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pH-responsive and antibacterial PANI-PEDOT:PSS fibres for wearable applications†

Rachel E. Smith,[a] Stella Totti,[a] Daniel Reid,[a] Suzanne M. Hingley-Wilson,[b] Eirini Velliou,[a] Paola Campagnolo,[b] Neil I. Ward,[a] John R. Varcoe[a] and Carol Crean *[a]

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With the emergence of wearable electronics for health and the prevalence of chronic wounds, there is demand for an effective wearable pH sensor. A simple method of solvent treating wet-spun poly(3,4-ethylenedioxythiophene)-poly(styrenesulfonate) (PEDOT:PSS) fibres proved effective in the development of highly conductive fibre substrates. With an optimised polyaniline coating, these fibres displayed a Nernstian response to changes in pH in the solid-state *vs.* a fabricated pseudoreference electrode across a wide pH range of 3.0 to 9.0. In addition, these biocompatible fibre electrodes displayed antibacterial properties. The fibre electrodes offer significant opportunities for future wearable textile-based sensors as well as the realisation of a real-time, on-body pH sensing device for monitoring skin and wound healing.

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1. Introduction

Biochemical reactions in the body are hugely influenced by pH, therefore wearable pH sensors could have a pivotal role to play in healthcare and personalised medicine. The information provided could help to track changes in health status, diagnose diseases, and manage treatment. Electrochemical pH sensors are beneficial due to the ease of miniaturization of the associated electronics.[1] This is best exemplified by wearable electrochemical blood glucose meters, which successfully facilitate diabetes management. Wearable pH sensors can be used to monitor health conditions such as skin disease, and for environmental hazard monitoring, in addition to measuring sports performance.[1] In healthcare, wound care is a significant burden, with chronic wounds causing an estimated annual cost of £5.3 billion in the UK alone.[2] Chronic wounds have a maximum of pH 9.0 and healthy, unwounded skin has a minimum of pH 4.0, (with sweat ranging from 4.5 to 7.0), therefore the pH range needed for analysis is pH 4.0 to 9.0.[3,4]

Current examples of pH sensors have primarily focused on materials such as hydrogen ionophores, metal oxides and polyaniline.[5] These materials were used to facilitate wearable sensing, with small form and flexibility allowing easier integration into devices in contrast to the traditional glass pH electrode. Hydrogen ionophores in a solid-state ion selective electrode have been used for pH sensing in a wearable format by Andrade *et al.* and Niu *et al.* Andrade *et al.* coated a cotton thread with carbon nanotubes followed by a hydrogen ion sensing membrane, which was flexible with an excellent pH response.[6] Niu's group used a similar strategy with a stretchable gold fibre electrode as the solid-state electrode, which had minor pH variation (−59.2 to −57.4 mV pH^{-1}) upon stretching between 0 and 200% strain.[7]

The development of micro–nano structures in metal oxides has enabled flexible and mechanically robust electrodes with good electrochemical response to pH. Iridium oxide (IrO_x) exhibits a wide range pH response and is the most commonly reported in flexible sensing applications, however, is expensive.[5] WO_3–based potentiometric pH sensors are an attractive alternative but need further research to address the sub-Nernstian behaviour and slow response time.[5]

Polyaniline (PANI) lends itself to wearable pH analysis because of its pH-sensitivity, good electrical conductivity and biocompatibility.[8] Guinovart *et al.* reported a screen–printed PANI electrode on a bandage that could detect pH of a wound with a sensitivity of −58.5 mV pH^{-1}, over the range of pH 5.5–8.0 with an RSD of 1.2%.[9] Wang *et al.* recently reported a stretchable gold coated styrene-ethylene/butylene-styrene fibre with an electropolymerized PANI layer for pH sensing. This fibre electrode showed a Nernstian response over the pH range of 4.0–8.0 even when stretched at 100% strain.[10]

Our previous work evaluated fibres made from different suppliers of the conducting polymer poly(3,4-ethylenedioxythiphene)-poly(styrenesulfonate) (PEDOT:PSS) and how solvent treatment

[a] *School of Chemistry and Chemical Engineering, University of Surrey, Guildford, GU2 7XH, UK. E-mail: c.crean@surrey.ac.uk*
[b] *Department of Microbial Sciences, University of Surrey, Guildford, GU2 7XH, UK*

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(both solvent and time) affected the fibres and their properties (size, electrical properties, mechanical properties, XPS studies, electrochemical surface area).[11] PEDOT:PSS can be easily spun into fibres by the industrially-scalable technique of wet-spinning. This helped us to determine that DMSO was superior to the other two solvent treatments because it improved the electrical properties of the fibres while maintaining and even slightly improving their mechanical properties.

In this study, we significantly extend that work by optimising the electrochemical deposition of PANI onto wet-spun PEDOT:PSS fibres *via* three different electropolymerisation methods, to allow for the real-time monitoring of pH. Key variables were optimised and the resulting fibre was used for small volume analysis. We further show that an optimised wearable pH sensor, consisting of PANI-PEDOT:PSS fibres *vs.* a wearable Ag/AgCl pseudoreference electrode, exhibits a Nernstian response over pH 3.0 to 9.0, even in pH-adjusted sweat matrices and human plasma. Microbiological and biocompatibility testing revealed that this biocompatible composite fibre also exhibited significant antibacterial properties.

2. Experimental

2.1. Materials

Aqueous PEDOT:PSS dispersion (Clevios PH 1000, 1 wt%) was procured from Heraeus (Germany). The following analytical grade chemicals were purchased from Sigma Aldrich (UK): dimethyl sulfoxide (DMSO), methanol, boric acid, phosphoric acid, acetic acid, sodium chloride, sodium hydroxide, ammonium chloride, magnesium chloride, pyruvic acid, glucose, urea, calcium carbonate, potassium nitrate, sodium nitrate, polyvinyl butyral (PVB) and *Escherichia coli* and nutrient broth. Human plasma mixed pool product was purchased from TCS Biosciences Ltd (Buckingham UK). The human keratinocytes (HaCaT) cells were purchased from CLS Cell Lines Service. Alamar Blue reagent was procured from ThermoFisher. The gauze fabric (100% cotton) was purchased from a local pharmacy. Ag/AgCl paste (C2130809D5) was obtained from Sun Chemicals® (Parsippany, NJ). An *E. coli* wild-type strain expressing yellow fluorescent protein (YFP; Balaban *et al.*[12]) was used in these experiments. Artificial sweat matrix was prepared containing 6 mM KCl, 0.08 mM $MgCl_2$, 0.18 mM pyruvic acid, 0.17 mM glucose, 5 mM NH_4Cl and 10 mM urea.

