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## Excited state deactivation in phytochemical flavonoids: astragalin and kaempferol†

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Femtosecond transient absorption experiments and *ab initio* and nonadiabatic dynamics (NAD) simulations have helped to explicate the photophysical dynamics of two ubiquitous members of the flavonoid family, astragalin and kaempferol. Differences between their excited state deactivation mechanisms have been observed and explained. Astragalin exhibits an ultrashort ( $\sim 10$  ps)  $S_1$  lifetime, whereas that of kaempferol extends approximately twice as long ( $\sim 20$  ps), explained by the systems' relative access to a  $S_1/S_0$  conical intersection following excited state intramolecular proton transfer. These findings expound the unexpected role played by sugar substituents in photoprotective flavonoids and justify the use of similar natural derivatives for UV photothermal materials.

## Introduction

Nature continues to inspire many of humanity's technological advancements.<sup>1</sup> Deference to billions of years of evolutionary development may provide the solution to the last century's global issues.<sup>2–4</sup> Active compounds in plants – phytochemicals – have seen use as preventative medicines and nutraceuticals,<sup>5,6</sup> whilst flora has informed the design of biobased materials.<sup>7</sup> Phytochemical photo-protectant molecules are of particular interest for biomimetic applications. Due to their dependence on photosynthesis, plants are especially sensitive to the effects of solar radiation and have therefore developed effective and sensitive defensive mechanisms.<sup>8</sup>

Amongst the multitude of natural products which have found use as pharmaceuticals is a group of phenolic species known as flavonoids.<sup>9</sup> Flavonoids are ubiquitous in fruits, vegetables, stems, flowers, and grains.<sup>9</sup> Kaempferol is a natural flavonol, the most abundant subgroup of flavonoids,<sup>9</sup> and has been identified as having antioxidant, antimicrobial, anticancer and neuroprotective properties.<sup>10</sup> The structural features of flavonols, including a  $C_2=C_3$  double bond

conjugated with a 4-oxo group and 3-hydroxyl group on the C ring, and 5,7-dihydroxylation on the A ring (see Fig. 1 for atom and ring labelling), are critical to their biological and antioxidant activities.<sup>11–13</sup>

In this study, we investigate the photochemistry of kaempferol and its phytochemical 3-O-glucoside, astragalin,<sup>14</sup> both experimentally and theoretically. Astragalin, found in a plethora of plant families,<sup>15</sup> retains many structural characteristics commonplace in bioactive flavonoids, and it was therefore judged as a suitable model for other plant-derived systems capable of antioxidative support against UV damage.<sup>16</sup> Astragalin's glycosyl substitution aids its aqueous solubility and may influence its biochemical properties but would not be expected to significantly influence its photochemistry. As pharmaceutical compounds already in use,<sup>14,15,17,18</sup> astragalin and kaempferol uniquely present themselves as models for

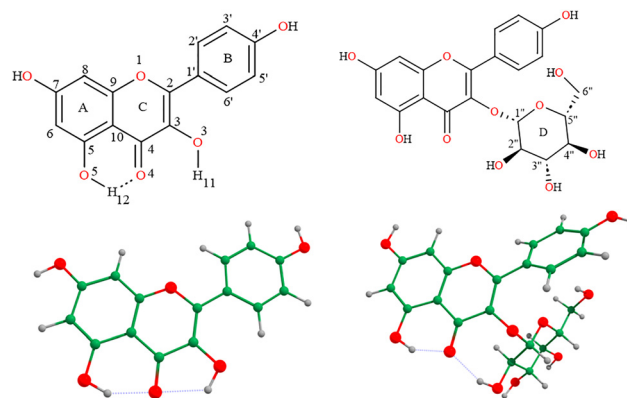


Fig. 1 Structures and atom numbering schema (top) and optimized geometries (bottom) of kaempferol (left) and astragalin (right).

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dual-purpose natural products, whose applications might bridge the divide between purely cosmeceutical (*e.g.*, antioxidant, anti-aging, cell protection) and specialised medical markets.<sup>19</sup>

This study evaluated whether the photochemistry of astragalin was distinct from kaempferol itself, or even from other flavonoids more generally.<sup>20</sup> By investigating both the photochemistry of astragalin and kaempferol in a polar (ethanol, dielectric constant  $\epsilon_r = 25.3^{21}$ ) and nonpolar (1,4-dioxane,  $\epsilon_r = 2.22^{21}$ ) solution, and rationalising our findings using computational chemistry, including exploration of ground and excited state geometries, electronic structure and nonadiabatic dynamics (NAD), we have also assessed their ability to act as photothermal materials.<sup>22</sup>

## Materials and methods

### UV-Visible absorption spectroscopy

UV-Visible absorption spectra of both solution samples were recorded using a Cary 60 UV/Vis spectrophotometer before and after irradiation with a simulated solar spectrum (LCS-100, Spectra-Physics, see Fig. S3, ESI†). Solutions of each molecule were made up to  $\sim 0.5$  absorbance at their UV-visible origin maximum ( $\lambda_{\max}$ ) in a 1 mm path length quartz cuvette for ultrafast experiments.

### Femtosecond transient electronic absorption spectroscopy

A detailed description of the femtosecond transient electronic absorption spectroscopy (fs-TEAS) setup has been described previously; a brief overview of the method used is therefore provided below.

