Organic & Biomolecular Chemistry



PAPER View Article Online
View Journal | View Issue



Cite this: *Org. Biomol. Chem.*, 2017, **15**, 7572

Intramolecular hydrogen bonding in conformationally semi-rigid α -acylmethane derivatives: a theoretical NMR study†

Antonio J. Mota, (10 ** Jürgen Neuhold, b Martina Drescher, c Sébastien Lemouzy, c Leticia González (10 ** and Nuno Maulide (10 ** c

Received 25th July 2017, Accepted 18th August 2017 DOI: 10.1039/c7ob01834g rsc.li/obc Conformational mobility is a core property of organic compounds, and conformational analysis has become a pervasive tool for synthetic design. In this work, we present experimental and computational (employing Density Functional Theory) evidence for unusual intramolecular hydrogen bonding interactions in a series of α -acylmethane derivatives, as well as a discussion of the consequences thereof for their NMR spectroscopic properties.

Introduction

Conformational flexibility is an intrinsic trait of most organic compounds.1 The remarkable ability of molecules to adopt multiple possible conformations is usually accompanied by a defined preference for certain of those conformations, a property that resides at the core of molecular interactions and molecular recognition.2 Although the distribution of conformational states can be strongly influenced by solvent effects,³ intramolecular attractive or repulsive interactions are often decisive factors that lead molecules to "choose" a specific spatial arrangement of atoms.4 Much, if not all, of the chemical machinery that sustains life as we know it hinges on subtle interactions of small thermodynamic value but enormous structural importance. Such interactions are deployed in multidirectional fashion and can on occasion result in robust, macroscopic 3D molecular scaffolds. Often, unforeseen attractive interactions can become important design elements with applications ranging from synthesis⁵ to catalysis⁶ and, ultimately, biology.7

N-Acyloxazolidinones, popularized through the seminal, textbook work of Evans and others⁸ remain, even in the 21st century, as cornerstone reagents for aldol- and related transformations. This is largely due, as in most cases of successful asymmetric induction (be it stoichiometric or catalytic), to

Herein, we present experimental and computational evidence for an unusual intramolecular hydrogen bonding interaction in a series of N-acyloxazolidinones and other α -acylmethane derivatives, with striking consequences to their NMR spectroscopic properties. The study also includes theoretical (density functional theory, DFT) 1 H- and 13 C-NMR determinations. Theoretical NMR determinations are often used in cases of problematic and dubious NMR assignments 9 and should be more frequently used as a tool to enable correct NMR analyses.

Results and discussion

During our recent studies on redox-neutral reactions of N-alkynyloxazolidinones (ynamides), 10 leading to the preparation of various α -arylated acyloxazolidinone products (1, Scheme 1), we noted the consistently and unusually low field 1 H-NMR shifts of the hydrogen marked in blue as Ha. For instance, in product 1-nBu (R = nButyl) the signal for Ha appeared at $\delta = 5.67$ ppm, at 5.68 ppm for 1-Cy

Scheme 1 Redox-neutral arylation of ynamides and unusual chemical shifts of Ha.

their adoption of predictable conformations in solution as governed (at a given temperature) by either chelation or dipole attraction/repulsion effects.

^aDepartment of Inorganic Chemistry, Faculty of Sciences, Avda. Fuentenueva s/n, 18002 Granada, Spain. E-mail: mota@ugr.es

^bUniversity of Vienna, Institute of Theoretical Chemistry, Währinger Strasse 17, 1090 Vienna, Austria. E-mail: leticia.gonzalez@univie.ac.at

^cUniversity of Vienna, Institute of Organic Chemistry, Währinger Strasse 38, 1090 Vienna Austria F-mail: nuno maulide@univie ac at

 $[\]dagger \, \text{Electronic}$ supplementary information (ESI) available. See DOI: 10.1039/c7ob01834g

(R = Cyclohexyl), and for **1-Ph** (R = Phenyl), the signal goes to 6.96 ppm, lying practically within the aromatic region.

In order to eliminate any possible effect of the sulfur residue on the aromatic ring, we compared the known compound **1-Dec** (R = Decyl) with its desulfurated analogue **2-Dec**. A decrease in chemical shift of 0.64 ppm (from 5.66 to 5.02 ppm, respectively) was found, whereas an equally high chemical shift value of $\delta = 5.11$ ppm has been reported¹¹ for the desulfurated methyl derivative **2-Me** (*cf.* Fig. 1a).

This far exceeds, for instance, the predicted value for this structure obtained by simple NMR-predicting software (the ChemOffice 13.0 suite, 12 for example, predicts a chemical shift, δ , of just 3.52 ppm). The value is all the more striking if one compares the acyloxazolidinone **2-Me** with other carbonyl analogues (*cf.* Fig. 1b). Indeed, the aldehyde **3-Me** (which could be expected to exert a comparable electron-withdrawing effect on the C–H bonds in α -position with respect to an acyloxazolidinone moiety), the ethylester **4-Me**, the methylthioester **5-Me** or the *N*-methylcarboxamide **6-Me** are all

Fig. 1 (a) Evaluation of the influence of an arylsulfur residue on the chemical shift of Ha, and on its desulfurated version, for oxazolidinone compounds. (b) Comparative analysis of analogous desulfurated carbonyl derivatives. (c) The different experimentally known α -acylmethane derivatives studied in this work, with the exception of **7-Me** that has been prepared by us for the first time.

known compounds reported to have chemical shifts for Ha inside a narrow window not exceeding a δ value of 3.92 ppm (vide infra Table 1).

