



and  $\beta$ -hydroxypyruvate to 2-amino-1,3,4-butanetriol (ABT),<sup>9b</sup> and the conversion of benzaldehyde and pyruvate to nor(pseudo)ephedrine N(P)E.<sup>9c</sup> However, to date, no reports have appeared in the literature detailing the asymmetric ring opening of racemic epoxides for the enantioselective synthesis of chiral  $\beta$ -amino alcohols *via* enzymatic catalysis, probably as a consequence of the difficulties in finding robust enzymes with excellent activity and enantioselectivity.<sup>10</sup> Besides that, how to solve the problem of cofactor regeneration in some cascade biocatalysis reactions is still a challenge.

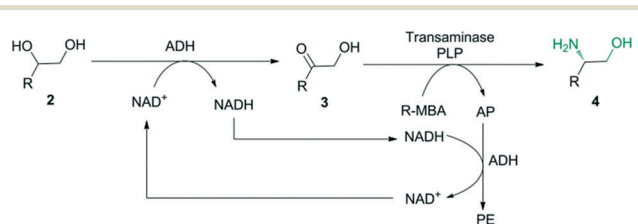
In this study, we envisioned a one-pot three-step enzymatic cascade reaction for the asymmetric ring opening of racemic epoxides to enantiopure  $\beta$ -amino alcohols (Scheme 1). The process includes the complete hydrolysis of a racemic epoxide to a vicinal diol by an epoxide hydrolase (EH), the oxidation of the vicinal diol intermediate to an  $\alpha$ -hydroxy ketone by two stereocomplementary alcohol dehydrogenases (ADH) and the asymmetric reductive amination of an  $\alpha$ -hydroxy ketone intermediate to an enantiopure  $\beta$ -amino alcohol by a transaminase (TA). ADH/(*R*)-(+)-1-phenylethylamine (*R*-MBA) was also utilized in the cascade system to drive the equilibrium towards the product and recycle the  $\text{NAD}^+$  cofactor.

In order to realize the novel cascade biocatalysis system, the transaminases for the conversion of  $\alpha$ -hydroxy ketones to enantiopure  $\beta$ -amino alcohols in the third step of the cascade biocatalysis system were selected. Based on our previously constructed high-throughput method for the screening of highly enantioselective amino alcohol specific transaminases,<sup>11</sup> (*R*)- $\omega$ -transaminase (MVTA) from *Mycobacterium vanbaalenii*<sup>12</sup> was identified as a robust enzyme with high activity and excellent enantioselectivity toward  $\beta$ -amino alcohols, and the enzyme was utilized to synthesise chiral  $\beta$ -amino alcohols from the corresponding  $\alpha$ -hydroxy ketones 3a–f for the first time. After 2 to 3 h of reaction time, the conversion reached 99% with the product in 97–99% ee (see the ESI,† Table S1). Due to (*R*)-MBA being used as an efficient amine donor of MVTA, the co-product acetophenone (AP) was produced in the asymmetric reductive amination process. In order to recycle the  $\text{NAD}^+$  in the second step of the cascade biocatalysis system, we designed an *in situ* self-sufficient cofactor regeneration system (Scheme 1) that involves the alcohol dehydrogenase catalyzed oxidation of vicinal diols and the same alcohol dehydrogenase catalyzed AP reduction. Then, the alcohol dehydrogenase (ADH)/(*R*)-MBA in the cascade system can be utilized to recycle the  $\text{NAD}^+$  cofactor and simultaneously drive the equilibrium towards the product. In order to realize this process, sixteen alcohol dehydrogenases were tested for diol 2c oxidation activity and AP reduction activity (see the ESI,† Table S2). BDHA from *Bacillus subtilis* and GoSCR from *Gluconobacter oxydans*<sup>13</sup> showed the highest activity toward the diols (*R*)-2c (0.80  $\text{U mg}^{-1}$ ) and (*S*)-2c (0.12  $\text{U mg}^{-1}$ ) and moderate activity toward AP (0.428  $\text{U mg}^{-1}$  for BDHA and 1.257  $\text{U mg}^{-1}$  for GoSCR), respectively. Therefore, BDHA and GoSCR were selected as promising candidates to catalyze the diol oxidation step of the cascade reaction. Substrate specific analysis revealed that BDHA and GoSCR have

