. Hernández-Santos and P. Fanjul-Bolado, *Electroanalysis*, 2018, **30**, 1095–1099.
- 132 O. J. R. Clarke, G. J. H. St Marie and C. L. Brosseau, *J. Electrochem. Soc.*, 2017, **164**, B3091–B3095.
- 133 C. N. Hernández, D. Martín-Yerga, M. B. González-García, D. Hernández-Santos and P. Fanjul-Bolado, *Talanta*, 2018, **178**, 85–88.
- 134 D. Martín-Yerga, A. Pérez-Junquera, M. B. González-García, J. V. Perales-Rondon, A. Heras, A. Colina, D. Hernández-Santos and P. Fanjul-Bolado, *Electrochim. Acta*, 2018, **264**, 183–190.
- 135 T. P. Lynk, O. J. R. Clarke, N. Kesavan and C. L. Brosseau, *Sens. Actuators, B*, 2018, **257**, 270–277.
- 136 A. M. Robinson, S. G. Harroun, J. Bergman and C. L. Brosseau, *Anal. Chem.*, 2012, **84**, 1760–1764.
- 137 R. A. Karaballi, A. Nel, S. Krishnan, J. Blackburn and C. L. Brosseau, *Phys. Chem. Chem. Phys.*, 2015, **17**, 21356–21363.
- 138 L. Zhao, J. Blackburn and C. L. Brosseau, *Anal. Chem.*, 2015, **87**, 441–447.
- 139 S. D. Bindesri, D. S. Alhatab and C. L. Brosseau, *Analyst*, 2018, **143**, 4128–4135.
- 140 G. Niaura, A. K. Gaigalas and V. L. Vilker, *J. Raman Spectrosc.*, 1997, **28**, 1009–1011.
- 141 R. Mažeikienė, G. Niaura and A. Malinauskas, *Chemija*, 2018, **29**, 81–88.
- 142 R. Mažeikienė, G. Niaura and A. Malinauskas, *Synth. Met.*, 2010, **160**, 1060–1064.
- 143 R. Mažeikienė, G. Niaura and A. Malinauskas, *J. Solid State Electrochem.*, 2019, **23**, 1631–1640.
- 144 R. Mažeikienė, G. Niaura and A. Malinauskas, *Synth. Met.*, 2019, **248**, 35–44.
- 145 R. Mažeikienė, G. Niaura and A. Malinauskas, *Synth. Met.*, 2018, **243**, 97–106.
- 146 R. Mažeikienė, G. Niaura and A. Malinauskas, *Spectrochim. Acta, Part A*, 2019, **221**, 117147.
- 147 R. Mažeikienė, G. Niaura, O. Eicher-Lorka and A. Malinauskas, *J. Colloid Interface Sci.*, 2011, **357**, 189–197.
- 148 R. Mažeikienė, G. Niaura and A. Malinauskas, *J. Electroanal. Chem.*, 2011, **660**, 140–146.
- 149 R. Mažeikienė, G. Niaura and A. Malinauskas, *Chemija*, 2017, **28**, 28–32.
- 150 R. Mažeikienė, G. Niaura and A. Malinauskas, *J. Solid State Electrochem.*, 2019, 2307–2316.
- 151 R. Mažeikienė, G. Niaura and A. Malinauskas, *Spectrochim. Acta, Part A*, 2017, **181**, 200–207.
- 152 R. Mažeikienė, G. Niaura and A. Malinauskas, *J. Electroanal. Chem.*, 2014, **719**, 60–71.
- 153 W. Kiefer and H. J. Bernstein, *Appl. Spectrosc.*, 1971, **25**, 609–613.
- 154 H. Yamada, T. Amamiya and H. Tsubomura, *Chem. Phys. Lett.*, 1978, **56**, 591–594.
- 155 P. Hildebrandt, K. A. Macor and R. S. Czernuszewicz, *J. Raman Spectrosc.*, 1988, **19**, 65–69.
- 156 A. J. McQuillan and R. E. Hester, *J. Raman Spectrosc.*, 1984, **15**, 15–19.
- 157 J. E. Pemberton and R. P. Buck, *Appl. Spectrosc.*, 1981, **35**, 571–576.