Detailed analysis of the structure of acyloxazolidinones 1-R/2-R revealed the possible intervention of an intramolecular hydrogen bonding interaction through a 6-membered ring ranging from the electron-rich oxazolidinone (Oxaz) carbonyl oxygen to the methinic hydrogen Ha, giving rise to the conformer shown in Fig. 2. We recognised that this structure-specific interaction might lie at the heart of the unusually high chemical shift observed for this particular compound (which is absent in the other derivatives, *cf.* Fig. 1b).

To validate this assumption, we required a tool to accurately model the two extreme conformational situations where this hydrogen bond is present and absent, compare their relative stability and simulate the NMR spectrum of both forms. We resorted to high-level DFT calculations since it is well-known that general purpose, theoretical ¹H-NMR spectra could be reliably covered by this level of theory. ¹³ This required accuracy aims not only at reliably reproducing the already known experimental data but also predict new values for unknown derivatives or non-detectable conformers. The latter is a crucial requirement as we need to compare experimentally assessed H-bonded situations with non-measurable conformers for which no H-bonding is operative and experimental quantification by NMR is not accessible.

We will initially focus on desulfurated α-acylmethane derivatives, *i.e.*, the known compounds **2-Me–6-Me** (*cf.* Fig. 1c). We further added an unknown compound, the *N*-methylimidazolidinone (Imid) 7-**Me**, to enlarge the predictability test of our model, and resynthesized **5-Me** to ascertain the ¹H-NMR chemical shift for Ha, since calculations found a discordant value with that reported in literature (see below in ref. 24).



Fig. 2 Possibility of six-membered intramolecular hydrogen bonding in 2-Me.

Table 1 Experimental and calculated ¹H-NMR chemical shifts (in ppm) of Ha for derivatives 2-Me-7-Me in chloroform

¹ H-NMR chemical shifts	2-Me (R1 = Oxaz)	3-Me $(R^1 = H)$	4-Me $(R^1 = OEt)$	$5-\mathbf{Me}\left(\mathbf{R}^{1}=\mathbf{SMe}\right)$	6-Me (R1 = NHMe)	7-Me ($R^1 = Imid$)
B3LYP	5.23	3.69	3.68	3.88	3.26	5.43
B3LYP-D3	5.02	3.70	3.68	3.91	3.33	5.17
CAM-B3LYP	5.16	3.62	3.63	3.85	3.21	5.36
M06-2X	5.12	3.79	3.82	4.22	3.48	5.20
LC-ωPBE Experimental	5.10 5.11 (ref. 11)	3.64 3.64 (ref. 23)	3.64 3.72 (ref. 24)	3.92 3.92 ^a (ref. 25)	3.30 3.55 (ref. 26)	5.29 5.28^a

^a This work (see ESI).

derivatives

Computational details

Calculations were performed using the GAUSSIAN09 suite of programs.14 Initial lowest-energy conformations were optimized by density functional theory (DFT) using the well-known hybrid B3LYP functional¹⁵ with the Pople's diffuse, polarized, split-valence, double-zeta 6-31+G* basis set. 16 From these geometries, the corresponding ¹H-NMR isotropic magnetic shielding values were calculated reoptimizing them with the larger triple-zeta 6-311+G(d,p) basis set¹⁷ (necessary to get a high accuracy in the determination of chemical shifts, 13 see the ESI,† for geometries) combined with different functionals. These include the hybrids B3LYP, which gives reliable ¹H-NMR chemical shifts for the most common compounds, their empirical dispersion, B3LYP-D3,18 and long-range corrected, CAM-B3LYP, 19 versions, and the Minnesota M06-2X functional,20 which accounts for non-covalent interactions. We have also included for comparison the non-hybrid GGA-functional LC-ωPBE, ^{21a} the long range corrected version of PBE^{21b} (another widely used functional for general purposes). All the calculations have been carried out including implicit solvent (in which the corresponding experimental spectra have been recorded, see below) through a Polarizable Continuum Model (PCM).²² Results obtained from these calculations for the chemical shift (in ppm) of the Ha hydrogen of the selected, lowest-energy conformers for compounds 2-Me-7-Me (Fig. 1c) are summarized in Table 1.

It should be noted that the precision in the ¹H-NMR chemical shift determination is fairly high along the different methods employed. Yet, for this set of molecules, the LC-ωPBE functional performs extremely well, achieving δ values in very close agreement to the experimental ones (see the ESI† for details), therefore we elected the LC-ωPBE functional for further analyses.

Interestingly, the calculated lowest-energy conformations for 2-Me and 7-Me correspond to that depicted in Fig. 2, in which a 6-membered hydrogen bond²⁷ is established between Ha and the carbonyl group of the heterocyclic moiety. We had originally postulated that this was at the origin of the unusual chemical shift values for Ha. With the help of DFT calculations, we are able to consider conformations in which the hydrogen bonding event is absent and recalculate the chemical shift associated to Ha in the same solvent (chloroform). Results for the two limiting conformations in each case (hydrogen bonded, 2-Me and 7-Me, and non-hydrogen bonded, 2b-Me and 7b-Me) are presented in Table 2. An important drop (about 1.5 ppm to high field) is observed affecting the δ value of Ha of the conformer in which the hydrogen bonding is not operating with respect to the one in which the hydrogen bond is established (lowest-energy conformer).

