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Cite this: *Environ. Sci.: Atmos.*, 2024, 4, 1255

Toxicological evaluation of SVOCs in exhaust emissions from light-duty vehicles using different fuel alternatives under sub-freezing conditions†

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Semivolatile organic compounds (SVOCs) in exhaust gas, though not directly regulated by emission standards, play a crucial role in assessing both conventional and alternative fuels. Our aim is to compare the differences in and toxicological effects of SVOC exhaust emissions from conventional and alternative fuels under sub-freezing conditions. High levels of NO_x, CO₂ and PAHs in SVOCs were observed in DI-E2 (EN590 winter-grade diesel), with E10 (gasoline with 10% ethanol) exhibiting higher CO₂ and PAH levels compared to E85 (high-blend ethanol with an 83/17% ethanol–gasoline ratio). SVOCs from DI-E6 (EN590 diesel) demonstrated significant cytotoxicity, while E10 resulted in higher inflammatory mediators and genotoxicity. Our findings show that SVOC composition and toxicity in exhaust gas differ based on the fuel type. Despite new emissions regulations reducing diesel vehicle emissions, SVOC toxicity remains unchanged. Toxicity from SVOCs in compressed natural gas and ethanol/gasoline vehicles is notable, with gasoline exhaust showing high inflammatory and genotoxic potential.

Received 16th May 2024
Accepted 29th September 2024

DOI: 10.1039/d4ea00062e

rsc.li/esatmospheres

Environmental significance

Due to the difficulty in collecting and detecting semivolatile organic compounds (SVOCs), they are not directly regulated by traffic emission standards. However, their presence in exhaust gas is crucial for evaluating both conventional and alternative fuels. We measured various exhaust concentrations of SVOCs emitted by newly regulated passenger vehicles using different fuels under low-temperature conditions, along with the cytotoxicity of SVOCs. We found that under the new emission standards, the toxicity of SVOCs emitted from both conventional and clean fuels cannot be ignored.

1. Introduction

Over the past few decades, the European Union (EU) has implemented a series of road emission standards aimed at controlling the release of carbon dioxide (CO₂), nitrogen dioxide (NO₂), and other gases, along with particulate matter (PM), from vehicles in traffic.¹ Various new technologies have reformed engines, fuels and exhaust aftertreatment to meet the emission standards. Diesel engines compliant with the 2007 standards, featuring exhaust gas recirculation (EGR), diesel oxidation catalysts, and diesel particulate filters (DPFs), have achieved

a significant reduction of 90% in PM, as well as over 90% in volatile organic compounds (VOCs) and semivolatile organic compounds (SVOCs) compared to older engine models.² The 2010 engine, fitted with urea selective catalytic reduction, achieved a reduction of over 90% in nitrogen oxides (NO_x) compared to the 2007 engine.² However, these stringent emission standards, new internal combustion engine technologies, and advancements in fuel and exhaust aftertreatment have not effectively improved air quality.^{3,4} This lack of improvement can be attributed to factors such as an aging fleet,⁵ an increase in the overall number of vehicles, and a transition from diesel to gasoline-powered vehicles. With the worsening environmental pollution and the depletion of natural resources, it is imperative to explore and use effective alternative fuels for diesel and gasoline. At present, there is insufficient research on the impact of alternative fuels, for example, ethanol and natural gas, on vehicle emissions, air pollution, and human health.

SVOCs are compounds that are volatile enough to exist as vapors in engine exhaust but can condense under atmospheric conditions. They are largely adsorbed on the surfaces of indoor materials⁶ and airborne particles.⁷ Both diesel and engine lubricating oil are primary components of diesel engine

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† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4ea00062e>



exhaust, with engine lubricating oil being a complex mixture of SVOCs.⁸ The oxidation of SVOC vapors, including engine lubricating oil and *n*-pentacosane, results in the significant formation of secondary organic aerosols.^{8–11} Despite the classification of SVOCs as indoor organic pollutants by the World Health Organization (WHO)⁶ and their exclusion from direct regulation by traffic emission standards,⁹ their presence in exhaust gas, alongside VOCs and PM, is essential for evaluating the harmful effects of conventional and alternative fuel combustion in engines.

Current studies indicate that compressed natural gas (CNG) fueled vehicles equipped with a three-way catalyst have extremely low levels of particle counts.¹² Furthermore, CNG-fueled vehicles demonstrate reduced levels of carcinogenic polycyclic aromatic hydrocarbons (PAHs) in PM when compared to gasoline or diesel vehicles.¹³ Limited research has been conducted on the generation of SVOCs from alternative fuels. Additionally, higher ethanol blends in gasoline have been found to decrease PM emissions in exhaust gas, with lower toxicological effects observed in PM-bound PAHs.^{14,15} This phenomenon has also been noticed at sub-freezing ambient temperatures.^{16,17} Therefore, besides differences in fuel, the running and start-up conditions also affect emissions, particularly at low temperatures, which are associated with higher emissions from passenger cars.^{18,19} However, although some studies have been conducted on this subject,^{5,16} the impact of low temperature on emission toxicity remains insufficiently explored.

Therefore, we collected SVOCs from the exhaust of one Euro 2 and four Euro 6 emission-regulated cars at -7°C . The Euro 2 diesel-powered car (DI-E2) used EN590 winter-grade diesel fuel, while the four Euro 6 cars included DI-E6 using regular EN590 diesel, CNG (compressed natural gas), E10 using gasoline with 10% ethanol, and E85 using high-blend ethanol fuel with an 83/17% ethanol–gasoline-ratio. We exposed RAW264.7 mouse macrophages to the collected SVOCs and evaluated their toxicological effects. DI-E2 and DI-E6 are two types of diesel vehicles that meet different EU standards, and they use different types of diesel fuel. E10 and E85 are ethanol–gasoline blends, with E10 being more commonly used in Finland. E85 and CNG are considered clean energy options and are also available in other European countries. Our aim is to compare the differences in and toxicological effects of SVOC exhaust emissions from conventional and alternative fuels under sub-freezing conditions.

