

RESEARCH ARTICLE

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2020, 4, 950**Pillar[5]arene-based tunable luminescent materials
via supramolecular assembly-induced Förster
resonance energy transfer enhancement†**Nan Song,^{ab} Xin-Yue Lou,^a Hao Yu,^a Paul S. Weiss,^{id} c Ben Zhong Tang^{id} b and
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Pillar[5]arene tetramers with tetraphenylethene cores and different lengths of alkyl ether chains (**H1-4C4P** and **H2-2C4P**) and a 9,10-distyrylanthracene-bridged neutral guest linker (**DSA-G**) are synthesized to fabricate tunable solid-state fluorescent materials through supramolecular assemblies of **DSA-G**⊂**H1-4C4P** and **DSA-G**⊂**H2-2C4P**. Their fluorescence emission is strongly enhanced and their colors are changed upon assembly, which can be ascribed to the supramolecular assembly-induced enhanced emission and Förster resonance energy transfer (FRET) processes between **H1-4C4P** (or **H2-2C4P**) and **DSA-G**. Both ensembles of **DSA-G**⊂**H1-4C4P** and **DSA-G**⊂**H2-2C4P** exhibit thermo and solvent dual-responsive features, while **DSA-G**⊂**H2-2C4P** shows higher sensitivity toward external stimuli as compared to **DSA-G**⊂**H1-4C4P** due to the shorter distance between fluorophores. The use of these fluorescent materials as inks confirms their efficiencies in the solid-state, paving the way for new potential applications of smart optical materials based on supramolecular assemblies.

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Introduction

Photoluminescent materials, especially solid-state materials with tunable wavelengths, have attracted significant attention due to their great potentials in fluorescent probes and sensors, optoelectronic devices, light-emitting diodes, bioimaging agents, molecular optical devices, data storage, and security printing.^{1–3} However, most of their applications have been impeded because of the intrinsic self-quenching properties of most luminescent dyes. Aggregation-induced emission (AIE)^{4,5} and aggregation-induced enhanced emission (AIEE)⁶ have been widely developed since 2001,⁷ which solved the drawback of self-quenching.^{8–10} Meanwhile, supramolecular assembly has proven to be a highly efficient way to fabricate smart luminescent materials with well-ordered architectures bearing typical

luminophores, new physical properties, optional assembly of entities, and reversible stimuli-responsiveness.^{11–15} Nevertheless, luminescent materials with tunable properties adjusted *via* pillar[*n*]arene-based supramolecular interactions both in solution and the solid state are less reported.

Pillar[*n*]arenes (pillararenes),^{16–19} consisting of *n* hydroquinone units linked by methylene bridges at *para* positions, have contributed significantly to the development of supramolecular chemistry and materials science since the first report by Ogoshi *et al.* in 2008.¹⁶ As a new class of important synthetic macrocycles,²⁰ pillararenes possess tailorable structures, facile functionality,^{21–23} and superior host-guest properties.^{24–29} A variety of pillararene-based supramolecular assemblies have been reported, which have been extensively used across a plethora of fields including molecular switches and molecular machines,^{30,31} metal-organic frameworks,³² controlled drug-delivery systems,^{33–38} artificial transmembrane channels,^{39–41} sensors,^{42–48} stimuli-responsive materials,^{49–55} hybrid absorbents,^{56–59} and biomedical applications,^{60–65} including as virus inhibitors and antimicrobials.^{64,65}

Several pillararene-based fluorescent systems combined with tetraphenylethene (TPE)^{66–68} and/or 9,10-distyrylanthracene (DSA)^{69–71} AIEgens have also been reported for potential applications as detectors, drug-delivery systems, *etc.*^{72–77} On the basis of our previous works,^{66,69} we envision that the pillararene-based supramolecular solid-state materials with tunable wavelengths can be constructed from host-guest

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Scheme 1 Schematic representation of the Förster resonance energy transfer (FRET) process facilitated by the combination of pillararene tetramer-based host–guest complexation and supramolecular assembly-induced emission enhancement.

complexation between TPE- and DSA-linked pillararene derivatives and guest compounds. Here, we designed and synthesized TPE-bridged pillararene tetramers and a DSA-bridged neutral guest linker for the construction of supramolecular fluorescent materials with tunable wavelengths through Förster resonance energy transfer (FRET), taking advantage of cooperative supramolecular assembly and host–guest interactions (Scheme 1).

