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Advances in steroid purification for novel techniques in carbon isotope ratio mass spectrometry of doping control

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Doping detection is a critical tool for maintaining fairness in competitive sports. Steroids are widely abused due to their ability to promote protein synthesis, enhance muscle growth, and improve athletic performance, leading to significant unfair competition. Carbon isotope ratios (CIRs) are effective in distinguishing between endogenous and exogenous steroid sources, making steroids an ideal target for isotope ratio analysis in anti-doping analysis. High-precision isotope ratio measurements using Gas Chromatography-Isotope Ratio Mass Spectrometry (GC-IRMS) require high separation between the target steroid and adjacent compounds to avoid interference. However, steroid concentrations in biological matrices like urine and blood are often trace and accompanied by a large number of coexisting interferents, making purification challenging. These limitations have partially restricted the use of compound-specific isotope analysis in doping detection. This review highlights the latest advancements over the past decade in sample preparation, liquid-phase purification, and both gas-phase and liquid-phase isotope ratio mass spectrometry (IRMS) techniques. By summarizing the application of these methods in anti-doping efforts and exploring future research directions, this review aims to enhance the precision and reliability of steroid doping detection technologies, providing scientific support for anti-doping efforts and identifying the abuse of steroids.

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1. Introduction

Anabolic steroids (AS) are compounds that enhance athletic performance by mimicking or amplifying the effects of endogenous hormones in the human body. The most common and widely studied category of these substances is anabolic-androgenic steroids (AAS).^{1,2} AAS are synthetic compounds with structures and functions similar to endogenous male hormones. AAS have two main effects: anabolic effects, which promote protein synthesis and increase muscle mass, strength, and bone density; and androgenic effects, which drive the development and maintenance of male secondary sexual characteristics.³ In addition to AAS, other substances with similar effects such as selective androgen receptor modulators (SARMs) are also considered steroid stimulants. While SARMs are not structurally steroids, they selectively bind to androgen receptors, producing anabolic effects similar to those of AAS.^{1,2} The

abuse of stimulants can be traced back to the mid-20th century, when reports first emerged of athletes using testosterone and its derivatives to enhance their physical performance.⁴ In modern competitive sports, the misuse of steroid stimulants is most prevalent in strength-based and explosive events, such as weightlifting and track and field throws,⁵ at the same time, there are also reports of abuse in endurance sports.⁶ These substances can significantly improve an athlete's muscle mass, strength, explosiveness, and potentially aid recovery, providing an unfair advantage in competition. Athletes typically use synthetic anabolic steroids through injection or oral administration, often in long-term or cyclical regimens. Despite ongoing updates to the World Anti-Doping Agency (WADA) prohibited list of substance and enhanced detection methods for anabolic agents, athletes continue to find new ways to evade detection.⁷ For instance, some athletes may use novel steroids not yet listed on the banned list or modify their drug usage cycles, employ masking agents, or use other techniques to reduce the concentration of steroids in their bodies and avoid detection.⁸ The abuse of AAS not only violates the principles of fair competition but also poses severe health risks to athletes. Long-term or high-dose use of AAS can result in liver damage, increased risk of cardiovascular diseases, reproductive system dysfunction and other serious adverse effects. It may also lead

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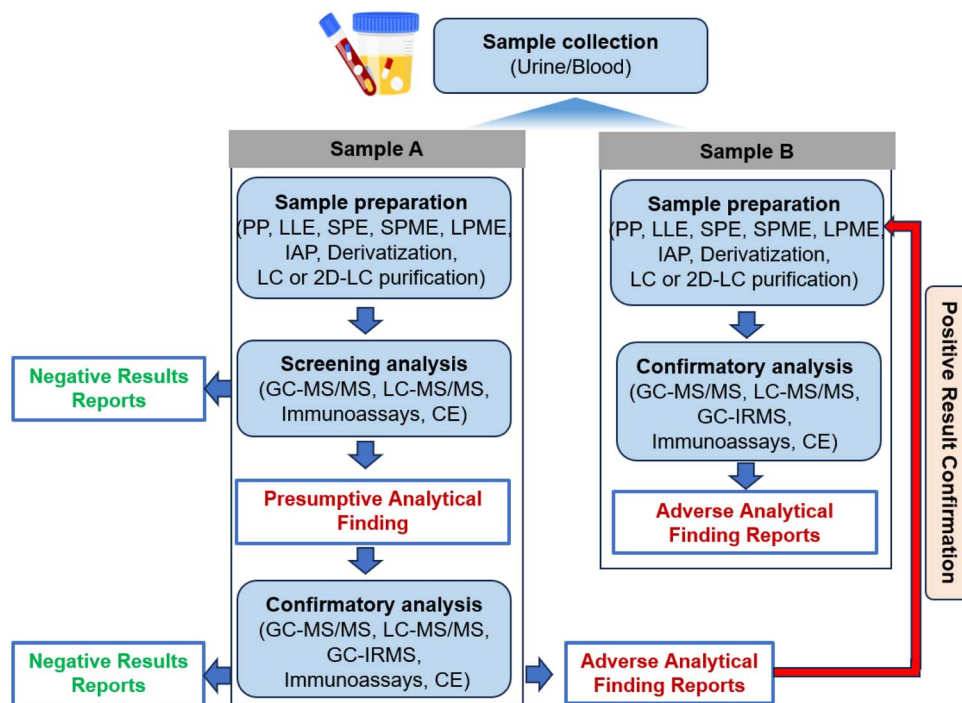


Fig. 1 Typical workflow of doping control analysis.

to psychological and physiological dependence, increasing the risk of sudden death.^{1,3,5}

