



Cite this: *Nanoscale Horiz.*, 2024, 9, 1683

Received 11th March 2024,
Accepted 8th July 2024

DOI: 10.1039/d4nh00107a

rsc.li/nanoscale-horizons

Luminescent carbon dots *versus* quantum dots and gold nanoclusters as sensors†

J. S. Anjali Devi,^{ib} *^{abc} S. Madanan Anju,^a G. M. Lekha,^a R. S. Aparna^a and Sony George^{ib} *^a

Ultra-small nanoparticles, including quantum dots, gold nanoclusters (AuNCs) and carbon dots (CDs), have emerged as a promising class of fluorescent material because of their molecular-like properties and widespread applications in sensing and imaging. However, the fluorescence properties of ultra-small gold nanoparticles (i.e., AuNCs) and CDs are more complicated and well distinguished from conventional quantum dots or organic dye molecules. At this frontier, we highlight recent developments in the fundamental understanding of the fluorescence emission mechanism of these ultra-small nanoparticles. Moreover, this review carefully analyses the underlying principles of ultra-small nanoparticle sensors. We expect that this information on ultra-small nanoparticles will fuel research aimed at achieving precise control over their fluorescence properties and the broadening of their applications.

^a Department of Chemistry, School of Physical and Mathematical Sciences, University of Kerala, Thiruvananthapuram 695581, Kerala, India.
E-mail: anjalidevijs@kannuruniv.ac.in, anjalidevijs@gmail.com, emailtosony@gmail.com; Tel: +91 8547481911, +91 9446462933

^b School of Chemical Sciences, Mahatma Gandhi University, Priyadarsini Hills P. O., Kottayam 686560, Kerala, India

^c Department of Chemistry, Kannur University, Swami Anandatheertha Campus, Payyanur, Edat P. O. Kannur 670327, Kerala, India

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4nh00107a>



J. S. Anjali Devi

J. S. Anjali Devi obtained her MSc in Chemistry from the University of Kerala with first rank and gold medal in 2013. Subsequently, she joined the Department of Chemistry, University of Kerala for her PhD under the supervision of Dr Sony George with a KSCSTE Research Fellowship 2013 (Govt. of Kerala), and DST-INSPIRE Fellowship 2015 (Govt. of India). She obtained her PhD degree in 2020. The thesis is entitled "Investigation on Photoluminescence Behaviour of Carbon Dots and Gold Nanoclusters for Sensing and Imaging Application." Then she joined Mahatma Gandhi University, Kerala as an Assistant Professor (Contract Faculty) from March 2021 to August 2023. Currently, she is working as an Assistant Professor at the Department of Chemistry, Kannur University, Kerala. She has 25 publications to her credit in international journals. Her research interests include physical chemistry, sensors, and photophysics.



S. Madanan Anju

S. Madanan Anju completed her MSc in Chemistry from the University of Kerala with the second rank in 2012. She did her MPhil Degree at the University of Kerala. She has qualified State Eligibility Test (Govt. of Kerala) and the National Eligibility Test (CSIR-UGC, Govt. of India). Currently, she is pursuing her Doctoral degree at the Department of Chemistry, University of Kerala under the guidance of Dr Sony George. She has over 30 publications to her credit in international journals. Her research interests include luminescent nanomaterials, cardiac biomarkers, explosive detection, and disease biomarker sensing.

1. General overview

Luminescence is the spontaneous emission of electromagnetic radiation, especially light waves, by substances in an electronic excited state.^{1,2} The word ‘luminescence’ derives from two Latin words, *lumen* (meaning ‘light’) and *escentia* (meaning ‘the process of’).³ The concept of luminescence is very different from incandescence so that the latter term deals with radiative emission from a hot substance. Luminescence is formerly classified by the difference in energy source as photoluminescence, cathodoluminescence, chemiluminescence, electroluminescence, mechanoluminescence, radioluminescence and thermoluminescence [Fig. S1 in ESI†].

Luminescence covers a wide array of topics. Fig. S1 (ESI†) display the depth of this overall topic. Thus, it is not practical to provide extensive coverage of all developments and features of this extremely broad area of research. Instead, we attempt to focus on fluorescence that arises from photon-induced excitation and emission.

Fluorescence refers to the spontaneous emission of radiation from an electronically excited state of a fluorescent species. It occurs through the electronic transition from the excited state to the ground state of the same spin multiplicity (Fig. S2, ESI†).^{1,2} The first report on fluorescence can be traced back to the observation of peculiar blue light in the infusion of a *Lignum nephriticum* (Latin for “kidney wood”) by Nicolás Monardes in 1565 and Franciscan Friar Bernardino de Sahagún in 1560.² This blue fluorescence is due to oxidation of flavonoids in “kidney wood” to highly fluorescent matlaline (from the Aztec word ‘matlali’ for blue) with an emission wavelength close to 466 nm.² Each subsequent innovation in fluorescence added wonderful features to the fluorescence signal, including intensity, lifetime, large wavelength separation (Stokes shift) between absorbing and emitting radiation as well as sensitivity to surrounding environmental factors.^{1,4} In many cases, fluorescence can be influenced by physicochemical aspects of the local environment (e.g.: viscosity, polarity, temperature, redox status, or pH) and interactions with quenchers.⁵ The interaction between a fluorophore and quenchers occurs *via* Förster resonance energy transfer (FRET), photoinduced electron transfer (PET), inner filter effect (IFE), aggregation, Dexter-type interaction, excimer formation or complex formation.^{4,6} These favourable features of fluorescence laid the foundations for fluorescent sensors which rely on a sensitive fluorescence response to specific analytes.⁷ To overcome the limitations of conventional organic-molecule-based fluorophores, tremendous attempts have been made to discover alternative fluorescent materials with a large Stokes shift, long lifetime, high biocompatibility, water solubility and so on. Every year, new evidence of fluorescent materials appears and interest in investigating the origin of fluorescence has risen. Owing to this intense evolution, fluorescence techniques using ultra-small nanoparticles have been developed as a powerful tool for analysis (see Fig. 1).^{8,9}

In this article, we summarise luminescent ultra-small nanoparticle sensors, especially those smaller than 10 nm, as they can be absorbed or excreted quickly by the human body but can



G. M. Lekha

and applications.’ Her research interests include magnetic and luminescent lanthanide-based nanoparticles.

G. M. Lekha received an MSc (2007), and PhD (2016) from the University of Kerala. In the year 2019, she completed a post-doctoral fellowship in the group of Dr Sony George at the University of Kerala. Since 2022, she has been a higher secondary teacher at MVHSS, Arumanoor, Kerala. She is the co-author of 7 publications in international journals, and a chapter in a book entitled ‘colloidal metal oxide nanoparticle synthesis, characterization,



R. S. Aparna

national journals. Her research interests include nanoscience, sensors, and bioimaging.

R. S. Aparna obtained her MSc in chemistry at the National Institute of Technology (NIT), Trichy, India. She pursued her PhD under the supervision of Dr Sony George at the Department of Chemistry, University of Kerala. She obtained her PhD degree in 2019. The thesis is entitled ‘Design and Development of Functionalized Copper Nanoclusters for Fluorescent Sensing and Imaging’. She has 22 publications to her credit in inter-



Sony George

sabbatical stint at the Department of Chemical Physics, LUND University, Sweden.

Sony George received his PhD at the Department of Chemistry, University of Kerala, India and he is currently working as an Associate Professor in the same institution. His area of interest is the design and development of magnetic and fluorescent nanomaterials for sensing and bioimaging. He had been a visiting professor at the A. A. Athinoula Centre for Biomedical Imaging, MGH affiliated to Harvard Medical School. He had a



Fig. 1 Number of publications on ultra-small nanoparticles based on Pubmed with the keyword "(ultrasmall OR carbon dots OR Gold nanocluster) AND sensing" in "Title/Abstract" section during the period 2003 to 2024 (data collected on 3 February 2024); total: 1648 results.

be circulated in the blood for a longer time.⁸ To date, many luminescent ultra-small nanosensors have been proposed for various applications. These materials include semiconductor nanocrystals (also known as quantum dots), metal nanoclusters (especially gold nanoclusters (AuNCs)) and carbon dots (CDs).^{8,10} Their photophysical properties, fluorescence origin and structural elucidation in previous reports have been given prime importance in this article. We summarise the use of these nanosensors to detect metal ions, neurotransmitters, biomarkers, biomolecules, pesticides, pH, temperature, polarity, viscosity, and hypoxia. Finally, the prospects and challenges in this rapidly maturing field are addressed.

1.1. How do they fluoresce?

Many luminescent materials exhibit two types of emission: downconversion and upconversion.⁸ Downconversion or Stokes shifted emission is the normal process in which particles absorb high-energy radiation and emit at lower energy. On the other hand, fluorescence upconversion occurs *via* an anti-Stokes phenomenon in which a high-energy photon is emitted through simultaneous multiple low-energy photon absorption or energy transfer processes.^{1,8} The Stokes shift is named after George Gabriel Stokes, who coined the term fluorescence in the 19th century.² The origin of the Stokes shift in conventional organic dyes can be illustrated simply with the aid of an energy level diagram known as the Jablonski diagram (Fig. S2, ESI†).¹ Many ultra-small nanoparticles exhibit upconversion fluorescence.^{11–13} Luminescent ultra-small nanoparticles have achieved larger Stokes shifts than conventional organic dyes. However, the detailed mechanism for fluorescence from ultra-small nanoparticles has not yet been completely understood.^{10,12,14}

Size-dependent fluorescence is widely observed in many ultra-small nanoparticles, especially semiconductor nanocrystals (quantum dots). Quantum dots are nanocrystals smaller than the typical Bohr exciton radius. At this nano-dimension, their energy levels become quantised with values directly related to the size of the quantum dots (Fig. 2).^{15–17} This phenomenon is called the quantum confinement effect. The absorption of a photon with energy above the semiconductor bandgap will trigger the generation of electron-hole pairs

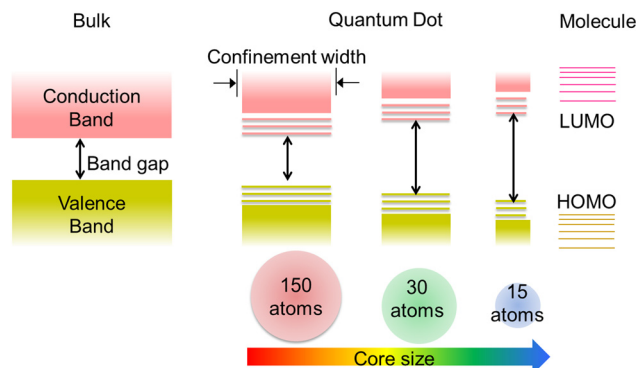


Fig. 2 Evolving quantum confinement effect within the energy level structure of semiconductor materials on tuning the dimensions from bulk to nano-dimensional. The bulk semiconductor possesses continuous conduction and valence bands separated by a constant bandgap. The limited number of atoms in quantum dots results in discrete 'molecule-like' energy levels depending upon the size of the particles. The quantum dots bridge the gap between cluster molecules and bulk materials.

(excitons). The subsequent radiative recombination of excitons results in narrow and symmetric fluorescence emission with a long lifetime.^{15,17}

CDs are widely visualised as quantum dots made of carbon.¹² That is why the quantum confinement effect in CDs has been comprehensively investigated over the last decade.^{12,18} Surprisingly, much research work has provided evidence of size-dependent fluorescence in CDs.^{19,20} Most importantly, Lee and co-workers reported bright blue, green, yellow and red fluorescence from CDs with increasing size.¹⁹ In another case, Lin and co-workers prepared green, blue and red luminescent nitrogen-doped carbon dots (NCDs) from *ortho*-, *meta*- and *para*-isomers of phenylenediamines.²⁰ They suspected that particle size and nitrogen content controlled the fluorescence colour of the CDs. From these experimental observations, researchers found some correlation between CD fluorescence and the quantum confinement effect in graphene oxide (Fig. 3). As a result, the theory of a bandgap transition within numerous isolated sp^2 -hybridised carbon nanodomains embedded on an sp^3 -hybridised carbon matrix of CDs has gained a lot of research attention.¹² According to this model, a single CD particle possesses numerous conjugated π -domains containing sp^2 -hybridised carbon clusters at its core (*i.e.*, the CD core is similar to the graphene oxide structure) and numerous polar surface functional groups of sp^3 -hybridised carbon. The fluorescence is the result of the quantum confinement effect. In contrast, researchers get confused when observing identical visible light fluorescence from CDs of ultra-small size as well as larger CDs with dimensions of several tens of nanometers.²¹ Subsequently, Eda *et al.* confirmed the probable presence of isolated sp^2 -hybridised carbon nanodomains within an sp^3 carbon matrix of CDs by theoretical calculations using Gaussian and time-dependent density functional theory (TD-DFT).²² They calculated that isolated π -domains within a carbon core of 3 nm can accommodate more than 100 aromatic benzene rings in such a way that CDs of this dimension should have a bandgap of only



Fig. 3 Scheme representing the electronic band structure of graphene oxide. The pink and blue bands indicate conduction and valence bands, respectively. Smaller sp^2 -hybridised carbon nanodomains exhibit a large bandgap due to the quantum confinement effect. The absorption of photons (E_{exc}) generates electron-hole pairs which on subsequent radiative recombination yield fluorescence (E_{PL}). The black arrows symbolise electronic transitions. DOS represents the density of states.