Results and discussion

DSA-G, including two triazole-cyano binding sites bridged by the yellow AIEgen DSA, functions as the energy acceptor.⁶⁸ **H2-2C4P**, possessing shorter alkyl chains compared with another pillararene tetramer (**H1-4C4P**),⁶⁶ was successfully synthesized and used for comparison to demonstrate the role of chain length in host compounds. **H3-4C1P**, **H4-2C1P**, **H5-4CM**, and **H6-2CM** have also been synthesized as control compounds. All the newly synthesized compounds have been fully characterized by ¹H NMR, ¹³C NMR, and MALDI-TOF MS spectroscopy (Fig. S1–S14, ESI[†]). **DSA-G** and **H2-2C4P** were endowed with AIE properties in mixed solvents, as expected (Fig. S15 and S16, ESI[†]). The sharp quenching of fluorescence of **DSA-G** upon increasing the fraction of unfavourable solvents to above 80% is ascribed to sedimentation due to its poor solubility.

The pillar[5]arene ring and triazole-cyano unit can form inclusion complex with a binding affinity of $1.46 \times 10^4 \text{ M}^{-1}$.⁶⁶

In order to develop their applications as supramolecular optical materials, the photophysical properties have been studied in chloroform *via* fluorescence spectroscopy, in which there is efficient host–guest binding affinity between the pillararene cavity and the triazole-cyano site. Ultraviolet-visible (UV-Vis) absorption spectra of **H1-4C4P**, **DSA-G**, and **DSA-G**⊂**H1-4C4P** are shown in Fig. S17 (ESI[†]). The fluorescent emission of **H1-4C4P** excited at 350 nm overlaps with the absorption spectrum of **DSA-G**, conforming to one of the basic principles of FRET donor–acceptor systems. Similarly, **H2-2C4P** can also be used to construct an optimal FRET system, upon assembly with **DSA-G**. As in Fig. S18 (ESI[†]), the conformational simulation of **DSA-G**, **H1-4C4P**, and **H2-2C4P** with energy minimization was simply conducted, respectively. The distances from the center of TPE core to the pillar[5]arene terminals of **H1-4C4P** and **H2-2C4P** is about 14.851 Å and 10.339 Å, respectively, while the distance from DSA core of **DSA-G** to the triazole-cyano binding site is about 15.294 Å. The efficient distance for FRET is *ca.* 1–10 nm, while the distances between host–guest complexes of **DSA-G**⊂**H1-4C4P** and **DSA-G**⊂**H2-2C4P** are *ca.* 3.0 nm and 2.6 nm. After the formation of supramolecular assemblies, the distance might decrease due to the twisted and interactional components, but still within the efficient distance for FRET.

Their fluorescence has been investigated in chloroform (Fig. 1a). Both **H1-4C4P** and **H2-2C4P** exhibited blue fluorescence with emission maxima at 481 nm. After the addition of **DSA-G**, red-shifted fluorescence emission can be detected at 510 nm. Upon continuous addition of **DSA-G** to the solution, their emission maxima underwent a ~25 nm red-shift and the intensity was enhanced dramatically. In the control systems, **DSA-G**⊂**H3-4C1P**, **DSA-G**⊂**H4-2C1P**, and individual **DSA-G** exhibited yellow emission centred at ~530 nm without obvious wavelength shifts.

