REVIEW

Nanomaterials-based electrochemical detection of chemical contaminants

Maduraiveeran Govindhan, Bal-Ram Adhikari and Aicheng Chen*

Owing to the high toxicity and detrimental effects of chemical contaminants to human health and the environment, public concerns over chemical contaminants in the environment and in foods have been mounting drastically. It is therefore significant to monitor contaminants *via* portable sensing devices, which encompass the demands of being low-cost and the potential for online environmental monitoring and food safety applications. This review will assess various concepts and recent advancements in design and the application of state-of-the-art nanomaterials through the incorporation of carbon nanomaterials, metallic and metallic oxide nanoparticles, titanium dioxide nanotubes, and dendrimers toward the development of electrochemical sensors for the detection of chemical contaminants in the environment and in foods. The development of nanomaterials based sensors facilitated by recent advances is having a major impact on sensor industries for environmental and food safety monitoring. Electrochemical sensing strategies have spurred intense interest in the research community as they have the capacity to serve as ideal sensor technology candidates, having such features as rapid response, robustness, high sensitivity and selectivity, low cost, miniaturization, and the potential for real-time monitoring. Nanomaterials have strong potential for increasing the competitiveness of new sensors for environmental monitoring and food safety applications through the combination of efficacious, yet simple fabrication techniques in the development of critical nanometric interfaces, and the optimization of their design and performance. Opportunities and future considerations for the use of nanomaterials in electrochemical sensors for producing advanced environmental and food sensing devices will also be addressed.

Department of Chemistry, Lakehead University, 955 Oliver Road, Thunder Bay, ON-P7B 5E1, Canada. E-mail: achen@lakeheadu.ca; Fax: +1 807 346 7775; Tel: +1 807 343 8318

Maduraiveeran Govindhan received his PhD in Chemistry from Madurai Kamaraj University, Tamil Nadu, India. He is currently working as a Postdoctoral Researcher in Department of Chemistry at Lakehead University, Canada. His research interests involve nanostructured metal particle assemblies and thin films, the design of optical and electrochemical sensors, fuel cells, electrochemical metal dissolution and water splitting, potentiometric high-temperature NO_x gas sensors and solid-state chemiresistive gas sensors. He has been conferred Senior Research Fellowship from the Council of Scientific and Industrial Research, India (2009) and Emerging Scientist Award from Lakehead University, Canada (2014).

Bal-Ram Adhikari is a PhD candidate in the Biotechnology programme at Lakehead University. He received his Master's degree in Microbiology from Tribhuvan University, Nepal. His research interests are in nanomaterials and nanochemistry for biological applications. His current research focuses primarily on the development of nanomaterials based sensors and biosensors for pharmaceutical, environmental and food safety applications.

1. Introduction

Enormous challenges currently confront environmental and food safety organizations that are responsible for the mitigation of risks associated with myriad complex mixtures of toxic chemical contaminants, which are constantly changing. Approximately 700 new chemicals are introduced into the marketplace annually in the US alone. Many of these substances are subsequently released into the ambient environment even though their toxicity and effects on human health and ecosystems are unknown, and where they are likely to be further combined with additional chemicals.[1] An expanding group of chemical contaminants of concern encompasses those that emanate primarily from industry, human and animal fecal waste, natural toxins, drinking water disinfection byproducts, through the corrosion of household plumbing systems, from food additives, personal care products, discarded pharmaceuticals, and *via* food preparation and packaging processes.[2] Chemical contaminants are of particular concern in that they may cumulatively cause severe damage to human health. Specifically, some chemicals are suspected of being cancer promoters, whereas other compounds have been associated with endocrine disrupting effects, or may be accumulated and biomagnified through the food chain.[3] The development of analytical methods and techniques in the environmental and food sciences have been developed parallel with growing consumer concerns as relates to food safety and security.[4] These research areas have been complicated by the increasing mobility and globalization of foodstuffs and associated raw materials, which are directly and indirectly exacerbating contamination events that are also growing on a global scale.[5] In an effort to decrease the complexity of food safety, food laboratories have been forced to replace their classical analytical methods with 21st century techniques, which may facilitate the provision of adequate responses to this expanding global demand. In EU countries, new European regulations (Regulation EC 258/97 or EN 29000 and subsequent issues), the Nutrition Labeling and Education Act in the U.S. as well as the Montreal Protocol, have had a major impact on food laboratory operations.[6] Dudarev *et al.*[7] have recently reported problems related to food security in the Russian Arctic *e.g.*, dietary imbalance, predominance of carbohydrates, shortage of dairy products, fruits and vegetables, chronic deficits of vitamins and microelements, and the presence of chemical, infectious agent, and parasitic food contaminants. Dudarev and his co-workers collected data from the Federal Automatic System "Social-Hygienic Monitoring", which contained data on ~178 000 analyzed food samples from all selected regions during 2007–2011. A chart related to the major chemical contaminants in foods is depicted in Fig. 1. Owing to the escalating demands on environmental and food monitoring, more sophisticated and appropriate instrumentation methods that have the capacity to provide improved qualitative and quantitative measurements are definitely warranted, as are increased sensitivity, precision, specificity, and/or speed of analyses.[8]

A variety of analytical techniques are commonly employed in environmental and food analysis. Typically, they encompass nuclear magnetic resonance (NMR), infrared (IR), atomic spectroscopy (AS), fluorescence, high-performance liquid chromatography (HPLC), gas chromatography (GC), capillary electrophoresis (CE), supercritical fluid chromatography (SFC), solid phase extraction (SPE), supercritical fluid extraction (SFE), headspace (HS), and flow injection analysis (FIA), *etc.* Electrochemical sensors are emerging to be a technology of choice in this respect as they are self-contained, compact and low cost, with minimal power requirements. These integrated devices

Aicheng Chen is a Professor of Chemistry and Canada Research Chair in Materials and Environmental Chemistry at Lakehead University. He received his PhD in electrochemistry at the University of Guelph in 1998. His research interests span the areas of electrochemistry, biosensors, green chemistry, materials science and nanotechnology. Chen is a receiptant of the Ontario Premier's Research Excellence Award, the Japan Society for the Promotion of Science Invitation Fellowship, the Fred Beamish Award of the Canadian Society for Chemistry, the Lash Miller Award of The Electrochemical Society Canadian Section, the Keith Laidler Award of the Canadian Society for Chemistry, the Lakehead University Distinguished Researcher Award, and the Canadian Catalysis Lectureship Award. In addition, he was elected as a Fellow of the Royal Society of Chemistry (UK) in 2011, and a Fellow of International Society of Electrochemistry in 2014.
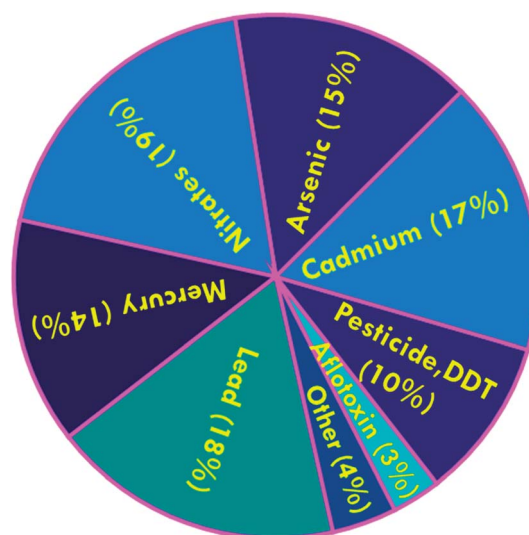

Fig. 1 Chart of chemical contaminants in food samples from all selected regions, percentage of total number of samples analyzed. Reproduced with permission from ref. 7. Copyright© 2013 International Journal of Circumpolar Health.

also offer high sensitivity, as they are capable of providing specific quantitative analytical information *via* the utilization of chemical/biological recognition elements, which can translate interfaced contaminant inputs into useable output signals.[9] Electrochemical sensors are generally easy to fabricate, while enabling the detection of single and multiple pollutants simultaneously. The primary drawback of these sensors for environmental and food applications is that they are not currently commercially available. Environmental and food samples are typically sent to laboratories, resulting in high costs and difficulties associated with real time monitoring.[10] The prerequisites of large sample analysis and the requirement for continuous monitoring in remote areas, make the development of these sensors even more challenging with respect to the establishment of first principles and specific applications.[11,12] A critical need for advanced and portable electrochemical sensors to facilitate environmental and food safety and security has been heightened, and is anticipated to expand tremendously over the next few years. The further integration of materials science, micro-fabrication, functional nanomaterials, and electrical engineering with sensor technologies will be required to advance wireless, real time environmental and food monitoring and analysis. Although some detection limits have been reported in the literature to be at femto-molar, ppt, or even lower concentrations,[13–15] in terms of ensuring the safety and security of food and the environment, an assessment on the detection levels that are suitable for these real-world applications will have to be made. The establishment of detection limits will be crucial in terms of risk assessment analyses. The desirable sensors should address the presence of specifical contaminants in environmental and food samples.

Over the last decade, nanomaterials have increasingly garnered attention in the disciplines of electrocatalysis and analytical chemistry, where they provide high surface area to volume ratios and can be tailored to encourage particular reaction pathways. Interestingly, morphologically dependent interatomic bond distances, melting points, chemical reactivity, as well as optical and electronic properties can have profound influences on the functionality of nanomaterials.[16] Nanomaterials may enable specific functionalities that are selected to align with targeted applications, which are derived from the interactivity of constituent nanometric elements with their immediate environments. The electrocatalytic activity of nanomaterials can be analyzed in detail using electrochemical measurements.[17] Nanomaterials make possible the potential development of novel devices with unique functionalities and processes that are activated on exposure to chemical contaminants, as well as the capacity for enhanced mobility and real time monitoring in environmental and food applications.[18] The evolution of nanostructured materials based electrochemical sensor technologies constitutes a very active and robust research area that is expected to provide next generation technologies for environmental and food science analyses, toward improving and/or supplanting conventional sensing strategies. This review will endeavor to elucidate the use of nanomaterials toward the design of potential electrochemical sensors and biosensors for the detection of chemical contaminants, in

facilitating efficacious environmental and food safety monitoring. One of its chief aims is to deliver an expansive overview and describe relevant advances in the field of nanomaterials based electrochemical sensing of chemical contaminants. Several core issues related to this rapidly emerging area must be taken into consideration for future research, and will be articulated in the last section.

2. Electrochemical methods in sensing

A variety of electrochemical methods have been explored for sensing chemical contaminants and biomolecules.[19–21] Typically, voltammetric techniques such as cyclic voltammetry (CV) and linear sweep voltammetry (LSV), differential pulse voltammetry (DPV), square wave voltammetry (SWV) *etc.*, amperometric and potentiometric techniques, and electrochemical impedance (EIS) techniques are utilized in the determination of analytes. For instance, electrochemical principles for the sensing of analytes are highly relevant in microfluidics and the broader field of separation science for the purposes of detection, injection, pumping, and valving. A number of microscopic techniques are coupled with electrochemical principles, such as scanning electrochemical microscopy (SECM) and chemically selective scanning tunneling microscopy (CSSTM), which are actually spatially resolved electrochemical sensors, even though they are classified as microscopic techniques. This operates *via* microdialysis probes that are coupled to an electrochemical detection system, which is an option following an on-line separation step.[22] An electrochemical sensor consists of two major components: (i) a chemical or biological recognition element; and (ii) a physical transducer (electrode) that transduces the analytical signal of the sensing event to an electronic circuit. Interactions between the sensing element of the substrate and the analytes are determined *via* the sensitivity, selectivity, speed of response and reversibility of the designed sensors.[23] Strong interactions are typically associated with higher sensitivity and selectivity, whereas perfect reversibility requires weak interactions. The critical parameters of electrochemical sensors are sensitivity, detection limit, dynamic range, selectivity, linearity, response time, and stability.[24] Fig. 2 displays a schematic illustration of an electrochemical sensor system, which illustrates the three primary elements, the sample (or analyte), the transduction platform, and the signal-processing step.

Electrochemistry based analytical methods have impacted multiple fields, including diagnostics, environmental analysis, food sciences, enzymatic kinetics and pharmacology.[25–27] In 2002, Wang reviewed miniaturizing electroanalytical systems and their incorporation into microfluidic devices for point of care applications, environmental analysis, and genetic testing.[28] In 2003, Baeumner[29] surveyed the application of electrochemical biosensors for environmental pollutants and food contaminants, which focused mainly on the detection of chemical and biological contaminants using electrochemical biosensors. Recently, the assembly of miniaturized
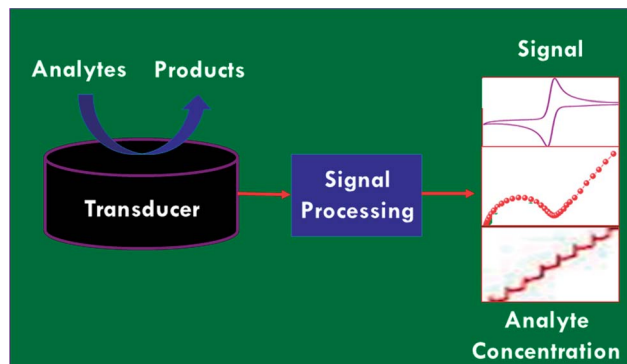
Fig. 2   Principal stages involved in the operation of an electrochemical sensor.

