Organic & Biomolecular Chemistry

ROYAL SOCIETY OF CHEMISTRY

PAPER

View Article Online
View Journal | View Issue

Check for updates

Cite this: *Org. Biomol. Chem.*, 2024, **22**, 5134

Trivalent dialkylaminopyridine-catalyzed site-selective mono-*O*-acylation of partially-protected pyranosides†

Kalyan Dey and Narayanaswamy Jayaraman *

Received 11th April 2024,
Accepted 22nd May 2024

DOI: 10.1039/d4ob00599f

rsc.li/obc

This work demonstrates trivalent tris-(3-*N*-methyl-*N*-pyridyl propyl)amine (**1**) catalyzing the site-selective mono-*O*-acylation of glycopyranosides. Different acid anhydrides were used for the acylation of monosaccharides, mediated by catalyst **1**, at a loading of 1.5 mol%; the extent of site-selectivity and the yields of mono-*O*-acylation products were assessed. The reactions were performed between 2 and 10 h, depending on the nature of the acid anhydride, where the bulkier pivalic anhydride required a longer duration for acylation. The glycopyranosides are maintained as diols and triols, and from a set of experiments, the site-selectivity of acylations was observed to follow the intrinsic reactivities and stereochemistry of hydroxy functionalities. The trivalent catalyst **1** mediates the reactions with excellent site-selectivities for mono-*O*-acylation product formation in the studied glycopyranosides, in comparison to the monovalent *N*,*N*-dimethylamino pyridine (DMAP) catalyst. This study illustrates the benefits of the multivalency of catalytic moieties in catalysis.

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

Introduction

The roles of hydroxy functionality-rich carbohydrates in biological functions are diverse.[1–3] As a result, an understanding of the structures and functions of carbohydrates has the greatest potential to uncover many intricate biological processes.[4] The presence of multiple hydroxy functionalities increases the repertoire of complex oligosaccharides and polysaccharides exponentially. Enzymes exquisitely streamline the assembly of oligosaccharides and polysaccharides, whereas the corresponding synthetic equivalents are also developed impressively.[5] The chemical synthesis of complex oligosaccharides encounters the critical roles of protecting groups that mask the reactivities of hydroxy functionalities not involved in the assembly. Protecting group methods enable the chemical synthesis of oligosaccharides and polysaccharides in the most achievable homogeneous manner.[6,7] The pK_a of primary and secondary hydroxy functionalities are within 16–18, implying that reactivity differences are minimal among these functionalities. As a result, development of methods for the site-selective modification of carbohydrates,[8] including one-pot synthetic methods,[9] is important. An attractive approach in this direction is exploiting the intramolecular hydrogen bonding of hydroxy functionalities, which increases nucleophilicity. Early demonstrations of this concept are (i) DMAP-catalyzed acylation reactions using Ac_2O for the regioselective formation of the *C*3-OAc surrogate of octyl-β-D-glucopyranoside, as reported by Yoshida and co-workers.[10a] (ii) Carboxylic or sulfonic acid-linked DMAP-catalyzed site selective acylation at the *C*6-OH group in octyl glucopyranosides and galactopyranosides.[10b] The effect of a counter ion in DMAP-catalyzed acylations was studied by Albert and Kattnig.[11] Acylation is favored at hydroxy functionalities either at *C*3, *C*4 or *C*6, depending on the nature of the counter anion through the formation of non-covalently linked cyclic transition state between acyl and hydroxy functionalities. The 'directed protecting group-induced regioselective functionalization' of partially protected carbohydrates was reported by Moitessier and co-workers, wherein the protecting group at *C*6 forms intramolecular hydrogen bonding preferentially at the *C*3 hydroxy functionality and facilitates regioselective acylation.[12] A good regioselectivity was observed for the pivaloylation of 4,6-*O*-benzylidene-protected glycoside derivatives.[13] Several other acylating agents, such as benzoyl imidazole,[14] BzCN,[15] and 1-(benzoyloxy)benzotriazole (BzOBt),[16] were developed in the regioselective acylation of 4,6-*O*-protected pyranoside derivatives. The selectivity was largely dependent on the reaction conditions, acidity and nucleophilicity of reactive functionalities. In the case of BzCN, benzoylation occurs at the axial hydroxy functionality in diols and triols in the presence of DMAP (10 mol%, −78 °C).[17] The formation

Indian Institute of Science, Bangalore 560012, India. E-mail: jayaraman@iisc.ac.in; Tel: +91-80-2293-2578

†Electronic supplementary information (ESI) available: HPLC traces of crude reaction mixture, ^{1}H and ^{13}C NMR spectra of all compounds. See DOI: https://doi.org/10.1039/d4ob00599f

This journal is © The Royal Society of Chemistry 2024

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

of monodentate hydrogen bonding with cyanide moiety and acylation at the acidic axial hydroxy functionality under kinetic control appeared to favor the regioselectivity, termed as the 'cyanide effect'.

Several structurally-modified DMAP derivatives have been reported to achieve the site-selective functionalization and enantioselective acylation of racemic alcohol.[18–26] Kawabata and co-workers[27] developed a 4-pyrrolidinopyridine (PPY)-based C_2-symmetric chiral catalyst based on L-tryptophan. A good site-selectivity at the hydroxyl functionality at *C*4 was observed for the installation of isobutyrate in octyl-β-D-glucopyranoside. Kirsch and co-workers generated a library of DMAP-based oligopeptides for the site-selective mono-*O*-benzoylation in favor of functionalization at *C*2 in benzylidene-protected α-D-glucopyranoside and at *C*3 in benzylidene-protected α-D-galactopyranoside derivatives.[28] Besides DMAP-derived catalysts, *N*-methyl imidazole (NMI) has been developed as an acyl transfer reagent. NMI-appended peptide-based catalysts have been developed for the regioselective functionalization of monosaccharides, as reported by Miller and co-workers.[29] Schreiner and co-workers recently reported site-selective acylation employing oligopeptide and photoswitchable azo-peptide NMI-catalyst.[30] NMI-based chiral oxazolidines catalyze the acylation of the hydroxyl functionality at *C*3 and the intrinsically less reactive functionality at the *C*2 carbon in carbohydrate derivatives.[31] Chiral benzotetramisole (BTM) catalyst was employed by Tang and co-workers[32a] to accomplish the regioselective acylation of the *trans*-1,2-diol moiety in pyranoside derivatives. The hydroxyl functionality at *C*2 or *C*3 carbon underwent the reaction selectively by choice of the enantiomeric catalyst. Regioselectivity was observed in this case due to the lone pair of electrons-cation interaction between the acylated catalyst intermediate and either an ether or hydroxyl oxygen of the substrate. In a subsequent study, *S*-adamantyl group-directed acylation using the same catalyst was also reported.[32b]

Chiral NHC catalysis[33] and chiral NHC-boronic acid dual catalysis[34] using aldehyde as an acyl source were applied successfully for the site-selective acylation of carbohydrates. Miller and co-workers developed the site-selective functionalization of methyl 4,6-*O*-benzylidene-protected pyranosides employing a chiral Ph-Box ligand with Cu(II).[35] Site-selectivity was dependent on the chirality of the ligands. Using similar Cu(II)–Ph–Box complexes, Dong and co-workers reported the site-selective acylation of pyranoside-derived triols and furanoside-derived diols.[36] Chiral phosphoric acid (CPA) catalyzed the site-selective acetalization of pyranoside-derived 1,2-diols.[37a] Recently, a polystyrene-supported reusable CPA was used for this purpose and variably protected saccharide derivatives were accomplished in an one-pot reaction.[37b] Methods were also developed for the site-selective functionalization of unprotected or partially protected pyranosides using salts of tin-,[38,39] iron-,[40] silicon-[41] and copper-[42] silver-based[43] reagents. Although organotin reagents are considered largely for the regioselective functionalization of unprotected carbohydrates, a major shortcoming is the inherent toxicity of tin reagents. Organoboron reagents, developed by Taylor and Shimada groups, are non-toxic and used for site-selective acylation,[44] sulfamoylation,[45a] sulfation,[45b] sulfonylation,[46,47] carbamoylation,[48] and glycosylation[49,50] for unprotected carbohydrates having the *cis*-1,2-diol moiety. The above developments illustrate the necessity to uncover methods that facilitate the site-selective derivatization of multi-functional carbohydrate moieties. An important functionalization in this direction is the protection of the hydroxyl functionality as esters, which are used routinely to derive, for example, partially-protected carbohydrate derivatives. The ester protecting groups are installed by treating the alcohol functionality with carboxylic acid anhydrides and acid chlorides in the presence of DMAP (cat.).[51] Several polymer-bound (dialkylamino)pyridine catalysts have been reported to act as nucleophilic catalysts in organic transformations.[52] Portnoy and co-workers developed a series of *p*-aminopyridine[53a] and imidazole-based[53b] nucleophilic organocatalysts, functionalized further with either oligoether or long alkyl moieties for the site-selective acylation and phosphorylation of amphiphilic diol and complex natural products.[53c,d] The choice of base and catalyst were emphasized in order to achieve orthogonal site-selectivities.

Multivalency in catalysis has gained importance in contemporary developments. The covalent attachment of multiple catalytic moieties onto a branched structure leads to efficiencies in catalysis that are over and above the statistical increases in the moieties.[54] Elegant examples are in C–C bond formations enabled by multivalent organometallic catalysts.[55] Multivalent organocatalysis by a variety of dendritic structures have been reported in a number of reactions.[56,57] Good to excellent selectivities were achieved, and such possibilities are not observed with monomeric organocatalysts alone.

The multivalent presentations of DMAP organocatalysts are previously reported, which demonstrated the catalytic properties of Fréchet-type benzyl ether and aliphatic ester dendritic polymers containing 4-(dialkylamino)pyridines in the interior environment.[58] Multivalent DMAP organocatalyst plays an important role for the selective labelling of biopolymer under mild and benign condition. Hamachi and co-workers devised an efficient method of the site-selective chemical acylation of different proteins enabled by multivalent affinity-guided DMAP (AGD) catalysts.[59] The combined effects of catalyst multivalency and affinity guidance imposed by the DMAP-attached ligand were the key factors for achieving site-selective protein acylation with acyl donors possessing the desired probe. Chelation and coordination assistance in the catalytic sites and substrates lead to increases in the catalytic activities in the studied instances. Such an opportunity is studied herein in the case of site-selectivities in multiple hydroxy functionalities of monosaccharides with the aid of a trivalent DMAP catalyst.

Selectivity herein pertains to the site-selectivity among the secondary hydroxy functionalities. The coordination of a trivalent catalyst with the hydroxyl functionalities might improve the reactivity at a particular site. A series of acylations was performed using the trivalent catalyst, involving several monosac-

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

charides that possess diols and triols and several carboxylic acid anhydrides, with variable steric and electronic features, as acylation agents. The details of the studies are described herein.

Results and discussion

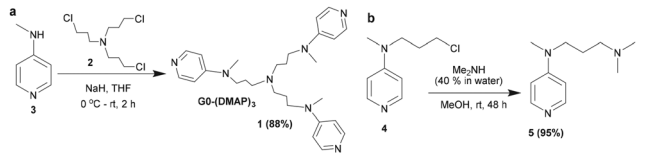
Synthesis of the catalyst

The site-selective functionalization of carbohydrates was undertaken with the aid of a trivalent (dialkylamino)pyridine (DAAP) catalyst. The synthesis of the new DAAP-based nucleophilic organocatalyst **1** was accomplished by the *N*-alkylation of tris-halide **2**[60] with 4-*N*-methylamino pyridine **3** in the presence of NaH in THF solution. The reaction occurred in a facile manner to afford the trivalent DAAP catalyst **1**, after purification by column chromatography (neutral Al_2O_3, $CHCl_3$/MeOH, linear gradient), in 88% yield (Scheme 1).

The corresponding monovalent catalyst **5** was secured through the *N*-alkylation of the corresponding halide **4**[61] with dimethyl amine (40% in water) in MeOH. The newly synthesized DAAP catalysts **1** and **5** were characterized by NMR spectroscopy and mass spectrometry; the details are given in the ESI.†

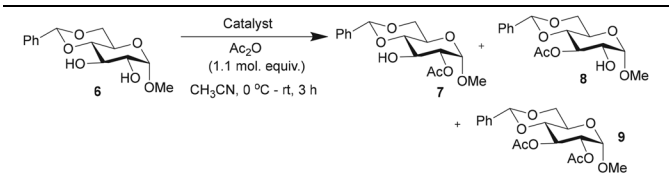
Study of the site-selective acylation of pyranoside-derived 1,2-diols

The efficacies of catalysts **1** and **5** as acylation transfer agents in monosaccharides having secondary 1,2-diol functionalities were studied initially. Reactivity-based differences are not obvious to equatorial secondary hydroxy functionalities, and in practice, scarcely attainable. For DMAP-catalyzed acylations, acyl halides or anhydrides are commonly employed. Trivalent catalyst **1** might facilitate the reaction of *N*-acyl-pyridinium intermediate with the hydroxy functionality than that with the monomeric DMAP catalyst. The effect of steric factors on the DMAP-catalyzed acylation was demonstrated by Zipse and co-workers.[62] The acylation of methyl 4,6-*O*-benzylidene-α-D-glucopyranoside (**6**), presenting a 1,2-*trans*-diol, was conducted using Ac_2O (1.1 molar equiv.), DAAP catalysts **1** (1.5 mol%), **5** (4.5 mol%) and DMAP (4.5 mol%) in MeCN, at 0 °C to room temperature for 3 h, and the formation of the acylation products was ascertained by ^{1}H NMR spectroscopy of the crude reaction mixtures. Mono-*O*-acylations and di-*O*-acylation were assessed, along with the overall yield of the reaction and site-selectivity towards the formation of the 2-*O*-acyl product (Table 1).