Solutions, as described *supra*, were used as prepared. The 'pump' (photoexcitation) wavelength was set to  $\lambda_{\max}$  for each analyte solution, as extracted from UV-visible absorption spectra (see Fig. 2); the pump power was set to  $\sim 500$   $\mu$ W. Our 'probe' pulse, a white light supercontinuum (WLS) which ranged from  $\sim 320$ – $720$  nm, was generated by focusing  $\sim 0.05$  W of a fundamental 800 nm beam into a translating CaF<sub>2</sub> window. The WLS probe was focused and overlapped with the pump beam inside the 1 mm sample cuvette to maximise observed signal. Pump-probe time delay ( $\Delta t$ ) was controlled by changing the pathlength of the probe pulse using a gold retro-reflector mounted onto a motorised delay stage. Every other pump pulse was blocked by a chopper spinning at 0.5 kHz to directly observe the 'pump-off' ( $I_0$ ) and 'pump-on' ( $I_{pu}$ ) probe absorbance. The resulting change in optical density ( $\Delta OD$ ) was recorded by focusing the probe beam into a fibre-coupled spectrometer (AvaSpec-ULS1650F, Avantes, Apeldoorn, The Netherlands), and calculated as:

$$\Delta OD(\lambda, t) = \log\left(\frac{I_0(\lambda)}{I_{pu}(\lambda, \Delta t)}\right)$$

### Computational details

Stable geometries of astragalin and kaempferol were identified using the conformer-rotamer ensemble sampling tool/

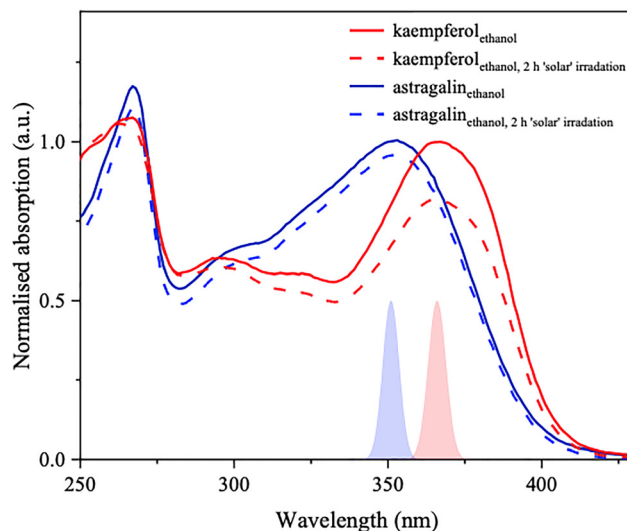


Fig. 2 UV-Visible absorption spectra of kaempferol (red) and astragalin (blue) in ethanol before and after 2 hours' 'solar' irradiation. Shaded plots are approximate profiles of the 'pump' pulses used in fs-TEAS for each solution.

version 6.4.0 (CREST).<sup>23,24</sup> CREST adopts a meta-dynamic (MTD) scheme for generating conformational ensembles, directly sampling at a semiempirical quantum mechanical (SQM) level. An energy window of 8.0 kcal mol<sup>-1</sup> was applied to screen conformational energies at the semiempirical level. This technique utilised the GFN2-xTB method.<sup>25</sup> A total of 649 conformers were identified for astragalin, while five were found for kaempferol: kaempferol lacks a glycoside moiety, resulting in fewer conformational degrees of freedom and thereby affording a more straightforward structural analysis. The 100 most stable conformers of astragalin and all conformers of kaempferol were subjected to geometry optimization using density functional theory (DFT) with the B3LYP functional, cc-pVDZ basis set, and D3 dispersion correction.<sup>26,27</sup> Subsequently, the 20 most stable DFT-optimized conformers of astragalin and all DFT-optimized conformers of kaempferol underwent a final optimization using the RI-MP2 theoretical model and cc-pVDZ basis set. The RI-ADC(2) and time dependent density functional theory (TD-DFT) methodologies (using  $\omega$ B97XD functional<sup>28</sup>) were used to predict the excitation energies and oscillator strengths associated with the four lowest energy singlet-singlet excitations, for the most stable identified ground state conformers of astragalin and kaempferol in both the gas and implicit solution phases. All calculations used the cc-pVDZ basis set. The polarizable continuum model (PCM)<sup>29</sup> and COSMO models were employed to incorporate implicit solvation effects when required.

The MP2/ADC(2)<sup>30–32</sup> calculations were done using the Turbomole suite (version 6.3 and 6.6) and DFT/TD-DFT calculations were performed using Gaussian 16.<sup>33</sup> We investigated nonradiative relaxation mechanisms and PESs by mapping reaction paths with relaxed scan and LIIC and deriving potential energy curves at the cost-effective ADC(2) theoretical



level.<sup>23–25</sup> In the relaxed scan, the O<sub>5</sub>–H bond length was selected as the reaction coordinate, whose value varies, in 0.06 Å intervals, from 0.99–1.72 Å and 0.99–1.78 Å for astragalín and kaempferol, respectively. The O<sub>5</sub>–H bond length was frozen while allowing the remaining degrees of freedom to be optimized. For both systems, the minimum energy S<sub>1</sub>/S<sub>0</sub> CI was optimized based on CASSCF(6,6)/6-31G\* method, implemented in the Gaussian 16 program.<sup>33</sup> The active space for these calculations comprised six electrons distributed across six molecular orbitals, including three occupied and three virtual orbitals (Fig. S14, ESI†); this active space was selected due to its reported accuracy and feasible computational cost.<sup>34–38</sup> To explore the implicit solvent effects (*i.e.*, of ethanol), the polarizable continuum model (PCM), implemented in Gaussian 16, and the COSMO model in Turbomole (version 6.6) were employed. To connect the S<sub>1</sub> optimized geometry to the CI structure, we used a Python script to generate 16 intervals to determine points on the LIIC potential energy surfaces.

UV-Visible absorption spectra and nonadiabatic dynamics simulations were performed using a mixed quantum-classical trajectory method incorporating decoherence correction ( $\alpha = 0.1$  Hartree), based on TD- $\omega$ B97XD/cc-pVDZ theory and implemented *via* Newton-X (v2.4)<sup>39,40</sup> interfaced with Gaussian 16.<sup>33</sup> The suitability of this computational framework for exploring excited-state deactivation processes has been validated in several prior studies.<sup>34,41–43</sup> To initiate the simulations, 500 initial conditions were sampled from a Wigner distribution (0 K) of normal modes from a harmonic oscillator around the ground-state minimum. The resulting nuclear ensemble was then employed to compute excitation energies and transition dipole moments, enabling simulation of the UV absorption spectrum for the bright S<sub>1</sub> (<sup>1</sup> $\pi\pi^*$ ) state.