Fig. 3   Schematic of the electrochemical detection of chemical contaminants with nanomaterials.

electrochemical devices using graphene oxide (GO) with ionic liquid (IL) composites has been demonstrated for the electrochemical detection of several electroactive targets of importance in food analyses, environmental monitoring, and clinical diagnoses.[30] Banks and co-workers[31] reviewed the electrochemical applications and advancements of graphene (GR) based materials for sensor applications. The environmental and biological monitoring of organophosphorus (OP) pesticides and nerve agents using nanomaterials based biosensors have reviewed by Zhang et al.[32] Cliffel and co-workers[33] examined the advances made in electrochemical and biochemical sensor development and usage, which primarily covered the areas of reference electrodes, potentiometric sensors, voltammetric sensors, amperometric sensors, biosensors, immunosensors, and mass sensors. Our research group has recently surveyed the design and the development of electrochemical sensors based on nanomaterials for biomedical applications,[34] the applications of SWV in electrochemical sensing,[35] and the synthesis and application of Pt based nanomaterials.[36] In the present review, we will focus on the development of electrochemical sensors and biosensors based on nanomaterials for the detection chemical contaminants and their promising applications in environmental monitoring and food safety.

3.   Nanomaterials based electrochemical sensing platforms

The schematic depiction of a variety of nanomaterials employed for the detection of chemical contaminants is illustrated in Fig. 3. Carbon based nanomaterials include single-walled carbon nanotubes (SWNTs), multiwalled carbon nanotubes (MWNTs), single-walled carbon nanohorns (SWCNHs), fullerenes (e.g., C_{60}), and graphene, among many others. Carbon has the capacity to hybridize into sp, sp^2 and sp^3 configurations, contingent on the bonding relationships with neighboring atoms, which have narrow gaps between their 2s and 2p electron shells. These associations are responsible for enabling the design of a wide assortment of nanoscale carbon-based materials that are very versatile, which exhibit distinct properties in terms of size, surface area, strength, photonics and electronics.
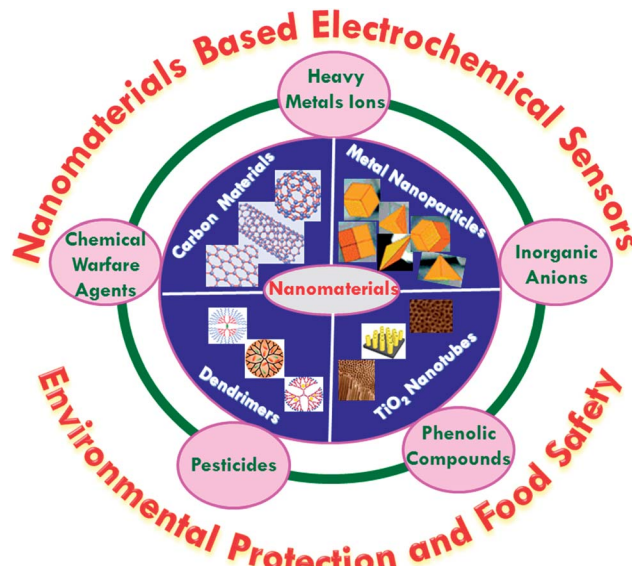
Carbon nanomaterials demonstrate many interesting attributes that make them ideal for serving as electrode materials in electrochemical sensor applications.[37–42] The presence of surface resident reactive groups, or edge-plane like sites that are situated at the ends of their structures, and at defect sites, are responsible for the excellent electrocatalytic activity of carbon nanomaterials. The advantages and limitations of carbon-based nanomaterials with the respect to analytical chemistry including sample preparation, separation and detection were reviewed by Scida et al. in 2011.[43] The functionalization of carbon nanotubes has been highlighted by a feature article authored by Huang and co-workers.[44] For instance, the chemically reduced GO modified GC electrode that functioned as the transducer for a potassium ion sensor with a PVC-based selective membrane, which showed superior performance in contrast to conductive polymer based sensors.[45] A potentiometric sensor has been developed by Michalska and co-workers[46] that employed functionalized graphene as a transducer and selective film for the detection of Zn^{2+} ions. In addition, the sensing metal ions was performed by the functionalization of a GO-based electrochemical sensor.[47] MWCNT modified electrodes were developed for the detection of europium(III) via differential pulse adsorptive stripping voltammetry (DPASV), where a wide linear range with a lower detection limit was obtained.[48] CNTs have been used as electrochemical sensing platforms for the detection of microbiological chemical toxins,[49] pesticides[50] and explosives.[51] A tyrosinase-immobilized carbon-felt-based flow biosensor was explored on the development of a highly sensitive sensor for monophenolic compounds.[52] SWCNH is another type of carbon-based nanomaterials, which has a conical structure with a base diameter of ~5 nm and a tubule length of ~50 nm. SWCNHs have been used for the detection of chemical contaminants such as chloramphenicol,[53] triclosan and bisphenol A.[54]

Metallic nanomaterials such as platinum, gold, silver and palladium with intriguing size dependent electrical, optical, magnetic, and chemical properties have been intensively studied for their fundamental scientific interest and toward many potential technological applications. Noble metal nanoparticles are under extensive study and have led to the development of numerous analytical techniques and methods for environmental monitoring and food safety applications.[55–57] The inhibitory enzyme–nanoparticles based biosensors were developed for food safety and environmental monitoring applications.[58] The inhibitory based biosensor can increase the sensor stability and render the regeneration of the enzyme easier and it may suffer from side reactions due to coexistence of several enzymes. The non-enzymatic electrochemical sensor for H_2O_2 was reported, which employed Au NPs that were embedded within a silicate sol–gel electrode.[59] Monometallic and bimetallic nanoparticles, such as gold nanoparticles (Au NPs), platinum nanoparticles (Pt NPs), palladium nanoparticles (Pd NPs), and platinum–palladium nanoparticles (Pt–Pd NPs) have been widely utilized in the development of electrochemical sensors for toxic chemicals.[60] Au NPs hosted by a water-soluble silsesquioxane polymer based electrochemical sensor was developed for the detection of nitrophenol isomers.[61] Metal oxide based nanostructures, such as ZnO, Fe_3O_4, NiO, SnO_2, ZrO_4, TiO_2, MgO and MnO_2, have also been widely used in the detection of chemical contaminants due to their unique nano-morphological, functionally biocompatible, non-toxic and catalytic properties.[62] Porous MgO nanoflowers have been synthesized for the fabrication of an electrochemical sensor for the detection of heavy metals. Radhakrishnan et al.[63] described a highly sensitive electrochemical sensor for nitrite detection, which was based on Fe_2O_3 nanoparticles that were decorated with reduced graphene oxide nanosheets, showing a linear concentration range of 50 nM to 0.78 mM, with a detection limit of 15 nM. Jena et al.[64] designed an electrochemical sensor that was based on ensembles of Au nanoelectrodes for the simultaneous electrochemical detection of ultra-trace concentrations of arsenic, mercury, and copper ions.

Titanium dioxide nanotubes (TiO_2 NTs) have garnered significant research interest due to their ease of preparation, high orientation, extensive surface area, high uniformity and excellent stability. The electrochemical behavior of TiO_2 is contingent on its crystalline structure, surface properties and textural properties, which include specific surface area, pore volume, pore dimensions and distribution.[65] TiO_2 NTs decorated with metal particles and enzymes have been employed for the development of electrochemical sensors and biosensors for the detection of a variety of species.[66–70] Au@Pd core–shell nanoparticles supported on amino-functionalized TiO_2 NTs were developed and tested as a potential electrochemical sensor for the detection of hydrazine.[71] Ozturk and his co-workers[72] reported rutile TiO_2 NTs for the detection of volatile organic compounds.

Dendrimers are a new class of polymeric materials, which are monodisperse, three-dimensional (3D) hyperbranched, nanoscale polymeric architectures with an extremely high surface functional group density.[73] The electrostatic interactions, complication reactions, various types of weaker forces, such as van der Waals, H-bonding, etc. are the prime drivers of guest encapsulation within dendrimers.[74,75] In recent years, dendrimer-encapsulated metal NPs and dendrimer-cored metal NPs have attracted considerable interest due to their potential use in catalysis and sensors. Fang et al.[76] investigated the electrochemical communication between Au NPs deposited onto adsorbed dendrimers of poly(amido amine) (PAMAM) and the gold or platinum electrode surface. Zheng and co-workers[77] have demonstrated an electrochemical sensor for the detection of nitrite using PAMAM dendrimer-stabilized Ag NPs. Siriviriyanun et al.[78] developed an electrochemical sensor approach that involved CNTs, Pt NPs, dendrimers, and enzymes that enabled effective and rapid detection of small volumes of analytes on potentially cost effective and disposable printed chips. The electrocatalytic activity of the prepared biosensors demonstrated the feasible detection of H_2O_2 at a low electrode potential, and the detection of an inhibitory biocontaminant (organophosphorus pesticide) over a wide range of concentrations. More details on the conducting polymer nanomaterials for sensing and biomedical applications may be found in the recent review articles.[79,80]

4. Applications in the detection of chemical contaminants

The development of electrochemical sensors is of great significance due to their capacity to resolve a potentially large number of analytical problems and challenges across very diverse areas, such as defense, homeland security, agriculture and food safety, environmental monitoring, medicine, pharmacology, industry, etc. A growing demographic of individuals with a high risk of diabetes and obesity, and the rising incidence of chronic diseases such as heart disease, stroke, cancer, chronic respiratory diseases, tuberculosis, etc., are driven by several factors, including medical and health issues that may be traced to significant problems with environmental monitoring. There also exist, of course, serious monitoring challenges for security and military and agriculture/food applications.[81] Many research articles have been published in recent years that discuss the application of electrochemical sensors for food analysis,[82,83] and the detection of specific analytes in environmental applications.[84–86] Nanomaterials based electrochemical sensing platforms have been developed for the recognition and quantification of chemical contaminants in water and food. The primary focus of this review is the application of electrochemical sensors for environmental and food safety and security. Chemical contaminants may be classified chiefly as heavy metals, small organic and inorganic pollutants, pharmaceutical and personal care products, toxins of microbial origin, and pathogens.[87,88] The detection of chemical contaminants may originate from a variety of sources, such as the improper use or storage of pesticides and veterinary drugs, and the creation of chemicals during processing techniques.[89] The high sensitive and selective analysis of food and water contaminants is vital, not only for human and wildlife protection, but also aids global

trade, by assisting with the specific identification of contaminated products, and thereby limiting its distribution.[90]

Chemical contaminants such as heavy metals, nitrates, sulphates, phenolics, pesticides and nerve agents, all have potentially serious health implications in terms of causing illness, or by limiting the efficacy of medicines through the development of drug resistance.[91,92] Physicochemical methods such as liquid chromatography-tandem mass spectrometry (LCMS/MS) are usually employed to detect the presence of these toxic compounds; however, these techniques tend to be expensive, complicated, and time consuming.[37] Electrochemical sensing methods have the potential to be integrated into portable and rapid detection tools to greatly reduce the expense and time for chemical contaminant analysis.

4.1. Detection of heavy metals

The increasing heavy metal contamination of fresh water and food systems in worldwide has become a core problem facing humanity. The detection and determination of heavy metal ion levels in potable water supplies and food samples is essential to human health and safety.[93] The real-time, high resolution measurement of heavy metals such as mercury, lead, cadmium, chromium and arsenic, *etc.* in water and foods at ultralow concentrations are required for maintaining safe water supplies.[94] Due to the non-biodegradable, non-thermo degradable, and toxic nature of heavy metals, they tend to accumulate in living organisms owing to their low clearance rates. Various health problems, including developmental disorders and organ failure are typically associated with exposure to these elements.[95] Mercury has become emphasized within the family of heavy metals, and its sources of contamination are derived mainly from coal combustion, industrial production, and agricultural chemicals. Water soluble mercuric ions ($Hg(II)$) are an important form of this heavy metal, which has a high toxicity; it can also exist in the environment in a variety of forms.[96] Mercuric ions can accumulate in the human body through the food chain and lead to permanent damage to the brain, and to other chronic diseases.[97] In 2005, the U.S. Environmental Protection Agency set the maximum permissible level of mercuric ions in potable water at 10 nM. The sensing of mercury is of critical importance in environmental and food safety, as

well as clinical toxicology. Zeng and co-workers[98] recently developed a three-dimensional electrochemical sensor, which was constructed using mercury-specific oligonucleotides, gold nanoclusters, and an anionic intercalator, as shown in Fig. 4. This sensor spatially captured electroactive mercuric ions to enable trace mercury measurements with high sensitivity due to the steric reaction field in the electrode surface microenvironment that was formed by the Au nanoclusters, thereby exhibiting strong environmental adaptability and high selectivity. A low detection limit was achieved at 0.01 nM with a linear range of 0.05 to 350 nM.

The Environmental Protection Agency has assessed that a portion of human exposure to lead and cadmium occurs through contaminated drinking water. The permissible level of lead and cadmium in drinking water is 3.0 and 10.0 mg L^{-1}, respectively, as established by the World Health Organization (WHO). Consequently, the accurate detection and determination of these heavy metals at trace levels is of particular importance. Kong *et al.*[99] developed a rapid and sensitive electrochemical sensor for the determination of trace amounts of $Pb(II)$ and $Cd(II)$ in aqueous solutions using fiber-in-tube solid-phase microextraction (SPME) in combination with anodic stripping voltammetry (ASV). These researchers investigated the major factors that influenced the preconcentration and extraction of the metal ions, pH of the aqueous solution, filling mode, sample flow rate, eluent concentration, eluent volume, and effluent flow rate with a limit of quantification of 0.001 μg L^{-1}.

