



Cite this: *Green Chem.*, 2021, **23**, 2507

Investigation of the catalytic activity and reaction kinetic modeling of two antimony catalysts in the synthesis of poly(ethylene furanoate)[†]

Lazaros Papadopoulos,^a Alexandra Zamboulis,^a Nejb Kasmi,^a Mohamed Wahbi,^a Christina Nannou,^b Dimitra A. Lambropoulou,^b Margaritis Kostoglou,^c George Z. Papageorgiou^d and Dimitrios N. Bikiaris^{*a}

In the last few decades, the interest in the synthesis and development of novel biobased polymers with interesting properties, able to compete with the existing petroleum-based polymers, has grown exponentially. 2,5-Furandicarboxylic acid (FDCA) is among the most studied biobased monomers and is currently the only aromatic one, when aromaticity is mandatory to obtain polymers with superior properties. However, furan-based polyesters are mainly prepared from 2,5-dimethyl furandicarboxylate (DMFD), by transesterification, adding a supplementary step in the synthesis and an extra barrier towards industrialization. Herein, we present the study of the polymerization of high-purity FDCA with EG using two different antimony catalysts (antimony oxide, Sb₂O₃, and antimony acetate, Sb(CH₃COO)₃) and different esterification and polycondensation temperatures by the traditional two-step polycondensation method. Each step was monitored by suitable characterization techniques, such as intrinsic viscosity measurements, carboxylic acid end-group analysis, nuclear magnetic resonance spectroscopy, infra-red spectroscopy and differential scanning calorimetry. Additionally, for the first time, liquid chromatography (LC) coupled with high resolution mass spectrometry (HRMS) was used in the study of FDCA polymerization. PEF oligomers produced during the esterification step were precisely identified by HRMS, providing new insights into the evolution of this step. Finally, theoretical studies were conducted to model the kinetics of the polymerization of PEF during both esterification and polycondensation stages.

Received 16th December 2020,
Accepted 2nd March 2021

DOI: 10.1039/d0gc04254d

rsc.li/greenchem

1. Introduction

The 21st century is characterized by a continuous strive of societies towards a greener, *i.e.* more sustainable, lifestyle. In polymer chemistry, this concern is implemented by the intensive research in the field of biobased polymers, *i.e.* polymers synthesized from monomers produced from renewable sources.^{1–5} Besides environmental concerns, this trend was further motivated by the progressive depletion of fossil fuels, the high oil price and its unpredictable fluctuations.⁶ In this

context, 2,5-furandicarboxylic acid (FDCA) has emerged as a versatile monomer, attracting the interest of both the academic and industrial communities.⁷ FDCA, which has been characterized as one of the twelve most important biobased monomers by the US Department of Energy,⁸ is considered an alternative to terephthalic acid due to their structural similarity.

Polymerization of FDCA with ethylene glycol (EG) affords poly(ethylene furanoate) (PEF).^{9,10} Compared to polyethylene terephthalate (PET), PEF exhibits superior thermal stability, a lower melting temperature, and significantly lower O₂ and CO₂ permeability, combined with excellent mechanical properties and good processability.^{11,12} PEF production is expected to reduce the non-renewable energy use by 40–50% and the greenhouse gas emissions by 45–55%.¹³ It has been estimated that, when produced at the same scale, FDCA and PEF production will be competitive with the production of terephthalic acid¹⁴ and PET,¹⁵ respectively. As a result of its biobased nature and superior physicochemical properties, there are real expectations that PEF will indeed replace PET in some applications. PEF is the spearhead of this new family of biobased

^aLaboratory of Chemistry and Technology of Polymers and Dyes, Department of Chemistry, Aristotle University of Thessaloniki, GR-541 24 Thessaloniki, Macedonia, Greece. E-mail: dbic@chem.auth.gr

^bLaboratory of Environmental Pollution Control, Department of Chemistry, Aristotle University of Thessaloniki, GR-541 24 Thessaloniki, Greece

^cLaboratory of Chemical and Environmental Technology, Aristotle University of Thessaloniki, GR-54124 Thessaloniki, Macedonia, Greece

^dDepartment of Chemistry, University of Ioannina, P.O. Box 1186, GR-45110 Ioannina, Greece

[†]Electronic supplementary information (ESI) available. See DOI: 10.1039/d0gc04254d



polyesters;^{16–21} however, a wide variety of promising polymers and copolymers have been obtained from FDCA.^{22–35} For example, poly(propylene furanoate) (PPF) and poly(butylene furanoate) (PBF) also exhibit a high gas barrier and mechanical properties.^{36–52}

The extended research on furan-based polyesters has resulted in a variety of starting monomers and polymerization techniques that can be employed for their synthesis.^{12,22,53–67}

Nevertheless, until now, the most popular strategy is utilizing dimethyl 2,5-furandicarboxylate (DMFD) in traditional melt polycondensations. Several catalytic systems have been screened for the polycondensation of DMFD by different research groups, and titanium catalysts exhibited the highest catalytic activity.^{68,69} This is why recent publications from academia are dominated by titanium catalysts. While this approach certainly offers some advantages (milder conditions and lower coloration when DMFD is the starting material),⁷⁰ it is not favored by the industry that would rather use FDCA as a starting monomer. Indeed, in this case, the esterification of FDCA to DMFD is bypassed. The atom efficiency for the polymerization of FDCA to PEF is higher compared to that for DMFD and water is produced instead of toxic methanol. Moreover, it demands a minimal adaptation of the existing PET production plants, where terephthalic acid, and not dimethyl terephthalate, is the starting monomer.⁷¹ Despite their efficiency, titanium-based catalysts result in higher coloration, compared to other metal-based catalysts, and faster degradation, while there are also some concerns regarding toxicity issues.^{68,69,72} Therefore, it is clear that more fundamental research needs to be conducted on the direct esterification of FDCA for the synthesis of PEF. Only a few kinetic studies have been conducted specifically on the esterification and polycondensation of FDCA.^{73–75} Among them, Brandão *et al.* studied the step-growth polymerization of PEF.⁷⁴ The apparent activation energies of the polymerization steps were calculated, and it was demonstrated that the esterification step is dependent on mass transfer, while the polycondensation is more affected by kinetic effects.

Herein, we present the kinetic study of the direct polymerization of high-purity FDCA with ethylene glycol (EG) with two different catalysts and at three different temperatures. We chose to study two antimony catalysts, antimony oxide, Sb₂O₃, and antimony acetate Sb(CH₃COO)₃, which are often neglected in PEF research in favor of titanate catalysts, despite their established suitability for food packaging applications. The polymerization of FDCA with ethylene glycol was monitored by traditional methods (intrinsic viscosity measurements, end-group analysis, nuclear magnetic resonance spectroscopy, infra-red spectroscopy, and differential scanning calorimetry). For the first time, to the best of our knowledge, further insight into the FDCA esterification at the molecular level was given by liquid chromatography coupled to high resolution mass spectrometry (LC-HRMS). The intermediate PEF oligomers that are formed during the esterification step of the polymerization were formally identified by HRMS providing a better understanding of the mechanism of polymerization. Finally, theoret-

ical studies were conducted to estimate the kinetic constants of the reactions involved in PEF synthesis by FDCA polymerization.

2. Materials and methods

2.1. Materials

2,5-Furandicarboxylic acid (FDCA, 99.2%, Sunshine Limited, China), ethylene glycol (anhydrous 99.8%), and antimony acetate (Sb(CH₃COO)₃) and antimony trioxide (Sb₂O₃) catalysts were purchased from Aldrich Co (Chemie GmbH, Steinheim, Germany). Phenol and 1,1,2,2-tetrachloroethane were purchased from Alfa Aesar (Kandel, Germany).

2.2. Synthesis

2.2.1. Esterification. 30.0 g (0.19 mol) of FDCA, 35.8 g of ethylene glycol (0.58 mol, 3 equiv.) and the appropriate catalyst (Sb(CH₃COO)₃ or Sb₂O₃ (400 ppm based on Sb metal)) were introduced into a polymerization reactor. The apparatus was evacuated and filled with nitrogen three times (in order to remove the existing oxygen). The reagents were heated at 160, 170 or 190 °C under nitrogen flow (50 mL min⁻¹) for 4 hours. At 30 minutes intervals, samples were taken from the polymerization reacting mixture for further analysis.

2.2.2. Polycondensation. 15.0 g of PEF oligomers prepared at 190 °C for 4 hours with each catalyst (Sb(CH₃COO)₃ or Sb₂O₃) were introduced into the polymerization reactor. The apparatus was evacuated and filled with nitrogen three times. The oligomers were heated at 220, 230 or 240 °C under high vacuum (5.0 Pa) for 3 hours. Samples were retrieved from the reaction mixture after 0.5, 1, 2 and 3 hours and characterized.

2.3. Characterization

2.3.1. Acid value (AV) measurements. The carboxyl end-group content was determined by titration using a methanolic solution of potassium hydroxide (1 M) and phenol red as an indicator. For each sample, the titration was repeated three times and the mean volume was further used.

The acid value (AV) is proportional to the unreacted acid groups and it is defined as the milligrams of potassium hydroxide required to neutralize one gram of sample. The AV was calculated according to the following equation (the concentration of the KOH solution is 1 M):

$$AV = \frac{56.11 \times V}{m} \quad (1)$$

where V is the volume of KOH solution used (in mL) and m is the mass of the sample (in g).

The conversion was calculated according to the following equation:

$$\text{Conversion} = \frac{AV_0 \times AV_t}{AV_0} \quad (2)$$

where AV_0 is the initial AV and AV_t is the AV value at each sampling interval.



2.3.2. Intrinsic viscosity (IV). Intrinsic viscosity $[\eta]$ was measured with an Ubbelohde viscometer (Schott Gerate GMBH, Hofheim, Germany) at 25 °C in a mixture of phenol and 1,1,2,2-tetrachloroethane (60/40, w/w). When the samples could not be dissolved at room temperature, they were slightly heated at 60 °C until complete dissolution. The intrinsic viscosity of polyester was calculated using the Solomon–Ciuta equation (eqn (3)) of a single point measurement:

$$[\eta] = \frac{\left[2 \left\{ \frac{t}{t_0} - \ln \left(\frac{t}{t_0} \right) - 1 \right\} \right]^{\frac{1}{2}}}{c} \quad (3)$$

where c is the concentration of the solution; t is the flow time of solution and t_0 is the flow time of pure solvent. For each sample, three measurements were conducted, and the average value was calculated. The number average molecular weight (M_n) was calculated from the IV values applying the modified Berkowitz equation:⁶⁹

$$M_n = 3.29 \times 10^4 [\eta]^{1.54}. \quad (4)$$

2.3.3. Fourier transform-infrared spectroscopy (FTIR). FTIR spectra were obtained using a Cary670 Agilent FTIR spectrometer, ATR unit: GladiATR Pike (diamond crystal) or a PerkinElmer FTIR spectrometer (PerkinElmer, Waltham, MA, USA), model Spectrum One, in absorbance mode and in the spectral region of 500–4000 cm^{-1} using a resolution of 4 cm^{-1} and 64 co-added scans.

2.3.4. Nuclear magnetic resonance (NMR). NMR spectra were recorded in deuterated dimethylsulfoxide (DMSO-d_6) on an Agilent 500 spectrometer (Agilent Technologies, Santa Clara, CA, USA), at room temperature. Spectra were calibrated using the residual solvent peaks.

2.3.5. Differential scanning calorimetry (DSC). Thermal analysis studies were carried out using a PerkinElmer Diamond DSC (PerkinElmer Corporation, Waltham MA, USA) updated to the DSC 8500 level, combined with an Intracooler IIP cooling system. For DSC analysis, the samples were briefly triturated with methanol to remove the excess EG. Samples of about 5 mg were used. In general, heating scans of the samples were conducted at 20 °C min^{-1} . T_g was recorded in the second heating run.

2.3.6. Liquid chromatography-high resolution mass spectrometry (LC-HRMS). HRMS was employed in order to achieve comprehensive structural characterization of the esterification reaction mixture. All analyses were performed on a Q Exactive Focus Orbitrap mass spectrometer, coupled with ultra-high-performance liquid chromatography (UHPLC), both from Thermo Scientific. The samples were dissolved in acetonitrile, filtered through PTFE syringe driven filters (0.22 μm) and injected into the LC Orbitrap MS/MS system. The separation of the oligomers was achieved on a Hypersil GOLD™ aQ column with dimensions 50 × 2.1 mm and a pore size of 1.9 μm (Thermo Scientific, MA, USA) at 40 °C. The mobile phase consisted of solvent A (water with 0.1% formic acid) and solvent B (methanol with 0.1% formic acid) at a flow rate of 200 μL

min^{-1} . The gradient elution program was set as follows: from 90% A (initial conditions) which stayed stable for 1.5 min to 40% in 4 min and then to 30% until 8 min. Afterwards, the mobile phase A turned to 0% within 3 min and stayed for additional 2 min, before returning to the initial conditions after 2 min with the re-equilibration of the column set at 1 min. The injection volume was set to 5 μL . The parameters of the heated electron-spray ion source (HESI) were: capillary temperature, 320 °C; S-lens RF level, 50; spray voltage, 3.5 kV; and the flow rate for the sheath, auxiliary and sweep gas, 45, 10 and 2 (au), respectively. All HESI parameters were optimized in the source auto-default option of the instrument after setting the LC flow rate at 200 $\mu\text{L min}^{-1}$.

