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P and N type copper phthalocyanines as effective semiconductors in organic thin-film transistor based DNA biosensors at elevated temperatures†

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Many health-related diagnostics are expensive, time consuming and invasive. Organic thin film transistor (OTFT) based devices show promise to enable rapid, low cost diagnostics that are an important aspect to enabling increased access and availability to healthcare. Here, we describe OTFTs based upon two structurally similar P (copper phthalocyanine – CuPc) and N (hexadecafluoro copper phthalocyanine – F₁₆-CuPc) type semiconductor materials, and demonstrate their potential for use as both temperature and DNA sensors. Bottom gate bottom contact (BGBC) OTFTs with either CuPc or F₁₆-CuPc semiconducting layers were characterized within a temperature range of 25 °C to 90 °C in both air and under vacuum. CuPc devices showed small positive shifts in threshold voltage (V_T) in air and significant linear increases in mobility with increasing temperature. F₁₆-CuPc devices showed large negative shifts in V_T in air and linear increases in mobility under the same conditions. Similar OTFTs were exposed to DNA in different hybridization states and both series of devices showed positive V_T increases upon DNA exposure, with a larger response to single stranded DNA. The N-type F₁₆-CuPc devices showed a much greater sensing response than the P-type CuPc. These findings illustrate the use of these materials, especially the N-type semiconductor, as both temperature and DNA sensors and further elucidate the mechanism of DNA sensing in OTFTs.

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Introduction

Organic thin films transistors (OTFTs) have shown promise as sensors for detecting various biological analytes such as glucose,¹ DNA,² thrombin,³ bovine serum albumin⁴ and brain injury markers.⁵ OTFTs and their functionally related cousins organic electrochemical transistors (OECTs) are well suited as biological sensors as they can be low-cost, disposable, and mechanically robust.^{6–8} While many OTFT biosensors are not able to match state of the art bioanalytical methods detection limits and sample complexities, they are well positioned to soon serve as rapid and low-cost point of care diagnostics.

DNA sequencing and sensing has rapidly developed since the first human genome was mapped.⁹ While many new technologies have been developed that have increased read times, decreased equipment footprint and lowered cost,^{10,11} there is still a wide margin for improvement. In particular, there are situations in which low cost, high throughput, point of care sensors that do not rely upon amplification would be useful,

such as in infectious disease detection.^{12,13} OTFT based DNA detection technology is well suited for these applications due to its potential for high sensitivity^{14,15} and low cost manufacturing.

Currently, various groups have investigated the use of OTFTs as DNA sensors. These devices detect target DNA strands by capturing double stranded DNA (dsDNA) onto the active layer of the OTFT. Physical adsorption of DNA,¹⁶ electro-immobilization,¹⁷ and chemical immobilization¹⁸ have all been investigated for fixing dsDNA, or single stranded DNA (ssDNA) probes, to the surface of an electrode or the semiconductor material itself. ssDNA has a linear structure comprised of four different bases which will bind with a complementary ssDNA strand to form a double helix (dsDNA) that orders the π orbitals of the bases. Upon applying a bias, charge hopping will occur across these bases and thus through the dsDNA.^{19,20} Additionally, each strand has a negatively charged phosphate backbone. When the probe and target ssDNA are captured at the semiconductor surface of an OTFT, there is an increase in negative charge at that surface, due to these phosphates, that can interact with the charge carriers present in the film. This results in a change in the electrical environment of the semiconducting active layer and therefore a measurable change in the OTFT's threshold voltage (V_T), field-effect mobility (μ) and/or on/off ratio ($I_{on/off}$). For example, Zhang and Subramanian found that upon exposure of a pentacene based OTFT to dsDNA, a positive shift in V_T

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of 19.6 V is observed.² Similarly Gui and Wang used pentacene OTFTs with an additional thin layer of CuPc as an interface for DNA to adsorb onto – they observed a positive shift in V_T of 8 V.¹⁶ Liu *et al.* exploited the net negative charge of ssDNA to improve sensitivity through an increase in immobilization efficiency by applying a positive bias during the immobilization period on a pentacene OTFT.¹⁷ In all these reports and others in the literature, the researchers used exclusively P-type semiconductors, such as pentacene, in a variety of OTFT DNA sensor device architectures.^{18,21-24}

An aspect of sensor design for most OTFT-based DNA sensors that has been overlooked is the required elevated temperature for DNA binding. To ensure specific binding of DNA, either at the point of detection or before, the strands must be in solution at a specific setpoint below the melting temperature (T_M) of the particular DNA sequences being investigated.²⁵⁻²⁷ Therefore it is imperative that the OTFT devices be operated at elevated temperature (typically optimal binding occurs between 40 °C to 70 °C depending on the particular DNA sequence) to ensure specific DNA binding and to reduce non-specific binding. Typically, a temperature of $T_M - 5$ °C is considered optimal for specific binding of complementary DNA strands. Currently only Gui *et al.* have investigated elevated temperature operation of OTFT DNA sensors.²⁸ They investigated the effect of hybridization times at $T = 20$ °C, 45 °C and 60 °C on sensor sensitivity. For their particular DNA sequence, they found that optimal sensor response occurred at 45 °C (13 °C below their sequence T_M). To the best of our knowledge there are no reports of OTFT-based DNA sensors which utilize N-type semiconductors or that operate just below the T_M value of the DNA analytes they are using.

Studies on the effect of operating temperature on the performance of organic semiconductors are uncommon,²⁹⁻³⁴ and the majority of the studies that have been conducted are utilizing P-type semiconductors such as pentacene. An improved and varied understanding of temperature effects on different charge carriers in OTFTs could lead to them being used in a myriad of important applications. For example, many medical therapies require precise temperature control at a patient surface, such as in hypothermia therapy,^{35,36} laser therapy,³⁷ and cryosurgery.³⁸ Often, IR thermography is used to measure patient surface temperature, but unfortunately this can be inaccurate, especially when various topical substances are used during treatment such as ultrasound gel.³⁹ Temperature sensing capabilities would also be useful in such applications as electronic skin, and smart fabrics. Most studies on varied OTFT operating temperature can be divided between low temperature (below room temperature), and high temperature (above room temperature) investigations. Many studies focusing on low temperature operation are doing so to investigate charge conduction models and factors affecting OTFT response to variable temperature operation. For example, Lin & Hung found that in pentacene OTFTs V_T increases positively with increasing temperature between $T = -150$ °C and $T = 25$ °C, a result they hypothesize is due to deep hole trapping.²⁹ Similarly, Sun *et al.* found that in the temperature range $T = -213$ °C to $T = 17$ °C pentacene OTFTs threshold voltage

increases linearly with increasing temperature and that humidity has significant effects on the number of deep hole traps.³⁰ Only a single study by Chesterfield R. J. *et al.* has investigated the performance of an N-type semiconductor (PTCDI-C₅) in OTFTs at variable low temperature operation ($T = -173$ °C to $T = 25$ °C) and it was found to have a thermally activated mobility.³² Finally, some groups have investigated elevated temperature operation of OTFTs and the use of OTFTs as temperature sensors. Jung *et al.*, among others, have investigated the use of pentacene based OTFTs at temperatures ranging from 0 °C to 90 °C and have observed measurable changes in the source/drain current (I_{DS}), μ , and V_T .^{34,40,41} Increasing temperature above room temperature in air led to decreased mobility, a more positive V_T , and increased off current in two P-type semiconducting polymers. This was hypothesized to occur due to polymer oxidation and gas adsorption.³¹ Most successfully, Ren *et al.* constructed a pentacene based flexible temperature sensor, measuring changes in I_{DS} , and enhancing sensitivity by incorporating a silver nanoparticle layer.³³ To the best of our knowledge, no reports have explored the investigation of N-type materials in OTFTs above 25 °C.