Doping detection is an important strategy for ensuring fairness in sports competitions and protecting athletes' health. Through rigorous testing, the use of prohibited substances by athletes can be identified and prevented in a timely. However, current doping detection still faces several technical and practical challenges,⁹ including the detection of low-concentration drug residues, interference from complex biological matrices, and the identification of metabolites.^{10,11} Firstly, detecting low concentrations of drug residues remains a significant challenge in the field of doping detection. Many banned substances and their metabolites are present in body fluids at extremely low concentrations, typically at the ng mL^{-1} or even at pg mL^{-1} level. This imposes strict requirements on the sensitivity and accuracy of detection techniques.¹² Secondly, the interference from endogenous substances in biological matrices of human body is another critical issue in doping detection. Biological samples such as blood and urine contain a wide range of endogenous substances that are chemically or physically similar to exogenous steroids, making it difficult to accurately distinguish between them.¹³ Moreover, steroids undergo complex metabolic processes in the body, generating various metabolites that closely resemble the parent compounds in structure. This further complicates detection and introduces additional uncertainties.¹⁴ Therefore, sample preparation and purification steps are crucial in the detection process, as they directly impact the accuracy, sensitivity, and reliability of the results. To address these challenges, optimizing existing technologies and developing novel detection methods will be the focus field for future doping analysis.

To address the various challenges in steroid doping detection, researchers have developed and applied a range of advanced techniques to enhance the sensitivity, specificity, and accuracy of detection methods¹⁵ (Fig. 1). For example, protein precipitation (PP), solid-phase extraction (SPE) and liquid-liquid extraction (LLE), which are commonly used sample preparation techniques, effectively remove interfering substances from biological matrices and concentrate the targets, thereby significantly improving the accuracy of subsequent analyses.¹⁶ These techniques in simplifying sample matrices, reducing potential interferences and improving analyte recovery, making them indispensable in the processing of complex biological samples such as urine and blood. In addition, IRMS has gained widespread application in steroid detection in recent years.^{17–19} By precisely measuring the CIRs ($^{13}\text{C}/^{12}\text{C}$) in steroid compounds, IRMS can effectively distinguish between endogenous and exogenous steroids, overcoming the limitations of traditional mass spectrometry techniques in this regard and greatly enhancing the specificity of detection. Meanwhile, the Athlete Biological Passport (ABP) has become an innovative tool for detecting doping abuse in the anti-doping field.²⁰ The ABP monitors long-term changes in an athlete's biomarkers, such as blood parameters and hormone levels, to establish an individual baseline.²¹ Unlike traditional one-time testing methods, the ABP can identify abnormal fluctuations that deviate from an athlete's normal physiological variations, thus indirectly indicating the use of performance-enhancing substances. This approach not only improves detection sensitivity but also compensates for the limitations of single-sample testing, providing robust support for steroid doping detection.

Table 1 List of abbreviations

| Abbreviation | Full term | Abbreviation | Full term |
|--------------|---|--------------|---|
| AS | Anabolic steroids | LPME | Liquid-phase microextraction |
| AAS | Anabolic-androgenic steroids | MIPs | Molecularly imprinted polymers |
| ABP | Athlete biological passport | MD-LC | Multidimensional liquid chromatography |
| AAFs | Adverse analytical findings | MSPE | Magnetic solid phase extraction |
| CIRs | Carbon isotope ratios | MSTFA | <i>N</i> -Methyl- <i>N</i> -(trimethylsilyl) trifluoroacetamide |
| DES | Deep eutectic solvents | MTBE | Methyl <i>tert</i> -butyl ether |
| DHEA | Dehydroepiandrosterone | PP | Protein precipitation |
| EpiA | Epiandrosterone | PSL | Prednisolone |
| EpiT | Epitestosterone | PS | Prednisone |
| ERC | Endogenous reference compound | SARMs | Selective androgen receptor modulators |
| GC-IRMS | Gas chromatography-isotope ratio mass spectrometry | SFE | Supercritical fluid extraction |
| GC-MS/MS | Gas chromatography-tandem mass spectrometry | SLE | Supported liquid extraction |
| HRMS | High-resolution mass spectrometry | SPE | Solid phase extraction |
| HTC | High-temperature conversion | SPME | Solid phase microextraction |
| ILs | Ionic liquids | T | Testosterone |
| IRMS | Isotope ratio mass spectrometry | TC | Target compound |
| LLE | Liquid-liquid extraction | VPDB | Vienna Pee dee belemnite |
| LC-IRMS | Liquid chromatography-isotope ratio mass spectrometry | WADA | World Anti-Doping Agency |
| LC-MS/MS | Liquid chromatography-tandem mass spectrometry | WCOC | Wet chemical oxidation conversion |
| LOD | Limit of detection | | |

This review aims to summarize the latest advancements in sample purification, analytical techniques, and IRMS in steroid doping detection. It will also explore the technical challenges and solutions encountered in steroid detection over the past decade. Furthermore, the review will look ahead to potential future developments to further enhance the scientificity and fairness of steroid detection. All the abbreviations used throughout the manuscript are listed in Table 1.

