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# Total synthesis of micrococcin P1 and thiocillin I enabled by Mo(vi) catalyst†

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Thiopeptides are a class of potent antibiotics with promising therapeutic potential. We developed a novel Mo(vi)-oxide/picolinic acid catalyst for the cyclodehydration of cysteine peptides to form thiazoline heterocycles. With this powerful tool in hand, we completed the total syntheses of two representative thiopeptide antibiotics: micrococcin P1 and thiocillin I. These two concise syntheses (15 steps, longest linear sequence) feature a C–H activation strategy to install the trisubstituted pyridine core and thiazole groups. The synthetic material displays promising antimicrobial properties measured against a series of Gram-positive bacteria.

The surge in drug-resistant strains of bacteria poses a major global health threat and the need for expedient discovery of new therapeutics characterized by novel modes of action represents a high priority.<sup>1</sup> The thiopeptide family of antibiotics contains ~150 members of post-translationally modified peptides of ribosomal origin that display antibacterial activities against drug-resistant pathogens, including methicillin-resistant *Staphylococcus aureus* (MRSA).<sup>2</sup> All thiopeptide antibiotics share a common molecular scaffold involving a nitrogen-containing heterocyclic core decorated with varying numbers of thiazol(in)e rings, assembled into macrocycles or acyclic chains of varying sizes and lengths.<sup>2c</sup> The macrocycle, whose size is correlated with molecular targets, is responsible for distinct modes of action, allowing thiopeptides to inhibit bacterial cell growth by means of blocking ribosomal protein synthesis.<sup>2d</sup> Because of their unique biological activities and intriguing structure, thiopeptides have received considerable attention from the synthetic community.<sup>3</sup> However, the lack of concise, scalable, and practical syntheses amenable for rapid molecular modifications limits access to these valuable natural products. In this paper, we report a general approach to the synthesis of thiopeptide antibiotics featuring a novel Mo(vi) oxide catalyst to promote the cyclodehydration of cysteine residues under mild and neutral conditions and is demonstrated by the preparation of D series thiopeptides micrococcin P1 (**1**)<sup>3u,3w,4</sup> and thiocillin I (**2**),<sup>3x,5</sup> This practical and scalable route is suitable for analogue preparation and site-selective modifications.<sup>6</sup>

Our retrosynthetic analysis of the synthetic targets led to the dissection of the macrocycle into two “top” and “bottom” fragments, which could be united through a peptide coupling and then macrocyclised at the thiazole carboxylic acid adjacent to valine to furnish the final product(s) (Fig. 1). For construction of the two fragments, we envisioned a convergent approach involving the strategic introduction of selected thiazole rings through a catalytic cysteine cyclodehydration. Catalytic methods for cysteine cyclodehydrations are scarce despite the myriad obstacles encountered when employing common stoichiometric reagents due to their high Lewis acidity, likely incompatibility with acid-sensitive groups, possible epimerization, and generation of undesired by-products.<sup>7</sup> We hypothesized that a MoO<sub>2</sub>(L)<sub>2</sub> complex with the appropriate ligands might minimize epimerization of the endo- and exocyclic methylene groups since the cyclization event occurs under neutral conditions. Furthermore, bidentate ligands may extend the lifetime of the catalyst and prevent precipitation of heterogeneous molybdenum particles.<sup>8</sup>

To validate the feasibility of this proposal, we evaluated a series of Mo(vi) oxide complexes (Table 1). Initially, we studied