All samples were analyzed in both positive and negative ionization modes. Full scan mass spectrum (FS-MS) data acquisition was performed in order to obtain the exact mass of each oligomer, setting the resolution to 70 000 FWHM, over a mass range of 60–900 m/z , and the mass error (Δ) below 5 ppm. Simultaneously, a data-dependent MS2 experiment was performed with the resolution set at 17 500 FWHM and the isolation width for the parent ion of interest at 1.0 m/z to obtain the fragmentation patterns of the formed oligomers and to confirm the findings of FS-MS. Technically, the molecular formulas of the expected oligomers were imported in an inclusion list that automatically generates the theoretical accurate mass, corresponding to their pseudo-molecular (protonated or deprotonated) ions. To induce the fragmentation, the collision energy (CE) in the high dissociation cell (HCD) was stepped at 20, 35 and 40 eV. All data and analyses were acquired and performed, respectively, using Thermo Xcalibur™ software (version 4.1).

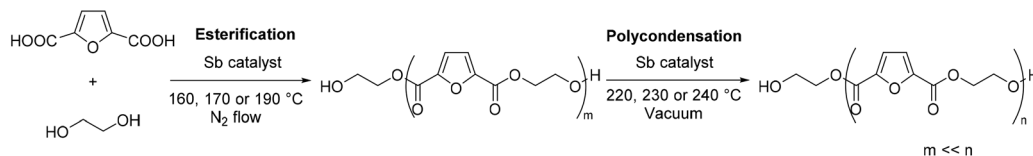
3. Results and discussion

In the two-step polycondensation polymerization of FDCA with ethylene glycol, in the first step, FDCA is esterified using EG while in the second step transesterification processes (polycondensation) take place, according to Scheme 1. Besides the catalyst used, the polymerization temperature is a crucial parameter that affects not only the extent of the reaction, but also the color of the final PEF polymer. It has been evidenced that FDCA decomposes at high temperature *via* decarboxylation processes, which is one of the causes of the coloration of PEF. Our group has previously studied the thermal stability of FDCA by thermogravimetric analysis (TGA) and pyrolysis-gas chromatography/mass spectrometry (Py-GC/MS).⁷⁶ It was found that the decomposition of FDCA starts at temperatures higher than 200 °C; however, mass losses were recorded at temperatures as low as 160 °C after 4 hours of isothermal heating. Therefore, we favored low temperatures (160, 170 and 190 °C) for the esterification step. Besides, it has recently been demonstrated that FDCA is satisfyingly soluble in EG above 140 °C.⁷⁷

3.1. Esterification

Esterification was performed in the presence of Sb_2O_3 or $\text{Sb}(\text{CH}_3\text{COO})_3$, at 160, 170 or 190 °C and samples were taken





Scheme 1 Polymerization of 2,5-furandicarboxylic acid with ethylene glycol. The first step, the esterification, affords oligomers. The second step, the polycondensation, yields poly(ethylene furanoate).

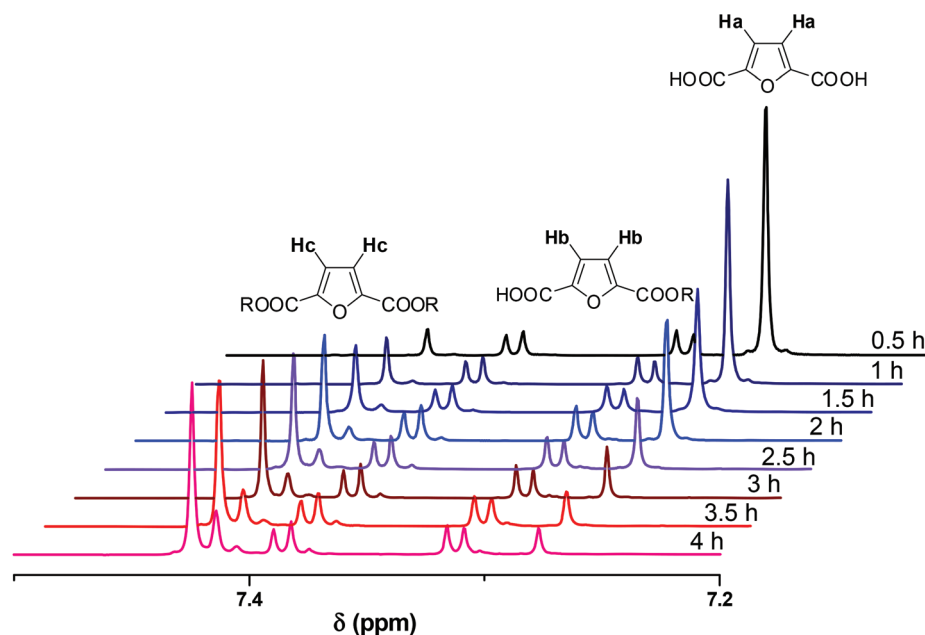


Fig. 1 ^1H NMR spectra of the crude reaction mixture of the esterification conducted at 160 °C, in the presence of Sb_2O_3 .

every 30 minutes to monitor the progress of the reaction. The consumption of the $-\text{COOH}$ groups of FDCA was evaluated by titration and ^1H NMR and the conversion was accordingly calculated. IR and DSC were also used to observe the evolution of the reacting mixture. Additionally, the monomers, dimers and trimers present in the reacting mixture were identified and quantified by LC-HRMS spectroscopy. Finally, based on these measurements, theoretical modelling was performed.

NMR spectra were recorded in DMSO-d_6 . A representative example of spectra of the crude reaction mixture is shown in Fig. 1. The aromatic region is appropriate to follow the progress of the reaction as the aromatic protons of the furanic unit are easily distinguishable. The aromatic protons of FDCA appear as a single peak at 7.27 ppm (protons *a*), while the corresponding protons of the di-esterified furanic ring appear at 7.42 ppm (protons *c*). The aromatic ring where only one COOH moiety has been esterified is detected by the two double peaks (one for each proton) at 7.31 and 7.39 ppm (protons *b*). As the reaction progressed, the peak at 7.27 ppm diminished. The peaks of the mono-ester initially increased, but, after a while, the mono-ester peaks tended to remain constant in favor of the peak of the di-ester which progressively dominated the spectrum. The conversion was calculated by

comparing the aromatic protons of the mono- and di-ester with the total amount of aromatic protons, according to eqn (5):

$$\text{conversion} = \frac{\frac{b}{2} + c}{a + b + c} \quad (5)$$

where *a* is the integral of the FDCA protons, *b* are the protons of the mono-ester and *c* are the protons of the di-ester.

The calculated conversions for each catalyst and each temperature are presented in Fig. 2 and in Tables S1 and S2.† The results obtained by titrimetry and NMR were in good agreement. At 160 °C Sb_2O_3 seemed to be slightly more active than $\text{Sb}(\text{CH}_3\text{COO})_3$; however the catalytic activity of the two catalysts is comparable at higher temperatures. We could say that, for the esterification step of the polymerization, the temperature, rather than the catalyst, has a higher impact on the conversion and that, within the studied timeframe (4 hours), a high temperature was necessary to achieve the complete conversion of FDCA. This is reasonable since this reaction is an autocatalyzed one (esterification reactions are catalyzed by acids) and EG is present in a large excess.^{78,79} The following characterization studies are indicatively illustrated by the polymerizations catalyzed by Sb_2O_3 .



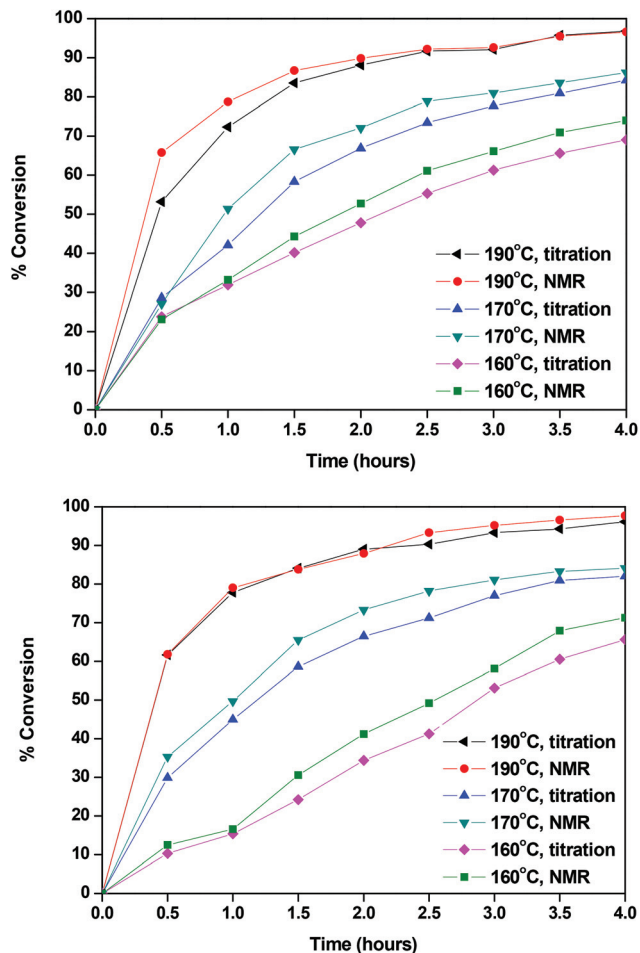


Fig. 2 Conversions calculated with Sb_2O_3 (up) and $\text{Sb}(\text{CH}_3\text{COO})_3$ (down).

The FTIR spectra of neat FDCA along with the spectra from various stages of the reaction are presented in Fig. 3. In the spectrum of FDCA, a broad absorption band, attributed to the OH stretching vibration of the carboxylic acid group of FDCA, is easily noticeable at $2500\text{--}3000\text{ cm}^{-1}$. As the reaction progresses, the intensity of the peak decreases, confirming the conversion of the end groups. Simultaneously, a new absorption band can be observed around 3360 cm^{-1} . This band corresponds to the $-\text{OH}$ end groups of the esterified intermediates. These results are in accordance with the titration and ^1H NMR trends shown above.

The peak that corresponds to the carbonyl groups is the second notable feature of these spectra. For FDCA, the peak that correlates with the $\text{C}=\text{O}$ stretching vibration of the carboxylic acid is observed at 1665 cm^{-1} . After half an hour, two overlapping peaks are clearly visible in the carbonyl region of the spectrum: at 1690 cm^{-1} (COOH) and at 1727 cm^{-1} (COOR), reflecting the conversion calculated by NMR and end-group analysis. As the reaction proceeds, the peak at 1690 cm^{-1} further decreases and the peak of the ester carbonyl groups at 1727 cm^{-1} dominates.

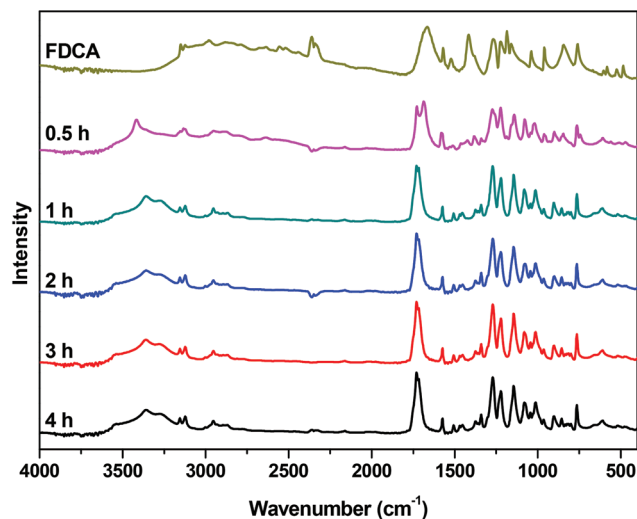


Fig. 3 FTIR spectra of the crude reaction mixture of the esterification catalyzed by Sb_2O_3 at $190\text{ }^\circ\text{C}$, at different reaction times.

The DSC thermographs are complementing the ^1H NMR data and acid value measurements. Indicatively, the DSC curves obtained at 170 and $190\text{ }^\circ\text{C}$ with the Sb_2O_3 catalyst are shown in Fig. 4. In all thermographs, at both reaction tempera-

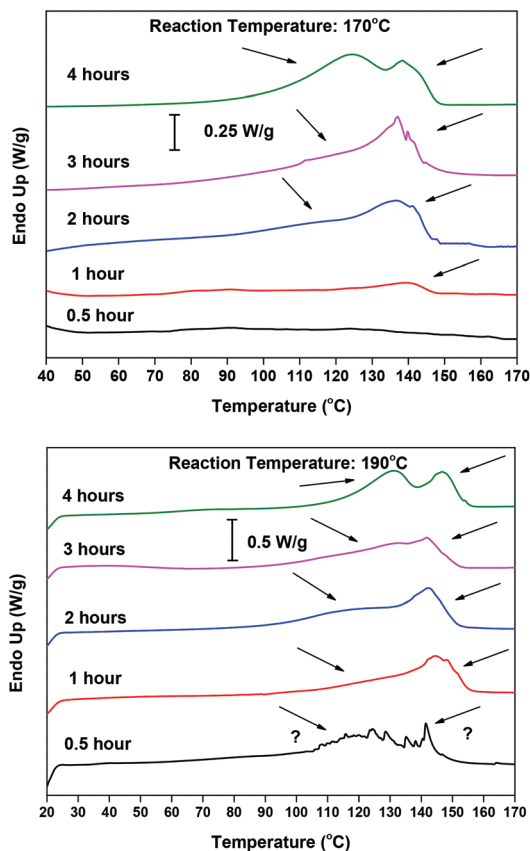


Fig. 4 DSC thermographs of the reaction samples of the esterification step, catalyzed by Sb_2O_3 at $170\text{ }^\circ\text{C}$ (up) and $190\text{ }^\circ\text{C}$ (down), at different reaction times.



