

Cite this: *Chem. Sci.*, 2021, 12, 6059

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 3rd February 2021
Accepted 5th March 2021

DOI: 10.1039/d1sc00660f

rsc.li/chemical-science

Chemoproteomic profiling of itacononations in *Salmonella*†

Yanling Zhang,^{ac} Wei Qin,^{ac} Dongyang Liu,^{ab} Yuan Liu^{ab} and Chu Wang^{id}*^{abc}

Itaconate is an immunoregulatory and anti-bacterial metabolite, and plays important roles in host–pathogen interactions. Chemoproteomic strategies have been used to explore the anti-inflammatory effects of itaconate on activated macrophages and it has been found that many key proteins in immune pathways were modified; however, how itaconate modulates pathogens was not fully understood. Here, we have designed and synthesized a series of itaconate-based bioorthogonal probes, which enable quantitative and site-specific profiling of itaconated proteins and sites in *Salmonella*. Among many proteins related to energy metabolism, we identified a key enzyme involved in the glyoxylate cycle, isocitrate lyase (ICL), as the most prominent target. Covalent modification of the active-site cysteine in ICL by itaconate abolishes the enzyme activity and suppresses bacterial growth. Our chemoproteomic study has uncovered the wide array of itaconation targets in *Salmonella* and provided a comprehensive resource for understanding the anti-bacterial function of this intriguing metabolite.

Introduction

Itaconate has been known as an anti-inflammatory and anti-bacterial metabolite involved in macrophage activation.^{1,2} It is generated from *cis*-aconitate by mitochondria-associated enzyme immune responsive gene 1 (IRG1) and its concentration is strikingly upregulated in activated macrophages.^{3,4} Since the first report of itaconate production in mammalian cells, most of the studies have been devoted to exploring its immunosuppressive function in macrophages.^{5,6} Itaconate was initially found to reduce cytokine secretion and regulate macrophage metabolism by inhibiting succinate dehydrogenase (SDH).¹ Owing to its weak electrophilic property, itaconate can also react with nucleophilic residues, thereby covalently modifying target proteins and affecting their functions. It has been reported that itaconate can exert anti-inflammatory function through the NRF2 and ATF3 signaling pathways by modifying the cysteine sidechains of KEAP1 and GSH, respectively.^{7,8} It can also modify NLRP3 and block the inflammasome activations.⁹ Our group has developed a couple of chemoproteomic strategies to globally identify the itaconate modifications (“itaconation”) in activated macrophages and revealed that

itaconate can covalently modify protein targets to modulate the glycolysis and necroptosis pathways.^{10,11} However, the global itaconation landscape in bacteria has not been thoroughly explored.

The knowledge of itaconate's anti-bacterial effect could be traced back to as early as 1971 when researchers found that itaconate could inhibit the growth of *Salmonella enterica* serovar *Typhimurium* (*S. enterica*) and *Mycobacterium tuberculosis* on a non-glucose-based carbon source.^{12,13} Later on, it was found that knocking out IRG1 in macrophages would lead to the rapid growth of pathogens during invasion. It is generally believed that itaconate can mimic substrate analogues and competitively inhibit the metabolic enzymes in pathogens to interfere with their energy metabolism, including methylisocitrate lyase (MICL) and isocitrate lyase (ICL) in the glyoxylate cycle, and propionyl-CoA carboxylase in the citramalate cycle.¹⁴ More recently, a detailed mechanistic study showed that the CoA derivative of itaconate acts as a suicide inhibitor of B12-dependent methylmalonyl-CoA mutase (MCM) in *Mycobacterium tuberculosis*.¹⁵ However, whether itaconate could regulate pathogen functions *via* direct itaconation is largely unknown.