Since the free-energy difference (ΔG) between the two limiting conformations (B3LYP/6-31+G(d)) is 4.46 kcal mol⁻¹ for 2-Me and 5.44 kcal mol⁻¹ for 7-Me, a conformational equilibrium of conformers 2b-Me and 7b-Me (without hydrogen bonding) can be excluded. This means that, besides other possible, energetically accessible conformations, the apparent

Table 2 Calculated ¹H-NMR chemical shift (in ppm) of Ha in conformers including and excluding hydrogen bonding for derivatives 2-Me and 7-Me, in chloroform, at the LC-ωPBE/6-311+G(d,p) level of theory

¹ H-NMR	2-Me	2b-Me	7-Me hydrogen bonding	7 b-Me
chemical	hydrogen	non-hydrogen		non-hydrogen
shifts	bonding	bonding		bonding
LC-ωPBE	5.10	3.64	5.29 5.28^{b}	3.53
Experimental	5.11 ¹¹	a		a

^a Non applicable. ^b This work (see ESI).

chemical shifts of Ha for 2-Me and 7-Me should be very close to the theoretical calculated values, as it is evidenced in Table 2.

At this juncture, we wished to dive into simulated spectra, with the aim to modulate the strength of the intramolecular H-bond by the calculated value for Ha. Accordingly, we elected several ad hoc unreported derivatives: the permethylated 8-Me and the perfluorinated 9-Me oxazolidinone analogues, and the acylcarbamate 10-Me, which constitutes the ring-strainless open-chain version of the oxazolidinone derivative 2-Me (Fig. 3). Calculations made on these compounds were performed for the conformations for which the 6-membered, intramolecular hydrogen bond is expected to be operative and gave rise to the set of δ values for Ha collected in Table 3 along with the calculated O···Ha distances.

From Table 3, it could be pointed out that, concerning the deshielding of Ha, the ureido derivative 7-Me and the open acylcarbamate 10-Me appear to be the more effective compounds establishing the intramolecular hydrogen bond, given the enhanced Lewis basicity (and hence stronger hydrogen bond-donor ability) of the carbonyl oxygen interacting with Ha. Contrarily to this, the fluorinated 9-Me derivative leads to a less electron-rich carbonyl, weakening the hydrogen bond and observing the corresponding δ value of Ha at higher field (by about 0.6 ppm, see Table 3). Interestingly, the incorporation of four methyl groups in 8-Me did not substantially affect the chemical shift of Ha with respect to the pure oxazolidinone moiety (2-Me). On the other hand, the hydrogen bonds (if available) expected to be stronger when presenting a higher δ value for Ha, also present, in general, shorter distances (Table 3).

Additionally, the LC-ωPBE chemical shifts correlate fairly well with the O···Ha equilibrium distances in compounds 2-Me and 7-Me-10-Me (see Fig. 4), with some small variations for shorter distances.²⁸

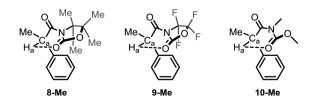


Fig. 3 Ad hoc structures created for the study of the 6-membered, intramolecular hydrogen bond in this series of compounds.

Table 3 Calculated ¹H-NMR chemical shifts of Ha for known structures 2-Me and 7-Me and unknown 8-Me-10-Me and O··· Ha distances

Parameters	2-Me (R1 = Oxaz)	$7\text{-Me}\left(R^1 = Imid\right)$	$8-\mathbf{Me} \left(\mathbf{R}^1 = \mathrm{Oxaz}(\mathbf{Me}_4) \right)$	$\mathbf{9\text{-}Me}\left(R^1 = \mathrm{Oxaz}(F_4)\right)$	10-Me (R^1 = open Oxaz)
δ (in ppm) Distance (in Å) ^a	5.10	5.29	5.21	4.75	5.31
Distance (in $Å$) ^{a}	2.243	2.198	2.201	2.295	2.173

^a O···Ha distances were extracted from the corresponding ¹H-NMR calculations (LC-ωPBE/6-311+G(d,p) in chloroform (PCM)).

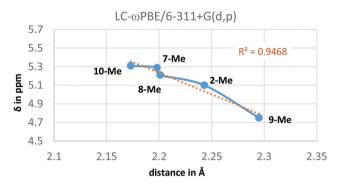


Fig. 4 Calculated ¹H-NMR chemical shifts vs. O···Ha distance for derivatives 2-Me and 7-Me-10-Me.

A comparison of these distances with the sum of the corresponding van der Waals radii for hydrogen and oxygen atoms, that is, 2.70 Å, 29 constitutes another observation pointing to the presence of intramolecular hydrogen bonding because its short distance, 30 as the oxazolidinone carbonyl group establishes another weak C=O···H-Ph interaction in **2-Me** at 2.777 Å, a distance slightly higher than the sum of the corresponding van der Waals radii. Interestingly, the remaining carbonyl group establishes three different close interactions: at 2.589 (C=O···CH₂), 2.644 (C=O···Me) and 2.792 Å (C=O···H-Ph) (Fig. 5), all them around the expected van der Waals distance.

In order to characterize this special interaction and evaluate its strength, we applied the quantum theory of atoms in molecules (QTAIM)³¹ over derivatives **2-Me** and **7-Me-10-Me** (from the corresponding LC- ω PBE/6-311+G(d,p) calculations) using the Multiwfn suite.³² This theory is a topological analysis able to identify bonding interactions within a molecule by means of the gradient vector field of the charge density, $\rho(\mathbf{r})$. Typical $\rho(\mathbf{r})$ values in shared interactions are 0.722, 0.551, and 0.252 a.u. for N₂, O₂, and C-C bond in ethane molecules, respectively, whereas in closed-shell interactions they are 0.046 and 0.036 a.u.