2.2. Instrumentation

All electrical resistance analyses were performed by mounting the fibres with silver paint on four electrical pins separated by 2 mm spaces. A constant current of 10 μA was applied across the two outer pins and the resulting voltage across the two inner pins was measured using a Keithley 2001 multimeter. Raman spectroscopy was performed on an inVia confocal Raman microscope (532 nm excitation, Renishaw UK). Scanning electron microscopy (SEM) images were obtained using a JEOL USA JSM-7100F analytical field emission electron microscope at 5.0 keV. Dynamic mechanical analysis (DMA) was performed using a TA Instruments Q800. Analysis was performed using 0.0001 N preload force, and an appropriate rate of force increase, chosen to provide sufficient data points for each sample. Fluorescence intensity measurements during the antibacterial tests were performed on an A1+ confocal Nikon microscope (Nikon A1M on Eclipse Ti-E). An EDAQ potentiostat EA163 and EDAQ e-corder 410 was used for the PANI electropolymerisation and potentiometric measurements.

2.3. Fabrication of conductive fibres

The PEDOT:PSS fibres were produced using the same wet-spinning method as Okuzaki *et al.*[13] Briefly, PEDOT:PSS ink (2 wt%) was prepared from an aqueous stock dispersion (1 wt%) *via* evaporation at 50 °C. The PEDOT:PSS ink was injected *via* syringe (using either a 16 G or 18 G needle with inner diameters of 1.19 mm and 0.18 mm respectively) into a rotating solution (10 rpm) of propan-2-ol and acetone (1 : 1) at the rate of 0.75 or 0.05 mL h^{-1} for diameter 75 μm and 25 μm fibres, respectively.

The resulting wet-spun fibres were either untreated (no solvent treatment post wet-spinning) or treated with DMSO. For DMSO treatment, the fibres were immersed in neat DMSO for 5 min and then dried for 30 min at 140 °C. DMSO-treated fibres with a diameter of 20 μm were selected as the base substrate for the final pH sensor (*vide infra*).

2.4. Fabrication and testing of the pH-sensitive fibre electrodes

To construct pH-sensitive fibres, polyaniline (PANI) was coated onto the fibre electrodes *via* cyclic voltammetry (CV), galvanostatic, and potentiostatic polymerisation in a solution of aniline (0.1 M) in HNO_3 (1.0 M) at room temperature. Solutions were degassed with N_2 before use. For polymerisation by CV, the potential was cycled from −0.2 to +1.0 V *vs.* Ag/AgCl (3 M KCl internal solution, BASi, UK) at 100 mV s^{-1}. For galvanostatic polymerisation, a current density of 2 mA cm^{-2} was applied, as used by Yang *et al.*[14] Potentiostatic polymerisation, was carried out at 0.8 V *vs.* Ag/AgCl.[15] CV cycles were varied between 5 and 20, while deposition times for galvanostatic and potentiostatic deposition were varied from 3 to 10 min to optimise the PANI coating. PANI coated fibres were subsequently rinsed with deionised water and air dried prior to further use. The PEDOT:PSS fibre (1 cm length) was employed as the working electrode with a Ag/AgCl reference electrode and a Pt mesh counter electrode (1 cm^2 geometric area).

Potentiometry was used for pH analysis and the potential of the PANI-PEDOT:PSS fibre electrodes (1 cm length) was measured against a Ag/AgCl double junction reference electrode (3 M NaCl internal solution, Alvatek, UK). Britton–Robinson pH buffered solutions were made from a mixture of 0.04 M boric acid, 0.04 M phosphoric acid, 0.04 M acetic acid and were adjusted to the desired pH (2.0 to 12.0) with 0.2 M NaOH.

The interference studies were performed using the method reported by Nyein *et al.*: interference solutions of 1 mM NH_4Cl, 1 mM $MgCl_2$, 1 mM $CaCO_3$, 8 mM KNO_3 and 20 mM $NaNO_3$ were subsequently added to a Britton–Robinson buffer solution of pH 4.[16] With the addition of an interference solution, there was a 20 s waiting period and then the voltage was measured.

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At the end of the study, the solution was changed to a buffer solution of pH 5 and the voltage was measured again.

2.5. Fabrication and testing of the wearable pseudoreference electrodes

For the quasi-real-world low volume pH analysis, a wearable Ag/AgCl pseudoreference electrode was fabricated. Two methods of constructing a Ag/AgCl pseudoreference electrode were used. In method 1, only a Ag/AgCl-ink was coated onto the substrate surface. In method 2, the already Ag/AgCl-coated substrate was then dip-coated with a reference ion-selective membrane (RF-ISM) as detailed below. Gauze fabric and carbon fibres (CF) were both investigated as substrates.

Method 1. A wearable Ag/AgCl pseudoreference gauze-based electrode was fabricated by applying 0.2 g Ag/AgCl paste to each side of a 1 cm^2 area of gauze fabric using a plastic pipette. The Ag/AgCl paste was applied to one side of the gauze, cured at 80 °C for 20 min, then applied to the other side and cured again. The CF-based pseudoreference electrode was fabricated by dipping a bundle of carbon fibres (length of 3 cm and diameter of 1 mm) into the Ag/AgCl paste for 10 s. The Ag/AgCl-CF electrode was then cured at 80 °C for 20 min.

Method 2. The RF-ISM contained 78 mg PVB and 50 mg NaCl in 1 mL methanol, as described by Parrilla *et al.*[17] An 8 mm portion of the Ag/AgCl-CF and the whole area of Ag/AgCl-gauze were dipped in the RF-ISM using a dipping cycle, in which they were submerged for 20 s before being dried at room temperature for 10 min. This constitutes one dip cycle and was performed up to 5 times. The RF-ISM-coated fibres were immersed for 12 h in 3 M KCl to equilibrate the chloride concentration and therefore stabilise the electrode response.