In this study, we employed quantitative chemoproteomics to survey proteins and sites that are covalently modified by itaconate in *S. enterica*. With a unique chemical probe that showed specific labeling in bacterial lysates, we were able to identify hundreds of proteins targeted by itaconate. We discovered that itaconate can covalently modify multiple cysteines in ICL including the active-site Cys195. Itaconation results in not only the inhibition of ICL's enzymatic activity but also the destabilization of the enzyme. We believe that the chemoproteomic profiling data would provide a comprehensive resource for

^aSynthetic and Functional Biomolecules Center, Beijing National Laboratory for Molecular Sciences, Key Laboratory of Bioorganic Chemistry and Molecular Engineering of Ministry of Education, China

^bCollege of Chemistry and Molecular Engineering, China

^cPeking-Tsinghua Center for Life Sciences, Peking University, Beijing 100871, China. E-mail: chuawang@pku.edu.cn

† Electronic supplementary information (ESI) available: Experimental details, synthetic procedures, characterization data for reaction products and additional figures. See DOI: 10.1039/d1sc00660f



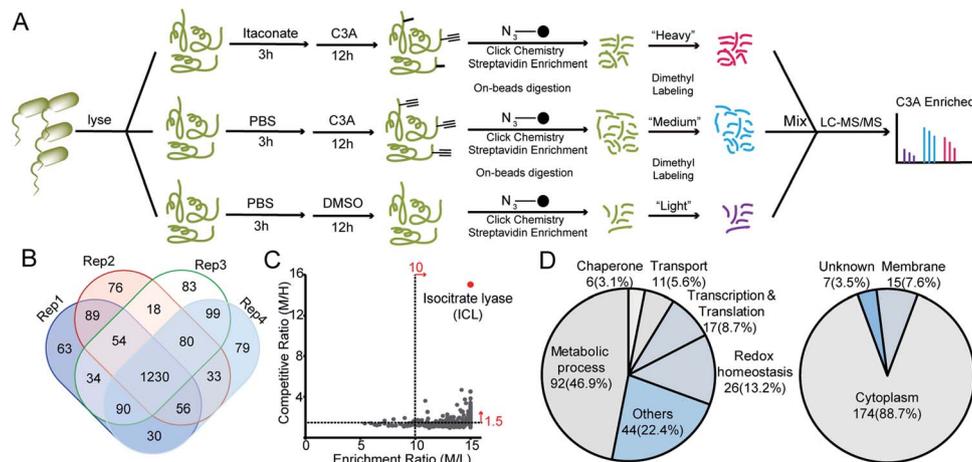


Fig. 2 Quantitative chemoproteomic profiling of itaconated proteins by C3A in *S. enterica* lysates. (A) Workflow for the triplex dimethylation-based quantitative chemoproteomic profiling of itaconated proteins in *S. enterica* by C3A. (B) Overlap of the itaconated proteins identified by C3A in four biological replicates. (C) Distribution of the averaged enrichment and competition ratios of the C3A-labeled proteins. (D) Bioinformatics analysis of the prominent 197 itaconated protein targets in terms of functional annotation and cellular location.

more than 2 sites (Fig. 3C). No other amino acid sites were detected beyond cysteines, suggesting the specific reactivity of C3A in the *S. enterica* proteome. We biochemically confirmed the itaconation sites on some of the targets including enolase (ENO), oligoribonuclease (ORN), ATP synthase gamma chain (ATPG), probable transcriptional regulatory protein (YEBC), putative oxidoreductase (YIEF), and putative intracellular proteinase (YHBO). C3A's labeling on these proteins was completely abolished by mutation of the corresponding cysteines to serines (Fig. 3D).