0.5 V. This calculated value of the bandgap could not provide emission in the visible region.

Metal nanoclusters are known as metal quantum clusters and a special case of the quantum confinement effect happens in certain metal nanoclusters (Fig. 4).²³ In particular, poly-amidoamine (PAMAM)-dendrimer-stabilised atomically precise AuNCs exhibit size-dependent fluorescence.^{24,25} This experimental evidence led to the formulation of a simple Jellium scaling law:

$$E_{\delta} = \frac{E_{Fermi}}{N^{1/3}}$$

where E_{δ} is the energy gap between the highest occupied molecular orbital (HOMO) and the lowest unoccupied molecular

orbital (LUMO) of AuNCs, E_{Fermi} is the Fermi energy of bulk gold and N is the number of gold atoms.²⁵ This model of PAMAM-protected AuNCs can be successfully applied for many other AuNCs protected by pepsin, bovine serum albumin or thiolate.^{26,27} However, the Jellium model holds good only for small AuNCs and it is violated for AuNCs with a large number of gold atoms as well as in the case of glutathione-stabilised AuNCs.^{14,28} To find the proper fluorescence mechanism of AuNCs, Zhu and coworkers performed TD DFT calculations on $[Au_{25}(SH)_{18}]^{-}$ clusters.²⁹ The results obtained suggest that a HOMO and three LUMOs are formed from the 6sp atomic orbitals of gold. Therefore, these molecular orbitals are collectively termed the sp band. On the other hand, HOMO-1 to HOMO-5 orbitals come from the 5d atomic orbitals of gold and these molecular orbitals are termed the d band. Thus they concluded that the electronic transition in $[Au_{25}(SH)_{18}]^{-}$ clusters is an intraband (sp $\rightarrow$ sp) HOMO-LUMO transition or an interband (d $\rightarrow$ sp) transition.

1.2. Excitation wavelength dependence of fluorescence

The observation of excitation-wavelength-independent fluorescence emission in ultra-small nanoparticles is a very common phenomenon following the classical Kasha-Vavilov rule.³⁰ This rule states that in any condensed phase (*i.e.* liquid or solution) fluorescence emission in appreciable yield occurs only from the lowest excited state of a given multiplicity (singlet or triplet) irrespective of the initial photoexcited state (see Fig. S2, ESI†).³⁰ Consequently, the fluorescence quantum yield should remain independent of excitation wavelength. An excellent demonstration of this law can be seen in many quantum dots, AuNCs protected by proteins or small molecules^{14,31,32} and a few CDs.^{33,34}

The fluorescence of CDs is more complicated than that of typical semiconductor quantum dots or AuNCs. Though there is a huge difference in the size and surface state of CDs prepared through different methods, excitation-dependent fluorescence emission is a common feature of CDs except for a few cases. However, the mechanism behind this phenomenon remains a topic of debate. Initially, this excitation-dependent fluorescence is correlated to variation in the quantum confinement effect due to different-sized particles present in the ensemble used for recording the fluorescence spectrum (Fig. 5).³⁵ Subsequently, several reports projected that different surface states or molecular states on a single particle, doping with an electronegative heteroatom, the synergic effect of quantum size, surface state and molecular state as well as solvent relaxation could have roles in multiple fluorescence centres involved in an excitation-dependent emission process. A comprehensive explanation of this matter can be found in the review article from 2016 by Gan *et al.*²¹

1.3. Factors affecting the fluorescence of ultra-small nanoparticles

It is well known that several parameters, including viscosity, polarity, temperature, redox status, pH and an adjacent heavy atom, can influence the photophysics of ultra-small



Fig. 4 Representations of a gold ring, a gold nanoparticle, a gold nanocluster solution in a cuvette under UV illumination and visible light, as well as a metallic gold atom, are given (from right to left) in the top panel. Bottom panel: bulk gold (as in a gold ring) and gold nanoparticles exhibit continuous conduction and valence energy bands (right). The limited number of gold atoms in gold nanoclusters results in discrete 'molecule-like' energy levels, a quantum confinement effect, and size-dependent fluorescence. Gold nanoclusters act as a 'missing link' between atomic gold and plasmonic gold nanoparticles.



Fig. 5 Schematic representation of surface-defect-state-derived excitation-independent fluorescence and different-sized sp^2 -hybridised carbon nanodomain-controlled excitation-dependent fluorescence in carbon dots.



Fig. 6 Scheme illustrating various environmental factors influencing the fluorescence performance of ultra-small nanoparticles.

nanoparticles (Fig. 6). Abnormal changes in these factors may affect the quality and fluorescence performance of ultra-small nanoparticles. Here, we outline the various environment-responsive fluorescent ultra-small nanoparticles that have been reported so far.

1.3.1. Viscosity. The viscosity of the dispersing medium will control the diffusion of ultra-small nanoparticles in the specified medium.⁵ Microviscosity (η) is related to the diffusion coefficient (D) and hydrodynamic radius (R_H) of particles *via* the Stokes-Einstein relationship:³⁶

$$\eta = \frac{k_B T}{6\pi R_H D}$$

where k_B is the Boltzmann constant and T is the absolute temperature. With an increase in the viscosity of the solvent, the diffusion and collision rate of a fluorophore will decrease to show less collisional non-radiative deactivation and enhanced fluorescence intensity. Using this principle, ultra-small nanoparticles of BSA-capped AuNCs have been proposed as a good

microviscosity-targeting sensor in single breast cancer cells (MCF7).³⁶ These authors found that the viscosity of breast cancer cells is much higher than that of normal breast cells. Based on the same principle, Mandal and co-workers used viscosity as a tool to determine the location of emissive species within the CD matrix.³⁷

1.3.2. Temperature. The fluorescence performance of a selected ultra-small nanoparticle for a particular application can be controlled by its fluorescence stability over the desired temperature range. With an increase in temperature, non-radiative transition pathways will be activated and this process will adversely affect the fluorescence intensity.¹ Yu and co-workers investigated the temperature dependence of fluorescence from quantum dots, AuNCs and CDs to gain insight into the secrets of their fluorescence.^{38–40} These experimental investigations established a decrease in fluorescence intensity with a rise in temperature. Among these interrogations, the fluorescence of AuNCs undergoes a small blue shift with rising temperature. In contrast, CDs exhibit a small red shift. On the other hand, the fluorescence band of semiconductor quantum dots shows a large red shift in the order of 100 meV due to strong exciton-phonon scattering. Moreover, from these results it is speculated that the π -electron-mediated fluorescence transition in CDs is closer to the electron transition in metallic quantum clusters rather than the quantum-confined electronic transition in semiconductor quantum dots.⁴⁰

1.3.3. Polarity. The optical properties of ultra-small nanoparticles can be strongly influenced by the local environment, including the polarity of the dispersing solvent. For instance, Leatherdale and Bawendi reported on the solvatochromic shift in the absorption spectrum of cadmium selenium quantum dots.⁴¹ This observation is made from the polarization energy of the quantum-confined exciton.

Few papers have been devoted to describing solvent effects on the photoluminescence of AuNCs. Typically, Chen and co-workers were the pioneer group who investigated atomically precise Au₈NCs formed by the etching of gold nanocrystals with biomolecules like deoxyribonucleic acid (DNA).⁴² Then they interrogated the fluorescence of these Au₈NCs in different organic solvents, including chloroform, tetrahydrofuran and *N,N*-dimethylformamide (DMF). They concluded that the surface properties of Au₈NCs play a major role in these solvent effects. Subsequently, Yi *et al.* explore the ligand- and solvent-dependent electronic relaxation dynamics of certain phenylethane-thiol-stabilised atomically precise monolayer-protected cluster-type AuNCs.⁴³ They found that the electron relaxation dynamics in these clusters are solvent independent and they determined that the electronic-vibrational coupling in these systems is significant.

Recently, many authors have used solvatochromism as an efficient tool for unravelling the fluorescence mechanisms of CDs. In most cases, they suggest that the fluorescence emission of CDs changes with the solvent. These reports unanimously identified the effect of hydrogen bonding of the solvent on the fluorescence of CDs.^{44–49}

1.3.4. Redox status. The close proximity of quantum dots to redox-active metal ions,^{50,51} organic molecules,^{52–54} or metal

complexes⁵⁵ causes fluorescence quenching. This fluorescence modulation has become the basis for several PET-directed sensors. This phenomenon is also used as an efficient tool for monitoring the photophysics of CDs^{56–58} and AuNCs.⁵⁹

1.3.5. pH. The fluorescence of ultra-small nanoparticles is sensitive to external parameters like the pH of the solution. Therefore, pH-elicited changes in the fluorescence of quantum dots can be sensibly tuned for designing effective sensors. In this respect, Raymo and coworkers designed an organic ligand which can be adsorbed on the surface of CdSe–ZnS core-shell quantum dots.⁶⁰ The [1,3]oxazine ring structure on this ligand is transformed into 4-nitrophenyl-azo phenolates on interaction with hydroxide anions. This transformation switches on an energy transfer pathway between the quantum dots and the adsorbed ligand. In this way, the modified quantum dots exhibit good pH-dependent fluorescence quenching. Ultra-small nanoparticles mostly exhibit maximum fluorescence at their optimum pH. For instance, Bao *et al.* reported that hydroxyl-terminated PAMAM-protected AuNCs are stable between pH 6 and pH 8.⁶¹ A breakthrough in AuNC synthesis was achieved by Xie *et al.*, who reported a BSA-directed bio-mineralisation-based AuNC synthesis. These biocompatible AuNCs show stable fluorescence over a broad range of pH from 3 to 12.³¹ Numerous pH-dependent studies were also done on the CD platform. In some cases, CDs exhibit the highest fluorescence intensity in neutral pH.⁶² On the other hand, pH-dependent changes in fluorescence intensity have been reported.⁶³ Moreover, a pH-dependent peak shift is observed in some cases, but it remains consistent in other cases.⁶⁴

1.3.6. Heavy atom effect. The fluorescence quenching of ultra-small nanoparticles *via* the heavy atom effect has wide application in sensor design. Usually, quenchers inducing a heavy atom effect are non-fluorescent paramagnetic species like inorganic halide ions or transition metal ions with a higher atomic number, which are either linked to fluorescent species or located in the vicinity of the fluorophore.¹ The heavy atom effect occurs *via* enhanced intersystem crossing in the presence of these paramagnetic heavy atoms. This is a dynamic quenching mechanism and is always accompanied by a decrease in fluorescence lifetime.

Many research groups reported the heavy atom effect on AuNCs. For instance, Durgadas *et al.* explored the heavy atom effect of copper(II) ions (Cu^{2+}) on BSA-capped AuNCs.⁶⁵ Among such research work, mercury(II) ion (Hg^{2+}) and silver(I) ion (Ag^+) induced quenching of protein-templated AuNCs is very interesting, because Hg^{2+} and Ag^+ can interact with AuNCs *via* the sulfur atoms of the ligand.^{11,66} Moreover, the metallophilic d^{10} – d^{10} interaction of Hg^{2+} or Ag^+ with Au^+ at the surface of AuNCs also plays an important role in their quenching mechanism. By realising the importance of the heavy atom effect on sensor design, Sony's group has tried the fluorescence quenching of 3,4-dihydroxy-L-phenylalanine (L-DOPA)-protected AuNCs with paramagnetic transition metal ions of the 3d series.⁶⁷ As evidence of the heavy atom effect by halide ions on AuNCs, Sony's group has explored iodide-ion-induced quenching on BSA-capped AuNCs and used the affinity between iodide ions

and amino acids for demonstrating an on-off-on mode sensor for cysteine and homocysteine.⁶⁸ Motivated by these results, Sony's group extended iodide-ion-induced quenching to a CD platform and proposed an iodide-ion-quenched CD as an on-off-off mode sensor for tyrosine.⁶⁹

The heavy atom effect appears to be a good tool with which to explore the fluorescence mechanism of CDs. In particular, the Kumbhakar group performed fluorescence quenching experiments on CDs with iodide ions to establish the self-assembly nature of CDs in solution.⁷⁰ Subsequently, Sony's group demonstrated the heavy atom effect of differently doped CDs with halide ions.⁷¹ In addition, there have been numerous reports of metal ion quenching on CDs.⁷² In contrast, detailed investigations of the heavy atom effect on quantum dots are scarce. Typically, Masteri-Farahani and Khademabbasi explored the sodium-halide-ion-induced heavy atom effect on mercaptoacetic-acid-capped CdS quantum dots.⁷³

1.3.7. Doping. Doping is a widely adopted technique to modulate the fluorescence of ultra-small nanoparticles.

Most importantly, doping with suitable non-toxic metal cations of copper, silver and manganese (transition metal ion series) can create intermediate energy levels between the discrete energy levels of quantum dots and cause enhanced quantum yield, improved lifetime, a large Stokes shift and new fluorescence bands.⁷⁴ [See the review article by Wu and Yan for details].

Doped or alloyed ultra-small luminescent AuNCs have unique optical and catalytic properties.⁷⁵ Interestingly, platinum- or palladium-doped AuNCs were found to be more stable than homo-AuNCs. In contrast, silver- or copper-doped AuNCs were less stable. However, silver doping is a widely adopted technique to improve the quantum yield of AuNCs.⁷⁶

Heteroatom doping with nitrogen, sulfur, phosphorous, boron, and metal ions of copper, manganese, iron, zinc, terbium, and gadolinium has been extensively investigated in the CD matrix.¹⁸ Oxygen-rich surface functional groups like –COOH or –OH are common in all types of CDs.⁷⁷ These functional groups (known as surface states) have an influence on the emission from CDs.⁷⁸ In this way, heteroatom doping has great potential for fluorescence tuning. In particular, the atomic size of nitrogen and carbon are comparable, so nitrogen doping in CD is a widely studied doping technique. Yu's group investigated sulfur and nitrogen co-doped CDs synthesised from citric acid and L-cysteine, and developed a model to understand the fluorescence mechanism of heteroatom-doped CDs (see Fig. 7).³³ They claimed that pristine CDs with oxygen-rich surface functional groups exhibited an array of energy levels matching various surface states (oxygen states). The surface states produced a broad UV-visible absorption band and excitation-dependent fluorescence. The heteroatom doping created a new form of surface state known as the nitrogen state. The excited electrons trapped in these nitrogen states induce high radiative recombination. If the density of the nitrogen states is proportional to that of the oxygen states, doped CDs will give wide and excitation-dependent fluorescence. Several authors have reported on the tuning of nitrogen



Fig. 7 Scheme representing the improved fluorescence quantum yield of carbon dots on doping with nitrogen and sulfur. The upward violet arrow indicates electronic excitation from the ground state. These excited electrons get trapped at oxygen-derived surface states and are returned to the ground state via non-radiative routes (curved blue arrow). In the presence of nitrogen states, the rate of radiative transition (downward violet arrow) from the excited state to the ground state is increased.

content on CDs for obtaining a desired fluorescence shift from the blue or green to the red or near infra-red region.²⁰ In particular, Xiong's group reported on the synthesis of NCD from urea and phenylenediamine and the separation of NCD fractions *via* silica column chromatography.⁷⁹ The separated fractions of the NCDs exhibit excitation-independent fluorescence varying from the blue to the red region. They attribute the red shift in emission to a decrease in bandgap with increased content of oxygen species in the surface state. In a recent report, Zhang *et al.* synthesised CDs from *o*-phenylenediamine and demonstrated simultaneous red fluorescence (at 620 nm) and two-photon absorption of near-infrared (NIR)-induced red fluorescence (630 nm and 680 nm) simply by protonating their NCDs.⁸⁰ They found that a molecular fluorophore named 2,3-diaminophenazine (known as the molecular state) formed on oxidation of the precursor played a significant part in the fluorescence of these doped CDs.