high activity and excellent enantioselectivity toward vicinal diol 2a–f, with an activity from 0.133 to 0.791  $\text{U mg}^{-1}$  protein for BDHA toward (*R*)-2a–f, and 0.067 to 0.119  $\text{U mg}^{-1}$  protein for GoSCR toward (*S*)-2a–f (see the ESI,† Table S3). For the first hydrolysis step, the epoxide hydrolase (SpeH) from *Sphingomonas* sp. HXN-200 was selected as a promising enzyme due to its remarkable activity in the enantioselective hydrolysis of epoxides.<sup>14</sup> The substrate specific and enantioselectivity of SpeH was further analyzed with six racemic epoxides 1a–f, with 20 mM of substrate. After reaction for 2 to 3 h, the substrates were completely converted to vicinal diols 2 (99% conversion), forming the product diol 2 in 3–40% ee (see the ESI,† Table S4). As the intermediate product 2 was formed in the first step of the cascade reaction with low ee values, it was then essential to employ two stereocomplementary alcohol dehydrogenases (2,3-butanediol dehydrogenase, BDHA, and polyol dehydrogenase, GoSCR) in the second step of the cascade biocatalysis system to ensure the complete conversion of both enantiomers of the vicinal diol intermediate.

After SpeH, BDHA, GoSCR and MVTA were selected as the best candidates for the new cascade biocatalysis system. The combination of the last two steps into a simultaneous one-pot in the cascade reaction was first tested with BDHA, GoSCR and MVTA (Scheme 2). The reaction conditions for the conversion of diols to chiral  $\beta$ -amino alcohols were optimized using 2c as a model substrate, and the highest conversion of 4c was obtained at pH 7.4 and 30 °C, with 0.5 mM of  $\text{NAD}^+$  added (see the ESI,† Tables S5–S7). The adjustment of the two enzyme concentrations at a ratio of 2 : 1 (20  $\text{mg mL}^{-1}$  of ADH and 10  $\text{mg mL}^{-1}$  of MVTA) resulted in the highest conversion of 4c (see the ESI,† Table S8). Under the optimized conditions, the enantioselective conversion of racemic diols 2c (50 mM) with the mixture of BDHA, GoSCR and MVTA (2 : 2 : 1) reached up to 99% with the product 4c in >99% ee. The generality of this cascade system (BDHA/GoSCR/MVTA) was further investigated using five other racemic diols (2a–b and 2d–f) and 31–99% conversions were obtained in 97–99% ee with a mixture of BDHA, GoSCR and MVTA (2 : 2 : 1) (see the ESI,† Table S9).

We next sought to combine the SpeH, BDHA, GoSCR and MVTA biocatalysts in a one-pot cascade to convert 1 to 4. Racemic 1c was exposed to SpeH followed by BDHA/GoSCR and MVTA, and the target (*S*)-4c was obtained in 86.5%–99% conversion and >99% ee (Table 1, entries 3–4). The generality of



**Scheme 2** Enantioselective conversion of racemic vicinal diol 2 to enantiopure  $\beta$ -amino alcohol 4 using a mixture of BDHA, GoSCR and MVTA.

**Table 1** Asymmetric ring opening of racemic epoxides **1** to (*S*)- $\beta$ -amino alcohol **4** with mixtures of cell-free extracts of *E. coli* (SpEH), *E. coli* (BDHA), *E. coli* (GoSCR) and *E. coli* (MVTA)

Entry	Sub.	Sub. conc [mM]	SpEH [mg mL <sup>-1</sup> ]	BDHA [mg mL <sup>-1</sup> ]	GoSCR [mg mL <sup>-1</sup> ]	MVTA [mg mL <sup>-1</sup> ]	Time [h]	Conv. to <b>4</b> <sup>a</sup> [%]	( <i>S</i> )- <b>4</b> ee <sup>b</sup> [%]
1	<b>1a</b>	20	10	20	20	10	24	47.3	97
2	<b>1b</b>	20	10	20	20	10	24	61.2	98
3	<b>1c</b>	20	10	20	20	10	16	99.0	>99
4	<b>1c</b>	30	20	40	40	20	16	86.5	>99
5	<b>1d</b>	20	10	20	20	10	24	96.9	>99
6	<b>1e</b>	20	10	20	20	10	24	70.5	>99
7	<b>1f</b>	20	10	20	20	10	24	34.0	>99