In this study, we report the first OTFT DNA sensors using an N-type semiconductor, copper hexadecafluorophthalocyanine (F₁₆-CuPc) (Fig. 1b), and compare it to a structurally similar P-

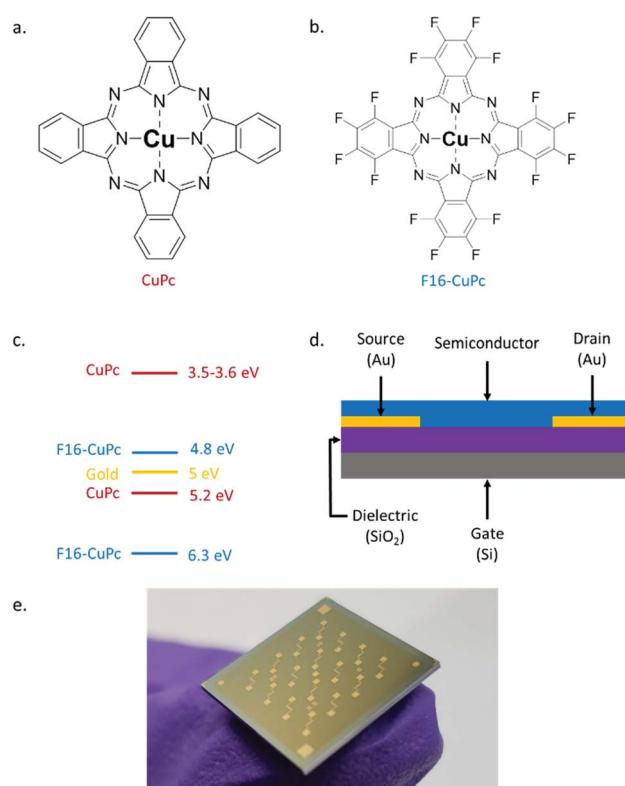


Fig. 1 (a) Structure of copper phthalocyanine (CuPc) (b) structure of copper hexadecafluorophthalocyanine (F₁₆-CuPc). (c) HOMO/LUMO levels of CuPc^{43,44} and F₁₆-CuPc⁴⁵ relative to the work function of gold (yellow). (d) Bottom gate bottom contact organic thin transistor (OTFT) structure. (e) Picture of one actual Fraunhofer device.



type material: copper phthalocyanine (CuPc) (Fig. 1a). We also investigate the effects of ambient temperature (between 25 °C to 90 °C) operation of OTFTs constructed with each of the semiconducting materials in both air and vacuum ($P < 0.1$ Pa). Both CuPc and F_{16} -CuPc are robust materials that make air-stable OTFTs that have been extensively researched and characterized,⁴² thus making good material candidates for investigation. As the negatively charged DNA backbone can affect the positive and negatively charged carriers differently, resulting devices are expected to have distinct operational responses which could then be detected separately or utilized together to enhance sensitivity/selectivity in a DNA sensor.

Results and discussion

Effect of OTFT operation temperature

Bottom gate, bottom contact (BGBC) OTFTs with an interfacial trichloro(octyl)silane (OTS) layer were fabricated by thermal vacuum deposition with either CuPc or F_{16} -CuPc as the semiconducting layer on heavily doped silicon substrates with a thermally grown silicon dioxide dielectric. The typical output and transfer curves for baseline devices characterized at 25 °C in air are shown in Fig. 2a-d for both CuPc and F_{16} -CuPc.

Our research group has recently found significant changes in P-type small molecules when tested in air compared to vacuum ($P < 0.1$ Pa)⁴⁶ and demonstrated differences in temperature

response under these conditions for two P-type semiconducting polymers.³¹ Therefore, we characterized these CuPc and F_{16} -CuPc devices at discrete temperatures ranging from $T = 25$ °C to $T = 90$ °C in air and from 25 °C to $T = 100$ °C in vacuum. Due to equipment heating limitations, we were unfortunately not able to test at temperatures greater than 90 °C in air. As temperature increased in air a noticeable increase in hole mobility (μ_H) and a slight positive shift in V_T was observed for CuPc. This change is well illustrated in Fig. 3a as a shift in the positive (+) direction and up in the μ_H as a function of gate voltage (V_{GS}) graph. For F_{16} -CuPc devices, a large negative shift in V_T and a small increase in electron mobility (μ_E) were observed (Fig. 3b). To determine the impact of environment, identical devices that had never been exposed to air were characterized under vacuum. The resulting plots can be found in Fig. 3c and d for CuPc and F_{16} -CuPc, respectively. The low pressure environment slightly improved the F_{16} -CuPc devices, increasing μ_E , while the opposite was true for CuPc devices. From this baseline, similar changes in V_T and μ_H were observed with increasing temperature for CuPc as in air. F_{16} -CuPc, on the other hand, lacked the large negative V_T shift observed in air with increasing temperature, demonstrating primarily a small increase in μ_E as with CuPc under both conditions.

The average μ_H in CuPc devices in air increased by about 1% °C⁻¹ increase in temperature from 25 °C to 90 °C with a coefficient of determination of $R^2 = 0.995$ (Fig. 4a). In the same