2. Luminescent ultra-small nanomaterials

The extremely small size regime of ultra-small nanoparticles induces many unique properties, such as high surface to volume ratio, fluorescence, catalytic activity as well as chirality. In addition, fluorescent AuNCs and CDs are widely known for their biocompatibility.^{12,81} Due to these unique features, ultra-small nanoparticles have attracted widespread research interest for a wide landscape of applications ranging from energy devices and catalysts to fluorescent sensors. This growing interest was established with the award of the 2023 Nobel Prize in Chemistry to Moungi G. Bawendi along with Louis E. Brus and Aleksey Yekimov "for the discovery and development of quantum dots."⁸²

2.1. Quantum dots

Fluorescent quantum dots are ultra-small nanoparticles of diameter 1 to 6 nm.⁸³ Quantum dots include semiconductor

nanoparticles from Group IV (silicon or germanium) as well as alloyed semiconductor nanoparticles with elemental combinations from Groups II–VI (CdE or ZnE, E = S/Se/Te), IV–VI (PbE), III–V (InP) and I–III–VI (CuInS₂, CuInSe₂).⁸ Among these alloyed quantum dots, CdSe- or CdTe-based core and core-shell quantum dots have received a lot of research attention for analytical applications due to their strong quantum confinement effect and high fluorescence quantum yield.¹⁶ Even though quantum dots were discovered in 1981,⁸⁴ their synthesis was a challenging task until 1993 when Bawendi and coworkers proposed a classical hot injection method for cadmium-based quantum dots from a precursor solution of dimethyl cadmium and trioctylphosphine chalcogenide or hexamethyldisilathiane in a high-boiling-point coordinating solvent (trioctylphosphine oxide).⁸⁵ However, the biocompatibility of quantum dots became an issue and a lot of research work concentrated on alternative cadmium precursors. Then quantum dot synthesis employing CdClO₄, cadmium oxide,⁸⁶ cadmium carbonate and cadmium acetate⁸⁷ was demonstrated.⁸⁸ Instead of toxic phosphine derivatives, oleic acid and octyl decyl amine have been used.^{88,89} Nowadays, quantum dots can be prepared in organic solvents or aqueous solutions.^{16,88} Alternatively, approaches based on artificial neural networks attempt to predict the photoluminescence emission spectra of CdSe quantum dots from different reaction conditions (*e.g.*: precursor composition, choice of ligand, reaction temperature, time) from a combined quantum dot synthesis data set from the literature.⁹⁰

Surface defects on quantum dots can adversely affect the quantum yield of quantum dots because an excited electron in the conduction band of the quantum dot can be nonradiatively transferred to these surface defects created by oxygen vacancies, trap states or defects.⁹¹ Photoblinking (a random transition from an on to an off state) is another problem which hinders the wide application of quantum dots in single-molecule experiments.⁹² The blinking behaviour of a single quantum dot is thought to be caused by the non-radiative recombination of electron-hole pair excitons in the off state.⁹³ The duration of the off state follows Lévy statistics and is marked by observable and long non-emissive periods.⁹⁴ So, there have been reports on the construction of non-blinking giant quantum dots by expanding the shell thickness of core-shell quantum dots up to a diameter of 13–15 nm.⁹² Similarly, high-quality quantum dots can be synthesised only in the organic phase. Such quantum dots have limited use in bioengineering.⁹³ To overcome these challenges, ligand exchange or surface coating with suitable hydrophilic and biocompatible ligands (*e.g.*: silica, amino acids, proteins, mercaptocarboxylic acids) are adopted for modifying the quantum dot surface and tuning the material performance for desired bio-related applications.^{88,91,95} Brus and Efros published a recent review narrating the story of quantum dots from the past to the modern era.⁹⁶

The photostability of quantum dots is a persistent challenge in nanochemistry due to the increased chances of oxidation, aggregation, and the creation of surface defects on the prolonged storage of quantum dots.⁸⁸ Zinc-based binary quantum



Fig. 8 Scheme representing the structures of Au⁺-thiolate complexes, conventional Au-thiolate nanoclusters with a shell of short Au⁺-thiolate motifs over an inner Au⁰ core and Au-thiolate nanoclusters exhibiting aggregation-induced emission (AIE) of an oligomeric Au⁺-thiolate complex over an inner Au⁰ core. The structure of gold nanoparticles (AuNPs) with a larger Au⁰ core is also given for comparison.

dots and ternary quantum dots could be a remedy for the toxicity issue in association with cadmium-based quantum dots. Even so, there is a strong demand for biocompatible counterparts to quantum dots.⁸

2.2. Gold nanoclusters

AuNCs are ultra-small gold nanoparticles of diameter 1–3 nm.⁹⁷ AuNCs show a broad UV-visible absorbance spectrum and molecule-like bright fluorescence over the visible to near-infrared region due to quantum confinement of conduction band electrons to the Fermi wavelength size regime of gold.⁶⁶ These AuNCs were first discovered in the gas phase.¹⁶ Detailed investigation of their unique properties and application for ultra-sensitive detection, bio-imaging and catalysis were hindered until researchers succeeded in synthesising these promising luminescent ultra-small materials in organic and aqueous phases. At present, they are generally prepared either by the etching of larger gold nanostructures or the reduction of a suitable gold salt in the presence of a stabilising agent like small thiolate or carboxylate ligands, proteins and polymers.⁸ The as-obtained atomically precise AuNCs are stabilised in the pockets of proteins, polymers or small ligands.¹⁶

The fluorescence emission from AuNCs is determined by several factors like Au⁰ core size, the Au⁺ content of the surface

shell over the inner core, surface ligands and the valence state of the AuNCs.¹⁰ Approaches based on machine learning evaluate the correlation between the physical properties of AuNCs and various reaction condition parameters by analysing a selected finite number from previous literature.^{98–100} However, optimisation of all variables and reaction parameters can be puzzling, owing to restricted knowledge about efficient combinations in the vast chemical space.¹⁰¹ Many thiol-capped AuNCs exhibit a large Stokes shift and a long microsecond lifetime.¹⁴ The unique behaviour of these thiol-capped AuNCs is correlated with the ligand-to-metal charge transfer (LMCT) interactions between sulfur atoms and the gold centre.¹⁴ Besides, the Au⁰@Au⁺-thiolate core-shell structure of these thiol-capped AuNCs holds oligomeric interlocked Au⁺-ligand motifs in the exterior shell over the inner Au⁰ core (Fig. 8). The aggregation-induced emission (AIE) of the oligomeric Au⁺-thiolate complex on the surface of the Au⁰ core makes a major contribution towards the long lifetime and large Stokes shift of their strong luminescence.²⁸ More specifically, this model claimed that the process behind the aggregated Au⁺-thiolate-motif-induced phosphorescence is an LMCT or ligand-to-metal-metal charge transfer (LMMCT), which produces radiative recombination of excitons *via* a metal-centered triplet state (see Fig. 9).¹⁰² Easy access to the triplet state is facilitated by improved spin-orbit coupling in the presence of a heavy inner Au⁰ core. Yet, the emission mechanism behind the AuNCs is not completely understood. Moreover, the fluorescence quantum yield of many monolayer-protected AuNCs is low. Therefore, researchers are driven by curiosity to explore AIE-type AuNCs in detail to achieve AuNCs with good quantum yield as a substitute for organic dyes or typical quantum dots for practical applications, even though precursors for AuNCs are rare and costly. Thus, there is a consistent demand for other biocompatible quantum dots from cheap and abundant precursor molecules.

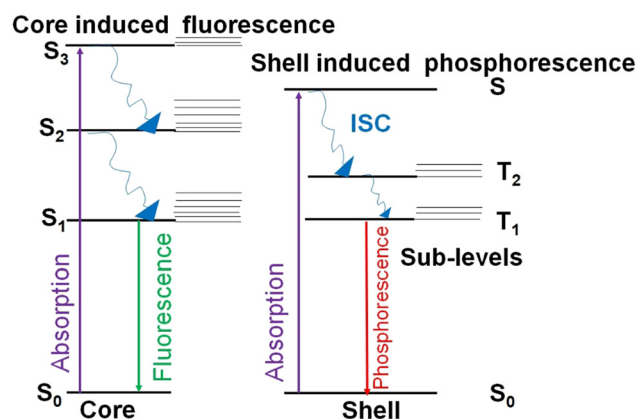


Fig. 9 Diagram showing radiative and nonradiative electronic transitions within aggregation-induced emission (AIE)-type AuNCs. The phosphorescence is generated by the AIE of the oligomeric Au⁺-thiolate complex on the surface shell. The fluorescence arises because of the Au⁰ core. S_n and T_n represent singlet state and triplet state, respectively. ISC stands for inter-system crossing.

2.3. Carbon dots

Fluorescent CDs are quasi-spherical nanoparticles with a diameter less than 10 nm which consist of a crystalline or amorphous sp^2 carbon core embedded in an sp^3 carbon matrix (Fig. 10).¹² CDs are proposed as a better substitute for classical fluorescent quantum dots because of their superior qualities like biocompatibility, chemical inertness, water-solubility, easy functionalization and low cost of synthesis.¹⁸ The discovery of fluorescent CDs in 2004 was serendipitous while attempting the separation and purification of single-walled carbon nanotubes prepared through an arc discharge method.¹⁰³

Subsequently, the discovery of a reliable synthetic methodology for CDs has become an interesting research topic. Sun and colleagues synthesised CDs *via* the laser ablation of carbonaceous material and subsequent surface functionalization of the obtained non-fluorescent nanoparticles with amine-terminated polyethylene glycol.³⁵ This group suggested the name 'carbon quantum dots' because they expected that the multiple excitation-dependent emission of the obtained CDs was evidence of quantum confinement and size effects. Next, the synthesis of CDs *via* electrochemical oxidation of graphite or multi-walled carbon nanotubes attracted tremendous attention because it was identified as an easy and cheap method to yield CDs in high quantity.^{18,104} However, these top-down approaches demand several complex, time-consuming and expensive post-synthetic treatments like surface functionalization. To overcome the limitations of top-down methods, bottom-up methods based on templates, the hydrothermal, microwave or ultrasound assisted decomposition of small molecules like glucose, citric acid, urea, or phenylenediamine⁸⁰ were widely adopted

to synthesise CDs.^{79,105–107} Some groups attempted machine learning (*e.g.*: artificial neural network) assisted CD synthesis.¹⁰⁸

Kasprzyk and coworkers isolated fluorescent molecules including several citrazinic acid derivatives from the reaction product formed on the decomposition of organic molecules.^{109,110} So substantial attention has been paid to a theory based on the molecular fluorescence of CDs (Fig. 10).^{37,111,112} Although, researchers were very curious as to why the excitation spectrum of CDs does not coincide with the absorption spectrum. Further, single-particle spectroscopic studies reveal that individual particles do not exhibit the same spectral or lifetime heterogeneity as their ensemble. To answer these contradictions, Demchenko *et al.* proposed an 'H-like emissive aggregate model' for CDs by considering the presence of surface-exposed fluorophores (Fig. 11).^{49,70,113–115} The theory of 'J and H molecular aggregates' was originally framed by Kasha and co-workers.^{30,116} The positive Coulomb coupling (parallel arrangement) between component fluorophores produced weakly emissive H-aggregates (abbreviated 'H for hypsochromic'), which are distinguished by a hypsochromic (blue) shift in the absorption/excitation spectrum, a marked red shift in suppressed fluorescence, and an increase in the rate of phosphorescence through intersystem crossing. Blue-shifted fluorescence and bathochromic shift (red shift) in the absorption/excitation spectrum are the hallmarks of superradiant J-aggregates, also known as Sheibe aggregates (J for E. E. Jelly, who discovered this form of aggregate in 1936). Yet, it is difficult to draw far-reaching conclusions on this issue and this latest J- and H-aggregate concept requires more cautious and careful investigation to achieve reliability of the emissive aggregate structure, especially for CDs from other precursor molecules. Besides, there are literature reports considering molecular fluorophores in CDs as being the consequence of impurities due to the use of inadequate purification techniques.¹¹⁷ Concurrently, multiple fluorescence emission bands from certain CDs are observed at a single excitation wavelength.¹¹⁸ These multiple emission bands are interpreted as individual contributions from the quantum confinement

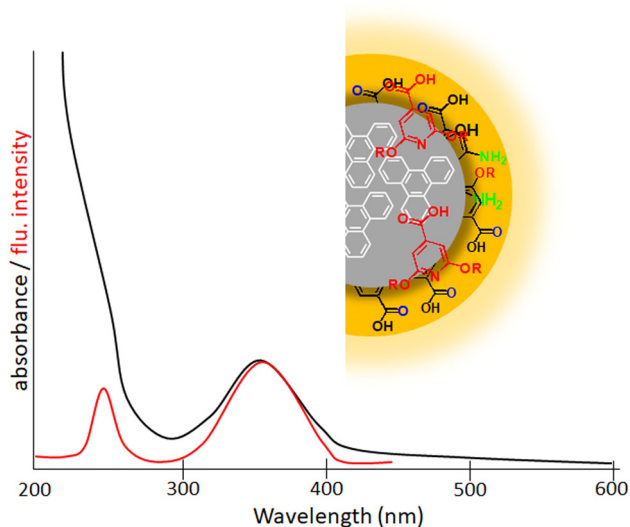


Fig. 10 Scheme representing the structure of a carbon nanodot. The inner carbon core of a carbon nanodot is composed of numerous conjugated π -domains. The surface functional groups connected to this inner core are termed the surface state. The fluorescent molecule (red molecular entities) connected to the surface or interior of the carbon core are known as the molecular state. A representation of a typical excitation spectrum (red line) and absorption spectrum (black line) of carbon dots is also given.