Among all the itaconated proteins identified by our chemoproteomic profiling, the most prominent one is isocitrate lyase (ICL, also known as ACEA) which has maximum ratios in both the enrichment and competition profiling experiments (Fig. 2B). ICL is a Mg^{2+} -dependent enzyme involved in the glyoxylate cycle that catalyzes the reversible conversion of isocitrate to succinate and glyoxylate²⁰ (Fig. S6A[†]). This catalysis bypasses two oxidative steps in the TCA cycle to synthesize succinate. Thus, the glyoxylate cycle is more carbon conserving and can help maintain an adequate supply of TCA intermediates under glucose limited conditions.²¹ Given its absence in vertebrates and conservation among most prokaryotes, ICL is considered as an ideal drug target candidate.²² Notably, a recent study independently discovered *via* the approaches of structural biology and medicinal chemistry that itaconate could serve as a covalent inhibitor for the two ICL isoforms from *Mycobacterium tuberculosis* by targeting their active-site cysteines.²³ Consistent with this finding, our chemoproteomic profiling identified 5 itaconation sites in the ICL of *S. enterica* including the catalytic Cys195 (Table S3[†]).

We recombinantly overexpressed the wild-type ICL and each of the 5 cysteine-to-serine single mutants with the N-terminal 6xHis tag in *S. enterica*. The C195S mutant almost lost all the labeling signal, indicating that it is the main itaconation site in ICL (Fig. 4A). We next purified each of the ICL variants (Fig. S6B[†]) and incubated the purified protein with itaconate to

confirm the modifications on five cysteine sites by LC-MS/MS (Fig. 4B and S6C[†]). We then measured the enzymatic activity of the wild-type and mutant ICLs following a previously reported assay.²⁴ Consistent with the knowledge that C195 serves as a catalytic residue to aid the product formation of succinate, the activity of C195S was completely abolished, while that of C318S was partially lost (Fig. 4C). We also observed a dose-dependent inhibition by itaconate of the activity of wild-type

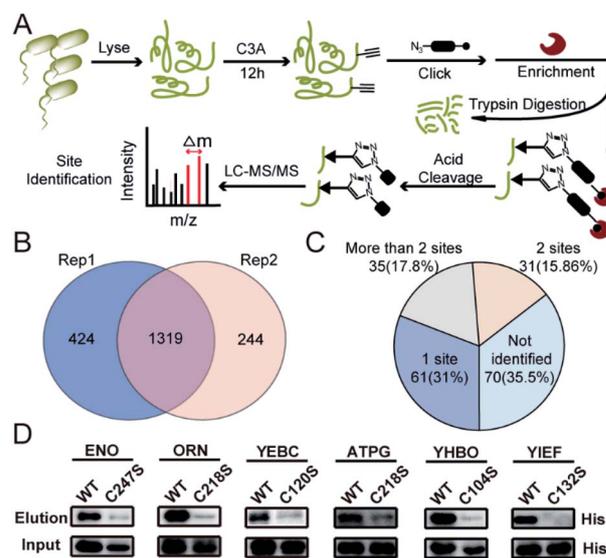


Fig. 3 Site-specific profiling of itaconated cysteines by C3A in *S. enterica* lysates. (A) The TOP-ABPP workflow for chemoproteomic profiling of itaconated sites in *S. enterica* lysates. (B) Overlap of the itaconated cysteines identified by C3A in two biological replicates. (C) Distribution of the itaconation targets in terms of the number of modification sites per protein. (D) Biochemical validation of the novel itaconation sites in ENO, ORN, YEBC, ATPG, YHBO, and YIEF by comparing the C3A labeling between the wild-type (WT) proteins and their cysteine-to-serine mutants.