2. Materials and methods

All the sampling methods and details concerning SVOCs are described in the Technical Research Center of Finland (VTT) report of Aakko-Saksa *et al.*² The following sections briefly describe some of the important factors.

2.1 Cars and fuels

SVOCs were collected from the exhaust of one older Euro 2-regulated diesel car and four new Euro 6 emissions-regulated

cars by VTT. SVOCs were collected from the diluted exhaust gas after PM filters. The Euro 2 diesel-powered car (DI-E2) used EN590 winter-grade diesel fuel, while the Euro 6 diesel car (DI-E6) used regular EN590 diesel. Compressed natural gas (CNG), gasoline with 10% ethanol (E10), and high-blend ethanol fuel with an 83/17% ethanol–gasoline-ratio (E85) were used in other Euro 6 cars. Detailed information on the Euro 6 and 2 diesel cars' engines and fuels is presented in the report² and our previous article.⁵

2.2 SVOC sampling and extraction

A standard PM sampling system was used for SVOC sampling. The collection and analysis of SVOCs were conducted using Empore SDB-XC disks (diameter 47 mm). SVOC Empore disks were positioned as back-up filters for SVOC sampling in a standard PM sampler, which used primary filters TX-40. The sample flow rate was 17.5 L min^{-1} , and the Empore disks were then subjected to analyses for PAHs and oxidative potential (dithiothreitol, DTT, $\text{HSCH}_2(\text{CH}(\text{OH}))_2\text{CH}_2\text{SH}$). Additionally, some SVOCs were collected into Tenax TA-Carbopack B tubes for chemical analyses. Tenax tubes were positioned after the high-capacity PM filters (fluorophore) in the PM sampling system, utilizing a side stream of diluted exhaust gas. A sample flow of approximately 0.1 L min^{-1} was drawn through Tenax TA-Carbopack B tubes using a separate pump throughout the entire driving cycle. One background Tenax-tube was collected, and ten SVOC samples were collected with Tenax-tubes. Three parallel tubes were collected for DI-E2, two for E85 and E10 and only one for CNG and DI-E6. Detailed sampling flow data and methods are provided in the report.² After the sampling filters were washed with high-performance liquid chromatography (HPLC)-gradient grade methanol. SVOC samples were obtained through rotary evaporation and nitrogen blowing. SVOC samples were then stored at -20°C .

2.3 Chemical characterization

Nitrogen oxides were analyzed by chemiluminescence (CLD). NO_x emissions refer to the sum of exhaust NO and NO_2 , calculated as NO_2 . CO and CO_2 were analyzed using the non-dispersive infrared principle. Total hydrocarbon (HC) emissions were measured using a flame ionization detector. Detection limits are provided in ESI Table A2.†

Twenty-four individual PAHs in SVOCs were analyzed from the fluorophore filters and Empore disks using the ISO 16000-6:2011 and EN 14662-4:2005 analysis methods. The disks were extracted with toluene using an ultrasonic bath and then analyzed. It was noticed that following toluene extraction of the Empore disks, the solution became cloudy (milky), and a sediment settled at the vial's bottom prior to analysis. This indicated potential partial degradation of the Empore disks during toluene extraction using an ultrasonic bath. However, the sediment didn't interfere with the PAH analyses. The detection limits ranged from 10 to $30\text{ ng (1 }\mu\text{g m}^{-3})$ per compound, and the measurement uncertainty was 30%.

The dithiothreitol ($\text{HSCH}_2(\text{CH}(\text{OH}))_2\text{CH}_2\text{SH}$, DTT) assay was employed to evaluate the oxidative potential of SVOCs. The DTT



assays were conducted at Biosafe Biological Safety Solutions Ltd according to the procedure outlined by Charrier & Anastasio,²⁰ with the modification of using 200 μM DTT in Chelex treated 0.10 M phosphate buffer (77.8 mM NaH_2PO_4 and 22.2 mM KH_2PO_4). OD 412 nm measurements were conducted at five, ten, fifteen, and twenty minutes. The DTT assay was performed under soft natural light conditions. Chelex-treated 0.1 M phosphate buffer served as the negative control, while 9,10-phenanthrenequinone (30 μM) was used as the positive control. The value of the negative sample control (methanol) was subtracted from the sample results. The analysis was conducted with three replicates. The positive control resulted in a DTT loss 3.5 times higher than that of the sample with the highest oxidative potential. DTT loss in phosphate buffer did not exceed the detection limit. DTT assay analysis began with DI-E2 samples, which were submitted for DTT analysis on a mass basis (SVOC mg per sample), concurrently with testing the reference Empore filter (SVOC). DTT samples of E10, E85, CNG, and DI-E6 were analyzed based on volume (m^3 exhaust gas per sample).

2.4 Cell culture and exposure experiments

The RAW264.7 mouse macrophage cell line was obtained from the American Type Culture Collection (ATCC). RAW264.7 mouse macrophages were cultured in RPMI 1640 cell culture medium (Life Technologies), supplemented with 10% heat-inactivated fetal bovine serum (Sigma, USA), 1% penicillin-streptomycin, and 1% L-glutamine (Life Technologies, USA) in a 5% CO_2 atmosphere at 37 °C. The day before the exposure experiment, the cell suspension was diluted to 2×10^5 cells per mL and 5×10^4 cells per mL and then cultured on 12-well plates and 96-well plates (Costar, Corning, NY, USA) for 24 hours, respectively. On the day of the exposure experiment, the medium was replaced with a fresh medium and the cells were allowed to stabilize for one hour.

The SVOC samples were dispersed in pyrogen-free water (Sigma W1503, USA) and dimethyl sulfoxide (DMSO) (final concentration of 0.3% v/v in the cell culture medium, with a dose of 150 mg mL^{-1}) and then sonicated for 30 min. The concentration of SVOC samples before exposure was 1000 L mL^{-1} . RAW264.7 cells were treated with the pyrogen free water control, the DMSO control, the blank membrane control, and three doses of each SVOC sample for 24 hours. For E10 and CNG, the doses were 0.005, 0.01, and 0.025 L mL^{-1} , while for E85, DI-E2 and DI-E6, the doses were 0.001, 0.005, and 0.01 L mL^{-1} . All experiments were conducted in triplicate with a minimum of three replicates.