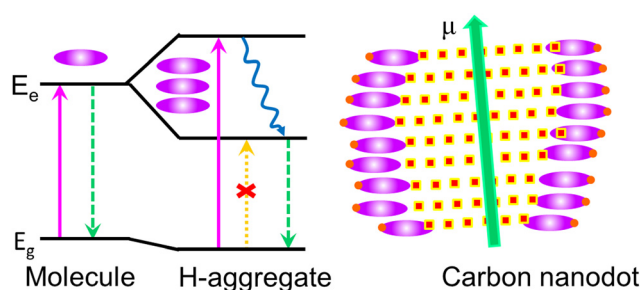


Fig. 11 The proposed 'H-like emissive aggregate model' for carbon dots. The H-like aggregates are formed by side-by-side arrangement of component fluorophores (represented by oval-shaped structures), which results in the splitting of excited-state energy levels into two. Among both these excited-state energy levels, excitation to higher excited level is the only allowed transition. However, there is excited electron non-radiative relaxation (curved arrow) to the lower excited level, and emission occurs from this lower excited level. This causes a large Stokes shift. The carbon dots are carbonised nanoparticles with numerous surface-exposed fluorophores assembled as H-aggregates.



Fig. 12 The fluorescence emission mechanism in multiemission carbon dots is described as individual contributions from the quantum size effect, surface state and molecular state.

effect, surface defects and molecular fluorescence (Fig. 12). In this context, the absence of a unified emission mechanism for CDs has hindered their fluorescence tuning to the desired wavelength region and complete fulfilment of their potential applications like bio-sensing and bioimaging.^{64,70}

3. Fluorescence quenching

A wide variety of substances, including metal ions, molecules and nanomaterials can cause fluorescence quenching (a decrease in fluorescence intensity) of ultra-small nanoparticles. The quenching process requires intimate contact between fluorescent ultra-small nanoparticles and quenchers. Consequently, quenching measurements can reveal the accessibility of fluorescent centres to quenchers. Thus, the occurrence of quenching depends on the quenching mechanism and the structure of the ultra-small nanoparticles. So, a detailed investigation of the fluorescence quenching of ultra-small nanoparticles provides a hint about the structural aspects of these materials. Four fundamental types of fluorescence quenching have been identified and form the main topic of discussion in this section (Fig. 13).



Fig. 13 The four fundamental types of fluorescence quenching are: dynamic quenching, static quenching, combined static and dynamic quenching, and amplified quenching. The representative kinetic curves are given to the side.

3.1. Dynamic quenching

Dynamic quenching of fluorescence is described by the Stern–Völmer equation:

$$\frac{I_0}{I} = 1 + k_q \tau_0 [Q] = 1 + K_D [Q]$$

In this equation, I_0 and I are the fluorescence intensities of a fluorescent material in the absence and presence of a quencher, respectively, k_q is the bimolecular quenching constant, τ_0 is the lifetime of a fluorescent material in the absence of a quencher, and K_D is the Stern–Völmer quenching constant. A linear plot of I_0/I versus concentration of a quencher give an intercept of one in the y-axis and a slope equal to K_D . For dynamic quenching, $\frac{\tau_0}{\tau} = \frac{I_0}{I}$, where τ_0 and τ are the fluorescence lifetime in the absence and presence of a quencher. Dynamic quenching affects only the excited state of the fluorescent material and has no effect on the absorption spectrum of the fluorescent material. The rate of dynamic quenching increases with temperature and decreases with viscosity because these parameters can influence the collision rate.

This type of quenching mechanism is observed for many ultra-small nanoparticles. Notably, the fluorescence quenching of homocysteine-capped CdTe quantum dots by Cu^{2+} ions illustrates this mechanism.¹¹⁹ To prove its dynamic quenching mechanism, quenching experiments at different temperatures (293 K, 298 K, 303 K) were conducted and a proportional rise in Stern–Völmer quenching constant was observed. This is due to enhanced collisions and dynamic quenching effect with a rise in temperature. Similarly, Ungor and colleagues conducted temperature-dependent fluorescence quenching experiments on λ -globulin-immunoprotein-stabilised red-emitting AuNCs by using L-kynurenine (a tryptophan metabolite) as a quencher.¹²⁰ They also observed evidence of dynamic quenching in this system. In another case, Zhang *et al.* reported the dynamic quenching of DNA-stabilised AuNCs with added glutathione.¹²¹ They confirmed the quenching mechanism using UV-visible absorption spectroscopy. These authors found a similar null effect in the absorbance spectrum of AuNCs after the addition of glutathione and concluded that it was an example of dynamic quenching. Furthermore, water molecules can cause the dynamic quenching of a CD dispersion in an organic solvent and this work was reported by Zheng and group members. They clarified the mechanism using time-resolved fluorescence spectroscopy and UV-visible absorption spectroscopy.¹²² They noticed that the UV-visible absorption spectra of CDs and a CD–water system are superimposed on one another. This indicates dynamic quenching. In addition, they confirmed the dynamic quenching mechanism by observing the variation in the fluorescence lifetime of the fluorophore in the presence and absence of water molecules.

3.2. Static quenching

The kinetics of static quenching can be represented as:

$$\frac{I_0}{I} = 1 + K_S [Q]$$

In this equation, K_S is the association constant. The measurement of fluorescence lifetime is the most appropriate method to distinguish static and dynamic quenching, because $\frac{\tau_0}{\tau} = 1$ for static quenching. The static quenching mechanism is the formation of a non-fluorescent ground-state complex between the fluorescent material and a quencher and it shows significant changes in the absorption spectrum of the fluorescent material.

The fluorescence quenching of a CdS quantum dot/N-methylpolypyrrole hybrid by picric acid is one static quenching experiment in the literature.¹²³ In this case, the static quenching mechanism is confirmed by observing the formation of a new absorption peak upon the addition of picric acid to this fluorescent material. Subsequently Ma *et al.* reported the static quenching of orange fluorescence of CDs by Ag^+ ions.¹²⁴ This group also found a notable difference between the absorption spectrum of the CDs before and after the addition of Ag^+ ions. However, the fluorescence lifetime of the CDs was 2.42 and 2.43 ns before and after the addition of Ag^+ ions. Then, Santhosh *et al.* demonstrated the static quenching of human-serum-albumin-stabilised AuNCs with bilirubin.¹²⁵ This research group also underlined that the fluorescence lifetime of a fluorescence material remains unchanged on static quenching.

3.3. Combined static and dynamic quenching

An upward-curving Stern–Völmer plot is strong evidence for combined static and dynamic quenching. The kinetics for this quenching process can be described as:

$$\frac{I_0}{I} = (1 + K_D[Q])(1 + K_S[Q])$$

This equation is second order in $[Q]$, which accounts for the upward-curving Stern–Völmer plot. More specifically, this type of quenching is observed on the interaction of CdTe quantum dots with protamine sulfate.¹²⁶ These authors noted a drastic increase in the slope of the Stern–Völmer plot with an increase in the temperature of the quenching experiments. This observation indicates dynamic quenching, but they did not find a corresponding decrease in the fluorescence lifetime proportional to the changes in fluorescence intensity. Hence, they monitored the zeta potential values of the quantum dots, which indicated a significant increase from -19.3 mV to $+19.8$ mV after the addition of protamine sulfate. The TEM image demonstrates aggregation in the quenching process. As a result, the authors proposed a combined static and dynamic quenching mechanism. Similarly, Anand *et al.* reported that the Stern–Völmer plot for the quenching of L-cysteine-functionalised CdS quantum dots with tetracycline displays a positive deviation and is concave to the y-axis.¹²⁷ Further, they observed a considerable decrease in the fluorescence lifetime on quenching. Then a linear plot of $\frac{\tau_0}{\tau}$ versus concentration of tetracycline gives a slope equal to K_D . From their fluorescence quenching data, they calculated the value of K_S using the standard method and found that $K_D > K_S$. Dynamic quenching predominates

over static quenching in this case. Subsequently, Lin and co-workers demonstrated the combined static and dynamic quenching of PAMAM-dendrimer-entrapped Au_8NCs by a paramagnetic nitroxide radical.¹²⁸ Similarly, this type of quenching mechanism was used by Gagoi and Khan for designing a CD-based fluorescence immunosensor for cardiac Troponin T.¹²⁹ This work illustrates FRET from CDs to a molybdenum disulfide nanosheet. However, plots of $\frac{I_0}{I}$ versus concentration of the molybdenum disulfide nanosheet and $\frac{\tau_0}{\tau}$ versus the concentration of molybdenum disulfide nanosheet do not coincide. This suggests the minor involvement of a static quenching mechanism. Moreover, the plot of $\frac{I_0}{I}$ versus quencher concentration displays an upward curvature.

3.4. Amplified quenching (or quenching sphere of action)

Amplified quenching is identified as evidence of aggregation, strong complex formation between the quencher and fluorescent material, as well as long-distance migration of a singlet exciton to the quencher trap state. Amplified quenching is achieved when the Stern–Völmer equation follows an exponential function,

$$\frac{I_0}{I} = (1 + K_D[Q]) \left(\exp \left(\frac{[Q]VN_0}{1000} \right) \right)$$

where V is the volume of the sphere and N_0 is Avogadro's number.¹ Importantly, fluorescence quenching of certain oleic-acid-capped CdSe quantum dots with nitroaromatic explosives follows this mechanism.¹³⁰ Interestingly, Giri and co-workers found that the fluorescence quenching of graphene quantum dots with single-walled carbon nanotubes followed an exponential Stern–Völmer plot.¹³¹ Anjali Devi *et al.* demonstrated an amplified fluorescence quenching effect on citric-acid-urea-derived CDs by transition metal ions (*e.g.*: Fe^{3+} , Ru^{3+} , Cr^{6+}) in polar solvents (DMSO and water).¹¹⁵

4. Quantitative parameters of luminescent sensors

The quality and performance of luminescence sensors are validated using different detection parameters.

4.1. Stern–Völmer quenching constant

The Stern–Völmer quenching constant (K_{SV}) is the product of k_q (the bimolecular quenching constant) and τ_0 (the lifetime of the fluorescent material in the absence of a quencher). The Stern–Völmer constant is widely adopted for describing the quenching of many ultra-small nanoparticles.

4.2. Limit of detection and quantification

The limit of detection (LOD) is also called the lower limit of detection.¹³² It is the smallest amount of analyte that can be

reliably measured with a given analytical method.⁷

$$\text{LOD} = \frac{3\sigma}{m}$$

where σ is the standard deviation of a low-concentration sample and m is the slope of the calibration curve.

The instrument detection limit is calculated by replicate ($n \geq 7$) measurements of an aliquot from one sample. The method detection limit will be greater than the instrument detection limit. The method detection limit is calculated by preparing different individual samples and analysing each at a one time.¹³²

Standard deviation is the degree of noise (random variation) in the blank or a small signal. When the signal is three-fold larger than noise, it is detectable. But it is not enough for accurate measurements. A signal which is ten-fold larger than noise is termed the limit of quantification (LOQ). LOQ is the smallest amount of analyte that can be accurately measured with a given analytical method:^{7,132}

$$\text{LOQ} = \frac{10\sigma}{m}$$

5. Underlying principles of luminescent ultra-small nanoparticle sensors

A survey of the literature on ultra-small nanoparticle sensors reveals that chemical interaction of the analyte with sensors occurs and this event will be transmitted to the observer as quantitative changes in the fluorescence signal. Several chemical interactions and fluorescence quenching mechanisms were responsible for changes in fluorescence intensity and the emission wavelength of ultra-small nanoparticles has the potential to be used as an indicator for analytes.

5.1. Non-covalent interactions

The non-covalent interaction between ultra-small nanoparticles and analyte molecules is very common. Here, we illustrate a small collection of examples discussing various non-covalent interactions in the literature (Fig. 14).

5.1.1. Hydrophobic association. Hydrophobic association is the association of different non-polar molecules in aqueous solution. This interaction is widely used for many cyclodextrin-functionalised ultra-small nanoparticle sensors. The cyclodextrin binds to hydrophobic guest molecules in its interior cavity. This binding event can affect the fluorescence emission signal of fluorescent nanoparticles attached nearby. The binding of the analyte can cause displacement of photophysically active guests from the cavity, or the analyte itself can undergo photophysical changes due to encapsulation in the cyclodextrin cavity. A pioneering report on this type of ultra-small nanoparticle sensor came from the Willner group in 2009.¹³³ They tried the competitive analysis of adamantane carboxylic acid and *p*-hydroxytoluene with Rhodamine-*B*-dye-incorporated



Fig. 14 Scheme representing various non-covalent interactions operating between ultra-small nanoparticles and analytes during the sensing process.