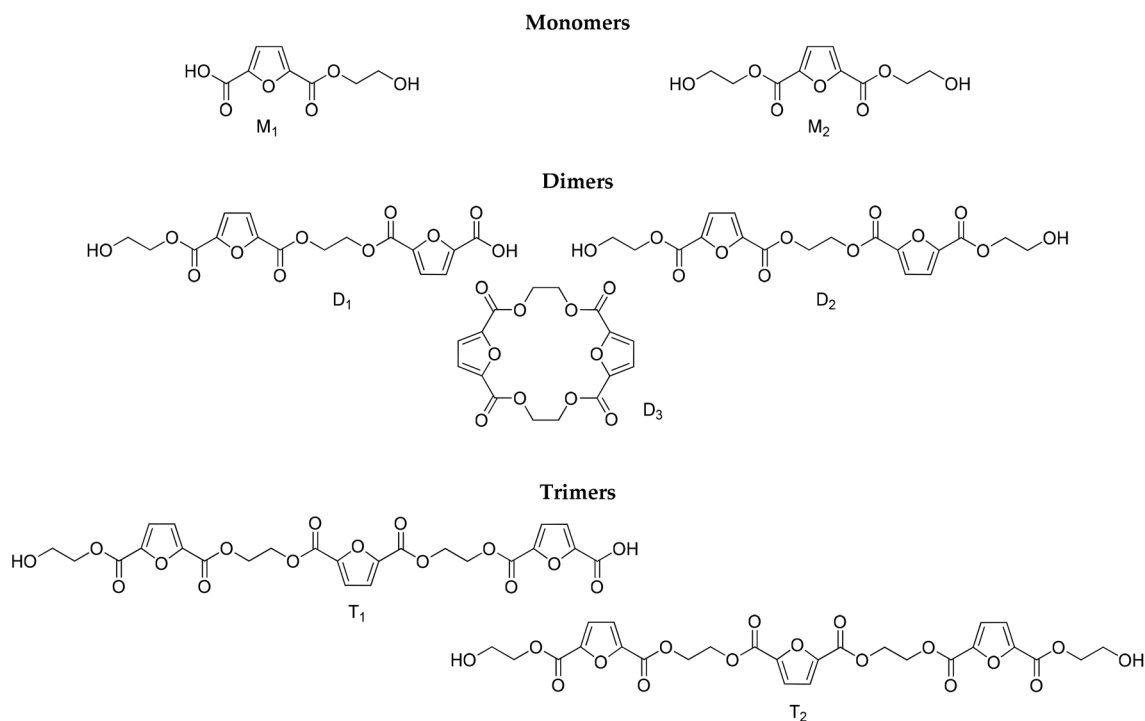
tures (170 and 190 °C), two main peaks can be seen. At 190 °C, in the early stages of esterification, a low intensity peak appears around 140 °C and, as the reaction progresses, its intensity decreases. We believe that this peak corresponds to 5-((2-hydroxyethoxy)carbonyl)furan-2-carboxylic acid, *i.e.* FDCA mono-ester with EG, monomer M1 in Scheme 2. Indeed, it is the first esterification product that is formed, and thus its presence is probable in the early stages of the reaction. Conversely, in the latest stages, as the conversion of the -COOH end groups tends to 100%, its concentration is dropping. The melting temperature depression compared with FDCA is rather big (m.p. 342 °C for FDCA); but comparing it with similar findings in the literature, mainly terephthalic acid (TA) and its mono-ester 4-((2-hydroxyethoxy)carbonyl)benzoic acid (m.p. 427 °C for TA, 180 °C for the ester)⁸⁰ and isophthalic acid (IA) and its mono-ester 3-((2-hydroxyethoxy)carbonyl)benzoic acid (m.p. 341 °C for IA, 132 °C for the ester),⁸¹ the T_m value seems reasonable. The melting peak at 120 °C could be attributed to the diester, bis(2-hydroxyethyl) furan-2,5-dicarboxylate (M2 in Scheme 2); however the reported m.p. in the literature is significantly lower (91 °C).⁸² Therefore, it probably corresponds to a dimer or trimer which is formed in the later stages of the esterification. Similar trends can be observed at 170 °C. The thermograms of the reaction at 160 °C (not shown) are dominated by a very broad peak around 140 °C, probably encompassing the peak at 120 °C. These results are in good agreement with the XRD patterns of the collected samples (please see Fig. S1 in the ESI†). At short reaction time, the recorded patterns indicate that almost amorphous materials are obtained, while their crystalline peaks are

increasing with time, as the esterification progresses. The patterns are almost identical for both catalysts.

To complement the results obtained by NMR and titrimetry, and to distinguish among the different structures that are progressively formed as the esterification proceeds, LC-HRMS analysis was performed. The structures of the oligomers that have been identified are depicted in Scheme 2. They have been divided into three groups (monomers, dimers and trimers) according to the number of furanic rings they bear. It is noteworthy that no traces of diethylene glycol by-products were detected.

Despite the unavailability of reference standards, LC-HRMS provided an array of key features to carry out a confident identification of the expected oligomers, namely, the accurate mass measurements (4 decimals, mass error <5 ppm), isotopic patterns, and characteristic fragments for each compound. For the comprehensive investigation of the obtained total ion chromatograms (TICs, please see ESI Fig. S2†) of the injected samples corresponding to various time intervals and different temperatures, in both positive and negative modes (ESI), mass filters were applied. Thus, the extrapolation of critical information was enabled with the aid of the generated extracted ion chromatograms (XICs). Table 1 summarizes the retention times, and the MS and MS2 data for the tentative identification of the oligomers.

Among the injected samples, those obtained at 190 °C exhibit more distinctive peaks, implying that higher amounts of the oligomers are formed compared to the lower temperatures. This is in good agreement with the findings of the rest of the instrumental analyses also indicating that the temperature plays a major role in the conversion of FDCA (Fig. 5).



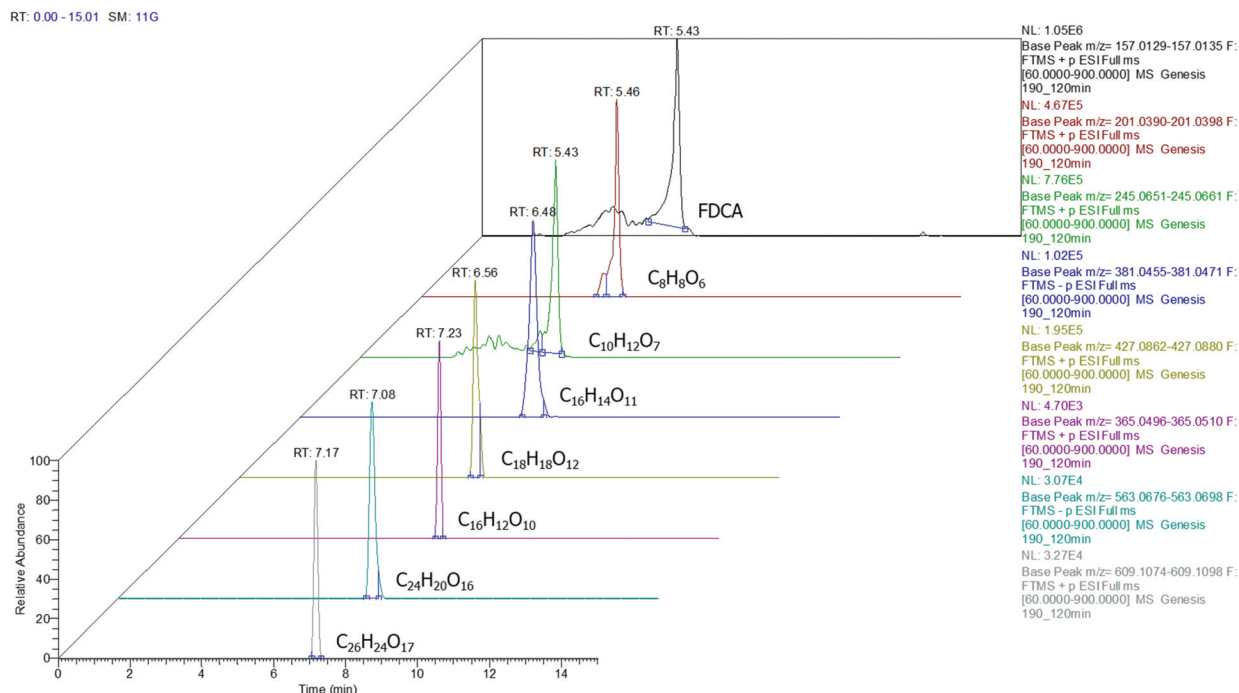
Scheme 2 Structures identified by LC-HRMS.



Table 1 Retention times, elemental composition, experimental and theoretical mass information, mass error deviation and double bond and ring equivalent number (RDB) of the detected oligomers

Formula	t_R (min)	Elemental composition	m/z (accurate mass)		RDBE	Δ (ppm)
			Theor.	Exp.		
<i>FDCA</i>						
C₆H₅O₅⁺	5.43	C₆H₅O₅⁺	157.0131	157.0132	4.5	0.3127
C ₅ H ₂ O ₂		C ₅ H ₃ O ₂ ⁺	95.0128	95.0132	4.5	4.4212
<i>Monomers</i>						
C₈H₈O₆⁺	5.46	C₈H₈O₆⁺	201.0394	201.0403	4.5	-2.800
C ₅ H ₂ O ₂		C ₅ H ₃ O ₂ ⁺	95.0128	95.0133	4.5	5.5454
C ₆ H ₂ O ₄	5.43	C ₆ H ₃ O ₄ ⁺	139.0026	139.0026	5.5	-0.1555
C₁₀H₁₂O₇⁺		C₁₀H₁₃O₇⁺	245.0656	245.0653	10	-1.1493
C ₆ H ₂ O ₄		C ₆ H ₃ O ₄ ⁺	139.0026	139.0025	5.5	-0.3750
C ₅ H ₂ O ₂		C ₅ H ₃ O ₂ ⁺	95.0128	95.0133	4.5	5.3045
<i>Dimers</i>						
C₁₆H₁₄O₁₁⁻	6.46	C₁₆H₁₃O₁₁⁻	381.0463	381.0470	10.5	3.265
C ₅ H ₂ O ₃		C ₅ H ₃ O ₃ ⁺	111.0077	111.0076	4.5	-0.2327
C ₉ H ₄ O ₃	6.56	C ₉ H ₅ O ₃	161.0233	161.0236	7.5	1.5791
C₁₈H₁₈O₁₂⁺		C₁₈H₁₉O₁₂⁺	427.0871	427.0867	9.5	-2.296
C ₆ H ₂ O ₄		C ₆ H ₃ O ₄ ⁺	139.0026	139.0025	5.5	-0.5946
C₁₆H₁₂O₁₀⁺	7.23	C₁₆H₁₃O₁₀⁺	365.0503	365.0503	10.5	-1.593
<i>Trimers</i>						
C₂₄H₂₀O₁₆⁻	7.08	C₂₄H₁₉O₁₆⁻	563.0679	563.0687	15.5	2.399
C₂₆H₂₄O₁₇⁺	7.17	C₂₆H₂₅O₁₇⁺	609.1086	609.1075	14.5	-2.863

Precursor ions in bold.

**Fig. 5** Extracted ion chromatograms for FDCA and oligomers detected at a sample corresponding to 190 °C (3D depiction).

According to the data presented in Fig. 5, FDCA is eluted at t_R 5.43 min, when its protonated ion [C₆H₅O₅]⁺ with a m/z of 157.0132 is detected, with a mass error of 0.3127 ppm. It is remarkable that for every positive finding of FDCA in the

recorded chromatograms from all injections, a subsequent fragmentation of the molecule has been induced, leading to the formation of the characteristic fragments [C₅H₃O₂]⁺ with m/z 95.0134, pronouncing the tentative identification of FDCA.