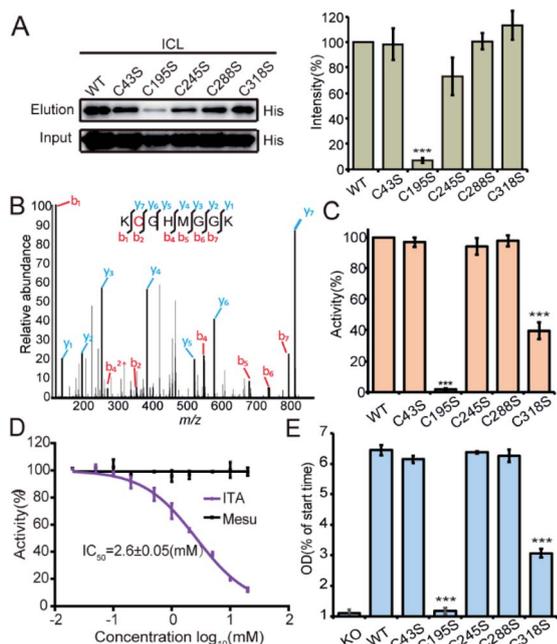


Fig. 4 Functional characterization of the itaconations on ICL. (A) Mutation of Cys195 abolished the C3A labeling on ICL (left). The quantification (right) of the labeling intensities was averaged from three repeats. (B) MS/MS spectrum of the peptide of ICL containing the itaconated Cys195. The purified ICL protein was incubated with 1 mM itaconate for 2 hours, and then the labeled proteins were digested by trypsin and analyzed by LC-MS/MS. (C) Enzyme activity of the purified wild-type (WT) ICL and the five cysteine-to-serine mutants. (D) Itaconate inhibits the enzyme activity of ICL with an IC_{50} of 2.6 mM. (E) Strains with overexpression of certain ICL mutants (C195S or C318S) showed growth defects.

ICL with an IC_{50} value around 2.6 mM (Fig. 4D). Lastly, we knocked out the endogenous ICL from *S. enterica* and complemented back with either the wild-type protein or each of the five mutants (Fig. 4E). The growth defects of these strains under the limited carbon source were highly consistent with the observed loss of the enzymatic activity, with the C195S and C318S strains showing complete and partial growth inhibition, respectively. These data collectively suggested that Cys195 is a functionally critical site for ICL and covalent modification of this specific site by itaconate mediated the metabolite's anti-bacterial activity.

Conclusions

In this study, we developed a novel bioorthogonal probe, C3A, for quantitative and site-specific chemoproteomic profiling of itaconate modifications in *S. enterica*. Compared to the ITalk probe used for profiling in living macrophages, C3A has an amide linkage and a shorter tail, which resulted in dramatically different labeling performance in bacterial lysates. While the exact reason for the enhanced and unique labeling in the *S. enterica* proteome by C3A remains elusive, the probe has enabled the first global profiling of itaconation events in bacteria, which will provide a rich resource for investigating the anti-bacterial role of the metabolite. In future,

the probe's permeability should be further improved for profiling in living pathogens. On a related note, a recent study showed that itaconate and its derivatives have different impacts on downstream pathways,²⁵ warranting the need to develop new chemoproteomic tools to map endogenous itaconation in both hosts and pathogens.

Itaconate has long been considered to inhibit ICL by competitive binding as a product analogue. An elegant study has recently provided unambiguous structural and biochemical evidence that itaconate indeed covalently modifies the active-site cysteine of ICL in *Mycobacterium tuberculosis* to inhibit the enzyme's activity,²³ which justifies the rationale of developing covalent inhibitors for this functionally important enzyme. Our profiling study independently discovered and confirmed such a covalent modification event. Since C3A showed specific labeling on the active-site Cys195 in the purified enzyme, we envision that the probe should be highly compatible with high-throughput screening platforms, such as Fluopol-ABPP,²⁶ to obtain more potent covalent inhibitors for the enzyme.

It is interesting to observe that the C318S mutant of ICL also loses partially the enzymatic activity although the site is distant to the catalytic center. Our preliminary data revealed that the mutation might affect the enzyme's stability (Fig. S7A†), but a structural biology approach would ultimately provide an answer for this mystery. Since Cys318 is also subjected to itaconate's modification as evidenced by the tandem mass spectra, it could in principle serve as another site for allosteric inhibitor development. Lastly, with probes available for monitoring itaconation in both macrophages and bacteria, the diversified roles of itaconate could be further explored in the context of host-pathogen interface in future.