After 24 hours of SVOC exposure, all the cell culture medium was collected and stored at -80°C . We recovered cells adhering to the bottom of each well using a cell scratcher with 600 μL of cold phosphate buffered saline (PBS). All the cell tubes were then centrifuged at 8000 rpm for 5 min at 4 °C, the supernatant was removed, and the cells were resuspended with 500 μL of cold PBS. The collected cells underwent toxicological analysis. For the detection of cell membrane integrity and intracellular oxidative potential, 200 μL of cell suspension was used. For cell cycle detection, 180 μL of cell suspension was used. For the detection of genotoxicity (comet assay), 120 μL of cell

suspension was used. The cell culture medium was employed for the detection of inflammatory mediators: TNF- α and MIP-2.

2.5 Toxicological analyses

2.5.1 Cellular mitochondrial activity (CMA). CMA was assessed using the MTT-test in 96-well plates. After exposure to SVOCs, 25 μL of MTT-solution was added to each well and incubated for 2 hours at 37 °C. Subsequently, 100 μL of sodium dodecyl sulfate (SDS) buffer was added to each well and incubated overnight at 37 °C. Absorbance was measured at 540 nm using a spectrophotometric plate reader (BioTek, SYNERGY H1, USA). The loss of cell mitochondrial activity was calculated as a percentage relative to the control group.

2.5.2 Cell membrane integrity and intracellular oxidative potential. Cell membrane integrity was assessed by the propidium iodide (PI) exclusion assay, and intracellular oxidative potential was determined by the 2',7'-dichlorofluorescein (DCF) assay. 100 μL of cell suspension was transferred to a 96-well plate, and 8 μL of DCF solution was added to each well. Absorbance was measured and recorded using the spectrophotometric plate reader at 485 nm three times, at 30-minute intervals, after incubation at 37 °C. Subsequently, 7.2 μL of PI solution was added to each well, stirred briefly with a plate shaker, and incubated for 20 min at 37 °C. We detected the absorbance using a spectrophotometric plate reader and recorded it at 540 nm. Following that, 20 μL of 10% Triton X-100 was added to each well and incubated for 20 min at room temperature and covered with foil, on a plate shaker. Finally, the absorbance was measured once more.

2.5.3 Inflammatory mediators. Tumor necrosis factor alpha (TNF- α) and macrophage inflammatory protein 2 (MIP-2) in the cell culture medium were immunochemically analyzed according to manufacturers' recommendations using commercial enzyme linked immunosorbent assay (ELISA) kits (R&D Systems, Minneapolis, MN, USA). We detected the absorbance with the spectrophotometric plate reader and recorded it at 450 nm. The assay range for mouse TNF- α and MIP-2 was 31.2–2000 pg mL^{-1} and 15.6–1000 pg mL^{-1} , respectively.

2.5.4 Cell cycle. A total of 180 μL of cell suspension was fixed in 4 mL of cold 70% ethanol and stored at 4 °C. On the day of the test, the cells (480 g, 5 min, 4 °C) were centrifuged and ethanol was removed. Subsequently, the cells were washed with 1 mL cold PBS and resuspended in 250 μL of cold PBS. To each tube, 7.5 μL of RNase A (10 mg mL^{-1}) was added, and the cells were vortexed and incubated at 50 °C for one hour. Afterward, 4 μL of PI solution was added to each tube, and the cells were vortexed again and incubated at 37 °C for 30 min. The cell cycle phases were analyzed by flow cytometric analysis using a BD FACSCanto™ II (BD Biosciences, San Jose, CA USA).

2.5.5 Genotoxicity. A total of 60 μL of cell suspension was mixed with 540 μL of freeze medium (40% of filtered FBS, 50% RPMI, 10% DMSO) and frozen at -80°C . Samples were thawed and centrifuged (8000 rpm, 5 min, 4 °C). A volume of 50 μL of medium was retained in the cell tube, and 20 μL of cell suspension was mixed with 80 μL of 0.5% low melting point



agarose (LMPA). Eighty μL of this cell LMPA mixture was then spread onto microscope slides coated with 1% normal melting point agarose (NMPA). A glass cover plate was added onto the microscope slide, and all the microscope slides were placed on an ice block for 5 min before removing the glass cover plate. Lysis buffer was then added to the microscope slides and left for 1 hour at 4 °C. Subsequently, all the microscope slides were placed into the electrophoresis chamber, and the electrophoresis buffer was added to the chamber. The slides were incubated in complete darkness for 40 min.

Next, the electrophoresis was set for 20 min at 24V/300 mA. Following that, we transferred all the microscope slides to a clean plate and rinsed them three times with neutralization buffer. DNA was fixed on the microscope slides using 99% ethanol for 10 min. After removing ethanol, all the microscope slides were dried overnight at room temperature and stored under dry, dark conditions. Seventy-five μL of diluted ethidium bromide dye was pipetted onto the microscope slides, covered with a glass plate, and left for 30 min to allow for DNA binding. The nuclei in ethidium bromide (100 cells per dose) were analyzed using a special image analysis system (Comet assay IV, Perceptive Instruments Ltd, Suffolk, UK). Tail intensity was used as the parameter in the statistical analysis.

2.6 Statistical analysis

The comparison of the results for all cell experiments in different groups was conducted using one-way analysis of variance (ANOVA), followed by the least squares difference (LSD) method to compare paired differences between the groups. A p -value < 0.05 was considered to be statistically significant.