- 158 J. E. Pemberton and R. P. Buck, *J. Electroanal. Chem. Interfacial Electrochem.*, 1982, **136**, 201–208.
- 159 L. Kavan, P. Janda, M. Krause, F. Ziegls and L. Dunsch, *Anal. Chem.*, 2009, **81**, 2017–2021.
- 160 L. Pospíšil, M. Gál, M. Hromádová, J. Bulíková, V. Kolivka, J. Cvaka, K. Nováková, L. Kavan, M. Zúkalová and L. Dunsch, *Phys. Chem. Chem. Phys.*, 2010, **12**, 14095–14101.



- 161 P. Damlin, C. Kvarnström, A. Petr, P. Ek, L. Dunsch and A. Ivaska, *J. Solid State Electrochem.*, 2002, **6**, 291–301.
- 162 A. Viinikanoja, J. Kauppila, P. Damlin, M. Suominen and C. Kvarnström, *Phys. Chem. Chem. Phys.*, 2015, **17**, 12115–12123.
- 163 M. L. McGlashen, M. E. Blackwood and T. G. Spiro, *J. Am. Chem. Soc.*, 1993, **115**, 2074–2075.
- 164 D. Ibañez, D. Plana, A. Heras, D. J. Fermin and A. Colina, *Electrochem. Commun.*, 2015, **54**, 14–17.
- 165 F. Bonhomme, J. C. Lassègues and L. Servant, *J. Electrochem. Soc.*, 2001, **148**, 450–458.
- 166 L. Kavan and L. Dunsch, *ChemPhysChem*, 2003, **4**, 944–950.
- 167 M. Kalbáč, L. Kavan, S. Gorantla, T. Gemming and L. Dunsch, *Chem. – Eur. J.*, 2010, **16**, 11753–11759.
- 168 M. Kalbac, L. Kavan, M. Zukalová and L. Dunsch, *J. Phys. Chem. C*, 2007, **111**, 1079–1085.
- 169 M. Kalbac, Y. P. Hsieh, H. Farhat, L. Kavan, M. Hofmann, J. Kong and M. S. Dresselhaus, *Nano Lett.*, 2010, **10**, 4619–4626.
- 170 M. Kalbáč, V. Vales, L. Kavan and L. Dunsch, *Nanotechnology*, 2014, **25**, 485706.
- 171 M. Kalbac, L. Kavan and L. Dunsch, *Diamond Relat. Mater.*, 2009, **18**, 972–974.
- 172 L. Kavan, P. Raptá and L. Dunsch, *Chem. Phys. Lett.*, 2000, **328**, 363–368.
- 173 J. Tarábek, L. Kavan, L. Dunsch and M. Kalbac, *J. Phys. Chem. C*, 2008, **112**, 13856–13861.
- 174 L. Kavan, O. Frank, A. A. Green, M. C. Hersam, J. Koltai, V. Zolyomi, J. Kürti and L. Dunsch, *J. Phys. Chem. C*, 2008, **112**, 14179–14187.
- 175 L. Kavan, L. Dunsch and H. Kataura, *Chem. Phys. Lett.*, 2002, **361**, 79–85.
- 176 M. Kalbáč, L. Kavan and L. Dunsch, *Anal. Chem.*, 2007, **79**, 9074–9081.
- 177 M. Kalbáč, L. Kavan, M. Zukalová and L. Dunsch, *Adv. Funct. Mater.*, 2005, **15**, 418–426.
- 178 C. S. L. Koh, H. K. Lee, G. C. Phan-Quang, X. Han, M. R. Lee, Z. Yang and X. Y. Ling, *Angew. Chem., Int. Ed.*, 2017, **56**, 8813–8817.
- 179 L. Zhang, V. Liao and Z. Yu, *Carbon*, 2010, **48**, 2582–2589.
- 180 D. Ibañez, E. C. Romero, A. Heras and A. Colina, *Electrochim. Acta*, 2014, **129**, 171–176.
- 181 M. Tominaga, N. Watanabe and Y. Yatsugi, *J. Electroanal. Chem.*, 2017, **800**, 156–161.
- 182 A. Colina, V. Ruiz, A. Heras, E. Ochoteco, E. Kauppinen and J. López-Palacios, *Electrochim. Acta*, 2011, **56**, 1294–1299.