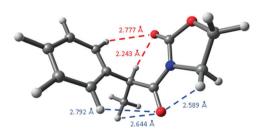


Fig. 5 Close interactions in the two carbonyl groups present in 2-Me.

for LiCl molecules and NaCl molecules, respectively. 31a,33 These bonding interactions can be classified in terms of the properties of the Laplacian of the electron density, $\nabla^2 \rho(\mathbf{r})$, into two broad general classes: shared ($\nabla^2 \rho(\mathbf{r}) < 0$, *i.e.* covalent bonds) and closed-shell ($\nabla^2 \rho(\mathbf{r}) > 0$, *i.e.* hydrogen bonds) interactions. 34 Concerning our study, the well-known criteria of the hydrogen bonding on the basis of AIM theory at the bond critical point (BCP), where the gradient vector field, $\nabla \rho(\mathbf{r})$, vanishes, are: (i) $\rho(\mathbf{r})$ between 0.002 and 0.034 a.u., and (ii) $\nabla^2 \rho(\mathbf{r})$ between +0.024 and +0.139 a.u. 35 Mata *et al.* 36 correlated the hydrogen-bonding energy, $E_{\rm HB}$, with the Lagrangian kinetic energy, $G(\mathbf{r})$, at the BCP as $E_{\rm HB} = 0.429 \times G(\mathbf{r})$.

Therefore, we calculated the BCPs and searched for those with a positive value of $\nabla^2 \rho(\mathbf{r})$, finding a BCP in all cases in between the C=O···Ha path (2-Me, as an example, in Fig. 6). Laplacians of the electron density, $\nabla^2 \rho(\mathbf{r})$, charge densities, $\rho(\mathbf{r})$, and Lagrangian kinetic energies, $G(\mathbf{r})$, at each C=O···Ha BCP, Ha chemical shifts, C=O···Ha equilibrium distances, and calculated $E_{\rm HB}$ energies are summarized in Table 4.

The data in Table 4 corroborates the presence of hydrogen bonds in the C=O···Ha paths for oxazolidinone- (2-Me and 8-Me-9-Me), imidazolidinone-based (7-Me) and the open-chain 10-Me compounds. The strongest hydrogen bond can be found in the latter structure 10-Me, with a rather short O···H distance, whereas the weakest one, as expected, appeared in the fluorinated derivative 9-Me.

Based on both the energy ($E_{\rm HB}$) and the O···Ha distances, these hydrogen bonds are positioned in the limit in between medium and weak and, hence, they present a bond contribution mostly electrostatic. ^{27b,37} As a reference, the calculated CCSD(T) interaction energy for a water dimer, a HF dimer, or a

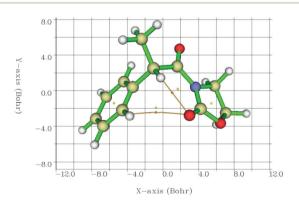


Fig. 6 Interatomic bond critical points (orange circles) identified by QTAIM and the corresponding paths between the concerned atoms. Circles in yellow correspond to ring critical points.

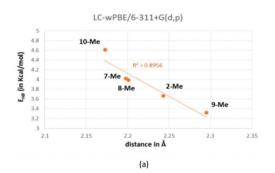
Table 4 Laplacians of the electron density, $\nabla^2 \rho(\mathbf{r})$, charge densities, $\rho(\mathbf{r})$, and Lagrangian kinetic energies, $G(\mathbf{r})$, at each C=O···Ha BCP, H_a chemical shifts, O···Ha equilibrium distances, and calculated $E_{\rm HB}$ energies for derivatives 2-Me and 7-Me-10-Me

Parameters	2-Me (R1 = Oxaz)	$7-\mathbf{Me}\left(\mathbf{R}^{1}=\mathbf{Imid}\right)$	8-Me $(R^1 = Oxaz(Me_4))$	$\mathbf{9-Me}\;\big(R^1 = \mathrm{Oxaz}\big(F_4\big)\big)$	10-Me (R^1 = open Oxaz)
$\nabla^2 \rho(\mathbf{r})$ (in a.u.)	0.06209	0.06791	0.06790	0.05658	0.07821
$\rho(\mathbf{r})$ (in a.u.)	0.01745	0.01899	0.01878	0.01586	0.02069
$G(\mathbf{r})$ (in a.u.)	0.01359	0.01493	0.01484	0.01234	0.01712
δ (in ppm)	5.10	5.29	5.21	4.75	5.31
distance (in Å)	2.243	2.198	2.201	2.295	2.173
$E_{\rm HB}$ (in kcal mol ⁻¹)	$3.66 (4.46)^a$	$4.02 (5.44)^a$	3.99	3.32	$4.61(5.14)^a$

^a In parenthesis, the B3LYP/6-31+G(d) calculated energy difference between the limiting conformations (those following an intramolecular C=0... Ha hydrogen bonding scheme and those that does not). Note that most part of this energy difference corresponds to the hydrogen

HCl dimer is 4.92, 4.52 and 1.90 kcal mol⁻¹, respectively.³⁸ Although the hydrogen-bond energy (E_{HB}) could be well correlated with the O···Ha distance (Fig. 7a), the best correlating descriptor with E_{HB} is the corresponding charge density $(\rho(\mathbf{r}))$ at the BCP,³⁹ as it is showed in Fig. 7b.

Solvent is usually an important parameter in NMR theoretical calculations and must be considered in order to accurately reproduce experimental NMR spectra.3,40 In hydrogen bonding, the election of the solvent for spectroscopic properties is not innocent, since high-polar solvents can theoretically disrupt or even break these types of interactions. 41 Thus, expecting that we could achieve different chemical shifts for Ha in polar solvents and have access to other molecular conformations than those stabilized by hydrogen bonding, we experimentally and theoretically (LC-ωPBE/6-311+G(d,p))



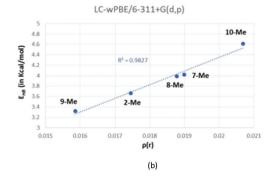


Fig. 7 Correlation between the calculated energy of the hydrogen bond (E_{HB}) and either the O···Ha distance (a) or the charge density (b).

studied the corresponding ¹H-NMR spectra for 7-Me (R¹ = Imid) in deuterated methanol and dimethylsulfoxide (Table 5).