2. Advances in sample preparation techniques

Purification and analysis methods for steroid stimulants are crucial for their detection and identification. Since steroids are often present at low concentrations in biological samples with complex matrices, using highly sensitive and specific analytical techniques is key to achieving accurate results. In recent years, advancements in sample preparation methods have not only improved sensitivity and specificity, but also simplified workflows, reduced processing time and lowered operational complexity (in Table 2). Common sample preparation techniques include protein PP, LLE, SPE, supercritical fluid extraction (SFE), and liquid-phase microextraction (LPME).^{22,23} PP and LLE remain classical sample pretreatment methods. PP involves the addition of organic solvents or salts to disrupt the hydration layer of proteins, leading to protein denaturation and precipitation, thereby releasing the analytes into the supernatant. On the other hand, LLE achieves separation and purification based on differences in partition coefficients of the target analytes between two immiscible solvents. In steroid detection, urine and plasma are often used as the aqueous phase, with appropriate organic solvents selected for protein precipitation and extraction. The solvents including acetonitrile, methanol and inorganic salts are the commonly reagents of PP methods,^{24–26} while the frequently organic solvents for LLE are mainly including

methyl *tert*-butyl ether (MTBE), ethyl acetate and dichloromethane and *etc.*^{26–28} Urine or blood samples often require enzymatic hydrolysis to convert conjugated steroids into their free forms, thereby improving extraction efficiency. For instance, Mazzarino *et al.* used β -glucuronidase to hydrolyze urine samples before employing LLE to extract endogenous corticosteroids, achieving a limit of detection (LOD) below 1 ng mL^{−1} and demonstrating excellent sensitivity for certain steroids.²⁹ Similarly, Makvandi *et al.* utilized methanol-acetonitrile (1 : 5) for PP, followed by methanol-based LLE, to detect steroids in serum samples. Compared to urine testing, their method identified anabolic-androgenic steroids (AAS) in 80% of serum samples, their method identified anabolic-androgenic steroids (AAS) in 80% of serum samples, highlighting the potential of serum as a complementary matrix for AAS detection.²⁵ Furthermore, Yuan *et al.* developed an MTBE-based LC-MS/MS method for detecting 12 steroid hormones in serum, achieving limits of quantification ranging from 0.005 ng mL^{−1} to 1 ng mL^{−1} and recovery rates between 86.4% and 115.0%, demonstrating high sensitivity and specificity.³⁰

However, traditional liquid-liquid extraction (LLE) has several limitations, including high solvent consumption, low selectivity and a tendency to form emulsions. To address these issues, researchers have begun exploring environmentally friendly solvent alternatives, such as deep eutectic solvents (DES),³¹ ionic liquids (ILs)³² and bio-based solvents.³³ These innovative solutions offer potential advantages in terms of sustainability and efficiency. Whereas, their application in doping detection remains limited, primarily due to the need for further validation of protocols and assessment of their compatibility with current analytical workflows.

SPE is another widely used sample preparation technique that separates and enriches target compounds based on their selective interactions with solid-phase adsorbents. It can effectively remove matrix interferences, concentrates target analytes





Table 2 Summary of sample pretreatment techniques

| Sample | Sample extraction | Solvent | Derivatization reagent | Analytical technique | Substances | Stimulants | Ref. | Cite |
|--------------|----------------------------------|---|------------------------------------|-----------------------|--------------------------------|------------|------|------|
| Serum | LLE | Dichloromethane | Dansyl chloride | LC-MS/MS | 15 Estrogens metabolites | No | 2014 | 56 |
| Saliva | SPME (PAN-HLB and PDMS-HLB) | — | — | LC-MS/MS | 6 Steroids | Yes | 2014 | 48 |
| Water | SPME (PDMS-DVB) | — | — | GC-MS/MS | 8 Steroids | Yes | 2014 | 47 |
| Urine | Hydrolysis, automated-SPE | Isopropyl alcohol-methanol (1 : 1) | — | LC-MS/MS | 18 Exogenous anabolic steroids | Yes | 2014 | 54 |
| Serum | LLE | Chloroform | MSTFA | GC-MS/MS | 4 Steroids | Yes | 2015 | 27 |
| Urine/plasma | (Hollow fiber)-LPME | Ethylene glycol, ((Me Ph)3PBr), ((Me Ph)3Pi) | — | HPLC-UV | 2 Steroids | Yes | 2017 | 50 |
| Urine | SPE-C18, hydrolysis, derivat. | Methanol | MSTFA | GC-MS/MS | 22 Endogenous steroids | Yes | 2017 | 55 |
| Urine | Hydrolysis, LLE, derivat. | MTBE (LLE) | MSTFA | GC-Orbitrap | 30 Exogenous steroids | Yes | 2018 | 57 |
| Serum | SLE | Dichloromethane | MSTFA | GC-MS/MS | 37 Steroids | Yes | 2018 | 44 |
| Foods | (Multiple monolithic fiber)-SPME | Methanol | — | HPLC-UV | 5 Steroids | Yes | 2019 | 41 |
| Urine | LLE, hydrolysis, LLE | MTBE (LLE) | — | LC-MS/MS | 8 Gglucocorticoids | Yes | 2019 | 29 |
| Urine | SPE-MIPs | Methanol | — | HPLC-UV | 2 Estrogens | Yes | 2019 | 43 |
| Serum | PP, LLE | Acetonitrile (PP), MTBE (LLE) | Isonicotinoyl chloride (INC) | LC-MS/MS | 12 Steroids | Yes | 2020 | 30 |
| Urine | Hydrolysis, SPE-HLB | Acetonitrile | — | LC-MS/MS | 23 Adrenal steroids | Yes | 2020 | 36 |
| Urine | SPE-C18, hydrolysis, LLE | Methanol (SPE), ether (LLE) | — | UHPLC-MS/MS | 9 Testosterone metabolites | Yes | 2020 | 35 |
| Serum | PP, LLE, derivat., SPE-C18 | Acetonitrile(PP), hexane/ethyl acetate (LLE) | Girard T (GT) and Girard P (GP) | LC-MS/MS | 14 Testosterone esters | Yes | 2021 | 61 |
| Hair | LLE, automated-SLE | Methanol (LLE), ethyl acetate (SLE) | — | LC-MS/MS | 5 Steroid hormones | Yes | 2021 | 45 |
| Serum | Magnetic-SPE | Methanol | Hydroxylamine hydrochloride (HAHC) | LC-MS/MS | 2 Steroids | Yes | 2022 | 42 |
| Urine | Hydrolysis SLE, derivat. | MTBE (SLE) | MSTFA | GC-MS/MS | 14 Steroids | Yes | 2022 | 46 |
| Serum | PP, LLE | Methanol-acetonitrile (1 : 5), methanol (LLE) | — | LC-MS/MS | 8 Anabolic steroids | Yes | 2023 | 25 |
| Serum | PP, derivat., LLE | Methanol-acetonitrile (1 : 1), MTBE (LLE) | Hydroxylamine hydrochloride (HAHC) | LC-MS/MS | 2 Steroids | Yes | 2023 | 26 |
| Urine/Serum | SPE-MIPs | Methanol-acetic acid (9 : 1) | — | HPLC-UV | Megestrol acetate | No | 2023 | 39 |
| Serum | Hydrolysis, LLE, SPE-C18 | Chloroform (LLE) | MSTFA | GC-MS/MS, UHPLC-MS/MS | 34 Ssteroids | Yes | 2024 | 58 |
| Plasma | Hydrolysis, (thin film)-SPME | — | — | LC-MS/MS | 4 Androgens | Yes | 2024 | 49 |
| Blood | PP, SPE-(C18 and PSA) | Water-methanol (95 : 5) (PP) | — | UHPSFCMS/MS | 9 Steroids | Yes | 2024 | 52 |
| Urine | Hydrolysis, SLE, derivat. | Dichloromethane (SLE) | Pyridine and acetic anhydride | UHPSFCMS/MS | 10 Steroids | Yes | 2024 | 53 |
| Serum | SPE-MIPs | Acetone-acetonitrile (4 : 1) | — | HPLC-UV | 4 Estrogens | No | 2024 | 40 |