3. Results

3.1 SVOC emissions and chemical composition

The averages of SVOC exhaust and NO_x emissions, as well as concentrations of SVOC-bound organic and inorganic constituents from exhaust gases of one Euro 2 and four Euro 6 engines, are shown in Table 1. DI-E2 exhibited the highest NO_x and CO_2 emissions, while E10 showed the least. E85 showed the highest CO exhaust and HC emissions, and CNG had the highest methane (CH_4) emissions, followed by E85. The total amount of SVOC-bound organic constituents, particularly PAHs, was the largest in E10, with DI-E6 having the least. The proportion of carcinogenic PAHs in total PAHs followed this order: E10 (59.22%), E85 (54.69%), DI-E2 (44.59%), DI-E6 (25.00%), and CNG (21.40%). SVOCs from DI-E2 produced more NO_x , CO, CO_2 , total HCs, and PAHs than DI-E6, while SVOCs from E85 emitted

Table 1 Emissions of SVOC exhaust, NO_x emissions, and SVOC-bound organic constituents from exhaust gases of one Euro 2 and four Euro 6 engines

	DI-E2 (Euro 2)	E10 (Euro 6)	CNG (Euro 6)	E85 (Euro 6)	DI-E6 (Euro 6)
Nitrogen oxides NO_x (mg km^{-1})	928.3	19.2	151.2	36.4	333.1
Carbon monoxide, CO (mg km^{-1})	472.1	463.9	42.1	2433.3	79.5
Carbon dioxide, CO_2 (g km^{-1})	241.0	176.4	129.1	158.6	187.7
Total hydrocarbons, HCs (mg km^{-1})	2.48	5.28	7.46	12.59	0.95
Methane, CH_4 (mg km^{-1})	3.8	7.5	95.9	55.8	9.2
Organic constituents in SVOCs: Polyaromatic hydrocarbons, PAHs					
Sum of total PAHs ($\mu\text{g km}^{-1}$)	23.37	51.03	2.29	25.89	0.04
Sum of total carcinogenic PAHs ($\mu\text{g km}^{-1}$)	10.42	30.22	0.49	14.46	0.01

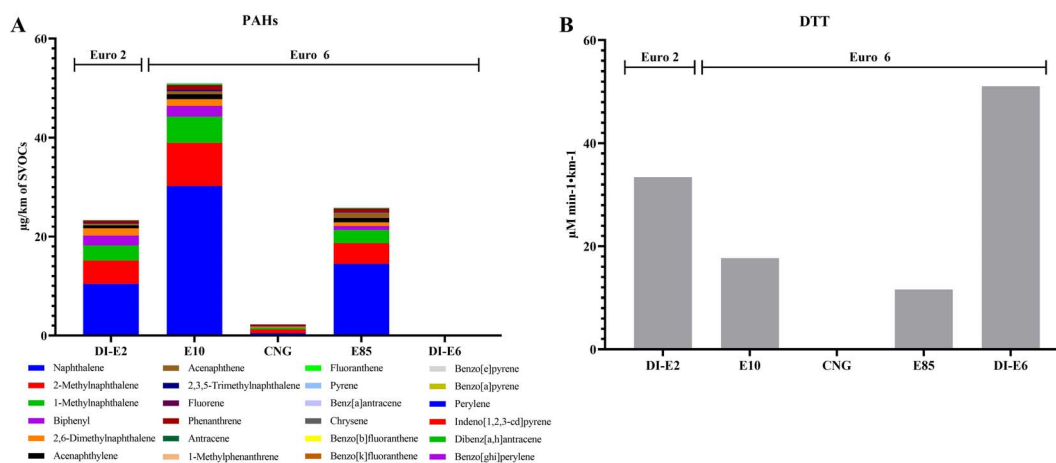


Fig. 1 Mass of SVOC-bound PAHs in diluted exhaust gas emitted during one kilometer of driving distance ($\mu\text{g km}^{-1}$) and oxidative potential of SVOCs from one Euro 2 and four Euro 6 engines. A: Mass concentrations of SVOC-bound PAHs; B: oxidative potential measured with DTT assay of SVOC samples.



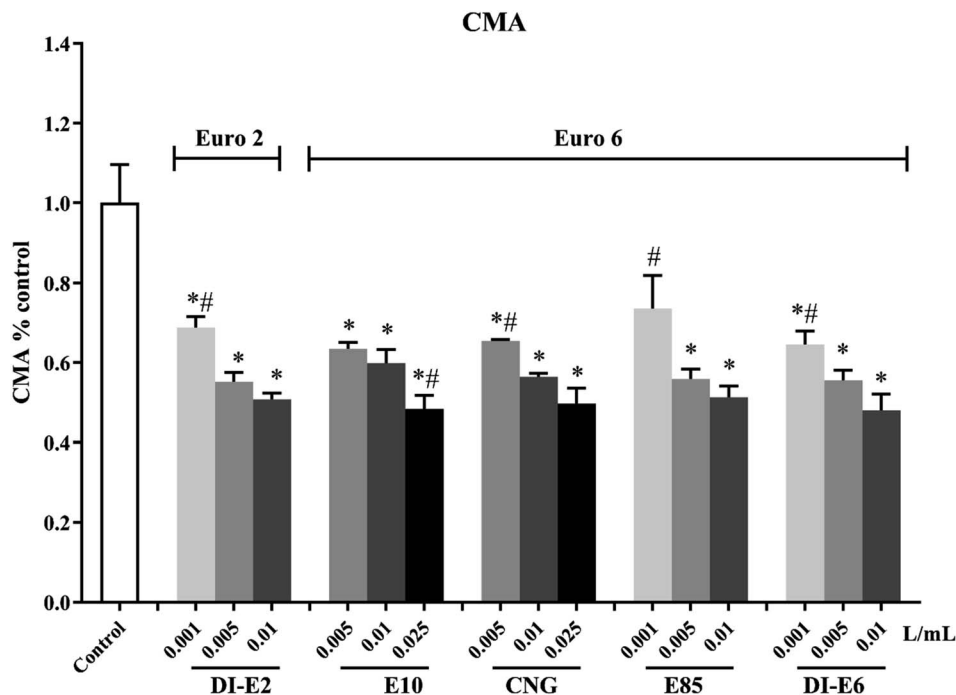


Fig. 2 Decrease of cellular mitochondrial activity (CMA) compared to control cells after RAW 264.7 macrophages exposure to exhaust gas volume-based doses of SVOC samples from one Euro 2 and four Euro 6 engines. * Significant differences between the exposure group and the control ($P < 0.05$). # Significant differences between the different exposure groups ($P < 0.05$).

higher NO_x , CO, total HCs, and CH_4 than E10. SVOCs from CNG emitted the least CO and CO_2 . The mass concentration of SVOC-bound PAHs in diluted exhaust gas and oxidative potential of SVOCs are shown in Fig. 1. SVOC-bound PAHs in E10, E85 and

DI-E2 were relatively high, with naphthalene being the most concentrated PAH. The oxidative potential of SVOCs followed this order: DI-E6 > DI-E2 > E10 > E85 > CNG.