Nonadiabatic dynamics simulations were conducted for the ground state (S<sub>0</sub>) and the two lowest excited singlet states (S<sub>1</sub> and S<sub>2</sub>), initiating the trajectories from S<sub>1</sub> due to its dominant role in UV absorption for both molecular systems. An additional set of 500 initial conditions was generated for S<sub>1</sub> using a Lorentzian line broadening with  $\delta = 0.05$  eV. Electronic propagation was carried out using the local diabaticization method.<sup>44,45</sup> When state hopping occurred, typically across a finite energy gap, total energy conservation was ensured by rescaling momenta along the direction of the nonadiabatic coupling vector.<sup>46,47</sup> In addition, a bond length criterion of O<sub>4</sub>–H<sub>12</sub>  $\leq 1.0$  Å, implemented *via* a custom Python script, was used to define proton transfer (PT) (see Fig. S10, ESI†).

Initial conditions for the nuclear ensemble method<sup>48</sup> were sampled within the spectral windows of  $3.5 \pm 0.1$  eV and  $3.7 \pm 0.1$  eV respectively for astragalín and kaempferol, as illustrated in Fig. S9 (ESI†). A total of 103 trajectories for astragalín and 92 trajectories for kaempferol were simulated. Each trajectory was propagated for a maximum of 1500 fs with a nuclear time step of 0.5 fs. Due to limitations inherent to TD-DFT, trajectories were terminated either when the energy gap between S<sub>1</sub> and S<sub>0</sub> dropped below 0.15 eV, or when the maximum simulation time was reached. Energy conservation was satisfactorily maintained throughout the simulations.

Solvent effects on the absorption spectra were investigated using the polarizable continuum model (PCM, water) as implemented in Gaussian 16. To characterize out-of-plane ring distortions during dynamics, Cremer–Pople puckering parameters<sup>49</sup> were calculated using the PLATON program.<sup>50</sup>

## Results

### UV-Visible absorption spectroscopy

As is characteristic of other flavonoid systems,<sup>51</sup> two distinct absorption bands (at  $\sim 250$  and  $\sim 350$  nm) are present in the UV region for kaempferol and astragalín, both in polar (ethanol, see Fig. 2) and nonpolar (1,4-dioxane, see Fig. S1, ESI†) solutions. Extinction coefficients  $\epsilon$  for kaempferol in ethanol at 366 nm and astragalín in ethanol at 351 nm were determined as  $22\,100 \pm 170$  and  $17\,320 \pm 30$  mol<sup>−1</sup> dm<sup>3</sup> cm<sup>−1</sup>, respectively (see Fig. S2, ESI†).

Photostability under a simulated solar spectrum was measured by recording UV-visible absorption spectra after 2 hours' exposure under the equivalent of 'one Sun' (shown in ethanol in Fig. 2; the simulated solar spectrum used is shown in Fig. S3, ESI†). Extremely high photostability is observed for astragalín, which is somewhat reduced for kaempferol; as is commonly reported in literature for UV filters, an area under curve index (AUCI) between 280 and 400 nm of 94% was calculated for astragalín and of 87% for kaempferol (see ESI,† Section S1.1), classifying each as photostable.<sup>52</sup>

### Ground state geometry determination

The most stable identified structures of ethanolic kaempferol and astragalín are shown in Fig. 1. As shown, the optimized ground state structures include the O<sub>5</sub>H<sub>12</sub> ··· O<sub>4</sub> hydrogen bond in both systems. Rotation of the C<sub>5</sub> hydroxyl group (O<sub>5</sub>H<sub>12</sub>) to face away from the C<sub>4</sub> carbonyl, removing the hydrogen bonding interactions between these moieties, destabilises kaempferol and astragalín by  $\sim 0.36$  and  $\sim 0.32$  eV, respectively (see ESI,† Sections S1.2 and S2.3 for further details). Similarly, in kaempferol, a geometry with both C<sub>3</sub> and C<sub>5</sub> hydroxyl groups facing the carbonyl oxygen is stabilized by  $\sim 0.18$  eV relative to a conformer with the C<sub>3</sub> hydroxyl oriented away. This appears to result from the loss of the hydrogen bonding interaction between the O<sub>3</sub>H<sub>11</sub> hydroxyl group and the C<sub>4</sub> carbonyl, and the diminished overlap of p orbitals associated with rings B and C: the steric hindrance between O<sub>3</sub>H<sub>11</sub> and the proton at C<sub>6</sub>/C<sub>2</sub>' causes a dihedral rotation of ring B  $\sim 40.2^\circ$  out of the plane of ring C.

### Vertical transition energies

Vertical transition energies and oscillator strengths at the ADC(2) level for the optimized structures of astragalín and kaempferol *in vacuo*, between the ground and four lowest-lying singlet excited states (S<sub>1</sub>–S<sub>4</sub>), are presented in Table 1. Selected valence molecular orbitals (MOs) of these two molecules are also presented in Fig. 3. For astragalín, the S<sub>1</sub>  $\leftarrow$  S<sub>0</sub> electronic transition is assigned as <sup>1</sup> $\pi\pi^*$ , originating from the