- 183 R. V. Salvatierra, L. G. Moura, M. M. Oliveira, M. A. Pimenta and A. J. G. Zarbin, *J. Raman Spectrosc.*, 2012, **43**, 1094–1100.
- 184 M. Kalbac, H. Farhat, J. Kong, P. Janda, L. Kavan and M. S. Dresselhaus, *Nano Lett.*, 2011, **11**, 1957–1963.
- 185 O. Frank, M. S. Dresselhaus and M. Kalbac, *Acc. Chem. Res.*, 2015, **48**, 111–118.
- 186 J. Binder, J. M. Urban, R. Stepniewski, W. Strupinski and A. Wyszomolek, *Nanotechnology*, 2016, **27**, 045704.
- 187 M. Bousa, O. Frank and L. Kavan, *Electroanalysis*, 2014, **26**, 57–61.
- 188 M. Bouša, O. Frank, I. Jirka and L. Kavan, *Phys. Status Solidi B*, 2013, **250**, 2662–2667.
- 189 S. Gupta, S. B. Carrizosa, J. Jasinski and N. Dimakis, *AIP Adv.*, 2018, **8**, 065225.
- 190 W. T. E. van den Beld, M. Odijk, R. H. J. Vervuurt, J. W. Weber, A. A. Bol, A. van den Berg and J. C. T. Eijkel, *Sci. Rep.*, 2017, **7**, 1–8.
- 191 R. Mažeikienė, G. Niaura and A. Malinauskas, *J. Electroanal. Chem.*, 2018, **808**, 228–235.
- 192 O. L. Gribkova, O. D. Iakobson, A. A. Nekrasov, V. A. Cabanova, V. A. Tverskoy, A. R. Tameev and A. V. Vannikov, *Electrochim. Acta*, 2016, **222**, 409–420.
- 193 B. Zangfronini, A. Colina, A. Heras, C. Zanardi, R. Seeber and J. López-Palacios, *Polym. Degrad. Stab.*, 2011, **96**, 2112–2119.
- 194 A. Blacha-Grzechnik, K. Piwowar, P. Zassowski, R. Motyka and J. Zak, *Electrochim. Acta*, 2017, **245**, 902–911.
- 195 S. Bilal, A. A. Shah and R. Holze, *Vib. Spectrosc.*, 2010, **53**, 279–284.
- 196 S. Shreepathi and R. Holze, *Langmuir*, 2006, **22**, 5196–5204.
- 197 A. A. Nekrasov, O. L. Gribkova, O. D. Iakobson, I. N. Ardabievskii, V. F. Ivanov and A. V. Vannikov, *Chem. Pap.*, 2017, **71**, 449–458.
- 198 Z. Morávková and E. Dmitrieva, *J. Raman Spectrosc.*, 2017, **48**, 1229–1234.
- 199 Z. Morávková and E. Dmitrieva, *ChemistrySelect*, 2019, **4**, 8847–8854.
- 200 M. Grzeszczuk, A. Kepas, C. Kvarnstrom and A. Ivaska, *Synth. Met.*, 2010, **160**, 636–642.
- 201 K. Crowley and J. Cassidy, *J. Electroanal. Chem.*, 2003, **547**, 75–82.
- 202 L. M. C. Ferreira, D. Grasseschi, M. S. F. Santos, P. R. Martins, I. G. R. Gutz, A. M. C. Ferreira, K. Araki, H. E. Toma and L. Angnes, *Langmuir*, 2015, **31**, 4351–4360.
- 203 A. J. Wilson and K. A. Willets, *Analyst*, 2016, **141**, 5144–5151.
- 204 C. Xu, H. Geng, R. Bennett, D. A. Clayton and S. Pan, *J. Phys. Chem. C*, 2013, **117**, 1849–1856.
- 205 P. Song, X. Guo, Y. Pan, Y. Wen, Z. Zhang and H. Yang, *J. Electroanal. Chem.*, 2013, **688**, 384–391.
- 206 T. Shegai, A. Vaskevich, I. Rubinstein and G. Haran, *J. Am. Chem. Soc.*, 2009, **131**, 14392–14398.
- 207 T. Itoh and R. L. McCreery, *J. Am. Chem. Soc.*, 2002, **124**, 10894–10902.