Furthermore, their detailed fluorescence behaviour along with control experiments have been investigated to test their supramolecular assembly-induced enhancement and FRET. **H1-4C4P** and **H2-2C4P** dissolved in chloroform exhibit weak blue fluorescence with emission maxima at 481 nm after excitation at 350 nm. However, the yellow fluorescence at the emission maximum of 535 nm is strongly enhanced upon the gradual addition of **DSA-G** to the solution with the same excitation, while the emission at 481 nm simultaneously weakens (Fig. S19A and B, ESI[†]). The fluorescent intensity of **DSA-G**⊂**H2-2C4P** is somewhat stronger than that of **DSA-G**⊂**H1-4C4P**. The rapid enhancement in fluorescence can be observed until stoichiometric ratios are 1:2, after which the fluorescence intensity increases slowly. In comparison, the fluorescence spectra of **DSA-G**⊂**H3-4C1P** (Fig. S19C, ESI[†]), **DSA-G**⊂**H4-2C1P** (Fig. S19D, ESI[†]), **DSA-G**/ **H5-4CM** (Fig. S19E, ESI[†]), **DSA-G**/ **H6-2CM** (Fig. S19F, ESI[†]), and individual **DSA-G** (Fig. S19G, ESI[†]) have also been measured at the same concentrations as **DSA-G**, and there is no obvious fluorescence enhancement in the above systems except the linear weak increase due to the increased concentration of **DSA-G**. Thus, there are three major features contributing to the remarkable fluorescence enhancement of **DSA-G**⊂**H1-4C4P** and **DSA-G**⊂**H2-2C4P** (Fig. 1b): (i) FRET occurs between the two

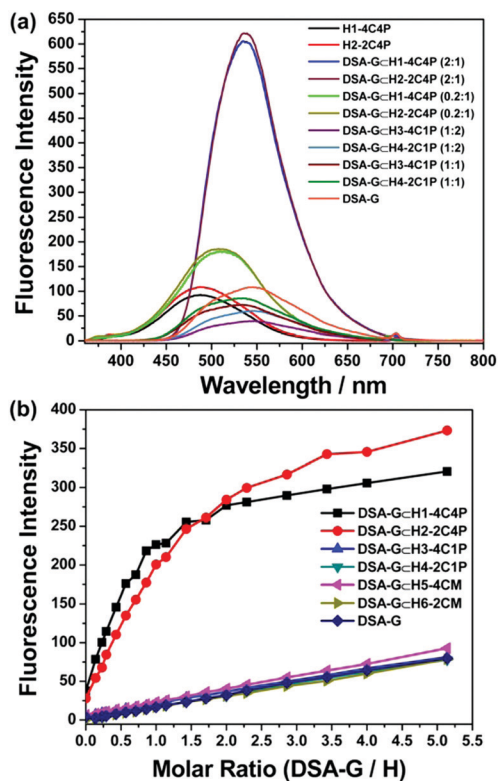


Fig. 1 (a) Fluorescence emission spectra ($\lambda_{\text{ex}} = 350$ nm; slit widths: ex. 5 nm, em. 5 nm; 25 °C, [H1-4C4P] = 50 μM ; [H2-4C4P] = 50 μM ; [DSA-G \subset H1-4C4P] = 100 μM \subset 50 μM ; [DSA-G \subset H2-2C4P] = 100 μM \subset 50 μM ; [DSA-G \subset H1-4C4P] = 10 μM \subset 50 μM ; [DSA-G \subset H2-2C4P] = 10 μM \subset 50 μM ; [DSA-G \subset H3-4C1P] = 100 \subset 200 μM ; [DSA-G \subset H4-2C1P] = 100 \subset 200 μM ; [DSA-G \subset H3-4C1P] = 100 \subset 2100 μM ; [DSA-G \subset H4-2C1P] = 100 \subset 100 μM ; [DSA-G] = 100 M) and (b) changes of fluorescence emission intensity of DSA-G \subset H1-4C4P, DSA-G \subset H2-2C4P, DSA-G \subset H3-4C1P, DSA-G \subset H4-2C1P, H5-4CM/DSA-G, H6-2CM/DSA-G and DSA-G (data gathered from Fig. S20, ESI †).