A facile electrochemical sensor was designed with functionalized GO sheets on a Au surface, resulting in the complexation-enhanced electrochemical detection of heavy metal ions, such as $Pb(II)$, $Cu(II)$ and $Hg(II)$, with improved low level detection limits.[100] Aneesh *et al.*[101] demonstrated the electrochemical synthesis of a gold atomic cluster–chitosan nanocomposite film modified gold electrode for the ultra-trace determination of mercury over a wider calibration range of 1 fM to 0.1 μM with a limit of detection (LOD) of 0.8 fM. Carcinogenic chromium ions ($Cr(VI)$) are considered as toxic species, and highly soluble and mobile in both biological and natural ambient systems.[102] Significant volumes of chromium compounds are discharged into the environment from various industrial processes such as mineral processing, pigments and electroplating. In USA, the allowable groundwater level of

Fig. 4   Detection strategies for a mercuric ion sensor using mercury–specific oligonucleotides, gold nanoclusters, and an anionic intercalator. Reproduced with permission from ref. 98 Copyright© 2014 Royal Society of Chemistry.

naturally occurring Cr(VI) can be up to 220 mg L^{-1} (4.2 μM).[103] A provisional limit of 50 mg L^{-1} (0.96 μM) in groundwater was set by The World Health Organization (WHO) in order to prevent Cr(VI) toxicity in humans and the environment.[104] Our group has recently developed a facile and effective electrochemical Cr(VI) sensor using Au NPs-decorated TiO$_2$ NTs.[70] Fig. 5A shows the SEM images of Au NPs decorated TiO$_2$ NTs. The Au NPs ($\sim$10 nm in diameter) were well dispersed on the as-prepared TiO$_2$ NTs. The diameter of these nanotubes was estimated to be $\sim$120 nm, with a wall thickness of $\sim$60 nm and a length of 3–4 μm. Due to its nanoparticle/nanotubular heterojunction infrastructure, the electrocatalytic activity of Ti/TiO$_2$NT/AuNPs electrode exhibited an almost 23-fold improvement in activity as

Fig. 5 (A) SEM image of the Au NPs decorated TiO$_2$NT electrode. (B) Amperometric responses at the Au nanoparticle decorated TiO$_2$NT electrode for the successive addition of Cr(VI) with a polarized potential of 0.28 V in 0.1 M HCl. (C) Calibration plot of corresponding amperometric response. Reproduced with permission from ref. 70 © 2014 Royal Society of Chemistry.

compared to a smooth polycrystalline gold electrode. The Ti/TiO$_2$NT/AuNPs electrode provided a wide linear concentration range of 0.1 μM to 105 μM, a low detection limit of 0.03 μM, and a high sensitivity of 6.91 μA μM^{-1} Cr(VI) using and amperometric technique, as shown in Fig. 5B and C. Furthermore, the Ti/TiO$_2$NT/AuNPs electrode exhibited good selectivity against interference from coexisting Cr(III) and other metal ions, and excellent recovery for Cr(VI) detection in both tap and lake water samples. The phosphinic acid functionalized CNT utilized as a sensing substrate for the sensitive and selective sensing of Cr(VI) in the range of 0.01 to 10 ppb, with a detection limit of 0.01 ppb.[105] Rajkumar et al.[106] demonstrated an electrochemical sensor for arsenic in controlled laboratory and drinking water samples using a nano Au–crystal violet film, which was fabricated on a glassy carbon electrode. A variety of electrochemical sensors and devices have been proposed and tested for the heavy metal assays in water.[107] Some of other developed electrochemical sensors for the detection and determination of heavy metal ions such as Hg(II),[108–111] Cr(VI),[69,109] Pb(II),[113,114] Cd(II),[114,115] As(III)[116–118] and Cu(II)[119] using the nanostructured materials are listed in Table 1.

4.2. Detection of inorganic anions and compounds

Environmental and food contaminants are the main vectors that may negatively impact both human and animal life. These contaminants impart mild to severe short-term, long term, or even deadly effects. The contamination of inorganic nitrite ions (NO$_2^-$) pollutants in water and food supplies continues to be a significant issue throughout the world. They can easily form carcinogenic nitrosamines by readily binding with hemoglobin, and causes methemoglobinemia, primarily in infants.[120] Nitrites are present in foods as a preservative compound in meats, cheeses, and natural vegetables.[121,122] The maximum permissible concentration of nitrite in portable water is 2.17 μmol L^{-1}.[123] Due to increasingly stringent environmental protection and public health measures, the quantitative analysis of nitrites in drinking water has more and more relevance. Miao et al.[124] developed an electrochemical sensor by employing platinum nanoparticles for the detection of nitrites with a detection resolution limit of 5 μM, and this method was successfully applied to the determination of nitrites with ambient lake water. The Au NPs coupled with MWCNT based sensing platform was prepared for nitrite detection incorporating various experimental parameters for the voltammetric response of nitrites.[125] The prepared sensor showed good selectivity in response to the most common ions and many environmental organic pollutants, with a sensitivity of 0.42 μA μM^{-1}, and a low detection limit of 0.01 μM. A bi-enzymatic biosensor was developed by Karunakaran and co-workers[126] for the simultaneous detection of nitrite and nitrate ions using copper, zinc superoxide dismutase (SOD1) and nitrate reductase (NaR) co-immobilized on a CNT–polypyrrole (PPy) nanocomposite modified electrode. Fig. 6 depicts the fabrication of a biosensor for the detection and determination of nitrite and nitrate ions.[126] A CNT integrated PPy composite was prepared by casting CNT onto an as-prepared PPy film modified Pt electrode.

Table 1 List of electrochemical sensors and biosensors based on nanomaterials for the detection of chemical contaminants[a]

Nanomaterials	Analytes	Samples	Detection methods	Linear range	Detection limit	Ref.
CILE–Au NPs	Mercury(II)	Waste and tap water	SWASV	0.01 to 50 ppb	15 ppt	108
MB–DNA–Au NPs	Mercury(II)	Tap water	DPV	0.2 to 100 ppb	0.064 ppb	109
Au–Pt NPs–NFs	Mercury(II)	River and tap water	SWASV	0.1 to 10 ppb	0.008 ppb	110
DNA–Au NPs	Mercury(II)	River water	DPV	0.5 to 120 nM	0.06 nM	111
Quercetin–MWCNT	Chromium(VI)	Mineral, river and tap water	ASV	1 to 20 μM	0.3 μM	112
Au NPs–TiO$_2$NT	Chromium(VI)	Lake and tap water	Amperometry	0.1 to 105 μM	0.03 μM	70
DNA–SWCNT	Lead(II)	River and tap water	DPV	1 nM to 10 nM	0.4 nM	113
MWCNT	Lead(II)	Tap water, waste water and food samples	SWASV	0.4 to 1100 ng mL^{-1}	0.25 ng mL^{-1}	114
	Cadmium(II)		SWASV	1 to 1200 ng mL^{-1}	0.74 ng mL^{-1}	
Stannum/p-ABSA–GR	Cadmium(II)	Waste, tap and farm land irrigation waster	SWASV	1.0 to 70 μg L^{-1}	0.05 μg L^{-1}	115
Ag NPs–GO	Arsenic(III)	River and ground water	SWASV	13.33 to 375.19 nM	0.24 nM	116
Au NPs–Te-hybrid	Arsenic(III)	N/A	SWASV	0.1 to 10 ppb	0.0026 ppb	117
Au NPs–Fe$_3$O$_4$	Arsenic(III)	Waste water	SWASV	0.01 to 1 ppb	0.00097 ppb	118
Aptamer–Au NPs	Copper(II)	Industrial electrolyte bath (gold, copper and brass)	SWV	0.1 nM to 10 μM	0.1 pM	119
MWCNT–iron(II) porphyrin	Cyanide	N/A	SWV	40 nM to 150 μM	8.3 nM	134
Au NPs/Co(II)MTpAP	Nitrites	Well water	Amperometry	0.5 μM to 4.7 mM	60 nM	135
CoPcF–MWCNT	Nitrites	N/A	Amperometry	0.096 μM to 340 μM	0.062 μM	136
Au NPs–MWCNT	Nitrites	Mineral water, tap water, sausage, salami and cheese	SWV	0.05 μM to 250 μM	0.01 μM	137
Fe$_2$O$_3$–RGO	Nitrites	Tap water	DPV	50 nM to 0.78 mM	15 nM	138
SOx–PB NPs–PPY	Sulphites	Red and white wine	CV	0.5 to 1000 μM	0.1 μM	139
Co NPs	Sulphites	Fruits, wine, tap and drinking water	CV	1 to 59 μM	0.4 μM	140
p-Aminophenol–MWCNT	Sulphites	Liquor, boiler, waste, river and drinking water	SWV	0.2 to 280 μM	0.09 μM	141
EDA–MWCNT	Nitric oxide	Rat liver tissue	DPV	95 nM to 11 μM	95 nM	142
Pt NPs–acetylene black	Nitric oxide	Rat liver tissue	DPV	0.18 to 120 μM	50 nM	143
RGO–Au–TPDT NRs	Nitric oxide	N/A	Amperometry	10 to 140 nM	6.5 nM	144
Nanoporous Au	Phenylamine	N/A	Amperometry	5 to 25 μM	0.5 μM	162
Nano-Zr–ZSM-5	p-Aminophenol	River and tap water	DPV	60 nM to 500 μM	26 nM	163
	o-Aminophenol			120 nM to 500	30 nM	
Aptamer–MWCNT	Polychlorinate biphenyl	Serum	CV	0.16 to 7.5 μM	10 nM	164
Pt–Au–OSi@CS	Catechol	N/A	Amperometry	0.06 to 90.98 μM	0.02 μM	165
	Hydroquinone			0.03 to 172.98 μM	0.01 μM	
RGO–TT–CNT	Hydroquinone	Tap water	DPV	0.01 to 200 μM	7.61 nM	166
	Catechol			0.5 to 200 μM	11.7 nM	
3D-GR	Catechol	Lake and tap water	DPV	0.25 μM to 12.9 μM	80 nM	167
Fe$_3$O$_4$–PGMA–HRP	Phenol	N/A	Amperometry	0.5 to 8.5 mM	0.028 mM	168
	Catechol			0.5 to 11.0 mM	0.046 mM	
CuNPs–HCF	Hydroquinone	N/A	Amperometry	10 μM to 0.2 mM	2.2 μM	169
ds DNA–PANI	Hydroquinone	Tap water	DPV	12.5 μM to 32 μM	0.9 μM	170
NNH–MWCNT	4-Chlorophenol	River, lake and tap water	DPV	1.0 to 750 μM	0.5 μM	171
MWCNT	Bisphenol A	Mineral water	CV	0.099 to 5.8 μM	0.032 μM	172
Pt–Pd NPs/CNT–RGO	1,3,5-Trinitrophenol	N/A	DPV	1 to 115 ppb	0.52 ppb	173
	Nitrobenzene			1 to 155 ppb	0.42 ppb	
OPH-bacteria–OMCs	Paraoxon	Sea, tap and sewage water	Amperometry	0.05 to 25 μM	9 nM	188
	Parathion			0.05 to 25 μM	15 nM	
	Methyl parathion			0.05 to 25 μM	10 nM	

Table 1 (*Contd.*)

Nanomaterials	Analytes	Samples	Detection methods	Linear range	Detection limit	Ref.
OMCs	Paraoxon	Sea, tap and sewage water	DPV	0.015 to 10 μM	1.9 nM	189
	Parathion			0.01 to 10 μM	2.1 nM	
	Methyl parathion			0.015 to 10 μM	3.4 nM	
GdHCF–GNs	Methyl parathion	River and tap water	DPV	0.008 to 10 μM	1 nM	190
MWCNT–CeO$_2$–Au NPs	Methyl parathion	River water and soil	DPV	0.1 nM to 0.1 μM	30.2 fM	191
ZrO$_2$NPs–GR	Methyl parathion	Garlic and cabbage	SWV	0.002 to 0.9 μg mL^{-1}	0.6 ng mL^{-1}	192
β-CD–GR	Methyl parathion	Sea water	DPV	0.3 to 500 ppb	0.05 ppb	193
CuO NWs–SWCNT	Malathion	N/A	DPV	0.3 to 600 nM	0.3 nM	144
PANI–SWCNT	Malathion	Tap water	DPV	0.2 μM to 1.4 μM	0.2 μM	194
CoO–RGO	Carbofuran	Grapes, orange, tomato and cabbages	DPV	0.2 to 70 μM	4.2 μg L^{-1}	195
AChE–Fe$_3$O$_4$–CH	Carbofuran	Cabbages	SWV	5.0 nM to 0.14 μM	3.6 nM	196
AChE–(3-APTES)–SWCNT	Sarin	N/A	LSV	20 to 60 nM	15 nM	206
	Diisopropyl fluorophosphate			20 to 80 nM	25 nM	
AChE–RGO	Dichlorvos	River water	Amperometry	5 ng mL^{-1} to 20 μg mL^{-1}	2 ng mL^{-1}	207
Au–ZrO$_2$NPs–SiO$_2$	Paraoxon-ethyl	N/A	SWV	1.0 to 500 ng mL^{-1}	0.5 ng mL	208
GR	Isoproturon	River and rice field water, tomato and lettuce	SWSV	0.02 to 10.0 mg L^{-1}	0.02 mg L^{-1}	209
	Carbendazim			0.5 to 10.0 mg L^{-1}	0.11 mg L^{-1}	

[a] CILE – carbon ionic liquid electrode; MB – methylene blue; NFs – nanofibres; *p*-ABSA – *p*-aminobenzene sulfonic acid; Co(II)MT*p*AP – *meso*-tetra(*para*-aminophenyl)porphyrinato cobalt(II); CoPcF – tetrakis(3-trifluoromethylphenoxy)phthalocyaninato cobalt(II); SO(x) – sulfite oxidase; PB NPs/PPY – Prussian blue nanoparticles/polypyrrole composite; TPDT – amine functionalized silicate sol–gel matrix; ZSM – metallosilicates; OSi@CS – organo silica@chitosan; EDA – ethylenediamine; TT – terthiophene; HCF – hexacyano ferrate; NNH – nano nickel hydroxide; PANI – polyaniline; PGMA – poly(glycidyl methacrylate); HRP – horseradish peroxidase; GNs – graphene nanosheets; β-CD – β-cyclodextrin; CH – cholinesterases; 3-APTES – aminopropyltriethoxy silane; AChE – acetylcholinesterase; OMCs – ordered mesopores carbons; LSV – linear sweep voltammetry; DPV – differential pulse voltammetry; SWV – square wave voltammetry; SWASV – square-wave anodic stripping voltammetry; ASV – anodic stripping voltammetry.