For low volume analysis, a planar side by side two-electrode set-up was used with the potential of the PANI-PEDOT:PSS fibres electrodes recorded against the fabricated Ag/AgCl pseudoreference electrode (Ag/AgCl-gauze were selected as the reference electrode for the final pH sensor). During analysis, a 200.0 μL drop of pH buffer (3.0 to 9.0) was placed between the two electrodes covering both electrodes. The drop was removed with absorbent tissue between measurements.

2.6. Antibacterial testing of final sensors

The antibacterial testing was performed using the same method as Smith *et al.*[18] *E.coli* cells were picked from a Luria Bertani agar plate, then grown in liquid culture with agitation at 300 rpm for 16 h at 37 °C. 300 μm lengths of thin DMSO PEDOT:PSS fibres (bare and PANI-coated) and 0.09 cm^2 areas of gauze (bare and Ag/AgCl-coated) were cut with a sterilised scalpel blade and sterilised by dipping in ethanol. The bacterial cells were then aliquoted (300 μL) at a final cell density of 1.28×10^8 cells mL^{-1} into wells containing the sensors. The 96-well plate was shaken at 150 rpm at 37 °C for 4 h. Subsequently, each "biofouled" fibre was transferred to a clean well (containing no solution or bacilli). Images were captured across a fixed area of 0.217 mm^2 and the mean intensity of fluorescence was measured with a Nikon confocal microscope using the fluorescein isothiocyanate (FITC) filter at 488 nm and using a 40× air objective lens (NA = 0.96).

2.7. Cell viability studies of final sensors

The biocompatibility testing was performed using the same method as Smith *et al.*[18] 500 μm lengths of thin, DMSO-treated PEDOT:PSS fibres (bare and PANI-coated) and 0.05 cm^2 area of Ag/AgCl-gauze (0.5 cm by 0.1 cm) were dipped in ethanol for 3 h, air-dried and then sterilised for 10 min in a UV/ozone generator (BioForce Nanosciences). The human keratinocyte cells HaCaT (300493, CLS, Germany) were seeded at a density of 10^4 cells per well into 96-well plate and allowed to adhere on the well surface for 24 h at 37 °C in a humidified atmosphere containing 5% CO_2 and 20% O_2. After this period, the sensors were then placed on top of the cells. The cell viability was assessed using the Alamar Blue® assay (Thermofisher Scientific, UK). This bioassay incorporated a redox indicator resulting in colour change of the culture medium according to cell metabolism. The toxicity of this assay is negligible, so it can be used multiple times on the same samples. The samples were incubated with the Alamar blue dye diluted in the cell culture medium according to the manufacturer's instructions for 1.5 h at different culture time points: 1, 3 and 5 d post fibre administration on the cells. The absorbance was measured with a spectrophotometer (Synergy, BioTek, VT, USA) at 570 nm and at 600 nm, as a reference wavelength. Finally, the cell viability was calculated using the dye molar extinction coefficients and appropriate absorbance equations provided by the manufacturer.

Statistical analysis was performed from $N = 3$ independent experiments with at least a triplicate ($n = 3$) per experiment. Error bars represent sample standard deviation. Analysis of variance (Two way ANOVA) was performed using GraphPad Prism® software with a p-value threshold 0.05 to evaluate whether there was a statistical difference between the experimental conditions.

3. Results and discussion

3.1. Development of conductive fibres

A major challenge with using PEDOT is its insolubility in water, therefore, PSS is used as a dopant to form water-dispersible PEDOT:PSS. The addition of insulating PSS, however, has a detrimental effect on the electronic conductivity. PEDOT can have conductivities above 1500 S cm^{-1}, whereas, the water-soluble copolymer PEDOT:PSS has a conductivity below 1 S cm^{-1}, too low to be used for electropolymerisation or sensing.[19] Previous studies have reported that treating PEDOT:PSS with solvents, for example, trifluoromethanesulfonic acid, sulfuric acid and DMSO can increase the conductivity because of the removal of insulating PSS and re-ordering of the polymer chains.[20,21] For example, Kara *et al.* observed that the conductivity of electrospun PEDOT:PSS nanofibres increased 15-fold with the addition of DMSO to the spinning solution.[22]

We previously examined two different suppliers of PEDOT:PSS, different fibre spinning conditions and solvent treatments and found that a combination of Clevios PEDOT:PSS and DMSO solvent treatment gave the best improvement in electrical conductivity with no detriment to the mechanical properties.[11] Further improvements in mechanical properties can be achieved

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Table 1 Summary data (mean ± sample stdev) of all PEDOT:PSS fibre types (n = 3), both before (diameter and conductivity) and after coating with PANI (pH response)

PEDOT:PSS fibre type	Diameter/ μm	Conductivity/ S cm^{-1}	pH response/ mV pH^{-1}
Thick (no treatment)	75 ± 13	125 ± 11	−45 ± 4
Thick (DMSO treatment)	77 ± 7	323 ± 35	−38 ± 7
Thin (no treatment)	21 ± 2	122 ± 18	−49 ± 5
Thin (DMSO treatment)	21 ± 2	802 ± 122	−56 ± 7

using fibre drawing methods post-spinning.[23] In this study, two different fibre dimensions were produced: thick and thin fibres with an average diameter of 70–80 μm and 20–25 μm diameter, respectively. No significant change in conductivity was observed for the different fibre diameters with 125 ± 11 S cm^{-1} and 122 ± 18 S cm^{-1} for thick and thin fibres (no solvent treatment), respectively. After DMSO treatment, the conductivity of thin fibres significantly increased, whereas the conductivity of the thick fibres increased to a lesser degree (Table 1). DMSO treatment removes the PSS on the fibres' surfaces. Proportionally, thin fibres contain less PSS than thick fibres. Therefore, removal of surface PSS by DMSO will have a greater net effect on the conductivity of the thin DMSO-treated fibres compared to the thick DMSO-treated fibres. The most conductive fibre produced was the thin PEDOT:PSS fibre with DMSO treatment, achieving a notable conductivity of 802 ± 122 S cm^{-1}.