In addition, two monomers, with formulas of the protonated ions $[C_8H_9O_6]^+$ and $[C_{10}H_{13}O_7]^+$, were eluted at similar retention times (5.46 and 5.43 min, respectively) in positive ionization mode, exhibiting a mass error below 3 ppm and at least two fragments each, a fact that increases the confidence level of the measurements. Despite the similarity of the retention times between the two monomers, HRMS facilitates the distinction and unambiguous identification, thereby proving its strength for such applications, thanks to the accurate mass measurement at a high resolution as well as the presence of different fragments. As seen in Table 1, the fragment ions of both monomers are the same, due to the similar structure and nature of the precursor molecules yet formed in different abundance and accompanied by different mass errors. Regarding the dimers, two out of three detected molecules were acquired in positive ESI ($C_{18}H_{19}O_{12}^+$, $C_{16}H_{13}O_{10}^+$) and one in negative ESI ($C_{16}H_{13}O_{11}^-$), at t_R 6.56, 7.23 and 6.53, respectively. As the t_R increases, compounds with higher molecular weight are identified; hence two trimers were identified later

in the chromatograms; more specifically, one trimer with m/z $[C_{24}H_{19}O_{16}]^-$ and one with $[C_{26}H_{25}O_{17}]^+$ at t_R 7.08 and 7.17, respectively. Both trimers were found at a slightly lower abundance compared to the other oligomers and as a result, the intensity threshold was not enough to induce fragmentation. A typical example of the process followed from the chromatogram to the spectrum of the fragment ions is given in Fig. 6. The mass spectra for all the investigated compounds are included in the ESI.†

In order to construct the kinetic profiles of monomers, dimers and trimers along with FDCA, their relative abundance expressed as the peak area was employed. The concentration profiles with time, based on the LC-HRMS analysis, are presented in Fig. 7. These results are in complete agreement with the ones reported until now. The amount of FDCA steadily decreases as the esterification progresses; FDCA is consumed significantly faster at 190 °C than at lower temperatures. As FDCA decreases, other species appear. Monomers, which include the mono- and di-ester of FDCA, are the first com-

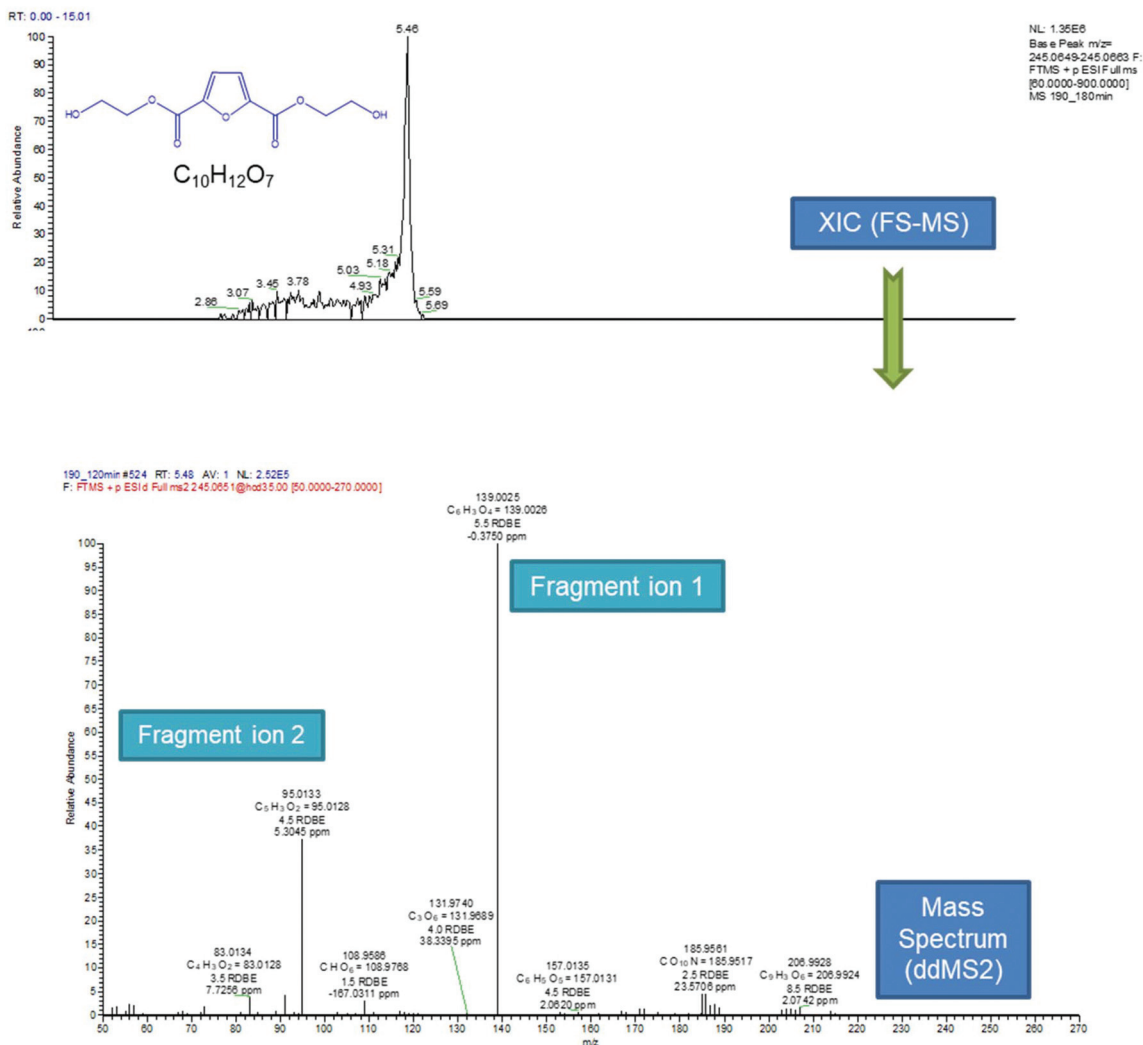


Fig. 6 Process followed from the extracted ion chromatogram to the spectrum of the fragment ions.



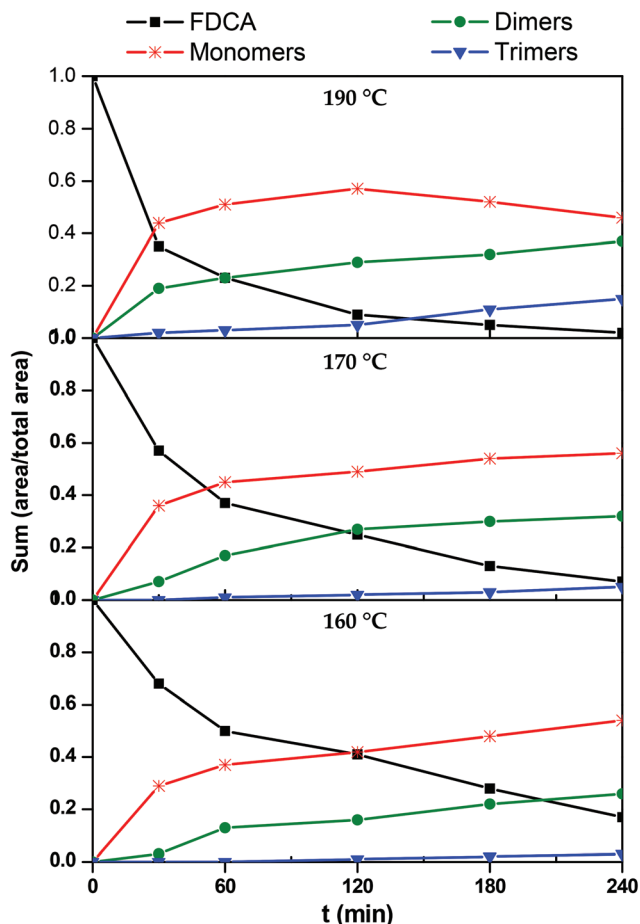


Fig. 7 Kinetic profiles of 2,5-furandicarboxylic acid (FDCA), monomers (M), dimers (D) and trimers (T) during the esterification reaction.

pounds to be formed. At 160 and 170 °C, the concentration of monomers increases rapidly initially and more slowly towards the end of the esterification step. The same trend is observed at 190 °C for the first two hours, while in the last couple of hours the monomer concentration decreases. This is expected

since monomers react with each other or with FDCA to yield longer oligomers. According to these results, monomers, and especially the bis(2-hydroxyethyl)furan-2,5-dicarboxylate, dominate the reacting mixture throughout the esterification step.

Progressively, as esterification takes place, dimers and trimers are formed. Their concentration increases steadily throughout the esterification step. The higher the temperature, the sooner the production of these oligomers is initiated. For example, at 190 °C trimers are detected in the reacting mixture from the first 30 minutes, while at 160 °C they are formed only after the first two hours. Similar remarks can also be made for dimers, though, not surprisingly, they form earlier and faster than trimers.

3.2. Transesterification – polycondensation

Once the esterification step of FDCA was studied, we investigated the second step of the polymerization where the molecular weight is being progressively built up. Since a higher conversion was achieved and longer oligomers were formed at 190 °C, this temperature was chosen to prepare two oligomer batches catalyzed by Sb_2O_3 and $\text{Sb}(\text{CH}_3\text{COO})_3$, respectively. These oligomers were further subjected to transesterification at 3 different temperatures: 220, 230 and 240 °C. Samples were retrieved from the polymerization reactor at 30, 60, 120 and 180 minutes. Intrinsic viscosity (IV) was measured and the samples were further characterized by DSC and IR.

The IV of the samples was measured in a phenol/1,1,2,2-tetrachloroethane mixture as indicated in the experimental part and the results are presented in Fig. 8. It is obvious that at all temperatures and regardless of the catalyst, IV increases with time. The higher the temperature, the higher the values of IV achieved. When comparing the two catalysts, we can see that the IV values obtained in the presence of Sb_2O_3 are much more important than the ones obtained when using $\text{Sb}(\text{CH}_3\text{COO})_3$. Therefore, according to IV measurements, Sb_2O_3 has a higher catalytic activity compared to $\text{Sb}(\text{CH}_3\text{COO})_3$. Overall, one could argue that the obtained IV values are not very high; however the aim of the present study was not to obtain high molecular weight PEF and additionally, the

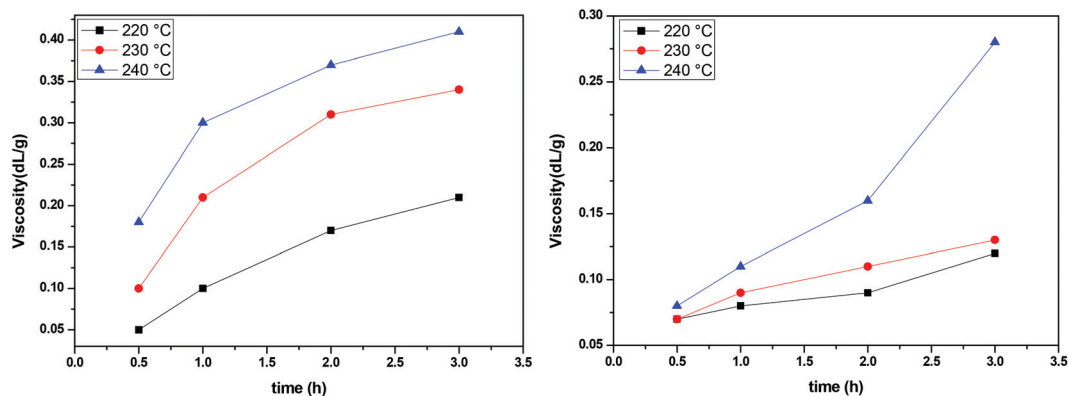


Fig. 8 Increase of intrinsic viscosity with time at different temperatures, catalyzed by Sb_2O_3 (left) and $\text{Sb}(\text{CH}_3\text{COO})_3$ (right).



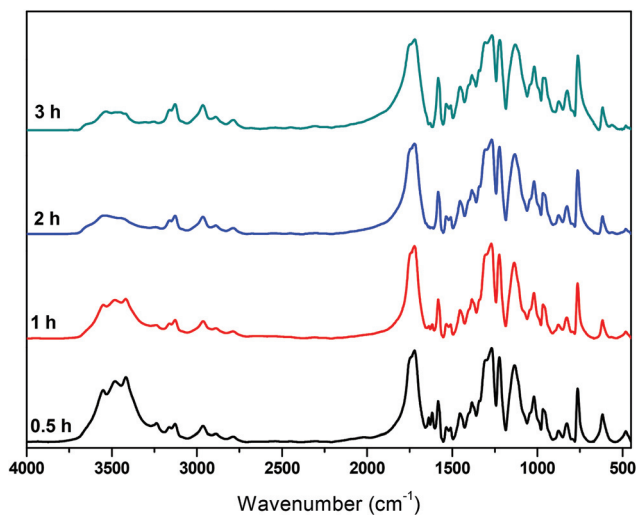


Fig. 9 FTIR spectra of the polycondensation catalyzed by Sb_2O_3 , at 190 °C, at different reaction times.

polymerization procedure was interrupted each time a sample was retrieved from the reacting mixture. Despite their brevity, those interruptions affected the overall procedure resulting in a lower molecular weight.