A 1 : 1 ratio mixture of enzyme solutions was coated onto a CNT–PPy–Pt electrode, resulting in a composite electrode. The electrocatalytic activity of SOD1 occurred toward nitrite oxidation at 0.8 V, and showed a linear range of 100 nM to 1 mM, with a detection limit of 50 nM, and sensitivity of 98.57 nA mM^{-1} cm^{-2}. The electrocatalytic reduction of nitrate ions occurred at −0.76 V using co-immobilized NaR and exhibited a linear range of 500 nM to 10 mM, with a detection limit of 200 nM, and sensitivity of 84.57 nA mM^{-1} cm^{-2}.

Sulphite ions (SO$_3{}^{2-}$) pollutants are widely utilized as a preservative, bleaching agent, reducing agent and antioxidant in the food industry, and also as a food additive in many countries. It may give rise to headaches, nausea, dizziness, asthma and other allergic reactions if sulphite levels are exceeded. The U.S. Food and Drug Administration (FDA) requires that sulfites must be declared on food labels when it is used as a food ingredient, a processing aid, or when present as an indirect food constituent (*e.g.*, dried fruit pieces), with a permissible concentration in foods of ≥10 ppm. The enzymes including sulfite oxidase, tetrathiafulvalene modified screen-printed carbon electrode based biosensors were developed for the determination of sulfites in wine.[127] The amperometric detection of sulfite *via* simple flow injection using a carbon nanotube–PDDA–Au NPs modified GC electrode was reported by Amatatongchai *et al.*[128] The linear working range for the determination of sulfite was 2–200 mg L^{-1} with a detection limit of 0.03 mg L^{-1} and the sensor was applied to the determination of sulfite in fruit juices and wines with a throughput of 23 samples per hour. The simultaneous electrochemical sensor was developed by Maduraiveeran *et al.*,[129] utilizing Au NPs, which were embedded in an amine functionalized silicate sol–gel (APS) modified electrode, was employed for the detection and quantification of toxic hydrazine, sulfite, and nitrite compounds in pH 7.2, as shown in Fig. 7. Fig. 7A displays a schematic depiction of Au nanoparticles dispersed in 3D network with the simultaneous electrocatalytic oxidation of hydrazine, sulfite and nitrite. The prepared Au NPs based electrode showed that the electrooxidation peaks appeared at 0.05, 0.2 and 0.55 V for hydrazine, sulfite and nitrite in pH 7.2, with a large decrease in the overpotentials to the extent of ~750 mV, ~600 mV, and ~250 mV, respectively, as shown in Fig. 7B. This figure also shows the LSV responses observed for the hydrazine, sulfite and nitrite mixture at the Au NPs modified GC electrode, with the successive addition of their concentrations. The linearly increased anodic currents were obtained with the elevated concentration of analytes (Fig. 7C).

The loading of Au on SnO$_2$ hollow microtubes for the non-enzymatic detection of H$_2$O$_2$ was reported using an amperometric method. The linear range was measured to be from 10 μM to 1 mM with the detection limit of 0.60 μM.[130] Furthermore,

Fig. 6 Construction of a NaR–SOD1–CNT–PPy–Pt electrode and reactions involved during the simultaneous determination of NO_2^- and NO_3^-. Reproduced with permission from ref. 126 © 2014 Elsevier.

this non-enzymatic sensor exhibited excellent selectivity for H_2O_2 when challenged with other normally co-existing electroactive species. The Pd NPs decorated MWNT (PdNPs/MWNT) electrocatalyst based electrochemical sensor was demonstrated for the determination of bromate ion by Li et al.[131] Bromate has been extensively used as a food additive in the production of fermented beverages and fish pastes, and also has been found in drinking water as a disinfection by-product (DBP).[132] The International Agency for Research on Cancer has classified bromate as a Group 2B carcinogen, as it has been reported that bromate ion may cause renal cell tumors.[133] The electrochemical reduction of bromate was obtained at between 0.15 and −0.25 V. The effects of different factors, including the scan rate, temperature and the initial concentration of bromate ions on bromate determination were studied in depth. An amperometric method showed that the PdNPs/MWNT modified electrode could be successfully employed as a sensor for bromate in the concentration range from 0.1 to 40 mM, with a sensitivity of 768.08 μA mM^{-1} cm^{-2}. These Pd NPs/MWNT electrodes have good potential for applications in fabrication of bromate sensors. Salimi et al.[134] demonstrated the use of MWNTs and an iron(III)–porphyrin film as a sensing substrate for the detection of chlorate (ClO_3^-), bromate (BrO_3^-) and iodate (IO_3^-). The greatly improved facilitation of electron transfer between Fe(III)P and CNTs that were immobilized on the electrode surface was the factor behind the enhanced electrocatalytic activity, with detection limits of 0.5 μM (chlorate), 0.6 (bromate), and 2.5 μM (iodate). This sensor possessed an excellent electrochemical reversibility of the redox couple, good reproducibility, high stability, a low detection limit, good longevity, rapid amperometric response time, wide linear concentration range, technical simplicity, and the possibility of rapid preparation and scale up. Furthermore, the electrochemical detection of inorganic anions including, cyanide,[135] nitrites,[136–138] sulphites[139–141] and nitric oxide[142–144] using nanomaterials are included in Table 1.

4.3. Detection of phenolic compounds

Phenolic compounds are known to be widespread in the environment from industrial waste due to their use as intermediates in the production of pharmaceuticals, wood preservatives, explosives, and in leather treatments. These substances have a toxic effect on humans, animals and plants, even at very low concentrations.[145] A variety of ~165 phenolic compounds are known to have a toxic effect on the environment. Phenolic compounds, as well as their vapors, are highly corrosive to the eyes, the skin, and the respiratory tract. Due to phenolics' caustic and anti-adipose properties, prolonged skin contact with phenol may cause dermatitis, or even second and third-degree burns. These compounds may also have harmful effects on the central nervous system and heart, resulting in dysrhythmia, seizures, coma and kidney trauma as well.[146] The U.S. Environmental Protection Agency has established a permissible level of 1 μg L^{-1} for all common phenolic compounds,[147] and the European Community (EC) Directive specifies a legal permissible level of 0.5 μg L^{-1} for phenolic compounds that are intended for human consumption.[148] Owing to their toxicity and persistence in the environment and in foods, the high resolution detection and quantification of phenolic compounds are of principal importance.[149,150] A variety of biological components, including microorganisms, enzymes, antibodies, antigens, nucleic acids, etc. may be used in the fabrication of biosensors for the detection and determination of phenolic compounds. Tyrosinase, peroxidase, laccase, glucose dehydrogenase, cellobiose dehydrogenase etc., are most commonly used enzymes for the fabrication of phenolic compound biosensors.

An enzymatic amperometric biosensor that was based on horseradish-peroxidase (HRP) and methylene blue (MB),[151] with chitosan on Au-modified TiO_2NT arrays, for the detection of trophenol, as shown in Fig. 8A. The dimensions of the TiO_2NT that were fabricated using anodization, were typically ~90 nm outer diameter, ~70 nm inner diameter, with a ~20 nm wall-thickness, as shown in Fig. 8B. The effective immobilization of biomolecules may be obtained by cumulative adhesion and
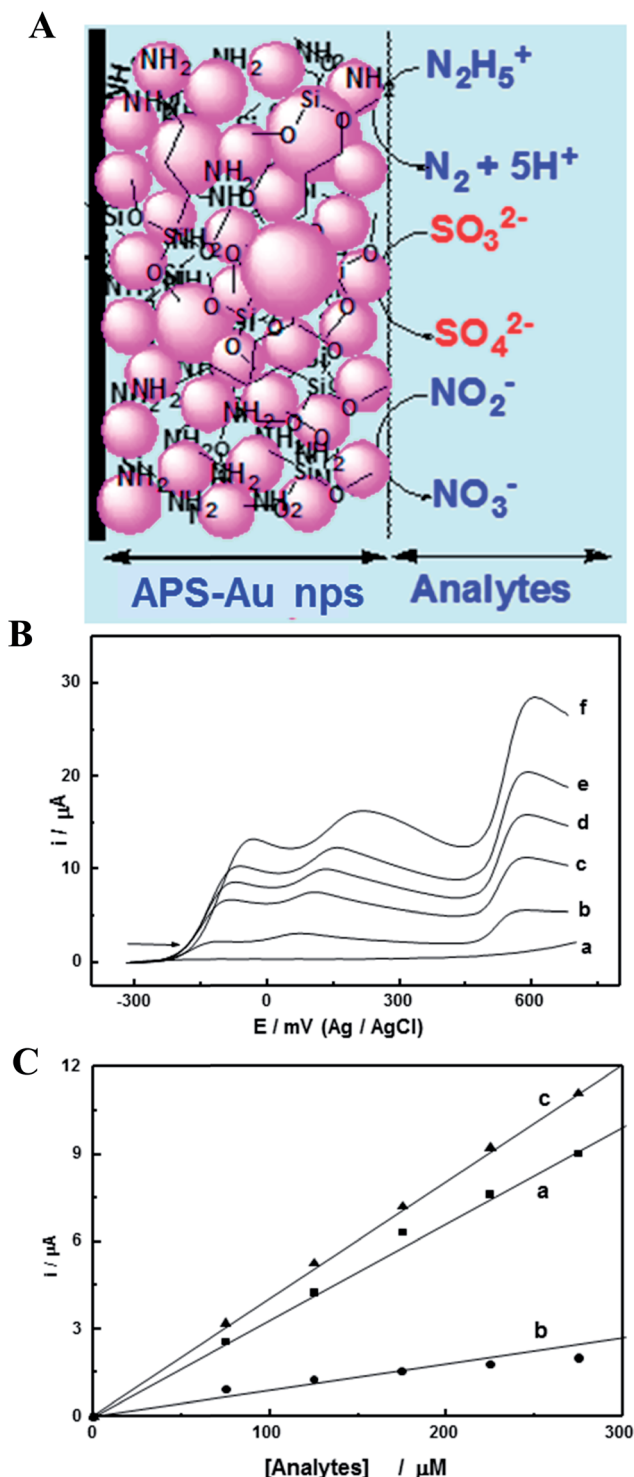
A

B

C

Fig. 7 (A) Schematic representation of an APS−Au NPs modified GC electrode and simultaneous electrocatalytic oxidation of N_2H_4, SO_3^{2-} and NO_2^-. (B) LSVs recorded at APS−AuNPs modified GC electrode for the mixture of N_2H_4, SO_3^{2-} and NO_2^- with the successive addition of concentrations in 0 (a), 200 μM (b), 300 μM (c), 400 μM (d), 500 μM (e) and 600 μM (f) in 0.1 MPBS (pH 7.2). (C) Corresponding calibration plots for N_2H_4 (a), SO_3^{2-} and NO_2^- (c). Reproduced with permission from ref. 129 © 2007 Elsevier.

integrity of the prepared TiO_2NT layer. Fig. 8C depicts the amperometric responses of three different electrodes (a–c) in the presence of 20 μM H_2O_2, for the addition of 3-nitrophenol, and the response of the TiO_2/Au/HRP–MB electrode was much higher than that of TiO_2NT/HRP and TiO_2NT/Au/HRP electrodes. The fabricated sensor in this study exhibited a linear response in the range from 0.3 μM to 120 μM for 3-nitrophenol (Fig. 8D), with a detection limit of 90 nM.[151] Hernandez et al.[152] demonstrated the electrochemical detection of phenolic compounds in environmental samples based on the horseradish peroxidase enzymatic reaction, by means of SWV. This sensor was coupled with a multivariate calibration method for the detection of five phenolic compounds including phenol, p-aminophenol, p-chlorophenol, hydroquinone and pyrocatechol. The LODs oscillated from 0.6 to 1.4 μM, based on the calibration model. A sensitive amperometric biosensor based on a GR–silkpeptide/tyrosinase nanointerface was developed for the detection and quantification of phenolic compounds.[153] The LODs were calculated as 0.23, 0.35 and 0.72 nM for catechol, phenol and BPA, respectively, with good repeatability and long-term stability. This group also tested preliminarily practical applications in the detection and determination of BPA that leaches from plastic drinking containers. The immobilizing of tyrosinase on poly(thionine)/nafion/MWCNT composite film electrode was fabricated for matairesinol biosensor. The CNTs uniformly integrated in the film formed a 3D electron conductive network and enabled the low detection limit of 37 nM.[154] The detection of phenolic compounds in plants using various electrochemical methods have been reviewed by Dobes et al.,[155] and the presence of phenolic compounds in foods was cataloged.