The reactions were conducted using 25–55 mM of the amine donor (*R*-MBA) in 100 mM sodium phosphate buffer (pH 7.5) containing 0.1 mM pyridoxal-5'-phosphate (PLP), 0.5 mM NAD<sup>+</sup>, 20–50 mM substrates, and 10% DMSO at 30 °C. <sup>a</sup> Conversion was determined by GC, error limit: <2% of the stated values. <sup>b</sup> ee values were determined by chiral GC.

**Table 2** Asymmetric ring opening of racemic epoxides **1** to (*S*)- $\beta$ -amino alcohol **4** with resting cells of *E. coli* (SpEH-BDHA-GoSCR-MVTA) (catalyst **3**) in an aqueous–organic two-phase system

Sub.	Sub. conc. [mM]	<i>R</i> (+)-MBA [mM]	Catalyst <b>3</b> [g cdw L <sup>-1</sup> ]	<i>T</i> [h]	Conv. to <b>4</b> <sup>a</sup> [%]	( <i>S</i> )- <b>4</b> ee <sup>b</sup> [%]
<b>1a</b>	20	25	18	5	99.0	97
<b>1b</b>	20	25	18	5	80.4	98
<b>1c</b>	50	55	20	16	92.0	>99
<b>1d</b>	20	25	18	5	99.0	>99
<b>1e</b>	20	25	18	5	97.1	>99
<b>1f</b>	20	25	18	5	79.6	>99

The reactions were conducted using 25–55 mM of the amine donor (*R*-MBA) in a two-liquid phase system consisting of sodium phosphate buffer (100 mM, pH 7.5) and hexadecane (1:1). The reactions were carried out using 0.1 mM PLP and 10–50 mM of the substrates at 30 °C. <sup>a</sup> Conversion was determined by GC, error limit: <2% of the stated values. <sup>b</sup> ee values were determined by chiral GC.

the cascade biocatalysis system was investigated by examining five other racemic epoxides (**1a–b** and **1d–f**) using SpEH, BDHA/GoSCR and MVTA (1:2:2:1) in the presence of 0.5 mM of NAD<sup>+</sup>, and the (*S*)- $\beta$ -amino alcohols **4a–b** were obtained in 97–99% ee and 47.3%–61.2% conversion from the racemic epoxides **1a–b** (20 mM) after 24 h of reaction time (Table 1, entries 1–2). For the halogen *para* substituted styrene oxide substrates **1d–f**, good to excellent conversions (34%–96.9%) of the (*S*)- $\beta$ -amino alcohols **4d–f** were obtained in >99% ee (Table 1, entries 4–6).

Compared to enzymatic catalysis, whole-cell biocatalysts provide unique properties,<sup>15</sup> such as that the intact cells can

protect the enzyme from rapid deactivation, and the cofactor NAD<sup>+</sup>/NADH in cells can be used for targeted reactions with co-factor recycling. Mixed resting cells of *E. coli* (SpEH-MVTA) and *E. coli* (BDHA-GoSCR) (Fig. S1†) were first examined for the conversion of epoxides **1a–f** to (*S*)- $\beta$ -amino alcohols **4a–f**. Combining *E. coli* (SpEH-MVTA) cells at 10 g cdw L<sup>-1</sup> and *E. coli* (BDHA-GoSCR) cells at 16 g cdw L<sup>-1</sup> led to the production of (*S*)-**4a–c** in 97–99% ee with 42.5%–68.1% conversion from 10 mM of **1a** and **1b** and 20 mM of **1c**, respectively. For the conversion of 20 mM of the epoxides **1d–f**, resting cells of *E. coli* (SpEH-MVTA) and *E. coli* (BDHA-GoSCR) were mixed in a 14:22 ratio, and after 24 h of reaction time, the products (*S*)-**4d–f** were obtained in >99% ee and 61.8–77.7% conversion (see the ESI,† Table S10).