Scheme 1 Synthesis of (a) trivalent DAAP catalyst **1** and (b) monovalent (**5**) DAAP catalysts.

Table 1 Acetylation of methyl 4,6-*O*-benzylidene-α-D-glucopyranoside **6** using **1**, **5** and DMAP catalysts[a]



Entry	Catalyst (mol%)	**7** (%)	**8** (%)	**9** (%)	Conversion (%)
1	**1** (1.5)	65[b]	10	25	80
2	**5** (4.5)	50	50	—	4
3	DMAP (4.5)	51	40	9	44

[a] Products ratio determined by comparing the anomeric proton integrations in the ^{1}H NMR spectrum. [b] Yield of the isolated product is 44%.

Reactions were conducted in MeCN as the substrates and catalysts are more soluble freely in MeCN than in $CHCl_3$, CH_2Cl_2 and THF solutions. After the desired duration, the reaction mixture was quenched with MeOH, solvents evaporated under reduced pressure and analyzed by ^{1}H NMR spectroscopy.

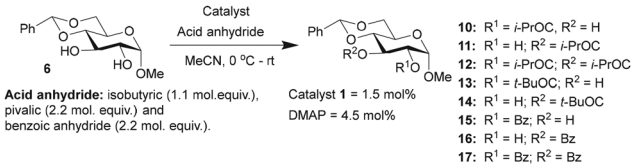
The acylation occurred at varying proportions, depending on the catalyst. Among the catalysts, trivalent DAAP **1** led to an efficient conversion to 2-*O*-acyl derivative **7** than monovalent **5** and DMAP catalysts. Further, an increase to 5 mol% of trivalent catalyst **1** led only to increased di-*O*-acylation product **9** (ESI†). As a result, the catalytic activities and site-selectivities of acylations mediated by multivalent catalysts were assessed using ~7 times reduced mol%. The monovalent catalyst **5** and DMAP were maintained at 4.5 mol%, whereas the trivalent catalyst **1** was maintained at one-third (1.5 mol%) throughout the present study. When DMAP derivative **3** was used as the catalyst (4.5 mol%), the conversion and site-selectivity were less, presumably due to acylation at the amine site of **3**. In light of the highest site-selectivity afforded by the trivalent catalyst **1**, further acylations were performed with this catalyst and in comparison, with DMAP. All the acylation experiments were conducted under auxiliary base-free condition.[63] Bases, such as Et_3N[64] and diisopropylethyl amine,[65] alone mediate the site-selective acylation of carbohydrate substrates.

Glucopyranoside **6** was subjected to acylation with few anhydrides of variable electronic and steric nature, namely, isobutyric anhydride ($(iPrCO)_2O$), pivalic anhydride ($(tBuCO)_2O$) and benzoic anhydride ($(PhCO)_2O$), under the optimized catalyst loading (1.5 mol%), and the observations are given in (Table 2).

The acylation of glucopyranoside **6** with isobutyric anhydride (1.1 molar equiv.) in the presence of catalyst **1** led to excellent site-selectivity for the 2-*O*-isobutyroyl product. The promising results of site-selective acetylation catalyzed by **1** warranted experiments using a sterically-hindered acylating agent, pivalic anhydride ($(tBuCO)_2O$). DMAP catalysis is sensitive to steric factors associated with reacting alcohols and car-

 This journal is © The Royal Society of Chemistry 2024

Table 2 Acylation of methyl 4,6-*O*-benzylidene-α-D-glucopyranoside **6** using variable acid anhydrides by **1** and DMAP catalysts[a]



Entry	Reagent	Time (h)	Catalyst	Products (%)			Conversion (%)	Yield[b] (%)
	$(i\text{-PrCO})_2O$	2		**10**	**11**	**12**		
1			**1**	75	10	15	90	61
2			DMAP	47	30	23	89	
	$(t\text{-BuCO})_2O$	16		**13**	**14**			
3			**1**	77	23	70		48
4			DMAP	64	36	45		
	Bz_2O	10		**15**	**16**	**17**		
5			**1**	54	12	34	90	42
6			DMAP	42	31	26	89	

[a] Products ratio determined by comparing the anomeric proton integrations in the ^{1}H NMR spectrum. [b] Yield of the isolated major product for **1**-catalyzed acylation reactions.

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

boxylic acid anhydrides. The same was observed in the pivaloylation of glucopyranoside **6** catalyzed by DMAP, which afforded an overall yield of 45% for the mono-*O*-acylated products, 2-pivaloyl (**13**) and 3-pivaloyl (**14**) products, in the ratio of 1.7 : 1. In these reactions, excess molar equiv. of pivalic anhydride (2.2 molar equiv.) was used, with increased duration of reaction to 16 h. Site-selectivity in benzoylation was conducted subsequently to secure the corresponding benzoates. The reaction was conducted using 2.2 mol. equiv. of benzoic anhydride $(PhCO)_2O$, catalyzed by either **1** or DMAP catalyst for 10 h. In this reaction, a considerable amount of 2,3-di-*O*-benzoyl product **17** also formed, presumably due to excess benzoic anhydride and longer reaction duration.

In all the cases, the overall conversion refers to the percentages of all the acylated products; the rest of the percentages were unreacted starting material, which were recovered during the purification of the crude reaction mixtures.

Acylation was then undertaken with methyl 4,6-*O*-benzylidene-α-D-galactopyranoside **18** possessing an equatorial 1,2-*trans* diol moiety. In this instance, the reactivity at *C*2 and *C*3 hydroxy functionalities are hardly distinguishable due to the 'axial oxy group effect'.[17b] The acylation of derivative **18** using Ac_2O (1.1 mol. equiv.) and catalyst **1** (1.5 mol%) afforded an overall yield of 99% in 2 h of reaction duration (Table 3). A ratio of 1 : 1 of products **19** and **20** was observed in this case. Similar results were observed in the isobutyration and benzoylation of derivative **18**; the site-selectivity was comparable in both cases. The facile site-selectivity for trivalent catalyst **1** might arise due to steric and electronic effects, favoring acylation at the *C*3 hydroxyl functionality.

Methyl 4,6-*O*-benzylidene-α-D-mannopyranoside **29**, possessing 1,2-*cis*-diol, was subjected to acylation subsequently. The results of the acylation of mannopyranoside **29** are summarized in Table 4. Slightly better yield (74%) was observed with catalyst **1** as compared to the DMAP catalyst in the case of *O*-acetylation occurring at the hydroxyl functionality at the *C*3 carbon. The lack of a considerable distinction in the site-selective *O*-acylation of mannopyranoside **29** is likely due to the higher reactivity of the *C*3 hydroxyl functionality of the 1,2-*cis*-diol. In pyranoside 1,2-diols, the site-selectivity appears to arise from the intrinsic reactivity difference. The hydrogen bonding at 1,2-*cis*-diol and the 'axial oxy group effect' might favor a site-selectivity towards the hydroxyl functionality at *C*3.

Methyl 4,6-*O*-benzylidene-α-D-glucopyranoside **6** afforded 2-*O*-acyl functionalized products, whereas methyl 4,6-*O*-benzylidene-α-D-galactopyranoside **18** and methyl 4,6-*O*-benzylidene-α-D-mannopyranoside **29** produced 3-*O*-acylation products. The synergistic effect of the hydrogen bonding, the disposition of the diols and steric factor associated with the acyl transfer process might play a role in the observed site-selectivities. Other than methyl 4,6-*O*-benzylidene-α-D-mannopyranoside **29** with 1,2-*cis*-diol disposition at *C*2 and *C*3 hydroxyl functionalities, the remaining pyranoside derivatives possess 1,2-*trans*-diol dispositions and afforded good yields and site-selectivities using trivalent catalyst **1**.

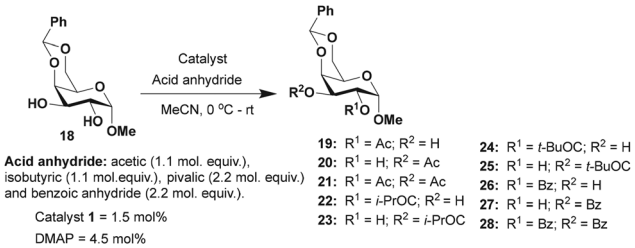
Acylation of 6-*O*-protected monosaccharides

The functionalization of the *C*6 primary hydroxyl functionality of pyranosides is facile in the presence of the secondary hydroxy functionalities. Advancements in the site-selective protection of 6-*O*-protected glycopyranosides are: (i) boron-mediated regioselective acylation of 6-*O*-protected carbohydrates;[44a] (ii) DMAP catalyst in the presence of *sym*-collidine by Kawabata and Muramatsu,[66] and (ii) Oriyama's chiral catalyst and TMEDA, as reported by Vasella and co-workers[67] (Scheme 2).

Site-selective acylation employing hydrogen bond acceptor-type protecting groups at the *C*6 hydroxyl functionality were also studied previously by Moitessier and co-workers.[12b,c]

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

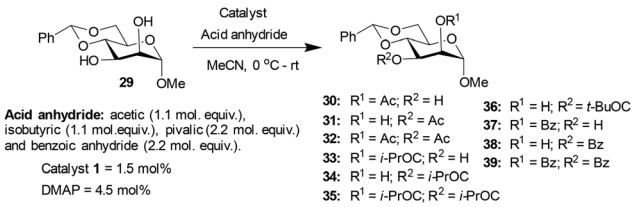
Table 3 Acylation of methyl 4,6-*O*-benzylidene-α-D-galactopyranoside **18** using variable acid anhydrides using **1** and DMAP catalysts[a]



Entry	Reagent	Time (h)	Catalyst	Products (%)			Conversion (%)	Yield[b] (%)
	Ac_2O	2		**19**	**20**	**21**		
1			**1**	18	61	20	99	51
2			DMAP	49	51	—	25	
	$(i\text{-}PrCO)_2O$	2		**22**	**23**			
3			**1**	21	79		93	64 (**22**/**23** = 30 : 70)
4			DMAP	34	66		71	
	$(t\text{-}BuCO)_2O$	16		**24**	**25**			
5			**1**	16	84		95	63 (**24**/**25** = 21 : 79)
6			DMAP	38	62		27	
	Bz_2O	10		**26**	**27**	**28**		
7			**1**	17	58	25	87	52 (**26**/**27** = 23 : 77)
8			DMAP	32	57	11	68	

[a] Products ratio determined by comparing the anomeric proton integrations in the ^{1}H NMR spectrum. [b] Yield of the isolated major product for **1**-catalyzed acylation reactions.

Table 4 Acylation of methyl 4,6-*O*-benzylidene-α-D-mannopyranoside **29** using variable acid anhydrides catalyzed by **1** and DMAP[a]



Entry	Reagent	Time (h)	Catalyst	Products (%)			Conversion (%)	Yield[b] (%)
	Ac_2O	3		**30**	**31**	**32**		
1			**1**	19	58	23	74	36
2			DMAP	16	75	9	60	
	$(i\text{-}PrCO)_2O$	2		**33**	**34**	**35**		
3			**1**	5	81	14	95	68
4			DMAP	7	82	11	91	
	$(t\text{-}BuCO)_2O$	16			**36**			
5			**1**		34		34	26
6			DMAP		28		28	
	Bz_2O	10		**37**	**38**	**39**		
7			**1**	11	83	6	78	56
8			DMAP	19	75	7	82	

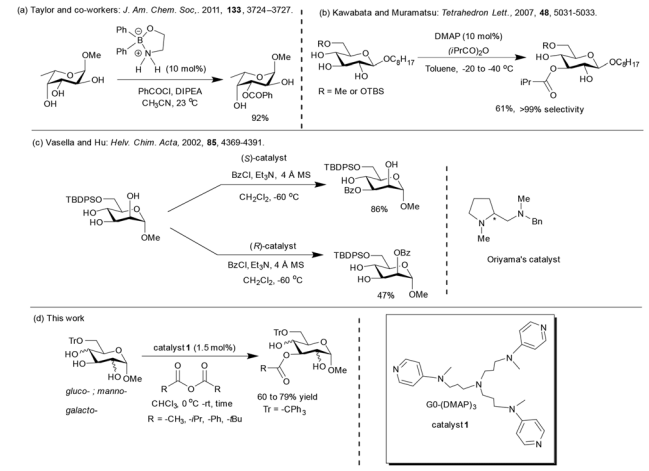
[a] Products ratio determined by comparing the anomeric proton integrations in the ^{1}H NMR spectrum. [b] Yield of the isolated major product for **1**-catalyzed acylation reactions.

Studies were undertaken to assess the site selectivity in 6-*O*-protected carbohydrate triols in the presence of trivalent **1** and DMAP catalysts. The reactions were conducted using methyl 6-*O*-trityl-α-D-glycopyranosides in $CHCl_3$ solution. The acylation of methyl 6-*O*-trityl-α-D-glucopyranoside (**40**) was conducted using catalyst **1** (1.5 mol%), Ac_2O (1.1 mol. equiv.) at 0 °C to room temperature for 45 min, followed by HPLC analysis. A new intense peak at a retention time of 7.6 min (56%) appeared, along with few other minor peaks. Unreacted starting material **40** appeared at a retention time of 27 min (Fig. 1).

The crude reaction mixture was subjected to column chromatography purification (SiO_2) (pet. Ether/EtOAc linear gradient) to secure a major product in 62% yield, which was identified as the 3-*O*-acylated product. The peak at 9.70 min

 This journal is © The Royal Society of Chemistry 2024

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .



Scheme 2 Site-selective acylation of pyranoside-derived triols.