and then enhance the sensitivity of detection. The choice of sorbent material is critical for SPE performance, with commonly used materials including reversed-phase C18 and amino-functionalized columns.³⁴

For example, Derly *et al.* employed C18 SPE cartridge to analyze testosterone (T) and hydroxylated metabolites in human urine, revealing age-dependent changes, with most metabolite levels decreasing in older men.³⁵ Similarly, Pussard *et al.* utilized Oasis HLB as an SPE adsorbent to extract and enrich endogenous anabolic steroids (EAAS) in human urine, including both free and conjugated forms of glucocorticoids and mineralocorticoids. This method demonstrated good linearity and accuracy, making it suitable for single-injection quantification of multiple steroids.³⁶

In recent years, researchers have focused on developing novel SPE materials to further improve the separation and purification efficiency of steroids.³⁷ One significant advancement is the application of molecularly imprinted polymers (MIPs) in SPE.³⁸ MIPs are functional polymers prepared through polymerization in the presence of template molecules (*i.e.*, target analytes), enabling highly specific recognition similar to antibodies. Using MIPs as SPE adsorbents allows for the selective adsorption of target steroid molecules, significantly improving purification efficiency and sensitivity. For instance, Pournamdari *et al.* synthesized MIPs for medroxyprogesterone using methacrylic acid and applied them to selectively adsorb the drug in human serum and urine, demonstrating the potential of MIPs-SPE for trace analysis in complex matrices.³⁹ Guo *et al.* utilized an MIPs-SPE method to efficiently enrich and quantify four estrogens in serum, enhancing the sensitivity of HPLC and reducing matrix interferences.⁴⁰ In addition to MIPs, researchers are actively exploring other innovative SPE materials. New adsorbents, such as graphene-based materials, nanomaterials, and magnetic media, show promise in steroid separation and purification due to their unique pore structures and surface chemistry.^{40–43} Magnetic solid-phase extraction (MSPE) has also garnered significant attention for its use of magnetic nanomaterials as sorbents. By applying an external magnetic field, MSPE enables to simplify the workflow and shorten sample preparation time.⁴² Additionally, supported-liquid extraction (SLE) has been applied as an alternative method. In SLE, sample solutions form a thin film on the surface of inert porous sorbents, and target analytes are extracted using a solvent immiscible with the film. This technique is particularly effective for extracting nonpolar and moderately polar compounds from biological fluids.^{44–46} These above growing array of advanced materials and techniques highlights the ongoing innovation in sample preparation for steroid analysis, offering solutions to improve efficiency, sensitivity and environmental sustainability in doping detection.

3. Advances in semi-preparative liquid chromatography purification techniques

Samples are typically derived from complex biological matrices, such as urine and serum, which contain numerous potential

interferences that pose significant challenges to subsequent analysis.^{51,59,60,62–64} Table 3 summarized the research progress of steroid purification and preparation. Semi-preparative HPLC is a powerful purification tool that effectively isolates and purifies target steroids and their metabolites from complex matrices, providing high-quality samples for structural identification, standard preparation and IRMS analysis. The principles of semi-preparative HPLC are similar to HPLC, relying on differences in partition coefficients of target compounds between the stationary and mobile phases. The key difference lies in the use of larger diameter columns and higher flow rates in semi-preparative HPLC, allowing for greater sample loading and improved separation efficiency to yield milligram-scale purified substances.⁶⁵