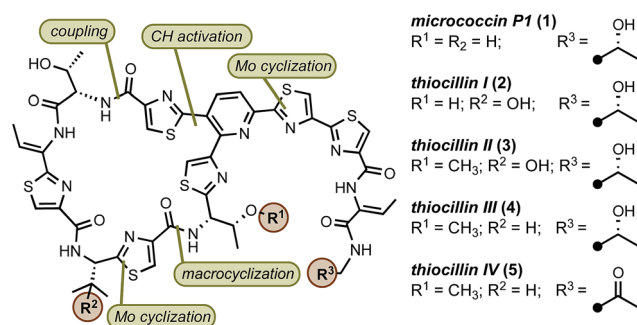
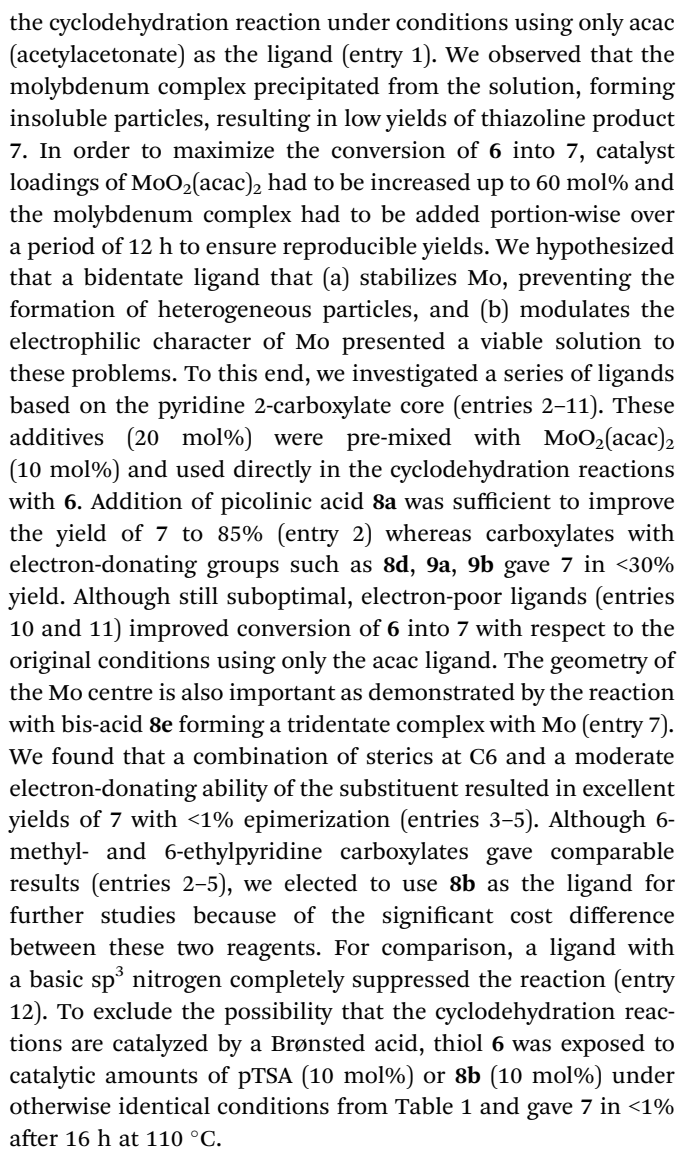


Fig. 1 Selected thiopeptide antibiotics of the D series.

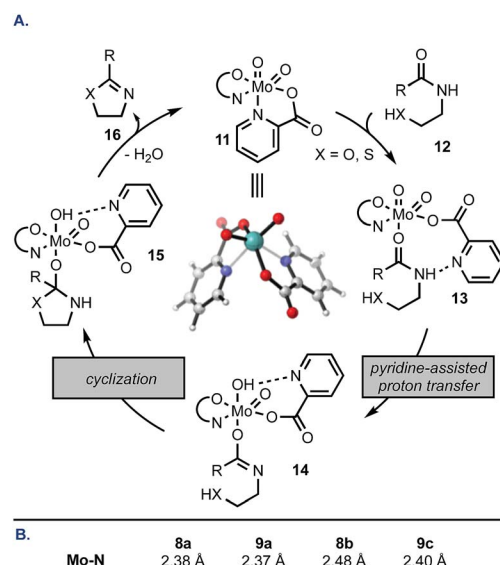
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Having established a robust method for the catalytic installation of select thiazole groups, we proceeded towards the preparation of bottom fragments **25** (Scheme 2). The optimized



**Scheme 1** (A) Proposed mechanism of Mo-catalysed cyclo-dehydration (second carboxylate ligand abbreviated for clarity). (B) Selected geometrical parameters of  $\text{MoO}_2(\text{X})_2$  complexes optimized at B3LYP/6-31G(d)/LANL2DZ level of theory. The optimized structure of the thermodynamic complex of  $\text{MoO}_2(\text{pic})_2$ , pic = 2-picolinate is presented in the middle of the catalytic cycle.

cyclodehydration conditions were applied to two dipeptides **17** followed by oxidation to thiazoles **18** in excellent yields for both substrates. The remaining portion of the bottom fragment was prepared *via* a modified Hantzsch method using thioamide **19** and bromopyruvate **20** in 94% yield. Coupling of **21** with serine **22** followed by a stereoselective (*dr* > 99 : 1) installation of the trisubstituted olefin with *MsCl* and a base (*DABCO*, *Et*<sub>3</sub>*NH*) resulted in thiazole **24**. With this key precursor in hand, the amine and carboxylate groups in **18** and **24** were liberated and the subsequent amide couplings resulted in gram-scale syntheses of bottom fragments **25** in seven steps for both micrococcin P1 and thiocillin I in 47% and 51% overall yield, respectively.