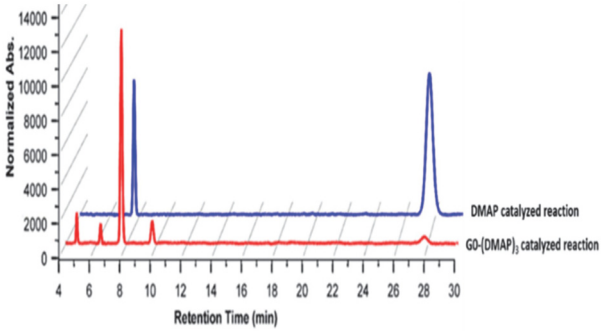


Fig. 1 HPLC chromatograms for the acetylation of glucopyranoside **40**. Chromatography conditions: Shimadzu Shim-pack (UC-X-Sil) 5 μm column, EtOAc–hexane = 60 : 40, flow rate 1 mL min^{-1}, PDA detector (254 nm), retention times: 7.6 min (major isomer **41**), 27.7 min (starting material **40**) and the remaining peaks correspond to the minor isomers.

corresponds to the 4-*O*-acetylated product **42**. The remaining peaks at 4.70 min and 6.29 min are related to multiple acetylated products, as evident from the mass spectrum of the crude reaction mixture. However, the identities of these minor products were not studied due to very less conversion (Fig. 1).

The observation reiterated the role of trivalent catalyst **1** in the elevated yields of the site-selective acylation product. Acylation glucopyranoside **40** was investigated further with varied acid anhydrides. Acetylation and isobutyration were conducted using acid anhydride (1.1 mol. equiv.), whereas benzoylation and pivaloylation were conducted using acid anhydride (2.2 mol. equiv.). In these instances too, catalyst **1** led to excellent overall yields of the mono-*O*-acylated products, with good site-selectivity in favor of the 3-*O*-acylated product. Except acetylation, other acylations afforded only 3-*O*-acyl and 4-*O*-acyl products (Table 5). In the case of bulky pivalic anhydride, the mono-3-*O*-pivaloate product **45** was formed in a good yield (84%) mediated by catalyst **1** than that mediated by the DMAP catalyst.

Site-selective acylations were also performed on methyl 6-*O*-trityl-α-D-mannopyranoside **49** under similar reaction conditions. In all the cases, the regio-isomers of the mono-*O*-acyl product were observed by HPLC analyses. The acetylation of mannopyranoside **49**, mediated by catalyst **1**, led to the preferential formation of mono 3-*O*-acetylated derivative. Mono *O*-acetylated products were isolated as an inseparable mixture of 2/3-*O*-acetylated products after purification (Table 6). Isobutyration also provided good yield of the mono-*O*-isobutyrated products, and the major 3-*O*-isobutyrated product **52** was isolated as a pure isomer. A good conversion of the pivaloylation also occurred, mediated by catalyst **1**, in comparison to DMAP catalysis. However, site-selectivity for acylation at the *C*3-hydroxyl functionality reduced due to the formation of a considerable amount of the *C*2-*O*-pivaloate **55**, for example, in the case of mono-*O*-benzoate **56**, at *C*3-hydroxyl functionality, in the presence of catalyst **1** (Table 6).

Galactopyranoside, namely, methyl 6-*O*-trityl-α-D-galactopyranoside-derived triol **58**, was subjected to *O*-acylation, wherein only two regioisomeric products were formed, as evident from the HPLC analysis of the crude reaction mixture. Isolation and NMR characterization suggested the formation of the *C*3-*O*-acylated product as the major regio-isomer (Table 7). In this instance too, catalyst **1** afforded the mono-*O*-acylation products in higher yields and site-selectivity, as compared to the DMAP catalyst. Isobutyration reaction provided excellent overall conversion of the mono-*O*-isobutyrate products, where *C*3-*O*-isobutyrate **61** was isolated as the major product. Pivaloylation also furnished good conversion with *C*3/*C*2-*O*-pivaloate (**63** and **64**) as an inseparable mixture after purification. Finally, the benzoylation reaction provided the mono-*O*-benzoyl derivatives **65** and **66**, catalyzed by catalyst **1**, in higher yields than that by the DMAP catalyst (Table 7).

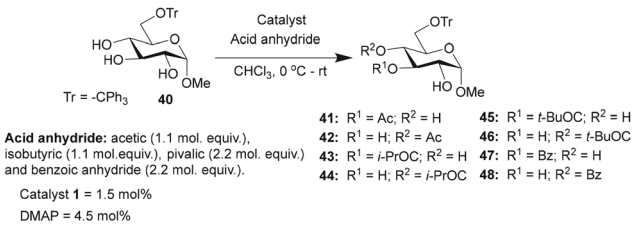
Although DMAP-catalyzed acylations of triols appear to provide an edge on the site-selectivity as compared to the trivalent catalyst **1**, however, striking differences are observed in the overall conversion and the yield of the major mono-*O*-acylated product. In all the cases, trivalent catalyst **1** mediated the acylation of pyranoside triol derivatives, with excellent overall conversion and yield of the major regioisomer.

The effect of base on the rate and site-selectivity of acylation reaction is studied previously for monosaccharides[11] and acyclic amphiphilic diols.[68] Besides being trivalent, catalyst **1** is also constituted with a tertiary amine functionality. In order to assess the role of tertiary amine, the acetylation of **6** was conducted with 4.5 mol% DMAP, 1.1 mol. equiv. Ac_2O, along with 1.1 mol. equiv. of Et_3N. Interestingly, the reactions of DMAP, with and without Et_3N, led to similar conversions and site-selectivities, as evident from the HPLC analyses of the reaction mixtures (Fig. S25†). Further, several unidentified products were observed in the reaction with Et_3N. The reaction with Et_3N suggests a minimal role, if any, on the enhanced site-selectivities and reaction rates by the trivalent catalyst **1**.

Ratios of the major and minor regioisomers were investigated at different time intervals for trivalent **1**-catalyzed acylation experiments for glucopyranoside diol **6** and triol **40**. At

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

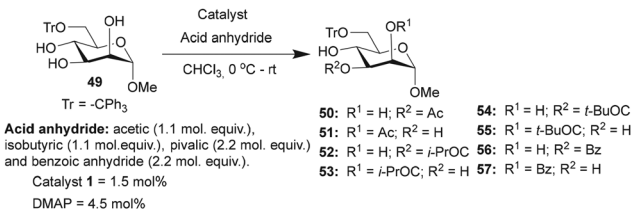
Table 5 Site-selective acylation of methyl 6-*O*-trityl-α-D-glucopyranoside **40** catalyzed by **1** and DMAP[a]



Entry	Reagent	Time	Catalyst	Products (%)		Conversion (%)	Yield[b] (%)
	Ac_2O	45 min		**41**	**42**		
1			**1**	74	9	94	62
2			DMAP	19	—	19	
	$(i\text{-PrCO})_2O$	30 min		**43**	**44**		
3			**1**	89	6	95	70
4			DMAP	35	3	38	
	$(t\text{-BuCO})_2O$	16 h		**45**	**46**		
5			**1**	84	14	98	72
6			DMAP	8	1	9	
	Bz_2O	8 h		**47**	**48**		
7			**1**	73	23	96	61
8			DMAP	12	3	16	

[a] Product ratios determined by HPLC analysis. [b] Yield of the isolated major product for **1**-catalyzed acylation reactions.

Table 6 Site-selective acylation of methyl 6-O-trityl-α-D-mannopyranoside **49** catalyzed by **1** and DMAP[a]



Entry	Reagent	Time	Catalyst	Products (%)		Conversion (%)	Yield[b] (%)
	Ac_2O	45 min		**50**	**51**		
1			**1**	59	11	91	52 (**50**/**51** = 70 : 30)
2			DMAP	21	4	26	
	$(i\text{-PrCO})_2O$	30 min		**52**	**53**		
3			**1**	65	23	97	58
4			DMAP	24	4	32	
	$(t\text{-BuCO})_2O$	16 h		**54**	**55**		
5			**1**	55	33	91	42
6			DMAP	11	1	13	
	Bz_2O	8 h		**56**	**57**		
7			**1**	63	24	86	47
8			DMAP	5	6	11	

[a] Product ratios determined by HPLC analysis. [b] Yield of the isolated major product for **1**-catalyzed acylation reactions.

different time intervals, an aliquot of the acylation reaction mixture was analyzed by HPLC after quenching with methanol. No significant change in the regioisomeric ratios was observed at different time intervals for a particular acylation reaction (ESI†).

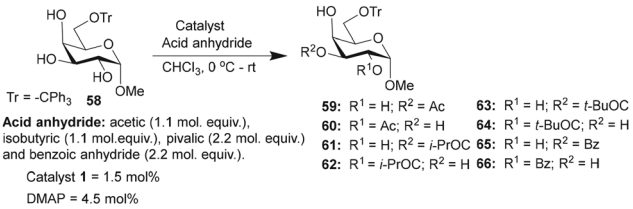
In order to verify the efficiency of the reaction, the turnover-frequency (TOF) was calculated for the major regioisomers in different acylations of substrates **6** and **40** catalyzed by trivalent catalyst **1**. The results of the experiments are tabulated in the ESI.† The TOF ranged between 7.8 after 4 h for pivaloylation of **6** and 184 after 15 min for the isobutylation of substrate **40** (ESI S7†). All the acylation experiments were conducted in the absence of any auxiliary base. We surmise that the tertiary amine site in the catalyst acts as the protonation site as a result.[52b]

The above demonstrations of the site-selective acylation of diols and triols mediated by the trivalent catalyst **1** illustrate that a trimeric catalyst gains benefit from both the overall con-

 This journal is © The Royal Society of Chemistry 2024

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

Table 7 Site-selective acylation of methyl 6-*O*-trityl-α-D-galactopyranoside **58** catalyzed by **1** and DMAP[a]



Entry	Reagent	Time	Catalyst	Products (%)		Conversion (%)	Yield[b] (%)
	Ac_2O	45 min		**59**	**60**		
1			**1**	40	27	71	20
2			DMAP	24	7	31	
	$(i\text{-PrCO})_2O$	30 min		**61**	**62**		
3			**1**	61	20	81	42
4			DMAP	27	10	38	
	$(t\text{-BuCO})_2O$	16 h		**63**	**64**		
5			**1**	57	15	72	45 (**63**/**64** = 77 : 23)
6			DMAP	5	1	6	
	Bz_2O	8 h		**65**	**66**		
7			**1**	61	25	86	44
8			DMAP	24	8	32	

[a] Product ratios determined by HPLC analysis. [b] Yield of the isolated major product for **1**-catalyzed acylation reactions.

version and site-selectivity. The site-selectivity is inferred due to the internal hydrogen bonding among the hydroxy functionalities and the associated increases in the nucleophilicity. The internal hydrogen bonding scheme, coupled with the acyl transfer efficiency of the trivalent catalyst **1**, contributes to the impressive site-selectivity of mono-*O*-acylation in the majority of instances. Increasing the number of catalytic moieties within a molecular scaffold increases the reactivities over and above the statistical increase of the catalytic moiety. Such trends of multivalent catalysis were observed in many occasions earlier in organometallic and organocatalysis reactions.[56,57,69] Higher reaction efficiencies by multivalent catalysts are accounted for by the stabilization of the active catalytic moiety, in addition to kinetic advantages. The trivalent catalyst **1** studied herein is an organocatalyst based on DMAP acyl transfer agent. The trivalent nature of catalyst **1** presumably leads to a statistical increase in the acyl transfer and, as a result, the increased reactivity. It is likely that the individual DAAP moiety in **1** might aid in the deprotonation of the pyridinium species and an efficient re-generation of catalytically active DAAP moiety, as advanced by Hamachi and co-workers on multivalent affinity-guided DMAP (AGD) catalysts.[59a] These combined phenomena are inferred as a plausibility for the observed significantly increased reactivity and site-selectivity of reactions catalyzed by trivalent catalyst **1**.

The α-anomeric methoxy moiety would participate in the formation of an internal hydrogen bonding with the hydroxyl functionality at *C*2 in glucopyranoside diol derivative **6**, which subsequently led to 2-*O*-acyl products predominantly. A similar plausible mechanism of internal hydrogen bonding in the substrate and an enhanced reactivity of the trivalent catalyst would account for the site-selective formation of 3-*O*-acylation product, in the case of galactopyranoside diol substrate **18**.

Catalyst development is critical to achieve both site-selectivity and conversion efficiencies. Reaction conditions, such as low temperatures and presence of a base, play an important role in achieving good selectivities and conversions. Peptide-based monovalent DMAP catalysts provide good selectivities for benzoylations, although with limited conversion efficiency.[28a] The selectivity and conversion efficiency of the present method are comparable in case of the Cu-bis(oxazoline)-catalyzed site-selective acetylation of glycoside **6** and benzoylation for galactopyranoside **18** using Cu-(*R*,*R*)-PhBox[35] catalysts. A further case in point is the isobutyration, an acylation encountered with the sterically hindered isobutyryl moiety. Trivalent catalyst **1** mediates this reaction with substrate **6** with good selectivity and yield of *C*2-isobutyroyl ester. In the case of pyranoside-derived triols, it is noteworthy that the trivalent catalyst **1** affords *C*3 *O*-acylated product in much higher yields than either DMAP or substrates possessing hydrogen bond acceptor type protecting group at the *C*6 hydroxyl functionality in the triol.[12b,c] Several acylations on 6-*O*-trityl-protected glucopyranoside triol substrate **40** yielded *C*3 *O*-acylated products in higher percentages than that known currently.[12b,c] Interestingly, the benzoylation of pyranoside derivatives **40**, **49** and **61** at *C*3, employing the catalyst **1**, was different from that observed by Evtushenko, using benzoic anhydride, 2,4,6-collidine and copper(II) trifluoroacetate as promoters.[42] Under this condition, benzoylation at *C*2 site was observed. $MoO_2(acac)_2$ as a catalyst promoted benzoylation at *C*3, as reported in an earlier study from the same group,[70] which was similar with the observation of the present studies. Without an amplification of the intrinsic reactivity differences, acylations occur equally at the available hydroxyl functionalities in diols and triols. We undertook to exploit the enhanced reactivity of multivalent catalysts. The beneficial effects of the valencies of catalytic sites

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

in a catalyst pertaining to site-selectivity and conversion efficiency are observed in the present study with varied acylations, ranging from acetyl, isobutyryl, pivaloyl and benzoyl esters and with diols and triols derived from D-glucopyranoside, galactopyranoside and mannopyranoside.