The choice of stationary phase is critical for the separation efficiency and purity achieved with semi-preparative HPLC. Common stationary phases include reversed-phase C18, C8, phenyl columns, normal-phase silica columns, amino columns, and other specialized functionalized materials.⁶⁶ For steroid separation, C18 columns are the most widely used, particularly for their hydrophobicity, which makes them suitable for separating most steroid compounds. For example, in the separation of hydrophobic steroids such as T and dehydroepiandrosterone (DHEA), C18 stationary phases effectively distinguish these compounds, ensuring high-purity separation in complex biological matrices such as urine and serum.^{67–71} Due to their superior resolution, C18 columns are commonly used for precise quantification and identification of steroids. C8 columns, with weaker hydrophobicity than C18 columns, are better suited for separating smaller or more polar steroid compounds.⁷² Normal-phase silica relies on the interaction of silanol groups (Si–OH) with the polar functional groups in steroid molecules, such as hydroxyl or keto groups, through hydrogen bonding.⁷³ However, C8 and normal-phase silica columns are less commonly used for steroid purification. Phenyl columns, which utilize hydrophobic interactions and π – π interactions are particularly effective for nonpolar and weakly polar compounds, especially those with aromatic rings.^{74–77} Amino columns, featuring medium polarity through aminopropyl-functionalized silica, are mainly used for separating steroids with hydroxyl or keto groups. The amino groups on the stationary phase form hydrogen bonds with these functional groups, enabling effective retention and high-efficiency separation. For example, amino columns have been used to separate glucocorticoids such as prednisone (PS) and prednisolone (PSL) and finally achieved efficient separation of these polar compounds.^{78,79} For steroids with chiral centers, chiral stationary phases are ideal for enantiomeric separation. For instance, in separating chiral molecules such as dexamethasone and betamethasone in urine and plasma, the Lux i-Cellulose-5 chiral column has been shown to achieve complete separation.⁸¹

Traditional one-dimensional liquid chromatography (1D-LC) often struggles to achieve sufficient resolution when separating structurally similar steroids and their metabolites with close polarity, which limit its ability to meet the demands of highly sensitive and selective analysis.^{75,79,80,82,83} Semi-preparative two-





Table 3 Summary of purification and IRMS techniques

| Sample | Sample extraction and purification | Methods | LC column | IRMS column | Ref. | Cite |
|--|---|---------|--|-------------|------|------|
| Urine (¹³ C-T) | Hydrolysis → LLE → derivatization → 1D-HPLC | GC-IRMS | Zorbas C18 (250 mm × 4.6 mm × 5 μm) | HP-5MS | 2014 | 67 |
| Urine (steroids in musk) | Hydrolysis → LLE → 1D-HPLC | GC-IRMS | Zorbas C18 (250 mm × 4.6 mm × 5 μm) | HP-5MS | 2017 | 98 |
| Urine (T and metabolites) | SPE → hydrolysis → LLE → SPE → derivatization → LLE → Offline-2D-HPLC | GC-IRMS | 1D: Shield RP18 (250 mm × 4.6 mm × 5 μm) 2D: Shield RP18 (250 mm × 4.6 mm × 5 μm) | HP ultra 1 | 2017 | 83 |
| Urine (5bAdiol, 5aAdiol) | LLE → hydrolysis → LLE → 1D-HPLC | GC-IRMS | ACE C18 (250 mm × 4.6 mm × 5 μm) | HP-5MS | 2017 | 70 |
| Urine (8 steroids) | SPE → LLE → hydrolysis → LLE → IAC → derivatization → SPE | GC-IRMS | — | DB-17MS | 2019 | 90 |
| Urine (9 steroids) | SPE → hydrolysis → LLE → online-2D-HPLC | GC-IRMS | 1D: Eclipse XDB phenyl (150 mm × 0.46 mm × 3.5 μm) 2D: Eclipse XDB phenyl (150 mm × 0.46 mm × 3.5 μm) | DB-5MS | 2020 | 77 |
| Urine (11 steroids including T and metabolites, B, BM, 19NA) | SPE → hydrolysis → LLE → derivatization → online-3D-HPLC | GC-IRMS | 1D: Poroshell 120 EC-C18 (100 mm × 4.6 mm × 2.7 μm) 2D: Poroshell 120 EC-CN (50 mm × 4.6 mm × 2.7 μm) 3D: Poroshell 120 EC-C18 (50 mm × 4.6 mm × 2.7 μm) | DB-17 MS | 2021 | 84 |
| Urine (T and metabolites) | LLE → hydrolysis → LLE → 1D-HPLC | GC-IRMS | ACE C18 (5 μm, 4.6 × 250 mm) | HP-5MS | 2022 | 68 |
| Urine (19NA) | SPE → hydrolysis → LLE → online-2D-HPLC | GC-IRMS | 1D: Eclipse XDB-phenyl (250 mm × 4.6 mm × 5 μm) 2D: Zorbas C18 (250 mm × 4.6 mm × 5 μm) | DB-5MS | 2020 | 88 |
| Urine (19NA, 19NE) | Hydrolysis → LLE → offline-2D-HPLC | GC-IRMS | 1D: Ascentis phenyl column (150 mm × 4.6 mm × 5 μm) 2D: ACE C18 (250 mm × 4.6 mm × 5 μm) | HP-5MS | 2021 | 82 |
| Urine (19NA, 19NE) | SPE → hydrolysis → LLE → online-2D-HPLC | GC-IRMS | 1D: Eclipse XDB-phenyl (250 mm × 4.6 mm × 5 μm) 2D: Zorbas C18 (250 mm × 4.6 mm × 5 μm) | DB-5MS | 2023 | 76 |
| Urine (19NA, 19NE) | SPE → hydrolysis → LLE → derivatization → online-3D-HPLC | GC-IRMS | 1D: Poroshell 120 EC-C18 (100 mm × 4.6 mm × 2.7 μm) 2D: Poroshell 120 EC-CN (50 mm × 4.6 mm × 2.7 μm) 3D: Poroshell 120 EC-C18 (50 mm × 4.6 mm × 2.7 μm) | DB-17 MS | 2024 | 85 |
| Urine (glucocorticoids) | LLE → hydrolysis → LLE → 1D-HPLC | GC-IRMS | ACE C18 (250 mm × 4.6 mm × 5 μm) | HP-5MS | 2015 | 86 |
| Urine (PS, PSL) | Hydrolysis → LLE → offline-2D-HPLC | GC-IRMS | 1D: ACE C18 (250 mm × 4.6 mm × 5 μm) 2D: ACE C18 amide (250 mm × 4.6 mm × 5 μm) | HP-5MS | 2019 | 79 |
| Urine (PS, PSL) | SPE → hydrolysis → LLE → online-2D-HPLC | GC-IRMS | 1D: ACE C18 (250 mm × 4.6 mm × 5 μm) 2D: Eclipse XDB-phenyl (250 mm × 4.6 mm × 5 μm) | DB-5MS | 2020 | 80 |
| Urine (PS, PSL) | SPE → hydrolysis → LLE → online-2D-HPLC | GC-IRMS | 1D: ACE C18 (250 mm × 4.6 mm × 5 μm) 2D: Eclipse XDB-phenyl (250 mm × 4.6 mm × 5 μm) | DB-5MS | 2024 | 74 |