AIEgens, *i.e.*, TPE (donor) and DSA (acceptor), which is facilitated by the host-guest interaction; (ii) supramolecular assemblies bring TPE and DSA cores close to each other, resulting in the large restriction of internal rotation of AIE chromophores that populates the decay to the ground state; and (iii) the gradual addition of DSA-G. The increasing fluorescent intensity of DSA-G \subset H3-4C1P and DSA-G \subset H4-2C1P is consistent with DSA-G, suggesting that the supramolecular assemblies contribute to the close distance between donors and acceptors.

Fluorescence quantum yields and fluorescence lifetimes have been measured using an integrating sphere and time-resolved fluorescence, respectively. The rate constants for radiative deactivation (k_r) and for non-radiative deactivation (k_{nr}) were elucidated. Compared with the control systems (DSA-G \subset H3-4C1P, DSA-G \subset H4-2C1P, individual H1-4C4P, H2-2C4P and DSA-G), the ensembles of DSA-G \subset H1-4C4P and DSA-G \subset H2-2C4P have higher quantum yields and k_r , indicating that the supramolecular assembly and FRET contribute to the enhanced fluorescence synergistically (Table S1 and Fig. S20, S21, ESI †). We calculated the energy transfer efficiency of DSA-G \subset H1-4C4P and DSA-G \subset H2-2C4P both to be 88.6% through



Fig. 2 Plots of fluorescence intensity changes of DSA-G \subset H1-4C4P and DSA-G \subset H2-2C4P (control experiments: individual H1-4C4P, H2-2C4P and DSA-G) at $\lambda_{\text{em}} = 535$ nm upon (A) elevating and (B) lowering the temperature ($\lambda_{\text{ex}} = 350$ nm; slit widths: ex. 3 nm, em. 5 nm; 25 °C; concentrations: [H1-4C4P] = [H2-2C4P] = 35 μM , [DSA-G] = 70 μM); histograms of fluorescent intensity changes of (C) DSA-G \subset H1-4C4P and (D) DSA-G \subset H2-2C4P influenced by solvents ($\lambda_{\text{ex}} = 350$ nm; slit widths: ex. 3 nm, em. 5 nm; 25 °C; concentrations: [H1-4C4P] = [H2-2C4P] = 14 μM , [DSA-G] = 28 μM ; all the solvents are mixed with CHCl_3 at a volume ratio of 1:1 to guarantee the solubility).

the eqn (1), in which I_D stands for the fluorescence intensity of the donor without the acceptor while I_{DA} represents the intensity of the mixture of donor and acceptor.

$$\phi_{\text{ET}} = 1 - \frac{I_D}{I_{DA}} \quad (1)$$

In the process of supramolecular assembly, the stabilities of host-guest interactions were always influenced by temperature changes because entropy governs their free energies of complexation. The fluorescence intensity of DSA-G \subset H1-4C4P and DSA-G \subset H2-2C4P was decreased upon gradually elevating the temperature from 0 to 45 °C (Fig. 2A, chloroform b.p.: 61.2 °C), while both increased upon lowering the temperature back to 0 °C (Fig. 2B and Fig. S23, ESI †), indicating their thermo-responsive properties. In control experiments, individual H1-4C4P, H2-2C4P, and DSA-G at the same concentrations exhibited no obvious variations in fluorescence intensity upon changing the temperature (Fig. S22, ESI †).

Considering the key roles of solvent composition in supramolecular assembly/disassembly, we also explored the influence of mixed solvents to the assemblies of DSA-G \subset H1-4C4P and DSA-G \subset H2-2C4P through changes in fluorescence emission intensity (Fig. 2C and D). Chloroform was chosen as the basic solvent to prepare the solutions of DSA-G \subset H1-4C4P and DSA-G \subset H2-2C4P (Fig. S24, ESI †). As shown in Fig. 2D, the fluorescence intensities of DSA-G \subset H2-2C4P in the mixed solvents of hexane, toluene, THF, and ethyl acetate with chloroform (v/v = 1:1) were enhanced compared with that in pure chloroform, while the fluorescent intensities of DSA-G \subset H1-4C4P were enhanced in mixed solvents