Conclusions

This work uncovers a catalytic, site-selective method for the mono-*O*-acylation of several pyranoside-derived diols and triols, employing a trivalent DAAP catalyst **1**. Varied acid anhydrides were used for the acylation; the acylation and the site-selectivity are largely dependent on the intrinsic reactivity of the carbohydrate hydroxyl functionalities. 4,6-*O*-Benzylidene-protected glucopyranoside diol furnished 2-*O*-acylated product, whereas the corresponding galactopyranoside diol enabled the formation of the 3-*O*-acylated derivative in good yield and site-selectivity, mediated by trivalent **1** catalyst. Experiments using monovalent DMAP led to considerably lesser conversions of the desired mono-*O*-acyl derivatives, with diminished site-selectivity in most instances. Among the glycopyranoside derivatives, 6-*O*-trityl-protected glycopyranoside triols furnished the 3-*O*-acylated derivatives in moderate to good yields and good site-selectivities using catalyst **1**.

Experimental

General methods

Unless otherwise noted, all reagents were purchased from commercial sources and used as received. Solvents were dried and distilled according to literature procedures. TLC analyses were performed on commercial Merck plates coated with silica gel or neutral alumina 60 F_{254} (type E, 0.2 mm), and visualizations were conducted by UV radiation or using I_2 as a staining agent. Silica gel (100–200 and 230–400 mesh) and neutral alumina were used for column chromatography purifications. Mass spectral characterizations were performed on ESI-QTOF operated in the positive ion mode on samples in either MeCN/water or MeOH/water solution. ^{1}H and ^{13}C NMR spectral analyses were performed on a spectrometer operating at 400 and 100 MHz, respectively. Processing of the FID data was performed on Bruker TopSpin™ and Mnova software with default settings. Chemical shifts are reported with respect to tetramethylsilane (TMS) for ^{1}H NMR and the central line (77.0 ppm) of $CDCl_3$ for ^{13}C NMR spectra. Coupling constants (J) are reported in Hertz. Standard abbreviations s, d, t, dd, td, br. s, m, quint. and app. refer to singlet, doublet, triplet, doublet of doublet, triplet of doublet, broad singlet, multiplet, quintet and apparent, respectively. HPLC analyses were performed using an analytical silica column attached with a photodiode array detector and using pet.ether/EtOAc binary gradient elution. The first 0 to ~3.5 min in the traces cover the void volume of each HPLC analysis, and the HPLC traces are given without the void volume.

Catalyst 1: [G0-(DMAP)$_3$]. A solution of 4-(*N*-methylamino) pyridine **3** (90.4 mg, 8.4 mmol) in THF (30 mL) was added dropwise to a suspension of sodium hydride (53.6 mg, 13.44 mmol) THF (10 mL) under N_2 atmosphere at 0 °C. The solution was stirred for an additional 2 h at room temperature. A solution of tris(3-chloropropyl)amine **2**[57] (0.28 g, 1.12 mmol) in THF (25 mL) was introduced to the mixture in a 0 °C ice bath *via* injection. The solution was stirred further for 24 h at 70 °C, filtered and the solution evaporated under reduced pressure. The resulting crude product was purified by column chromatography (neutral Al_2O_3) (eluant: MeOH/CH_2Cl_2 1 : 99) to secure **1** (0.46 g, 88%) as a colorless gum. ^{1}H NMR (400 MHz, $CDCl_3$) δ 8.16 (app. d, J = 6.4 Hz, 6 H), 6.44 (app. d, J = 6.8 Hz, 6 H), 3.33 (t, J = 7.2 Hz, 6 H), 2.91 (s, 9 H), 2.42 (t, J = 7.2 Hz, 6 H), 1.68 (quint., J = 7.2 Hz, 6 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 153.4, 149.8, 106.6, 51.4, 49.5, 37.5, 24.4. HRMS (ESI) m/z: $[M + H]^+$ calcd for $C_{27}H_{40}N_7$, 462.3345; found 462.3345.

4-[*N*-Methyl-*N*-(3-dimethylaminopropyl)amino]pyridine (5). A solution of 4-[*N*-methyl-*N*-(3-hydroxypropyl)amino]pyridine[58] (0.15 g, 0.92 mmol) and $SOCl_2$ (0.1 mL, 1.38 mmol) in $CHCl_3$ (40 mL) was stirred at 55 °C for 3 h, and the solvents were removed under reduced pressure. The crude product was dissolved in EtOH (5 mL), warmed for 10 min and solvents removed under reduced pressure. The resulting residue was dissolved in EtOAc (40 mL) and washed with aq. $NaHCO_3$ brine (3 × 5 mL), dried and evaporated under reduced pressure, and the crude product was purified (alumina, CH_2Cl_2/MeOH 99 : 1) to secure intermediate **4** (0.14 g, 82%) as a colorless liquid. ^{1}H NMR (400 MHz, $CDCl_3$) δ 8.18 (app. d, J = 6.4 Hz, 2 H), 6.48 (app. d, J = 6.4 Hz, 2 H), 3.55 (t, J = 6.2 Hz, 2 H), 3.50 (t, J = 7 Hz, 2 H), 2.96 (s, 3 H), 2.02 (qn, J = 6.8 Hz, 2 H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 153.2, 149.9, 106.6, 48.5, 42.3, 37.8, 29.6. HRMS (ESI) m/z: $[M + H]^+$ calcd for $C_9H_{14}N_2Cl$, 185.0849; found 185.0846.

A solution of **4** (0.11 g, 0.59 mmol) in MeOH (20 mL) and Me_2NH (40% in water) (0.25 mL, 1.47 mmol) was stirred for 48 h at room temperature. Solvents were removed under reduced pressure, the resulting residue diluted with $CHCl_3$ (25 mL), washed with saturated aq. $NaHCO_3$, brine, dried, evaporated under reduced pressure and the resulting crude product was purified (alumina, CH_2Cl_2/MeOH 99 : 1) to secure **5** (0.11 g, 95%) as a colorless liquid. ^{1}H NMR (400 MHz, D_2O) δ 7.88 (d, J = 4 Hz, 2 H), 6.40 (d, J = 8.0 Hz, 2 H), 3.16 (t, J = 7.2 Hz, 2 H), 2.75 (s, 3 H), 2.15 (t, J = 8 Hz, 2 H), 2.02 (s, 6 H), 1.53 (quint, J = 8 Hz, 2 H). ^{13}C NMR (100 MHz, D_2O): δ 154.0, 148.2, 106.7, 55.8, 48.8, 43.8, 36.7, 23.6. HRMS (ESI) m/z: $[M + H]^+$ calcd for $C_{11}H_{20}N_3$, 194.1657, found 194.1658.

Acetylation with variable catalysts. The monosaccharide substrate **6** (20 mg, 0.070 mmol) and the desired catalyst (1.5 mol% of trivalent DMAP catalyst (**1**), 4.5 mol% of monovalent DMAP catalyst (**5**) or 4.5 mol% DMAP catalyst) in MeCN (1 mL) was stirred at 0 °C, 1.1 molar equiv. of acetic anhydride was added, and stirred at 0 °C to room temperature. After being stirred for 3 h, the reaction was quenched with MeOH (1 ml) and stirred for further 30 min. The solvent was evapor-

 This journal is © The Royal Society of Chemistry 2024

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

ated under reduced pressure, the crude residue was dissolved in $CDCl_3$, and the regioselectivity and yield were determined by ^{1}H NMR spectra signal integrals.

General procedure A: regioselective acylation of pyranoside diols catalyzed by trivalent 1 or DMAP. The carbohydrate starting material (1 molar equiv.) and either 1.5 mol% of trivalent DMAP catalyst (**1**) or 4.5 mol% of normal DMAP catalyst were dissolved in MeCN, desired carboxylic acid anhydride (1.1 molar equiv. of acetic and isobutyric anhydride) and 2.2 molar equiv. of pivalic and benzoic anhydride was added and stirred at 0 °C to room temperature for a desired time. Afterwards, the reaction was quenched with MeOH (1 mL) and stirred for a further 30 min. The solvent was removed under reduced pressure and the crude product purified by column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc) to secure the major regioisomeric product.

Methyl 2-*O*-acetyl-4,6-*O*-benzylidene-α-D-glucopyranoside (7). A mixture of methyl 4,6-*O*-benzylidene-α-D-glucopyranoside (**6**) (30 mg, 0.106 mmol), **1** (0.92 mg, 0.002 mmol) and Ac_2O (11.0 μL, 0.12 mmol) was stirred in MeCN (1 mL) for 3 h and worked-up, as given in the general procedure A. Compound **7** [30a] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 1 : 1) as a colorless gum (15.2 mg, 44%). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.51–7.49 (m, 2 H), 7.38–7.37 (m, 3 H), 5.55 (s, 1 H), 4.95 (d, *J* = 3.6 Hz, 1 H), 4.80 (dd, *J* = 9.6, 3.6 Hz, 1 H), 4.30 (dd, *J* = 10, 4.4 Hz, 1 H), 4.18 (t, *J* = 9.6 Hz, 1 H), 3.85 (td, *J* = 10, 4.4 Hz, 1 H), 3.76 (t, *J* = 10.2 Hz, 1 H), 3.57 (t, *J* = 9.2 Hz, 1 H), 3.41 (s, 3 H), 2.16 (s, 3 H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 170.8, 137.0, 129.5, 128.5, 126.4, 102.2, 97.7, 81.5, 73.7, 69.0, 68.8, 62.1, 55.6, 21.1. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd for $C_{16}H_{20}O_7Na$, 347.1107, found 347.1107.

Methyl 4,6-*O*-benzylidene-2-*O*-isobutyryl-α-D-glucopyranoside (10). A mixture of methyl 4,6-*O*-benzylidene-α-D-glucopyranoside (**6**) (30 mg, 0.106 mmol), **1** (0.92 mg, 0.002 mmol) and isobutyric anhydride (19.4 μL, 0.116 mmol) was stirred in MeCN (1 mL) for 75 min and worked-up as given in the general procedure A. Compound **10** [30a] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 4 : 1), as a colorless gum (22.8 mg, 61%). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.50–7.48 (m, 2 H), 7.38–7.37 (m, 3 H), 5.54 (s, 1 H), 4.94 (d, *J* = 3.6 Hz, 1 H), 4.77 (dd, *J* = 9.6, 4 Hz, 1 H), 4.29 (dd, *J* = 10, 4.4 Hz, 1 H), 4.18 (t, *J* = 9.6 Hz, 1 H), 3.84 (td, *J* = 10, 4.8 Hz, 1 H), 3.75 (t, *J* = 10.2 Hz, 1 H), 3.55 (t, *J* = 9.2 Hz, 1 H), 3.39 (s, 3 H), 2.66 (sept, *J* = 6.8 Hz, 1 H), 1.19 (dd, *J* = 6.8, 2.4 Hz, 6 H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 177.0, 137.1, 129.4, 128.5, 126.4, 102.1, 97.7, 81.5, 73.4, 69.0, 68.9, 62.1, 55.6, 33.9, 19.1, 18.9. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd for $C_{18}H_{24}O_7Na$, 375.1420, found 375.1420.

Methyl 4,6-*O*-benzylidene-2-*O*-pivaloyl-α-D-glucopyranoside (13). A mixture of methyl 4,6-*O*-benzylidene-α-D-glucopyranoside (**6**) (30 mg, 0.106 mmol), **1** (0.92 mg, 0.002 mmol) and pivalic anhydride (47.2 μL, 0.233 mmol) was stirred in MeCN (1 mL) for 16 h and worked-up as given in the general procedure A. Compound **13** [13] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 3 : 1), as a colorless gum (18.7 mg, 48%). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.50–7.48 (m, 2 H), 7.37–7.36 (m, 3 H), 5.54 (s, 1 H), 4.93 (d, *J* = 3.6 Hz, 1 H), 4.73 (dd, *J* = 9.6, 3.6 Hz, 1 H), 4.29 (dd, *J* = 10, 4.4 Hz, 1 H), 4.18 (t, *J* = 9.6 Hz, 1 H), 3.84 (td, *J* = 10, 4.8 Hz, 1 H), 3.75 (t, *J* = 10.2 Hz, 1 H), 3.55 (t, *J* = 9.4 Hz, 1 H), 3.38 (s, 3 H), 1.24 (s, 9 H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 178.3, 137.1, 129.4, 128.4, 126.4, 102.1, 97.8, 81.5, 73.5, 69.0, 68.9, 62.1, 55.7, 39.0, 27.1. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd for $C_{19}H_{26}O_7Na$, 389.1576, found 389.1574.