β -cyclodextrin-modified CdSe-ZnS core-shell quantum dots. Besides, they demonstrated the chiroselective response of this sensor towards D,L-phenylalanine and D,L-tyrosine. Feng and co-workers synthesised β -cyclodextrin-modified CDs for a fluorometric assay of β -galactosidase activity with a detection limit of 0.6 UL^{-1} .¹³⁴ The fluorescence of the as-designed CDs will be quenched on the formation of an inclusion complex with glycosylated *p*-nitrophenol through a static quenching mechanism. There were reports on a hydrophobic inclusion complex of cyclodextrin-functionalised CDs with organic entities like C_{60} fullerene,¹³⁵ methyl viologen,¹³⁶ or testosterone.¹³⁷

5.1.2. Electrostatic interactions. Electrostatic interaction is the electric force of attraction/repulsion between different/similarly charged molecules. Examples illustrating electrostatic interaction in luminescent ultra-small nanoparticle sensors use ionic polyelectrolytes or organic molecules as surface ligands to bind oppositely charged analytes and transduce that binding into noticeable fluorescence changes. More specifically, Saberi *et al.* demonstrated the fluorescence quenching of cationic CDs from cetrimonium bromide with a negatively charged acetamidiprid aptamer *via* electrostatic interaction.¹³⁸ They used this system for the fluorescence turn-on detection of acetamidiprid (a pesticide) with a detection limit as low as 0.3 nM. Similarly, Vaishnavi and Renganathan demonstrated the cationic porphyrin-induced quenching of anionic thioglycolic-acid-capped CdTe quantum dots and used this system as a turn on-off-on mode sensor for anionic calf thymus DNA.¹³⁹ Pyo *et al.* used the fluorescence quenching of cationic azadioxatriangulenium dye by electrostatic interaction with various anionic *p*-mercaptobenzoic-acid-capped AuNCs. The fluorescence of the dye can be restored by tuning the surface charge of AuNCs with pH, thereby creating a pH sensor.¹⁴⁰

5.1.3. Hydrogen bonding. Hydrogen bonding is a special type of dipole-dipole interaction. It is a non-covalent bonding

interaction where a hydrogen atom is covalently bonded to two or more electronegative atoms like O, N or halogen.

Chen and co-workers demonstrate hydrogen bonding and electrostatic interaction between dopamine and (3-amino-propyl)trimethoxysilane-capped silicon nanoparticles.¹⁴¹ Similarly, Ma *et al.* reported on electrostatic and hydrogen bonding interactions between heparin and silicon nanoparticles prepared from 3-aminopropyltriethoxysilane and L-ascorbic acid.¹⁴²

Sciortino *et al.* reported that citric-acid-urea-co-derived CDs show a strong response towards the hydrogen bonding interactions of the dispersing solvent.⁴⁶ They proposed these CDs as an example of a solvent polarity indicator. Subsequently, Anjali Devi *et al.* reported the hydrogen-bonding-directed J/H-aggregate-like nature of citric-acid-urea-based CDs in polar solvents.⁴⁹

Heintzelman and co-workers investigated the fluorescence of particular phosphine-protected AuNCs {i.e., $[\text{Au}_{11}(\text{BINAP})_4\text{X}_2]^+$ where X = Cl, Br} in different solvents like dichloromethane, methanol, or ethanol. They observed that hydrogen bonding between halogen in the surface of the AuNCs and the dispersing solvent is crucial in intense photoluminescence and produced a 2-fold reduction in vibration-mediated non-radiative pathways. They noted the quenching of these AuNCs in a solvent which has no favourable hydrogen bonding.¹⁴³

5.2. Mechanism of luminescence sensing

This section is devoted to outlining various fluorescence quenching mechanisms reported in the literature for designing selective, sensitive and fast-response fluorescence detection strategies using ultra-small nanoparticles (Fig. 15). These mechanisms are not mutually exclusive, so that several reports interrogate whether fluorescence quenching occurs *via* combinations of these mechanisms.

5.2.1. FRET. FRET is the non-radiative energy transfer from excited fluorophores to neighbouring acceptors *via* long-range

dipole-dipole interaction. It is widely used for bioanalytical applications.

5.2.1.1. Quantum dots. In many FRET sensors, quantum dots act as a donor fluorescent material. Importantly, Miao *et al.* reported a FRET-based aptasensor for chloramphenicol, which works on the energy transfer from CdSe quantum dots to DNA-labelled iron-oxide-linked silica-coated gold nanoparticles (AuNPs). This method has a linear range of $0.001\text{--}10\text{ ng mL}^{-1}$ with a detection limit of 0.0002 ng mL^{-1} .¹⁴⁴ Similarly, Chang *et al.* used FRET between glucosamine-coated manganese-doped ZnS quantum dots and mercaptophenyl-boronic-acid-capped AuNPs for the detection of glycoproteins like immunoglobulin G, transferrin, $\alpha 1$ -acid glycoprotein and horseradish peroxidase.¹⁴⁵ FRET-based sensors are efficient enough to accomplish accurate bioanalysis. Subsequently, many authors reported FRET sensors for gene detection. For instance, Sham-sipur *et al.* designed a FRET nanosensor for a BCR/ABL fusion gene in patients with chronic myelogenous leukemia.¹⁴⁶ They used 18-mer-oligonucleotide-conjugated CdTe quantum dots as a donor and methylene blue as an acceptor. The hybridisation of DNA on the quantum dot surface to target the BCR/ABL fusion gene ensures the proximity of methylene blue and quantum dots and this proximity enables FRET. This FRET nanosensor provides fluorescence enhancement of methylene blue on photoexcitation of the quantum dots as the signal for quantitative detection. In many other cases, quantum dots act as an acceptor in a FRET sensor. For instance, Borghei *et al.* used FRET from DNA-conjugated silver nanoclusters to thio-glycolic-acid-capped CdTe quantum dots for detecting the BRCA1 gene in human breast cancer cells (MCF-7).¹⁴⁷

5.2.1.2. CDs. There have been numerous efforts to design FRET sensors working on the fluorescence of CDs. For instance, Shu *et al.* tried the ratiometric fluorescence imaging of intracellular zinc in MCF-7 cells using FRET between citric-acid-polyethylenimine-co-derived CDs and fluorescein.¹⁴⁸ Ge *et al.* designed a FRET-based strategy for analysing the antibiotic molecule, D-penicillamine.¹⁴⁹ This strategy is based on energy transfer from citric-acid-urea-derived CDs to pH-mediated aggregates of AuNPs. Cheng *et al.* reported a FRET-based aptasensor for adenosine triphosphate (ATP) using CDs as donor and graphene oxide as acceptor.¹⁵⁰ Similarly, a graphene-oxide-CD FRET system was used for the detection of Mucin-1 protein.¹⁵¹ Subsequently, Qaddare *et al.* proposed a CD and AuNP based FRET-aptasensor for homogeneous HIV-1 gene detection.¹⁵² They used a nanocomposite consisting of AuNPs and graphene oxide as acceptor in FRET to achieve amplification of the signal. Chawre *et al.* constructed a FRET assembly composed of CDs and gold nanorods for the detection of H_2O_2 , glucose, glutathione, and acetylcholinesterase.¹⁵³

5.2.1.3. AuNCs. AuNPs are widely accepted as efficient quenchers for FRET sensors. There have been several reports regarding the tuning of the structure and shape of AuNPs to achieve a desired absorption property in a suitable wavelength region. For instance, Qin *et al.* designed a FRET system working



Fig. 15 The fluorescence quenching mechanisms used for designing fluorescence detection strategies with ultra-small nanoparticles.

in the near-infrared region for the turn-on fluorescence sensing of Glutathione-S-transferase. In this case, AuNCs act as a donor and gold nanorods act as an acceptor.¹⁵⁴

5.2.2. PET. In the photoinduced electron transfer (PET) mechanism, a complex is formed between an electron donor and an electron acceptor. The excited ultra-small nanoparticles can act either as an electron donor or an electron acceptor, depending upon the oxidation and reduction potential of the ground and excited states.¹ Numerous PET-mediated quenching cases on ultra-small nanoparticles can be traced in previous literature. In the oxidative PET mechanism, a photo-excited luminescent material transfers an excited electron from its ground-state LUMO to the vacant LUMO of the quencher (acceptor). In this case, the ground-state LUMO of the donor luminescent material should be higher in energy than the LUMO of the acceptor. On the other hand, reductive PET occurs when the quencher donates an electron from its ground-state HOMO to the singly occupied ground-state HOMO of a photo-excited luminescent material (see Fig. 15).

5.2.2.1. Quantum dots. Specifically, Jose *et al.* employed PET-mediated fluorescence quenching of mercaptosuccinic acid-capped CdTe/Cds quantum dots for the detection of pralidoxime.¹⁵⁵ In this case, the oxidation and reduction potentials of the quantum dots were 1.22 V and -1.19 V, respectively. Consequently, the HOMO and LUMO energy levels of the quantum dots were at -5.83 eV and -3.69 eV. The LUMO of pralidoxime is at -4.09 eV. From these values, it is very evident that the LUMO of pralidoxime lies within the HOMO-LUMO energy gap of the quantum dots and this facilitates electron transfer from the LUMO of the quantum dots to the LUMO of the quencher molecule. Huang *et al.* reported the PET-based off-on mode detection of a ruthenium anticancer drug and calf thymus DNA using CdTe quantum dots.¹⁵⁶

5.2.2.2. CDs. The PET mechanism is also common in CDs. The pioneering report on PET-mediated quenching on CDs came from Wang *et al.* in 2009, who performed fluorescence quenching experiments on CDs with nitroaromatics.⁵⁶ Subsequently, Qu *et al.* demonstrated the PET-mediated fluorescence quenching of dopamine-derived CDs with Fe³⁺ ions.⁵⁸ Then, many authors assigned the fluorescence quenching of CDs with Fe³⁺ ions as an illustration of the PET mechanism.¹⁵⁷

5.2.2.3. AuNCs. Interestingly, Huang *et al.* reported the oxidative PET-mediated fluorescence quenching of dithiothreitol-carboxylated-chitosan-co-modified AuNCs with doxorubicin.¹⁵⁸ Qu's group and Yun's group demonstrated the dopamine-induced fluorescence quenching of BSA-stabilised AuNCs *via* a PET mechanism.^{59,159} Yun's group found that dopamine can transfer a proton to BSA-stabilised AuNCs to form dopaquinone and the fluorescence of AuNCs was quenched *via* reductive PET in the presence of dopaquinone.

5.2.3. Inner filter effect. The inner filter effect (IFE) was considered an inevitable error in fluorescence spectroscopy. In recent years, it has found profound application in fluorescence sensors. IFE is a radiative energy transfer pathway due to

the absorption of an excitation or emission beam for a fluorescence material by an absorber in the sensing system.¹⁶⁰

5.2.3.1. Quantum dots. Khezri *et al.* designed an IFE-based fluorescence sensor for arginine using L-cysteine-capped CdS quantum dots and gold nanoparticle quenchers.¹⁶¹ Arginine (pI 10.8) has a positive charge and citrate-capped gold nanoparticles have a negative charge at the working pH of the sensor (pH 9.6). The introduction of arginine causes aggregation of the gold nanoparticles to give corresponding colorimetric signals. Meanwhile, IFE on the quantum dots by gold nanoparticles gets broken, which in turn switches on quantitative off-on mode fluorescence signals.

5.2.3.2. AuNCs. IFE-based AuNC sensors have also been widely reported. For instance, Chang *et al.* used poly(vinylpyrrolidone)-protected gold nanoparticles as absorbers of the excitation beam of BSA-capped AuNCs.¹⁶² These authors employed *in situ* syntheses of poly(vinylpyrrolidone)-protected gold nanoparticles by the direct mixing of an HAuCl₄ solution and poly(vinylpyrrolidone) in the presence of hydrogen peroxide (H₂O₂) as a catalyst. They employed this method for the detection of H₂O₂ and cholesterol.

5.2.3.3. CDs. A remarkable report on IFE with CDs is the fluorescence quenching of citrate-based CDs with Cr⁶⁺ ions.¹⁶³ Similarly, Sun *et al.* showed IFE with CDs using vitamin B₁₂ as an absorber.¹⁶⁴ Xu and co-workers designed a machine-learning-assisted fluorescence sensor array based on two different CDs to recognize four variants of tetracycline *via* an IFE sensing mechanism.¹⁶⁵

5.2.4. Chelation-induced fluorescence enhancement. Fluorescence enhancement may happen on complex formation between an analyte and the surface ligands of the nanoparticle. More specifically, Liu *et al.* synthesised boronic acid-amine-bi-functionalised CDs by the hydrothermal decomposition of aminophenylboronic acid so that these CDs can complex with dopamine and this binding event is transmitted to the observer as a quantitative enhancement in fluorescence signal.¹⁶⁶ Likewise, Chaiendoo *et al.* reported the fluorescence enhancement of CDs synthesised from lactose in a heated alkaline medium upon polydopamine coating over its surface.¹⁶⁷ In addition, Ag⁺-induced enhancement of CDs was also reported.¹⁶⁸

5.2.5. Chelation-induced fluorescence quenching. Fluorescence sensors can work on fluorescence quenching in association with non-fluorescent complex formation between the sensor entity and the analyte molecule. A detailed discussion of this type of sensor is given in the section entitled "static quenching".