A non-enzymatic sensor was developed for the detection of p-nitrophenol using a poly(methylene blue)-modified GC electrode with a detection limit of 90 nM.[156] The proposed sensor was applied to detect p-nitrophenol in real water samples using a DPV method. TiO_2-modified graphene nanocomposite was used for the detection of p-chlorophenol with a low detection limit of 0.02 μM.[157] Furthermore, an electrogenerated chemiluminescent sensor was demonstrated,[158] which was based on a hybrid GR/MWCNT/Au nanocluster, for the detection and determination of phenolic compounds. The sensor signal increased linearly with elevated concentrations of catechol, ranging from 1.0 to 80 μM, with a detection limit of 0.3 μM. Moreover, the prepared sensor was applied to other phenolic compounds, such as hydroquinone, resorcinol, p-cresol, p-chlorophenol and 2-bromophenol, and exhibited a similar enhancement effect on the ECL at the GR/MWCNT/AuNCs/GC; however, it cannot be applied for the simultaneous/multiple determination and discrimination of these compounds using this method. The composite of MWCNT–poly(diphenylamine) modified GC electrode was used for the amperometric detection of p-nitrophenol in pH 7.[159] The composite electrode showed a good electrocatalytic reduction of p-nitrophenol, and exhibited a linear range from 8.9 μM to 1.43 mM with a sensitivity of 0.6322 μA μM^{-1} cm^{-2}. A chitosan–Fe_3O_4 nanocomposite based sensor was fabricated for the detection and determination of bisphenol-A (BPA) using a DPV method. Chitosan is a
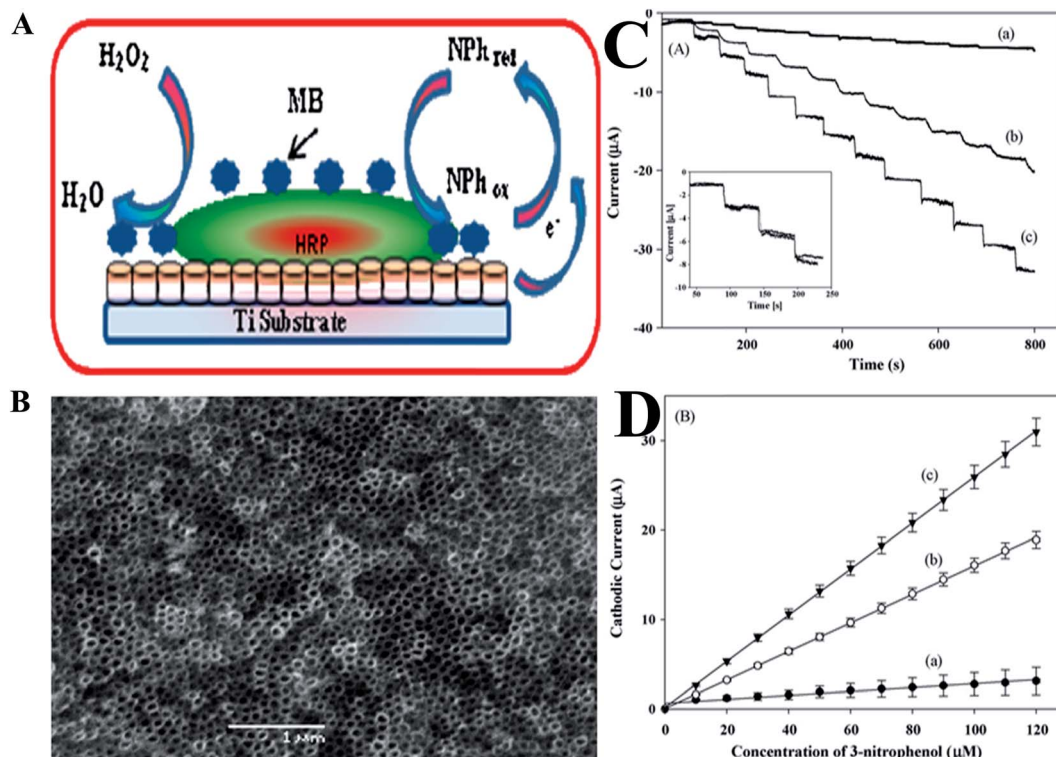
Fig. 8 (A) Scheme for designing a biosensor for nitrophenol detection. (B) SEM image of TiO$_2$NT arrays grown directly on Ti substrates. (C) Amperometric response of TiO$_2$NT/HRP (a), TiO$_2$NT/Au/HRP (b), and TiO$_2$NT/Au/HRP−MB (c) on Ti electrodes with the addition of 10 μM 3-nitrophenol in 0.1 MPBS (pH 7.0) in the presence of 20 μM H$_2$O$_2$, E_{app} = −0.05 V. Inset: amperometric response of the TiO$_2$NT/Au/HRP−MB electrode following 45 days under identical experimental conditions (solid line); (D) calibration plots of the proposed sensors for 3-nitrophenol. Reproduced with permission from ref. 151 © 2009 Elsevier.

polyaminosaccharide, which provides for a stable and homogenous suspension of Fe$_3$O$_4$ due to their chemical structures during sensor fabrication.[160] Yang *et al.*[161] prepared an electrochemical sensor based on Au NPs–GR nanohybrid bridged 3-amino-5-mercapto-1,2,4-triazole-functionalized MWCNT for the simultaneous determination of hydroquinone, catechol, resorcinol, and nitrite, as shown in Fig. 9. Due to the synergistic effects that exist between MWCNT–SH and Au NPs–GR, along with the excellent electrocatalytically active film forming ability of MWCNT–SH@Au–GR, the prepared MWCNT–SH@Au–GR sensor was used for the simultaneous determination of the toxic compounds, hydroquinone, catechol, resorcinol, and nitrite with LODs of 4.17 μM, 1.00 μM, 7.80 μM and 23.5 μM, respectively. The proposed simultaneous sensor showed excellent sensitivity, high selectivity and good stability with practical applications in the assessment of water samples. Nanostructured materials based electrochemical sensing platforms for phenolic compounds including phenylamine,[162] *o* and *p*-aminophenol,[163] polychlorinate biphenyl,[164] catechol,[165–169] hydroquinone,[166,167,170] *p*-chlorophenol,[171] bisphenol A,[172] phenol,[168] 1,3,5-trinitrophenol,[173] nitrobenzene,[173] are also comprised and listed in Table 1.

4.4. Detection of pesticides

Pesticides are regularly employed in agriculture worldwide, leading to the widespread contamination of air, water, soil and agricultural products. Due to their ability to accumulate, coupled with their potentially long-term effects on living organisms, pesticides are considered as the most dangerous of environmental contaminants.[174] In particular, the presence of pesticides in the environment is especially hazardous, leading to several serious human health problems such as asthma attacks, skin rashes, chronic disorders, and neurological diseases.[175] Intense research efforts have been invested in the development of sensitive and selective sensors for the detection and quantification of pesticides.[176] Details on nanomaterials based electrochemical and optical sensors for the detection of pesticides and pathogens may be found in the comprehensive review articles authored by Merkoc and co-workers.[177,178] State-of-art receptor selection, the use of different transduction techniques, rapid screening strategies, and the application of various biosensors have been developed for food and environmental safety and security applications. An important principle for the development of pesticide biosensors is derived from the correlation between the toxicity of pesticides and decreased enzymatic activity. The quantitative measurement of enzymatic activity, prior to, and following exposure to a target analyte relies on the development of biosensors, in particular, (i)
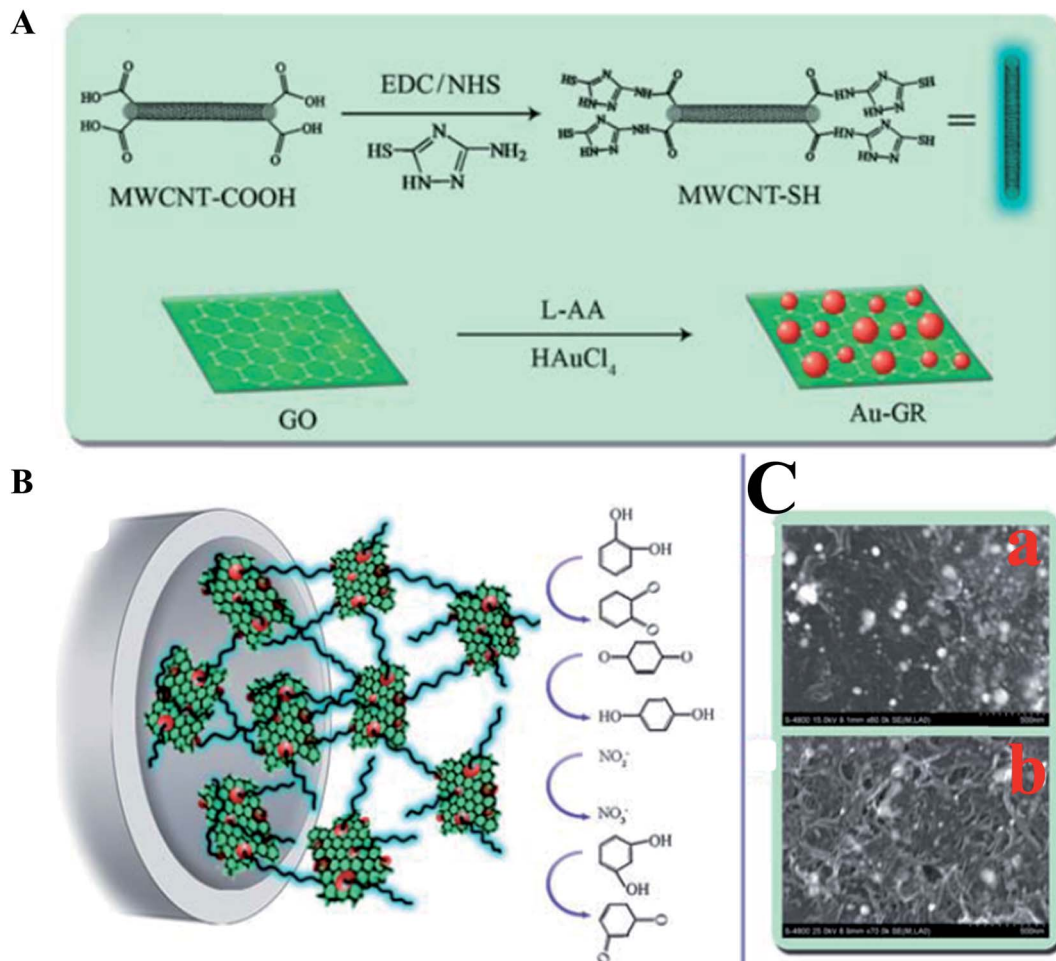
Fig. 9 (A) Preparation process for MWCNT−SH and Au−GR. (B) Schematic depiction of the MWCNT−SH@Au−GR electrode. (C) SEM images of Au−GR nanohybrid (a) and MWCNT−SH@Au−GR hybrid material (b). Reproduced with permission from ref. 161 © 2013 Royal Society of Chemistry.

determination of initial enzymatic activity, (ii) incubation of a biosensor in a solution that contains pesticides, and (iii) final measurement of residual activity. Amperometry and potentiometric transduction techniques are typically utilized to measure the activity of biosensing systems.[179,180]

Acetylcholinesterase (AChE) based biosensors have been reviewed by Hooda and co-workers[169] for the electrochemical detection of organophosphorus compound based pesticides. These researchers discussed a number of characteristic AChE based biosensor features, in terms of fabrication, detection limits, linearity range, incubation time, and storage stability. AChE based biosensors for the detection of OP compounds is an intensive field of research, with myriad applications for environmental monitoring, human health concern, and food industries. An electrochemical sensing platform was reported for the highly sensitive detection of organophosphate pesticides, based on carbon nanotube assemblies deposited on a nanoporous gold electrode, as shown in Fig. 10.[181] The
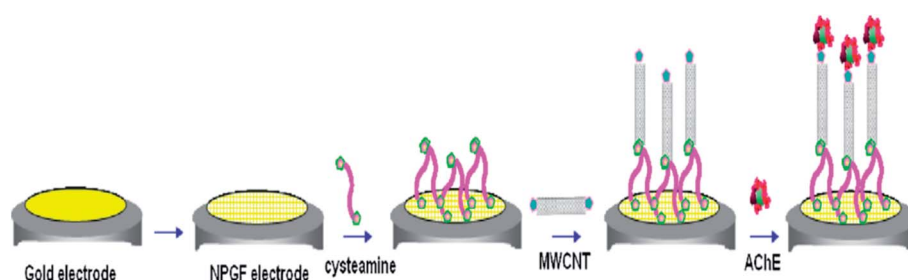

Fig. 10 Scheme for the fabrication of AChE−MWCNT−CA−NPG electrode. Reproduced with permission from ref. 181 © 2014 Elsevier.