Table 3 (Contd.)

| Sample | Sample extraction and purification | Methods | LC column | IRMS column | Ref. | Cite |
|------------------------------|---|-------------------|--|-----------------------|------|------|
| Urine (6 α -OH-Adion) | SPE \rightarrow hydrolysis \rightarrow LLE \rightarrow online-2D-HPLC | GC-IRMS | 1D: Zorbax Extend-C18 (250 mm \times 4.6 mm \times 5 μ m) 2D: Gemini C6-Phenyl (150 mm \times 4.6 mm \times 5 μ m) | DB-17 MS | 2014 | 75 |
| Urine (EpiA-S) | LLE \rightarrow 1D-HPLC | GC-IRMS | Hypersil Gold C18 (150 mm \times 4.6 mm \times 5 μ m) | DB-17 MS | 2020 | 71 |
| Urine (EpiA-S) | LLE \rightarrow hydrolysis \rightarrow LLE \rightarrow derivatization \rightarrow LLE | 2D-GC-IRMS | — | Optima 1 DB-17 MS | 2020 | 101 |
| Urine (10 steroids) | SPE \rightarrow LLE \rightarrow hydrolysis \rightarrow derivatization | 2D-GC-IRMS | — | Optima 1 DB-17 MS | 2018 | 100 |
| Serum (A, EpiA) | PP \rightarrow LLE \rightarrow derivatization \rightarrow 1D-HPLC | GC-IRMS | Eclipse XDB-C18 (150 mm \times 4.6 mm \times 5 μ m) | DB-17 MS | 2024 | 69 |
| Urine (Bo, BoM) | Hydrolysis \rightarrow LLE \rightarrow 1D-HPLC \rightarrow derivatization | Online-LC-GC-IRMS | ACE C18 (250 mm \times 4.6 mm \times 5 μ m) | Silica capillary tube | 2014 | 107 |
| Standards (5 steroids) | — | LC-IRMS | — | Xbridge BEH300C4 | 2014 | 104 |
| Standards (steroids) | — | LC-IRMS | — | Sachtopore-RP | 2024 | 105 |
| Urine (4 steroids) | SPE \rightarrow hydrolysis \rightarrow LLE \rightarrow derivat. \rightarrow online-2D-HPLC \rightarrow hydrolysis \rightarrow SPE | LC-IRMS | 1D: Poroshell 120 EC-C18 (100 mm \times 4.6 mm \times 2.7 μ m) 2D: Poroshell 120 EC-CN (50 mm \times 4.6 mm \times 2.7 μ m) | Sachtopore-RP | 2024 | 87 |

dimensional liquid chromatography (2D-LC) and multi-dimensional liquid chromatography (MD-LC) have emerged as powerful tools for steroid purification, significantly enhancing peak capacity and resolution by integrating orthogonal separation mechanisms.^{74,76–78,84–88} The core principle of 2D-LC is the use of two chromatographic columns with different separation mechanisms, such as a combination of reversed-phase chromatography with hydrophilic interaction chromatography (HILIC) or ion exchange chromatography (IEC). Fractions separated in the first dimension are automatically collected and transferred to the second dimension for further separation. This orthogonal approach effectively removes matrix interferences and enables the separation of structurally similar steroid stimulants.⁸⁹ For example, Lalonde *et al.* developed an automated 2D-HPLC method for the purification of steroids in urine samples. Using an XDB Phenyl column in the first dimension and a reversed-phase C18 column in the second dimension, the method achieved efficient separation anabolic steroids and their metabolites, including 9 urinary steroids such as T and DHEA.⁷⁷ MD-LC builds upon the foundation of 2D-LC by incorporating more than two dimensions and combining multiple separation mechanisms. This approach allows for highly efficient purification of trace steroid stimulants from complex samples. For instance, Honesova *et al.* and Polet *et al.* established a three-dimensional semi-preparative LC purification method for the analysis of T and its metabolites in urine samples as well as the purification of nandrolone metabolites.^{84,85} This method used two six-port switching valves to enable two independent, non-interfering loops for multidimensional chromatographic collection and rapid sample cleanup. While multidimensional LC systems offer higher peak capacity and resolution, these systems are inherently more complex and require more professional operating skills and method development experience. To address these challenges, immunoaffinity chromatography (IAC) has been developed as an alternative sample preparation technique for gas chromatography-combustion-isotope ratio mass spectrometry (GC-C-IRMS) analysis.⁹⁰ However, HPLC methods has its advantages in stability and purification quality, so it is still the preferred method for precise analytical workflows.