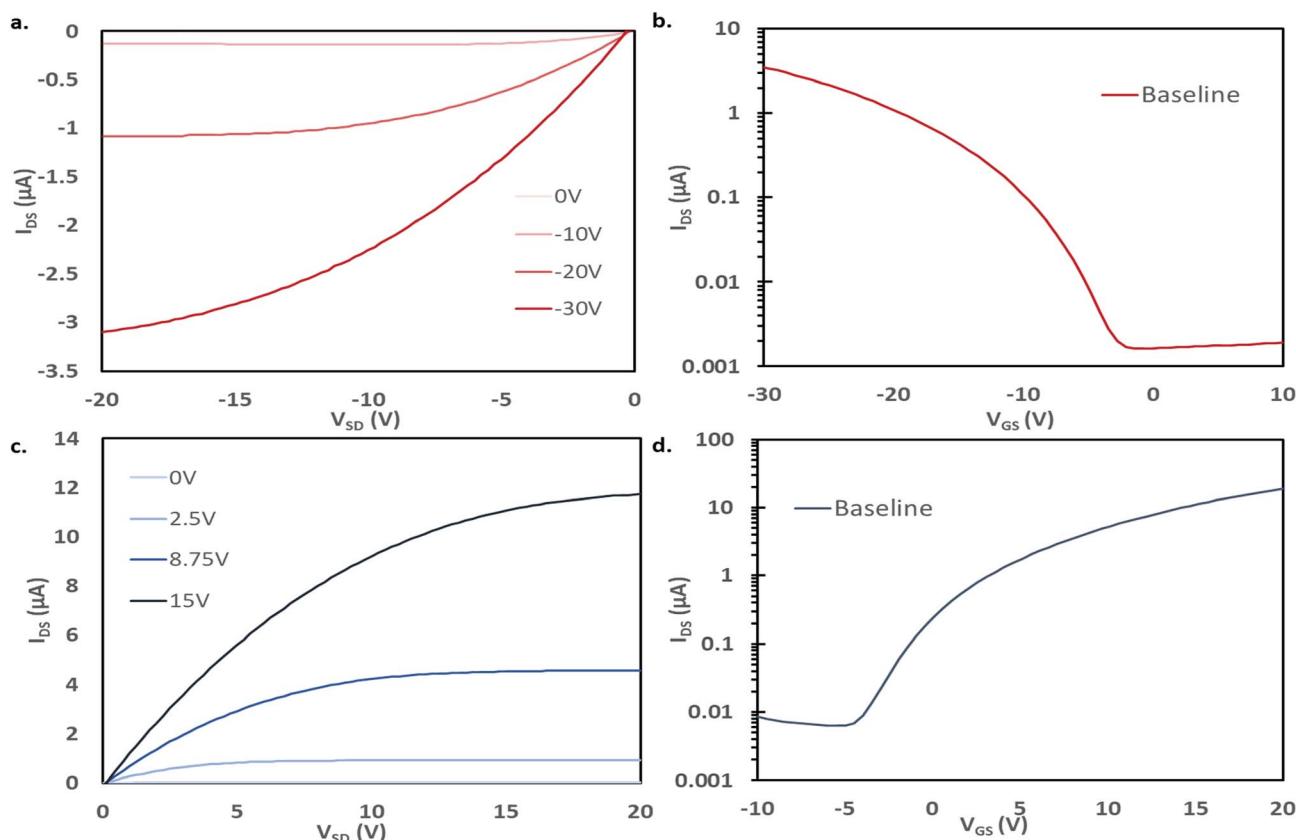


Fig. 2 Output and transfer curves for baseline BGBC OTFTs tested at 25 °C. (a) Output curve and (b) transfer curve ($V_{DS} = -20$ V) for CuPc devices. (c) Output curve and (d) transfer curve ($V_{DS} = 20$ V) for F_{16} -CuPc devices.



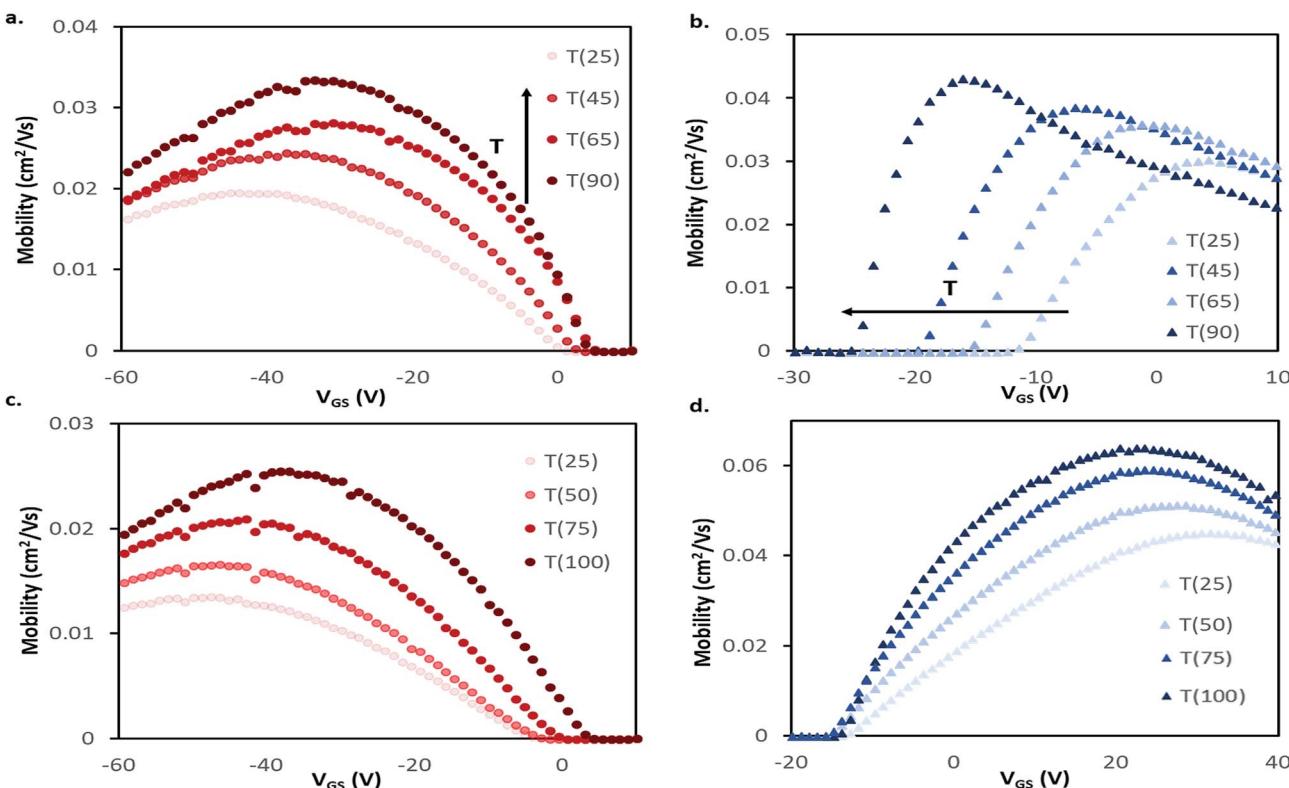


Fig. 3 Field-effect mobility for (a and c) CuPc and (b and d) F₁₆-CuPc BGBC devices deposited at $T = 140$ °C with respect to applied gate-source voltage (V_{GS}) for characteristic devices at varied temperatures in air (a and b) and vacuum (c and d). This mobility was calculated between adjacent points in the transfer data using eqn (2). Devices were tested in the range of $T = 25$ °C to $T = 90$ °C in air, and $T = 25$ °C to $T = 100$ °C in vacuum.

temperature range, V_T shifted from about -7.6 V to -1.8 V, which correlates to a change of about $0.11\text{ V}^{\circ}\text{C}^{-1}$. This value in the units of $\text{V}^{\circ}\text{C}^{-1}$ can be defined as the sensitivity of the device as it shows how much the output value (V) changes with changing input that is being measured (°C), this also holds for the mobility % change per °C values. The change in μ_e of F₁₆-CuPc based OTFTs was smaller, about $0.1\% \text{ }^{\circ}\text{C}^{-1}$ with $R^2 = 0.785$. However, F₁₆-CuPc based OTFTs experienced a significant change in V_T between $T = 25$ °C and 90 °C as seen in Fig. 4b. A V_T shift was measured from -9.9 V to -26.4 V, correlating to a shift of $-0.25\text{ V}^{\circ}\text{C}^{-1}$. As expected, the $I_{on/off}$ ratios for both materials changed similarly to the changes in mobility. Similar effects were observed under vacuum (Fig. 4d-f) except F₁₆-CuPc devices appeared to have little or no significant change in V_T .

Identical experiments were performed on devices that were fabricated through thermal evaporation on heated substrates ($T = 25$ °C), which is significantly cooler than those described above which were fabricated with substrates heated to 140 °C during deposition. The results obtained from these devices can be seen in Fig. S1 and S2.† These devices displayed similar trends except for a shift in V_T for the F₁₆-CuPc devices in air that was not as large, $\Delta V_T = 12.1$ V (it shifted with a rate of $0.19\text{ V}^{\circ}\text{C}^{-1}$).