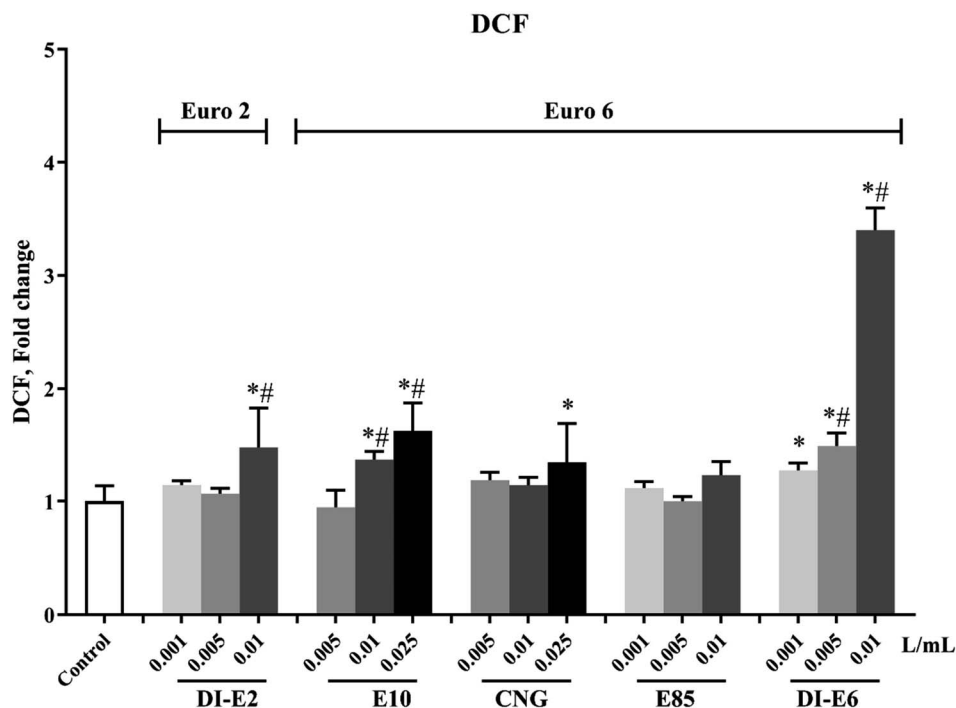


Fig. 3 Intracellular oxidative potential of RAW 264.7 macrophages after exposure to exhaust gas volume-based doses of SVOC samples from one Euro 2 and four Euro 6 engines. * Significant differences between the exposure group and the control ($P < 0.05$). # Significant differences between the different exposure groups ($P < 0.05$).



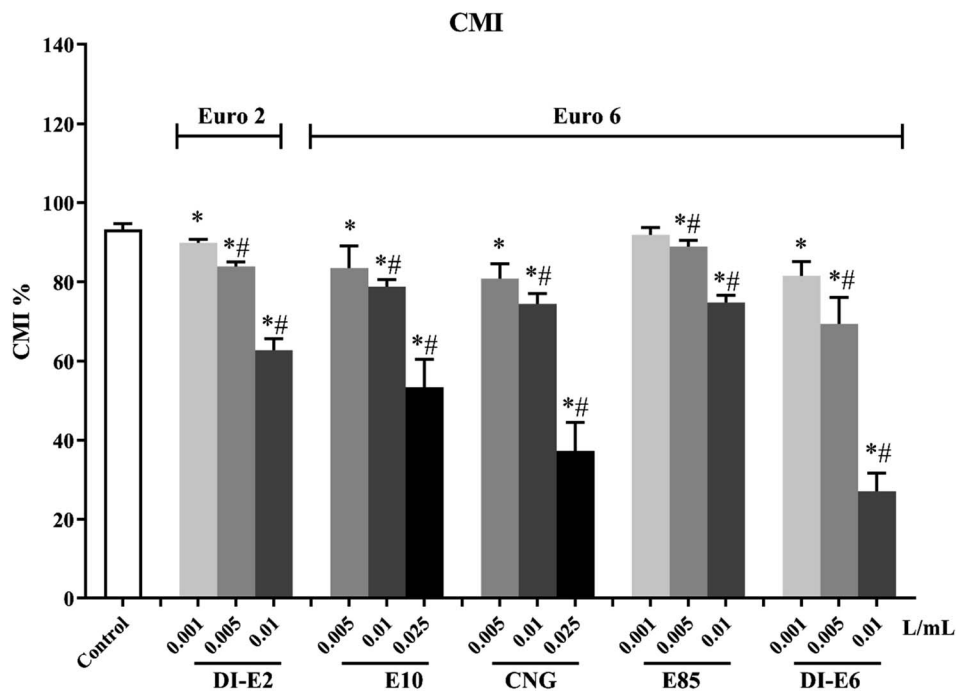


Fig. 4 Cell membrane integrity and vitality decrease of RAW 264.7 macrophages after exposure to exhaust gas volume-based doses of SVOC samples from one Euro 2 and four Euro 6 engines. * Significant differences between the exposure group and the control ($P < 0.05$). # Significant differences between the different exposure groups ($P < 0.05$).

3.2 CMA

The decrease in CMA compared to control cells after RAW264.7 macrophages were exposed to exhaust gas volume-based doses of SVOC samples is shown in Fig. 2. Exposure to SVOCs decreased all the levels of CMA. The 0.01 L mL^{-1} group of E85, DI-E2 and DI-E6 caused a similar level of CMA decline as the 0.025 L mL^{-1} group of E10 and CNG.

3.3 Intracellular oxidative potential and cell membrane integrity

The intracellular oxidative potential of RAW264.7 macrophages after exposure to exhaust gas volume-based doses of SVOC

samples is shown in Fig. 3. Exposure to E10 and DI-E6 samples induced a dose-dependent increase in intracellular oxidative potential, especially in the 0.01 L mL^{-1} group of DI-E6, where the level of intracellular oxidation potential was more than three times that of the control group.