Methyl 4,6-*O*-benzylidene-2-*O*-benzoyl-α-D-glucopyranoside (15). A mixture of methyl 4,6-*O*-benzylidene-α-D-glucopyranoside (**6**) (30 mg, 0.106 mmol), **1** (0.92 mg, 0.002 mmol) and benzoic anhydride (52.8 mg, 0.23 mmol) was stirred in MeCN (1 mL) for 10 h and worked-up as given in the general procedure A. Compound **15** [28b] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 3 : 1), as a colorless gum (17.2 mg, 42%). ^{1}H NMR (400 MHz, $CDCl_3$): δ 8.11–8.09 (m, 2 H), 7.60–7.57 (m, 1 H), 7.53–7.51 (m, 2 H), 7.48–7.44 (m, 2 H), 7.39–7.38 (m, 3 H), 5.58 (s, 1 H), 5.07 (d, *J* = 6.4 Hz, 1 H), 5.04 (dd, *J* = 9.6, 4 Hz, 1 H), 4.38–4.31 (m, 2 H), 3.92 (td, *J* = 10, 4.8 Hz, 1 H), 3.80 (t, *J* = 10.4 Hz, 1 H), 3.64 (t, *J* = 9.6 Hz, 1 H), 3.40 (s, 3 H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 166.4, 1371., 133.5, 130.1, 129.6, 129.4, 128.6, 128.5, 126.5, 102.2, 97.9, 81.6, 74.2, 69.1, 69.0, 62.2, 55.6. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd for $C_{21}H_{22}O_7Na$, 409.1263, found 409.1264.

Methyl 3-*O*-acetyl-4,6-*O*-benzylidene-α-D-galactopyranoside (20). A mixture of methyl 4,6-*O*-benzylidene-α-D-galactopyranoside (**18**) (20 mg, 0.07 mmol), **1** (0.46 mg, 0.001 mmol) and Ac_2O (7.4 μL, 0.078 mmol) was stirred in MeCN (1 mL) for 2 h and worked-up as given in the general procedure A. Compound **20** [30a] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 3 : 2) as a colorless gum (11.7 mg, 51%).^{1}H NMR (400 MHz, $CDCl_3$): δ 7.51–7.49 (m, 2 H), 7.38–7.36 (m, 3 H), 5.51 (s, 1 H), 5.12 (dd, *J* = 10.4, 2.8 Hz, 1 H), 4.95 (d, *J* = 3.2 Hz, 1 H), 4.38 (d, *J* = 2.8 Hz, 1 H), 4.28 (d, *J* = 12.8 Hz, 1 H), 4.18–4.16 (m, 2 H), 4.07 (d, *J* = 12.4 Hz, 1 H), 3.73 (s, 3 H), 2.14 (s, 3 H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 171.5, 137.7, 129.1, 128.3, 126.3, 100.9, 100.4, 74.4, 71.8, 69.3, 66.8, 62.6, 55.8, 21.3. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd for $C_{16}H_{20}O_7Na$, 347.1107, found 347.1106.

Methyl 4,6-*O*-benzylidene-2/3-*O*-isobutyryl-α-D-galactopyranoside (22/23). A mixture of methyl 4,6-*O*-benzylidene-α-D-galactopyranoside (**18**) (20 mg, 0.07 mmol), **1** (0.46 mg, 0.001 mmol) and isobutyric anhydride (13.0 μL, 0.078 mmol) was stirred in MeCN (1 mL) for 2 h and worked-up, as given in the general procedure A. Compound **22**/**23** was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 2 : 1) as a colorless gum (15.9 mg, 64%) and as an inseparable 2/3-*O*-isobutyrate regioisomers (30 : 70). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.50–7.48 (m, 2.6 H), 7.36–7.35 (m, 3.9 H), 5.57 (s, 0.44 H), 5.52 (s, 1 H), 5.14 (dd, *J* = 10, 2.8 Hz, 0.43 H), 5.09 (dd, *J* = 10.4, 3.2 Hz, 1 H), 4.98 (d, *J* = 3.2 Hz, 0.45 H), 4.95 (d, *J* = 3.2 Hz, 1 H), 4.38 (d, *J* = 3.2 Hz, 1 H), 4.32–4.27 (m, 1.95 H), 4.18 (dd, *J* = 10, 3.2 Hz, 1 H), 4.13–4.05 (m, 2 H), 3.73 (s, 1.42 H), 3.47 (s, 3 H), 3.42 (s, 0.69 H), 2.66 (sept, *J* = 7 Hz, 1.44 H), 1.19 (d, *J* = 6.8 Hz, 7.88 H). ^{13}C NMR (100 MHz, $CDCl_3$): δ 177.6, 177.5, 137.8, 137.5, 129.4, 129.0, 128.4, 128.3, 126.4,

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

126.1, 101.5, 100.7, 100.4, 98.4, 77.3, 76.2, 74.3, 71.6, 71.2, 69.4, 67.4, 66.9, 62.7, 62.6, 55.9, 55.8, 34.1, 34.0, 19.3, 19.1, 19.0, 18.9. HRMS (ESI) *m/z*: [M + Na]$^+$ calcd for $C_{18}H_{24}O_7Na$, 375.1420, found 375.1422.

Methyl 4,6-*O*-benzylidene-2/3-*O*-pivaloyl-α-D-galactopyranoside (24/25). A mixture of methyl 4,6-*O*-benzylidene-α-D-galactopyranoside (**18**) (20 mg, 0.07 mmol), **1** (0.46 mg, 0.001 mmol) and pivalic anhydride (31.6 μL, 0.16 mmol) was stirred in MeCN (1 mL) for 16 h and worked-up as given in the general procedure A. Compound **24/25** [28b] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 2 : 1) as a colorless gum (16.3 mg, 63%) and as an inseparable 2/3-*O*-pivaloate regioisomers (21 : 79). ^{1}H NMR (400 MHz, $CDCl_3$): δ 7.51–7.48 (m, 2.5 H), 7.37–7.34 (m, 3.6 H), 5.57 (s, 0.3 H), 5.52 (s, 1 H), 5.06 (dd, *J* = 10.4, 3.2 Hz, 1 H), 4.97 (d, *J* = 3.6 Hz, 0.3 H), 4.93 (d, *J* = 3.6 Hz, 1 H), 4.38 (d, *J* = 3.2 Hz, 1 H), 4.30–4.27 (m, 1.5 H), 4.18 (dd, *J* = 10.4, 3.2 Hz, 1 H), 4.10–4.05 (m, 1.4 H), 3.80 (s, 0.14 H), 3.73 (s, 1 H), 3.47 (s, 3 H), 3.41 (s, 0.63 H), 1.24 (s, 10.9 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 178.9, 178.8, 137.8, 137.5, 130.0, 129.4, 128.9, 128.4, 128.2, 126.4, 125.9, 101.4, 100.5, 100.4, 98.5, 76.2, 74.2, 71.6, 71.2, 69.3, 67.4, 66.9, 62.6, 62.5, 55.9, 55.8, 39.1, 39.0, 27.2, 27.1. HRMS (ESI) *m/z*: [M + Na]$^+$ calcd for $C_{19}H_{26}O_7Na$, 389.1576, found 389.1574.

Methyl 4,6-*O*-benzylidene-2/3-*O*-benzoyl-α-D-galactopyranoside (26/27). A mixture of methyl 4,6-*O*-benzylidene-α-D-galactopyranoside (**18**) (20 mg, 0.07 mmol), 1 (0.46 mg, 0.001 mmol) and benzoic anhydride (35.3 mg, 0.16 mmol) was stirred in MeCN (1 mL) for 10 h and worked-up as given in the general procedure A. Compound **26/27** [71] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 3 : 2) as a colorless gum (14.2 mg, 52%) and as an inseparable 2/3-*O*-benzoate regioisomers (77 : 23). ^{1}H NMR (400 MHz, $CDCl_3$): δ 8.11–8.09 (m, 2.7 H), 7.63–7.54 (m, 1.5 H), 7.52–7.48 (m, 2.6 H), 7.46–7.42 (m, 2.6 H), 7.39–7.35 (m, 3.2 H), 5.60 (s, 0.34 H), 5.54 (s, 1 H), 5.39 (dd, *J* = 10, 3.6 Hz, 1.4 H), 5.13 (d, *J* = 3.2 Hz, 0.3 H), 5.01 (d, *J* = 3.6 Hz, 1 H), 4.50 (d, *J* = 3.2 Hz, 1 H), 4.36–4.30 (m, 3 H), 4.14–4.09 (m, 1.5 H), 3.80 (s, 1.4 H), 3.50 (s, 3 H), 3.44 (s, 0.8 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 166.9, 166.8, 137.8, 137.5, 133.8, 133.4, 130.3, 130.0, 129.9, 129.4, 129.0, 128.8, 128.6, 128.5, 128.4, 128.2, 126.4, 126.2, 101.6, 100.7, 100.5, 98.5, 76.3, 74.6, 72.3, 72.0, 69.4, 69.2, 67.5, 67.1, 62.8, 62.6, 55.9, 55.8. HRMS (ESI) *m/z*: [M + Na]$^+$ calcd For $C_{21}H_{22}O_7Na$, 409.1263, found 409.1263.

Methyl 3-*O*-acetyl-4,6-*O*-benzylidene-α-D-mannopyranoside (31). A mixture of methyl 4,6-*O*-benzylidene-α-D-mannopyranoside (**29**) (25 mg, 0.09 mmol), **1** (0.60 mg, 0.0013 mmol) and Ac_2O (9.2 μL, 0.097 mmol) was stirred in 1 mL of CH_3CN for 3 h and worked-up as given in the general procedure A. Compound **31** [30a] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 3 : 1) as a colorless gum (10.3 mg, 36%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.46–7.44 (m, 2 H), 7.36–7.26 (m, 3 H), 5.55 (s, 1 H), 5.34 (dd, *J* = 10.4, 3.2 Hz, 1 H), 4.75 (s, 1 H), 4.29 (dd, *J* = 9.6, 4.4 Hz, 1 H), 4.15 (brs, 1 H), 4.08 (t, *J* = 9.6 Hz, 1 H), 3.93 (td, *J* = 10, 4.4 Hz, 1 H), 3.86 (t, *J* = 10 Hz, 1 H), 3.41 (s, 3 H), 2.13 (s, 3 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 169.9, 137.3, 129.9, 129.2, 128.4, 126.3, 102.0, 101.4, 76.2, 70.8, 69.9, 68.9, 63.8, 55.2, 21.2. HRMS (ESI) *m/z*: [M + Na]$^+$ calcd for $C_{16}H_{20}O_7Na$, 347.1107, found 347.1105.

Methyl 4,6-*O*-benzylidene-3-*O*-isobutyryl-α-D-mannopyranoside (34). A mixture of methyl 4,6-*O*-benzylidene-α-D-mannopyranoside (**29**) (25 mg, 0.09 mmol), **1** (0.60 mg, 0.001 mmol) and isobutyric anhydride (16.2 μL, 0.097 mmol) was stirred in MeCN (1 mL) for 2 h and worked-up as given in the general procedure A. Compound **34** [30a] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 3 : 1) as a colorless gum (21.2 mg, 68%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.45–7.43 (m, 2 H), 7.35–7.34 (m, 3 H), 5.56 (s, 1 H), 5.34 (dd, *J* = 10, 2.8 Hz, 1 H), 4.76 (s, 1 H), 4.29 (dd, *J* = 9.6, 4 Hz, 1 H), 4.14 (br, 1 H), 4.09 (t, *J* = 9.6 Hz, 1 H), 3.95–3.83 (m, 2 H), 3.41 (s, 3 H), 2.62 (sept, *J* = 7.2 Hz, 1 H), 1.18 (dd, *J* = 6.8, 4 Hz, 6 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 176.1, 137.4, 129.0, 128.3, 126.1, 101.7, 101.5, 76.4, 70.4, 70.1, 68.9, 63.7, 55.2, 34.2, 19.3, 19.0. HRMS (ESI) *m/z*: [M + Na]$^+$ calcd for $C_{18}H_{24}O_7Na$, 375.1420, found 375.1423.

Methyl 4,6-*O*-benzylidene-3-*O*-pivaloyl-α-D-mannopyranoside (36). A mixture of methyl 4,6-*O*-benzylidene-α-D-mannopyranoside (**29**) (25 mg, 0.09 mmol), **1** (0.60 mg, 0.0013 mmol) and pivalic anhydride (39.3 μL, 0.19 mmol) was stirred in MeCN (1 mL) for 16 h and worked-up as given in the general procedure A. Compound **36** [13] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 3 : 1) as a colorless gum (8.4 mg, 26%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.43–7.42 (m, 2 H), 7.35–7.33 (m, 3 H), 5.57 (s, 1 H), 5.33 (dd, *J* = 10.4, 3.2 Hz, 1 H), 4.75 (s, 1 H), 4.30 (dd, *J* = 9.6, 4 Hz, 1 H), 4.13–4.07 (m, 2 H), 3.94–3.85 (m, 2 H), 3.40 (s, 3 H), 1.23 (s, 9 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 177.3, 137.4, 128.9, 128.3, 126.0, 101.6, 101.5, 76.4, 70.5, 70.1, 68.9, 63.7, 55.2, 39.2, 27.3. HRMS (ESI) *m/z*: [M + Na]$^+$ calcd for $C_{19}H_{26}O_7Na$, 389.1576, found 389.1576.

Methyl 4,6-*O*-benzylidene-3-*O*-benzoyl-α-D-mannopyranoside (38). A mixture of methyl 4,6-*O*-benzylidene-α-D-mannopyranoside (**29**) (25 mg, 0.09 mmol), **1** (0.60 mg, 0.0013 mmol) and benzoic anhydride (43.9 mg, 0.19 mmol) was stirred in MeCN (1 mL) for 10 h and worked-up as given in the general procedure A. Compound **38** [72] was secured after column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc 3 : 1) as a colorless solid (19.1 mg, 56%).^{1}H NMR (400 MHz, $CDCl_3$) δ 8.10–8.06 (m, 2 H), 7.63–7.55 (m, 1 H), 7.49–7.42 (m, 4 H), 7.32–7.31 (m, 3 H), 5.60 (s, 1 H), 5.56 (dd, *J* = 10, 3.2 Hz, 1 H), 4.80 (s, 1 H), 4.34–4.25 (m, 3 H), 4.00 (td, *J* = 10, 4.8 Hz, 1 H), 3.91 (t, *J* = 10.4 Hz, 1 H), 3.44 (s, 3 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 165.6, 137.4, 133.4, 130.3, 129.9, 129.1, 128.6, 128.3, 126.3, 101.9, 101.6, 76.3, 71.6, 69.9, 69.0, 63.9, 55.3. HRMS (ESI) *m/z*: [M + Na]$^+$ calcd for $C_{21}H_{22}O_7Na$, 409.1263, found 409.1262.