For the P-type CuPc, the small positive shift in V_T is similar to what has been reported for pentacene.³⁴ This shift in V_T could be explained by a decreased number of positive carriers

being confined to a trap state at higher temperature, as charge carriers have been shown to be affected by thermal activation.⁴¹ The increase in mobility with temperature is known to occur as the generally accepted charge conduction mechanisms of OTFTs are temperature dependent.^{32,47} The same mechanism might explain the increase in electron mobility with rising temperature observed for F₁₆-CuPc in both air and vacuum. However, a large negative change in V_T with increasing temperature is observed in air, shifting the devices to an “on” state without a gate bias. Since the shift is not observed in vacuum, it is likely caused by some component of air such as oxygen or water. Changes in V_T can be attributed to energetically deep traps, or gate bias stress effects,³² with negative changes associated with hole traps or accumulation of positive charge. Typically, oxygen suppresses electron transport and can act as a hole dopant in some materials, so it is difficult to explain how it might shift the operating bias negatively. It has been reported that devices with SiO₂ dielectrics have positive threshold voltage shifts upon exposure to ambient air due to water interacting with dangling -OH groups at the insulator surface.⁴⁸ Thus, it's possible that upon increasing the temperature of the devices, water desorption is shifting the threshold voltage of the devices negatively. Although this phenomena would affect both CuPc and F₁₆-CuPc devices, the differing chemical structure or morphology of the semiconductors may explain the differing response however further investigation is warranted.



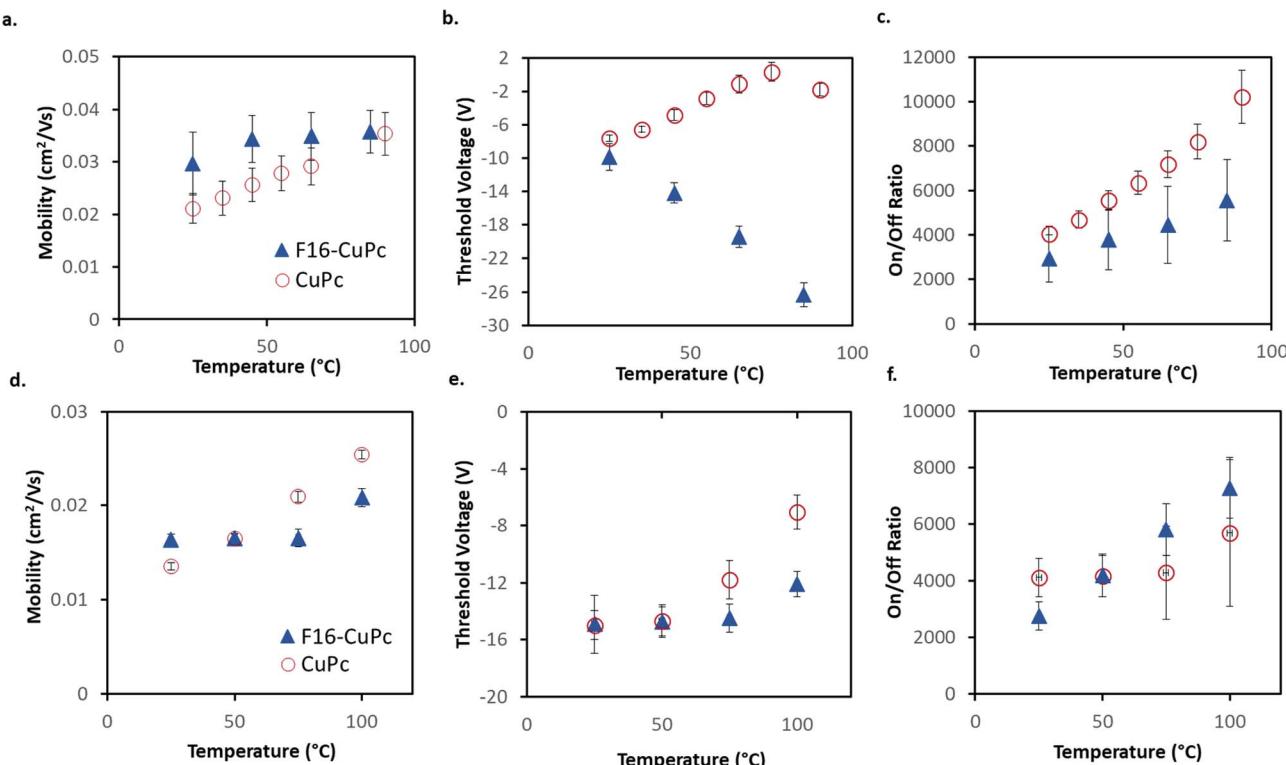


Fig. 4 Performance of BGBC CuPc and F₁₆-CuPc devices (deposited at $T = 140$ °C) in air (a, b and c) or vacuum (d, e and f) at various temperatures. (a and d) Field-effect mobility. (b and e) Threshold voltage. (c and f) On/off ratio. Presented are the averages for four devices with error bars representing the standard deviation. The legend in (a and d) is the same as in (b and e) and (c and f). Devices were tested in the range of $T = 25$ °C to $T = 90$ °C in air, and $T = 25$ °C to $T = 100$ °C in vacuum.

To investigate film morphology further, AFM images were taken of 15 nm films of both CuPc and F₁₆-CuPc deposited at both $T = 140$ °C and $T = 25$ °C. These images can be seen in Fig. 5. The above films were found to have root mean square (R_q) values of 1.84 nm, and 1.21 nm for high temperature (140 °C) deposited CuPc and F₁₆-CuPc films respectively. The low temperature (25 °C) films had R_q values of 1.93 nm (CuPc) and 1.53 nm (F₁₆-CuPc). Thus, the F₁₆-CuPc are smoother in both cases, while the smoothest films were obtained at the higher deposition temperatures. It is also apparent that high temperature deposition of F₁₆-CuPc (Fig. 5b) films have larger grains than the CuPc films (Fig. 5a). Grain sizes between the low temperature films are difficult to distinguish by visual inspection (Fig. 5c and d). Between the two F₁₆-CuPc samples, the smoother sample and the sample with the larger grain size (Fig. 5b) experience a larger ΔV_T over the temperature range. This is also true for the CuPc samples, although the ΔV_T is in the positive direction. These changes in surface area for gas adsorption appear to reflect a greater electronic sensitivity towards temperature in air.

These results discussed illustrate that both CuPc and F₁₆-CuPc based OTFTs can operate between $T = 25$ °C and 90 °C with controlled and predictable changes in performance as a function of operating temperature. This indicates that both molecules could function as components of OTFT-based temperature sensors.