Fig. 11 Schematic illustration of fabrication of simultaneous electrochemical immunosensor array for endosulfan and paraoxon. Reproduced with permission from ref. 185 © 2014 Elsevier.

immobilized AChE based biosensor model showed excellent activity in relation to its substrate, and enabled the quantitative measurement of organophosphate pesticides in the range from 0.001 to 0.5 μg mL^{-1} with a LOD of 0.5 ng mL^{-1}. This sensitive electrochemical AChE biosensor, comprised of polyaniline (PANI) and a MWCNT core–shell modified glassy carbon electrode, demonstrated the detection of carbamate pesticides in fruits and vegetables (apple, broccoli and cabbage).[182]

CNT/conductive polymer composites have exhibited a synergetic effect, leading to the augmentation of the electronic and mechanical characteristics of the constituent components.[183] An electrochemical carbofuran pesticide biosensor has been reported recently, which was based on esterases from Eupenicillium shearii FREI-39 endophytic fungi.[184] This enzymatic biosensor was prepared using the physical adsorption of the isolated endophytic fungus Eupenicillium shearii FREI-39 esterase on halloysite clay using graphite powder, MWCNT and mineral oil, for the determination of carbofuran pesticides *via* the inhibition of the esterase using SWV. A linear plot was obtained for carbofuran detection in the range from 5.0 to 100.0 μg L^{-1} with a LOD of 1.69 μg L^{-1}. Fig. 11 depicts the preparation of an electrochemical immunosensor based on an assembly of SWNT that was formed *via* micro contact printing (MCP) on GC substrates for the simultaneous detection of endosulfan and paraoxon.[185] Ferrocenedimethylamine (FDMA) and pyrroloquinoline quinone (PQQ) redox probes were loaded into a four-channel poly(dimethylsiloxane) (PDMS) patterning template. The change in the environment, due to the attenuation of electron transfer to the redox molecules and the association and dissociation of antibodies at the sensing interface, causes the modulation of the electrochemical behaviors of the redox molecules. This strategy has potential to lead to the design of portable sensing devices for the on-site detection of pesticides in environmental monitoring. A hybrid copper oxide nanowire (CuO NW)/SWCNT based non-enzymatic electrochemical sensor was developed by Huo et al.[186] for the detection of organophosphorus pesticides (OPs). A DPV technique was applied for a malathion sensor that showed a wide dynamic

range, with a LOD of 0.1 ppb (0.3 nM), with good selectivity against other common pesticides, inorganic ions and sugars, as well as good stability and reproducibility. A potentiometric biosensor was described for the pesticide atrazine, using urease biomolecules that were immobilized onto an insulator–semi-conductor electrode with various additional materials, such as glutaraldehyde (as a cross-linking agent), bovine serum albumin, coated Fe_3O_4 nanoparticles, cationic poly(allylamine hydrochloride), and anionic poly(sodium 4-styrenesulfonate) polyelectrolytes, as shown in Fig. 12.[187] The sensing of atrazine was enabled *via* inhibitory activity on the urease biosensor, with a LOD of 0.13 μM. Some other recently demonstrated electrochemical sensors and biosensors for the detection and determination of pesticides such as paraoxon,[188,189] parathion,[188,189] methyl parathion,[188,190–193] malathion,[144,194] carbofuran,[195,196] and carbaryl[195] are contained in Table 1.

4.5. Detection of chemical warfare agents

Chemical warfare agents (CWAs) are toxic chemicals that have been used historically in an effort to gain military superiority. They have been utilized on the battlefield as lachrymatory agents (causing respiratory irritation) during the World War I, and by the French and German armies in recent modern history.[197] The discovery of far more potent agents has rendered these early non-lethal chemicals obsolete as warfare agents, and thus they are currently used in riot control applications. Derived from the toxic effects of CWAs on humans, CWAs are classified as seven types: nerve, blister or vesicant, chocking or pulmonary, vomiting, asphyxiant or blood, tear or lachrymatory, and incapacitating agents.[198] The periodic use of chemical weapons of war and terror over history, has created an urgent need for the development of rapid CWA detection devices. Prior to the World War II, the presence of CWA vapors or other toxic chemicals, was detected *via* biological indicators, such as the human nose, or through the use of small animals. Modern approaches employ physical-electronic devices in their operation. Electrochemical systems are typically faster, more selective and
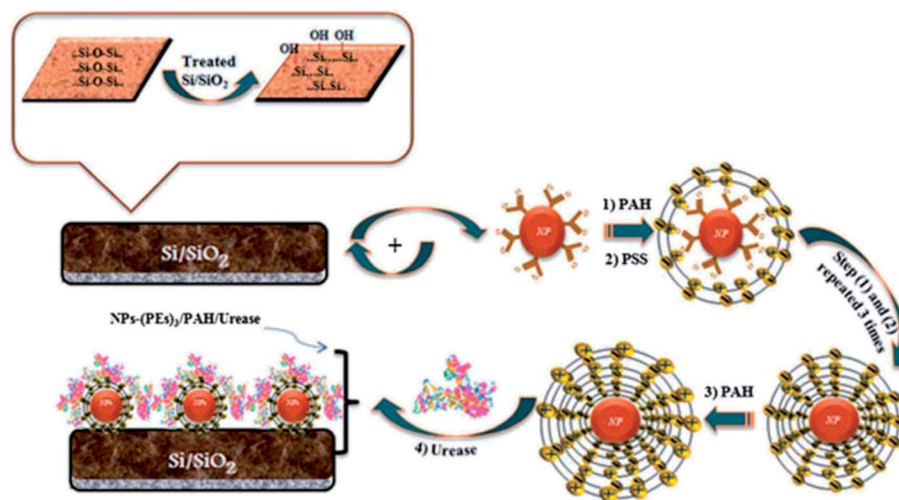
Fig. 12 Biosensor construction using Fe_3O_4 and polyelectrolyte materials for atrazine detection. Reproduced with permission from ref. 187 © 2013 Royal Society of Chemistry.

flexible, as they may be adapted to detect multiple compounds or entire classes of compounds. In USA, a miniaturized CWA detector, dubbed as an individual chemical agent detector (ICAD), has been developed through electrochemical methods. This detector consists of a reusable electronic module (processor, audio alarm, and warning light) and a disposable sensor module that contains a battery and the sensor cells. Black[199] undertook a brief historical overview of the development of bioanalytical methods for CWA, with a detailed summary of developing methods for the detection of nerve agents. Bhowmick et al.[200] reviewed the detection of chemical warfare agents using surface-immobilized molecules with advanced functionalities. The design and fabrication of smart surfaces may have the capacity to immobilize functional molecules, which perform specific functions under the input of
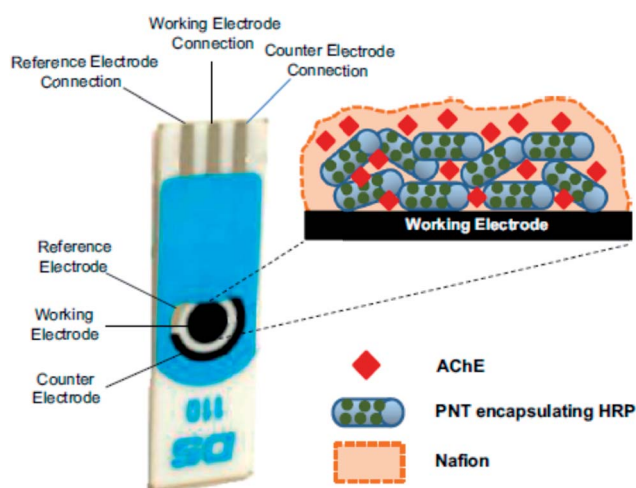

Fig. 13 Schematic illustration of the peptide nanotube (PNT)-modified gold electrode containing encapsulated HRP and the surface attached AChE enzymes. Reproduced with permission from ref. 202 © 2014 Elsevier.

external stimuli, and describes their adequacy as sensors for the detection of CWAs in environmental applications.

A biosensor was reported that was based on cholinesterase inhibition,[201] which was assembled on a miniaturized prototype for a paraoxon nerve agent analog. Nerve agents such as Sarin, exhibit more robust inhibitory behaviors than paraoxon, thus the system might be suitable for the detection of lower concentrations of nerve agents. The OP biosensor was developed on the basis of peptide nanotubes (PNTs), and encapsulated horseradish peroxidase, surface coated with AChE, which were attached to gold screen printed electrodes, as shown in Fig. 13.[202] In this biosensor, AChE catalyzed the hydrolysis of ATCh to produce thiocholine, which is then oxidized by HRP to produce dithiocholine. PNTs facilitated direct electron transfer between the HRP and the electrode. The prepared biosensor had the capacity to detect OP vapors at low concentrations below 25 ppb. A multidimensional FeO–OH nanoneedle-decorated hybrid polypyrrole nanoparticle (PFF) based electrochemical sensor was demonstrated using a dual nozzle electrospray and heat stirring process for the detection of chemical nerve gas agents.[203] A simple amperometric biosensor was fabricated for sarin using Prussian blue modified silver screen-printed electrode was developed using butyrylthiocholine (BTCh) and butyrylcholinesterase (BChE), in the presence of sarin, using an amperometric method with a LOD of 12 ppb.[204] Liu et al.[205] have developed an indirect detection biosensor using AChE with an amperometric method to monitor the decrease in choline concentrations due to CWA exposure. A variety of nanomaterials based electrochemical sensors for the detection CWA including sarin,[205,206] diisopropyl fluorophosphate,[206] dichlorvos,[207] paraoxon-ethyl,[208] isoproturon,[209] and carbendazim,[209] are summarized in Table 1.

5.   Conclusions and perspectives

This review has discussed and highlighted recent advancements in nanomaterials based electrochemical sensors for

environmental analyses and food safety applications. Electro-chemical sensors based on nanostructured materials provide a crucial analytical tool, as the demand for the ultrasensitive, rapid, and selective determination of analytes increases. They may be easily adapted to selective and multiple sensing a range of chemical contaminants inexpensively, in contrast to spectroscopic and chromatographic instrumentation. Some unique physicochemical and electrochemical properties of nanomaterials may facilitate the fabrication of potentially robust electrochemical sensor systems, with attributes such as good biocompatibility and outstanding electrical conductivity. The architectures of nanometric interfaces also afford a suitable environment for biomolecule immobilization, overpotential decreases, and improvements in sensitivity and long-term stability. Additionally, nanomaterial enabled sensors are capable of being embedded into/integrated with portable or miniaturized devices for specific applications, due to the selective catalytic activity, stability and biocompatibility of the sensor design. The design of sensitive and selective simultaneous electrochemical sensor strategies for chemical contaminants that are present in the environment and in foods also warrants exploitation.

Recent advances in synthesis of nanomaterials and electrochemical analytical methods provide a strong and promising future toward the realization of sensitive analytical devices for sensor researchers and manufacturers. The production and commercialization of low-cost nanomaterials based sensing devices remain key issues in the development of analytical devices for the environmental monitoring and food safety industries. The combination of controlled nanostructured architectural design, nanomaterials synthesis, sensor fabrication, processing, integration, miniaturization and testing techniques may offer a possible solution to existing analytical challenges in the fabrication of practical sensing devices.

One of the core issues that requires immediate resolution toward the development of future advanced, highly selective and accurate, portable/miniaturized, rapid (ideally real time), multi-detection environmental and food monitoring sensors, is the significant bottleneck that remains between research laboratory benchtop prototypes and their widespread commercial applications. The highly complex nature of myriad contaminants that increasingly continue to be released into the ambient environment and in foods, will require sophisticated and versatile sensing platforms that have an extremely high degree of selectivity and sensitivity. In the ambient environment, many chemicals have the capacity to combine with others to further exacerbate existing challenges in regard to their precise identification and toxicity assessment. Therefore, extensive collaborative efforts between multidisciplinary science and engineering disciplines (*e.g.*, environmental sciences, nanosciences, limnology, physics, chemistry, electrochemistry, mechanical engineering, electronics, software development, *etc.*), will be indispensible, and indeed, are urgently warranted for the successful design, development, and implementation of optimized, "market ready" sensors. No longer can these disciplines operate in isolation in view of the immense and highly complex challenges that we presently confront. With continual

and rapid advancements across many facets of sensor development, it is anticipated that unprecedented breakthroughs in the emergence of nanomaterials based sensing devices will be realized to seriously address the widespread chemical contamination of the environment and foods in the near future.

Acknowledgements

References

1 TSCA Chemical Substance Inventory, Section 8(b), United States Environmental Protection Agency, 2014.
2 V. I. Pye and R. Patrick, *Science*, 1983, **221**, 713.
3 L. Kantiani, M. Llorca, J. Sanchis, M. Farre and D. Barcelo, *Anal. Bioanal. Chem.*, 2010, **398**, 2413.
4 E. Ibanez and A. Cifuentes, *Crit. Rev. Food Sci. Nutr.*, 2001, **41**, 413.
5 M. A. Hamburg, *Science*, 2011, **331**, 987.
6 V. Garcia-Canâas, C. Simo, M. Herrero, E. Ibanâez and A. Cifuentes, *Anal. Chem.*, 2012, **84**, 10150.
7 A. A. Dudarev, E. V. Dushkina, Y. N. Sladkova, P. R. Alloyarov, V. S. Chupakhin, V. M. Dorofeyev, T. A. Kolesnikova, K. B. Fridman, B. Evengard and L. M. Nilsson, *Int. J. Circumpolar Health*, 2013, **72**, 1.
8 S. Rodriguez-Mozaz, M. P. Marco, M. J. L. de Alda and D. Barcelo, *Anal. Bioanal. Chem.*, 2004, **378**, 588.
9 G. Hanrahan, D. G. Patil and J. Wang, *J. Environ. Monit.*, 2004, **6**, 657.
10 J. Parellada, A. Narváez, M. A. López, E. Domínguez, J. J. Fernández, V. Pavlov and I. Katakis, *Anal. Chim. Acta*, 1998, **362**, 47.
11 A. Baeumner, *Anal. Bioanal. Chem.*, 2003, **377**, 434.
12 R. Chen, Y. Li, K. Huo and P. K. Chu, *RSC Adv.*, 2013, **3**, 18698.
13 A. Bahrami, A. Besharati-Seidani, A. Abbaspour and M. Shamsipur, *Electrochim. Acta*, 2014, **118**, 92.
14 J. Ma, L. Adornato, R. H. Byrne and D. Yuan, *TrAC, Trends Anal. Chem.*, 2014, **60**, 1.
15 B. Chen, Z. Wang, D. Hu, Q. Ma, L. Huang, C. Xv, Z. Guo and X. Jiang, *Sens. Actuators, B*, 2014, **200**, 310.
16 A. N. Goldstein, C. M. Echer and A. P. Alivisatos, *Science*, 1992, **256**, 1425.
17 S. E. F. Kleijn, S. C. S. Lai, M. T. M. Koper and P. R. Unwin, *Angew. Chem., Int. Ed.*, 2014, **53**, 3558.
18 V. J. Nagaraj, M. Jacobs, K. M. Vattipalli, V. P. Annam and S. Prasad, *Environ. Sci.: Processes Impacts*, 2014, **16**, 135.
19 J. Kirsch, C. Siltanen, Q. Zhou, A. Revzin and A. Simonian, *Chem. Soc. Rev.*, 2013, **42**, 8733.
20 R. X. Xu, X. Y. Yu, C. Gao, J. H. Liu, R. G. Compton and X. J. Huang, *Analyst*, 2013, **138**, 1812.