General procedure B: regioselective acylation of the 6-*O*-protected pyranoside triols. The carbohydrate starting material (1 mol. equiv.) and desired catalyst (either 1.5 mol% of **1** or 4.5 mol% DMAP catalyst) were dissolved in $CHCl_3$, desired carboxylic acid anhydride (1.1 molar equiv. of acetic anhydride, isobutyric anhydride and 2.2 molar equiv. of pivalic and benzoic anhydride) was added and stirred from 0 °C to room temperature for a desired time. Afterwards, 1 mL of MeOH was

 This journal is © The Royal Society of Chemistry 2024

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

added and stirred for a further 30 min to quench the reaction. The solvent was evaporated under reduced pressure and the residue analyzed by HPLC (Chromatography condition: analytical silica column (5 μm), *n*-hexane/EtOAc eluant system, flow rate 1 mL min^{-1}, PDA detector (254 nm)) to determine the ratio of the regioisomers. The crude product was purified by column chromatography (SiO_2) (eluant: *n*-hexane/EtOAc) to secure the major regioisomeric product.

Methyl 3-*O*-acetyl-6-*O*-trityl-α-D-glucopyranoside (41). A mixture of methyl 6-*O*-trityl-α-D-glucopyranoside (**40**) (25 mg, 0.057 mmol), **1** (0.39 mg, 0.001 mmol) and acetic anhydride (6.3 μL, 0.063 mmol) was stirred in $CHCl_3$ (1 mL) for 45 min and worked-up as given in the general procedure B. Compound **41** [73] was secured after column chromatography (SiO_2) (eluant: *n*-hexane–EtOAc 3 : 2), as a colorless gum (16.9 mg, 62%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.46–7.44 (m, 6 H), 7.33–7.29 (m, 6 H), 7.26–7.23 (m, 3 H), 5.05 (t, *J* = 9.4 Hz, 1 H), 4.78 (d, *J* = 4 Hz, 1 H), 3.71–3.66 (m, 1 H), 3.61 (t, *J* = 9.2 Hz, 2 H), 3.44 (s, 3 H), 3.41–3.39 (m, 2 H), 2.15 (s, 3 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 172.4, 143.8, 128.8, 128.1, 127.3, 99.3, 87.4, 76.5, 71.0, 70.7, 70.3, 64.2, 55.4, 21.2. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd For $C_{28}H_{30}O_7Na$, 501.1889, found 501.1888.

Methyl 3-*O*-isobutyryl-6-*O*-trityl-α-D-glucopyranoside (43). A mixture of methyl 6-*O*-trityl-α-D-glucopyranoside (**40**) (25 mg, 0.057 mmol), **1** (0.39 mg, 0.001 mmol) and acetic anhydride (10.5 μL, 0.063 mmol) was stirred in $CHCl_3$ (1 mL) for 45 min and worked-up as given in the general procedure B. Compound **43** was secured after column chromatography (SiO_2) (eluant : *n*-hexane–EtOAc 7 : 3), as a colorless gum (20.3 mg, 70%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.47–7.45 (m, 6 H), 7.33–7.29 (m, 6 H), 7.24–7.22 (m, 3 H), 5.03 (t, *J* = 9.4 Hz, 1 H), 4.79 (d, *J* = 4 Hz, 1 H), 3.73–3.68 (m, 1 H), 3.59–3.54 (m, 2 H), 3.46 (s, 3 H), 3.43–3.35 (m, 2 H), 2.63 (sept, *J* = 7 Hz, 1 H), 1.19 (dd, *J* = 6.8, 5.2 Hz, 6 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 178.7, 143.8, 128.8, 128.0, 127.3, 99.3, 87.3, 76.4, 71.0, 70.6, 70.5, 64.2, 55.4, 34.3, 19.1, 19.0. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd For $C_{30}H_{34}O_7Na$, 529.2202, found 529.2200.

Methyl 3-*O*-pivaloyl-6-*O*-trityl-α-D-glucopyranoside (45). A mixture of methyl 6-*O*-trityl-α-D-glucopyranoside (**40**) (30 mg, 0.068 mmol), **1** (0.47 mg, 0.001 mmol) and pivalic anhydride (30.3 μL, 0.15 mmol) was stirred in $CHCl_3$ (1 mL) for 16 h and worked-up as given in the general procedure B. Compound **45** was secured after column chromatography (SiO_2) (eluant : *n*-hexane–EtOAc 4 : 1) as a colorless gum (25.7 mg, 72%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.48–7.46 (m, 6 H), 7.33–7.29 (m, 6 H), 7.26–7.22 (m, 3 H), 5.02 (t, *J* = 9.4 Hz, 1 H), 4.80 (d, *J* = 3.6 Hz, 1 H), 3.76–3.71 (m, 1 H), 3.62 (br, 1 H), 3.54–3.52 (m, 1 H), 3.48 (s, 3 H), 3.45–3.44 (m, 1 H), 3.36 (dd, *J* = 10, 5.6 Hz, 1 H), 1.23 (s, 9 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 180.2, 143.9, 128.8, 128.0, 127.2, 99.2, 87.2, 76.6, 71.0, 70.6, 70.5, 64.1, 55.3, 39.1, 27.2. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd for $C_{31}H_{36}O_7Na$, 543.2359, found 543.2357.

Methyl 3-*O*-benzoyl-6-*O*-trityl-α-D-glucopyranoside (47). A mixture of methyl 6-*O*-trityl-α-D-glucopyranoside (**40**) (25 mg, 0.057 mmol), **1** (0.39 mg, 0.001 mmol) and benzoic anhydride (28.3 mg, 0.125 mmol) was stirred in $CHCl_3$ (1 mL) for 8 h and worked-up as given in the general procedure B. Compound **47** was secured after column chromatography (SiO_2) (eluant : *n*-hexane–EtOAc 3 : 2) as a colorless gum (18.8 mg, 61%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 8.10–8.06 (m, 2 H), 7.63–7.54 (m, 1 H), 7.49–7.47 (m, 6 H), 7.45–7.41 (m, 2 H), 7.33–7.29 (m, 6 H), 7.25–7.23 (m, 3 H), 5.32 (t, *J* = 9.2 Hz, 1 H), 4.85 (d, *J* = 3.6 Hz, 1 H), 3.82–3.75 (m, 3 H), 3.48 (s, 3 H), 3.43 (td, *J* = 12, 4 Hz, 2 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 167.9, 143.8, 133.4, 130.2, 130.0, 128.7, 128.4, 128.0, 127.2, 99.3, 87.2, 76.8, 71.0, 70.7, 70.4, 64.1, 55.4. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd for $C_{33}H_{32}O_7Na$, 563.2046, found 563.2046.

Methyl 2/3-*O*-acetyl-6-*O*-trityl-α-D-mannopyranoside (51/50). A mixture of methyl 6-*O*-trityl-α-D-mannopyranoside (**49**) (30 mg, 0.068 mmol), **1** (0.47 mg, 0.001 mmol) and acetic anhydride (7.0 μL, 0.074 mmol) was stirred in $CHCl_3$ (1 mL) for 45 min and worked-up as given in the general procedure B. Compound **51/50** [74] was secured after column chromatography (SiO_2) (eluant : *n*-hexane–EtOAc 3 : 2) as a colorless foam (17.1 mg, 52%) and as an inseparable regioisomeric mixture of 2-OAc/3-OAc (70 : 30). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.31–7.3 (m, 7 H), 7.28 (br, 10 H), 7.27–7.26 (m, 2 H), 5.08 (d, *J* = 4 Hz, 1 H), 5.06 (d, *J* = 3.2 Hz, 0.4 H), 4.71 (s, 1 H), 4.68 (s, 0.5 H), 4.02 (br, 1 H), 3.97–3.96 (m, 2 H), 3.93–3.91 (m, 1.4 H), 3.87–3.80 (m, 1 H), 3.64–3.57 (m, 1.4 H), 3.38 (s, 3 H), 3.36 (s, 1 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 171.1, 170.9, 143.8, 143.7, 128.8, 128.7, 128.1, 128.0, 127.3, 127.1, 100.6, 100.5, 87.5, 87.2, 74.3, 71.9, 70.8, 70.7, 70.0, 69.4, 69.3, 67.8, 64.7, 62.8, 55.13, 55.10, 21.3, 21.2. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd for $C_{28}H_{30}O_7Na$, 501.1889, found 501.1886.

Methyl 3-*O*-isobutyryl-6-*O*-trityl-α-D-mannopyranoside (52). A mixture of methyl 6-*O*-trityl-α-D-mannopyranoside (**49**) (30 mg, 0.068 mmol), **1** (0.47 mg, 0.001 mmol) and isobutyric anhydride (12.3 μL, 0.074 mmol) was stirred in $CHCl_3$ (1 mL) for 30 min and worked-up as given in the general procedure B. Compound **52** was secured after column chromatography (SiO_2) (eluant : *n*-hexane–EtOAc 7 : 3) as a colorless gum (20.1 mg, 58%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.47–7.45 (m, 6 H), 7.32–7.28 (m, 7 H), 7.26–7.22 (m, 2 H), 5.06 (dd, *J* = 10, 3.4 Hz, 1 H), 4.72 (d, *J* = 1.2 Hz, 1 H), 3.98 (br, 1 H), 3.91–3.87 (m, 1 H), 3.76–3.71 (m, 1 H), 3.45 (t, *J* = 4.6 Hz, 2 H), 3.41 (s, 3 H), 2.64 (sept, *J* = 6.8 Hz, 1 H), 1.19 (d, *J* = 8 Hz, 6 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 177.3, 143.8, 128.7, 128.0, 127.3, 100.5, 87.4, 74.1, 71.0, 69.4, 67.9, 64.6, 55.1, 34.1, 19.15, 19.12. HRMS (ESI) *m*/*z*: $[M + Na]^+$ calcd for $C_{30}H_{34}O_7Na$, 529.2202, found 529.2206.

Methyl 3-*O*-pivaloyl-6-*O*-trityl-α-D-mannopyranoside (54). A mixture of methyl 6-*O*-trityl-α-D-mannopyranoside (**49**) (30 mg, 0.068 mmol), **1** (0.47 mg, 0.001 mmol) and pivalic anhydride (30.3 μL, 0.149 mmol) was stirred in $CHCl_3$ (1 mL) for 16 h and worked-up as given in the general procedure B. Compound **54** was secured after column chromatography (SiO_2) (eluant : *n*-hexane–EtOAc 4 : 1) as a colorless gum (15.0 mg, 42%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.48–7.46 (m, 6 H), 7.32–7.28 (m, 7 H), 7.24–7.22 (m, 2 H), 5.03 (dd, *J* = 9.6, 3.2 Hz, 1 H), 4.73 (d, *J* = 1.6 Hz, 1 H), 3.98–3.96 (m, 1 H), 3.89–3.82

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

(m, 1 H), 3.76–3.71 (m, 1 H), 3.49–3.44 (m, 1 H), 3.42 (s, 3 H), 3.40 (s, 1 H), 1.23 (s, 9 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 178.7, 143.8, 128.8, 128.0, 127.2, 100.5, 87.3, 74.2, 71.1, 69.4, 55.1, 39.1, 27.3. HRMS (ESI) *m*/*z*: [M + K] $^+$ calcd For $C_{31}H_{36}O_7K$, 559.2098, found 559.2095.

Methyl 3-*O*-benzoyl-6-*O*-trityl-α-D-mannopyranoside (56). A mixture of methyl 6-*O*-trityl-α-D-mannopyranoside (**49**) (30 mg, 0.068 mmol), **1** (0.47 mg, 0.001 mmol) and benzoic anhydride (33.7 mg, 0.15 mmol) was stirred in $CHCl_3$ (1 mL) for 8 h and worked-up as given in the general procedure B. Compound **56** [70] was secured after column chromatography (SiO_2) (eluant : *n*-hexane–EtOAc 4 : 1) as a colorless gum (17.4 mg, 47%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 8.12–8.07 (m, 4 H), 7.63–7.55 (m, 2 H), 7.49–7.42 (m, 8 H), 7.32–7.29 (m, 4 H), 7.24–7.22 (m, 2 H), 5.33 (dd, *J* = 9.6, 3.2 Hz, 1 H), 4.77 (d, *J* = 1.2 Hz, 1 H), 4.15–4.07 (m, 2 H), 3.84–3.79 (m, 1 H), 3.49 (d, *J* = 4.8 Hz, 2 H), 3.43 (s, 3 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 166.6, 143.7, 133.8, 130.3, 130.0, 128.7, 128.6, 128.1, 127.3, 100.6, 87.5, 75.0, 70.8, 69.4, 68.0, 64.7, 55.2. HRMS (ESI) *m*/*z*: [M + K]$^+$ calcd for $C_{33}H_{32}O_7K$, 579.1785, found 579.1786.