5.2.6. Aggregation-induced emission. AIE in fluorescent materials was discovered by Tang's group in 2001. The restricted intramolecular motion (RIM) in association with aggregate formation is the pivotal reason behind this phenomenon.¹⁴

5.2.6.1. AuNCs. Notably, Xie's group investigated the fluorescence of glutathione-capped AuNCs and found evidence of AIE in these materials.²⁸ They detected the Au⁰@Au⁺-thiolate

complex structure of glutathione-capped AuNCs. They stated that these structures are formed by aggregation of an Au^+ -thiolate complex over an Au^0 core. Chang *et al.* also found a similar aggregated structure in L-cysteine-capped AuNCs.¹⁶⁹ In another pioneering report, Huang *et al.* showed AIE enhancement of glutathione-capped AuNCs with human serum albumin.¹⁷⁰ Similarly, You *et al.* demonstrated the AIE enhancement of glutathione-capped AuNCs by peptides like AG73 peptide and glycine-arginine repeat peptide.¹⁷¹ Fu and co-workers reported poly(allylamine hydrochloride)-mediated AIE in (D-/L)-penicillamine-stabilised AuNCs.¹⁷²

5.2.6.2. CDs. Interestingly, Gigoi *et al.* reported AIE enhancement behaviour of CDs while diluting CD dispersions in tetrahydrofuran/water medium with water. Further, they witnessed the fluorescence enhancement of CDs in the presence of cholesterol.¹⁷³

5.2.6.3. Quantum dots. Similarly, Mou *et al.* demonstrated the AIE property of Cu-In-S ternary quantum dots while changing the solvent from water to 90% (by volume) DMSO.¹⁷⁴ Parthan *et al.* tried to explore magnetic graphene quantum dots and their AIE enhancement effect for designing a turn-on sensor for As^{3+} ions.¹⁷⁵

6. Self-assembly nature in AuNCs and CDs

6.1. AuNCs

The first report on AIE-type AuNCs came from the Xie group in 2012.^{14,28,102} They synthesised a series of $\text{Au}^0@ \text{Au}^+$ -thiolate core-shell nanoclusters with intense luminescence due to the presence of oligomeric aggregates of Au^+ -SR motifs in the surface shell.^{28,176} Strategies to enhance the luminescence from these AIE-type AuNCs are as follows: (1) creating elongated or interlocked Au^+ -ligand oligomeric structures in the exterior shell of the AuNCs; (2) increasing the amount of Au^+ leads to stronger aurophilic interactions;¹⁷⁷ (3) enhancing Au-ligand bonding to facilitate LMCT or LMMCT; (4) enhancing the stiffness of the protective shell by surface engineering,¹⁷⁸ (5) achieving tight inter-cluster stacking by self-assembly or crystallisation.^{28,179,180} This charge transfer mechanism is substantiated by its long average lifetime in microsecond units. The luminescence decay spectra of thiol-capped AuNCs follows a bi- or tri-exponential fit, which can be resolved into a long microsecond component (phosphorescence) and short nanosecond components (delayed fluorescence^{181,182} and prompt fluorescence).¹⁰² The luminescence from this family of AuNCs can be varied from the visible to the NIR regions simply by increasing the self-assembly or length of the Au^+ -ligand motifs on the surface shell.¹⁰² On the other hand, reductions in the self-assembly pattern or length of Au^+ -ligand motifs can shift the cluster luminescence from AIE-type phosphorescence to Au^0 core-directed fluorescence (Fig. 9).^{102,183}

6.2. CDs

One of the main topics of contemporary debates is where the CD fluorescence originates. The amount and arrangement of atoms that make up the core and surface of the CD ensemble must be heterogeneous even after extensive purification, and this influences their features, including optical properties. By the spectroscopic analysis of CDs at the single-particle level and ensemble-averaged level, Demchenko *et al.* deduced the 'H-aggregate model' where CDs are visualised as particles that hold surface-exposed fluorophores and can self-assemble to excitonic H-type aggregates very similar to conventional molecular fluorophore aggregates.¹¹³ Further, Kumbhakar's group assigned the huge excitation-dependent/independent emission spectra of citric-acid-derived CDs to multiple discrete electronic states of different aggregates. In this regard, the surface functional groups can determine the pattern and degree of aggregation.^{70,114} Subsequently, Anjali Devi *et al.* validated this H-aggregate theory for CD fluorescence by conducting spectroscopic analysis in solvents of different polarity or viscosity.⁴⁹ They observed evidence for J-type aggregates, H-type aggregates, and higher-order assemblies controlled by dipole-dipole, hydrogen bonding, and charge transfer interactions. Then, an amplified fluorescence quenching experiment on citric-acid-derived CDs with transition metal ions reveals that pyridinic nitrogen bases play a pivotal role in the construction of J-like emissive aggregates.¹¹⁵ Likewise, oxygen-rich surface functional groups are involved in H-like aggregates. The most intense emission, in this case, is provided by the J-like assembly of H aggregates. The heterogeneous aggregate structure in solution is substantiated by the fluorescence decay spectra of citric-acid-derived CDs, following a multi-exponential fit that can be resolved into nanosecond components. These results demand further detailed and cautious investigation on the aggregate chemistry of CDs originating from other precursors.

7. Conclusions and perspectives

Recent studies have highlighted the unique features, facile synthesis, and surface modification of ultra-small nanoparticles, fuelling their popularity. These materials offer significant technological benefits over traditional fluorophores, leading to several potential applications, ranging from sensors to catalysts and energy devices. Nevertheless, despite these advantageous features, ultra-small nanomaterial sensors face issues concerning their large-scale fabrication and commercialisation. (See Table 1 for some patented ultra-small nanoparticle sensors.) There is anxiety regarding the safety of ultra-small nanoparticle constituents as well as their assay reproducibility.

It is necessary to implement proper standard synthetic protocols for CDs and AuNCs, ensuring consistent size, shape, composition, and surface modification to facilitate better immobilisation, enhanced sensitivity, and specific interactions. The relationship between structure and optical properties in these new materials is unclear. The formation of excited states that affect absorption and fluorescence emission spectra is not

Table 1 Examples of ultra-small nanoparticles for fluorescence sensing that have been patented. (Results acquired from Espacenet)

Sl no.	Title	Organisation	Ultra-small nanoparticle and its amendments	Analyte	Mechanism of sensing/detection strategy	Point of care device	Patent No.
1.	Method and compositions for biosensing	University of North Carolina at Charlotte, USA	Metal-doped core-shell quantum dots (Groups II-VI/ Groups III-V)	Analyte (eg: nucleic acid)	Fluorescence turn-on/turn-off	—	US2020240992A1
2.	Biocompatible quantum dot sensor	Veinot Jonathan, Robidillo Christopher Jayt	Green fluorescent protein and silicon quantum dots	Analyte	Fluorescence turn-off	—	US2022050099A1
3	Gold/quantum dot nanoprobe for detecting active ricin in complex matrix and use thereof	Beijing Center for Disease Prevention and Control	Gold nanoparticle quenched quantum dots	Active ricin	Fluorescence turn-on	—	WO2020147735A1
4	Method for the detection of silica nanoparticles and/or graphene nanoplates	Fondazione st Italiano Tecnologia; Inail Istituto Naz per Lassicurazione Contro Gli Infortuni Sul Lavoro; Italy	Carbon dots	Silica nanoparticle/graphene nanoplates	Fluorescence turn-off	—	WO2022219461A1
5	Nanocomposite fluorescent probe for visibly detecting quercetin and preparation method thereof	Infotumi Sul Lavoro; China	Carbon dots/rhodamine 6G	Quercetin	Fluorescence turn-off	Filter paper	WO2020186503A1
6	Detection kit and method of detecting abused drugs	National Taiwan University, Taiwan	Carbon dots from D-phenylalanine	Nimetazepam, flunitrazepam, clonazepam, nitrazepam, and nitro-substituted benzodiazepine	Fluorescence turn-on	Detection kit	US11360082B2
7	Fluorescence test paper for detecting mercury ions and detection method using the same	Jiangsu University, China	Carbon dots and copper nanoclusters	Mercury ions	Fluorescence ratiometric sensor	Test paper	US11378519B2
8	Carbon quantum dots with improved optical properties	Juame I University, Spain	Carbon dots (boronic acid functionalised)	Monosaccharide, (eg: glucose)	Fluorescence sensor (turn-off)	—	WO2020254550A1
9	Gold nanoclusters, dopamine biosensors including them, and methods for diagnosing neurological diseases using the same	Industry Academic Cooperation Foundation of Yonsei University, Korea	Glutathione stabilised gold nanocluster (functionalised with benzyl chlorformate and aminobenzoboroxole)	Dopamine, glycoproteins	Fluorescence sensor (turn-on)	—	US20230341420A1
10	Device for visual detection of bilirubin	Indian Institute of Technology, Guwahati, India	Cu ²⁺ ion quenched chitosan stabilised gold nanocluster	Bilirubin	Fluorescence sensor (turn-on)	Polymer membrane strip	US11268967

fully understood. There is a significant drive to identify a unified emission mechanism hidden behind these nano-materials to achieve reliable control over their material performance. Remarkably, the excitation-dependent emission of CDs remained an active debate topic before the research community. In this aspect, an emissive aggregate model for fluorescence is still a growing area of research not only for CDs but also for AuNCs.

With the advancement in machine learning technology, many ultra-small luminescent sensors are being applied in portable, artificial-intelligence-based smart devices.¹⁸⁴ However, such on-site devices suffer from issues of biofouling and limited recyclability, which restrict their usage in regular real-time monitoring.¹⁸⁵ Existing research should be directed to transfer overall analyte detection strategies from the benchtop in the laboratory to simple smart on-site devices to develop lab-independent, decentralized, personalised analytical techniques that are economic, quick, high output, and mobile. Smart and innovative devices can also be made available which can be easily linked to smartphones and tablets. Future research will shed more light on their limitations and shortcomings.

Conflicts of interest

The author(s) declare no competing financial interest.

Acknowledgements

The authors gratefully acknowledge the encouragement, generous support, and facilities from the Professor & Head of the host institutions. The authors are grateful to Ms R. R. Anjana for her assistance in developing pictorial representations in the manuscript. J. S. Anjali Devi acknowledges the Department of Science and Technology, Govt. of India for the DST-INSPIRE Fellowship (IF 150393).

References

- 1 J. R. Lakowicz, *Principles of Fluorescence Spectroscopy*, Springer US, 2013.
- 2 M. G. Lagorio, G. B. Cordon and A. Iriel, *Photochem. Photobiol. Sci.*, 2015, **14**, 1538–1559.
- 3 Granite, Fluorescence, Phosphorescence and Photoluminescence Differences, <https://www.edinst.com/blog/photoluminescence-differences/>, (accessed 14 February 2024).
- 4 W. J. Peveler and W. R. Algar, *ACS Chem. Biol.*, 2018, **13**, 1752–1766.
- 5 Z. Yang, J. Cao, Y. He, J. H. Yang, T. Kim, X. Peng and J. S. Kim, *Chem. Soc. Rev.*, 2014, **43**, 4563–4601.
- 6 X. Sun, Y. Wang and Y. Lei, *Chem. Soc. Rev.*, 2015, **44**, 8019–8061.
- 7 T. L. Mako, J. M. Racicot and M. Levine, *Chem. Rev.*, 2019, **119**, 322–477.
- 8 Z. Li, Q. Sun, Y. Zhu, B. Tan, Z. P. Xu and S. X. Dou, *J. Mater. Chem. B*, 2014, **2**, 2793–2818.
- 9 S. Li, J. Wei, Q. Yao, X. Song, J. Xie and H. Yang, *Chem. Soc. Rev.*, 2023, **52**, 1672–1696.
- 10 Y. Huang, L. Fuksman and J. Zheng, *Dalton Trans.*, 2018, **47**, 6267–6273.
- 11 L. Zhang and E. Wang, *Nano Today*, 2014, **9**, 132–157.
- 12 S. Y. Lim, W. Shen and Z. Gao, *Chem. Soc. Rev.*, 2015, **44**, 362–381.
- 13 C. I. M. Santos, I. F. A. Mariz, S. N. Pinto, G. Gonçalves, I. Bdkin, P. A. A. P. Marques, M. G. P. M. S. Neves, J. M. G. Martinho and E. M. S. Maçôas, *Nanoscale*, 2018, **10**, 12505–12514.
- 14 N. Goswami, Q. Yao, Z. Luo, J. Li, T. Chen and J. Xie, *J. Phys. Chem. Lett.*, 2016, **7**, 962–975.
- 15 X. Michalet, F. F. Pinaud, L. A. Bentolila, J. M. Tsay, S. Doose, J. J. Li, G. Sundaresan, A. M. Wu, S. S. Gambhir and S. Weiss, *Science*, 2005, **307**, 538.
- 16 V. Biju, *Chem. Soc. Rev.*, 2014, **43**, 744–764.
- 17 M. G. Bawendi, M. L. Steigerwald and L. E. Brus, *Annu. Rev. Phys. Chem.*, 1990, **41**, 477–496.
- 18 Q. Xu, T. Kuang, Y. Liu, L. Cai, X. Peng, T. Sreenivasan Sreepasad, P. Zhao, Z. Yu and N. Li, *J. Mater. Chem. B*, 2016, **4**, 7204–7219.
- 19 H. Li, X. He, Z. Kang, H. Huang, Y. Liu, J. Liu, S. Lian, C. H. A. Tsang, X. Yang and S.-T. Lee, *Angew. Chem., Int. Ed.*, 2010, **49**, 4430–4434.
- 20 K. Jiang, S. Sun, L. Zhang, Y. Lu, A. Wu, C. Cai and H. Lin, *Angew. Chem., Int. Ed.*, 2015, **54**, 5360–5363.
- 21 Z. Gan, H. Xu and Y. Hao, *Nanoscale*, 2016, **8**, 7794–7807.
- 22 G. Eda, Y.-Y. Lin, C. Mattevi, H. Yamaguchi, H.-A. Chen, I.-S. Chen, C.-W. Chen and M. Chhowalla, *Adv. Mater.*, 2010, **22**, 505–509.
- 23 M. S. Mathew, A. Baksi, T. Pradeep and K. Joseph, *Biosens. Bioelectron.*, 2016, **81**, 68–74.
- 24 J. Zheng, J. T. Petty and R. M. Dickson, *J. Am. Chem. Soc.*, 2003, **125**, 7780–7781.
- 25 J. Zheng, C. Zhang and R. M. Dickson, *Phys. Rev. Lett.*, 2004, **93**, 077402.
- 26 H. Kawasaki, K. Hamaguchi, I. Osaka and R. Arakawa, *Adv. Funct. Mater.*, 2011, **21**, 3508–3515.
- 27 Y. Yu, Z. Luo, C. S. Teo, Y. N. Tan and J. Xie, *Chem. Commun.*, 2013, **49**, 9740–9742.
- 28 Z. Luo, X. Yuan, Y. Yu, Q. Zhang, D. T. Leong, J. Y. Lee and J. Xie, *J. Am. Chem. Soc.*, 2012, **134**, 16662–16670.
- 29 M. Zhu, C. M. Aikens, F. J. Hollander, G. C. Schatz and R. Jin, *J. Am. Chem. Soc.*, 2008, **130**, 5883–5885.
- 30 A. P. Demchenko, V. I. Tomin and P.-T. Chou, *Chem. Rev.*, 2017, **117**, 13353–13381.
- 31 J. Xie, Y. Zheng and J. Y. Ying, *J. Am. Chem. Soc.*, 2009, **131**, 888–889.
- 32 J. Annie Ho, H.-C. Chang and W.-T. Su, *Anal. Chem.*, 2012, **84**, 3246–3253.
- 33 Y. Dong, H. Pang, H. B. Yang, C. Guo, J. Shao, Y. Chi, C. M. Li and T. Yu, *Angew. Chem., Int. Ed.*, 2013, **52**, 7800–7804.
- 34 X. Li, S. Zhang, S. A. Kulinich, Y. Liu and H. Zeng, *Sci. Rep.*, 2014, **4**, 4976.