21 D. Tang, J. Tang, B. Su and G. Chen, *J. Agric. Food Chem.*, 2010, **58**, 10824.

22 E. Bakker, *Anal. Chem.*, 2004, **76**, 3285.

23 A. Hierlemann and R. Gutierrez-Osuna, *Chem. Rev.*, 2008, **108**, 563.

24 M. Bonizzoni and E. V. Anslyn, *J. Am. Chem. Soc.*, 2009, **131**, 14597.

25 M. Amalraj and S. A. John, *Electrochim. Acta*, 2014, **117**, 360.

26 H. S. Toh, K. Tschulik, C. Batchelor-McAuley and R. G. Compton, *Analyst*, 2014, **139**, 3986.

27 J. P. Metters, R. O. Kadara and C. E. Banks, *Analyst*, 2011, **136**, 1067.

28 J. Wang, *TrAC, Trends Anal. Chem.*, 2002, **21**, 226.

29 A. Baeumner, *Anal. Bioanal. Chem.*, 2003, **377**, 434.

30 F. Valentini, M. Carbone and G. Palleschi, *Anal. Bioanal. Chem.*, 2013, **405**, 3449.

31 D. A. C. Brownson and C. E. Banks, *Analyst*, 2010, **135**, 2768.

32 W. Zhang, A. M. Asiri, D. Liu, D. Du and Y. Lin, *TrAC, Trends Anal. Chem.*, 2014, **54**, 1.

33 D. W. Kimmel, G. LeBlanc, M. E. Meschievitz and D. E. Cliffel, *Anal. Chem.*, 2011, **84**, 685.

34 A. Chen and S. Chatterjee, *Chem. Soc. Rev.*, 2013, **42**, 5425.

35 A. Chen and B. Shah, *Anal. Methods*, 2013, **5**, 2158.

36 A. Chen and P. Holt-Hindle, *Chem. Rev.*, 2010, **110**, 3767.

37 B. Shah, T. Lafleur and A. Chen, *Faraday Discuss.*, 2013, **164**, 135.

38 S. Chatterjee, J. Wen and A. Chen, *Biosens. Bioelectron.*, 2013, **42**, 349.

39 A. Ahmadalinezhad, G. Wu and A. Chen, *Biosens. Bioelectron.*, 2011, **30**, 287.

40 S. Chatterjee and A. Chen, *Biosens. Bioelectron.*, 2012, **35**, 302.

41 S. Chatterjee and A. Chen, *Anal. Chim. Acta*, 2012, **751**, 66.

42 A. K. Wanekaya, *Analyst*, 2011, **136**, 4383.

43 K. Scida, P. W. Stege, G. Haby, G. A. Messina and C. D. García, *Anal. Chim. Acta*, 2011, **691**, 6.

44 C. Gao, Z. Guo, J.-H. Liu and X.-J. Huang, *Nanoscale*, 2012, **4**, 1948.

45 J. Ping, Y. Wang, J. Wu and Y. Ying, *Electrochem. Commun.*, 2011, **13**, 1529.

46 E. Jaworska, W. Lewandowski, J. Mieczkowski, K. Maksymiuk and A. Michalska, *Analyst*, 2012, **137**, 1895.

47 X. Gong, Y. Bi, Y. Zhao, G. Liu and W. Y. Teoh, *RSC Adv.*, 2014, **4**, 24653.

48 S. Yuan, Q. He, S. Yao and S. Hu, *Anal. Lett.*, 2006, **39**, 373.

49 I. Balan, I. G. David, V. David, A. I. Stoica, C. Mihailciuc, I. Stamatin and A. A. Ciucu, *J. Electroanal. Chem.*, 2011, **654**, 8.

50 L. G. Zamfir, L. Rotariu and C. Bala, *Biosens. Bioelectron.*, 2011, **26**, 3692.

51 L. S. Fifield and J. W. Grate, *Carbon*, 2010, **48**, 2085.

52 Y. Wang and Y. Hasebe, *Anal. Bioanal. Chem.*, 2011, **399**, 1151.

53 W. Tu, J. Lei, L. Ding and H. Ju, *Chem. Commun.*, 2009, 4227.

54 H. Daia, L. Gong, G. Xu, S. Zhang, S. Lu, Y. Jiang, Y. Lin, L. Guo and G. Chen, *Electrochim. Acta*, 2013, **111**, 57.

55 L. Rassaei, F. Marken, M. Sillanpaa, M. Amiri, C. M. Cirtiu and M. Sillanpaa, *TrAC, Trends Anal. Chem.*, 2011, **30**, 1704.

56 L. Cui, J. Wu and H. Ju, *Biosens. Bioelectron.*, 2015, **63**, 276.

57 K. Warriner, S. M. Reddy, A. Namvar and S. Neethirajan, *Trends Food Sci. Technol.*, 2014, DOI: 10.1016/j.tifs.2014.07.008.

58 A. Amine, H. Mohammadi, I. Bourais and G. Palleschi, *Biosens. Bioelectron.*, 2006, **21**, 1405.

59 G. Maduraiveeran and R. Ramaraj, *J. Electroanal. Chem.*, 2007, **608**, 52.

60 D. W. Kimmel, G. LeBlanc, M. E. Meschievitz and D. E. Cliffel, *Anal. Chem.*, 2011, **84**, 685.

61 P. S. R. D. Silva, B. C. Gasparini, H. R. A. Magosso and A. Spinelli, *J. Hazard. Mater.*, 2014, **273**, 70.

62 Y. Wei, R. Yang, X. Y. Yu, L. Wang, J. H. Liu and X. J. Huang, *Analyst*, 2012, **137**, 2183.

63 S. Radhakrishnan, K. Krishnamoorthy, C. Sekar, J. Wilson and S. J. Kim, *Appl. Catal., B*, 2014, **148**, 22.

64 B. Kumar Jena and C. Retna Raj, *Anal. Chem.*, 2008, **80**, 4836.

65 A. Chen, *Can. J. Chem.*, 2014, **92**, 581.

66 P. Benvenuto, A. K. M. Kafi and A. Chen, *J. Electroanal. Chem.*, 2009, **627**, 76.

67 A. K. M. Kafi, G. Wu and A. Chen, *Biosens. Bioelectron.*, 2008, **24**, 566.

68 D. Reyter, D. Bélanger and L. Roué, *J. Electroanal. Chem.*, 2008, **622**, 64.

69 A. Ahmadalinezhad and A. Chen, *Biosens. Bioelectron.*, 2011, **26**, 4508.

70 W. Jin, G. Wu and A. Chen, *Analyst*, 2014, **139**, 235.

71 X. Chen, W. Liu, L. Tang, J. Wang, H. Pan and M. Du, *Mater. Sci. Eng., C*, 2014, **34**, 304.

72 E. Sennik, N. Kilinc and Z. Z. Ozturk, *J. Alloys Compd.*, 2014, **616**, 89.

73 E. Buhleier, W. Wehner and F. Vogtle, *Synthesis*, 1978, **1978**, 155.

74 R. M. Crooks, M. Zhao, L. Sun, V. Chechik and L. K. Yeung, *Acc. Chem. Res.*, 2000, **34**, 181.

75 M. Hasanzadeh, N. Shadjou, M. Eskandani, J. Soleymani, F. Jafari and M. de la Guardia, *TrAC, Trends Anal. Chem.*, 2014, **53**, 137.

76 P. P. Fang, O. Buriez, E. Labbé, Z. Q. Tian and C. Amatore, *J. Electroanal. Chem.*, 2011, **659**, 76.

77 D. Ning, H. Zhang and J. Zheng, *J. Electroanal. Chem.*, 2014, **717**, 29.

78 A. Siriviriyanun, T. Imae and N. Nagatani, *Anal. Biochem.*, 2013, **443**, 169.

79 H. Yoon, *Nanomaterials*, 2013, **3**, 524.

80 W.-K. Oh, O. S. Kwon and J. Jang, *Polym. Rev.*, 2013, **53**, 407.

81 J. Kirsch, C. Siltanen, Q. Zhou, A. Revzin and A. Simonian, *Chem. Soc. Rev.*, 2013, **42**, 8733.

82 H. J. Salavagione, A. M. Diez-Pascual, E. Lazaro, S. Vera and M. A. Gomez-Fatou, *J. Mater. Chem. A*, 2014, **2**, 14289.

83 M. Campas, D. Garibo and B. Prieto-Simon, *Analyst*, 2012, **137**, 1055.

84 J. P. Metters, R. O. Kadara and C. E. Banks, *Analyst*, 2012, **137**, 896.

85 Z. Xiao-hong, S. Bao-dong, S. Han-chang, L. Lan-hua, G. Hong-li and H. Miao, *Sens. Actuators, B*, 2014, **198**, 150.

86 F. A. Esteve-Turrillas and A. Abad-Fuentes, *Biosens. Bioelectron.*, 2013, **41**, 12.

87 J. K. Fawell and G. Stanfield, in *Pollution: Causes, Effects and Control*, ed. R. M. Harrison, RSC, 2001, vol. 4, pp. 59–81.

88 *An Introduction to Pollution Science*, ed. R. M. Harrison, RSC, 2006, pp. 77–121.

89 T. F. McGrath, C. T. Elliott and T. L. Fodey, *Anal. Bioanal. Chem.*, 2012, **403**, 75.

90 S. Viswanathan, H. Radecka and J. Radecki, *Monatsh. Chem.*, 2009, **140**, 891.

91 S. J. Kim, K. V. Gobi, H. Iwasaka, H. Tanaka and N. Miura, *Biosens. Bioelectron.*, 2007, **23**, 701.

92 T. McGrath, A. Baxter, J. Ferguson, S. Haughey and P. Bjurling, *Anal. Chim. Acta*, 2005, **529**, 123.

93 J. Gong, T. Liu, X. Wang, X. Hu and L. Zhang, *Environ. Sci. Technol.*, 2011, **45**, 6181.

94 G. Aragay, J. Pons and A. Merkoçi, *Chem. Rev.*, 2011, **111**, 3433.

95 F. Fu and Q. Wang, *J. Environ. Manage.*, 2011, **92**, 407.

96 H. H. Harris, I. J. Pickering and G. N. George, *Science*, 2003, **301**, 1203.

97 I. Onyido, A. R. Norris and E. Buncel, *Chem. Rev.*, 2004, **104**, 5911.

98 Y. Zhang, G. M. Zeng, L. Tang, Y. P. Li, Z. M. Chen and G. H. Huang, *RSC Adv.*, 2014, **4**, 18485.

99 Z. Y. Kong, J. F. Wei, Q. J. Fan, Y. H. Li, K. Y. Zhao, H. T. Guo and C. M. Wei, *Anal. Methods*, 2013, **5**, 4905.

100 X. Gong, Y. Bi, Y. Zhao, G. Liu and W. Y. Teoh, *RSC Adv.*, 2014, **4**, 24653.

101 P. K. Aneesh, S. R. Nambiar, T. P. Rao and A. Ajayaghosh, *Phys. Chem. Chem. Phys.*, 2014, **16**, 8529.

102 L. E. Eary and D. Rai, *Environ. Sci. Technol.*, 1987, **21**, 1187.

103 L. McNeill, J. McLean, M. Edwards and J. Parks, in *State of the Science of Hexavalent Chromium in Drinking Water*, Water Research Foundation, 2012.

104 C. M. Welch, O. Nekrassova and R. G. Compton, *Talanta*, 2005, **65**, 74.

105 A. Deep, A. L. Sharma, S. K. Tuteja and A. K. Paul, *J. Hazard. Mater.*, 2014, **278**, 559.

106 M. Rajkumar, S. Thiagarajan and S. M. Chen, *Int. J. Electrochem. Sci.*, 2011, **6**, 3164.

107 L. Pujol, D. Evrard, K. Groenen-Serrano, M. Freyssinier, A. Ruffien-Cizsak and P. Gros, *Front. Chem.*, 2014, **2**, 19.

108 A. Safavi and E. Farjami, *Anal. Chim. Acta*, 2011, **688**, 43.

109 X. Liu, C. Sun, H. Wu, Y. Zhang, J. Jiang, G. Shen and R. Yu, *Electroanalysis*, 2010, **22**, 2110.

110 J. Gong, T. Zhou, D. Song, L. Zhang and X. Hu, *Anal. Chem.*, 2009, **82**, 567.

111 S. Tang, P. Tong, W. Lu, J. Chen, Z. Yan and L. Zhang, *Biosens. Bioelectron.*, 2014, **59**, 1.

112 S. Sadeghi and A. Garmroodi, *Mater. Sci. Eng., C*, 2013, **33**, 4972.

113 Y. Lian, M. Yuan and H. Zhao, *Fullerenes, Nanotubes, Carbon Nanostruct.*, 2014, **22**, 510.

114 A. Afkhami, H. Ghaedi, T. Madrakian and M. Rezaeivala, *Electrochim. Acta*, 2013, **89**, 377.

115 Z. Wang, H. Wang, Z. Zhang, X. Yang and G. Liu, *Electrochim. Acta*, 2014, **120**, 140.

116 R. A. Dar, N. G. Khare, D. P. Cole, S. P. Karna and A. K. Srivastava, *RSC Adv.*, 2014, **4**, 14432.

117 D. Wang, Y. Zhao, H. Jin, J. Zhuang, W. Zhang, S. Wang and J. Wang, *ACS Appl. Mater. Interfaces*, 2013, **5**, 5733.