Nevertheless, the conclusion that is drawn from Table 5 is that no solvent effect is observed in the chemical shift of Ha, even in the presence of methanol, a polar protic solvent. Therefore, since in Table 2 was clearly shown that the nonhydrogen bonded form of 7-Me (7b-Me) presented a chemical shift of 3.53 ppm, we must conclude that, even with polar solvents, the conformational equilibrium at room temperature keeps this specially stabilized conformation the most part of the time, avoiding the expected shielding of the chemical shift of Ha.

Traditionally, organic chemists rely mostly on ¹H-NMR spectra, with relatively little attention being paid to ¹³C-NMR analysis. However, 13C-NMR constitutes a crucial axis to determining and ascertaining structures mainly due to the fact that it presents a much larger window allowing the appreciation of even small variations and rendering the collapsing of two different signals unlikely. In addition, H-H coupling often leading to broad/multiplet bands and often complicating assignment is absent in 13C-NMR. Therefore, 13C-NMR provides, from a theoretical point of view, 13d,42 an ideal ground where many organic products could be unambiguously characterized by a unique fingerprint. We thus set out to complete our findings and reinterpret the problem at hand based on 13 C-NMR data for the Ca carbon (the methinic carbon to which Ha is bonded), see Fig. 1 and 3.

Table 6 compiles the chemical shift values for Ca for compounds 2-Me and 2b-Me, 3-Me-6Me, and 7-Me and 7b-Me, using the five functionals initially considered (Table 1). Nevertheless, the basis set used was Pople's polarized splitvalence double-zeta 6-31G(d,p), 16 since it is known that Pople's double-zeta basis sets perform better than the triple-zeta ones

Table 5 Theoretical and experimental solvent effects on the ¹H-NMR chemical shift of Ha for compound 7-Me

¹ H-NMR chemical shifts	CDCl_3	MeOD	DMSO- d_6
Theoretical ^a	5.29	5.31	5.30
Experimental ^b	5.28	5.26	5.24

 $[^]a$ LC-ωPBE/6-311+G(d,p). b This work (see ESI).

Table 6 Experimental and calculated values of the ¹³C-NMR chemical shifts (in ppm) of Ca for derivatives **2-Me-7-Me** (along with **2b-Me** and **7b-Me**) in chloroform

¹³ C-NMR chemical shifts	B3LYP	B3LYP-D3	CAM-B3LYP	M06-2X	LC-ωPBE	Experimental
2-Me	45.86	46.16	44.33	48.59	43.08	42.6 (ref. 11 <i>a</i>) 42.8 ^a (ref. 11 <i>b</i>)
2b-Me	51.18	51.17	49.38	53.20	47.55	_b
3-Ме	57.22	56.91	55.23	58.89	52.94	53.2 (ref. 23 <i>a</i>) 53.11 (ref. 23 <i>b</i> and <i>c</i>)
4-Me	49.15	48.82	47.39	50.03	45.46	45.59 (ref. 24 <i>a</i>) 45.5 (ref. 24 <i>b</i>)
5-Me	58.00	57.75	55.75	60.32	53.46	54.23^{c}
6-Me	51.61	50.73	49.72	51.68	47.23	47.2 (ref. 26 <i>a</i>)
7-Me	45.45	45.67	43.91	47.83	42.61	42.69^{c}
7b-Me	50.63	50.71	48.91	52.47	47.14	_b

^a Carbon non-assigned in the experimental spectrum. ^b Non-applicable. ^c This work (see ESI).

for 13 C-NMR calculations, probably due to some cancellation errors. 43

Table 6 also reflects a traditional problem associated with the theoretical determination of 13 C-NMR chemical shifts, namely that the obtained values are very sensitive to the functional used. This variability is perhaps the main reason why theoretical calculations on 13 C-NMR spectra are less common. In this case, although CAM-B3LYP and LC- ω PBE are again the more reliable functionals, LC- ω PBE clearly outperforms, delivering chemical shifts very close to the experimental values.

Furthermore, it is important to highlight the difference of *ca.* 11 ppm between the chemical shifts calculated for Ca in the imidazolidinone (7-**Me**) and the methylthio (5-**Me**) derivatives. This showcases the large spectral window made possible by ¹³C-NMR.

Calculations for the *ad hoc* structures **8-Me-10-Me** led to the results collected in Table 7. Owing to the fact that 13 C-NMR is quite sensitive to steric factors (about an order of magnitude more than 1 H-NMR) 44 the reported δ values calculated for Ca do not correlate with any property treated in this text. This is in the line presented just above, for which 13 C-NMR determinations are better oriented to the unambiguous assignment of spectroscopic data and, hence, the accurate prediction of spectra for unknown products.

Finally, and as expected given the results achieved for ¹H-NMR, solvent effects on compound **7-Me** were almost negligible, as it could be drawn from Table 8, indicating that the hydrogen-bonded conformer should be largely favoured within the timescale of the experiment.

Table 7 Calculated 13 C-NMR chemical shifts (in ppm) of Ca for known structures 2-Me and 7-Me and unknown 8-Me-10-Me a

¹³ C-NMR chemical shifts	2-Me	7-Me	8-Me	9-Me	10-Me
δ (in ppm)	43.08	42.61	44.63	45.57	45.44

^a Structures referred here were calculated at the LC-ωPBE/6-31G(d,p) level of theory and using chloroform as solvent (PCM).