A recombinant *E. coli* strain that co-expresses all of the necessary enzymes is often used for cascade biocatalysis since this avoids having to cultivate cells of multiple strains.<sup>16</sup> Resting cells of *E. coli* (SpEH-BDHA-GoSCR-MVTA) co-expressing four enzymes were thus used for the one-pot conversion of the epoxides **1a–f** to the (*S*)-amino alcohols **4a–f**. The *E. coli* (SpEH-BDHA-GoSCR-MVTA) cells showed specific activities of 14.8–40.6 U g<sup>-1</sup> cdw for the conversion of **1c** to **4c** after growth for 2–16 h (see the ESI,† Fig. S3). The SDS-PAGE of the cell-free extracts of the *E. coli* cells harvested at 4–16 h is shown in Fig. S2.† Reactions of 10–20 mM of the racemic epoxide **1c** with *E. coli* (SpEH-BDHA-GoSCR-MVTA) at 10–20 g cdw L<sup>-1</sup> afforded (*S*)-**4c** in >99% ee and 99% conversion. When the concentration of the racemic epoxide **1c**



**Fig. 1** Time course of the conversion of styrene oxide **1c** (50 mM) to (*S*)-**4c** with resting cells of *E. coli* (SpEH-BDHA-GoSCR-MVTA) (20 g cdw L<sup>-1</sup>) in a two-liquid phase system. ♦ = the concentration of **1c**; ■ = the concentration of **2c**; × = the concentration of **3c**; and ▲ = the concentration of (*S*)-**4c**.

was increased to 50 mM, only a trace amount of the desired product (*S*)-**4c** was formed (5.6% conversion) (see the ESI,† Table S11). We attribute this to the low solubility and high toxicity of the epoxide substrate in aqueous phase. To circumvent the solubility and toxicity problem of the epoxides, an aqueous–organic two-phase system consisting of sodium phosphate buffer (100 mM, pH 7.5) and hexadecane at 1:1 (v/v) was used as the reaction medium. Reactions of 20 mM of epoxides **1a** and **1b** with *E. coli* (SpeH–BDHA–GoSCR–MVTA) at 18 g cdw L<sup>−1</sup> afforded (*S*)-**4a** and (*S*)-**4b** in 97%–98% ee and 80.4–99% conversion within 5 h of reaction time. The reaction of epoxide **1c** (50 mM) with the same catalysts at 20 g cdw L<sup>−1</sup> gave (*S*)-**4c** in >99% ee and 92% conversion (Table 2). The time course of this reaction is shown in Fig. 1. At 2 h, epoxide **1c** was totally converted to 1-phenyl-1, 2-ethanediol **2c** due to the high activity of SpeH, 33.5 mM diol **2c** remained in the reaction mixture, and the remainder of diol **2c** was converted to 15.5 mM of (*S*)-2-amino-2-phenylethanol **4c** in >99% ee by BDHA/GoSCR and MVTA. From 2 to 16 h, the concentration of (*S*)-**4c** increased linearly upon a linear decrease in the concentration of **2c**. At 16 h, 46 mM of (*S*)-**4c** was produced in >99% ee, and a trace amount of **2c** remained in the reaction mixture. In the whole reaction process, almost no 2-hydroxyacetophenone **3c** was detected. These results show that the  $\alpha$ -hydroxy ketone produced in the second step was quickly converted to an  $\beta$ -amino alcohol by MVTA in the third step of the cascade reaction. Further conversion of epoxides **1d–f** (20 mM) with *E. coli* (SpeH–BDHA–GoSCR–MVTA) at 18 g cdw L<sup>−1</sup> gave (*S*)-**4d–f** in >99% ee and 79.6–99% conversion.

The applicability of this new type of cascade biocatalysis was tested in a 100 mL scale preparative experiment. After 24 h of reaction times, pure (*S*)-**4c** was isolated in >99% ee and 80% isolated yield after extraction with ethyl acetate and flash chromatography (see the ESI,† Fig. S17 and S18). This example demonstrates the facile and green application of this new type of cascade biocatalysis for the asymmetric ring opening of epoxides to enantiopure  $\beta$ -amino alcohols in high-yield.