4. Advances in IRMS techniques

IRMS can distinguish endogenous steroids from exogenous synthetic steroids by measuring the CIRs, providing robust evidence for doping detection.⁹¹ To enhance the interpretability of isotope ratio data, CIRs are typically reported using the delta notation ($\delta^{13}\text{C}$), expressed in per mil (‰) relative to the international standard Vienna Pee Dee Belemnite (VPDB).⁹² The delta value is calculated using the formula: $\delta^{13}\text{C} = [(R_{\text{sample}}/R_{\text{standard}}) - 1] \times 1000$, where R is the ratio of $^{13}\text{C}/^{12}\text{C}$ in the sample or standard.⁹³ Recent updates in reference materials for $\delta^{13}\text{C}$ calibration should be noted. The certified reference material VPDB has undergone re-evaluation, which slightly affects the absolute isotope ratio scales and reinforces the importance of precise two-point calibration using well-characterized reference gases.⁹⁴ In anti-doping analysis, the



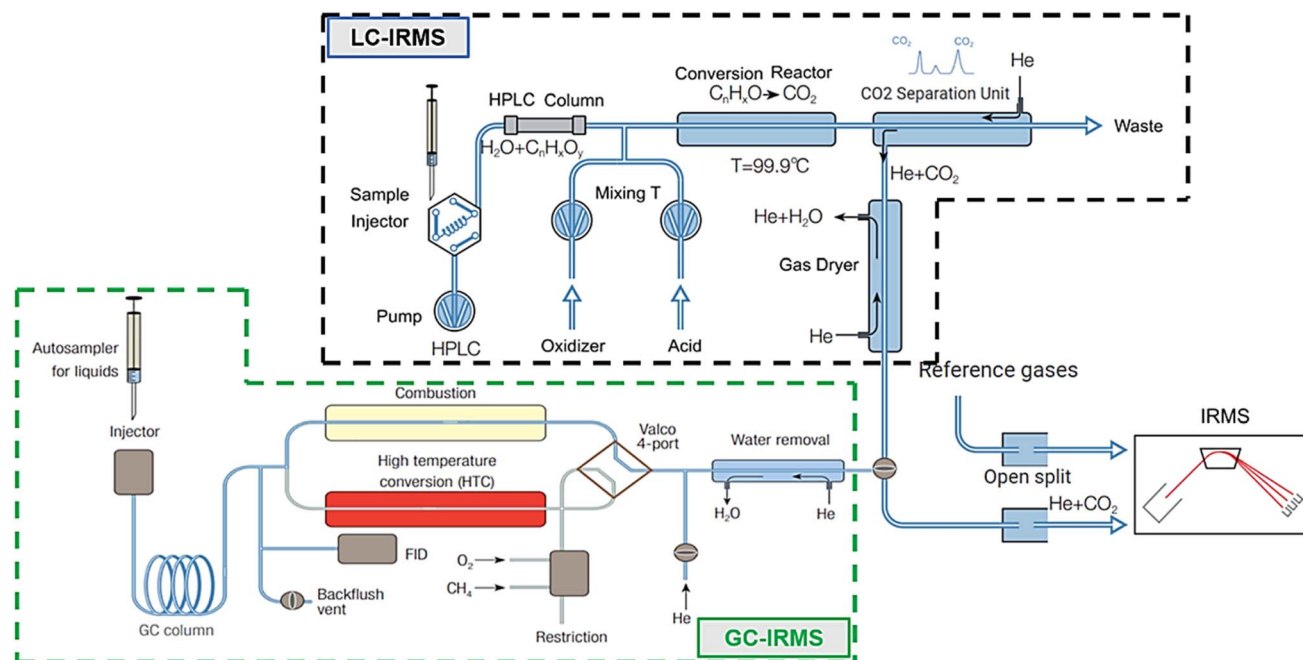


Fig. 2 Principles of the GC-IRMS and LC-IRMS interfaces.^{96,102}

differentiation between endogenous and exogenous steroids relies not on absolute $\delta^{13}\text{C}$ values, but on Δ -values, defined as the isotopic difference between a target compound (TC) and an endogenous reference compound (ERC): $\Delta = \delta^{13}\text{C}_{\text{ERC}} - \delta^{13}\text{C}_{\text{TC}}$. WADA recommends using a Δ -value greater than 3‰ to indicate exogenous origin, due to the small but consistent differences in carbon isotope ratios between synthetic steroids (typically $< -27\text{‰}$) and endogenous compounds (usually between -16‰ and -26‰).⁹⁵ According to the sample introduction methods, IRMS is mainly classified into GC-IRMS and liquid-phase IRMS (LC-IRMS)⁹⁶ (Fig. 2). GC-IRMS combines gas chromatography (GC) with IRMS and is suitable for analyzing volatile or derivatized compounds. In doping control, target compounds often require derivatization to enhance volatility for GC separation. The separated compounds are then introduced into a combustion or high-temperature pyrolysis furnace, where they are converted into simple gases (e.g., CO_2 , H_2 , O_2) for isotopic ratio measurements by IRMS.⁹⁷ For example, Wang *et al.* utilized GC-C-IRMS to investigate the effects of musk intake on anti-doping controls. They found that varying doses of musk altered steroid profiles and CIRs, potentially leading to abnormal analytical findings (AAFs). This study was the first to confirm that musk intake could result in positive doping test results.⁹⁸ In terms of other element determination, such as H and O isotopes, environmental factors during sample pretreatment and separation can lead to significant isotope fractionation. Therefore, it requires very delicate operations during sample pretreatment to ensure the accuracy and reproducibility of H and O isotopes.⁹⁹