The cell membrane integrity and vitality decrease of RAW264.7 macrophages after exposure to exhaust gas volume-based doses of SVOC samples are shown in Fig. 4. Almost all SVOC exposure affected cell membrane integrity. The highest dose, the 0.01 L mL^{-1} group of DI-E6, resulted in less than 30% membrane integrity, while the 0.025 L mL^{-1} group of CNG caused less than 40% membrane integrity.

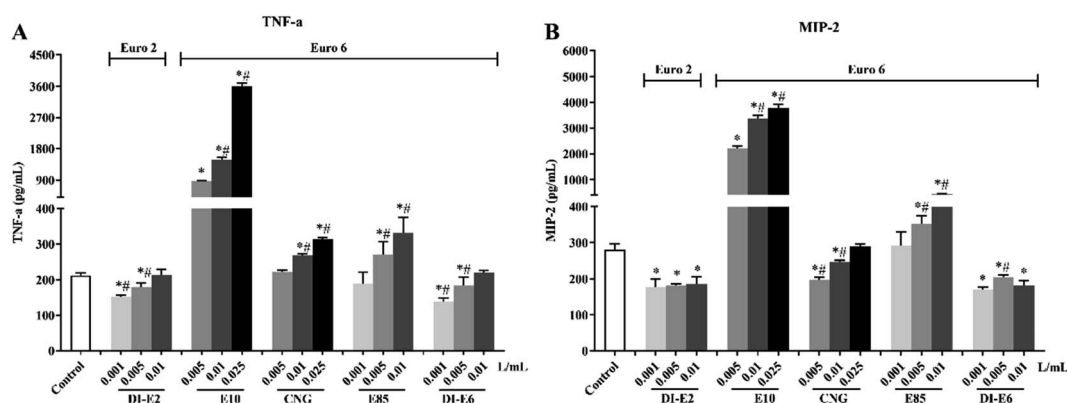


Fig. 5 Tumor necrosis factor alpha (TNF-α) and macrophage inflammatory protein 2 (MIP-2) production in RAW 264.7 macrophages after exposure to exhaust gas volume-based doses of SVOC samples from one Euro 2 and four Euro 6 engines. * Significant differences between the exposure group and the control ($P < 0.05$). # Significant differences between the different exposure groups ($P < 0.05$).



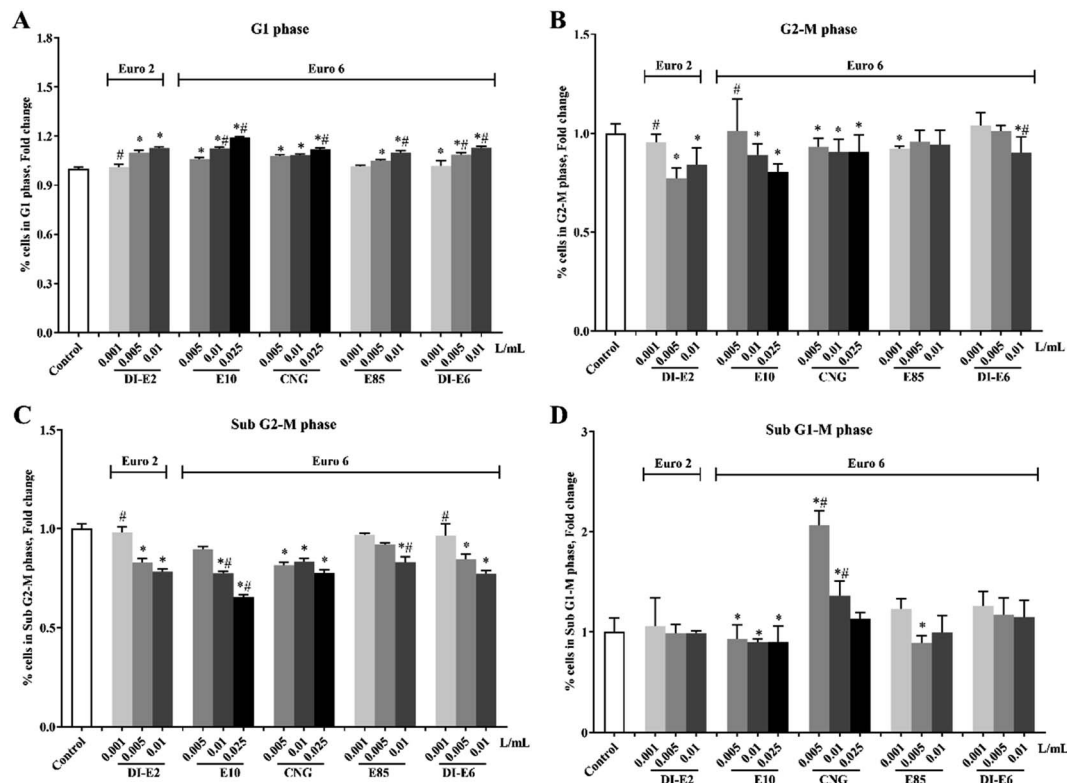


Fig. 6 Cells with DNA damage as measured by flow cytometric cell cycle phase analysis on RAW 264.7 macrophages after exposure to exhaust gas volume-based doses of SVOC samples from one Euro 2 and four Euro 6 engines. * Significant differences between the exposure group and the control ($P < 0.05$). # Significant differences between the different exposure groups ($P < 0.05$).