Author contributions

Chu Wang, Yanling Zhang and Wei Qin conceived the project. Yanling Zhang conducted most of the experiments unless specified otherwise. Dongyang Liu carried out the C8A synthesis. Yuan Liu helped perform MS data analysis. Yanling Zhang and Chu Wang analyzed the data and wrote the manuscript with input from all the authors.

Conflicts of interest

The authors declare no competing interests.

Acknowledgements

We thank the Computing Platform of the Center for Life Science for supporting the LC-MS/MS data analysis. This work is supported by grants from Ministry of Science and Technology of People's Republic of China (No. 2016YFA0501500) and the National Natural Science Foundation of China (No. 91953109, 21778004 and 21925701) to C. W.

Notes and references

- V. Lampropoulou, A. Sergushichev, M. Bambouskova, S. Nair, E. E. Vincent, E. Loginicheva, L. Cervantes-



- Barragan, X. Ma, S. C. Huang, T. Griss, C. J. Weinheimer, S. Khader, G. J. Randolph, E. J. Pearce, R. G. Jones, A. Diwan, M. S. Diamond and M. N. Artyomov, *Cell Metab.*, 2016, **24**, 158–166.
- 2 L. A. J. O'Neill and M. N. Artyomov, *Nat. Rev. Immunol.*, 2019, **19**, 273–281.
- 3 C. L. Strelko, W. Lu, F. J. Dufort, T. N. Seyfried, T. C. Chiles, J. D. Rabinowitz and M. F. Roberts, *J. Am. Chem. Soc.*, 2011, **133**, 16386–16389.
- 4 A. Michelucci, T. Cordes, J. Ghelfi, A. Pailot, N. Reiling, O. Goldmann, T. Binz, A. Wegner, A. Tallam, A. Rausell, M. Buttini, C. L. Linster, E. Medina, R. Balling and K. Hiller, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 7820–7825.
- 5 Z. Zasłona and L. A. J. O'Neill, *Mol. Cell*, 2020, **78**(5), 814–823.
- 6 T. Cordes, A. Michelucci and K. Hiller, *Annu. Rev. Nutr.*, 2015, **35**, 451–473.
- 7 E. L. Mills, D. G. Ryan, H. A. Prag, D. Dikovskaya, D. Menon, Z. Zasłona, M. P. Jedrychowski, A. S. H. Costa, M. Higgins, E. Hams, J. Szpyt, M. C. Runtsch, M. S. King, J. F. McGouran, R. Fischer, B. M. Kessler, A. F. McGettrick, M. M. Hughes, R. G. Carroll, L. M. Booty, E. V. Khatko, P. J. Meakin, M. L. J. Ashford, L. K. Modis, G. Brunori, D. C. Sévin, P. G. Fallon, S. T. Caldwell, E. R. S. Kunji, E. T. Chouchani, C. Frezza, A. T. Dinkova-Kostova, R. C. Hartley, M. P. Murphy and L. A. O'Neill, *Nature*, 2018, **556**, 113–117.
- 8 M. Bambouskova, L. Gorvel, V. Lampropoulou, A. Sergushichev, E. Loginicheva, K. Johnson, D. Korenfeld, M. E. Mathyer, H. Kim, L. H. Huang, D. Duncan, H. Bregman, A. Keskin, A. Santeford, R. S. Apte, R. Sehgal, B. Johnson, G. K. Amarasinghe, M. P. Soares, T. Satoh, S. Akira, T. Hai, C. de Guzman Strong, K. Auclair, T. P. Roddy, S. A. Biller, M. Jovanovic, E. Klechevsky, K. M. Stewart, G. J. Randolph and M. N. Artyomov, *Nature*, 2018, **556**, 501–504.