DNA hybridization sensing

To investigate DNA hybridization sensing with OTFTs, a series of analytes were pipetted onto either CuPc or F₁₆-CuPc based devices. For each of these analytes, the base device was first characterized, followed by the analyte deposition, rinsing with deionized water, drying under vacuum for 3 minutes, and then re-characterized. During this series of experiments, the OTFTs were operated at 51.1 °C or 25.0 °C. The elevated temperature is equal to $T_M - 5$ °C for the specific complementary probe and target DNA sequences used. As outlined, this is important to ensure specific and efficient hybridization of the complementary strands. If the temperature at which binding occurs is too high, then binding will not occur due to it being thermodynamically unfavourable. If the temperature is too low, then unspecific binding can occur. This procedure facilitated the comparison of affects across different analytes on the devices. The charge mobilities for both CuPc and F₁₆-CuPc decreased between 60–70% from their initial values no matter the analyte added. There was no statistically significant difference between buffer, ssDNA, and dsDNA; suggesting the buffer and the procedure itself impacts or degrades the semiconducting material/structure and function. As such, the on/off ratios for each analyte also decreased but with no significant differences between these analytes. However, the change in V_T values did vary significantly between different analytes. Fig. 6a and



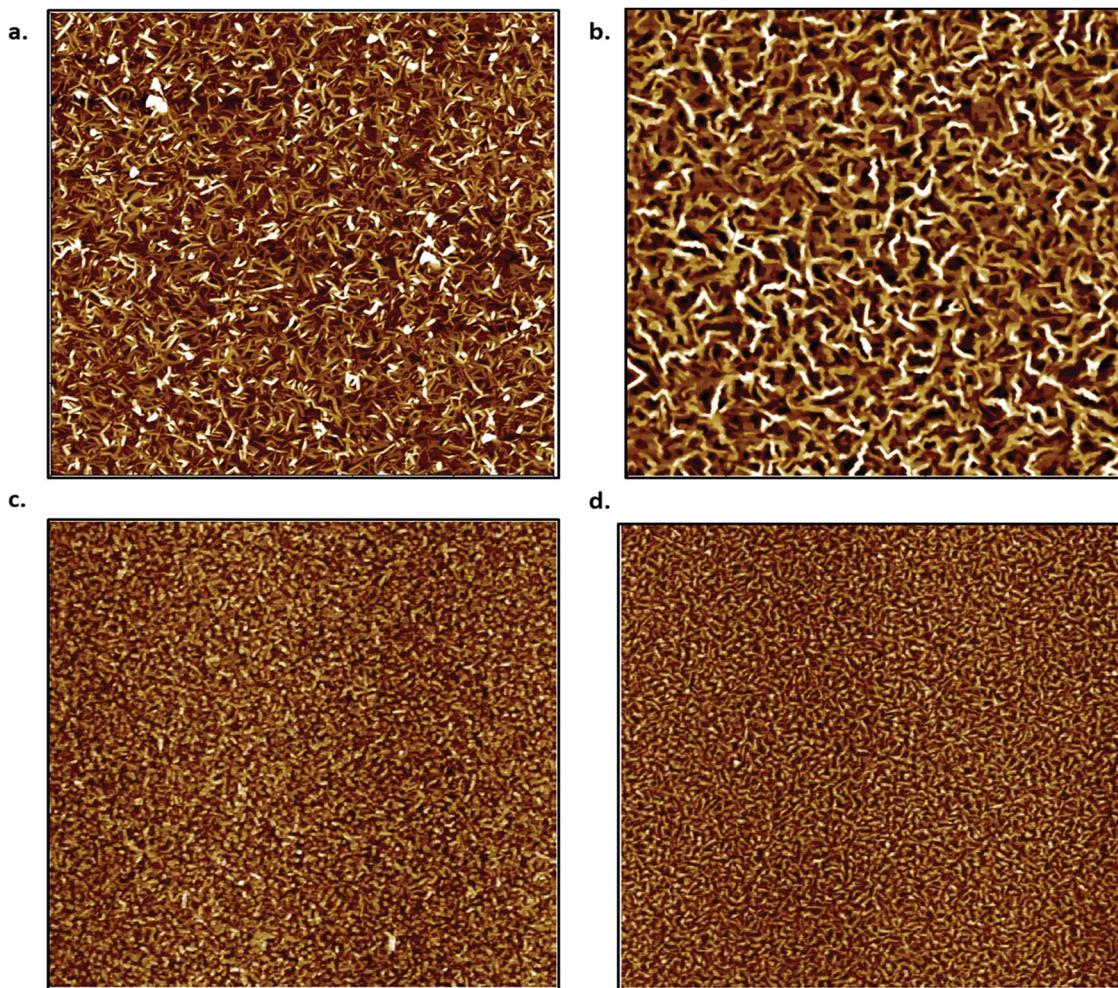


Fig. 5 AFM images of CuPc (a and c) and F_{16} -CuPc (b and d) thin films deposited on SiO_2 substrates at $140\text{ }^\circ\text{C}$ (a and b) and $20\text{ }^\circ\text{C}$ (c and d). Images (a and b) are $5\text{ }\mu\text{m} \times 5\text{ }\mu\text{m}$ while (c and d) are $2.5\text{ }\mu\text{m} \times 2.5\text{ }\mu\text{m}$.

b show the changes in the transfer curve for CuPc and F_{16} -CuPc respectively. These curves illustrate clear shifts in V_T as well as changes in μ and I_{DS} with exposure to different DNA analytes but the difference between ssDNA and dsDNA were determined as statistically insignificant. At $51.1\text{ }^\circ\text{C}$, an average positive shift of $V_T = +2.8\text{ V}$ (from baseline) was observed for CuPc based OTFT after buffer addition and an average positive shift of $V_T = +8.8\text{ V}$ (from baseline) after ssDNA addition and $V_T = +2.6\text{ V}$ after dsDNA addition (from baseline) (Fig. 6c). Similarly to CuPc, F_{16} -CuPc based OTFTs also experienced a positive shift compared to baseline $\Delta V_T = +2.3\text{ V}$, $+21.5\text{ V}$ and $+12.5\text{ V}$ with the addition of buffer, ssDNA and dsDNA, respectively. Additionally, the exact same experiments were performed on F_{16} -CuPc devices but operated at $25.0\text{ }^\circ\text{C}$. This is seen in the inset bars of Fig. 6d. The threshold voltage shifts of ssDNA and dsDNA were $V_T = +6.4\text{ V}$, and $+8.8\text{ V}$, respectively, but were determined as statistically insignificant.

Similarly to what is observed for CuPc devices, reports employing pentacene as a semiconductor also found small positive V_T shifts and decreases in I_{DS} with the addition of

ssDNA.^{49,50} Gui *et al.* reported larger V_T shifts around 8 V for a pentacene device that utilized a thin CuPc layer that acted as an adsorption site for DNA as well as environmental protection for the pentacene semiconducting layer.¹⁶ A positive shift in V_T is often associated with electron trapping, but more directly it is associated with the accumulation of negative charges which would electrostatically facilitate the injection of holes and oppose that of electrons.⁵¹ Thus, positive changes in V_T upon addition of DNA could be explained by the negative charge associated with the phosphate groups in the DNA backbone. The addition of dsDNA to CuPc OTFTs resulted in a positive shift of $V_T = +2.6\text{ V}$ (from baseline) which is much less than the increase seen with ssDNA, $V_T = +8.8\text{ V}$ (Fig. 6c). At surfaces, ssDNA has a much higher effective density than dsDNA due to its more flexible structure.⁵² This difference is due to the relative chain rigidity and intermolecular coulombic repulsions of dsDNA.⁵³ The higher chain density for ssDNA might lead to a higher charge density at the surface of the semiconducting material compared to dsDNA, regardless of dsDNA having double the negative charge per molecule. This might explain the larger threshold voltage shift observed for ssDNA compared to dsDNA.