- 35 Y.-P. Sun, B. Zhou, Y. Lin, W. Wang, K. A. S. Fernando, P. Pathak, M. J. Meziani, B. A. Harruff, X. Wang, H. Wang, P. G. Luo, H. Yang, M. E. Kose, B. Chen, L. M. Veca and S.-Y. Xie, *J. Am. Chem. Soc.*, 2006, **128**, 7756–7757.
- 36 S. Chatteraj and K. Bhattacharyya, *J. Phys. Chem. C*, 2014, **118**, 22339–22346.
- 37 A. Das, D. Roy, C. K. De and P. K. Mandal, *Phys. Chem. Chem. Phys.*, 2018, **20**, 2251–2259.
- 38 X. Wen, A. Sitt, P. Yu, Y.-R. Toh and J. Tang, *Phys. Chem. Chem. Phys.*, 2012, **14**, 3505–3512.
- 39 P. Yu, X. Wen, Y.-R. Toh and J. Tang, *J. Phys. Chem. C*, 2012, **116**, 6567–6571.
- 40 P. Yu, X. Wen, Y.-R. Toh and J. Tang, *J. Phys. Chem. C*, 2012, **116**, 25552–25557.
- 41 C. A. Leatherdale and M. G. Bawendi, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 2001, **63**, 165315.
- 42 R. Zhou, M. Shi, X. Chen, M. Wang and H. Chen, *Chem. – Eur. J.*, 2009, **15**, 4944–4951.
- 43 C. Yi, H. Zheng, P. J. Herbert, Y. Chen, R. Jin and K. L. Knappenberger, *J. Phys. Chem. C*, 2017, **121**, 24894–24902.
- 44 N. Basu and D. Mandal, *J. Phys. Chem. C*, 2018, **122**, 18732–18741.
- 45 C. J. Reckmeier, Y. Wang, R. Zboril and A. L. Rogach, *J. Phys. Chem. C*, 2016, **120**, 10591–10604.
- 46 A. Sciortino, E. Marino, B. van Dam, P. Schall, M. Cannas and F. Messina, *J. Phys. Chem. Lett.*, 2016, **7**, 3419–3423.
- 47 S. Mukherjee, E. Prasad and A. Chadha, *Phys. Chem. Chem. Phys.*, 2017, **19**, 7288–7296.
- 48 T. Zhang, J. Zhu, Y. Zhai, H. Wang, X. Bai, B. Dong, H. Wang and H. Song, *Nanoscale*, 2017, **9**, 13042–13051.
- 49 J. S. Anjali Devi, R. S. Aparna, R. R. Anjana, J. Nebu, S. M. Anju and S. George, *J. Phys. Chem. A*, 2019, **123**, 7420–7429.
- 50 M. J. Ruedas-Rama and E. A. H. Hall, *Anal. Chem.*, 2008, **80**, 8260–8268.
- 51 Y. Lou, Y. Zhao, J. Chen and J.-J. Zhu, *J. Mater. Chem. C*, 2014, **2**, 595–613.
- 52 S. J. Clarke, C. A. Hollmann, Z. Zhang, D. Suffern, S. E. Bradforth, N. M. Dimitrijevic, W. G. Minarik and J. L. Nadeau, *Nat. Mater.*, 2006, **5**, 409–417.
- 53 K. Abha, I. S. Sumithra, S. Suji, R. R. Anjana, J. S. Anjali Devi, J. Nebu, G. M. Lekha, R. S. Aparna and S. George, *Anal. Bioanal. Chem.*, 2020, **412**, 5671–5681.
- 54 K. Abha, J. Nebu, J. S. Anjali Devi, R. S. Aparna, R. R. Anjana, A. O. Aswathy and S. George, *Sens. Actuators, B*, 2019, **282**, 300–308.
- 55 K. Palaniappan, S. A. Hackney and J. Liu, *Chem. Commun.*, 2004, 2704–2705.
- 56 X. Wang, L. Cao, F. Lu, M. J. Meziani, H. Li, G. Qi, B. Zhou, B. A. Harruff, F. Kermarrec and Y.-P. Sun, *Chem. Commun.*, 2009, 3774–3776.
- 57 J. Gong, X. Lu and X. An, *RSC Adv.*, 2015, **5**, 8533–8536.
- 58 K. Qu, J. Wang, J. Ren and X. Qu, *Chem. – Eur. J.*, 2013, **19**, 7243–7249.
- 59 Y. Tao, Y. Lin, J. Ren and X. Qu, *Biosens. Bioelectron.*, 2013, **42**, 41–46.
- 60 M. Tomasulo, I. Yildiz and F. M. Raymo, *J. Phys. Chem. B*, 2006, **110**, 3853–3855.
- 61 Y. Bao, C. Zhong, D. M. Vu, J. P. Temirov, R. B. Dyer and J. S. Martinez, *J. Phys. Chem. C*, 2007, **111**, 12194–12198.
- 62 Y. Guo, Z. Wang, H. Shao and X. Jiang, *Carbon*, 2013, **52**, 583–589.
- 63 W. Kong, H. Wu, Z. Ye, R. Li, T. Xu and B. Zhang, *J. Lumin.*, 2014, **148**, 238–242.
- 64 S. Dutta Choudhury, J. M. Chethodil, P. M. Gharat and P. K. Praseetha, *J. Phys. Chem. Lett.*, 2017, **8**, 1389–1395.
- 65 C. V. Durgadas, C. P. Sharma and K. Sreenivasan, *Analyst*, 2011, **136**, 933–940.
- 66 L.-Y. Chen, C.-W. Wang, Z. Yuan and H.-T. Chang, *Anal. Chem.*, 2015, **87**, 216–229.
- 67 J. S. Anjali Devi, S. Salini, A. H. Anulekshmi, G. L. Praveen and G. Sony, *Sens. Actuators, B*, 2017, **246**, 943–951.
- 68 J. Nebu, J. S. Anjali Devi, R. S. Aparna, B. Aswathy, G. M. Lekha and G. Sony, *Anal. Bioanal. Chem.*, 2019, **411**, 997–1007.
- 69 J. Nebu, K. S. Anu, J. S. Anjali Devi, R. S. Aparna, A. O. Aswathy, G. M. Lekha and G. Sony, *Microchem. J.*, 2019, **146**, 12–19.
- 70 A. Sharma, T. Gadly, S. Neogy, S. K. Ghosh and M. Kumbhakar, *J. Phys. Chem. Lett.*, 2017, **8**, 1044–1052.
- 71 A. O. Aswathy, S. M. Anju, J. Jayakrishna, N. S. Vijila, J. S. Anjali Devi, B. Anjitha and S. George, *J. Fluoresc.*, 2020, **30**, 1337–1344.
- 72 Y. Dong, J. Cai, X. You and Y. Chi, *Analyst*, 2015, **140**, 7468–7486.
- 73 M. Masteri-Farahani and K. Khademabbasi, *J. Lumin.*, 2018, **204**, 130–134.
- 74 P. Wu and X.-P. Yan, *Chem. Soc. Rev.*, 2013, **42**, 5489–5521.
- 75 R. Jin and K. Nobusada, *Nano Res.*, 2014, **7**, 285–300.
- 76 Q. Zhou, Y. Lin, M. Xu, Z. Gao, H. Yang and D. Tang, *Anal. Chem.*, 2016, **88**, 8886–8892.
- 77 Z. Gan, H. Xu and Y. Hao, *Nanoscale*, 2016, **8**, 7794–7807.
- 78 L. Bao, C. Liu, Z. Zhang and D. Pang, *Adv. Mater.*, 2015, **27**, 1663–1667.
- 79 H. Ding, S.-B. Yu, J.-S. Wei and H.-M. Xiong, *ACS Nano*, 2016, **10**, 484–491.
- 80 Q. Zhang, R. Wang, B. Feng, X. Zhong and K. (Ken) Ostrikov, *Nat. Commun.*, 2021, **12**, 6856.
- 81 Y. Tao, M. Li, J. Ren and X. Qu, *Chem. Soc. Rev.*, 2015, **44**, 8636–8663.
- 82 The Nobel Prize in Chemistry 2023, <https://www.nobelprize.org/prizes/chemistry/2023/summary/>, (accessed 7 February 2024).
- 83 U. Resch-Genger, M. Grabolle, S. Cavaliere-Jaricot, R. Nitschke and T. Nann, *Nat. Methods*, 2008, **5**, 763.
- 84 A. Onushchenko and A. Ekimov, *JETP Lett.*, 1981, **34**, 345.
- 85 C. B. Murray, D. J. Norris and M. G. Bawendi, *J. Am. Chem. Soc.*, 1993, **115**, 8706–8715.
- 86 Z. A. Peng and X. Peng, *J. Am. Chem. Soc.*, 2001, **123**, 183–184.
- 87 L. Qu, Z. A. Peng and X. Peng, *Nano Lett.*, 2001, **1**, 333–337.
- 88 H. R. Chandan and R. G. Balakrishna, *Sens. Actuators, B*, 2018, **258**, 1191–1214.