- 208 W. F. Paxton, S. L. Kleinman, A. N. Basuray, J. F. Stoddart and R. P. Van Duyne, *J. Phys. Chem. Lett.*, 2011, **2**, 1145–1149.
- 209 M. B. Pomfret, J. J. Pietron and J. C. Owrutsky, *Langmuir*, 2010, **26**, 6809–6817.



- 210 R. A. Karaballi, S. Merchant, S. R. Power and C. L. Brosseau, *Phys. Chem. Chem. Phys.*, 2018, **20**, 4513–4526.
- 211 K. E. R. McLeod, T. P. Lynk, C. S. Sit and C. L. Brosseau, *Anal. Methods*, 2019, **11**, 924–929.
- 212 D. Ibañez, A. Santidrian, A. Heras, M. Kalbáč and A. Colina, *J. Phys. Chem. C*, 2015, **119**, 8191–8198.
- 213 B. H. C. Greene, D. S. Alhatab, C. C. Pye and C. L. Brosseau, *J. Phys. Chem. C*, 2017, **121**, 8084–8090.
- 214 T. Yuan, L. Le Thi Ngoc, J. Van Nieuwkastele, M. Odijk, A. van Den Berg, H. Permentier, R. Bischoff and E. T. Carlen, *Anal. Chem.*, 2015, **87**, 2588–2592.
- 215 H. Le-The, J. J. A. Lozeman, M. Lafuente, P. Muñoz, J. G. Bommer, H. Duy-Tong, E. Berenschot, A. van den Berg, N. R. Tas, M. Odijk and J. C. T. Eijkel, *Nanoscale*, 2019, **11**, 12152–12160.
- 216 H. Le-The, R. M. Tiggelaar, E. Berenschot, A. van den Berg, N. Tas and J. C. T. Eijkel, *ACS Nano*, 2019, **13**, 6782–6789.
- 217 J. V. Perales-Rondon, S. Hernandez, A. Heras and A. Colina, *Appl. Surf. Sci.*, 2019, **473**, 366–372.
- 218 J. V. Perales-Rondon, S. Hernandez, D. Martin-Yerga, P. Fanjul-Bolado, A. Heras and A. Colina, *Electrochim. Acta*, 2018, **282**, 377–383.
- 219 A. J. Bard and M. Stratmann, *Encyclopedia of electrochemistry*, 2007, ISBN: 9783527610426, DOI: 10.1002/9783527610426.
- 220 C. Zong, C. J. Chen, M. Zhang, D. Y. Wu and B. Ren, *J. Am. Chem. Soc.*, 2015, **137**, 11768–11774.
- 221 C. Shi, W. Zhang, R. L. Birke and J. R. Lombardi, *J. Electroanal. Chem.*, 1997, **423**, 67–81.
- 222 D. H. Murgida and P. Hildebrandt, *J. Am. Chem. Soc.*, 2001, **123**, 4062–4068.
- 223 D. H. Murgida and P. Hildebrandt, *Angew. Chem., Int. Ed.*, 2001, **40**, 728–731.
- 224 C. Shi, W. Zhang, J. R. Lombardi and R. L. Birke, *J. Phys. Chem.*, 1992, **96**, 10093–10096.
- 225 W. Zhang, A. Vivoni, J. R. Lombardi and R. L. Birke, *J. Phys. Chem.*, 1995, **99**, 12846–12857.
- 226 C. Shi, W. Zhang, R. L. Birke and J. R. Lombardi, *J. Phys. Chem.*, 1990, **94**, 4766–4769.
- 227 J. Wessel, *J. Opt. Soc. Am. B*, 1985, **2**, 1538–1541.
- 228 R. M. Stöckle, Y. D. Suh, V. Deckert and R. Zenobi, *Chem. Phys. Lett.*, 2000, **318**, 131–136.
- 229 M. S. Anderson, *Appl. Phys. Lett.*, 2000, **76**, 3130–3132.
- 230 N. Hayazawa, Y. Inouye, Z. Sekkat and S. Kawata, *Opt. Commun.*, 2000, **183**, 333–336.
- 231 B. Pettinger, G. Picardi, R. Schuster and G. Ertl, *Electrochemistry*, 2000, **68**, 942–949.