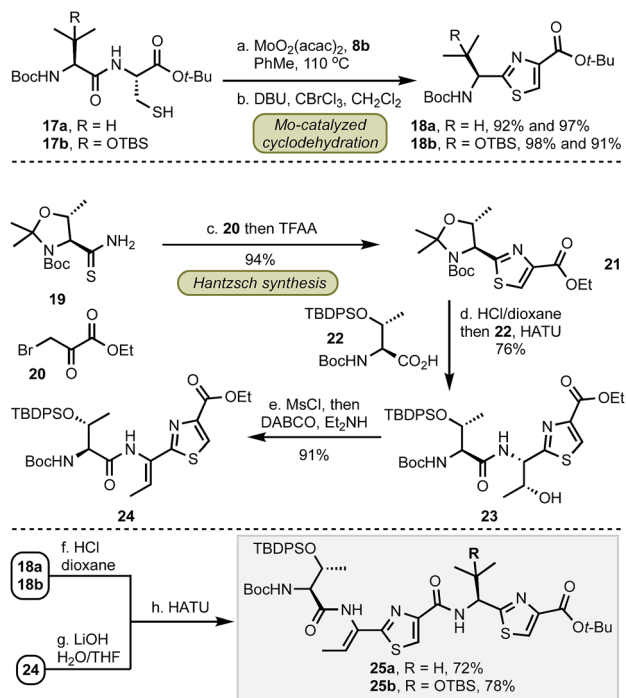
The synthesis of top fragment **37**, common to D series thiopeptides, started from a Pd-catalyzed C–H activation of thiazole **26** (Scheme 3). We found that the yield of this reaction was critically dependent on the size of the ester group and the methyl group was incompatible with the cross-coupling conditions. The reaction of ethyl thiazole ester **26b** with *Pd(OAc)*<sub>2</sub> (5 mol%) and *CyJohnPhos* (15 mol%) gave, after conversion from **28a**, methyl ester **28b**<sup>9</sup> which was then coupled with amine **30** following the two-step, selective chlorination of the pyridine core (97%). At this stage, the sulfur protective group was removed, and *MoO*<sub>2</sub>(*acac*)<sub>2</sub>-catalyzed cyclodehydration and oxidation afforded **32** in 86% yield. We found that the pyridine

scaffold in **31** was sufficient to stabilize the *Mo(VI)* catalyst and prevented decomposition of the *Mo* complex in the cyclodehydration reaction thus alleviating the need of further stabilization by additional ligands. Next, the Stille coupling with ethyl vinyl ether **33** followed by bromination with *NBS* afforded bromoketone **34**. Inspired by the prior work of Williams,<sup>10</sup> we converted **34** into thiazole **36** in 86%. Because the exocyclic methylene group in the Hantzsch intermediate can undergo epimerization, we optimized the cyclization step by buffering the conditions with a bulky base (2,6-lutidine) whereas larger bases (*e.g.*, 2,6-di-*t*-butyl-pyridine) were ineffective in promoting this transformation. To confirm the optical purity of the newly formed thiazole **36**, the carbamate was removed, and the resultant amine was converted into a Mosher amide giving a 95 : 5 *dr* (determined by <sup>19</sup>F NMR). Finally, site-selective hydrolysis of the methyl ester in **36** proceeded without concomitant removal of the *i*-Pr group to yield top fragment **37** in 73%.<sup>31</sup>

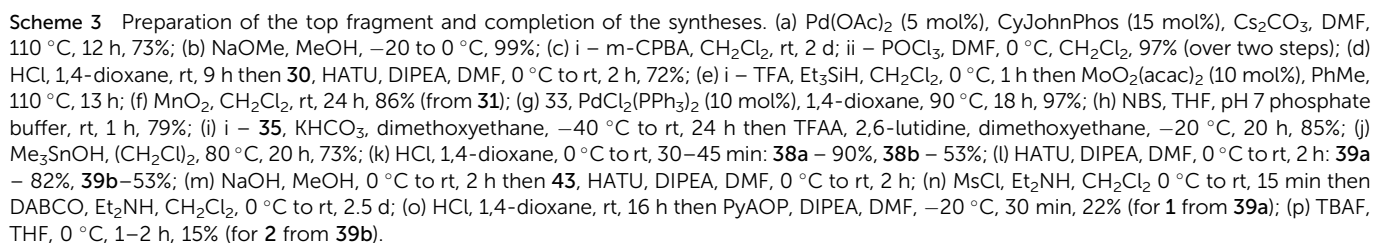
In preparation for the union of the two fragments, the *Boc* protective group was removed from bottom segments **25** to yield free amines **38** which were then coupled with top unit **37** to afford oligopeptides **39**. Next, the *i*-Pr ester in **39** was removed under more forcing conditions (*NaOH/H*<sub>2</sub>*O*), and the resultant acid was merged with dipeptide **43**. The installation of the olefin was accomplished by elimination of the hydroxyl group in threonine with *MsCl* in exclusive *Z* selectivity. The final steps of the synthesis involved global deprotection with *HCl* to cleave all alcohol, amine, and carboxylic acid protective groups in one step, followed by macrocyclization of the resulting amino acid intermediate with *PyAOP* furnished micrococcin P1 **1** in 22% yield. The similarity of micrococcin P1 and thiocillin I, which differ only by a mutation of *L*-valine into 3-hydroxy-*L*-valine, led us to adopt identical conditions for both syntheses, with the exception of the deprotection for thiocillin I. We found that acidic conditions alone were insufficient and resulted in only partial cleavage of the silicon group at the hydroxyvaline position (*ca.* 30%); thus, following the final macrocyclization, the resultant precursor was subjected to *TBAF* to reveal thiocillin I (**2**) in 15%. Micrococcin P1 was completed in 14 steps and thiocillin I was completed in 15 steps from known commercial materials (longest linear sequence).