In summary, a new type of cascade biocatalysis reaction was successfully developed for the asymmetric ring opening of racemic epoxides to useful and valuable chiral  $\beta$ -amino alcohols in high ee via epoxide hydrolysis, diol oxidation and  $\alpha$ -hydroxy ketone asymmetric reductive amination. *In vitro* cascade biocatalysis with mixtures of recombinant SpeH, BDHA, GoSCR and MVTA was conducted in the presence of a trace amount of NAD<sup>+</sup> to convert the epoxides **1a–f** to the corresponding (*S*)- $\beta$ -amino alcohols **4a–f** in 97–99% ee and 34–99% conversion. The alcohol dehydrogenase (ADH)/(*R*)-(+)-1-phenylethylamine (*R*-MBA) in the cascade biocatalysis system was utilized to drive the equilibrium towards the product and recycle the NAD<sup>+</sup> cofactor. Whole-cell based cascade biocatalysis was also achieved using the engineered recombinant *E. coli* cells without any additional NAD<sup>+</sup> cofactor, and the produced (*S*)- $\beta$ -amino alcohols were obtained in good to excellent conversion (79–99%) and 97–99% ee. Preparative biotransformation was also demonstrated on a 100 mL scale with cells of *E. coli* (SpeH–BDHA–GoSCR–MVTA). The

designed cascade process might be generally applicable to access a variety of different chiral  $\beta$ -amino alcohols starting from the corresponding inexpensive racemic epoxides by combining appropriate enzymes.

This study was supported by the National Natural Science Foundation of China (Grant No. 21772141), the Shanxi Province Science Foundation for Youths (grant No. 201701D221042) and the Open Funding Project of the State Key Laboratory of Bioreactor Engineering.

## Conflicts of interest

There are no conflicts to declare.

## Notes and references

- (a) R. E. Lee, M. Protopopova, E. Crooks, R. A. Slayden, M. Terrot and C. E. Barry III, *J. Comb. Chem.*, 2003, **5**, 172–187; (b) A. V. Malkov, M. A. Kabeshov, M. Bella, O. Kysilka, D. A. Malyshev, K. Pluháčková and P. Kočovský, *Org. Lett.*, 2007, **9**, 5473–5476; (c) J. Guo, M. Zhu, T. Wu, C. Hao, K. Wang, Z. Yan, W. Huang, J. Wang, D. Zhao and M. Cheng, *Bioorg. Med. Chem.*, 2017, **25**, 3500–3511; (d) Y. Hamada, T. Nakanishi, K. Suzuki, R. Yamaguchi, T. Hamada, K. Hidaka, S. Ishiura and Y. Kiso, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 4640–4644; (e) X. Cattoën and M. A. Pericàs, *Tetrahedron*, 2009, **65**, 8199–8205.
- (a) K. B. Sharpless, D. W. Patrick, L. K. Truesdale and S. A. Biller, A new reaction, *J. Am. Chem. Soc.*, 1975, **97**, 2305–2307; (b) G. Li, H. Chang and K. B. Sharpless, *Angew. Chem., Int. Ed. Engl.*, 1996, **35**, 451–454; (c) J. A. Birrell and E. N. Jacobsen, *Org. Lett.*, 2013, **15**, 2895–2897; (d) C. X. Ye, Y. Y. Melcamu, H. H. Li, J. T. Cheng, T. T. Zhang, Y. P. Ruan, X. Zheng, X. Lu and P. Q. Huang, *Nat. Commun.*, 2018, **9**, 410.
- (a) E. J. Jacobsen, *Acc. Chem. Res.*, 2000, **33**, 421–431; (b) C. Wang, L. Luo and H. Yamamoto, *Acc. Chem. Res.*, 2016, **49**, 193–204.
- (a) A. K. Chakraborti, S. Rudrawar and A. Kondaskar, *Org. Biomol. Chem.*, 2004, **2**, 1277–1280; (b) S. Regati, Y. He, M. Thimmaiah, P. Li, S. Xiang, B. Chen and J. C. G. Zhao, *Chem. Commun.*, 2013, **49**, 9836–9838; (c) M. Kumar, R. I. Kureshy, S. Saravanan, S. Verma, A. Jakhar, N. H. Khan, S. H. R. Abdi and H. C. Bajaj, *Org. Lett.*, 2014, **16**, 2798–2801; (d) Z. Sun, J. Chen, Y. Liu and T. Tu, *Adv. Synth. Catal.*, 2017, **359**, 494–505.
- A. Rouf, G. P. Upta, M. A. Aga, B. Kumar, R. Parshad and S. C. Taneja, *Tetrahedron: Asymmetry*, 2011, **22**, 2134–2143.
- (a) M. S. Malik, E. S. Park and J. S. Shin, *Green Chem.*, 2012, **14**, 2137–2140; (b) H. L. Wu, J. D. Zhang, C. F. Zhang, X. J. Fan, H. H. Chang and W. L. Wei, *Appl. Biochem. Biotechnol.*, 2017, **181**, 972–985; (c) A. Nobili, F. Steffen-Munsberg, H. Kohls, I. Trentin, C. Schulzke, M. Höhne and U. T. Bornscheuer, *ChemCatChem*, 2015, **7**, 757–760.
- R. Patel, A. Banerjee, J. Howell, C. McNamee, D. Brozowski, D. Mirfakhrae, V. Nanduri, J. Thottathil and L. Szarka, *Tetrahedron: Asymmetry*, 1993, **4**, 2069–2084.