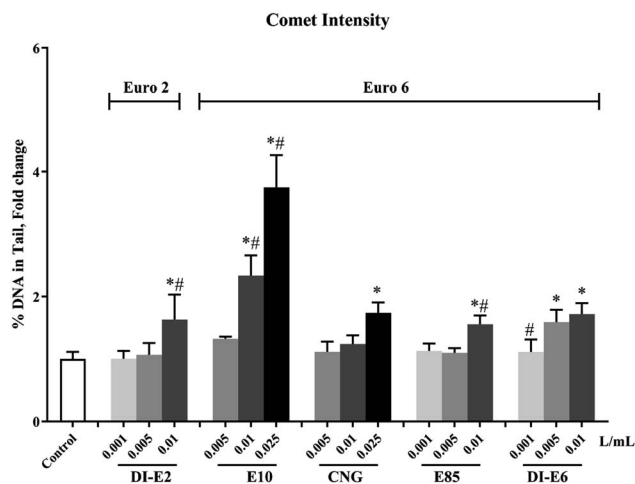


Fig. 7 Genotoxicity of RAW 264.7 macrophages after exposure to exhaust gas volume-based doses of SVOC samples from one Euro 2 and four Euro 6 engines. * Significant differences between the exposure group and the control ($P < 0.05$). # Significant differences between the different exposure groups ($P < 0.05$).

3.4 Inflammatory mediators

The production of TNF- α and MIP-2 in RAW264.7 macrophages after exposure to exhaust gas volume-based doses of SVOC samples is shown in Fig. 5. Exposure to E10, CNG and E85

exhaust caused increases in TNF- α production, especially in E10, while decreases in TNF- α levels were observed with DI-E2 and DI-E6 exposure. E10 and E85 exhaust exposure caused increases in MIP-2 levels, especially E10, whereas CNG, DI-E2, and DI-E6 exposure resulted in reductions in MIP-2 levels.

3.5 Cell cycle

Cells with DNA damage, as measured by flow cytometric cell cycle phase analysis on RAW264.7 macrophages after exposure to exhaust gas volume-based doses of SVOC samples, are shown in Fig. 6. Exposure to all the SVOCs resulted in an increased amount of DNA in the G1 phase and decreased DNA in the sub G2-M phase. Notably, the low-dose 0.005 and 0.01 L mL⁻¹ groups of CNG exhaust caused significant increases in the sub G1-M phase, and the 0.005 L mL⁻¹ group exhibited a higher level than the 0.01 L mL⁻¹ group, with no significant difference observed between the 0.025 L mL⁻¹ group and the control group.

3.6 Genotoxicity

Genotoxicity in RAW264.7 macrophages after exposure to exhaust gas volume-based doses of SVOC samples is shown in Fig. 7. Notably, the 0.01 and 0.025 L mL⁻¹ groups in E10, 0.025 L mL⁻¹ group in CNG, 0.01 L mL⁻¹ group in E85 and DI-E2, and 0.005 and 0.01 L mL⁻¹ groups in DI-E6 induced significant increases in DNA tail intensity, with E10 exhibiting the most pronounced effect. The increases in DNA tail intensity caused

by CNG, E85, DI-E2 and DI-E6 samples were less than twice that of the control group.

4. Discussion

SVOCs are typically present as vapors in engine exhaust and can readily attach to PM. The exhaust gas, composition and toxicological effects in PM from one Euro 2 regulated car and four Euro 6 emission regulated cars were also evaluated in previous study.⁵ With the exception of Euro 2 car DI-E2 and Euro 6 car DI-E6, the PAHs in SVOCs produced by E10, CNG, and E85 are much higher than those in PM. Furthermore, similar to the findings of this study, the total amount of PAHs in PM produced by DI-E2, E10 and E85 was also relatively high.⁵ PAHs are produced when insufficient oxygen or other factors cause incomplete combustion of organic matter, as observed in engines.² Diesel cars are known for emitting PAHs; however, it is noteworthy that gasoline cars may also emit high concentrations of PAHs, particularly at low temperatures.²¹ Euro 2 diesel fuel car DI-E2 lacked any exhaust gas aftertreatment system, which may explain the high PAH concentrations in PM and SVOCs emitted by DI-E2. Temperature might play a crucial role in PAH emissions from gasoline fueled cars E10 and E85. Diesel-fueled cars emit significant amounts of PM, especially ultrafine particles, which strongly adsorb PAHs. Although gasoline-fueled cars emit nearly the same amount of PM as diesel-fueled cars,^{22–24} vehicles using gasoline with high ethanol content have lower emissions.¹⁵ The reduction in PM emissions led to a decrease in the number of PM-bound PAHs, resulting in an increase in PAHs in SVOCs. Additionally, the content of PAHs in PM⁵ and SVOCs emitted by DI-E6 using diesel fuel was very low. This reduction may be attributed to the DPF and EGR systems equipped in the Euro 6 diesel car DI-E6, which effectively decreased both PM and PAH emissions in the exhaust gas.

DTT is commonly used to assess particle oxidative potential outside cell environments.²⁵ We found that despite the Euro 6 diesel vehicle DI-E6 being equipped with DPF and EGR systems, SVOCs emitted by it exhibited the highest oxidative potential compared to those emitted by all other vehicles. In contrast to the findings for PM, DI-E2 emitted PM with the highest oxidative potential, whereas DI-E6 emitted PM with the lowest oxidative potential.⁵ In PM, the trends in DTT, PM mass and number concentration, and PAHs emitted by each vehicle were found to be similar.⁵ The cleaning system equipped with DI-E6 reduced PM emissions and may further reduce the oxidative potential of PM emitted by DI-E6. However, in the SVOCs from the five vehicles in this study, the change in DTT was not consistent with any SVOC exhaust gas. The only similarity observed in this study was that SVOCs emitted by DI-E6 also caused a significant increase in intracellular oxidative potential, but SVOCs from other vehicles did not induce a similar change in intracellular oxidative potential. Previous studies have found that SVOCs might exert a dominant effect on the DTT response of these particle types after heating diesel or ultrafine particles in the environment.²⁶ However, we found that among SVOCs, PM and PM + SVOCs, the DTT in PM + SVOC was the highest (ESI Fig. A1†); especially the DTT in PM + SVOCs of the DI-E2 car

was significantly higher than that of SVOCs and PM (4–6 times). Some studies have found that DTT loss is associated with components such as metals and PAHs in PM, although many of these correlations were not causal.^{20,27} There were almost no metals in SVOCs, and the content of PAHs in DI-E6 was very low in this study. The high DTT of SVOCs from the DI-E6 car may be attributed to the limited presence of substances that oxidize DTT. Currently, we lack conclusive proof or a clear explanation for the variation in the oxidative potential of SVOCs emitted by different vehicles, necessitating further exploration.