Raman characterisation of solvent treatment. Raman spectroscopy was performed to confirm the chemical structure of the PEDOT:PSS fibres before and after DMSO solvent treatment. PEDOT:PSS peaks were observed, which are consistent with previously published spectra. The peaks at 1257, 1367, 1507 and 1566 cm^{-1} correspond to Cα–Cα inter-ring stretching, Cβ–Cβ stretching, asymmetric Cα=Cβ stretching vibrations and aromatic chain vibrations of PSS, respectively (Table 2 and Fig. S2, ESI†).[24] The spectra of the wet-spun PEDOT:PSS fibres showed a change post-DMSO treatment. The peak observed at 1435 cm^{-1} in the untreated 25 μm fibres shifted to 1428 cm^{-1} following solvent treatment (the spectral resolution of the Raman spectrometer was <1.0 cm^{-1}); this peak is attributed to symmetric stretching of the five-membered thiophene ring, which can be present as either the benzoid (Cα=Cβ) or quinoid (Cα–Cβ) resonant structure depending on the oxidation state and the level of doping.[25] This shift is considerably less prominent in the thick fibres from 1436 cm^{-1} (untreated) to 1433 cm^{-1} on solvent post-treated. When the polymer is solvent treated, there is a conformational change of the PEDOT chains from a coiled benzoid structure to a more linear structure. This enhances the mobility of the charge carriers in the PEDOT:PSS, explaining the observed increase in conductivity with DMSO treatment.[26]

Scanning electron microscopy. SEM was performed to determine the surface morphology of the fibres (Fig. S1, ESI†). The surface of the thick fibres is less ridged than the thin fibres, which may be due to the thin wet-spun fibres drying in air more quickly than the thick fibres (smaller exposed surface area), causing a rougher surface. The surface of the untreated fibres is visibly smoother and less ridged than the DMSO-treated fibres. This is because the solvent treatment rearranges the polymer chains and removes some of the electrically insulating PSS.[13]

3.2. Development of pH-sensitive fibres

Different electropolymerisation approaches were trialled to find the ideal conditions for pH sensing PEDOT:PSS fibres. Experimental methods for electrodepositing PANI include cyclic voltammetry (CV), potentiostatic and galvanostatic techniques. Following PANI deposition *via* CV onto the four types of PEDOT:PSS fibres, the pH sensitivity was assessed against a double-junction Ag/AgCl electrode (Column 3 in Table 1). The thick PEDOT:PSS fibres, without and with the solvent treatment, gave the poorest pH sensitivity with a slope of −45 ± 4 and −38 ± 7 mV pH^{-1}, respectively. As established by the conductivity values (Table 1), the thick fibres have a higher ratio of hydrophilic PSS to PEDOT, which leads to more water being absorbed and more fibre swelling during pH analysis that affects the resistance of the electrode and results in a poor quality PANI deposition.[27] The thin fibres without solvent treatment showed improved pH sensitivities (−49 ± 5 mV pH^{-1}). The best pH response was achieved by the thin fibres with DMSO treatment with a Nernstian response (−56 ± 7 mV pH^{-1}). As this DMSO-treated thin PEDOT:PSS fibre achieved the highest conductivity and a Nernstian response when PANI coated, it was down-selected for further optimisation of the PANI coating.

Optimisation of PANI coating. PANI was electrodeposited onto the thin DMSO-treated fibres by CV, potentiostatic and galvanostatic deposition. With potentiostatic deposition, the post polymerisation CV of the PANI-PEDOT:PSS electrode shows redox peaks associated with PANI (Fig. 1(a)). The first redox couple (a in Fig. 1a, anodic peak potential (Epa) at 0.36 V (potentiostatic) and 0.42 V (CV) *vs.* Ag/AgCl) corresponds to the redox interconversion between leucoemeraldine base and emeraldine salt, and the second (c in Fig. 1a, Epa at 0.86 V (potentiostatic) and 0.93 V

Table 2 Raman spectra assignments for wet-spun PEDOT:PSS fibres (n = 3) before and after the DMSO solvent treatment

75 μm (no DMSO)	75 μm (DMSO)	25 μm (no DMSO)	25 μm (DMSO)	
Raman shift/cm^{-1}				Assignments
1257	1257	1257	1257	Cα–Cα inter-ring stretching
1367	1367	1367	1367	Cβ–Cβ stretching
1436	1433	1435	1428	Cα=Cβ stretching
1507	1507	1507	1507	Asymmetric Cα=Cβ stretching
1566	1566	1566	1566	Aromatic chain PSS

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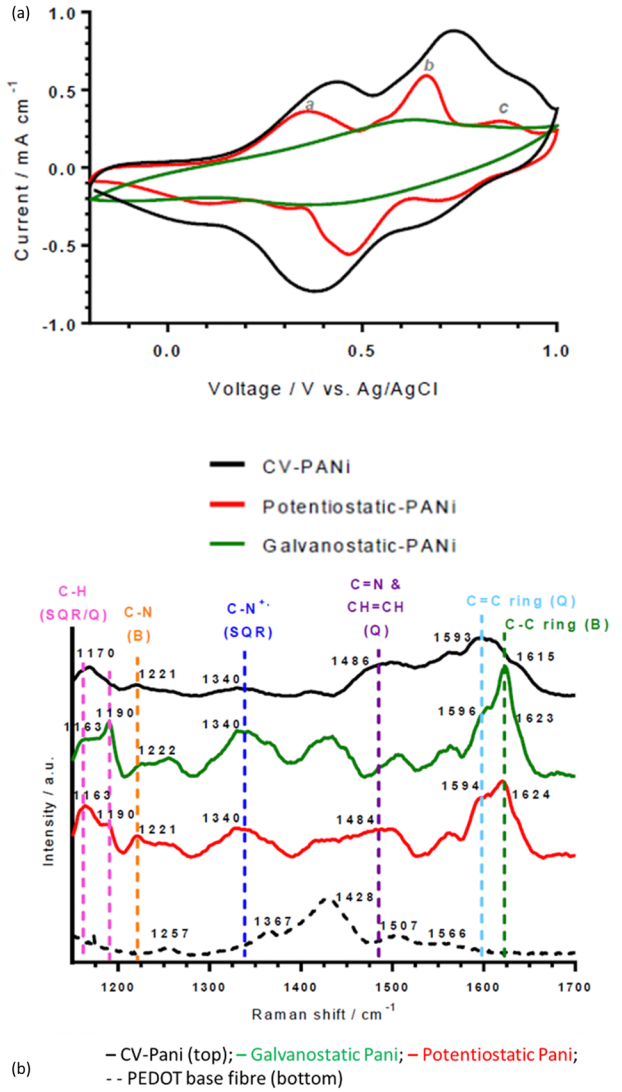