- 232 F. Shao and R. Zenobi, *Anal. Bioanal. Chem.*, 2019, **411**, 37–61.
- 233 D. Kourouski, M. Mattei and R. P. Van Duyne, *Nano Lett.*, 2015, **15**, 7956–7962.
- 234 Z. C. Zeng, S. C. Huang, D. Y. Wu, L. Y. Meng, M. H. Li, T. X. Huang, J. H. Zhong, X. Wang, Z. L. Yang and B. Ren, *J. Am. Chem. Soc.*, 2015, **137**, 11928–11931.
- 235 J. F. Li, Y. F. Huang, Y. Ding, Z. L. Yang, S. B. Li, X. S. Zhou, F. R. Fan, W. Zhang, Z. Y. Zhou, D. Y. Wu, B. Ren, Z. L. Wang and Z. Q. Tian, *Nature*, 2010, **464**, 392–395.
- 236 J. F. Li, Y. J. Zhang, A. V. Rudnev, J. R. Anema, S. B. Li, W. J. Hong, P. Rajapandian, J. Lipkowski, T. Wandlowski and Z. Q. Tian, *J. Am. Chem. Soc.*, 2015, **137**, 2400–2408.
- 237 L. A. Colnago, F. D. Andrade, A. A. Souza, R. B. V. Azeredo, A. A. Lima, L. M. Cerioni, T. M. Osán and D. J. Pusiol, *Chem. Eng. Technol.*, 2014, **37**, 191–203.
- 238 L. M. Silva Nunes, T. B. Moraes, L. L. Barbosa, L. H. Mazo and L. A. Colnago, *Anal. Chim. Acta*, 2014, **850**, 1–5.
- 239 T. Kobayashi, P. K. Babu, J. H. Chung, E. Oldfield and A. Wieckowski, *J. Phys. Chem. C*, 2007, **111**, 7078–7083.
- 240 P. K. Babu, J. H. Chung, E. Oldfield and A. Wieckowski, *Electrochim. Acta*, 2008, **53**, 6672–6679.
- 241 P. K. Babu, H. S. Kim, J. H. Chung, E. Oldfield and A. Wieckowski, *J. Phys. Chem. B*, 2004, **108**, 20228–20232.
- 242 J. B. Day, P. A. Vuissoz, E. Oldfield, A. Wieckowski and J. P. Ansermet, *J. Am. Chem. Soc.*, 1996, **118**, 13046–13050.
- 243 P. K. Babu, H. S. Kim, E. Oldfield and A. Wieckowski, *J. Phys. Chem. B*, 2003, **107**, 7595–7600.
- 244 Y. Y. Tong, S. K. Hee, P. K. Babu, P. Waszczuk, A. Wieckowski and E. Oldfield, *J. Am. Chem. Soc.*, 2002, **124**, 468–473.
- 245 Y. Y. Tong, C. Rice, A. Wieckowski and E. Oldfield, *J. Am. Chem. Soc.*, 2000, **122**, 1123–1129.
- 246 Y. Y. Tong, C. Rice, A. Wieckowski and E. Oldfield, *J. Am. Chem. Soc.*, 2000, **122**, 11921–11924.
- 247 Y. Tong, C. Rice, N. Godbout, A. Wieckowski and E. Oldfield, *J. Am. Chem. Soc.*, 1999, **121**, 2996–3003.
- 248 C. Rice, Y. Tong, E. Oldfield and A. Wieckowski, *Electrochim. Acta*, 1998, **43**, 2825–2830.
- 249 H. Yano, J. Inukai, H. Uchida, M. Watanabe, P. K. Babu, T. Kobayashi, J. H. Chung, E. Oldfield and A. Wieckowski, *Phys. Chem. Chem. Phys.*, 2006, **8**, 4932–4939.
- 250 Y. Y. Tong, A. Wieckowski and E. Oldfield, *J. Phys. Chem. B*, 2002, **106**, 2434–2446.
- 251 P. A. Vuissoz, J. P. H. Ansermet and A. Wieckowski, *Phys. Rev. Lett.*, 1999, **83**, 2457–2460.
- 252 P. J. Slezak and A. Wieckowski, *J. Electroanal. Chem.*, 1992, **339**, 401–410.
- 253 S. Klod, F. Ziegls and L. Dunsch, *Anal. Chem.*, 2009, **81**, 10262–10267.