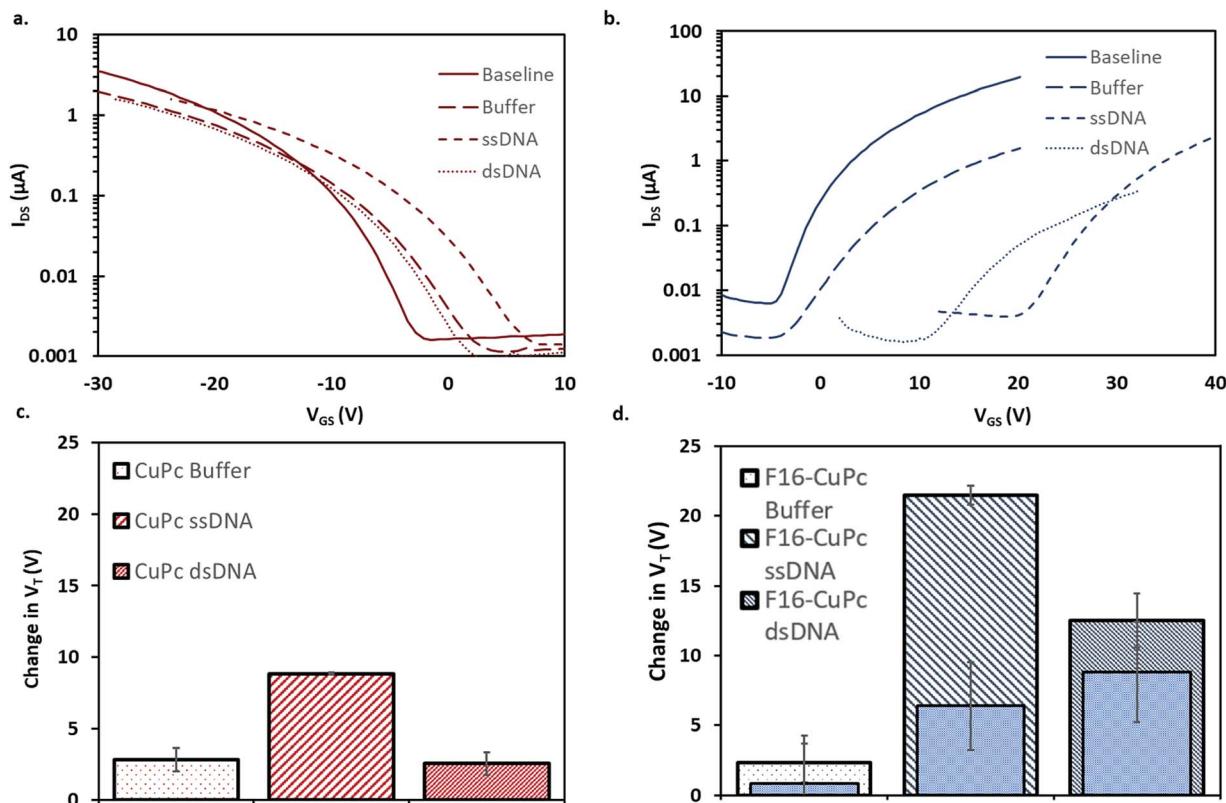


Fig. 6 Transfer curves (a and b) and absolute change in threshold voltage (c and d) of 1 μ L of 1 μ M ssDNA, 1 μ M dsDNA or buffer were added to baseline of CuPc OTFT (a and c) and F₁₆-CuPc (b and d). All characterization was performed at 51.1 °C in air except for the superimposed inner bars in (d) (filled blue rectangle) which were characterized at 25.0 °C.

F₁₆-CuPc based OTFTs also experienced a positive shift compared to baseline. These results, shown in Fig. 6b and d, again show a significant difference in ΔV_T between ssDNA and dsDNA, suggesting reduced charge density at the semiconductor surface for dsDNA analytes. A larger ΔV_T is seen in F₁₆-CuPc than for CuPc. This mirrors the larger response to temperature seen in F₁₆-CuPc in air, although the threshold voltage shift is in the opposite direction. Perhaps the film morphology or energetics of the N-type F₁₆-CuPc make it more susceptible to electron trapping upon addition of various aqueous analytes. The larger grains, and seemingly less dense film, may be more permeable to strands of DNA, increasing the relative changes observed.

Finally, as seen in the superimposed inner bars in Fig. 6d, the sensing response between ssDNA and dsDNA when sample injection and device operation is at $T = 25$ °C, is not significantly different. This is expected as DNA requires an elevated temperature approaching its T_M to bind specifically. Thus it's expected that the two samples are in similar binding states even though one sample contains complementary DNA (dsDNA label) while the other does not (ssDNA label). Of note, is that the ssDNA response at lower temperature is much lower than the ssDNA response at higher temperature. Thus it appears that higher temperature device operation alone could increase sensing response for DNA. This could be due to differing analyte interactions at the surface of the film at different operation temperatures.

Lastly, we investigated the effect of dsDNA concentration on the change in V_T for F₁₆-CuPc based OTFTs. It was found that the effective concentration range of the F₁₆-CuPc sensor is at least between 0.01 μ M and 0.1 μ M, and as the concentration of DNA increases, so too does the magnitude of the change in V_T within the specified range. Further experiments at various concentrations is required to establish the full operating window, including the limit of detection. Further steps could be necessary to maximize this operating window, such as modifying the electrode design, channel geometry or materials. Regardless, these results illustrate that N-type semiconductors such as F₁₆-CuPc can be utilized as the sensor element for dsDNA through simple physical adsorption.

Experimental

Materials

Copper phthalocyanine (CuPc, 90%), and copper(II) 1,2,3,4,8,9,10,11,15,16,17,18,22,23,24,25-hexadecafluoro-29H,31H-phthalocyanine (F₁₆-CuPc, >99.9%) were obtained from TCI Chemicals. CuPc was purified twice by train sublimation before use. All chemicals were used as received unless otherwise specified. The following single stranded deoxyribonucleic acid (DNA) oligonucleotides were purchased from Integrated DNA Technologies.

DNA	Sequence (5'-3', 20bp)	T_M °C ^a
Probe	CAC ACG GAA CTG AAC AAG GTC	56.1
Target (complementary to probe)	GAC CTT GTT CAG TTC CGT GTG	56.1
Random control	GAG TCT TAA TAA GAA TGC ATC	46.3

^a T_M = melting temperature of DNA strand.

The DNA oligonucleotides were resuspended in water to a concentration of 100 μ M. The DNA was aliquoted and frozen until use. DNA solutions were made to the desired concentration for each experiment with 5 \times saline-sodium citrate (SSC) buffer/0.1% Tween-20. A stock 20 \times SSC solution consisting of 3 M NaCl, 300 mM trisodium citrate, with pH adjusted to 7 with HCl was used.

Preparation of devices

Heavily n-doped silicon substrates with a 230 nm SiO_2 dielectric and prefabricated gold source-drain electrodes from Fraunhofer IPMS ($W = 2000 \mu\text{m}$, $L = 20 \mu\text{m}$) were washed with acetone and dried with nitrogen. They were then treated with oxygen plasma for 15 minutes to clean and hydrolyze the surface. Substrates were then rinsed with water and isopropanol, before a 1 hour surface treatment in 1% v/v octyltrichlorosilane (OTS) in toluene at 70 °C. Silane-treated substrates were washed with toluene and isopropanol and dried for 1 hour at 70 °C under vacuum. CuPc and $\text{F}_{16}\text{-CuPc}$ were deposited using physical vapor deposition in an Angstrom EvoVac thermal evaporator with a target thickness of 150 Å and a rate of 0.3 Å s⁻¹ at 140 °C. Heated substrates were allowed to cool to room temperature before being removed from the vacuum chamber, usually overnight.