**Table 1** Vertical transition energies and oscillator strengths for lowest lying electronic transitions in astragalín and kaempferol *in vacuo* and in implicit ethanol. ADC(2) results were determined based on the MP2 optimized ground state geometries, and TD-DFT results were determined based on the  $\omega$ B97XD optimized ground state geometries for each system

| Transition energy/eV                                                |                               | ADC (2)/cc-pVDZ |               | TD- $\omega$ B97XD/cc-pVDZ |
|---------------------------------------------------------------------|-------------------------------|-----------------|---------------|----------------------------|
| Molecule                                                            | Excited state                 | Gas phase       | COSMO/ethanol | PCM/ethanol                |
| Astragalín (experimental $\lambda_{\text{max}}$ : 351 nm (3.53 eV)) | S <sub>1</sub> ( $\pi\pi^*$ ) | 3.84 (0.3450)   | 3.76 (0.4482) | 3.98 (0.5444)              |
|                                                                     | S <sub>2</sub> ( $n\pi^*$ )   | 4.12 (0.0038)   | 4.14 (0.1468) | 4.52 (0.0955)              |
|                                                                     | S <sub>3</sub> ( $\pi\pi^*$ ) | 4.21 (0.1774)   | 4.16 (0.0003) | 4.75 (0.0049)              |
|                                                                     | S <sub>4</sub> ( $\pi\pi^*$ ) | 4.58 (0.0956)   | 4.49 (0.1413) | 4.94 (0.1109)              |
| Kaempferol (experimental $\lambda_{\text{max}}$ : 366 nm (3.39 eV)) | S <sub>1</sub> ( $\pi\pi^*$ ) | 3.61 (0.5306)   | 3.56 (0.5417) | 3.66 (0.6726)              |
|                                                                     | S <sub>2</sub> ( $\pi\pi^*$ ) | 4.29 (0.1166)   | 4.26 (0.1133) | 4.45 (0.0708)              |
|                                                                     | S <sub>3</sub> ( $n\pi^*$ )   | 4.36 (0.0000)   | 4.31 (0.0034) | 4.83 (0.0072)              |
|                                                                     | S <sub>4</sub> ( $\pi\pi^*$ ) | 4.62 (0.0447)   | 4.58 (0.1109) | 4.86 (0.0000)              |

highest occupied molecular orbital to the lowest unoccupied molecular orbital (LUMO  $\leftarrow$  HOMO) (64%) and LUMO  $\leftarrow$  HOMO-1 (30%) single electron transition. The S<sub>2</sub>  $\leftarrow$  S<sub>0</sub> transition corresponds to an optically dark  $^1n\pi^*$  state, dominated by a LUMO  $\leftarrow$  HOMO-7 transition. The third and fourth singlet electronic transitions are also assigned as optically bright  $^1\pi\pi^*$  states, arising from LUMO  $\leftarrow$  HOMO-1 (51%) and LUMO  $\leftarrow$  HOMO-2 (65%) for the S<sub>3</sub>  $\leftarrow$  S<sub>0</sub> and S<sub>4</sub>  $\leftarrow$  S<sub>0</sub> transitions respectively. Similar electronic transition assignments hold for kaempferol, except for the character of S<sub>2</sub> and S<sub>3</sub> swapping, and we therefore neglect further discussion on this system (see ESI,<sup>†</sup> Section S1.3 for more information).

We also determined the four lowest lying electronic transitions of the two molecules at ADC(2) level, considering the effect of an implicit solvent based on the COSMO model,<sup>53</sup> the results of which are present in Table 1. The S<sub>1</sub> ( $^1\pi\pi^*$ )  $\leftarrow$  S<sub>0</sub> electronic transition of both molecules exhibits a slight shift in this implicit model *versus in vacuo*, and the oscillator strength of subsequent excited states changes. Finally, time-dependent density functional theory (TD-DFT) calculations were performed with ethanol as an implicit solvent. Overall, TD-DFT results are consistent with ADC(2) (see Table 1), demonstrating reliability in describing the electronic structures and photophysics of the systems. Our transition assignments using TD-DFT confirm that the S<sub>1</sub> ( $^1\pi\pi^*$ ) state is responsible for UV absorption in both flavonoids. These results also suggest that solvent interactions play only a minor role in modulating the

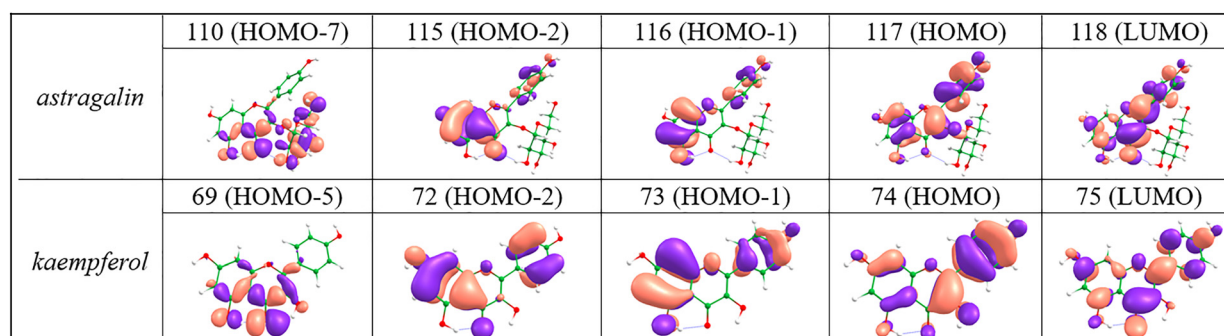
electronic properties of the systems. Optimized geometries of both systems in the ground (at MP2/cc-pVDZ) and S<sub>1</sub> (at ADC(2)/cc-pVDZ) states are presented in Fig. S4 (ESI<sup>†</sup>). Both astragalín and kaempferol exhibit a local S<sub>1</sub> minimum, the properties of which may govern their respective photophysics.