- 254 J. Z. Hu, N. R. Jaegers, M. Y. Hu and K. T. Mueller, *J. Phys.: Condens. Matter*, 2018, **30**, 463001.
- 255 R. D. Webster, *Anal. Chem.*, 2004, **76**, 1603–1610.
- 256 X. Zhang and J. W. Zwanziger, *J. Magn. Reson.*, 2011, **208**, 136–147.
- 257 S. Klod and L. Dunsch, *Magn. Reson. Chem.*, 2011, **49**, 725–729.
- 258 P. Raptá, K. Haubner, P. Machata, V. Lukeš, M. Rosenkranz, S. Schiemenz, S. Klod, H. Kivelä, C. Kvarnström, H. Hartmann and L. Dunsch, *Electrochim. Acta*, 2013, **110**, 670–680.



- 259 M. Zalibera, P. Machata, T. T. Clikeman, M. Rosenkranz, S. H. Strauss, O. V. Boltalina and A. A. Popov, *Analyst*, 2015, **140**, 7209–7216.
- 260 L. Huang, E. G. Sorte, S. G. Sun and Y. Y. J. Tong, *Chem. Commun.*, 2015, **51**, 8086–8088.
- 261 U. Bussy, P. Giraudeau, V. Silvestre, T. Jaunet-Lahary, V. Ferchaud-Roucher, M. Krempf, S. Akoka, I. Tea and M. Boujtita, *Anal. Bioanal. Chem.*, 2013, **405**, 5817–5824.
- 262 R. Boisseau, U. Bussy, P. Giraudeau and M. Boujtita, *Anal. Chem.*, 2015, **87**, 372–375.
- 263 B. Ferreira Gomes, P. Ferreira da Silva, C. M. Silva Lobo, M. da Silva Santos and L. A. Colnago, *Anal. Chim. Acta*, 2017, **983**, 91–95.
- 264 E. J. Kelly, *J. Electrochem. Soc.*, 1977, **124**, 987–994.
- 265 B. Ferreira Gomes, L. M. Silva Nunes, C. M. Silva Lobo, L. F. Cabeça and L. A. Colnago, *Anal. Chem.*, 2014, **86**, 9391–9393.
- 266 L. M. A. Monzon and J. M. D. Coey, *Electrochem. Commun.*, 2014, **42**, 38–41.
- 267 D. W. Mincey, M. J. Popovich, P. J. Faustino, M. M. Hurst and J. A. Caruso, *Anal. Chem.*, 1990, **62**, 1197–1200.
- 268 S. H. Cao, Z. R. Ni, L. Huang, H. J. Sun, B. Tang, L. J. Lin, Y. Q. Huang, Z. Y. Zhou, S. G. Sun and Z. Chen, *Anal. Chem.*, 2017, **89**, 3810–3813.
- 269 E. G. Sorte and Y. J. Tong, *J. Electroanal. Chem.*, 2016, **769**, 1–4.
- 270 E. G. Sorte, S. Jilani and Y. Y. J. Tong, *Electrocatalysis*, 2017, **8**, 95–102.
- 271 X. P. Zhang, W. L. Jiang, S. H. Cao, H. J. Sun, X. Q. You, S. H. Cai, J. L. Wang, C. Sen Zhao, X. Wang, Z. Chen and S. G. Sun, *Electrochim. Acta*, 2018, **273**, 300–306.
- 272 J. Wang, X. Cao, L. Fang, X. You, K. Wong, S. Cao, C. Xiao, S. Cai, Y. Huang, X. Zhang and Z. Chen, *Int. J. Hydrogen Energy*, 2019, **44**, 16411–16423.
- 273 J. Wang, X. You, C. Xiao, X. Zhang, S. Cai, W. Jiang, S. Guo, S. Cao and Z. Chen, *Appl. Catal., B*, 2019, **259**, 118060.
- 274 S. H. Cao, S. Liu, H. J. Sun, L. Huang, Z. R. Ni, W. L. Jiang, M. Zhan, Z. Y. Zhou, S. G. Sun and Z. Chen, *Anal. Chem.*, 2019, **91**, 1686–1691.