Table 8 Theoretical and experimental solvent effects on the ¹³C-NMR chemical shift (in ppm) of Ca for compound **7-Me**

¹³ C-NMR chemical shifts	$CDCl_3$	MeOD	DMSO- d_6
- h	42.61	43.58	43.60
	42.69	43.78	42.13

 $[^]a$ LC-ωPBE/6-31G(d,p). b This work (see ESI).

Conclusions

In summary, we have presented experimental and computational evidence for an unusual intramolecular hydrogen bonding interaction in a series of N-acyloxazolidinones and other α-acylmethane derivatives, with striking consequences to their NMR spectroscopic properties. In the course of this study, theoretical (DFT) 1H- and 13C-NMR determinations, along with QTAIM analysis, were employed to clearly identify the nature of such special interaction, among others found in these structures. Of special interest is the finding of the reliability and accuracy of the LC-ωPBE functional, which works fairly well in this domain, at least for this kind of derivatives. Crucial has been the possibility to calculate conformers not accessible by synthesis and that emphasize the stronger interaction involved in this series of compounds. Therefore, results presented here hint at possible general applications to the prediction of highly accurate NMR spectral properties for organic compounds.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This research was supported by the ERC (FLATOUT 278872), and the University of Vienna. Calculations were partially performed at the Vienna Scientific Cluster (VSC). We also would like to thank the Centro de Supercomputación de la Universidad de Granada (Alhambra, CSIRC) for computation resources.

Notes and references

- 1 (a) E. L. Eliel, N. L. Allinger, S. J. Angyal and G. A. Morrison, Conformational Analysis, Interscience Publishers, New York, 1965; (b) D. Barton, Nobel Lecture 1967, Nobel Lectures, Chemistry 1963-1970, Elsevier Publishing Company, Amsterdam, 1972; (c) R. Hubert, Nature, 1979, 280, 538; (d) F. G. Ridell, The conformational analysis of heterocyclic compounds, Academic Press, London, 1980; (e) G. W. Gokel and A. Nakano, Feeble Forces and Flexible Frameworks, in Crown Compounds: Toward Future Applications, ed. S. Cooper, Verlag Chemie, 1992, ch. 1; (f) C. Gilon, C. Mang, E. Lohof, A. Friedler and H. Kessler, Synthesis of Cyclic Pepetides, in Houben-Weyl Methods of Organic Chemistry Vol. E 22b, 4th Edition Supplement: Synthesis of Peptides and Peptidomimetics, ed. A. Felix, L. Moroder and C. Toniolo, Georg Thieme Verlag, Stuttgart, 2004; (g) M. C. Misuraca, T. Grecu, Z. Freixa, V. Garavini, C. A. Hunter, P. W. N. M. van Leeuwen, M. D. Segarra-Maset and S. M. Turega, J. Org. Chem., 2011, 76, 2723; (h) Structure Elucidation in Organic Chemistry: The Search for the Right Tools, ed. M.-M. Cid and J. Bravo, John Wiley & Sons, Weinheim, 2015; (i) V. M. Burger, D. J. Arenas and C. M. Stultz, Sci. Rep., 2016, 6, 29040; (j) B. Szyszko, M. J. Białek, E. Pacholska-Dudziak and L. Latos-Grażyński, Chem. Rev., 2017, 117, 2839.
- 2 (a) D. J. Chadwick and K. Widdows, Host-Guest Molecular Interactions: From Chemistry to Biology, Ciba Foundation Symposium, John Wiley & Sons, Chichester, 1991; (b) H.-J. Böhm and G. Klebe, Angew. Chem., Int. Ed. Engl., 1996, 35, 2588; (c) R. A. Laskowski, N. M. Luscombe, M. B. Swindells and J. M. Thornton, Protein Sci., 1996, 5, 2438; (d) K. K. Frederick, M. S. Marlow, K. G. Valentine and A. J. Wand, Nature, 2007, 448, 325; (e) P. Hobza and J. Řezáč, *Chem. Rev.*, 2016, **116**, 4911; (f) D. Chatterji, *Basics* of Molecular Recognition, CRC Press, Boca Raton, 2016.
- 3 (a) K.-J. Liu and J. L. Parsons, Macromolecules, 1969, 2, 529; Reviews in Computational Chemistry II, K. B. Lipkowitz and D. B. Boyd, John Wiley & Sons, New York, 1991; (c) Carbohydrates: Structures, Syntheses and Dynamics, ed. P. Finch, Springer Science+Business Media, Dordrecht, 1999; (d) Ch. Cappelli, S. Corni and J. Tomasi, J. Phys. Chem. A, 2001, 105, 10807; (e) Handbook of solvents, ed. G. Wypych, ChemTec Publishing, Toronto-New York, 2001; (f) Zh. Xu, Hs. Tsai, Hs.-L. Wang and M. Cotlet, J. Phys. Chem. B, 2010, 114, 11746; (g) P. I. Nagy, J. Phys. Chem. A, 2012, 116, 7726; (h) Ch. Wang, F. Ying, W. Wu and Y. Mo, J. Org. Chem., 2014, 79, 1571; (i) I. V. Alabugin, Stereoelectronic Effects: A Bridge Between Structure and Reactivity, John Wiley & Sons, Chichester, 2016.
- 4 (a) M. Oki, The Chemistry of Rotational Isomers, Springer-Verlag, Berlin-Heidelberg, 1993; (b) J. E. Anderson, V. Bru-Capdeville, P. A. Kirsch and J. S. Lomas, J. Chem. Soc.,