### Femtosecond transient electronic absorption spectroscopy

Following measurements in the steady-state regime, spectroscopy was done on the ultrafast (femtosecond) timescale. Solutions of both astragalín and kaempferol in ethanol and 1,4-dioxane were analysed using fs-TEAS,<sup>54</sup> a method long implemented for studying the time-resolved photochemistry of biological chromophores. Each solution was photo-excited at its respective origin band maximum ( $\lambda_{\text{max}}$ , see Fig. 2). Spectral dynamics following photoexcitation were similar in each system, as were kinetics between solvents. Collected transient absorption (TA) spectra, as shown in Fig. 4 for astragalín and kaempferol in ethanol (and in Fig. S5 in 1,4-dioxane, ESI<sup>†</sup>), were fitted using a global fitting procedure (see ESI,<sup>†</sup> Section S1.4 for instrument response function (IRF), decay associated spectra (DAS), and residuals from fitting);<sup>55</sup> lifetimes extracted from this fitting are shown in Table 2.

### Photophysics: nonadiabatic dynamics (NAD) simulations and conical intersections

To explore the dynamical behaviour of astragalín and kaempferol following photoexcitation, we undertook NAD simulations



**Fig. 3** Selected valence MOs which play a prominent role in the lowest lying electronic transitions of astragalín and kaempferol.





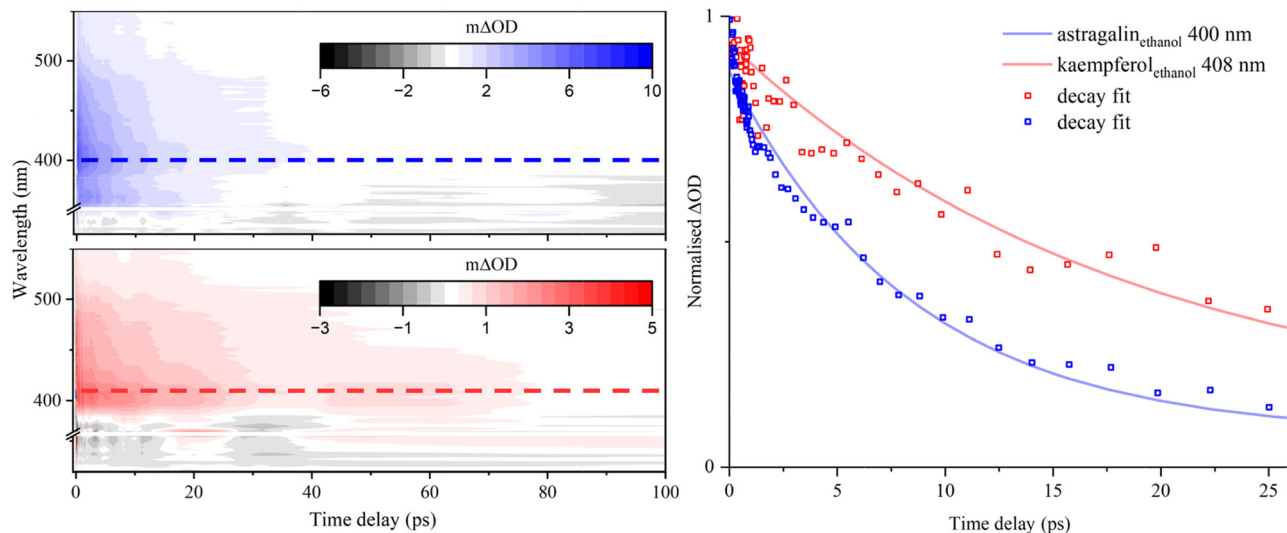


Fig. 4 Collected TA heatmaps (left) and kinetic traces at excited state absorption maximum (right) of astragalin (blue) and kaempferol (red) in ethanol after photoexcitation with 351 nm and 366 nm light, respectively.

Table 2 Lifetimes extracted from global fitting of TA spectra collected following photoexcitation at  $\lambda_{\text{pump}}$

| Lifetime                     | Astragalin      |                  | Kaempferol       |                  |
|------------------------------|-----------------|------------------|------------------|------------------|
| Solvent                      | Ethanol         | 1,4-Dioxane      | Ethanol          | 1,4-Dioxane      |
| $\lambda_{\text{pump}}$ (nm) | 351             | 347              | 366              | 364              |
| $\tau_1$ (ps)                | $8.44 \pm 0.07$ | $11.05 \pm 0.05$ | $18.40 \pm 0.24$ | $16.84 \pm 0.09$ |
| $\tau_2$ (ps)                | > 2500          | > 2500           | > 2500           | > 2500           |
| IRF (FWHM) (fs)              | $136 \pm 6$     | $106 \pm 12$     | $160 \pm 18$     | $109 \pm 8$      |

based on the TD- $\omega$ b97XD/cc-pVDZ theoretical model,<sup>30,56</sup> initialized from the most stable conformer of each molecule. Simulated UV-visible absorption spectra of these two systems (see Fig. S9, ESI<sup>†</sup>), were generated using the nuclear ensemble approach.<sup>48</sup> To add from the outset, preliminary TD-DFT calculations (see ESI<sup>†</sup>, Section S2.5) showed that intersystem crossing (ISC) was unlikely to contribute significantly to the relaxation mechanisms of either system; the energies of triplet states ( $T_1$ – $T_3$ ) corresponding to configurations generated by relaxing the Franck–Condon (FC) geometry into an energetically accessible local  $S_1$  minimum (where ISC would be most feasible) were further than 0.26 eV away from the vertical  $S_1$  energy. NAD simulations were initialized from the  $S_1$  ( $^1\pi\pi^*$ ) excited state, using the decoherence-corrected fewest switches surface-hopping (DC-FSSH) approach.<sup>57,58</sup> Starting in the optically bright  $S_1$  state, 103 trajectories were generated for astragalin, and 92 for kaempferol; selected trajectories are shown in Fig. 5. Trajectories were run up to 1500 fs for astragalin and 2000 fs for kaempferol. For astragalin, 80% of trajectories accessed a  $S_1/S_0$  CI within 1500 fs; however, for kaempferol, only 10% of the trajectories reached a CI before 2000 fs simulation, indicating the excited state lifetime is substantially longer than this. This difference agrees with what is observed experimentally.