of hexane/chloroform and toluene/chloroform. We ascribed their solvent-responsiveness to the cooperative impact of the polarities, viscosities, and solvent solubility, since the solubility of **DSA-G** \subset **H1-4C4P** and **DSA-G** \subset **H2-2C4P** and the polarities have significant influence on their molecular recognition events. On the basis of the above data, we conclude that the assembly of **DSA-G** \subset **H2-2C4P** exhibits relatively higher sensitivity toward both temperature and solvent composition as compared to that of **DSA-G** \subset **H1-4C4P**, suggesting that the assembly of **DSA-G** \subset **H2-2C4P** has been affected more by the greater degree of restriction and relaxation of phenyl units in TPE cores led by the shorter lengths of carbon chains. Additional data including UV-Vis absorption spectra, quantum yields, and time-resolved fluorescence decay curves have also been obtained (Fig. S25–S28 and Tables S2, S3, ESI[†]).

Photographs of solutions under natural light and 365 UV lamp showed color changes with different stoichiometric ratios of guest and host, which are consistent with the results of the fluorescence spectra in Fig. 1. The color changes of the fluorescence can be attributed to the extent of energy transfer between the donor-acceptor pairs at different stoichiometries. As in Fig. 3, individual **DSA-G** molecules without donor molecules exhibit yellow fluorescence but are apparently dimmer than when the molecular ratio of **DSA-G** \subset **H1-4C4P** is 2:1, which means the energy transfer from **H1-4C4P** plays a significant role. Yet when the stoichiometry is 0.2:1, part of the energy of the excited donor molecules has been released through emission, instead of being transferred to **DSA-G**. Partial energy transfer also manifests as shifts in the fluorescence emission spectra (Fig. 1).

Variable-temperature NMR analysis of **DSA-G** \subset **H1-4C4P** and **DSA-G** \subset **H2-2C4P** was performed (Fig. S29 and S30, ESI[†]). The split peaks changed into broaden peak, indicating the formation of supramolecular architectures. The concentration-dependent DOSY



Fig. 3 Photographs (above) under natural light and (below) ultraviolet irradiation (lamp $\lambda_{em} = 365$ nm); (A) **H1-4C4P** ([**H1-4C4P**] = 50 μ M); (B) **DSA-G** ([**DSA-G**] = 100 μ M); (C) **DSA-G** \subset **H1-4C4P** ([**H1-4C4P**] = 50 μ M, [**DSA-G**] = 100 μ M); (D) **DSA-G** \subset **H3-4C1P** ([**H3-4C1P**] = 200 μ M, [**DSA-G**] = 100 μ M); (E) **H2-2C4P** ([**H2-4C4P**] = 50 μ M); (F) **DSA-G** \subset **H2-2C4P** ([**H2-2C4P**] = 50 μ M, [**DSA-G**] = 100 μ M); (G) **DSA-G** \subset **H4-2C1P** ([**H4-2C1P**] = 200 μ M, [**DSA-G**] = 100 μ M); (H) **DSA-G** \subset **H1-4C4P** ([**H1-4C4P**] = 50 μ M, [**DSA-G**] = 10 μ M); (I) **DSA-G** \subset **H2-2C4P** ([**H2-2C4P**] = 50 μ M, [**DSA-G**] = 10 μ M); (J) **DSA-G** \subset **H3-4C1P** ([**H3-4C1P**] = 100 μ M, [**DSA-G**] = 100 μ M); (K) **DSA-G** \subset **H4-2C1P** ([**H4-2C1P**] = 100 μ M, [**DSA-G**] = 100 μ M).