Two-dimensional gas chromatography (2D-GC) combined with IRMS enables higher-resolution analysis of steroids in complex samples. By employing two chromatographic columns with different separation mechanisms (e.g., polarity and non-

polarity), 2D-GC significantly enhances separation efficiency and analytical capability.^{100,101} Putz *et al.* have reported the application of 2D-GC-IRMS for steroid analysis.¹⁰⁰ The first GC dimension used a low-polarity column (Optima 1, 30 m length, 0.25 mm ID) for preliminary purification of urinary steroids, while the second dimension employed a moderately polar column (DB-17 MS, 30 m length, 0.25 mm ID) for compound separation. This method was applied to 74 routine doping control samples to establish population-based thresholds, reducing manual workload and time on sample preparation. Epiandrosterone (EpiA), which is sulfated in the body, requires enzymatic hydrolysis followed by acidic solvolysis for analysis, making its sample preparation highly complex. In order to simplify the sample preparation of sulfonated compounds, Piper *et al.* tested the enzymatic cleavage of arylsulfatase from *Pseudomonas aeruginosa* and further simplified the pretreatment process by using MD-gas chromatography to ensure the peak purity required for CIRs.¹⁰¹ While GC-IRMS is a mature technique with high sensitivity and excellent reproducibility, derivatization steps can introduce isotope fractionation effects, potentially affecting measurement accuracy. Additionally, thermally unstable steroid compounds may degrade during derivatization or GC separation, limiting the applicability of this method.

LC-IRMS overcomes the limitations of GC-IRMS, as it eliminates the need for derivatization, allowing direct analysis of non-volatile and thermally unstable compounds.¹⁰² The LC-IRMS workflow begins with LC separation, where different steroid compounds in complex samples are separated based on their physicochemical properties. The eluate is then introduced into an online interface that converts the target analytes into gas forms detectable by IRMS. Common interface technologies

include wet chemical oxidation (WCOC) and high-temperature conversion (HTC).

WCOC method employs strong oxidizing agents (*e.g.*, sodium persulfate) to oxidize organic compounds into CO₂, while HTC pyrolyzes compounds at high temperatures into CO₂ and other small gas molecules. The generated CO₂ is subsequently introduced into the IRMS for measurement of the ¹³C/¹²C isotope ratio.¹⁰³ Zhang *et al.* developed a novel high-temperature liquid chromatography coupled with photodiode array detection and isotope ratio mass spectrometry (HT-LC/PDA/IRMS) for the CIRs analysis of non-derivatized steroid hormones. Using a C4 column at high temperatures with ultrapure water as the sole mobile phase, the method achieved complete separation of a mixture of five steroids, including T and epitestosterone (EpiT). The applicability of the method was validated using testosterone-containing ointments.¹⁰⁴ Recently, Honesova *et al.* evaluated six analytical columns for their suitability in separating steroids using similar conditions. They identified ZirChrom-PBD and Sachtopore-RP as the only two columns suitable for steroids separation.¹⁰⁵ Subsequently, the team employed a 2D-LC purification method, combining Sachtopore-RP columns with conditions up to 200 °C and pure water as the mobile phase. This approach successfully measured the CIRs of steroids in urine, enabling differentiation between endogenous and synthetic steroids.⁸⁷ Since LC-IRMS converts analytes into CO₂ *via* wet oxidation and accurately determines CIRs using water as the mobile phase, the current design of LC interfaces imposes strict limitations on flow rates and particularly the composition of the mobile phase. Addressing these limitations will require the development of novel, high-efficiency interface technologies combined with advanced multi-dimensional chromatography methods to achieve precise analysis of trace steroid stimulants in complex matrices.^{106,107} Additionally, coupling IRMS with high-resolution mass spectrometry (HRMS) has emerged as a promising approach. This combination provides both structural and isotopic ratio information, offering more comprehensive evidence for doping detection and enhancing the reliability of analytical results.¹⁰⁸

5. Conclusion

This review provides a summary of the latest research advancements in the purification, analysis, and application of IRMS techniques for steroid doping detection. In the field of purification and analysis, the combination of separation techniques such as HPLC and GC with mass spectrometry has significantly improved the sensitivity and specificity. These techniques enable rapid separation of targets from complex biological samples and differentiate between endogenous and exogenous steroids. With the optimization of sample preparation, particularly the combined use of LLE and SPE, detection efficiency has been further enhanced. Meanwhile, the application of IRMS techniques (*e.g.*, GC-C-IRMS and LC-IRMS) has provided a scientific basis for distinguishing between endogenous and exogenous steroids through the measurement of CIRs, thus further strengthening the ability to prevent detection

evasion. However, given the complex structures of novel steroid stimulants and increasingly abuse strategies, existing detection methods still require further optimization, particularly in the areas of rapid screening, sensitivity enhancement, and the identification of unknown stimulants. Future advancements include the development of innovative sample preparation materials, such as molecularly imprinted polymers, for detecting new synthetic steroids. Additionally, integrating AI-assisted mass spectrometry for enhanced screening capabilities, optimizing mass spectrometry parameters and exploring new biomarkers will help improve the sensitivity for detecting micro-dose stimulants. Moreover, enhancing international collaboration and establishing standardized testing protocols and databases are essential for advancing the global standardization of doping detection technologies.

Data availability

Data will be made available on request.

Author contributions

Zhongquan Li: research concept, literature review, data collection, data analysis and interpretation, statistical analysis, writing of the manuscript; Jiahui Cheng: literature review, data analysis and interpretation, visualization; Chaomin Zhao: supervision, writing-review, project administration; Qing Chen: writing-review, project administration (corresponding author). Bing Liu: research concept, supervision, writing-review, project administration (corresponding author); Peijie Chen: supervision, writing-review, project administration (corresponding author).

Conflicts of interest

The authors declare that they have no known competing financial interests.

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