Having access to two representative members of the thiopeptide family, we evaluated the *in vitro* antimicrobial activity against a panel of recent Gram-positive bacterial strains (Table 2). Micrococcin P1 and thiocillin I exhibited similar potencies, showing 2- to 4-fold difference in their minimum inhibitory concentration (MIC) values in nearly all of the test bacterial isolates. An exception to this trend was observed for *B. subtilis* ATCC 6633, where micrococcin P1 was inactive at 16 μg mL<sup>−1</sup> while thiocillin I displayed activity at concentrations as low as 4 μg mL<sup>−1</sup>, a result comparable to a previously published report showing thiocillin I as active at 1.56 μg mL<sup>−1</sup> against *B. subtilis* PCI 219.<sup>5</sup> Additionally, micrococcin P1 and thiocillin I were more potent than comparator antibiotics against vancomycin-resistant *E. faecalis*.

In summary, we described a general synthetic route toward thiopeptide antibiotics belonging to the D series. Given the



**Scheme 2** Synthesis of bottom fragments **25**. (a) *MoO*<sub>2</sub>(*acac*)<sub>2</sub> (10 mol%), **8b** (20 mol%), PhMe, 110 °C, 2.5 h: **18a** – 92%, **18b** – 98%; (b) DBU, *CBRCl*<sub>3</sub>, *CH*<sub>2</sub>*Cl*<sub>2</sub>, 0 °C, 1 h: **18a** – 97%, **18b** – 91%; (c) **20**, *NaHCO*<sub>3</sub>, THF, 0 °C to rt, 16 h then pyridine, TFAA, 0 °C, 20 min, 94%; (d) *HCl*, 1,4-dioxane, rt, 7 h then **22**, HATU, DIPEA, DMF, 0 °C to rt, 1 h, 76%; (e) *MsCl*, *Et*<sub>3</sub>*N*, *CH*<sub>2</sub>*Cl*<sub>2</sub>, 0 °C, 1 h then *DABCO*, *Et*<sub>3</sub>*NH*, 0 °C to rt, *CH*<sub>2</sub>*Cl*<sub>2</sub>, 16 h, 91%; (f) *HCl*, 1,4-dioxane, 0 °C to rt, 45 min; (g) *Bu*<sub>2</sub>*SnO*, MeOH, 80 °C, 4 h then *LiOH*, *H*<sub>2</sub>*O*/THF, 90% (over two steps); (h) HATU, DIPEA, DMF, 0 °C to rt, 2 h: **25a** – 72%, **25b** – 78%.



|                    |            | MIC ( $\mu\text{g mL}^{-1}$ ) |     |            |               |
|--------------------|------------|-------------------------------|-----|------------|---------------|
| Isolate            |            | 1                             | 2   | Vancomycin | Ciprofloxacin |
| <i>S. aureus</i>   | ATCC 29213 | 4                             | 2   | 1          | 0.5           |
|                    | 1974149    | 2                             | 2   | 0.5        | 0.25          |
|                    | 1974148    | 8                             | 2   | 1          | >16           |
| <i>E. faecalis</i> | 1674621    | 1                             | 0.5 | 0.5        | 1             |
|                    | 1674614    | 1                             | 0.5 | >16        | >16           |
| <i>B. subtilis</i> | ATCC 6633  | >16                           | 4   | 0.25       | 0.06          |
| <i>S. pyogenes</i> | 1744264    | 1                             | 0.5 | 0.5        | 0.25          |

for stoichiometric reagents and generates water as the only by-product. Moreover, the modular approach lends itself into future SAR studies to attain compounds with desirable properties for pre-clinical and clinical development.

There are no conflicts to declare.

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