In our simulations of astragalin, excited state intramolecular proton transfer (ESIPT) from  $O_5$  to  $O_4$  was observed in 97% of trajectories; the remaining 3% showed ESIPT from the sugar's  $O_2$ -H to  $O_4$ . In kaempferol, ESIPT occurred from  $O_5H_{12}$  in 86% of trajectories, and from  $O_3H_{11}$  in the remaining 14%. To evaluate the ESIPT timescale from NAD simulations in both systems (*i.e.*, the time taken for PT to occur from  $O_5H_{12}$ ), an exponential decay was fitted to  $O_5H_{12}$  proton occupation (see Fig. S10, ESI<sup>†</sup>).<sup>59</sup> From this, ESIPT lifetimes of  $45 \pm 5$  fs for astragalin and  $75 \pm 2$  fs for kaempferol were extracted, the latter agreeing with the results of Ji *et al.*<sup>60</sup> Both lifetimes are within the IRF for our fs-TEAS experiments (see Table 2) and therefore cannot be observed.

The  $S_1$  lifetime was evaluated from NAD trajectories in a similar way to ESIPT (see Fig. S11, ESI<sup>†</sup>),<sup>60</sup> yielding  $984 \pm 67$  fs for astragalin. Conversely, as shown in Fig. 5(b), it is predicted that most of kaempferol's excited population remains trapped in the  $S_1$  excited state; state crossings between  $S_1$  and  $S_0$  were infrequent within 2000 fs. Monitoring astragalin's structure (in trajectories that culminate in  $S_1/S_0$  crossing before 2000 fs) identified key molecular motions that facilitate nonadiabatic coupling. An out-of-plane  $C_2$  distortion, continuing to the end of dynamics, puckers the benzopyran moiety. This ring deformation coordinate steadily increases the ground state energy, until  $S_1/S_0$  state degeneracy is reached, and thus embodies the primary deactivation pathway in astragalin. The distribution of trajectories in the space of Cremer–Pople parameters ( $\theta$ ,  $\Phi$ ) at  $S_1/S_0$  crossings in astragalin, defined by out-of-plane deformation of atoms in the C ring, is shown in Fig. S12 (ESI<sup>†</sup>).<sup>49</sup> The  $C_2$  puckering deformation experienced by astragalin's benzopyran moiety is defined by the parameters  $\theta$  and  $\Phi$ ;<sup>61–63</sup> most  $S_1/S_0$  crossings appear within the range  $270^\circ > \Phi > 180^\circ$ , yielding an envelope conformation for the ring structure.

To investigate the excited state deactivation of astragalin and kaempferol further, we have resolved the potential energy



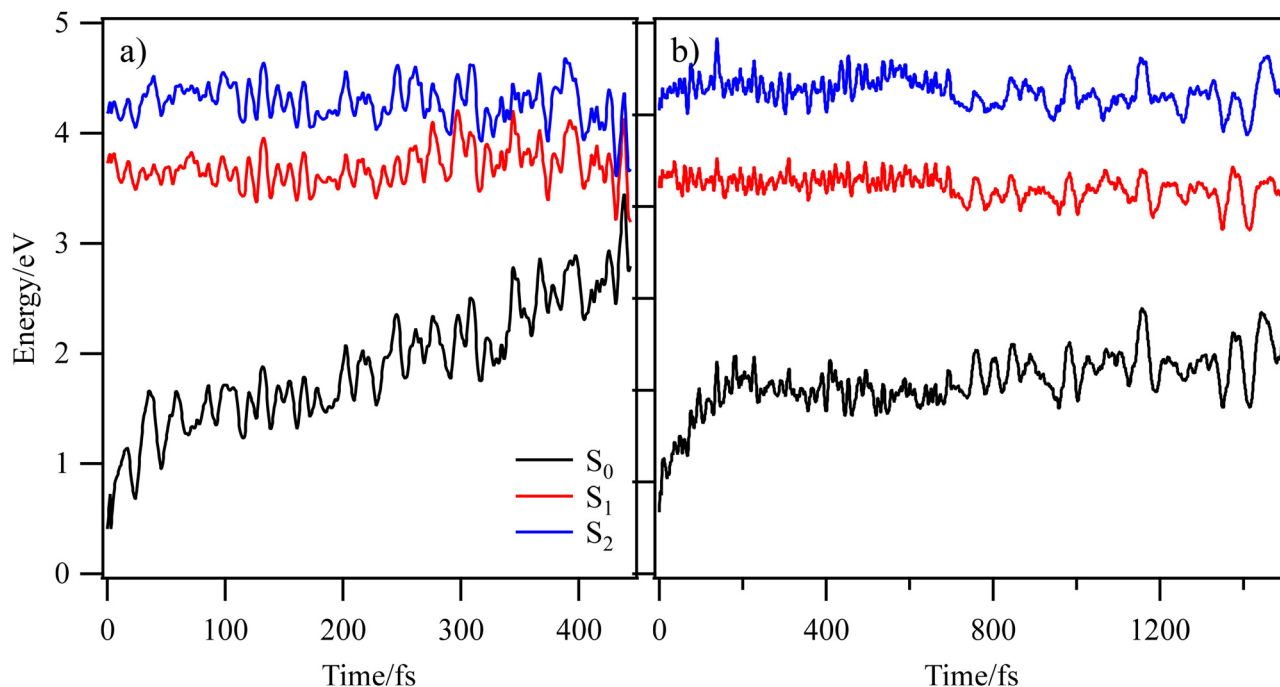


Fig. 5 Energy profiles from selected NAD trajectories of astragalin (a) and kaempferol (b). The black, red and blue curves respectively indicate the ground ( $S_0$ ),  $S_1$  and  $S_2$  state energies.

surfaces (PESs) and located possible CIs in both systems, using the complete active space self-consistent field (CASSCF) theoretical method (Fig. S4, ESI†). As shown in Table 1,  $S_1$  ( $^1\pi\pi^*$ ) has significant oscillator strength; it is therefore responsible for both astragalin and kaempferol's absorption in the UV region. Consequently, it is anticipated to play a governing role in their photophysics. Relaxing each molecule in  $S_1$  from its FC geometry is accompanied by ESIPT from  $O_5H_{12}$  to  $O_4$ . To further explore this, we generated approximate 1D minimum energy paths (MEPs) for  $S_0$  and  $S_1$  through relaxed scans along the proton transfer (PT) coordinate in each system (see Fig. S13, ESI†). As shown, in both systems ESIPT follows a decreasing, and barrierless,  $S_1$  PES. ESIPT should therefore be exothermic, justifying its rapidity.