- 8 (a) J. M. Sperl and V. Sieber, *ACS Catal.*, 2018, **8**, 2385–2396; (b) J. D. Zhang, S. Wu, J. Wu and Z. Li, *ACS Catal.*, 2015, **5**, 51–58; (c) S. Gandomkar, A. Dennig, A. Dordic, L. Hammerer, M. Pickl, T. Haas, M. Hall and K. Faber, *Angew. Chem., Int. Ed.*, 2018, **57**, 427–430.
- 9 (a) P. Gupta and N. Mahajan, *New J. Chem.*, 2018, **42**, 12296–12327; (b) L. Rios-Solis, P. Morris, C. Grant, A. O. O. Odeleye, H. C. Hailes, J. M. Ward, P. A. Dalby, F. Baganz and G. J. Lye, *Chem. Eng. Sci.*, 2015, **122**, 360–372; (c) T. Sehl, H. C. Hailes, J. M. Ward, R. Wardenga, E. von Lieres, H. Offermann, R. Westphal, M. Pohl and D. Rother, *Angew. Chem., Int. Ed.*, 2013, **52**, 6772–6775.
- 10 I. V. Pavlidis, M. S. Weiß, M. Genz, P. Spurr, S. P. Hanlon, B. Wirz, H. Iding and U. T. Bornscheuer, *Nat. Chem.*, 2016, **8**, 1076–1082.
- 11 J. D. Zhang, H. L. Wu, T. Meng, C. F. Zhang, X. J. Fan, H. H. Chang and W. L. Wei, *Anal. Biochem.*, 2017, **518**, 94–101.
- 12 M. Höhne, S. Schätzle, H. Jochens, K. Robins and U. T. Bornscheuer, *Nat. Chem. Biol.*, 2010, **6**, 807–813.
- 13 Z. M. Cui, J. D. Zhang, X. J. Fan, G. W. Zheng, H. H. Chang and W. L. Wei, *J. Biotechnol.*, 2017, **243**, 1–9.
- 14 S. Wu, A. Li, Y. S. Chin and Z. Li, *ACS Catal.*, 2013, **3**, 752–759.
- 15 (a) F. Garzón-Posse, L. Becerra-Figueroa, J. Hernández-Arias and D. Gamba-Sánchez, *Molecules*, 2018, **23**, 1265; (b) C. C. C. R. de Carvalho, *Microb. Biotechnol.*, 2017, **10**, 250–263.
- 16 (a) Y. Zhou, S. Wu and Z. Li, *Angew. Chem., Int. Ed.*, 2016, **55**, 11647–11650; (b) H. Chen, R. Huang and Y.-H. P. Zhang, *Appl. Microbiol. Biotechnol.*, 2017, **101**, 4481–4493.