Fig. 4 Fluorescence intensity changes upon changes in temperature and solvents. (A) Application of **DSA-G** \subset **H1-4C4P** (a, h, c) and (B) **DSA-G** \subset **H2-2C4P** (e, i, f) written on filter paper, (left: under natural light, right: under ultraviolet light, $\lambda_{em} = 365$ nm) as tunable fluorescent inks. (C) Fluorescent inks of **DSA-G** \subset **H1-4C4P** (a, h, c) and **DSA-G** \subset **H2-2C4P** (e, i, f) on the same white paper; (a): **H1-4C4P** ([**H1-4C4P**] = 50 μ M); h: **DSA-G** \subset **H1-4C4P** ([**H1-4C4P**] = 50 μ M, [**DSA-G**] = 10 μ M); c: **DSA-G** \subset **H1-4C4P** ([**H1-4C4P**] = 50 μ M, [**DSA-G**] = 100 μ M); e: **H2-2C4P** ([**H2-4C4P**] = 50 μ M); i: **DSA-G** \subset **H2-2C4P** ([**H2-2C4P**] = 50 μ M, [**DSA-G**] = 10 μ M); f: **DSA-G** \subset **H2-2C4P** ([**H2-2C4P**] = 50 μ M, [**DSA-G**] = 100 μ M)). Images of (D) the solutions and (E) solution-loaded pens.

spectra (Fig. S31 and S32, ESI[†]) of **DSA-G** \subset **H1-4C4P** and **DSA-G** \subset **H2-2C4P** indicated the changes of their diffusion coefficients. Compared with individual **DSA-G** with the diffusion coefficient of $(2.87 \pm 0.08) \times 10^{-9} \text{ m}^2 \text{ s}^{-1}$, the diffusion coefficient decreased upon addition of **H1-4C4P** or **H2-2C4P**, suggesting that they gradually assembled into large polymeric architectures.

Similar color changes can also be observed in fluorescence microscopy images, which indicate their solid-state fluorescent properties (Fig. S33–S36, ESI[†]). CIE coordinates showed in Fig. S37 (ESI[†]) also confirmed the color changes as blue, green, and yellow of the supramolecular ensembles (*i.e.*, **DSA-G** \subset **H1-4C4P**, **DSA-G** \subset **H2-2C4P**, and individual **DSA-G**, respectively).

In order to determine the solid-state fluorescence emission, measurements of fluorescent inks have also been performed (Fig. 4). Considering that chloroform can make plastics swell, the solutions were loaded into glass pens and written on filter paper (Fig. 4E). Similarly, the colours of the characters also changed from blue to green, and then to yellow upon the gradual addition of **DSA-G**. The handwriting on the coated papers lasted for at least 24 h, indicating that the tunable fluorescence of **DSA-G** \subset **H1-4C4P** and **DSA-G** \subset **H2-2C4P** were the same and efficient both in solution and the solid state.

Conclusions

In summary, tunable solid-state fluorescent materials were fabricated through supramolecular assembly of **DSA-G** \subset **H1-4C4P** and **DSA-G** \subset **H2-2C4P**, accompanied by supramolecular assembly-induced emission enhancement and FRET facilitated by molecular recognition. The remarkable fluorescence emission enhancement and the tunable fluorescence, ranges from blue to green to yellow, resulting from different stoichiometric ratios of **DSA-G**. Both of

the ensembles of **DSA-G**⊂**H1-4C4P** and **DSA-G**⊂**H2-2C4P** exhibited thermo- and solvent-responsive properties, showing great potential for smart optical switches. **DSA-G**⊂**H2-2C4P** possesses relative higher sensitivity to external stimuli compared with **DSA-G**⊂**H1-4C4P**. Furthermore, fluorescence microscopy images and experiments on fluorescent inks of these assemblies confirmed that their tunable fluorescence emission was efficient, even in the solid state. These tunable solid fluorescent materials possess great potential in smart optical devices. Efforts are ongoing in our laboratory to develop tunable solid-state fluorescent materials with extended wavelengths, taking advantage of supramolecular assembly and molecular machinery, to endow them with reversible stimuli-responsiveness.

Conflicts of interest

There are no conflicts to declare.

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