Taking the final geometries resulting from PT, the minimum  $S_1/S_0$  CIs were determined at the CASSCF (6,6)/6-31G\* theoretical level for astragalin and kaempferol (see Fig. S4, ESI†). The active space for CASSCF level consists of three occupied and three virtual orbitals (see Fig. S14, ESI†). As locating CIs does not guarantee a physically related non-radiative deactivation mechanism for the systems, we have determined  $S_0$  and  $S_1$  PESs of both systems along a linear interpolation of internal coordinates (LIIC), connecting the  $S_1$  minimum of each system to the located minimum energy CI (see Fig. S15, ESI†). For both systems the  $S_1/S_0$  CI is accessible, albeit following dissimilar  $S_1$  PES topographies. Thus, following photoexcitation, each molecule can undergo PT on an ultra-short timescale and, following this, approach a  $S_1/S_0$  CI by slight out-of-plane deformation in the benzopyran ring (at  $C_2$ ). This provides a significant route for internal conversion (IC) by

returning the ring to planarity and finally by back-PT to recover the ground state minimum energy structure.

## Discussion

Firstly, a brief treatise on properties of the UV-visible absorption spectrum of each system is necessary. From the apparent lack of substantial solvatochromism for astragalin's absorption maximum at  $\sim 350$  nm (between in ethanol and in 1,4-dioxane, see Fig. 2 and Fig. S1 respectively, ESI†), we may conclude that the transition corresponding to this band does not generate a dipole-dependent charge transfer (CT) state,<sup>64</sup> but is instead consistent with local excitation (LE), most likely arising around the central conjugated flavonoid moiety. This is supported by transition densities generated for the  $S_1 \leftarrow S_0$  excitation (see ESI,† Section S2.1). Kaempferol's electronic origin band (at  $\sim 365$  nm) is similarly insensitive to solvent polarity (see Fig. 2 and Fig. S1, ESI†). The sugar side chain would normally be expected to act as a spectator, but the significant blue shift of astragalin's absorption maximum with respect to kaempferol refutes this.

As mentioned *supra*, the two absorption bands seen in the UV-visible spectra are ubiquitous in flavonoid systems.<sup>65</sup> From consideration of the above, it is prudent to assume that photodynamics following excitation to similar respective bands in astragalin and kaempferol would be alike. Vertical excitation energies and oscillator strengths predicted by TD-DFT (Table 1) also explain the origin of experimentally observed absorption between the two most significant absorption bands. Rather



than being purely vibronic in origin, nonzero absorption within this region can be attributed to  $S_{2-4} \leftarrow S_0$  transitions, which are (variably) predicted to be moderately bright for ethanolic astragalin and kaempferol. The oscillator strengths associated with these transitions, however, are far lower than those predicted for the corresponding system's  $S_1 \leftarrow S_0$  transition. Exclusively elucidating those relaxation pathways that are followed upon population of  $S_1$  can still therefore offer insight into the overall systems' photophysics.

Under a simulated solar spectrum, astragalin preserves good stability, with little reduction in observed absorption (Fig. 2). This photostability indicates a predominant relaxation pathway which does not lead to permanent chemical change. Kaempferol's reduced photostability may be explained by an alternative dissociative pathway in the excited state (not explicitly identified), which can be accessed while population resides in  $S_1$ .

At first glance, results from ultrafast measurements indicate similar spectral dynamics between flavonoids and solvents. Global fitting of TA data for each system yields two lifetimes, both of which are similar between solvents (Table 2). A broad excited state absorption (ESA) feature, centred at  $\sim 400$  nm, is seen in each solvent, for both molecules. The excited state dynamics of both are kinetically uncomplicated.  $\tau_1$  is assigned to a convolution of fast ESIPT (potentially within our experimental IRF) and continuous geometry relaxation towards and through the  $S_1/S_0$  CI into the ground state, incorporating back-PT to reform the starting material. Astragalin's faster relaxation in ethanol *versus* 1,4-dioxane may be explained by enhanced coupling between its glucoside and the surrounding polar environment effecting more efficient vibrational energy dissipation; this would not be seen in the non-glycosylated kaempferol.  $\tau_2$  captures unrecovered excited state population. The relaxation pathway identified in NAD simulations of astragalin indicates a strong propensity for efficient regeneration. The identified CI, which 80% of trajectories encountered within the simulation time of 1500 fs, afforded passage to a barrierless

path along the ground state PES, featuring steep gradient(s) and 3.48 eV stabilization, back to the FC geometry (see Fig. 6(a)).

From both experimental and theoretical results (*i.e.*, from *ab initio* PESs and NAD simulations), we summarise the photodynamics of astragalin and kaempferol in Fig. 6. Both systems, when initialized at the FC geometry, undergo ultrafast ESIPT (within 45 and 75 fs for astragalin and kaempferol, respectively). Subsequently, both systems experience further conformational change, involving the displacement of  $C_2$ , which leads to a puckered benzopyran moiety. This motion drives the system toward an energetically accessible  $S_1/S_0$  CI, where a nonadiabatic transition to the ground state occurs, followed by back-PT in  $S_0$ . The net effect of this process is conversion of deleterious UV light to heat, a key criterion for UV filters.