Besides IV measurements, all samples were characterized by IR. The trends observed in all spectra are similar and one of them is indicatively presented in Fig. 9. The band corresponding to the carbonyl of the furan ring, at *ca.* 1720 cm^{-1} , is not affected by the polycondensation reaction, while the band around 3500 cm^{-1} is progressively decreasing. This is expected since most of the oligomers formed at the end of the esterification step are esters with hydroxyl-ended chains. The ester carbonyl groups are not affected by the transesterification reaction, while the concentration of hydroxyl groups decreases progressively as oligomers react to form polymers and EG is released (since the polycondensation takes place under vacuum, EG is immediately removed from the reacting mixture).

Finally, these results were complemented with DSC measurements, Fig. 10. As the polycondensation progresses and the length and molecular weight of the polymeric chains increase, T_g and T_m progressively shift to higher temperatures. When the reaction is catalyzed by $\text{Sb}(\text{CH}_3\text{COO})_3$, at 220 °C and at short reaction time (0.5 h) T_g is around 54 °C. By increasing reaction time and temperature T_g reaches approximately 76 °C (3 h, 240 °C). Similarly, T_m increases from 185 °C to 195 °C. A corresponding trend is observed when the polycondensation is catalyzed by Sb_2O_3 , though higher temperatures were recorded. After 3 h at 240 °C, PEF with a T_g at 85 °C and a T_m at 203 °C is obtained. The higher T_g and T_m obtained with Sb_2O_3 indicate a higher molecular weight. These data are in accordance with the IV results, where PEF of higher molecular weight is obtained when the reaction is catalyzed by Sb_2O_3 compared to $\text{Sb}(\text{CH}_3\text{COO})_3$, suggesting the superior catalytic performance of Sb_2O_3 .

4. Kinetic modeling of the processes

There is a whole hierarchy of methods for modeling the pair of esterification and transesterification reactions. The simplest approach is to assume a global reaction with an n -th order kinetics of disappearance of $-\text{COOH}$.⁷⁶ However, the majority of the modeling efforts attempts to model separately the two reactions, so this is the approach that has been adopted in the present study.

4.1. Esterification reaction

There are three general approaches for modeling the esterification reaction in the literature. They are (in order of decreasing complexity and detail):⁸³ (i) the molecular species model which gives the exact concentration for all species produced during the reaction; (ii) the functional group model which gives information only on the global degree of oligomerization; and (iii) the overall effective reaction model which describes only the consumption of the reactants. The fact that the esterification products have been identified allows to write down the exact set of reactions occurring during esterification. Let us designate as F and A the two reactants (FDCA and EG respectively), while M, D, and T will stand for the products, monomers, dimers and trimers, respectively. Finally, the subscript under the letter denotes the specific type of the i -mers (see the description in Scheme 2). There is a system of 9 reactions that gives the evolution of all the identified components. A category (i) approach implies the direct solution of the 9 ordinary differential equations that describe the reaction set. This means that 9 kinetic constants must be estimated from a limited set of data (underspecified problem). The standard approach to overcome this difficulty in reaction engineering is the model reduction (called lumping) in order to ensure compatibility with the experimental data, allowing the extraction of kinetic parameters.

Another complication in modeling esterification regards the removal of water and A due to evaporation during the reaction. The removal model may be quite complicated, involving thermodynamic equilibrium and mass transfer considerations.⁷⁷ This may be very important in the case of removal of A. However, in the present experimental set-up only water is removed and since there is no direct measurement of it, the removal process is included in the lumping procedure. In other words, approach (iii) will be followed in this study. It was found, employing the conversion degree data, that the single reaction model used for different esterification reactions is not appropriate to describe the present data. A two-step lumped model is thus proposed here:



This model reflects the finding that $-\text{COOH}$ reacts at a different rate in a molecule with two $-\text{COOH}$ (F) compared to a molecule with one $-\text{COOH}$ (M_1). In the following discussion, all the concentrations are normalized by the initial concen-



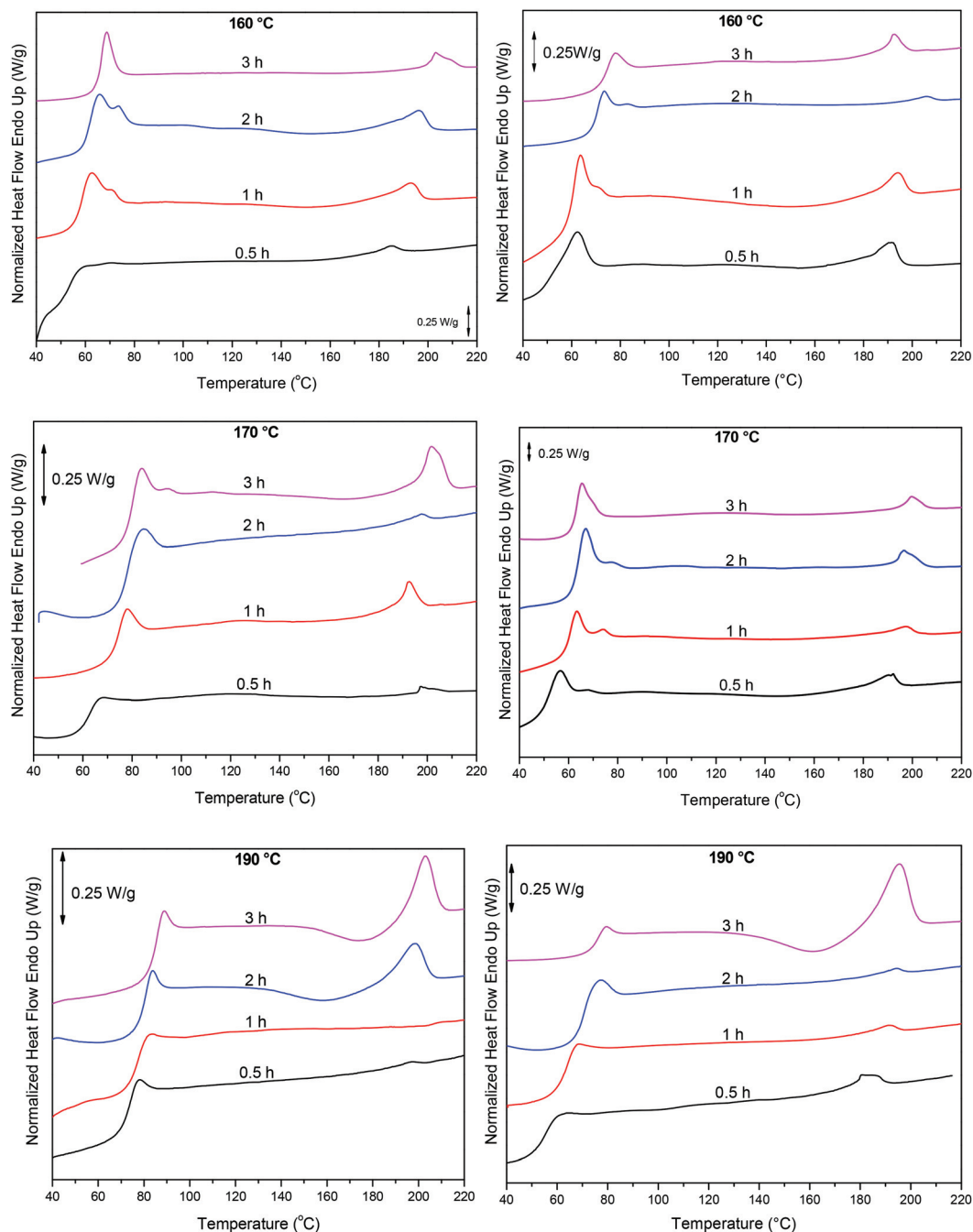


Fig. 10 DSC thermograms of the condensation step at different reaction times and temperatures, catalyzed by Sb_2O_3 (left) and $\text{Sb}(\text{CH}_3\text{COO})_3$ (right).

tration of F (denoted as $[\text{F}]_0$) in order to render the analysis independent from initial concentrations. The equations that describe the evolution of the concentrations are:

$$\frac{d[\text{F}]}{dt} = -K_1[\text{F}][\text{A}] \quad (7a)$$

$$\frac{d[\text{A}]}{dt} = -K_1[\text{F}][\text{A}] \quad (7b)$$

$$\frac{d[\text{M}_1]}{dt} = K_1[\text{F}][\text{A}] - K_2[\text{M}_1][\text{A}] \quad (7c)$$

$$\frac{d[\text{M}_2]}{dt} = K_2[\text{M}_1][\text{A}] \quad (7d)$$

The fractional conversion of COOH is computed as $1 - [\text{F}] - [\text{M}_1]/2$ since each M_1 molecule has half COOH groups than each F molecule. The pseudo-first order reaction constants K_1 and K_2 are related to the intrinsic second order reaction con-



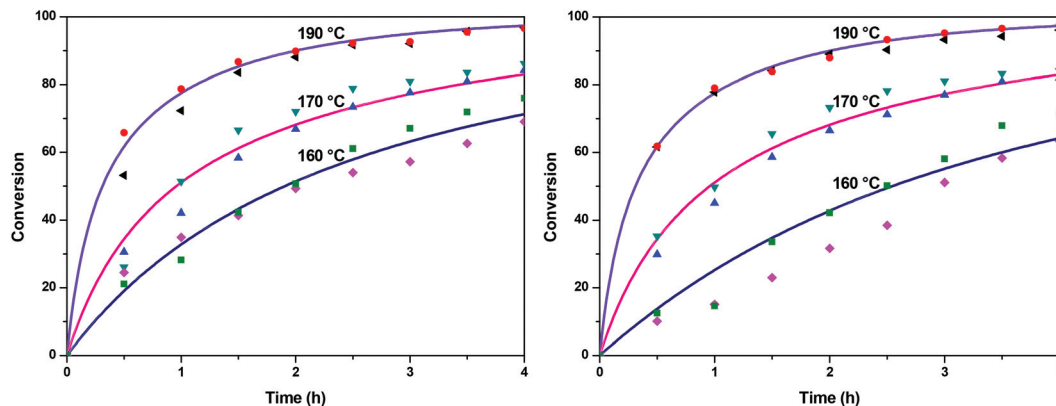


Fig. 11 Comparison between model (solid lines) and experimental (symbols) conversions of esterification reaction with (a) Sb_2O_3 and (b) $\text{Sb}(\text{CH}_3\text{COO})_3$.

stants $K_{1,\text{in}}$ and $K_{2,\text{in}}$ by $K_1 = K_{1,\text{in}}[\text{F}]_0$, $K_2 = K_{2,\text{in}}[\text{F}]_0$. The above system of ordinary differential equations has initial conditions (at $t = 0$) $[\text{F}] = 1$, $[\text{A}] = 3$, $[\text{M}_1] = [\text{M}_2] = 0$ and is solved numerically using the explicit Euler method. The coefficients K_1 and K_2 are chosen (fitted in a general sense since no exact objective function can be constructed) in order to give a qualitative agreement to the conversion data derived by the two experimental techniques. The comparison between model results and experimental data appears in Fig. 11. The selected values are $K_1 = 0.2, 0.7, 2.1 \text{ h}^{-1}$ and $K_2 = 0.15, 0.2$ and 0.6 h^{-1} for temperature T equal to 160, 170 and 190 °C, respectively. The same values were found for the second catalyst except K_1 at 160 °C which is 0.3 instead of 0.2. It is clear that the second reaction is a few times slower than the first one. Assuming a mean (for the two catalysts) value of K_1 at 160 °C equal to 0.25, the Arrhenius plots (appearing in Fig. 12) were constructed and the two constants K_1 and K_2 were found to exhibit an Arrhenius temperature dependence $K_i = K_{i_0} \exp(-E_i/R_g T)$ ($i = 1, 2$) with $E_1/R_g = 13\,844 \text{ K}$, $E_2/R_g = 9520 \text{ K}$ (R_g is the gas constant).