Methyl 3-*O*-acetyl-6-*O*-trityl-α-D-galactopyranoside (59). A mixture of methyl 6-*O*-trityl-α-D-galactopyranoside (**58**) (30 mg, 0.068 mmol), **1** (0.47 mg, 0.001 mmol) and acetic anhydride (7.0 μL, 0.074 mmol) was stirred in $CHCl_3$ (1 mL) for 45 min and worked-up as given in the general procedure B. Compound **59** was secured after column chromatography (SiO_2) (eluant: *n*-hexane–EtOAc 3 : 2) as a colorless gum (6.5 mg, 20%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.45–7.43 (m, 6 H), 7.32–7.28 (m, 6 H), 7.24–7.22 (m, 3 H), 5.03 (dd, *J* = 10, 2.8 Hz, 1 H), 4.86 (d, *J* = 4 Hz, 1 H), 4.09 (app. d, *J* = 2 Hz, 1 H), 4.05 (dd, *J* = 10, 3.6 Hz, 1 H), 3.86 (t, *J* = 5.2 Hz, 1 H), 3.44 (s, 3 H), 3.42–3.34 (m, 2 H), 2.16 (s, 3 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 171.0, 143.5, 128.5, 127.9, 127.2, 99.6, 87.2, 73.3, 69.1, 68.7, 67.3, 63.5, 55.4, 21.2. HRMS (ESI) *m*/*z*: [M + Na]$^+$ calcd for $C_{28}H_{30}O_7Na$, 501.1889, found 501.1890.

Methyl 3-*O*-isobutyryl-6-*O*-trityl-α-D-galactopyranoside (61). A mixture of methyl 6-*O*-trityl-α-D-galactopyranoside (**58**) (30 mg, 0.068 mmol), **1** (0.47 mg, 0.001 mmol) and isobutyric anhydride (12.3 μL, 0.074 mmol) was stirred in $CHCl_3$ (1 mL) for 30 min and worked-up as given in the general procedure B. Compound **61** was secured after column chromatography (SiO_2) (eluant : *n*-hexane–EtOAc 3 : 2) as a colorless gum (14.6 mg, 42%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.46–7.44 (m, 6 H), 7.32–7.28 (m, 6 H), 7.24–7.22 (m, 3 H), 5.02 (dd, *J* = 10.4, 2.8 Hz, 1 H), 4.86 (d, *J* = 4 Hz, 1 H), 4.05 (app. d, *J* = 2 Hz, 2 H), 3.87 (t, *J* = 5.4 Hz, 1 H), 3.45 (s, 3 H), 3.38 (d, *J* = 5.2 Hz, 2 H), 2.66 (sept, *J* = 7 Hz, 1 H), 1.20 (dd, *J* = 6.8, 5.2 Hz, 6 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 177.3, 143.7, 128.7, 128.0, 127.3, 99.8, 87.3, 73.1, 69.2, 68.9, 67.5, 63.6, 55.5, 34.1, 19.2, 19.1. HRMS (ESI) *m*/*z*: [M + Na]$^+$ calcd for $C_{30}H_{34}O_7Na$, 529.2202, found 529.2206.

Methyl 3/4-*O*-pivaloyl-6-*O*-trityl-α-D-galactopyranoside (63/64). A mixture of methyl 6-*O*-trityl-α-D-galactopyranoside (**58**) (30 mg, 0.068 mmol), **1** (0.47 mg, 0.001 mmol) and pivalic anhydride (30.3 μL, 0.149 mmol) was stirred in 1 mL of $CHCl_3$ for 16 h and worked-up as given in the general procedure B. Compound **63**/**64** was secured after column chromatography (SiO_2) (eluant : *n*-hexane/EtOAc 3 : 2) as a colorless gum and as an inseparable mixture of 3-*O*-pivaloyl/2-*O*-pivaloyl (73 : 27) regioisomers (15.6 mg, 45%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 7.47–7.44 (m, 8 H), 7.32–7.28 (m, 9 H), 7.24–7.22 (m, 2 H), 5.00 (dd, *J* = 10.4, 3.2 Hz, 1 H), 4.94 (dd, *J* = 10, 3.6 Hz, 0.37 H), 4.90 (d, *J* = 3.6 Hz, 0.36 H), 4.85 (d, *J* = 4 Hz, 1 H), 4.04–3.94 (m, 3 H), 3.89–3.85 (m, 1.52 H), 3.46 (s, 3 H), 3.42–3.38 (m, 1.27 H), 3.37 (s, 0.9 H), 3.36–3.33 (m, 0.6 H), 1.24 (s, 9 H), 1.23 (s, 2.50 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 179.1, 178.5, 143.8, 143.7, 128.7, 128.1, 128.0, 127.3, 127.2, 99.7, 97.5, 87.2, 73.2, 71.7, 70.3, 69.1, 69.0, 68.6, 68.5, 67.6, 63.5, 55.6, 55.5, 39.1, 39.0, 27.3, 27.1. HRMS (ESI) *m*/*z*: [M + Na]$^+$ calcd for $C_{31}H_{36}O_7Na$, 543.2356, found 543.2358.

Methyl 3-*O*-benzoyl-6-*O*-trityl-α-D-galactopyranoside (65). A mixture of methyl 6-*O*-trityl-α-D-galactopyranoside (**58**) (30 mg, 0.068 mmol), **1** (0.47 mg, 0.001 mmol) and benzoic anhydride (33.7 mg, 0.149 mmol) was stirred in $CHCl_3$ (1 mL) for 8 h and worked-up as given in the general procedure B. Compound **65** [70] was secured after column chromatography (SiO_2) (eluant : *n*-hexane–EtOAc 3 : 2) as a colorless gum (16.3 mg, 44%). ^{1}H NMR (400 MHz, $CDCl_3$) δ 8.11–8.09 (m, 3 H), 7.63–7.54 (m, 2 H), 7.49–7.41 (m, 9 H), 7.32–7.29 (m, 4 H), 7.23–7.22 (m, 2 H), 5.26 (dd, *J* = 10, 2.8 Hz, 1 H), 4.92 (d, *J* = 4 Hz, 1 H), 4.24–4.20 (m, 2 H), 3.94 (dd, *J* = 10.8, 5.6 Hz, 1 H), 3.48 (s, 3 H), 3.41 (dd, *J* = 5.2, 2 Hz, 2 H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 166.6, 143.6, 133.4, 130.3, 130.0, 128.7, 128.5, 128.0, 127.3, 99.8, 87.3, 74.1, 69.2, 68.9, 67.5, 63.5, 55.6. HRMS (ESI) *m*/*z*: [M + Na]$^+$ calcd for $C_{33}H_{32}O_7Na$, 563.2046, found 563.2048.

Author contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

This research is supported by a generous funding by Science and Engineering Research Board, DST, New Delhi. K. D. acknowledges Council of Industrial and Scientific Research, New Delhi, for a research fellowship.

References

1 A. Varki, *Glycobiology*, 2017, **27**, 3–49.
2 Y. van Kooyk and G. A. Rabinovich, *Nat. Immunol.*, 2008, **9**, 593–601.

 This journal is © The Royal Society of Chemistry 2024

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

3 S. Liedtke, H. Geyer, M. Wuhrer, R. Geyer, G. Frank, R. Gerardy-Schahn, U. Zahringer and M. Schachner, *Glycobiology*, 2001, **11**, 373–384.

4 (*a*) P. H. Seeberger and D. B. Werz, *Nat. Rev. Drug Discovery*, 2005, **4**, 751–763; (*b*) S. S. Shivatare, V. S. Shivatare and C.-H. Wong, *Chem. Rev.*, 2022, **122**, 15603–15671; (*c*) M. Panza, S. G. Pistorio, K. J. Stine and A. V. Demchenko, *Chem. Rev.*, 2018, **118**, 8105–8150; (*d*) L. Krasnova and C.-H. Wong, *J. Am. Chem. Soc.*, 2019, **141**, 3735–3754; (*e*) C.-W. Cheng, Y. Zhou, W.-H. Pan, S. Dey, C.-Y. Wu, W.-L. Hsu and C.-H. Wong, *Nat. Commun.*, 2018, **9**, 5202; (*f*) O. J. Plante, E. R. Palmacci and P. H. Seeberger, *Science*, 2001, **291**, 1523–1527.

5 (*a*) Y. Maki, R. Okamoto, M. Izumi, T. Murase and Y. Kajihara, *J. Am. Chem. Soc.*, 2016, **138**, 3461–3468; (*b*) X. Qian, K. Sujino, A. Otter, M. M. Palcic and O. Hindsgaul, *J. Am. Chem. Soc.*, 1999, **121**, 12063–12072; (*c*) K. Moremen, A. Ramiah, M. Stuart, J. Steel, L. Meng, F. Forouhar, H. A. Moniz, G. Gahlay, Z. Gao, D. Chapla, S. Wang, J.-H. Yang, P. K. Prabhakar, R. Johnson, M. D. Rosa, C. Geisler, A. V. Nairn, J. Seetharaman, S.-C. Wu, L. Tong, H. J. Gilbert, J. LaBaer and D. J. Jarvis, *Nat. Chem. Biol.*, 2018, **14**, 156–162; (*d*) C. E. Council, K. J. Kilpin, J. S. Gusthart, S. A. Allman, B. Linclau and S. S. Lee, *Org. Biomol. Chem.*, 2020, **18**, 3423–3451.

6 (*a*) P. G. M. Wuts and T. W. Greene, *Greene's Protective Groups in Organic Synthesis*, Wiley, Hoboken, NJ, 4th edn, 2007; (*b*) P. G. Goekjian and S. Vidal, Regioselective Protection at the Secondary Positions of Carbohydrates with Acyclic Protecting Groups, in *Protecting Groups – Strategies and Applications in Carbohydrate Chemistry*, ed. S. Vidal, Wiley-VCH, Weinheim, 2019, ch. 4, pp. 109–144.

7 (*a*) B. Dhakal and D. Crich, *J. Am. Chem. Soc.*, 2018, **140**, 15008–15015; (*b*) A. A. Hettikankanamalage, R. Lassfolk, F. S. Ekholm, R. Leino and D. Crich, *Chem. Rev.*, 2020, **120**, 7104–7151.

8 (*a*) V. Dimakos and M. S. Taylor, *Chem. Rev.*, 2018, **118**, 11457–11517; (*b*) K. Yamatsugu and M. Kanai, *Chem. Rev.*, 2023, **123**, 6793–6838; (*c*) M. Jäger and A. J. Minnaard, *Chem. Commun.*, 2016, **52**, 656–664; (*d*) M. D. Witte and A. J. Minnaard, *ACS Catal.*, 2022, **12**, 12195–12205; (*e*) K. Dey and N. Jayaraman, *Chem. Commun.*, 2022, **58**, 2224–2227; (*f*) K. Dey and N. Jayaraman, *Carbohydr. Res.*, 2022, **520**, 108610.

9 (*a*) A. Francais, D. Urban and J.-M. Beau, *Angew. Chem., Int. Ed.*, 2007, **46**, 8662–8665; (*b*) S. S. Kulkarni, C.-C. Wang, N. M. Sabbavarapu, A. R. Podilapu, P.-H. Liao and S.-C. Hung, *Chem. Rev.*, 2018, **118**, 8025–8104; (*c*) Z. Zhang, I. R. Ollmann, X.-S. Ye, R. Wischnat, T. Baasov and C.-H. Wong, *J. Am. Chem. Soc.*, 1999, **121**, 734–753.

10 (*a*) T. Kurahashi, T. Mizutani and J.-i. Yoshida, *J. Chem. Soc., Perkin Trans. 1*, 1999, 465–473; (*b*) T. Kurahashi, T. Mizutani and J.-i. Yoshida, *Tetrahedron*, 2002, **58**, 8669–8677.

11 E. Kattnig and M. Albert, *Org. Lett.*, 2004, **6**, 945–948.

12 (*a*) J. Lawandi, S. Rocheleau and N. Moitessier, *Tetrahedron*, 2016, **72**, 6283–6319; (*b*) N. Moitessier, P. Englebienne and Y. Chapleur, *Tetrahedron*, 2005, **61**, 6839–6853; (*c*) N. Moitessier and Y. Chapleur, *Tetrahedron Lett.*, 2003, **44**, 1731–1735; (*d*) S. Rocheleau, J. Pottel, I. Huskic and N. Moitessier, *Eur. J. Org. Chem.*, 2017, 646–656; (*e*) J. Lawandi, S. Rocheleau and N. Moitessier, *Tetrahedron*, 2011, **67**, 8411–8420.

13 L. Jiang and T.-H. Chan, *J. Org. Chem.*, 1998, **63**, 6035–6038.

14 F. A. Carey and K. O. Hodgson, *Carbohydr. Res.*, 1970, **12**, 463–465.

15 S. A. Abbas and A. H. Haines, *Carbohydr. Res.*, 1975, **39**, 358–363.

16 S. Kim, H. Chang and W. J. Kim, *J. Org. Chem.*, 1985, **50**, 1751–1752.

17 (*a*) P. Peng, M. Linseis, R. F. Winter and R. R. Schmidt, *J. Am. Chem. Soc.*, 2016, **138**, 6002–6009; (*b*) T. Li, T. Li, M. Linseis, F. Wang, R. F. Winter, R. R. Schmidt and P. Peng, *ACS Catal.*, 2020, **10**, 11406–11416.

18 G. Ma, J. Deng and M. P. Sibi, *Angew. Chem., Int. Ed.*, 2014, **53**, 11818–11821.

19 (*a*) M. R. Heinrich, H. S. Klisa, H. Mayr, W. Steglich and H. Zipse, *Angew. Chem., Int. Ed.*, 2003, **42**, 4826–4828; (*b*) R. Tandon, T. Unzner, T. A. Nigst, N. De Rycke, P. Mayer, B. Wendt, O. R. P. David and H. Zipse, *Chem. – Eur. J.*, 2013, **19**, 6435–6442.