Fig. 1 (a) Cyclic voltammograms and (b) Raman spectra of wet-spun thin, DMSO-treated PEDOT:PSS fibres PANI-coated *via* cyclic voltammetry, potentiostatic and galvanostatic techniques.

(CV) *vs.* Ag/AgCl) to the redox reaction between emeraldine salt and pernigraniline salt.[28]

The middle peak (b in Fig. 1a, Epa at 0.66 V (potentiostatic), 0.74 V (CV) and 0.63 (galvanostatic) *vs.* Ag/AgCl) is normally due to the redox couple of benzoquinone/hydroquinone, which is a degradation product of PANI.[29] However, characteristic Raman peaks for benzoquinone are absent (1644 and 1651 cm^{-1} for C═C and C═O bands, respectively). Thus, the peak may be due to the dimer intermediate, *p*-aminodiphenylamine, which is found at ~0.70 V *vs.* Ag/AgCl and results from head-to-tail coupling of aniline. It will contribute to the intensity of Raman bands attributed to C–C ring stretching (~1620 cm^{-1}) and C–N stretching (~1220 cm^{-1}) in the Raman spectra of the PANI-deposited fibres (Fig. 1(b)). In contrast to the other deposition types, the galvanostatic-deposited PANI fibre has one broad, ill-defined middle peak centred at 0.63 V *vs.* Ag/AgCl (that may be obscuring the other conventional PANI redox peaks).

Interestingly, variations in redox potentials and Raman spectra were observed between the three deposition techniques. CV regularly cycles the electrode potential throughout the deposition (−0.2 V to 1.0 V), cycling the electropolymerised PANI from its non-conducting forms (leucoemeraldine and pernigraniline salts) to its conducing form (emeraldine base, between 0.8 and 1.0 V). Thus, the resulting PANI is in a less-conducting form and is disordered, confirmed by the large oxidation peak at (0.74 V).[30] The presence of this peak at a significantly higher current density to the oxidation peaks corresponding to conducting PANI (peaks labelled a and c in Fig. 1(a)) explains the lower pH sensitivity of the CV-deposited fibres (*vide infra*). This difference in oxidation peaks is also observed in the galvanostatic-deposited fibres, suggesting an explanation for the lower pH sensitivity for these also (see SEM analysis below for a further explanation).

Raman characterisation of deposited PANI. The Raman spectra of the PANI-PEDOT:PSS fibres (potentiostatic-deposited) all exhibited the expected PANI bands situated at 1624, 1594, 1484, 1335, 1221, 1190, 1163 cm^{-1}, which have been assigned to C–C ring stretching (benzenoid diamine; emeraldine base/salt), C═C ring stretching (quinoid diimine; emeraldine base/salt), C═N and CH═CH stretching (quinoid diimine; emeraldine base), C–N^{+} stretching (semiquinone radical cation segment; emeraldine salt), C–N stretching (benzenoid diamine; emeraldine base/salt), C–H bending of both the semiquinone radical cation segment (polaron; emeraldine salt) and quinoid diimine segment (bipolaron; emeraldine salt), respectively (Fig. 1(b) and Table S1, ESI†).[31] Raman shifts corresponding to both emeraldine base and salt forms were present because the emeraldine base to emeraldine salt transition takes place between pH 3.0 and pH 4.0 and the Raman spectra were obtained on fibres which had previously been in solution at pH 3.5.[32]

There are subtle differences between the Raman spectra of the fibres made from the different electrochemical deposition techniques (Fig. 1(b)). The difference in maximum intensity between the peaks at *ca.* 1163 and 1190 cm^{-1} (assignments in ESI†) gives an indication of the global oxidation level of the emeraldine salt.[33] In the CV and potentiostatic deposited fibres, there is a greater intensity of the peak at *ca.* 1163 cm^{-1} (C–H bending in the bipolaron form) than at 1190 cm^{-1} (C–H bending in the polaron form), showing that the deposited PANI is present mainly in the bipolaron form. In contrast, in the galvanostatic deposited PANI, there is a greater intensity of the peak at 1190 cm^{-1} than the peak at *ca.* 1163 cm^{-1}, indicating a higher proportion of polarons than bipolarons present. This is supported by the large peak at 1340 cm^{-1} attributed to C–N^{+} stretching in the polaron form.[30] Interestingly, this follows the pH sensing trend, in which a greater and more repeatable pH sensing capability was achieved by the PANI containing the higher proportion of bipolarons than polarons (potentiostatic and CV-deposited compared to galvanostatic-deposited PANI).