It appears that the first reaction is more sensitive to temperature than the second one. The resulting Arrhenius temperature dependence confirms that the selected reaction scheme is not an empirical one and has a physical basis (*i.e.* it accounts for the dominant steps of the complete reaction scheme). It is noted that the employed reaction scheme is the same as the one employed in ref. 77. However, the choice there was to assume that $K_2 = K_1$ and to use as additional fitting parameters the mass transfer coefficients of A and W from liquid to gas phase. The fact that the (resulting from fitting) mass transfer coefficient of W was found to be independent of experimental conditions while the mass transfer coefficient of A differs up to 30 times among the experiments raises questions about the validity of the approach in.⁷⁷

The above global reaction scheme cannot predict the evolution of the concentration of the identified oligomers. This could be done in principle by the detailed reaction system for oligomers. However, the experimental information is still not enough to allow simultaneous determination of the 9 kinetic coefficients. Instead a different lumped reaction scheme focused on oligomer evolution is employed, with a smaller number of kinetic constants to make their determination from the experimental data feasible. The following reaction scheme is considered (produced water does not appear in the following scheme because it does not participate in the kinetic equations and due to lumping its exact stoichiometric coefficient is unknown):



where M refers to $\text{M}_1 + \text{M}_2$, D refers to $\text{D}_1 + \text{D}_2 + \text{D}_3$ and T refers to $\text{T}_1 + \text{T}_2$. The evolution of concentrations can be found from the following system of ordinary differential equations.

$$\frac{d[\text{F}]}{dt} = -k_1[\text{F}][\text{A}] \quad (9a)$$

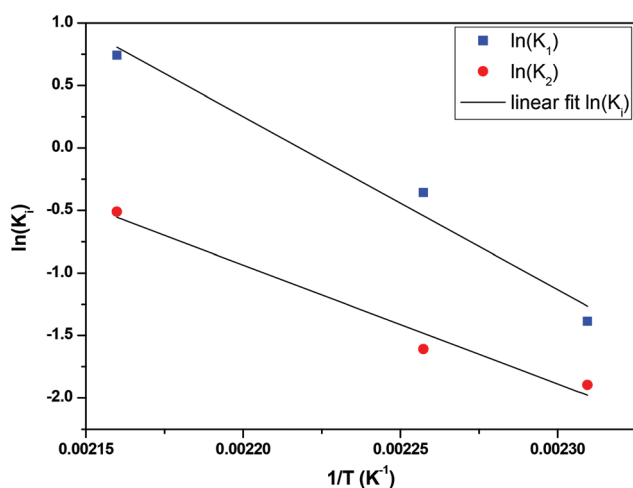


Fig. 12 Arrhenius plot for the constants K_1 and K_2 of esterification reaction.



$$\frac{d[A]}{dt} = -k_1[F][A] \quad (9b)$$

$$\frac{d[M]}{dt} = k_1[F][A] - k_2[M]^2 \quad (9c)$$

$$\frac{d[D]}{dt} = k_2[M]^2 - k_3[D][M] \quad (9d)$$

$$\frac{d[T]}{dt} = k_3[D][M] \quad (9e)$$

where the concentrations are once more normalized to $[F]_0$ and the coefficients k_i (inverse time units) are equal to the intrinsic second order reaction coefficients multiplied by $[F]_0$. The initial conditions for the above system (at $t = 0$) are $[F] = 1$, $[A] = 3$, $[M] = [D] = [T] = 0$. It is solved numerically using the explicit Euler method. The normalized concentrations can be used to find the molar fractions in the reaction mixture (dividing each of them with their sum).

In order to compare the experimentally determined weight fractions with the model results, they must be transformed to molar fractions. This is done by using the average molecular weight of the different types of monomers, dimers and trimers appearing in Scheme 2. This is only an approximation since the exact concentration of each species is not known but the error is not appreciable since the molecular weights do not vary a lot among the different forms of i-mers. The parameters k_1 , k_2 , and k_3 were extracted by fitting the model to the data. The comparison between experimental and model molar fractions is shown in Fig. 13 for the three temperatures. The comparison for the trimers is not shown due to their small fractions but it is stated that there is an agreement up to a molar fraction of 0.01. The deviation is somewhat larger only for the highest temperature and time and the model overpredicts the trimer concentration by not taking into account the possibility of (a small amount) of larger molecules. The values of the extracted constants are for 160 °C: $k_1 = 0.19$, $k_2 = 1.3$, $k_3 = 0.105 \text{ h}^{-1}$; for 170 °C: $k_1 = 0.29$, $k_2 = 1.5$, $k_3 = 0.12 \text{ h}^{-1}$; for 190 °C: $k_1 = 0.6$, $k_2 = 3.45$, $k_3 = 0.24 \text{ h}^{-1}$. We calculated the index R^2 to quantify the quality of fitting. Its average value for the three curves of each temperature is $R^2 = 0.92, 0.94, 0.92$ for $T = 190 \text{ °C}, 170 \text{ °C}$ and 160 °C respectively. By observing the experimental data in Fig. 13 it can be argued that the larger part of deviation of R^2 from unity is due to the data uncertainty (scatter) rather than to the performance of the model.

The kinetic constants increase with temperature, so they were placed in an Arrhenius plot and it was found that an Arrhenius temperature dependence is closely followed. These plots are shown in Fig. 14. The corresponding activation energies found for the three reactions constants are $E_1/R_g = 6780 \text{ K}$, $E_2/R_g = 7655 \text{ K}$, and $E_3/R_g = 5726 \text{ K}$ respectively. Summarizing a consistent set of 5 reaction constants following Arrhenius dependence was derived from the existing experimental data by constructing two properly lumped reaction schemes. Such an achievement would not be possible using the detailed reaction scheme due to a large number of undetermined para-

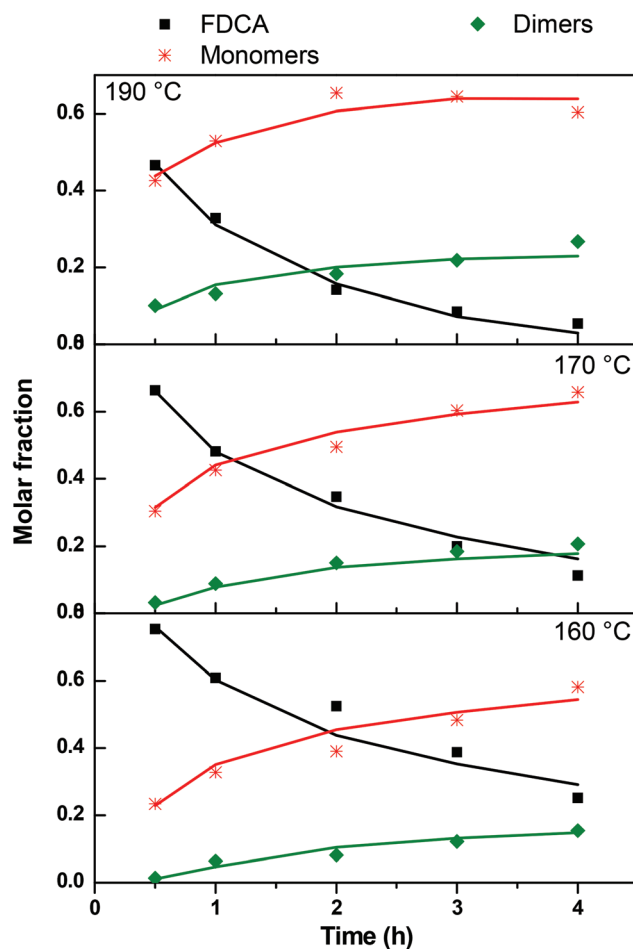


Fig. 13 Comparison between model (solid lines) and experimental data (symbols) of molar fraction evolution of FDCA (F), monomers (M), dimers (D) and trimers (T) during the esterification reaction.

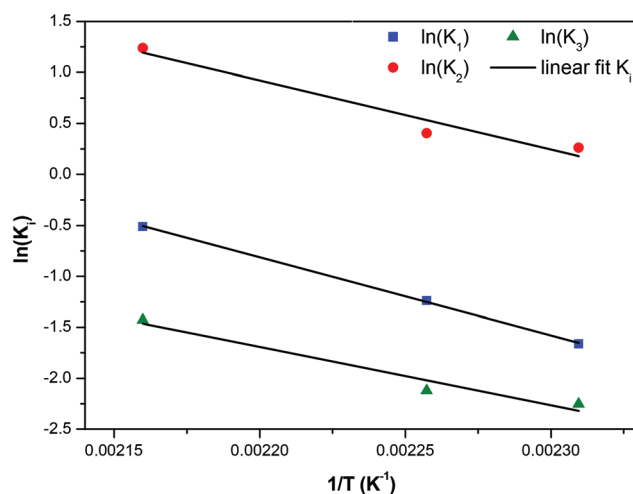


Fig. 14 Arrhenius plot for the constants of reaction scheme (8).



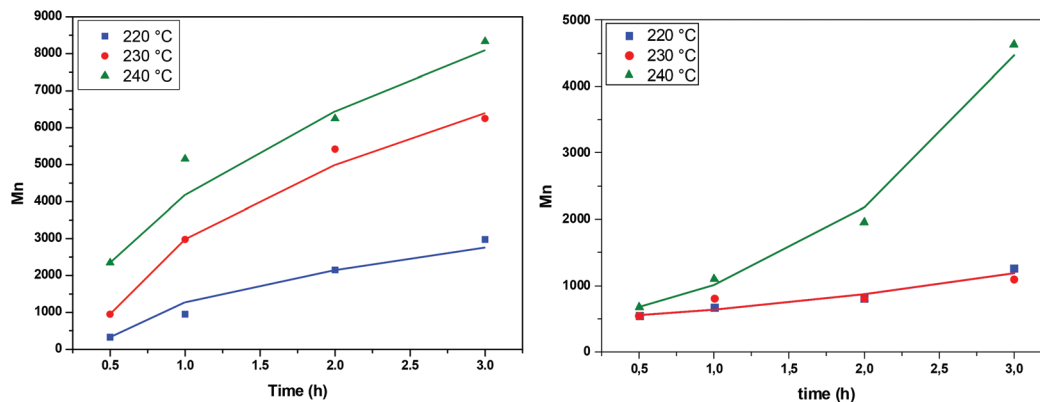


Fig. 15 Evolution of average molecular weight during the transesterification step at different temperatures, catalyzed by Sb₂O₃ (left) and Sb(CH₃COO)₃ (right). Comparison between model (solid lines) and experimental data (symbols).

mers. The successful modelling and parameter estimation using experimental data for the *i*-mer evolution of the esterification reaction of FDCA with EG is an essential innovation of the present work.

4.2. Transesterification – polycondensation reaction

In the present case of reactants there is only one functional group that reacts, so the functional group and the empirical global reaction approaches coincide. Typically, a second order global reaction is assumed.⁸⁴ As an example, in ref. 74 a second order reaction is employed combined to a thermodynamic-mass transfer model for EG (necessary for the experimental setup therein). The fact that the fit leads to the initial mass transfer coefficient varying up to 12 orders of magnitude among the employed experimental conditions suggests that a more general reaction kinetic model must be considered. The transesterification reaction is a linear chain polymerization reaction and the second order implies that the reaction constants do not depend on the chain length of the reactant molecules. This assumption was relaxed by assuming a rate constant decreasing with the chain length of reactants as follows $K_{ij} = K_0(i^{-\lambda} + j^{-\lambda})$ where *i* and *j* are the chain lengths of the reacting molecules. The coagulation population balance with the above rate was derived, and the monodisperse method of moments was applied in order to end up with the following equation for the evolution of the average molecular weight X_w .⁶⁹

$$\frac{dX_w}{dt} = K_R X_w^{-\lambda} \quad (10)$$

where $K_R = K_0 C_0 M_{w_0}$, M_{w_0} is the monomer molecular weight and C_0 is the initial monomer concentration. In order to compare the model results to the experimental ones, the molecular weight X_{w_0} at time $t = t_0$ (equal to 0.5 h in the present case) is considered known and equal to the experimental one. Its subsequent evolution must be described by eqn (9) in which equation can be integrated from t_0 to t to give:

$$X_w = (X_{w_0}^{1+\lambda} + (1+\lambda)K_R(t-t_0))^{1/(1+\lambda)} \quad (11)$$

The above expression is fitted to the experimental molecular weight evolution curves (comparison between the model and experimental data appears in Fig. 15). The averaged fitting coefficient R^2 for the three temperatures is 0.96 and 0.92 for Sb₂O₃ and Sb(CH₃COO)₃, respectively, which are rather acceptable values. The results for antimony oxide are quite expected. A temperature independent value of $\lambda = 1$ (typical of this type of reaction⁶⁹) is found with values of K_R increasing with temperature as follows: $K_R = 1.5 \times 10^6 \text{ h}^{-1}$, $8 \times 10^6 \text{ h}^{-1}$ and $12 \times 10^6 \text{ h}^{-1}$ for $T = 220, 230$ and $240 \text{ }^\circ\text{C}$, respectively. This exponent denotes the decrease of the mobility of the molecules as their molecular weight increases. A quite strange behavior is observed for antimony acetate. The reaction proceeds very slowly (and quite similarly) for 220 °C and 230 °C. In this case the small extent of the reaction and scatter of the data allows fitting using any value of λ (ill-posed problem). The simplest choice is to assume $\lambda = 0$. Surprisingly the reaction becomes accelerated (like a self-catalyzed one) at 240 °C leading to a quite unusual value of $\lambda = -0.9$. The second catalyst appears to have not only small yield but also uncontrollable behavior and for these reasons it is not appropriate for the present process.

5. Conclusions

In the present work, the catalytic activity of two industrial, safe for food packaging applications, antimony catalysts, antimony trioxide, Sb₂O₃, and antimony acetate, Sb(CH₃COO)₃, in the two-step polymerization of FDCA with EG for the production of PEF was investigated. The progress of the esterification step was monitored by end-group analysis (titration of –COOH groups), as well as NMR and FTIR spectroscopy. Based on the obtained conversions, it was deduced that, for the first step of the polymerization, *i.e.* the esterification of FDCA with EG to afford PEF oligomers, Sb₂O₃ seems more active than Sb(CH₃COO)₃ at lower temperatures (160 °C). Increasing the temperature smoothens the differences between the catalysts and at 190 °C satisfactory conversions are obtained. For the first time the structure of the PEF oligomers formed during



the first step of the polymerization was identified by LC-HRMS and the evolution of their concentrations with reaction time was determined. HRMS investigations demonstrated that at 190 °C, in the presence of Sb₂O₃, dimers and trimers had already formed within the first 30 minutes. In the second step of the polymerization, the difference between the two catalysts is much more pronounced, with Sb₂O₃ exhibiting the highest activity. Not surprisingly, a higher temperature also contributes to the formation of higher molecular weight polymers. These experimental observations were complemented by theoretical kinetic investigations and the rate constants for the different reactions taking place during the polymerization were calculated.