OTFT testing & electrical characterization

Contact with the source-drain electrodes was made with BeCu alloy probe tips. Output curves were obtained by fixing the gate voltage (V_{GS}) at discrete values and sweeping the source-drain voltage (V_{SD}). Electrical measurements were performed using a custom electrical probe station with a chamber allowing for controlled atmosphere, oesProbe A10000-P290 (Element Instrumentation Inc. & Kreus Design Inc.) with a Keithley 2614B to control source-drain voltage (V_{DS}), gate voltage (V_{GS}), and measure source-drain current (I_{DS}). V_{DS} was held constant while V_{GS} was varied to obtain measurements of I_{DS} . From these measurements, saturation region field-effect mobility, on/off current ratio, and threshold voltage were determined. The general expression relating current to field-effect mobility and gate voltage in the saturation mode is given in eqn (1):

$$I_{DS} = \frac{\mu C_i W}{2L} (V_{GS} - V_T)^2 \quad (1)$$

where I_{DS} is the source-drain voltage, μ is the field-effect mobility of the particular material, C_i is the capacitance density, W is the width of the channel, L is the length of the channel, V_{GS} is the gate-source voltage, and V_T is the threshold

voltage. To obtain a linear relation, the square root of eqn (1) is taken, giving eqn (2), so that the mobility and threshold voltage can be calculated directly from the slope and x -intercept of an $\sqrt{I_{DS}}$ vs. V_{GS} curve.

$$\sqrt{I_{DS}} = \sqrt{\frac{\mu C_i W}{2L}} (V_{GS} - V_T) \quad (2)$$

Finally, the on/off ratio is determined by eqn (3):

$$\text{On/off ratio} = \frac{I_{on}}{I_{off}} \quad (3)$$

where I_{on} and I_{off} are the highest and lowest currents, respectively, measured in the characterized gate voltage range.

DNA experiments

During testing of the devices used in DNA sensing experiments, the devices were operated at 51.1 °C (as this is the melting temperature ($T_M - 5$ °C) or 25 °C. The devices were tested with no analytes to establish a baseline at 51.1 °C, then either 2 μL of buffer, or one of the DNA solutions were pipetted directly onto the source/drain channel. Then, the droplets were left to evaporate (2 minutes), then the devices were rinsed with deionized water. They were then dried with nitrogen and placed under vacuum for 3 minutes. The device was then retested. DNA solutions were made by first heating the individual ssDNA to 95 °C for 15 seconds, then either mixing the probe and target strands (complementary) or the probe and control strands (non-complementary) in the desired concentration, and then pipetting onto the transistor surface.

AFM

A Park NX10 system was used in non contact mode with a PPP-NCH-20 tip, with a scan rate of 0.7 Hz and image size of 512 \times 512 pixels. The images were produced with the XEI software version 1.8.2.

Conclusions

Bottom gate bottom contact OTFT temperature and DNA sensors were fabricated using both CuPc or $\text{F}_{16}\text{-CuPc}$ as the P or N type semiconductor layer, respectively. Within a temperature range of 25 °C to 90 °C CuPc devices in air showed little change in V_T but significant and linear increases in mobility. Under the same conditions, $\text{F}_{16}\text{-CuPc}$ showed a linear and significant negative change in V_T , with an increase in mobility as well. Under vacuum, devices with both materials varied similarly with increasing temperature, exhibiting almost no change in V_T and an increase in mobility. Both CuPc and $\text{F}_{16}\text{-CuPc}$ devices responded differently when treated with ssDNA versus dsDNA. The negative charge originating from DNA effects shifted the V_T in the positive direction for both P and N type materials, with a greater shift observed for ssDNA compared to dsDNA. While similar observations have been reported for P-type semiconductors, detection using an N-type organic semiconductor is unprecedented, more sensitive, and further



supports one of the proposed mechanisms for dsDNA detection by OTFTs. Hybrid detection could be possible by examining the changes in V_T for each material and future efforts will focus on amplification of signal response through device engineering and material selection.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 J. Liu, M. Agarwal and K. Varahramyan, Glucose sensor based on organic thin film transistor using glucose oxidase and conducting polymer, *Sens. Actuators, B*, 2008, **135**, 195–199.
- 2 Q. Zhang and V. Subramanian, DNA hybridization detection with organic thin film transistors: Toward fast and disposable DNA microarray chips, *Biosens. Bioelectron.*, 2007, **22**, 3182–3187.
- 3 M. L. Hammock, O. Knopfmacher, B. D. Naab, J. B.-H. Tok and Z. Bao, Investigation of Protein Detection Parameters Using Nanofunctionalized Organic Field-Effect Transistors, *ACS Nano*, 2013, **7**, 3970–3980.
- 4 H. U. Khan, J. Jang, J.-J. Kim and W. Knoll, Effect of passivation on the sensitivity and stability of pentacene transistor sensors in aqueous media, *Biosens. Bioelectron.*, 2011, **26**, 4217–4221.
- 5 W. Huang, *et al.*, Label-free brain injury biomarker detection based on highly sensitive large area organic thin film transistor with hybrid coupling layer, *Chem. Sci.*, 2014, **5**, 416–426.
- 6 S. R. Forrest, The path to ubiquitous and low-cost organic electronic appliances on plastic, *Nature*, 2004, **428**, 911–918.
- 7 C. Reese, M. Roberts, M. Ling and Z. Bao, Organic thin film transistors, *Mater. Today*, 2004, **7**, 20–27.
- 8 L. Basiricò, *et al.*, Electrical characteristics of ink-jet printed, all-polymer electrochemical transistors, *Org. Electron.*, 2012, **13**, 244–248.
- 9 W. J. Ansorge, Next-generation DNA sequencing techniques, *N. Biotechnol.*, 2009, **25**, 195–203.
- 10 J. M. Rothberg, *et al.*, An integrated semiconductor device enabling non-optical genome sequencing, *Nature*, 2011, **475**, 348–352.
- 11 M. Jain, *et al.*, Nanopore sequencing and assembly of a human genome with ultra-long reads, *Nat. Biotechnol.*, 2018, **36**, 338–345.
- 12 B. Y. C. Ng, E. J. H. Wee, N. P. West and M. Trau, Rapid DNA detection of *Mycobacterium tuberculosis*-towards single cell sensitivity in point-of-care diagnosis, *Sci. Rep.*, 2015, **5**, 15027.
- 13 A. Niemz, T. M. Ferguson and D. S. Boyle, Point-of-care nucleic acid testing for infectious diseases, *Trends Biotechnol.*, 2011, **29**, 240–250.
- 14 G. Schwartz, *et al.*, Flexible polymer transistors with high pressure sensitivity for application in electronic skin and health monitoring, *Nat. Commun.*, 2013, **4**, 1859.
- 15 S. Takao, D. Ananth, G. Alan, E. K. Howard and Z. Bao, Integration and Response of Organic Electronics with Aqueous Microfluidics, *Langmuir*, 2002, **18**, 5299–5302.
- 16 H. Gui, B. Wei and J. Wang, High sensitivity and air stability in an organic transistor-based biosensor by inserting a CuPc layer, *Phys. Status Solidi A*, 2014, **211**, 2499–2502.
- 17 N. Liu, *et al.*, A label-free, organic transistor-based biosensor by introducing electric bias during DNA immobilization, *Org. Electron.*, 2012, **13**, 2781–2785.
- 18 X. Chen, H. Gui, B. Wei and J. Wang, A label-free biosensor based on organic transistors by using the interaction of mercapto DNA and gold electrodes, *Mater. Sci. Semicond. Process.*, 2015, **35**, 127–131.
- 19 M. Bixon, *et al.*, Long-range charge hopping in DNA, *Proc. Natl. Acad. Sci. U. S. A.*, 1999, **96**, 11713–11716.
- 20 D. Porath, A. Bezryadin, S. de Vries and C. Dekker, Direct measurement of electrical transport through DNA molecules, *Nature*, 2000, **403**, 635–638.
- 21 J.-M. Kim, *et al.*, A flexible pentacene thin film transistors as disposable DNA hybridization sensor, *J. Ind. Eng. Chem.*, 2012, **18**, 1642–1646.
- 22 M. Demelas, *et al.*, An organic, charge-modulated field effect transistor for DNA detection, *Sens. Actuators, B*, 2012, **171–172**, 198–203.
- 23 S. P. White, K. D. Dorfman and C. D. Frisbie, Label-Free DNA Sensing Platform with Low-Voltage Electrolyte-Gated Transistors, *Anal. Chem.*, 2015, **87**, 1861–1866.
- 24 H. U. Khan, M. E. Roberts, O. Johnson, W. Knoll and Z. Bao, The effect of pH and DNA concentration on organic thin-film transistor biosensors, *Org. Electron.*, 2012, **13**, 519–524.
- 25 J. Meinkoth and G. Wahl, Hybridization of nucleic acids immobilized on solid supports, *Anal. Biochem.*, 1984, **138**, 267–284.
- 26 G. Steger, Thermal denaturation of double-stranded nucleic acids: prediction of temperatures critical for gradient gel electrophoresis and polymerase chain reaction, *Nucleic Acids Res.*, 1994, **22**, 2760–2768.
- 27 P. Sykacek, *et al.*, The impact of quantitative optimization of hybridization conditions on gene expression analysis, *BMC Bioinf.*, 2011, **12**, 73.
- 28 H. Gui, B. Wei and J. Wang, The hybridization and optimization of complementary DNA molecules on organic field-effect transistors, *Mater. Sci. Semicond. Process.*, 2014, **30**, 250–254.
- 29 Y.-J. Lin and C.-C. Hung, Temperature-dependent hole transport for pentacene thin-film transistors with a SiO₂