Scanning electron microscopy. There were notable differences observed between the surfaces of PANI-PEDOT:PSS coated using CV, galvanostatic and potentiostatic polymerisation as analysed by SEM (Fig. 2). A possible explanation can be found for this by considering the variation in the deposition

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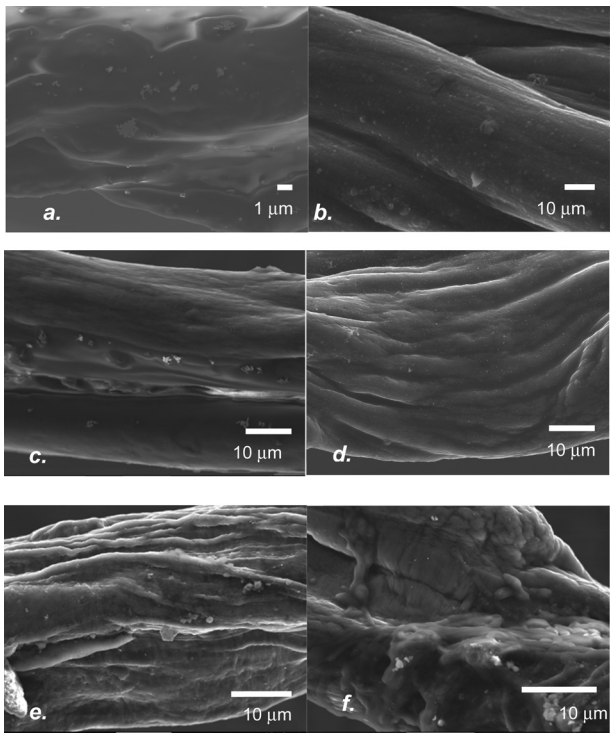

Fig. 2 Scanning electron microscopy images of the PEDOT:PSS fibres: (a) thick with no treatment (PANI coated by CV), (b) thick with DMSO treatment (PANI coated by CV), (c) thin with no treatment (PANI coated by CV), (d) thin with DMSO treatment (PANI coated by CV), (e) thin with DMSO treatment (PANI coated by galvanostatic deposition) and (f) thin with DMSO treatment (PANI coated by potentiostatic deposition).

conditions. During CV, the potential was cycled between −0.2 V and 1.0 V and the oxidation of aniline to PANI only occurs between 0.8 and 1.0 V.[25] Thus, there is limited growth in each cycle and the nucleation of PANI begins afresh each time, resulting in a layer-by-layer deposition. Additionally, the intermediates formed during polymerisation, such as dimers and short chain oligomers, could be released or dissolved into the electrolyte during this period of interruption in the polymerisation (as opposed to galvanostatic and potentiostatic deposition).[34] This can explain the relatively smooth surface of PANI covering the fibre (Fig. 2(d)). On the other hand, in galvanostatic/potentiostatic conditions, the constant current/potential leads to uninterrupted polymerisation resulting in improved PANI growth as observed in the clusters on the surface of the fibre (Fig. 2(e) and 3(f)).

pH sensitivity. The PANI-PEDOT:PSS fibres were assessed for their pH sensitivity. All polymerisation methods achieved near Nernstian responses; however, the highest linearity (R^2 = 0.995), and most repeatable pH sensing, was achieved after potentiostatic deposition (Fig. 3). Therefore, potentiostatic deposition was chosen for further development of the wearable pH sensor. This linear, Nernstian pH response presents a comparable response to the previous screen-printed pH sensor successfully developed by Vacca *et al.*, which consisted of electropolymerised PANI on ink-jet printed PEDOT:PSS, achieving 59 mV pH^{-1} from pH 2 to 14 (repeatability not reported).[9]

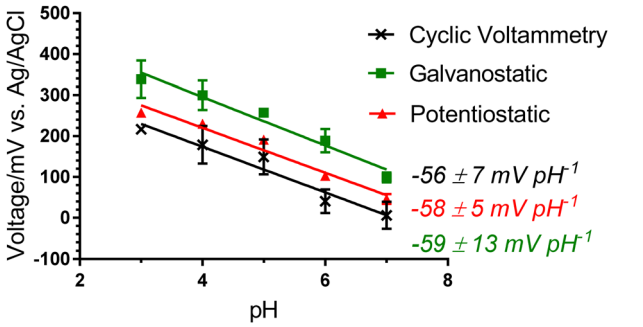

Fig. 3 pH sensitivity of thin, DMSO-treated, wet-spun PEDOT:PSS fibres PANI-coated by CV (10 cycles), potentiostatic (600 s) and galvanostatic (300 s) techniques. The error bars represent the sample stddev of n = 3 replicate measurements.

3.3. Development of wearable sensor

A wearable Ag/AgCl pseudoreference electrode was fabricated to develop a solid-state pH sensor. A requirement of a pseudoreference electrode is to present a constant voltage regardless of variation in pH and other sweat/wound-related interferences.[35] Two main approaches have been adopted previously in the literature for the fabrication of pseudoreference electrodes. The first approach coated Ag/AgCl-ink onto a surface, and the second is to further dip-coat the Ag/AgCl-coated substrate with a reference ion-selective membrane (RF-ISM). Both approaches were undertaken in this research to determine the optimal pseudoreference electrode for pH analysis.

The Ag/AgCl-gauze demonstrated the best and most repeatable response with a slope of −2 ± 0 mV pH^{-1}, compared to −5 ± 1 mV pH^{-1} for Ag/AgCl-CF (Fig. S3, ESI†). Furthermore, the gauze fabric is more similar to typical bandages than CF. Hence, Ag/AgCl-gauze was selected for further solid-state analysis.

The wearable Ag/AgCl pseudoreference electrode (Ag/AgCl-gauze) was connected to the PANI-PEDOT:PSS fibre electrode and the potential was measured as interfering ions (NH_4^+, Mg^{2+}, Ca^{2+}, K^+ and Na^+) at physiologically relevant concentrations were added sequentially to a pH 4 buffered solution (Fig. 4(a)). The largest voltage change observed was ±2 mV, which was significantly smaller than 53 mV after the change to pH 5 (Fig. 4(a)) thus demonstrating pH selectivity. This shows that the fibre can sense pH effectively regardless of the sweat and wound-related interferences present.

A Nernstian response of −58 ± 1 mV pH^{-1} was achieved over the pH range 3.0 to 9.0 with a 200.0 μL drop of analyte solution covering the electrodes of the solid-state system (Fig. 4(b)). This pH range is across the physiologically relevant pH range for skin. Likewise, the electrodes obtained a Nernstian response in both a pH-adjusted sweat matrix (common sweat interferences, *e.g.*, 6 mM KCl) and pH-adjusted human plasma (common wound-related interferences, *e.g.*, proteins) (Fig. 4(b)). Small volume (200.0 μL) analysis over the physiologically critical pH range for skin and wounds (pH 4.0 to 9.0) demonstrates the potential of the PANI-PEDOT:PSS fibre electrode in an epidermal patch for sweat and wound monitoring.