20 M. R. Crittall, H. S. Rzepa and D. R. Carbery, *Org. Lett.*, 2011, **13**, 1250–1253.

21 S. Singh, G. Das, O. V. Singh and H. Han, *Org. Lett.*, 2007, **9**, 401–404.

22 N. D. Rycke, G. Berionni, F. Couty, H. Mayr, R. Goumont and O. R. P. David, *Org. Lett.*, 2011, **13**, 530–533.

23 (*a*) J. C. Ruble, H. A. Latham and G. C. Fu, *J. Am. Chem. Soc.*, 1997, **119**, 1492–1493; (*b*) J. C. Ruble, J. Tweddell and G. C. Fu, *J. Org. Chem.*, 1998, **63**, 2794–2795.

24 (*a*) T. Kawabata, M. Nagato, K. Takasu and K. Fuji, *J. Am. Chem. Soc.*, 1997, **119**, 3169–3170; (*b*) H. Mandai, K. Fujii, H. Yasuhara, K. Abe, K. Mitsudo, T. Korenaga and S. Suga, *Nat. Commun.*, 2016, **7**, 11297.

25 (*a*) A. C. Spivey, T. Fekner and S. E. Spey, *J. Org. Chem.*, 2000, **65**, 3154–3159; (*b*) E. Larionov, M. Mahesh, A. C. Spivey, Y. Wei and H. Zipse, *J. Am. Chem. Soc.*, 2012, **134**, 9390–9399.

26 R. P. Wurz, *Chem. Rev.*, 2007, **107**, 5570–5595.

27 (*a*) T. Kawabata, W. Muramatsu, T. Nishio, T. Shibata and H. Schedel, *J. Am. Chem. Soc.*, 2007, **129**, 12890–12895; (*b*) Y. Ueda, W. Muramatsu, K. Mishiro, T. Furuta and T. Kawabata, *J. Org. Chem.*, 2009, **74**, 8802–8805; (*c*) M. Yanagi, Y. Ueda, R. Ninomiya, A. Imayoshi, T. Furuta, K. Mishiro and T. Kawabata, *Org. Lett.*, 2019, **21**, 5006–5009.

28 (*a*) F. Huber and S. F. Kirsch, *Chem. – Eur. J.*, 2016, **22**, 5914–5918; (*b*) M. L. Tong, F. Huber, E. S. T. Kaptouom, T. Cellnik and S. F. Kirsch, *Chem. Commun.*, 2017, **53**, 3086–3089.

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .

29 (*a*) K. S. Griswold and S. J. Miller, *Tetrahedron*, 2003, **59**, 8869–8875; (*b*) M. Sánchez-Roselló, A. L. A. Puchlopek, A. J. Morgan and S. J. Miller, *J. Org. Chem.*, 2008, **73**, 1774–1782.
30 (*a*) A. Seitz, R. C. Wende, E. Roesner, D. Niedek, C. Topp, A. C. Colgan, E. M. McGarrigle and P. R. Schreiner, *J. Org. Chem.*, 2021, **86**, 3907–3922; (*b*) A. Seitz, R. C. Wende and P. R. Schreiner, *Chem. – Eur. J.*, 2023, **29**, e202203002; (*c*) D. Niedek, F. R. Erb, C. Topp, A. Seitz, R. C. Wende, A. K. Eckhardt, J. Kind, D. Herold, C. M. Thiele and P. R. Schreiner, *J. Org. Chem.*, 2020, **85**, 1835–1846.
31 X. Sun, H. Lee, S. Lee and K. L. Tan, *Nat. Chem.*, 2013, **5**, 790–795.
32 (*a*) G. Xiao, G. A. Cintron-Rosado, D. A. Glazier, B. Xi, C. Liu, P. Liu and W. Tang, *J. Am. Chem. Soc.*, 2017, **139**, 4346–4349; (*b*) S. A. Blaszczyk, G. Xiao, P. Wen, H. Hao, J. Wu, B. Wang, F. Carattino, Z. Li, D. A. Glazier, B. J. McCarty, P. Liu and W. Tang, *Angew. Chem., Int. Ed.*, 2019, **58**, 9542–9546.
33 D. L. Cramer, S. Bera and A. Studer, *Chem. – Eur. J.*, 2016, **22**, 7403–7407.
34 W.-X. Lv, H. Chen, X. Zhang, C. C. Ho, Y. Liu, S. Wu, H. Wang, Z. Jin and Y. R. Chi, *Chem*, 2022, **8**, 1518–1534.
35 C. L. Allen and S. J. Miller, *Org. Lett.*, 2013, **15**, 6178–6181.
36 I.-H. Chen, K. G. M. Kou, D. N. Le, C. M. Rathbun and V. M. Dong, *Chem. – Eur. J.*, 2014, **20**, 5013–5018.
37 (*a*) E. Mensah, N. Camasso, W. Kaplan and P. Nagorny, *Angew. Chem., Int. Ed.*, 2013, **52**, 12932–12936; (*b*) S. Wang, O. Zhelavskyi, J. Lee, A. J. Arguelles, Y. Y. Khomutnyk, E. Mensah, H. Guo, R. Hourani, P. M. Zimmerman and P. Nagorny, *J. Am. Chem. Soc.*, 2021, **143**, 18592–18604.
38 (*a*) Z. Zhang and C.-H. Wong, *Tetrahedron*, 2002, **58**, 6513–6519; (*b*) Y. Zhou, K. S. Liao, T. Y. Chen, Y. S. Y. Hsieh and C.-H. Wong, *J. Org. Chem.*, 2023, **88**, 7141–7151.
39 (*a*) Y. Demizu, Y. Kubo, H. Miyoshi, T. Maki, Y. Matsumura, N. Moriyama and O. Onomura, *Org. Lett.*, 2008, **10**, 5075–5077; (*b*) F. Iwasaki, T. Maki, O. Onomura, W. Nakashima and Y. Matsumura, *J. Org. Chem.*, 2000, **65**, 996–1002; (*c*) W. Muramatsu, *J. Org. Chem.*, 2012, **77**, 8083–8091.
40 J. Lv, T. Luo, Y. Zhang, Z. Pei and H. Dong, *ACS Omega*, 2018, **3**, 17717–17723.
41 Y. X. Zhou, O. Ramstrom and H. Dong, *Chem. Commun.*, 2012, **48**, 5370–5372.
42 E. V. Evtushenko, *Carbohydr. Res.*, 2012, **359**, 111–119.
43 H. Wang, J. She, L.-H. Zhang and X.-S. Ye, *J. Org. Chem.*, 2004, **69**, 5774–5777.
44 (*a*) D. Lee and M. S. Taylor, *J. Am. Chem. Soc.*, 2011, **133**, 3724–3727; (*b*) N. Shimada, Y. Nakamura, T. Ochiai and K. Makino, *Org. Lett.*, 2019, **21**, 3789–3794.
45 (*a*) D. J. Gorelik, J. A. Turner and M. S. Taylor, *Org. Lett.*, 2022, **24**, 5249–5253; (*b*) D. Gorelik, Y. C. Lin, A. L. Briceno-Strocchia and M. S. Taylor, *J. Org. Chem.*, 2019, **84**, 900–908.
46 D. Lee, C. L. Williamson, L. Chan and M. S. Taylor, *J. Am. Chem. Soc.*, 2012, **134**, 8260–8267.
47 S. Kuwano, Y. Hosaka and T. Arai, *Chem. – Eur. J.*, 2019, **25**, 12920–12923.
48 J. Alsarraf, L. Petitpoisson and A. Pichette, *Org. Lett.*, 2021, **23**, 6052–6056.
49 N. Shimada, T. Sugimoto, M. Noguchi, C. Ohira, Y. Kuwashima, N. Takahashi, N. Sato and K. Makino, *J. Org. Chem.*, 2021, **86**, 5973–5982.
50 (*a*) K. A. D'Angelo and M. S. Taylor, *J. Am. Chem. Soc.*, 2016, **138**, 11058–11066; (*b*) C. Gouliaras, D. Lee, L. Chan and M. S. Taylor, *J. Am. Chem. Soc.*, 2011, **133**, 13926–13929.
51 (*a*) B. Neises and W. Steglich, *Angew. Chem., Int. Ed. Engl.*, 1978, **17**, 522–524; (*b*) S. Xu, B. Kempf, H. Mayr, W. Steglich and H. Zipse, *Chem. – Eur. J.*, 2005, **11**, 4751–4757.
52 (*a*) A. Deratani, G. D. Darling, D. Horak and J. M. J. Fréchet, *Macromolecules*, 1987, **20**, 767–772; (*b*) A. Deratani, G. D. Darling and J. M. J. Fréchet, *Polymer*, 1987, **28**, 825–830; (*c*) E. J. Delaney, L. E. Wood and I. M. Klotz, *J. Am. Chem. Soc.*, 1982, **104**, 799–807; (*d*) F. M. Menger and D. J. McCann, *J. Org. Chem.*, 1985, **50**, 3928–3930; (*e*) G.-J. Wang and W. K. Fife, *J. Am. Chem. Soc.*, 1998, **120**, 883–887; (*f*) H. T. Chen, S. Huh, J. W. Wiench, M. Pruski and V. S. Y. Lin, *J. Am. Chem. Soc.*, 2005, **127**, 13305–13311.
53 (*a*) A. Fallek, M. Weiss-Shtofman, M. Kramer, R. Dobrovetsky and M. Portnoy, *Org. Lett.*, 2020, **22**, 3722–3727; (*b*) A. Fallek, O. Fleischer, F. Saady and M. Portnoy, *Eur. J. Org. Chem.*, 2022, e202201089; (*c*) N. Ashush, R. Fallek, A. Fallek, R. Dobrovetsky and M. Portnoy, *Org. Lett.*, 2020, **22**, 3749–3754; (*d*) R. Fallek, N. Ashush, A. Fallek, O. Fleischer and M. Portnoy, *J. Org. Chem.*, 2022, **87**, 9688–9698.
54 (*a*) P. I. Kitov and D. Bundle, *J. Am. Chem. Soc.*, 2003, **125**, 16271–16284; (*b*) L. L. Kiessling, J. E. Gestwicki and L. E. Strong, *Angew. Chem., Int. Ed.*, 2006, **45**, 2348–2368; (*c*) M. Mammen, S. K. Choi and G. M. Whitesides, *Angew. Chem., Int. Ed.*, 1998, **37**, 2754–2794.
55 G. Jayamurugan and N. Jayaraman, *Adv. Synth. Catal.*, 2009, **351**, 2379–2390.
56 A. M. Caminade, A. Ouali, M. Keller and J. P. Majoral, *Chem. Soc. Rev.*, 2012, **41**, 4113–4125.
57 B. Rasmussen and J. B. Christensen, *Org. Biomol. Chem.*, 2012, **10**, 4821–4835.
58 B. Helms, C. O. Liang, C. J. Hawker and J. M. J. Fréchet, *Macromolecules*, 2005, **38**, 5411–5415.
59 (*a*) H. Wang, Y. Koshi, D. Minato, H. Nonaka, S. Kiyonaka, Y. Mori, S. Tsukiji and I. Hamachi, *J. Am. Chem. Soc.*, 2011, **133**, 12220–12228; (*b*) T. Hayashi, Y. Sun, T. Tamura, K. Kuwata, Z. Song, Y. Takaoka and I. Hamachi, *J. Am. Chem. Soc.*, 2013, **135**, 12252–12258; (*c*) Y. Koshi, E. Nakata, M. Miyagawa, S. Tsukiji, T. Ogawa and I. Hamachi, *J. Am. Chem. Soc.*, 2008, **130**, 245–251.
60 F. E. Hah and S. Rupprecht, *Chem. Ber.*, 1991, **124**, 481–486.
61 W. Storck and G. Manecke, *J. Mol. Catal.*, 1985, **30**, 145–169.

 This journal is © The Royal Society of Chemistry 2024

62 B. Fischer, S. Xu and H. Zipse, *Chem. – Eur. J.*, 2006, **12**, 5779–5784.
63 A. Sakakura, K. Kawajiri, T. Ohkubo, Y. Kosugi and K. Ishihara, *J. Am. Chem. Soc.*, 2007, **129**, 14775–14779.
64 X.-A. Lu, C.-H. Chou, C.-C. Wang and S.-C. Hung, *Synlett*, 2003, 1364–1366.
65 B. Ren, L. Gan, L. Zhang, N. Yana and H. Dong, *Org. Biomol. Chem.*, 2018, **16**, 5591–5597.
66 W. Muramatsu and T. Kawabata, *Tetrahedron Lett.*, 2007, **48**, 5031–5033.
67 G. Hu and A. Vasella, *Helv. Chim. Acta*, 2002, **85**, 4369–4391.
68 R. Fallek, N. Ashush, A. Fallek and M. Portnoy, *Synlett*, 2021, **32**, 1849–1854.
69 R. S. Bagul and N. Jayaraman, *Inorg. Chim. Acta*, 2014, **409**, 34–52.
70 E. V. Evtushenko, *J. Carbohydr. Chem.*, 2010, **29**, 369–378.
71 N. Dang, V. R. N. Munasinghe and W. G. Overend, *J. Chem. Soc., Perkin Trans. 1*, 1983, 257–264.
72 V. Cachatra, A. Almeida, J. Sardinha, S. D. Lucas, A. Gomes, P. D. Vaz, M. H. Florencio, R. Nunes, D. Vila-Vicosa, M. J. Calhorda and A. P. Rauter, *Org. Lett.*, 2015, **17**, 5622–5625.
73 C. G. Casinovi, M. Framondino, G. Randazzo and F. Siani, *Carbohydr. Res.*, 1974, **36**, 67–73.
74 D. A. MacManus and E. N. Vulfson, *Carbohydr. Res.*, 1995, **279**, 281–291.

Published on 24 2024. Downloaded on 16/08/2025 12:08:06 .