- 275 P. Ferreira da Silva, B. Ferreira Gomes, C. M. Silva Lobo, L. H. K. Queiroz Júnior, E. Danieli, M. Carmo, B. Blümich and L. A. Colnago, *Microchem. J.*, 2019, **146**, 658–663.
- 276 J. B. Richter, C. Eßbach, I. Senkovska, S. Kaskel and E. Brunner, *Chem. Commun.*, 2019, **55**, 6042–6045.
- 277 H. Wang, T. K. J. Köster, N. M. Trease, J. Ségalini, P. L. Taberna, P. Simon, Y. Gogotsi and C. P. Grey, *J. Am. Chem. Soc.*, 2011, **133**, 19270–19273.
- 278 B. Ferreira Gomes, C. M. Silva Lobo and L. A. Colnago, *Appl. Sci.*, 2019, **9**, 498.
- 279 A. P. Bruins, *TrAC, Trends Anal. Chem.*, 2015, **70**, 14–19.
- 280 C. Dass, *Fundamentals of Contemporary Mass Spectrometry*, Wiley, 2007.
- 281 W. Plieth, G. S. Wilson and C. Gutiérrez De La Fe, *Pure Appl. Chem.*, 1998, **70**, 1395–1414.
- 282 O. Kasian, S. Geiger, K. J. J. Mayrhofer and S. Cherevko, *Chem. Rec.*, 2019, **19**, 2130–2142.
- 283 S. O. Klemm, A. A. Topalov, C. A. Laska and K. J. J. Mayrhofer, *Electrochem. Commun.*, 2011, **13**, 1533–1535.
- 284 S. O. Klemm, A. Karschin, A. K. Schuppert, A. A. Topalov, I. Katsounaros and K. J. J. Mayrhofer, *J. Electroanal. Chem.*, 2013, **693**, 127.
- 285 E. M. Yuill and L. A. Baker, *ChemElectroChem*, 2017, **4**, 806–821.
- 286 X. Xu, W. Lu and R. B. Cole, *Anal. Chem.*, 1996, **2700**, 4244–4253.
- 287 F. T. G. van den Brink, L. Büter, M. Odijk, W. Olthuis, U. Karst and A. van den Berg, *Anal. Chem.*, 2015, **87**, 1527–1535.
- 288 H. Faber, M. Vogel and U. Karst, *Anal. Chim. Acta*, 2014, **834**, 9–21.
- 289 R. Zabel and G. Weber, *Anal. Bioanal. Chem.*, 2016, **408**, 1237–1247.
- 290 M. Odijk, W. Olthuis, A. van den Berg, L. Qiao and H. Girault, *Anal. Chem.*, 2012, **84**, 9176–9183.
- 291 L. Qiao, Y. Lu, B. Liu and H. H. Girault, *J. Am. Chem. Soc.*, 2011, **133**, 19823–19831.
- 292 K. Yu, H. Zhang, J. He, R. N. Zare, Y. Wang, L. Li, N. Li, D. Zhang and J. Jiang, *Anal. Chem.*, 2018, **90**, 7154–7157.
- 293 J. Jiang, H. Zhang, M. Li, M. T. Dulay, A. J. Ingram, N. Li, H. You and R. N. Zare, *Anal. Chem.*, 2015, **87**, 8057–8062.
- 294 R. Qiu, X. Zhang, H. Luo and Y. Shao, *Chem. Sci.*, 2016, **7**, 6684–6688.
- 295 W. Guo, H. Ding, C. Gu, Y. Liu, X. Jiang, B. Su and Y. Shao, *J. Am. Chem. Soc.*, 2018, **140**, 15904–15915.
- 296 J. L. Owens, H. A. Marsh and G. Dryhurst, *J. Electroanal. Chem.*, 1978, **91**, 231–247.
- 297 Z. Jusys, H. Massong and H. Baltruschat, *J. Electrochem. Soc.*, 1999, **146**, 1093–1098.
- 298 O. Wolter and J. Heitbaum, *Ext. Abstr., Meet. – Int. Soc. Electrochem.*, 1984, **6**, 454–456.
- 299 O. Wolter and J. Heitbaum, *Phys. Chem. Chem. Phys.*, 1984, **88**, 6–10.