Funding

This publication is based upon work from COST Action FUR4Sustain, CA18220, supported by COST (European Cooperation in Science and Technology).

Conflicts of interest

There are no conflicts to declare.

References

- 1 H. Storz and K.-D. Vorlop, *Landbauforsch. Appl. Agric. For. Res.*, 2013, **63**, 321.
- 2 G. Walther, High-performance polymers from nature: catalytic routes and processes for industry, *ChemSusChem*, 2014, **7**, 2081.
- 3 D. K. Schneiderman and M. A. Hillmyer, 50th anniversary perspective: there is a great future in sustainable polymers, *Macromolecules*, 2017, **50**, 3733.
- 4 R. Mülhaupt, Green polymer chemistry and bio-based plastics: dreams and reality, *Macromol. Chem. Phys.*, 2013, **214**, 159.
- 5 A. Llevot, P. K. Dannecker, M. Von Czapiewski, L. C. Over, Z. Söyler and M. A. R. Meier, Renewability is not enough: recent advances in the sustainable synthesis of biomass-derived monomers and polymers, *Chem. – Eur. J.*, 2016, **22**, 11510.
- 6 G. Q. Chen and M. K. Patel, Plastics derived from biological sources: present and future: a technical and environmental review, *Chem. Rev.*, 2012, **112**, 2082.
- 7 W. P. Dijkman, D. E. Groothuis and M. W. Fraaije, Enzyme-catalyzed oxidation of 5-hydroxymethylfurfural to furan-2,5-dicarboxylic acid, *Angew. Chem., Int. Ed.*, 2014, **53**, 6515.
- 8 J. J. Bozell and G. R. Petersen, Technology Development for the Production of Biobased Products from Biorefinery Carbohydrates - The US Department of Energy's "Top 10" Revisited, *Green Chem.*, 2010, **12**(4), 539.
- 9 A. Gandini, A. J. D. Silvestre, C. P. Neto, A. F. Sousa and M. Gomes, The furan counterpart of poly(ethylene terephthalate): An alternative material based on renewable resources, *J. Polym. Sci., Part A: Polym. Chem.*, 2009, **47**(11), 295.
- 10 J. G. van Berkel, N. Guigo, H. A. Visser and N. Sbirrazzuoli, Chain Structure and Molecular Weight Dependent Mechanics of Poly(ethylene 2,5-furandicarboxylate) Compared to Poly(ethylene terephthalate), *Macromolecules*, 2018, **51**(21), 8539.
- 11 X. Fei, J. Wang, J. Zhu, X. Wang and X. Liu, Bio-based Poly(ethylene 2,5-furanoate): No Longer an Alternative, but an Irreplaceable Polyester in the Polymer Industry, *ACS Sustainable Chem. Eng.*, 2020, **8**(23), 8471.
- 12 K. Loos, R. Zhang, I. Pereira, B. Agostinho, H. Hu, D. Maniar, N. Sbirrazzuoli, A. J. D. Silvestre, N. Guigo and A. F. Sousa, A Perspective on PEF Synthesis, Properties, and End-Life, *Front. Chem.*, 2020, **8**, 585.
- 13 A. J. J. E. Eerhart, A. P. C. Faaij and M. K. Patel, Replacing fossil based PET with biobased PEF; process analysis, energy and GHG balance, *Energy Environ. Sci.*, 2012, **5**, 6407.
- 14 E. de Jong, M. A. Dam, L. Sipos and G.-J. M. Gruter, *Furandicarboxylic Acid (FDCA), A Versatile Building Block for a Very Interesting Class of Polyesters*, ACS Symp. Ser., 2012, vol 1105, pp. 1–13.
- 15 A. J. J. E. Eerhart, M. K. Patel and A. P. C. Faaij, Fuels and plastics from lignocellulosic biomass via the furan pathway: an economic analysis, *Biofuels, Bioprod. Biorefin.*, 2015, **9**(3), 307.
- 16 C. F. Araujo, M. M. Nolasco, P. J. A. Ribeiro-Claro, S. Rudić, A. J. D. Silvestre, P. D. Vaz and A. F. Sousa, Inside PEF: Chain Conformation and Dynamics in Crystalline and Amorphous Domains, *Macromolecules*, 2018, **51**(9), 3515.
- 17 E. Forestier, C. Combeaud, N. Guigo, G. Monge, J.-M. Haudin, N. Sbirrazzuoli and N. Billon, Strain-induced crystallization of poly(ethylene 2,5-furandicarboxylate). Mechanical and crystallographic analysis, *Polymer*, 2020, **187**, 122126.
- 18 J. G. van Berkel, N. Guigo, J. J. Kolstad and N. Sbirrazzuoli, Biaxial Orientation of Poly(ethylene 2,5-furandicarboxylate): An Explorative Study, *Macromol. Mater. Eng.*, 2018, **303**(3), 1700507.
- 19 D. G. Papageorgiou, N. Guigo, V. Tsanaktis, S. Exarhopoulos, D. N. Bikiaris, N. Sbirrazzuoli and G. Z. Papageorgiou, Fast Crystallization and Melting Behavior of a Long-Spaced Aliphatic Furandicarboxylate Biobased Polyester, Poly(dodecylene 2,5-furanoate), *Ind. Eng. Chem. Res.*, 2016, **55**(18), 5315.
- 20 E. Forestier, C. Combeaud, N. Guigo, N. Sbirrazzuoli and N. Billon, Understanding of strain-induced crystallization developments scenarios for polyesters: Comparison of poly(ethylene furanoate), PEF, and poly(ethylene terephthalate), PET, *Polymer*, 2020, **203**, 122755.
- 21 L. Maini, M. Gigli, M. Gazzano, N. Lotti, D. N. Bikiaris and G. Z. Papageorgiou, Structural Investigation of Poly(ethylene furanoate) Polymorphs, *Polymer*, 2018, **10**(3), 296.
- 22 G. Z. Papageorgiou, D. G. Papageorgiou, Z. Terzopoulou and D. N. Bikiaris, Production of bio-based 2,5-furan dicar-



- boxylate polyesters: Recent progress and critical aspects in their synthesis and thermal properties, *Eur. Polym. J.*, 2016, **83**, 202.
- 23 Z. Terzopoulou, L. Papadopoulos, A. Zamboulis, D. G. Papageorgiou, G. Z. Papageorgiou and D. N. Bikiaris, Tuning the Properties of Furandicarboxylic Acid-Based Polyesters with Copolymerization: A Review, *Polymer*, 2020, **12**(6), 1209.
- 24 A. F. Sousa, N. Guigo, M. Pożycka, M. Delgado, J. Soares, P. V. Mendonça, J. F. J. Coelho, N. Sbirrazzuoli and A. J. D. Silvestre, Tailored design of renewable copolymers based on poly(1,4-butylene 2,5-furandicarboxylate) and poly(ethylene glycol) with refined thermal properties, *Polym. Chem.*, 2018, **9**, 722.
- 25 M. Soccio, M. Costa, N. Lotti, M. Gazzano, V. Siracusa, E. Salatelli, P. Manaresi and A. Munari, Novel fully bio-based poly(butylene 2,5-furanoate/diglycolate) copolymers containing ether linkages: Structure-property relationships, *Eur. Polym. J.*, 2016, **81**, 397.
- 26 H. Xie, L. Wu, B.-G. Li and P. Dubois, Modification of Poly(ethylene 2,5-furandicarboxylate) with Biobased 1,5-Pentanediol: Significantly Toughened Copolyesters Retaining High Tensile Strength and O₂ Barrier Property, *Biomacromolecules*, 2019, **20**(1), 353.
- 27 M. J. Soares, P.-K. Dannecker, C. Vilela, J. Bastos, M. A. R. Meier and A. F. Sousa, Poly(1,20-eicosanediyl 2,5-furandicarboxylate), a biodegradable polyester from renewable resources, *Eur. Polym. J.*, 2017, **90**, 301.
- 28 L. Genovese, N. Lotti, V. Siracusa and A. Munari, Poly(Neopentyl Glycol Furanoate): A Member of the Furan-Based Polyester Family with Smart Barrier Performances for Sustainable Food Packaging Applications, *Materials*, 2017, **10**(9), 1028.
- 29 M. Matos, A. F. Sousa, A. C. Fonseca, C. S. R. Freire, J. F. J. Coelho and A. J. D. Silvestre, A New Generation of Furanic Copolyesters with Enhanced Degradability: Poly(ethylene 2,5-furandicarboxylate)-co-poly(lactic acid) Copolyesters, *Macromol. Chem. Phys.*, 2014, **215**(22), 2175.
- 30 Y. Jiang, D. Maniar, A. J. J. Woortman and K. Loos, Enzymatic synthesis of 2,5-furandicarboxylic acid-based semi-aromatic polyamides: enzymatic polymerization kinetics, effect of diamine chain length and thermal properties, *RSC Adv.*, 2016, **6**, 67941.
- 31 L. Genovese, M. Soccio, N. Lotti, A. Munari, A. Szymczyk, S. Paszkiewicz, A. Linares, A. Nogales and T. A. Ezquerro, Effect of chemical structure on the subglass relaxation dynamics of biobased polyesters as revealed by dielectric spectroscopy: 2,5-furandicarboxylic acid vs. trans-1,4-cyclohexanedicarboxylic acid, *Phys. Chem. Chem. Phys.*, 2018, **20**, 15696.
- 32 D. Maniar, K. F. Hohmann, Y. Jiang, A. J. J. Woortman, J. van Dijken and K. Loos, Enzymatic Polymerization of Dimethyl 2,5-Furandicarboxylate and Heteroatom Diamines, *ACS Omega*, 2018, **3**(6), 7077.
- 33 G. Guidotti, L. Genovese, M. Soccio, M. Gigli, A. Munari, V. Siracusa and N. Lotti, Block Copolyesters Containing 2,5-Furan and trans-1,4-Cyclohexane Subunits with Outstanding Gas Barrier Properties, *Int. J. Mol. Sci.*, 2019, **20**(9), 2187.
- 34 Y. Jiang, D. Maniar, A. J. J. Woortman, G. O. R. Alberda Van Ekenstein and K. Loos, Enzymatic Polymerization of Furan-2,5-Dicarboxylic Acid-Based Furanic-Aliphatic Polyamides as Sustainable Alternatives to Polyphthalamides, *Biomacromolecules*, 2015, **16**(11), 3674.
- 35 G. Guidotti, M. Soccio, M.-C. García-Gutiérrez, E. Gutiérrez-Fernández, T. A. Ezquerro, V. Siracusa, A. Munari and N. Lotti, Evidence of a 2D-Ordered Structure in Biobased Poly(pentamethylene furanoate) Responsible for Its Outstanding Barrier and Mechanical Properties, *ACS Sustainable Chem. Eng.*, 2019, **7**(21), 17863.
- 36 S. Chen, R. Zou, L. Li, J. Shang, S. Lin and J. Lan, Preparation of biobased poly(propylene 2,5-furandicarboxylate) fibers: Mechanical, thermal and hydrolytic degradation properties, *J. Appl. Polym. Sci.*, 2021, **138**(18), e50345.
- 37 H. Hu, R. Zhang, A. Sousa, Y. Long, W. B. Ying, J. Wang and J. Zhu, Bio-based poly(butylene 2,5-furandicarboxylate)-b-poly(ethylene glycol) copolymers with adjustable degradation rate and mechanical properties: Synthesis and characterization, *Eur. Polym. J.*, 2018, **106**, 42.
- 38 M. Soccio, D. E. Martínez-Tong, A. Alegría, A. Munari and N. Lotti, Molecular dynamics of fully biobased poly(butylene 2,5-furanoate) as revealed by broadband dielectric spectroscopy, *Polymer*, 2017, **128**, 24.
- 39 P. A. Klonos, L. Papadopoulos, D. Tzetzis, A. Kyritsis, G. Z. Papageorgiou and D. N. Bikiaris, Thermal, nanoindentation and dielectric study of nanocomposites based on poly(propylene furanoate) and various inclusions, *Mater. Today Commun.*, 2019, **20**, 100585.
- 40 M. Matos, A. F. Sousa, N. H. C. S. Silva, C. S. R. Freire, M. Andrade, A. Mendes and A. J. D. Silvestre, Furanoate-Based Nanocomposites: A Case Study Using Poly(Butylene 2,5-Furanoate) and Poly(Butylene 2,5-Furanoate)-co-(Butylene Diglycolate) and Bacterial Cellulose, *Polymer*, 2018, **10**(8), 810.
- 41 P. Skoczinski, M. K. Espinoza Cangahuala, D. Maniar, R. W. Albach, N. Bittner and K. Loos, Biocatalytic Synthesis of Furan-Based Oligomer Diols with Enhanced End-Group Fidelity, *ACS Sustainable Chem. Eng.*, 2020, **8**(2), 1068.
- 42 M. Matos, A. F. Sousa, P. V. Mendonça and A. J. D. Silvestre, Co-Polymers based on Poly(1,4-butylene 2,5-furandicarboxylate) and Poly(propylene oxide) with Tuneable Thermal Properties: Synthesis and Characterization, *Materials*, 2019, **12**(2), 328.
- 43 G. Guidotti, M. Soccio, N. Lotti, M. Gazzano, V. Siracusa and A. Munari, Poly(propylene 2,5-thiophenedicarboxylate) vs. Poly(propylene 2,5-furandicarboxylate): Two Examples of High Gas Barrier Bio-Based Polyesters, *Polymer*, 2018, **10**(7), 785.
- 44 Y. Jiang, A. J. J. Woortman, G. O. R. Alberda van Ekenstein and K. Loos, A biocatalytic approach towards sustainable furanic-aliphatic polyesters, *Polym. Chem.*, 2015, **6**, 5198.