- 89 J. Jasieniak, C. Bullen, J. van Embden and P. Mulvaney, *J. Phys. Chem. B*, 2005, **109**, 20665–20668.
- 90 Y. Orimoto, K. Watanabe, K. Yamashita, M. Uehara, H. Nakamura, T. Furuya and H. Maeda, *J. Phys. Chem. C*, 2012, **116**, 17885–17896.
- 91 J. Zhou, Y. Liu, J. Tang and W. Tang, *Mater. Today*, 2017, **20**, 360–376.
- 92 B. Mahler, P. Spinicelli, S. Buil, X. Quelin, J.-P. Hermier and B. Dubertret, *Nat. Mater.*, 2008, **7**, 659–664.
- 93 Isnaeni, K. H. Kim, D. L. Nguyen, H. Lim, P. T. Nga and Y.-H. Cho, *Appl. Phys. Lett.*, 2011, **98**, 012109.
- 94 X. Brokmann, J.-P. Hermier, G. Messin, P. Desbailles, J.-P. Bouchaud and M. Dahan, *Phys. Rev. Lett.*, 2003, **90**, 120601.
- 95 P. Reiss, J. Bleuse and A. Pron, *Nano Lett.*, 2002, **2**, 781–784.
- 96 A. L. Efros and L. E. Brus, *ACS Nano*, 2021, **15**, 6192–6210.
- 97 R. Jin, C. Zeng, M. Zhou and Y. Chen, *Chem. Rev.*, 2016, **116**, 10346–10413.
- 98 Y. Gu, S. Tang, X. Liu, X. Liang, Q. Zhu, H. Wu, X. Yang, W. Jin, H. Chen, C. Liu, Y. Zhu and J. Ma, *J. Mater. Chem. A*, 2024, **12**, 4460–4472.
- 99 J. Li, T. Chen, K. Lim, L. Chen, S. A. Khan, J. Xie and X. Wang, *Adv. Intell. Syst.*, 2019, **1**, 1900029.
- 100 T. Chen, J. Li, P. Cai, Q. Yao, Z. Ren, Y. Zhu, S. Khan, J. Xie and X. Wang, *Nano Res.*, 2023, **16**, 4188–4196.
- 101 O. Voznyy, L. Levina, J. Z. Fan, M. Askerka, A. Jain, M.-J. Choi, O. Ouellette, P. Todorović, L. K. Sagar and E. H. Sargent, *ACS Nano*, 2019, **13**, 11122–11128.
- 102 Z. Wu, Q. Yao, O. J. H. Chai, N. Ding, W. Xu, S. Zang and J. Xie, *Angew. Chem.*, 2020, **132**, 10020–10025.
- 103 X. Xu, R. Ray, Y. Gu, H. J. Ploehn, L. Gearheart, K. Raker and W. A. Scrivens, *J. Am. Chem. Soc.*, 2004, **126**, 12736–12737.
- 104 J. Zhou, C. Booker, R. Li, X. Zhou, T.-K. Sham, X. Sun and Z. Ding, *J. Am. Chem. Soc.*, 2007, **129**, 744–745.
- 105 Z. Ma, H. Ming, H. Huang, Y. Liu and Z. Kang, *New J. Chem.*, 2012, **36**, 861–864.
- 106 S. Qu, X. Wang, Q. Lu, X. Liu and L. Wang, *Angew. Chem., Int. Ed.*, 2012, **51**, 12215–12218.
- 107 K. Holá, M. Sudolská, S. Kalytchuk, D. Nachtigallová, A. L. Rogach, M. Otyepka and R. Zbořil, *ACS Nano*, 2017, **11**, 12402–12410.
- 108 R. D. Senanayake, X. Yao, C. E. Froehlich, M. S. Cahill, T. R. Sheldon, M. McIntire, C. L. Haynes and R. Hernandez, *J. Chem. Inf. Model.*, 2022, **62**, 5918–5928.
- 109 W. Kasprzyk, S. Bednarsz and D. Bogdał, *Chem. Commun.*, 2013, **49**, 6445.
- 110 W. Kasprzyk, T. Świergosz, S. Bednarsz, K. Walas, N. V. Bashmakova and D. Bogdał, *Nanoscale*, 2018, **10**, 13889–13894.
- 111 J. Schneider, C. J. Reckmeier, Y. Xiong, M. von Seckendorff, A. S. Susha, P. Kasák and A. L. Rogach, *J. Phys. Chem. C*, 2017, **121**, 2014–2022.
- 112 A. Das, V. Gude, D. Roy, T. Chatterjee, C. K. De and P. K. Mandal, *J. Phys. Chem. C*, 2017, **121**, 9634–9641.
- 113 A. P. Demchenko and M. O. Dekaliuk, *Nanoscale*, 2016, **8**, 14057–14069.
- 114 A. Sharma, T. Gadly, A. Gupta, A. Ballal, S. K. Ghosh and M. Kumbhakar, *J. Phys. Chem. Lett.*, 2016, **7**, 3695–3702.
- 115 J. S. Anjali Devi, R. S. Aparna, R. R. Anjana, N. S. Vijila, J. Jayakrishna and S. George, *Photochem. Photobiol. Sci.*, 2020, **19**, 207–216.
- 116 N. J. Hestand and F. C. Spano, *Chem. Rev.*, 2018, **118**, 7069–7163.
- 117 J. B. Essner, J. A. Kist, L. Polo-Parada and G. A. Baker, *Chem. Mater.*, 2018, **30**, 1878–1887.
- 118 P. Zhu, K. Tan, Q. Chen, J. Xiong and L. Gao, *Chem. Mater.*, 2019, **31**, 4732–4742.
- 119 L. Hao, Y. Shen, X. Chen, X. Yang, Q. Du, Y. Bian, L. Chen, K. Tang, R. Zhang, Y. Zheng and S. Gu, *Microchem. J.*, 2019, **148**, 684–690.
- 120 D. Ungor, K. Horváth, I. Dékány and E. Csapó, *Sens. Actuators, B*, 2019, **288**, 728–733.
- 121 Z. Zhang, T. Liu, S. Wang, J. Ma, T. Zhou, F. Wang, X. Wang and G. Zhang, *J. Photochem. Photobiol., A*, 2019, **370**, 89–93.
- 122 J. Wei, Y. Yuan, H. Li, D. Hao, C. Sun, G. Zheng and R. Wang, *New J. Chem.*, 2018, **42**, 18787–18793.
- 123 F. Abbasi, A. Akbarinejad and N. Alizadeh, *Spectrochim. Acta, Part A*, 2019, **216**, 230–235.
- 124 Y. Ma, W. Lv, Y. Chen, M. Na, J. Liu, Y. Han, S. Ma and X. Chen, *New J. Chem.*, 2019, **43**, 5070–5076.
- 125 M. Santhosh, S. R. Chinnadayaala, A. Kakoti and P. Goswami, *Biosens. Bioelectron.*, 2014, **59**, 370–376.
- 126 F. Xue, L. Liu, Y. Mi, H. Han and J. Liang, *RSC Adv.*, 2016, **6**, 10215–10220.
- 127 S. K. Anand, U. Sivasankaran, A. R. Jose and K. G. Kumar, *Spectrochim. Acta, Part A*, 2019, **213**, 410–415.
- 128 C.-P. Liu, T.-H. Wu, C.-Y. Liu, H.-J. Cheng and S.-Y. Lin, *J. Mater. Chem. B*, 2015, **3**, 191–197.
- 129 S. Gogoi and R. Khan, *Phys. Chem. Chem. Phys.*, 2018, **20**, 16501–16509.
- 130 G. H. Shi, Z. B. Shang, Y. Wang, W. J. Jin and T. C. Zhang, *Spectrochim. Acta, Part A*, 2008, **70**, 247–252.
- 131 R. Das, G. Rajender and P. K. Giri, *Phys. Chem. Chem. Phys.*, 2018, **20**, 4527–4537.
- 132 D. C. Harris, *Quantitative chemical analysis*, Macmillan, 2010.
- 133 R. Freeman, T. Finder, L. Bahshi and I. Willner, *Nano Lett.*, 2009, **9**, 2073–2076.
- 134 C. Tang, J. Zhou, Z. Qian, Y. Ma, Y. Huang and H. Feng, *J. Mater. Chem. B*, 2017, **5**, 1971–1979.
- 135 A. Cayuela, M. Laura Soriano and M. Valcárcel, *Analyst*, 2016, **141**, 2682–2687.
- 136 S. Mondal and P. Purkayastha, *J. Phys. Chem. C*, 2016, **120**, 14365–14371.
- 137 M. Luo, Y. Hua, Y. Liang, J. Han, D. Liu, W. Zhao and P. Wang, *Biosens. Bioelectron.*, 2017, **98**, 195–201.
- 138 Z. Saberi, B. Rezaei and A. A. Ensafi, *Microchim. Acta*, 2019, **186**, 273.
- 139 E. Vaishnavi and R. Renganathan, *Analyst*, 2014, **139**, 225–234.
- 140 K. Pyo, M. F. Matus, S. Malola, E. Hulkko, J. Alaranta, T. Lahtinen, H. Häkkinen and M. Pettersson, *Nanoscale Adv.*, 2022, **4**, 4579–4588.

- 141 X. Zhang, X. Chen, S. Kai, H.-Y. Wang, J. Yang, F.-G. Wu and Z. Chen, *Anal. Chem.*, 2015, **87**, 3360–3365.
- 142 S. Ma, Y. Chen, J. Feng, J. Liu, X. Zuo and X. Chen, *Anal. Chem.*, 2016, **88**, 10474–10481.
- 143 D. J. Heintzelman, S. A. Nelson and K. L. Knappenberger, *J. Phys. Chem. Lett.*, 2024, **15**, 2951–2956.
- 144 Y. Miao, N. Gan, T. Li, Y. Cao, F. Hu and Y. Chen, *Sens. Actuators, B*, 2016, **222**, 1066–1072.
- 145 L. Chang, X. He, L. Chen and Y. Zhang, *Sens. Actuators, B*, 2017, **250**, 17–23.
- 146 M. Shamsipur, V. Nasirian, A. Barati, K. Mansouri, A. Vaisi-Raygani and S. Kashanian, *Anal. Chim. Acta*, 2017, **966**, 62–70.
- 147 Y.-S. Borghei, M. Hosseini, M. R. Ganjali and S. Hosseinkhani, *J. Pharm. Biomed. Anal.*, 2018, **152**, 81–88.
- 148 Y. Shu, N. Zheng, A.-Q. Zheng, T.-T. Guo, Y.-L. Yu and J.-H. Wang, *Anal. Chem.*, 2019, **91**, 4157–4163.
- 149 H. Ge, K. Zhang, H. Yu, J. Yue, L. Yu, X. Chen, T. Hou, K. A. Alamry, H. M. Marwani and S. Wang, *J. Fluoresc.*, 2018, **28**, 1405–1412.
- 150 X. Cheng, Y. Cen, G. Xu, F. Wei, M. Shi, X. Xu, M. Sohail and Q. Hu, *Microchim. Acta*, 2018, **185**, 144.
- 151 Y. Ding, J. Ling, H. Wang, J. Zou, K. Wang, X. Xiao and M. Yang, *Anal. Methods*, 2015, **7**, 7792–7798.
- 152 S. H. Qaddare and A. Salimi, *Biosens. Bioelectron.*, 2017, **89**, 773–780.
- 153 Y. Chawre, M. L. Satnami, A. B. Kujur, K. K. Ghosh, R. Nagwanshi, I. Karbhal, S. Pervez and M. K. Deb, *ACS Appl. Nano Mater.*, 2023, **6**, 8046–8058.
- 154 L. Qin, X. He, L. Chen and Y. Zhang, *ACS Appl. Mater. Interfaces*, 2015, **7**, 5965–5971.
- 155 A. R. Jose, A. E. Vikraman and K. Girish Kumar, *New J. Chem.*, 2017, **41**, 10828–10834.
- 156 S. Huang, F. Zhu, H. Qiu, Q. Xiao, Q. Zhou, W. Su and B. Hu, *Colloids Surf., B*, 2014, **117**, 240–247.
- 157 X. Yue, W. Zhu, S. Ma, S. Yu, Y. Zhang, J. Wang, Y. Wang, D. Zhang and J. Wang, *J. Agric. Food Chem.*, 2016, **64**, 706–713.
- 158 K.-Y. Huang, H.-X. He, S.-B. He, X.-P. Zhang, H.-P. Peng, Z. Lin, H.-H. Deng, X.-H. Xia and W. Chen, *Sens. Actuators, B*, 2019, **296**, 126656.
- 159 S. Govindaraju, S. R. Ankireddy, B. Viswanath, J. Kim and K. Yun, *Sci. Rep.*, 2017, **7**, 40298.
- 160 S. Chen, Y.-L. Yu and J.-H. Wang, *Anal. Chim. Acta*, 2018, **999**, 13–26.
- 161 S. Khezri, M. Bahram and N. Samadi, *Anal. Methods*, 2017, **9**, 6513–6524.
- 162 H.-C. Chang and J. A. Ho, *Anal. Chem.*, 2015, **87**, 10362–10367.
- 163 M. Zheng, Z. Xie, D. Qu, D. Li, P. Du, X. Jing and Z. Sun, *ACS Appl. Mater. Interfaces*, 2013, **5**, 13242–13247.
- 164 X. Y. Sun, M. J. Yuan, B. Liu and J. S. Shen, *RSC Adv.*, 2018, **8**, 19786–19790.
- 165 Z. Xu, Z. Wang, M. Liu, B. Yan, X. Ren and Z. Gao, *Spectrochim. Acta, Part A*, 2020, **232**, 118147.
- 166 X. Liu, X. Hu, Z. Xie, P. Chen, X. Sun, J. Yan and S. Zhou, *Anal. Methods*, 2016, **8**, 3236–3241.
- 167 K. Chaiendoo, S. Ittisanronnachai, V. Promarak and W. Ngeontae, *Carbon*, 2019, **146**, 728–735.
- 168 Z. Qian, J. Ma, X. Shan, H. Feng, L. Shao and J. Chen, *Chem. – Eur. J.*, 2014, **20**, 2254–2263.
- 169 H.-Y. Chang, Y.-T. Tseng, Z. Yuan, H.-L. Chou, C.-H. Chen, B.-J. Hwang, M.-C. Tsai, H.-T. Chang and C.-C. Huang, *Phys. Chem. Chem. Phys.*, 2017, **19**, 12085–12093.
- 170 Z. Huang, M. Wang, Z. Guo, H. Wang, H. Dong and W. Yang, *ACS Omega*, 2018, **3**, 12763–12769.
- 171 J.-G. You and W.-L. Tseng, *Anal. Chim. Acta*, 2019, **1078**, 101–111.
- 172 J. Fu, Y. Miao, D. Zhang, Y. Zhang, L. Meng, X. Ni, J. Shen and W. Qi, *Langmuir*, 2023, **39**, 13316–13324.
- 173 N. Gogoi, D. S. Agarwal, A. Sehgal, D. Chowdhury and R. Sakhuja, *ACS Omega*, 2017, **2**, 3816–3827.
- 174 M. Mou, Y. Wu, Q. Niu, Y. Wang, Z. Yan and S. Liao, *Chem. Commun.*, 2017, **53**, 3357–3360.
- 175 S. Pathan, M. Jalal, S. Prasad and S. Bose, *J. Mater. Chem. A*, 2019, **7**, 8510–8520.
- 176 Y. Yu, Z. Luo, D. M. Chevrier, D. T. Leong, P. Zhang, D. Jiang and J. Xie, *J. Am. Chem. Soc.*, 2014, **136**, 1246–1249.
- 177 Z. Wu, Y. Du, J. Liu, Q. Yao, T. Chen, Y. Cao, H. Zhang and J. Xie, *Angew. Chem., Int. Ed.*, 2019, **58**, 8139–8144.
- 178 K. Pyo, V. D. Thanthirige, K. Kwak, P. Pandurangan, G. Ramakrishna and D. Lee, *J. Am. Chem. Soc.*, 2015, **137**, 8244–8250.
- 179 T. Chen, S. Yang, J. Chai, Y. Song, J. Fan, B. Rao, H. Sheng, H. Yu and M. Zhu, *Sci. Adv.*, 2017, **3**, e1700956.
- 180 Z. Wu, Q. Yao, S. Zang and J. Xie, *ACS Mater. Lett.*, 2019, **1**, 237–248.
- 181 X. Wen, P. Yu, Y.-R. Toh, A.-C. Hsu, Y.-C. Lee and J. Tang, *J. Phys. Chem. C*, 2012, **116**, 19032–19038.
- 182 Z. Liu, L. Luo, J. Kong, E. Kahng, M. Zhou and R. Jin, *Nanoscale*, 2024, **16**, 7419–7426.
- 183 M. Sugiuchi, J. Maeba, N. Okubo, M. Iwamura, K. Nozaki and K. Konishi, *J. Am. Chem. Soc.*, 2017, **139**, 17731–17734.
- 184 S. Wei, B. wang, H. Zhang, C. Wang, S. Cui, X. Yin, C. Jiang and G. Sun, *Chem. Eng. J.*, 2023, **466**, 143103.
- 185 Y. Fan, X. Wang, T. Funk, I. Rashid, B. Herman, N. Bompoti, M. S. Mahmud, M. Chrysochoou, M. Yang, T. M. Vadas, Y. Lei and B. Li, *Environ. Sci. Technol.*, 2022, **56**, 13546–13564.