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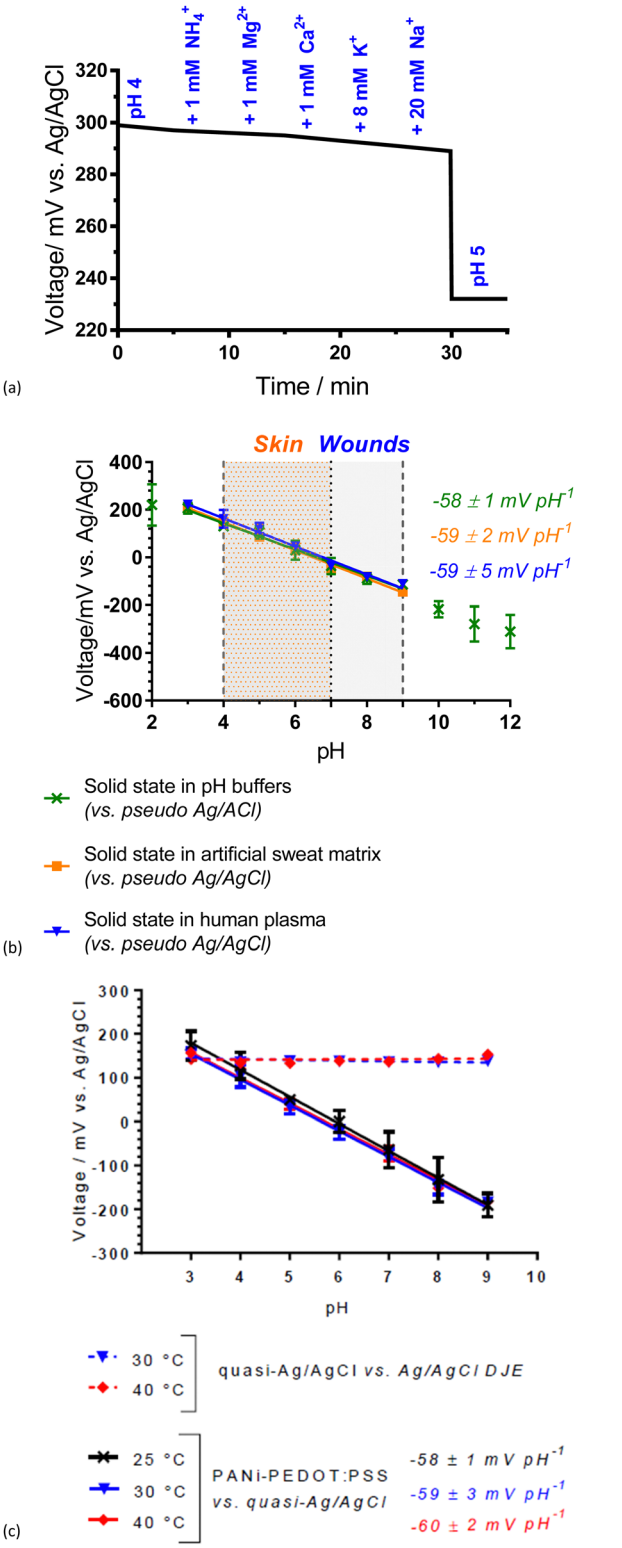

Fig. 4 Potentiometric response of solid-state pH analysis: (a) Selectivity when various sweat-related interferences (NH_4^+, Mg^{2+}, Ca^{2+}, K^+ and Na^+) were added to the pH 4 aqueous samples (20 mL). (b) Response in buffered solution to varying pH levels, in pH-adjusted artificial sweat matrix and human plasma (200 μl) (*vs.* Ag/AgCl pseudoreference electrode). (c) Potentiometric (open circuit) response of PANI-PEDOT:PSS fibres and quasi-reference electrode in buffered solutions to varying pH levels in temperatures 25, 30 and 40 °C. Error bars represent the standard deviation of n = 3 replicate measurements.

Table 3 Mean ultimate tensile strength, breaking strain and Young's modulus values for the electrodes in this article. The errors represent the sample standard deviation of three replicate measurements

Fibre electrode	Ultimate Tensile Strength/MPa	Breaking Strain/%	Young's Modulus/GPa
PEDOT:PSS fibre	33 ± 21	13 ± 2	0.6 ± 0.1
PANI-PEDOT:PSS fibre	32 ± 11	4 ± 1	2.0 ± 0.9
Bare gauze fabric	141 ± 25	9 ± 5	1.7 ± 0.7
Ag/AgCl-gauze	139 ± 11	<3 ± 1	23.0 ± 5.0

A near-Nernstian response of the pH sensing fibres was maintained at temperatures relevant to skin (between 25 and 40 °C), with non-significant variation (Fig. 4(c)). An increase in slope with temperature is expected from the Nernst equation (−59.2 mV pH^{-1} at 25 °C and −62.1 mV pH^{-1} at 40 °C). Wound/skin temperatures range between 31 and 37 °C and therefore the PANI-PEDOT:PSS fibres have good potential to effectively monitor skin/wound beds while being worn on the body.

Mechanical properties. The mechanical strength of the PANI-PEDOT:PSS sensor fibre and Ag/AgCl-gauze pseudo-reference, was assessed by dynamic mechanical analysis (DMA) and the results can be found in Table 3. The PANI-PEDOT:PSS fibres were robust enough to be handled and could be integrated within a bandage system, but did not have as much breaking strength as the control gauze fabric. Both the breaking strain and Young's modulus of the PEDOT:PSS fibre and gauze fabric increased after coating with PANI and Ag/AgCl paste respectively (Table 3), indicating the lower flexibility and increased brittleness of the coated electrode materials.

Antibacterial properties. Antibacterial properties are an important requirement for a wearable sensor. Increased microbial growth would hinder and discourage the healing of a wound. The antibacterial properties of the materials used here were evaluated with wild type *E. coli* expressing yellow fluorescent protein (YFP) as a model system for the skin/wound surface. A bacterial cell suspension was incubated with the sensors, after which the growth or attachment of the bacteria was evaluated by measuring fluorescence intensity (FITC, 488 nm), which gave a quantitative indication of the extent of the bacteria on the sensors.