gate dielectric modified by $(\text{NH}_4)_2\text{S}_x$ treatment, *Microelectron. Reliab.*, 2018, **81**, 90–94.

30 Q.-J. Sun, X. Gao and S.-D. Wang, Understanding temperature dependence of threshold voltage in pentacene thin film transistors, *J. Appl. Phys.*, 2013, **113**, 194506.

31 S. Bixi, O. A. Melville, N. T. Boileau and B. H. Lessard, The influence of air and temperature on the performance of PBDB-T and P3HT in organic thin film transistors, *J. Mater. Chem. C*, 2018, **6**, 11972–11979.

32 R. J. Chesterfield, *et al.*, Variable temperature film and contact resistance measurements on operating *n*-channel organic thin film transistors, *J. Appl. Phys.*, 2004, **95**, 6396–6405.

33 X. Ren, P. K. L. Chan, J. Lu, B. Huang and D. C. W. Leung, High Dynamic Range Organic Temperature Sensor, *Adv. Mater.*, 2013, **25**, 1291–1295.

34 S. Jung, T. Ji and V. K. Varadan, Point-of-care temperature and respiration monitoring sensors for smart fabric applications, *Smart Mater. Struct.*, 2006, **15**, 1872–1876.

35 C. Servadio and Z. Leib, Hyperthermia in the treatment of prostate cancer, *Prostate*, 2007, **5**, 205–211.

36 T. Uray, *et al.*, Surface Cooling for Rapid Induction of Mild Hypothermia After Cardiac Arrest: Design Determines Efficacy, *Acad. Emerg. Med.*, 2010, **17**, 360–367.

37 T. Sugiura, *et al.*, Photothermal therapy of tumors in lymph nodes using gold nanorods and near-infrared laser light with controlled surface cooling, *Nano Res.*, 2015, **8**, 3842–3852.

38 H. Takeda, S. Maruyama, J. Okajima, S. Aiba and A. Komiya, Development and estimation of a novel cryoprobe utilizing the Peltier effect for precise and safe cryosurgery, *Cryobiology*, 2009, **59**, 275–284.

39 V. Bernard, E. Staffa, V. Mornstein and A. Bourek, Infrared camera assessment of skin surface temperature – effect of emissivity, *Phys. Medica*, 2013, **29**, 583–591.

40 D. Kawakami, Y. Yasutake, H. Nishizawa and Y. Majima, Bias Stress Induced Threshold Voltage Shift in Pentacene Thin-Film Transistors, *Jpn. J. Appl. Phys.*, 2006, **45**, L1127–L1129.

41 H.-K. Chen, P.-T. Liu, T.-C. Chang and S.-L. Shy, Variable Temperature Measurement on Operating Pentacene-Based OTFT, *MRS Proc.*, 2008, **1091**, 1091-AA07–91.

42 O. A. Melville, B. H. Lessard and T. P. Bender, Phthalocyanine-Based Organic Thin-Film Transistors: A Review of Recent Advances, *ACS Appl. Mater. Interfaces*, 2015, **7**, 13105–13118.

43 I. G. Hill and A. Kahn, Energy level alignment at interfaces of organic semiconductor heterostructures, *Appl. Phys. Lett.*, 1998, **84**, 913.

44 T. Chassé, C.-I. Wu, I. G. Hill and A. Kahn, Band alignment at organic-inorganic semiconductor interfaces: α -NPD and CuPc on InP(110), *J. Appl. Phys.*, 1999, **85**, 6589.

45 W. Chen, D.-C. Qi, H. Huang, X. Gao and A. T. S. Wee, Organic-Organic Heterojunction Interfaces: Effect of Molecular Orientation, *Adv. Funct. Mater.*, 2011, **21**, 410–424.

46 N. A. Rice, F. Magnan, O. Melville, J. L. Brusso and B. H. Lessard, Organic Thin Film Transistors Incorporating Solution Processable Thieno[3,2-b]thiophene Thienoacenes, *Materials*, 2017, **11**, 8.

47 P. Stallinga and H. L. Gomes, Thin-film field-effect transistors: The effects of traps on the bias and temperature dependence of field-effect mobility, including the Meyer–Neldel rule, *Org. Electron.*, 2006, **7**, 592–599.

48 D. Kumaki, T. Umeda and S. Tokito, Influence of and on threshold voltage shift in organic thin-film transistors: Deprotonation of SiOH on gate-insulator surface, *Appl. Phys. Lett.*, 2008, **92**, 93309.

49 Q. Zhang, L. Jagannathan and V. Subramanian, Label-free low-cost disposable DNA hybridization detection systems using organic TFTs, *Biosens. Bioelectron.*, 2010, **25**, 972–977.

50 J. M. Kim, S. K. Jha, R. Chand, D. H. Lee and Y. S. Kim, DNA hybridization sensor based on pentacene thin film transistor, *Biosens. Bioelectron.*, 2011, **26**, 2264–2269.

51 H. Sirringhaus, Reliability of Organic Field-Effect Transistors, *Adv. Mater.*, 2009, **21**, 3859–3873.

52 A. N. Rao and D. W. Grainger, Biophysical properties of nucleic acids at surfaces relevant to microarray performance, *Biomater. Sci.*, 2014, **2**, 436–471.

53 Z.-J. Tan and S.-J. Chen, Nucleic acid helix stability: effects of salt concentration, cation valence and size, and chain length, *Biophys. J.*, 2006, **90**, 1175–1190.