- Chem. Commun., 1994, 1077; (c) Y. Cheng, Y. Qi, Y. Tang, Ch. Zheng, Y. Wan, W. Huang and R. Chen, J. Phys. Chem. Lett., 2016, 7, 3609.
- 5 (a) C. Altavilla and E. Ciliberto, *Inorganic Nanoparticles:* Synthesis, Applications and Perspectives, CRC Press, Boca Raton, 2011; (b) Ch. F. Chen and Y.-X. Ma, Yptycenes Chemistry: From Synthesis to Applications, Springer-Verlag, Berlin-Heidelberg, 2013.
- 6 (a) Food Proteins: Properties and Characterization, ed. Sh. Nakai and H. W. Modler, Wiley-VCH, New York, 1996; (b) M. N. Khan, Micellar catalysis, CRC Press, Boca Raton, 2007
- 7 (a) D. Leckband and J. Israelachvili, Q. Rev. Biophys., 2001, 34, 105; (b) J. McMurry, Organic Chemistry with Biological Applications, Brooks/Cole CENGAGE Learning, 2010; (c) M. J. R. Yunta, Using Molecular Modelling to Study Interactions Between Molecules with Biological Activity, in Bioinformatics, ed. H. Pérez-Sánchez, CC by 3.0 license, 2012, ch. 8.
- Rhodium-Catalyzed 8 Modern Organic Reactions. ed. P. A. Evans, Wiley-VCH, Weinheim, 2005.
- 9 (a) S. D. Rychnovsky, Org. Lett., 2006, 8, 2895; (b) F. Sánchez-Izquierdo, P. Blanco, F. Busqué, R. Alibés, P. de March, M. Figueredo, J. Font and T. Parella, Org. Lett., 2007, 9, 1769.
- 10 (a) B. Peng, X. Huang, L.-G. Xie and N. Maulide, Angew. Chem., Int. Ed., 2014, 53, 8718; (b) L.-G. Xie, S. Niyomchon, A. J. Mota, L. González and N. Maulide, Nat. Commun., 2016, 7, 10914; (c) D. Kaldre, B. Maryasin, D. Kaiser, O. Gajsek, L. González and N. Maulide, Angew. Chem., Int. Ed., 2017, 56, 2212.
- 11 (a) E. Coulbeck and J. Eames, Tetrahedron: Asymmetry, 2007, 18, 2313; (b) A. Bigot, A. Williamson and M. J. Gaunt, J. Am. Chem. Soc., 2011, 133, 13778.
- 12 ChemOffice suite v. 13.0. CambridgeSoft, PerkinElmer Inc., 2012.
- 13 (a) G. Schreckenbach and T. Ziegler, Theor. Chem. Acc., 1998, **99**, 71; (b) E. Y. Pankratyev, A. R. Tulyabaev and L. M. Khalilov, J. Comput. Chem., 2011, 32, 1993; (c) I. A. Konstantinov and L. J. Broadbelt, J. Phys. Chem. A, 2011, **115**, 12364; (d) M. Elyashberg, A. Williams and K. Blinov, Contemporary Computer-Assisted Approaches to Molecular Structure Elucidation, Royal Society of Chemistry, 2011; (e) D. Fleig, M. Maurer, M. Hanni, K. Braunger, L. Kick, M. Thubauville and Ch. Ochsenfeld, J. Chem. Theory Comput., 2014, 10, 572; (f) D. E. Hill, N. Vasdev and J. P. Holland, Comput. Theor. Chem., 2015, 1051, 161.
- 14 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand,

- K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, N. J. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, Ö. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, *Gaussian 09, Revision B.01*, Gaussian, Inc., Wallingford, CT, 2010.
- 15 (a) A. D. Becke, *Phys. Rev. A*, 1988, 38, 3098; (b) C. Lee,
 W. Yang and R. G. Parr, *Phys. Rev. B: Condens. Matter*, 1988,
 37, 785; (c) A. D. Becke, *J. Chem. Phys.*, 1993, 98, 5648.
- 16 (a) M. M. Francl, W. J. Pietro, W. J. Hehre, J. S. Binkley,
 D. J. DeFrees, J. A. Pople and M. S. Gordon, *J. Chem. Phys.*,
 1982, 77, 3654; (b) V. A. Rassolov, M. A. Ratner, J. A. Pople,
 P. C. Redfern and L. A. Curtiss, *J. Comput. Chem.*, 2001, 22,
 976.
- (a) K. Raghavachari, J. S. Binkley, R. Seeger and J. A. Pople,
 J. Chem. Phys., 1980, 72, 650; (b) D. McLean and
 G. S. Chandler, J. Chem. Phys., 1980, 72, 5639.
- 18 S. Grimme, J. Comput. Chem., 2004, 25, 1463.
- (a) Y. Tawada, T. Tsuneda, S. Yanagisawa, T. Yanai and K. Hirao, *J. Chem. Phys.*, 2004, **120**, 8425; (b) T. Yanai, D. P. Tew and N. C. Handy, *Chem. Phys. Lett.*, 2004, **393**, 51.
- 20 Y. Zhao and D. G. Truhlar, Theor. Chem. Acc., 2008, 120, 215.
- 21 (a) O. A. Vydrov and G. E. Scuseria, J. Chem. Phys., 2006, 125, 234109; (b) J. P. Perdew, K. Burke and M. Ernzerhof, Phys. Rev. Lett., 1996, 77, 3865.
- 22 (a) S. Miertus, E. Scrocco and J. Tomasi, J. Chem. Phys., 1981, 55, 117; (b) R. Cammi and J. Tomasi, J. Comput. Chem., 1995, 16, 1449; (c) J. Tomasi and M. Persico, Chem. Rev., 1994, 94, 2027.
- 23 (a) R. Srirambalaji, S. Hong, R. Natarajan, M. Yoon, R. Hota, Y. Kim, Y. H. Ko and K. Kim, *Chem. Commun.*, 2012, 48, 11650; (b) A. T. Straub, M. Otto, I. Usui and B. Breit, *Adv. Synth. Catal.*, 2013, 355, 2071; (c) S. H. Christensen, E. P. K. Olsen, J. Rosenbaum and R. Madsen, *Org. Biomol. Chem.*, 2015, 13, 938.
- 24 (a) D. Katayev, V. Matousek, R. Koller and A. Togni, *Org. Lett.*, 2015, 17, 5898; (b) P. H. Gehrtz, V. Hirschbeck and I. Fleischer, *Chem. Commun.*, 2015, 51, 12574.
- 25 For spectral data for 5-Me in CCl₄, see: (a) D. Seebach and R. Bürstinghaus, *Synthesis*, 1975, 461. For a prior misassigned spectrum of 5-Me in deuterated chloroform, see: (b) M. Clericuzio, I. Degani, S. Dughera and R. Fochi, *Synthesis*, 2002, 921. See the ESI† for ¹H- and ¹³C-NMR spectra of 5-Me, in chloroform, newly synthesized for this work.
- 26 (a) Sh. Hanada, T. Ishida, Y. Motoyama and H. Nagashima, J. Org. Chem., 2007, 72, 7551; (b) J. L. Barneto, M. Avalos, R. Babiano, P. Cintas, J. L. Jiménez and J. C. Palacios, Org. Biomol. Chem., 2010, 8, 857.
- 27 (a) J. Catalán, J. C. del Valle, J. Palomar, C. Díaz and J. L. G. de Paz, J. Phys. Chem. A, 1999, 103, 10921;
 (b) G. R. Desiraju and T. Steiner, The weak hydrogen bond in