In this study, although the amount of exhaust gas in the SVOCs emitted by DI-E2 was high, it did not cause strong cellular toxicological effects. In contrast, the oxidative and cell membrane damage induced by SVOCs emitted by DI-E6 were more serious. This may be related to the high DTT of DI-E6. PM emitted by DI-E2, characterized by high DTT, also led to a more significant expression of intracellular oxidative potential.⁵ The new emissions regulations for the Euro 6 passenger diesel car DI-E6 have efficiently reduced SVOC exhaust NO_x, CO, CO₂, HC, and PAH emissions. However, despite these reductions, oxidative potential and cytotoxicity remain in low-emission SVOCs in this study. Moreover, the NO_x levels in SVOCs emitted by DI-E6 were still significantly higher than those of other Euro 6 vehicles. The harmful effects of NO_x on humans have been confirmed by many studies.²⁸ Therefore, diesel vehicles subject to new emissions regulations still require additional methods to further improve their exhaust emissions.

E10 and E85 were both gasoline-fueled vehicles with ethanol in this study. The cytotoxicity caused by SVOCs emitted by E10 and E85 was similar, but E10 stimulated cells to secrete higher levels of inflammatory mediators and exhibited more severe genotoxicity. This difference is related to the PAH content in their SVOCs. Naphthalene is the predominant PAH in ambient air.²⁹ The exposure of naphthalene in rodents has been shown to cause nasal tract inflammation and nasal epithelium tumors.^{30–32} In this study, SVOCs from E10 had the highest levels of PAHs, particularly naphthalene, and induced the production of more inflammatory mediators. Previous research has also indicated that exposure to PM rich in PAH content results in elevated levels of inflammatory mediators.⁵ Several PAHs, including benzo(a)pyrene, with their metabolites being highly carcinogenic and mutagenic.² Therefore, exposure to SVOCs from E10 also caused the most significant genotoxicity in this study. This observation aligns with the results of Hakkarainen *et al.*,⁵ who reported a consistent trend between the mutagenicity in PM and the PAHs. Despite some exhaust gas components in the SVOCs emitted by E85 being higher than those of E10, the level of cell damage caused by SVOC exposure in E85 was still lower than that in E10 in this study. Therefore, the use of alternative fuels with high ethanol/gasoline ratios may potentially mitigate the adverse effects of vehicle exhaust on human health.

CNG is generated by compressing conventional natural gas, primarily composed of CH₄, to less than 1% of its original volume under standard atmospheric pressure.³³ In CNG engines, most of the HC emissions are attributed to CH₄.³⁴ This occurs due to incomplete combustion and the inefficiency of



the oxidation catalyst in effectively removing CH₄.³⁴ Therefore, in this study, the levels of CH₄ and HC in the SVOC exhaust gas from CNG engines were high. Currently, CNG is considered one of the best solutions to replace fossil fuels because of its inherent clean burning properties.^{35,36} Research has found that, compared with diesel engines, CNG engines emit nearly 50 times fewer carcinogenic PAHs, 20 times less formaldehyde, and more than 30 times less PM and experience significant reductions in NO_x, with genetic toxic activity reduced by up to 20–30 times.³⁷ While reducing HC emissions is essential, CNG vehicles exhibit significant advantages in exhaust emissions reduction compared to diesel and gasoline technologies.^{5,34} However, the environmental and climate impacts of high HC emissions from CNG engines may outweigh the benefits gained in terms of human health.

Previous studies have indicated that the toxicity of CNG exhaust gas, concerning reactive oxygen species, inflammatory biomarkerd (CYP1A1, IL8, and COX-2), and mutagenicity is very low.³⁸ Additionally, PM from CNG has been found to not cause cytotoxic effects and mutagenicity.^{5,13} However, in our study, SVOCs from CNG not only reduced cellular mitochondrial activity and damaged cell membrane integrity but also stimulated cells to secrete TNF- α , interfered with DNA replication in Sub-G1-M, and caused cellular genotoxicity.

Similarly, Seagrave *et al.*³⁹ found that, although the lung toxicity of CNG emission samples was lower than that of gasoline and diesel samples, its mutagenic potency was similar to that of gasoline and diesel emission samples. While most current studies on the toxicity of CNG engine emissions report negative or low toxicity results, the low emissions of CNG engines do not completely eliminate the toxicity of the organic compounds they emit. This is because, in addition to fuel combustion itself, engine lubricating oil is a significant source of SVOCs,⁸ and CNG engines cannot control the evaporation of engine lubricating oil, leading to the generation of SVOCs. Therefore, the toxicity of organic compounds emitted by CNG engines cannot be ignored.

5. Conclusions

Our study fills the gap in the chemical and toxicological characterization of SVOC emissions from passenger vehicles using different fuels under sub-freezing conditions. While new technology has successfully reduced the emissions of diesel vehicles, the toxicity of SVOCs emitted by them remains unchanged. The toxicity of SVOCs from CNG and ethanol/gasoline blended vehicles cannot be ignored.

Data availability

The data are not publicly available, but are available from the corresponding author upon reasonable request.

Author contributions

Mo Yang: conceptualization, methodology, validation, formal analysis, investigation, writing-original draft, writing-review

and editing, resources, data curation, supervision. Päivi Aakko-Saksa: methodology, validation, formal analysis, data curation. Henri Hakkarainen: methodology, writing-review and editing. Topi Rönkkö: methodology, validation, formal analysis, data curation. Päivi Koponen: methodology, validation, formal analysis, data curation. Xiao-Wen Zeng: writing-review and editing. Guang-Hui Dong: writing-review and editing. Pasi Jalava: conceptualization, methodology, validation, writing-original draft, writing-review and editing, funding acquisition, resources, supervision.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

The authors thank Piritta Roslund for her technical assistance. This work was supported by the European Union's Horizon 2020 research and innovation programme under grant agreement No. 814978 (TUBE) and Academy of Finland, grants: 287982, 294081 and 319245.

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