- 300 A. A. Abd-El-Latif, C. J. Bondue, S. Ernst, M. Hegemann, J. K. Kaul, M. Khodayari, E. Mostafa, A. Stefanova and H. Baltruschat, *TrAC, Trends Anal. Chem.*, 2015, **70**, 4–13.
- 301 D. B. Trimarco, S. B. Scott, A. H. Thilsted, J. Y. Pan, T. Pedersen, O. Hansen, I. Chorkendorff and P. C. K. Vesborg, *Electrochim. Acta*, 2018, **268**, 520–530.
- 302 M. Heinen, Y. X. Chen, Z. Jusys and R. J. Behm, *Electrochim. Acta*, 2007, **52**, 5634–5643.
- 303 S. Brimaud, Z. Jusys and R. Jürgen Behm, *Beilstein J. Nanotechnol.*, 2014, **5**, 735–746.
- 304 J. Schnaidt, M. Heinen, Z. Jusys and R. J. Behm, *J. Phys. Chem. C*, 2012, **116**, 2872–2883.
- 305 J. Schnaidt, M. Heinen, Z. Jusys and R. J. Behm, *Catal. Today*, 2013, **202**, 154–162.
- 306 R. Reichert, J. Schnaidt, Z. Jusys and R. J. Behm, *ChemPhysChem*, 2013, **14**, 3678–3681.
- 307 M. Gorlin, J. F. De Araujo, H. Schmies, D. Bernsmeier, S. Dresch, M. Gliech, Z. Jusys, P. Chernev, R. Kraehnert, H. Dau and P. Strasser, *J. Am. Chem. Soc.*, 2017, **139**, 2070–2082.



- 308 D. B. Trimarco, T. Pedersen, O. Hansen, I. Chorkendorff and P. C. K. Vesborg, *Rev. Sci. Instrum.*, 2015, **86**, 075006.
- 309 C. Roy, B. Sebok, S. B. Scott, E. M. Fiordaliso, J. E. Sørensen, A. Bodin, D. B. Trimarco, C. D. Damsgaard, P. C. K. Vesborg, O. Hansen, I. E. L. Stephens, J. Kibsgaard and I. Chorkendorff, *Nat. Catal.*, 2018, **1**, 820–829.
- 310 A. Winiwarter, L. Silvioli, S. B. Scott, K. Enemark-Rasmussen, M. Sariç, D. B. Trimarco, P. C. K. Vesborg, P. G. Moses, I. E. L. Stephens, B. Seger, J. Rossmeisl and I. Chorkendorff, *Energy Environ. Sci.*, 2019, **12**, 1055–1067.
- 311 B. Liu, X. Y. Yu, Z. Zhu, X. Hua, L. Yang and Z. Wang, *Lab Chip*, 2014, **14**, 855–859.
- 312 Z. Wang, Y. Zhang, B. Liu, K. Wu, S. Thevuthasan, D. R. Baer, Z. Zhu, X. Y. Yu and F. Wang, *Anal. Chem.*, 2017, **89**, 960–965.
- 313 J. Yu, Y. Zhou, X. Hua, S. Liu, Z. Zhu and X. Y. Yu, *Chem. Commun.*, 2016, **52**, 10952–10955.
- 314 T. C. Rohner, J. S. Rossier and H. H. Girault, *Anal. Chem.*, 2001, **73**, 5353–5357.
- 315 T. C. Rohner, J. S. Rossier and H. H. Girault, *Electrochem. Commun.*, 2002, **4**, 695–700.
- 316 G. J. Van Berkel, J. M. E. Quirke, R. A. Tigani, A. S. Dilley and T. R. Covey, *Anal. Chem.*, 1998, **70**, 1544–1554.
- 317 Á. I. López-Lorente, J. Izquierdo, C. Kranz and B. Mizaikoff, *Vib. Spectrosc.*, 2017, **91**, 147–156.
- 318 W. Cheuquepán, J. M. Pérez, J. M. Orts and A. Rodes, *J. Phys. Chem. C*, 2014, **118**, 19070–19084.
- 319 M. Dunwell, X. Yang, Y. Yan and B. Xu, *J. Phys. Chem. C*, 2018, **122**, 24658–24664.
- 320 A. Zimmermann, U. Künzelmann and L. Dunsch, *Synth. Met.*, 1998, **93**, 17–25.