- 9 A. Hooftman, S. Angiari, S. Hester, S. E. Corcoran, M. C. Runtsch, C. Ling, M. C. Ruzek, P. F. Ruzek, A. F. McGettrick, K. Banahan, M. M. Hughes, A. D. Irvine, R. Fischer and L. A. J. O'Neill, *Cell Metab.*, 2020, **32**, 468–478.
- 10 W. Qin, K. Qin, Y. Zhang, W. Jia, Y. Chen, B. Cheng, L. Peng, N. Chen, Y. Liu, W. Zhou, Y. L. Wang, X. Chen and C. Wang, *Nat. Chem. Biol.*, 2019, **15**, 983–991.
- 11 W. Qin, Y. Zhang, H. Tang, D. Liu, Y. Chen, Y. Liu and C. Wang, *J. Am. Chem. Soc.*, 2020, **142**, 10894–10898.
- 12 J. O. Williams, T. E. Roche and B. A. McFadden, *Biochemistry*, 1971, **10**, 1384–1390.
- 13 B. A. McFadden and S. Purohit, *J. Bacteriol.*, 1977, **131**, 136–144.
- 14 I. A. Berg, L. V. Filatova and R. N. Ivanovsky, *FEMS Microbiol. Lett.*, 2002, **216**, 49–54.
- 15 M. Ruetz, G. C. Campanello, M. Purchal, H. Shen, L. McDevitt, H. Gouda, S. Wakabayashi, J. Zhu, E. J. Rubin, K. Warncke, V. K. Mootha, M. Koutmos and R. Banerjee, *Science*, 2019, **366**, 589–593.
- 16 R. A. Kulkarni, D. W. Bak, D. Wei, S. E. Bergholtz, C. A. Briney, J. H. Shrimp, A. Alpsy, A. L. Thorpe, A. E. Bavari, D. R. Crooks, M. Levy, L. Florens, M. P. Washburn, N. Frizzell, E. C. Dykhuizen, E. Weerapana, W. M. Linehan and J. L. Meier, *Nat. Chem. Biol.*, 2019, **15**, 391–400.
- 17 J. D. McKinney, K. H. zu Bentrup, E. J. Muñoz-Eliás, A. Miczak, B. Chen, W.-T. Chan, D. Swenson, J. C. Sacchettini, W. R. Jacobs and D. G. Russell, *Nature*, 2000, **406**, 735–738.
- 18 H. Mi, A. Muruganujan, X. Huang, D. Ebert, C. Mills, X. Guo and P. D. Thomas, *Nat. Protoc.*, 2019, **14**, 703–721.
- 19 E. Weerapana, A. E. Speers and B. F. Cravatt, *Nat. Protoc.*, 2007, **2**, 1414–1425.
- 20 S. Hillier and W. T. Charnetzky, *J. Bacteriol.*, 1981, **145**, 452–458.
- 21 F. C. Fang, S. J. Libby, M. E. Castor and A. M. Fung, *Infect. Immun.*, 2005, **73**, 2547–2549.
- 22 D. G. Russell, *Nat. Rev. Mol. Cell Biol.*, 2001, **2**, 569–577.
- 23 B. X. C. Kwai, A. J. Collins, M. J. Middleditch, J. Sperry, G. Bashiri and I. K. H. Leung, *RSC Med. Chem.*, 2021, **12**(1), 57–61.
- 24 E. M. el-Mansi, C. MacKintosh, K. Duncan, W. H. Holms and H. G. Nimmo, *Biochem. J.*, 1987, **242**, 661–665.
- 25 A. Swain, M. Bambouskova, H. Kim, P. S. Andhey, D. Duncan, K. Auclair, V. Chubukov, D. M. Simons, T. P. Roddy, K. M. Stewart and M. N. Artyomov, *Nat. Metab.*, 2020, **2**, 594–602.
- 26 D. A. Bachovchin, S. J. Brown, H. Rosen and B. F. Cravatt, *Nat. Biotechnol.*, 2009, **27**, 387–394.