- structural chemistry and biology, Oxford University Press, Oxford, 1999; (c) J. Chen, P. G. Willis, S. Parkin and Cammers, Eur. J. Org. Chem., 2005, S. J. Grabowski, Challenges and advances in (d) Computational Chemistry and Physics, 3: Hydrogen Bonding -New Insights, Springer, Dordrecht, 2006; (e) Y.-Y. Zhu, G.-T. Wang, R.-X. Wang and Zh.-T. Li, Cryst. Growth Des., 2009, 9, 4778; (f) P. S. Kalsi, Organic Reactions and their Mechanisms, New Age International, 2009; (g) L. Quiquempoix, E. Bogdan, N. J. Wells, J.-Y. Le Questel, J. Graton and B. Linclau, Molecules, 2017, 22, 518.
- 28 It should be noted that ¹H chemical shift values could be influenced by different factors others than pure inductive effect of the surrounding groups or the chosen solvent, *i.e.* anisotropy, van der Waals deshielding (steric factors), other hydrogen bonding-like or electrostatic interactions. See: L. D. S. Yadav, *Organic Spectroscopy*, Springer Science+Business Media, Dordrecht, 2005.
- 29 S. Alvarez, Dalton Trans., 2013, 42, 8617.
- 30 P. A. Kollman and L. C. Allen, Chem. Rev., 1972, 72, 283.
- 31 (a) R. F. W. Bader, Atoms in Molecules A quantum theory, Oxford University Press, New York, 1990; (b) P. S. V. Kuma, V. Raghavendra and V. Subramanian, J. Chem. Sci., 2016, 128, 1527.
- 32 T. Lu and F. Chen, J. Comput. Chem., 2012, 33, 580.
- 33 Y. Aray, J. Rodríguez and D. Vega, *J. Phys. Chem. B*, 2000, **104**, 4608.
- 34 R. F. W. Bader and H. Essen, J. Chem. Phys., 1984, 80, 1943.
- 35 (a) U. Koch and P. L. A. Popelier, J. Phys. Chem., 1995, 99, 9747; (b) D. Mani and E. Arunan, Noncovalent Forces, Challenges and Advances in Computational Chemistry and Physics Series, ed. J. Leszcznski, Springer Int'l Pub., Switzerland, 2015, vol. 19; (c) J. H. Lee, J.-H. Lee, E.-H. Kong and H. M. Jang, Sci. Rep., 2016, 6, 21687.
- 36 I. Mata, I. Alkorta, E. Espinosa and E. Molins, *Chem. Phys. Lett.*, 2011, **507**, 185.
- 37 A. Nangia, CrystEngComm, 2002, 4, 93.
- 38 K. Wendler, J. Thar, S. Zahn and B. Kirchner, *J. Phys. Chem. A*, 2010, **114**, 9529.
- 39 A. T. Ayoub, J. Tuszynski and M. Klobukowski, *Theor. Chem. Acc.*, 2014, **133**, 1520.
- 40 M. Dračínský and P. Bouř, J. Chem. Theory Comput., 2010, 6, 288.
- 41 *Hydrogen Bonding and Transfer in the Excited State*, ed. K.-L. Han and G.-J. Zhao, John Wiley & Sons, Chichester, 2011.
- 42 (a) Calculation of NMR and EPR Parameters. Theory and Applications, ed. M. Kaupp, M. Bühl and V. G. Malkin, Wiley-VCH, Weinheim, 2004; (b) C. Timmons and P. Wipf, J. Org. Chem., 2008, 73, 9168.
- 43 D. J. Giesen and N. Zumbulyadis, *Phys. Chem. Chem. Phys.*, 2002, 4, 5498.
- 44 G. W. Buchanan, J. B. Stothers and G. Wood, *Can. J. Chem.*, 1973, **51**, 3746.