PEDOT:PSS fibre (thin, DMSO-treated) showed moderate antibacterial activity (intensity = 138 ± 47 a.u.), compared to the bare gauze fabric (intensity = 945 ± 101 a.u.) that is commonly used in wound bandages (Fig. 5(a)). The antibacterial activity of the fibres was increased further with the PANI-coating, which lowered bacterial intensity further to 74 ± 2 a.u. PANI is known for its antibacterial activity due to the charged nitrogen in the PANI backbone attacking the bacterial cell wall.[36] PANI has been previously used as an antibacterial agent in textiles.[33]

The Ag/AgCl-gauze exhibited a notably lower fluorescence intensity (98 ± 27 a.u.) than bare gauze fabric (941 ± 101 a.u.), demonstrating a high antibacterial activity (Fig. 5(a)). This was expected as recent evidence suggested that AgCl slowly releases silver ions, which are antibacterial.[37] Previously, this antibacterial effect was proved to accelerate wound healing[34] and was used as an antibacterial agent in wound dressings.[38]

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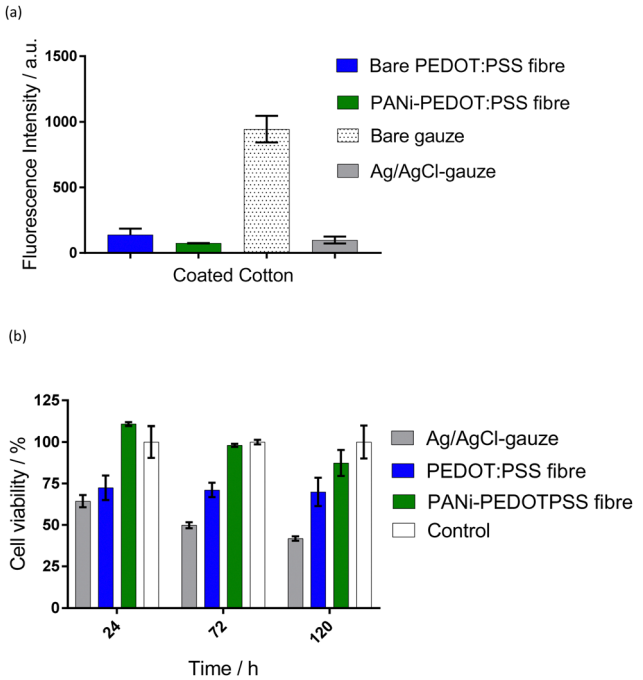

Fig. 5 (a) Mean intensity of fluorescence (FITC, 488 nm) across a fixed area (0.217 mm^2) of PANI-coated PEDOT:PSS fibre and Ag/AgCl-gauze biofouled with *E. coli* wild type strain expressing yellow fluorescent protein (YFP). Error bars represent the standard errors of three independent experiments. (b) Cell viability compared to control taken at three time points: days 1, 2 and 4 post sensor administration. Wells without sensors were used as controls and were considered 100% viable. Error bars represent the standard errors of three independent experiments.

Fluorescence imaging of the bare gauze fabric showed bacterial cells growing within the fabric (data not shown). However, the Ag/AgCl-coating has discouraged this growth or attachment of the bacteria, protecting the surface of the reference electrode from bacterial growth. These results highlight the potential of both the PANI-PEDOT:PSS fibres and Ag/AgCl-gauze in reducing bacteria in an epidermal patch during exercise and/or reducing the likelihood of an infection in wounds.

Cell viability studies. The biocompatibility of these materials is important for wearable applications to avoid harm or irritation to the wearer. Thus, a proof-of-concept study was performed to investigate the biocompatibility of the sensor materials. Human skin cells were seeded on a 96-well plate and the fibres/gauze plated on top. Cell viability was assessed on days 1, 3 and 5 to give an initial indication of cytotoxicity, using Alamar Blue® assay. The colour change as a result of cell metabolism was measured (at absorbance; 570 nm) and the cell viability was calculated from this. Wells without sensor materials were used as controls and were considered as 100% viable cells.

The cell viability on the PEDOT:PSS fibres (thin, DMSO-treated) showed a significant decrease ($p < 0.05$) of 29% after 24 h compared to the empty well control (Fig. 5(b)). The PANI-coated PEDOT:PSS fibres demonstrated no significant change in cell viability compared to the control. There average cell viability number seems to be lower at longer timepoints and therefore further experiments need to be conducted to confirm the biocompatibility of these materials. Ag/AgCl-gauze induced a toxic reaction in the cells, reducing the viability by 37% after 24 h compared to the control ($p < 0.05$). A possible explanation is the presence of methyl carbitol in the Ag/AgCl ink used to coat the gauze since previous studies have shown that Ag/AgCl is not toxic.[39] Although future studies are needed to investigate the biocompatibility of the solid-state pair, this proof-of-concept study shows promise for the use in wearable sensors.

4. Conclusions

Successful pH-sensing fibre electrodes have been developed based on PANI and PEDOT:PSS. PEDOT:PSS fibres were made by wet-spinning followed by solvent treatment with DMSO. Subsequent potentiostatic electropolymerisation of PANI onto the surface facilitated pH sensing. The formulation of a wearable Ag/AgCl pseudoreference involved curing Ag/AgCl ink onto gauze fabric (analogous to wound dressings). The pH sensing electrode (PANI-PEDOT:PSS) was successful when measured in a low volume sensing configuration *vs.* the wearable pseudoreference electrode. The fibre and gauze electrodes demonstrated a selective, Nernstian response in both pH-adjusted sweat matrices (common sweat interferences, *e.g.*, 6 mM KCl) and pH-adjusted human plasma (common wound-related interferences, *e.g.*, proteins). Significant antibacterial properties were demonstrated for both the pH sensing and reference electrode. Additionally, the PANI-PEDOT:PSS fibres showed promising biocompatible properties. Next steps include enhancing the strength of the PANI-PEDOT:PSS fibres in addition to further biocompatibility investigation.

Conflicts of interest

There are no conflicts to declare.

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