118 H. Cui, W. Yang, X. Li, H. Zhao and Z. Yuan, *Anal. Methods*, 2012, **4**, 4176.

119 M. Noroozifar, M. Khorasani-Motlagh and A. Taheri, *J. Hazard. Mater.*, 2011, **185**, 255.

120 I. A. Wolff and A. E. Wasserman, *Science*, 1972, **177**, 15.

121 X.-H. Pham, C. A. Li, K. N. Han, B.-C. Huynh-Nguyen, T.-H. Le, E. Ko, J. H. Kim and G. H. Seong, *Sens. Actuators, B*, 2014, **193**, 815.

122 D. Zhang, H. Ma, Y. Chen, H. Pang and Y. Yu, *Anal. Chim. Acta*, 2013, **792**, 35.

123 S. M. da Silva and L. H. Mazo, *Electroanalysis*, 1998, **10**, 1200.

124 P. Miao, M. Shen, L. Ning, G. Chen and Y. Yin, *Anal. Bioanal. Chem.*, 2011, **399**, 2407.

125 A. Afkhami, F. Soltani-Felehgari, T. Madrakian and H. Ghaedi, *Biosens. Bioelectron.*, 2014, **51**, 379.

126 T. Madasamy, M. Pandiaraj, M. Balamurugan, K. Bhargava, N. K. Sethy and C. Karunakaran, *Biosens. Bioelectron.*, 2014, **52**, 209.

127 B. A. Molinero-Abad, M. A. N. Alonso-Lomillo, O. Dominguez-Renedo and M. J. Arcos-Martinez, *Anal. Chim. Acta*, 2014, **812**, 41.

128 M. Amatatongchai, W. Sroysee, S. Chairam and D. Nacapricha, *Talanta*, 2014, DOI: 10.1016/j.talanta.2014.07.055.

129 G. Maduraiveeran and R. Ramaraj, *Electrochem. Commun.*, 2007, **9**, 2051.

130 S. Liu, B. Yu, F. Li, Y. Ji and T. Zhang, *Electrochim. Acta*, 2014, **141**, 161.

131 D. d. Zhou, L. Ding, H. Cui, H. An, J. p. Zhai and Q. Li, *Chem. Eng. J.*, 2012, **200**, 32.

132 H. S. Weinberg, C. A. Delcomyn and V. Unnam, *Environ. Sci. Technol.*, 2003, **37**, 3104.

133 L. Xie and C. Shang, *Water Sci. Technol.: Water Supply*, 2006, **6**, 131.

134 A. Salimi, H. MamKhezri, R. Hallaj and S. Zandi, *Electrochim. Acta*, 2007, **52**, 6097.

135 P. Muthukumar and S. Abraham John, *J. Colloid Interface Sci.*, 2014, **421**, 78.

136 P. Li, Y. Ding, A. Wang, L. Zhou, S. Wei, Y. Zhou, Y. Tang, Y. Chen, C. Cai and T. Lu, *ACS Appl. Mater. Interfaces*, 2013, **5**, 2255.

137 A. Afkhami, F. Soltani-Felehgari, T. Madrakian and H. Ghaedi, *Biosens. Bioelectron.*, 2014, **51**, 379.

138 S. Radhakrishnan, K. Krishnamoorthy, C. Sekar, J. Wilson and S. J. Kim, *Appl. Catal., B*, 2014, **1489**, 22.

139 R. Rawal and C. S. Pundir, *Int. J. Biol. Macromol.*, 2012, **51**, 449.

140 S. Devaramani and P. Malingappa, *Electrochim. Acta*, 2012, **85**, 579.

141 A. A. Ensafi, H. Karimi-Maleh and M. Keyvanfard, *Int. J. Environ. Anal. Chem.*, 2013, **93**, 650.

142 L. Wang, P. Hu, X. Deng, F. Wang and Z. Chen, *Biosens. Bioelectron.*, 2013, **50**, 57.

143 D. Zheng, X. Liu, D. Zhou and S. Hu, *Microchim. Acta*, 2012, **176**, 49.

144 S. Jayabal, P. Viswanathan and R. Ramaraj, *RSC Adv.*, 2014, **4**, 33541.

145 R. Gatermann, H. Hühnerfuss, G. Rimkus, M. Wolf and S. Franke, *Mar. Pollut. Bull.*, 1995, **30**, 221.

146 F. Karim and A. N. M. Fakhruddin, *Rev. Environ. Sci. Biotechnol.*, 2012, **11**, 261.

147 U.S. EPA Title 40, Chapter 1, Part 141, National Primary Drinking Water Regulation, http://www.law.cornell.edu/cfr/text/40/141.11, accessed 17 August, 2014.

148 Drinking Water Directive 80/778/EEC, Commission of the European Communities, 1980.

149 T. C. Canevari, P. A. Raymundo-Pereira, R. Landers and S. A. S. Machado, *Eur. J. Inorg. Chem.*, 2013, 5746.

150 J. S. Lee, S. G. Kim, J. Jun, D. H. Shin and J. Jang, *Adv. Funct. Mater.*, 2014, **24**, 6145.

151 A. K. M. Kafi and A. Chen, *Talanta*, 2009, **79**, 97.

152 S. R. Hernandez, S. V. Kergaravat and M. I. Pividori, *Talanta*, 2013, **106**, 399.

153 Y. Qu, M. Ma, Z. Wang, G. Zhan, B. Li, X. Wang, H. Fang, H. Zhang and C. Li, *Biosens. Bioelectron.*, 2013, **44**, 85.

154 J. A. Rather, S. Pilehvar and K. D. Wael, *Analyst*, 2013, **138**, 204.

155 J. Dobes, O. Zitka, J. Sochor, B. Ruttkay-Nedecky, P. Babula, M. Beklova, J. Kynicky, J. Hubalek, B. Klejdus and R. Kizek, *Int. J. Electrochem. Sci.*, 2013, **8**, 4520.

156 K. Giribabu, R. Suresh, R. Manigandan, S. Munusamy, S. P. Kumar, S. Muthamizh and V. Narayanan, *Analyst*, 2013, **138**, 5811.

157 X. Bai, X. Huang, X. Zhang, Z. Hu, C. Wang, Q. Qin and Q. Zhang, *RSC Adv.*, 2014, **4**, 13461.

158 D. Yuan, S. Chen, R. Yuan, J. Zhang and W. Zhang, *Analyst*, 2013, **138**, 6001.

159 Y. L. Yang, B. Unnikrishnan and S. M. Chen, *Int. J. Electrochem. Sci.*, 2011, **6**, 2902.

160 C. Yu, L. Gou, X. Zhou, N. Bao and H. Gu, *Electrochim. Acta*, 2011, **56**, 9056.

161 C. Yang, Y. Chai, R. Yuan, W. Xu and S. Chen, *Anal. Methods*, 2013, **5**, 666.

162 B. T. P. Quynh, J. Y. Byun and S. H. Kim, *Sens. Actuators, B*, 2014, **193**, 1.

163 B. Kaur and R. Srivastava, *Electrochim. Acta*, 2014, **141**, 61.

164 S. Pilehvar, J. Ahmad Rather, F. Dardenne, J. Robbens, R. Blust and K. De Wael, *Biosens. Bioelectron.*, 2014, **54**, 78.

165 D. Yuan, S. Chen, F. Hu, C. Wang and R. Yuan, *Sens. Actuators, B*, 2012, **168**, 193.

166 H. S. Han, J. M. You, H. Seol, H. Jeong and S. Jeon, *Sens. Actuators, B*, 2014, **194**, 460.

167 J. Du, L. Ma, D. Shan, Y. Fan, L. Zhang, L. Wang and X. Lu, *J. Electroanal. Chem.*, 2014, **722**, 38.

168 E. Cevik, M. Senel, A. Baykal and M. F. Abasiyanik, *Sens. Actuators, B*, 2012, **173**, 396.

169 H. Wu, J. Hu, H. Li and H. Li, *Sens. Actuators, B*, 2013, **182**, 802.

170 W. Tang, M. Zhang, W. Li and X. Zeng, *Talanta*, 2014, **127**, 262.

171 J. Zolgharnein, T. Shariatmanesh and A. Babaei, *Sens. Actuators, B*, 2013, **186**, 536.

172 L. Zhang, Y. P. Wen, Y. Y. Yao, Z. F. Wang, X. M. Duan and J. K. Xu, *Chin. Chem. Lett.*, 2014, **25**, 517.

173 C. X. Yuan, Y. R. Fan, Z. Tao, H. X. Guo, J. X. Zhang, Y. L. Wang, D. L. Shan and X. Q. Lu, *Biosens. Bioelectron.*, 2014, **58**, 85.

174 T. Pradeep, Anshup and M. S. Bootharaju, *Nanotechnology Applications for Clean Water*, 2nd edn, 2014, ch. 16, pp. 241–270.

175 M. Tankiewicz, J. Fenik and M. Biziuk, *TrAC, Trends Anal. Chem.*, 2010, **29**, 1050.

176 S. Liu, Z. Zheng and X. Li, *Anal. Bioanal. Chem.*, 2013, **405**, 63.

177 B. Perez-Lopez and A. Merkoc, *Trends Food Sci. Technol.*, 2011, **22**, 625.

178 G. Aragay, F. Pino and A. Merkoc, *Chem. Rev.*, 2012, **112**, 5317.

179 G. G. Guilbault, M. H. Sadar, S. S. Kuan and D. Casey, *Anal. Chim. Acta*, 1970, **52**, 75.

180 V. Dhull, A. Gahlaut, N. Dilbaghi and V. Hooda, *Biochem. Res. Int.*, 2013, 1.

181 J. Ding, H. Zhang, F. Jia, W. Qin and D. Du, *Sens. Actuators, B*, 2014, **199**, 284.

182 I. Cesarino, F. C. Moraes, M. R. Lanza and S. A. Machado, *Food Chem.*, 2012, **135**, 873.

183 L. Agüí, C. Peña-Farfal, P. Yáñez-Sedeño and J. M. Pingarrón, *Electrochim. Acta*, 2007, **52**, 7946.

184 G. F. Grawe, T. R. de Oliveira, E. de Andrade Narciso, S. K. Moccelini, A. J. Terezo, M. A. Soares and M. Castilho, *Biosens. Bioelectron.*, 2015, **63**, 407.

185 G. Liu, W. Guo and D. Song, *Biosens. Bioelectron.*, 2014, **52**, 360.

186 D. Huo, Q. Li, Y. Zhang, C. Hou and Y. Lei, *Sens. Actuators, B*, 2014, **199**, 410.

187 Y. Braham, H. Barhoumi and A. Maaref, *Anal. Methods*, 2013, **5**, 4898.

188 X. Tang, T. Zhang, B. Liang, D. Han, L. Zeng, C. Zheng, T. Li, M. Wei and A. Liu, *Biosens. Bioelectron.*, 2014, **60**, 137.

189 T. Zhang, L. Zeng, L. Han, T. Li, C. Zheng, M. Wei and A. Liu, *Anal. Chim. Acta*, 2014, **822**, 23.

190 Y. Li, M. Xu, P. Li, J. Dong and S. Ai, *Anal. Methods*, 2014, **6**, 2157.

191 J. Dong, X. Wang, F. Qiao, P. Liu and S. Ai, *Sens. Actuators, B*, 2013, **186**, 774.

192 J. Gong, X. Miao, H. Wan and D. Song, *Sens. Actuators, B*, 2012, **162**, 341.

193 S. Wu, X. Lan, L. Cui, L. Zhang, S. Tao, H. Wang, M. Han, Z. Liu and C. Meng, *Anal. Chim. Acta*, 2011, **699**, 170.

194 S. Ebrahim, R. El-Raey, A. Hefnawy, H. Ibrahim, M. Soliman and T. M. Abdel-Fattah, *Synth. Met.*, 2014, **190**, 13.

195 M. Wang, J. Huang, M. Wang, D. Zhang and J. Chen, *Food Chem.*, 2014, **151**, 191.

196 T. Jeyapragasam and R. Saraswathi, *Sens. Actuators, B*, 2014, **191**, 681.

197 P. A. D'Agostino and C. L. Chenier, Technical Report March 2006, DRDC Suffield TR 2006-022.

198 S. Royo, R. Martinez-Manez, F. Sancenon, A. M. Costero, M. Parra and S. Gil, *Chem. Commun.*, 2007, 4839.

199 R. M. Black, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2010, **878**, 1207.

200 I. Bhowmick and Neelam, *Analyst*, 2014, **139**, 4154.

201 F. Arduini, D. Neagu, S. Dall'Oglio, D. Moscone and G. Palleschi, *Electroanalysis*, 2012, **24**, 581.

202 P. A. Baker, M. N. Goltz, A. M. Schrand, D. Y. Yoon and D. S. Kim, *Biosens. Bioelectron.*, 2014, **61**, 119.

203 J. S. Lee, D. H. Shin, J. Jun and J. Jang, *ACS Nano*, 2013, **7**, 10139.

204 F. Arduini, A. Amine, D. Moscone, F. Ricci and G. Palleschi, *Anal. Bioanal. Chem.*, 2007, **388**, 1049.

205 G. Liu and Y. Lin, *Anal. Chem.*, 2006, **78**, 835.

206 S. P. Sharma, L. N. S. Tomar, J. Acharya, A. Chaturvedi, M. V. S. Suryanarayan and R. Jain, *Sens. Actuators, B*, 2012, **166**, 616.

207 S. Wu, F. Huang, X. Lan, X. Wang, J. Wang and C. Meng, *Sens. Actuators, B*, 2013, **177**, 724.

208 Y. Yang, H. Tu, A. Zhang, D. Du and Y. J. Lin, *J. Mater. Chem.*, 2012, **22**, 4977.

209 P. Noyrod, O. Chailapakul, W. Wonsawat and S. Chuanuwatanakul, *J. Electroanal. Chem.*, 2014, **719**, 54.