A difference in experimental excited state lifetimes was measured for these two systems ( $\sim 10$  ps and  $\sim 20$  ps for astragalin and kaempferol respectively). Guided by our *ab initio* calculations' predictions for  $S_1/S_0$  CIs in both compounds, we seek to explain this. In astragalin, the identified  $S_1/S_0$  CI was predicted to locate lower in energy (0.48 eV) than the vertical  $S_1 \leftarrow S_0$  electronic transition energy, meaning there is no barrier on the  $S_1$  PES between the FC geometry and  $S_1/S_0$  CI (see Fig. 6(a)). In kaempferol, we predict that the  $S_1/S_0$  CI locates slightly higher in energy (0.45 eV) than the vertical  $S_1 \leftarrow S_0$  transition energy (see Fig. 6(b)); the  $S_1/S_0$  CI cannot be accessed in kaempferol without surmounting this barrier. Expectedly, kaempferol's excited state deactivation, although still facile, is significantly slower than in astragalin. Excited population can experience trapping in the  $S_1$  minimum, from which it could conceivably decay to the ground state (although an emission quantum yield of only  $\sim 1\%$  was observed) or undergo excited state photochemistry, accounting for its degradation.

The shorter excited state lifetime of astragalin, seen both experimentally and computationally, is effected by additional structural complexity conferred by the sugar moiety. The

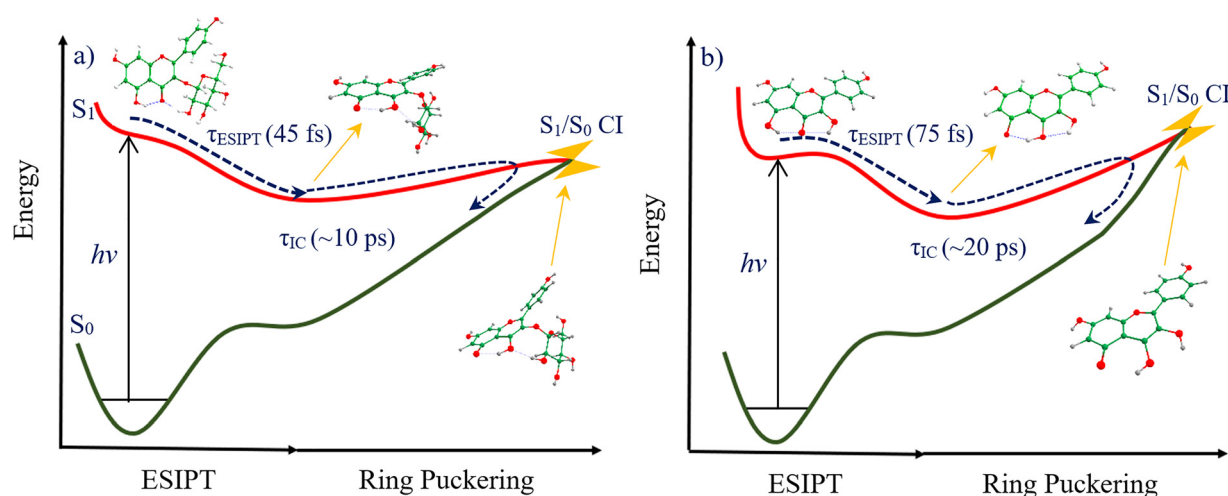


Fig. 6 Summary of the nonradiative decay pathway of astragalin (a) and kaempferol (b) photoexcited to  $S_1$ . Geometries of relevant optimized structures are inset. Time scales corresponding to excited state intramolecular proton transfer (ESIPT) and internal conversion (IC) are also shown.



out-of-plane deformation at C<sub>2</sub> is aided, albeit marginally, by the steric interaction of the glucosyl with the B ring. Judicial substitution of saccharides to flavonoids, or selection of appropriately glycosylated natural derivatives, could therefore enhance the efficiency of nonradiative relaxation in these systems.

## Conclusion

In this study, we have comprehensively investigated the intrinsic photostability and dynamics of astragalin and kaempferol using a combined experimental and theoretical approach. While both compounds have appreciable UV absorption and undergo excited state intramolecular proton transfer, their ultrafast dynamics differ. Specifically, kaempferol's excited state lifetime is approximately twice that of astragalin, evidenced by prolonged excited state absorption. In astragalin, no potential energy barrier has been predicted before a S<sub>1</sub>/S<sub>0</sub> CI, enabling more efficient nonradiative relaxation. In contrast, for kaempferol, a small barrier hinders excited population approaching the S<sub>1</sub>/S<sub>0</sub> CI, explaining slower deactivation. These results are qualitatively reflected in nonadiabatic surface hopping dynamics simulations. The shorter excited state lifetime of astragalin compared to kaempferol arises from additional structural complexity introduced by the sugar moiety, aiding the out-of-plane motion of C<sub>2</sub> (benzopyran).

Our results demonstrate the significant role of sugar in enhancing excited state deactivation of flavonoids, comparable to the effect of sugars on the photophysical behaviour of nucleobases.<sup>36,66–69</sup> These findings enhance our understanding of nonradiative pathways in flavonoids and the role of substitution, offering biomimetic insight into designing more photostable materials. Moreover, as two members of the flavonoid family, our results confirm that these systems fulfil the conditions for model nature-inspired UV filters, due both to their strong UV absorption and ultrafast ground state recovery.

## Author contributions

S. P. and V. G. S. conceived the presented idea. M. H. carried out the fs-TEAS and steady-state experiments. J. E., M. S. and R. O. performed the computational calculations. J. E. undertook the preliminary computational study. M. H., J. E., M. S., R. O. and V. G. S. analysed the results. M. H., J. E., M. S. and R. O. designed the figures. M. H. wrote the manuscript with input from all authors. R. O., S. P. and V. G. S. supervised the project.

## Conflicts of interest

There are no conflicts to declare.

## Data availability

The data that support the findings of this study are available upon request.

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