- 45 M. Gigli, F. Quartinello, M. Soccio, A. Pellis, N. Lotti, G. M. Guebitz, S. Licoccia and A. Munari, Enzymatic hydrolysis of poly(1,4-butylene 2,5-thiophenedicarboxylate) (PBTF) and poly(1,4-butylene 2,5-furandicarboxylate) (PBF) films: A comparison of mechanisms, *Environ. Int.*, 2019, **130**, 104852.
- 46 B. Robles-Hernández, M. Soccio, I. Castrillo, G. Guidotti, N. Lotti, Á. Alegría and D. E. Martínez-Tong, Poly(alkylene 2,5-furanoate)s thin films: Morphology, crystallinity and nanomechanical properties, *Polymer*, 2020, **204**, 122825.
- 47 N. Pouloupoulou, N. Guigo, N. Sbirrazzuoli, D. G. Papageorgiou, D. N. Bikiaris, G. N. Nikolaidis and G. Z. Papageorgiou, Towards increased sustainability for aromatic polyesters: Poly(butylene 2,5-furandicarboxylate) and its blends with poly(butylene terephthalate), *Polymer*, 2020, 123157.
- 48 G. Guidotti, M. Soccio, M. C. García-Gutiérrez, T. Ezquerro, V. Siracusa, E. Gutiérrez-Fernández, A. Munari and N. Lotti, Fully Biobased Superpolymers of 2,5-Furandicarboxylic Acid with Different Functional Properties: From Rigid to Flexible, High Performant Packaging Materials, *ACS Sustainable Chem. Eng.*, 2020, **8**(25), 9558.
- 49 M. Vannini, P. Marchese, A. Celli and C. Lorenzetti, Fully biobased poly(propylene 2,5-furandicarboxylate) for packaging applications: excellent barrier properties as a function of crystallinity, *Green Chem.*, 2015, **17**, 4162.
- 50 J. C. Morales-Huerta, A. Martínez de Ilarduya and S. Muñoz-Guerra, Sustainable Aromatic Copolyesters via Ring Opening Polymerization: Poly(butylene 2,5-furandicarboxylate-co-terephthalate)s, *ACS Sustainable Chem. Eng.*, 2016, **4**(9), 4965.
- 51 S. Mahmud, J. Wang, N. Shao, Z. Xiong, R. Zhang and J. Zhu, Nucleation and crystallization of poly(propylene 2,5-furan dicarboxylate) by direct blending of microcrystalline cellulose: improved tensile and barrier properties, *Cellulose*, 2020, **27**, 9436.
- 52 J. C. Morales-Huerta, A. Martínez de Ilarduya, S. León and S. Muñoz-Guerra, Isomannide-Containing Poly(butylene 2,5-furandicarboxylate) Copolyesters via Ring Opening Polymerization, *Macromolecules*, 2018, **51**(9), 3340.
- 53 A. F. Sousa, M. Matos, C. S. R. Freire, A. J. D. Silvestre and J. F. J. Coelho, New copolyesters derived from terephthalic and 2,5-furandicarboxylic acids: A step forward in the development of biobased polyesters, *Polymer*, 2013, **54**(2), 513.
- 54 A. F. Sousa, A. C. Fonseca, A. C. Serra, C. S. Freire, A. J. D. Silvestre and J. Coelho, New unsaturated copolyesters based on 2,5-furandicarboxylic acid and their cross-linked derivatives, *Polym. Chem.*, 2016, **7**, 1049.
- 55 M. Soccio, N. Lotti, A. Munari, E. Rebollar and D. E. Martínez-Tong, Wrinkling poly(trimethylene 2,5-furanoate) free-standing films: Nanostructure formation and physical properties, *Polymer*, 2020, **202**, 122666.
- 56 M. Matos, A. F. Sousa and A. J. D. Silvestre, Improving the Thermal Properties of Poly(2,5-furandicarboxylate)s Using Cyclohexylene Moieties: A Comparative Study, *Macromol. Chem. Phys.*, 2017, **218**(5), 1600492.
- 57 J. C. Morales-Huerta, A. Martínez de Ilarduya and S. Muñoz-Guerra, Partially Renewable Poly(butylene 2,5-furandicarboxylate-co-isophthalate) Copolyesters Obtained by ROP, *Polymer*, 2018, **10**(5), 483.
- 58 J. C. Morales-Huerta, A. Martínez de Ilarduya and S. Muñoz-Guerra, Poly(alkylene 2,5-furandicarboxylate)s (PEF and PBF) by ring opening polymerization, *Polymer*, 2016, **87**, 148.
- 59 N. Lotti, A. Munari, M. Gigli, M. Gazzano, V. Tsanakis, D. N. Bikiaris and G. Z. Papageorgiou, Thermal and structural response of in situ prepared biobased poly(ethylene 2,5-furan dicarboxylate) nanocomposites, *Polymer*, 2016, **103**, 288.
- 60 D. Maniar, F. Silvianti, V. M. Ospina, A. J. J. Woortman, J. van Dijken and K. Loos, On the way to greener furanic-aliphatic poly(ester amide)s: Enzymatic polymerization in ionic liquid, *Polymer*, 2020, **205**, 122662.
- 61 A. F. Sousa, J. F. J. Coelho and A. J. D. Silvestre, Renewable-based poly((ether)ester)s from 2,5-furandicarboxylic acid, *Polymer*, 2016, **98**, 119.
- 62 G. Z. Papageorgiou, N. Guigo, V. Tsanakis, D. G. Papageorgiou, S. Exarhopoulos, N. Sbirrazzuoli and D. N. Bikiaris, On the bio-based furanic polyesters: Synthesis and thermal behavior study of poly(octylene furanoate) using fast and temperature modulated scanning calorimetry, *Eur. Polym. J.*, 2015, **68**, 115.
- 63 V. Tsanakis, D. N. Bikiaris, N. Guigo, S. Exarhopoulos, D. G. Papageorgiou, N. Sbirrazzuoli and G. Z. Papageorgiou, Synthesis, properties and thermal behavior of poly(decylene-2,5-furanoate): a biobased polyester from 2,5-furan dicarboxylic acid, *RSC Adv.*, 2015, **5**, 7459.
- 64 A. Martínez de Ilarduya and S. Muñoz Guerra, Ring opening polymerization of macrocyclic oligoesters derived from renewable sources, *Polym. Chem.*, 2020, **11**, 4850.
- 65 D. Maniar, Y. Jiang, A. J. J. Woortman, J. van Dijken and K. Loos, Furan-Based Copolyesters from Renewable Resources: Enzymatic Synthesis and Properties, *ChemSusChem*, 2019, **12**(5), 990.
- 66 J. C. Morales-Huerta, A. Martínez de Ilarduya and S. Muñoz-Guerra, Blocky poly(ϵ -caprolactone-co-butylene 2,5-furandicarboxylate) copolyesters via enzymatic ring opening polymerization, *J. Polym. Sci., Part A: Polym. Chem.*, 2018, **56**(3), 290.
- 67 J. C. Morales-Huerta, C. B. Ciulik, A. Martínez de Ilarduya and S. Muñoz-Guerra, Fully bio-based aromatic-aliphatic copolyesters: poly(butylene furandicarboxylate-co-succinate)s obtained by ring opening polymerization, *Polym. Chem.*, 2017, **8**, 748.
- 68 G. J. M. Gruter, L. Sipos and M. A. Dam, Accelerating Research into Bio-Based FDCA-Polyesters by Using Small Scale Parallel Film Reactors, *Comb. Chem. High Throughput Screening*, 2012, **15**(2), 180.
- 69 Z. Terzopoulou, E. Karakatsianopoulou, N. Kasmi, V. Tsanakis, N. Nikolaidis, M. Kostoglou,



- G. Z. Papageorgiou, D. A. Lambropoulou and D. N. Bikiaris, Effect of catalyst type on molecular weight increase and coloration of poly(ethylene furanoate) bio-based polyester during melt polycondensation, *Polym. Chem.*, 2017, **8**(44), 6895.
- 70 R. J. I. Knoop, W. Vogelzang, J. van Haveren and D. S. van Es, High molecular weight poly(ethylene-2,5-furanoate); critical aspects in synthesis and mechanical property determination, *J. Polym. Sci., Part A: Polym. Chem.*, 2013, **51**(19), 4191.
- 71 M. B. Banella, J. Bonucci, M. Vannini, P. Marchese, C. Lorenzetti and A. Celli, Insights into the Synthesis of Poly(ethylene 2,5-Furandicarboxylate) from 2,5-Furandicarboxylic Acid: Steps toward Environmental and Food Safety Excellence in Packaging Applications, *Ind. Eng. Chem. Res.*, 2019, **58**(21), 8955.
- 72 Z. Terzopoulou, E. Karakatsianopoulou, N. Kasmi, M. Majdoub, G. Z. Papageorgiou and D. N. Bikiaris, Effect of catalyst type on recyclability and decomposition mechanism of poly(ethylene furanoate) biobased polyester, *J. Anal. Appl. Pyrolysis*, 2017, **126**, 357.
- 73 M. Lomeli-Rodriguez, M. Martin-Molina, M. Jimenez-Pardo, Z. Nasim-Afzal, S. I. Cauet, T. E. Davies, M. Rivera-Toledo and J. A. Lopez-Sanchez, Synthesis and kinetic modeling of biomass-derived renewable polyesters, *J. Polym. Sci., Part A: Polym. Chem.*, 2016, **54**(18), 2876.
- 74 A. L. T. Brandão, B. F. Oechsler, F. W. Gomes, F. G. Souza Jr. and J. C. Pinto, Modeling and parameter estimation of step-growth polymerization of poly(ethylene-2,5-furandicarboxylate), *Polym. Eng. Sci.*, 2018, **58**(5), 729.
- 75 M. Toshiaki, K. Masayuki, K. Takeyuki and M. Norifumi, *US Pat*, US20140024793A1, 2013.
- 76 V. Tsanaktsis, G. Z. Papageorgiou and D. N. Bikiaris, A Facile Method to Synthesize High-Molecular-Weight Biobased Polyesters from 2,5-Furandicarboxylic Acid and Long-Chain Diols, *J. Polym. Sci., Part A: Polym. Chem.*, 2015, **53**(22), 2617.
- 77 A. S. Joshi, N. Alipourasiabi, Y.-W. Kim, M. R. Coleman and J. G. Lawrence, Role of enhanced solubility in esterification of 2,5-furandicarboxylic acid with ethylene glycol at reduced temperatures: energy efficient synthesis of poly(ethylene 2,5-furandicarboxylate), *React. Chem. Eng.*, 2018, **3**, 447.
- 78 T. Yamada and Y. Imamura, Simulation of Continuous Direct Esterification Process between Terephthalic Acid and Ethylene Glycol, *Polym.-Plast. Technol. Eng.*, 1989, **28**(7–8), 811.
- 79 H. Patel, G. Feix and R. Schomäcker, Modeling of Semibatch Esterification Process for Poly(ethylene terephthalate) Synthesis, *Macromol. React. Eng.*, 2007, **1**(4), 502.
- 80 M. Krumpolc and J. Malek, Esterification of Benzenecarboxylic Acids with Ethylene Glycol. 4. Kinetics of the initial stage of polyesterification of terephthalic acid with ethylene glycol catalyzed by zinc oxide, *Makromol. Chem.*, 1973, **171**(1), 69.
- 81 A. E. Ardis and A. A. Vaitekunas, *DE Pat*, DE1198343B, 1965.
- 82 M. Gomes, A. Gandini, A. J. D. Silvestre and B. Reis, Synthesis and Characterization of Poly(2,5-furan dicarboxylate)s Based on a Variety of Diols, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**(17), 3759.
- 83 D. N. Bikiaris and D. S. Achilias, Synthesis of poly(alkylene succinate) biodegradable polyesters I. Mathematical modelling of the esterification reaction, *Polymer*, 2006, **47**(13), 4851.
- 84 D. N. Bikiaris and D. S. Achilias, Synthesis of poly(alkylene succinate) biodegradable polyesters, Part II: Mathematical modelling of the polycondensation reaction, *Polymer*, 2008, **49**(11), 3677.

