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C-Glycosylation enabled by N-(glycosyloxy) acetamides†

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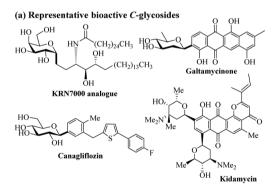
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The C-glycosylation of C-nucleophiles including allyltrimethylsilane, silyl enol ethers and phenols with N-(glycosyloxy)acetamides as glycosyl donors has been realized. This protocol provides a convenient and practical route for the synthesis of alkyl C-glycosides and aryl 2-deoxy- β -C-glycosides under mild reaction conditions.

C-Glycosides are a class of important compounds with a carbohydrate unit linked to another carbohydrate unit or an aglycone through a carbon-carbon bond. Compared with the corresponding O-glycosides and N-glycosides, C-glycosides show better enzyme stability and hydrolysis resistance.² On the other hand, C-glycosides are present in a diverse range of biologically active natural products and drugs such as a KRN7000 analogue,³ Canagliflozin,⁴ Kidamycin,⁵ and Galtamycinone⁶ (Scheme 1a). Therefore, the synthesis of C-glycosides has been one of the significant and critical topics in organic chemistry. 2d,7 Among the numerous methods for the construction of the C-C glycosidic bond, the coupling reaction of glycosyl donors with C-nucleophiles in the presence of an activator is a straightforward and highly efficient route. For instance, glycosyl halides, glycals, glycosyl acetates, thioglycosides, thioglycosides, sulfoxides, 12 1,2-anhydro sugars, 13 and glycosyl phosphates 14 have been widely employed for C-glycosylation. Although a lot of elegant strategies and methods in this field have been available over the last few decades, 15 the development of efficient and stereoselective protocols for the synthesis of C-glycosides remains challenging. New approaches for the synthesis of diverse C-glycosides are highly desirable.

Recently, we developed a new O-glycosylation method by the use of N-(glycosyloxy)acetamides as glycosyl donors and a Lewis acid $[Cu(OTf)_2]$ or $SnCl_4$ as a promoter (Scheme 1b). The easily accessible and stable donors as well as mild reaction conditions sparked our interest in exploring other applications of this method. As part of our continuing effects on the synthetic methodologies toward C-glycosyl compounds, Therein we report the C-glycosylation of C-nucleophiles with N-(glycosyloxy)acetamides (Scheme 1c).

We commenced our study by investigating the C-glycosylation of allyltrimethylsilane (2a) with perbenzyl N-(glucosyloxy)acetamide (1a). Initially, following the optimized O-glycosylation conditions, the reaction was conducted in CH_2Cl_2 with $Cu(OTf)_2$ as the activator. Unfortunately, the desired allyl C-glycoside 3a was obtained in only 10% yield (Table 1, entry 1). Subsequently, various solvents, including



(b) N-(Glycosyloxy)acetamides-based O-glycosylation

(c) This work: N-(Glycosyloxy)acetamides-based C-glycosylation

Scheme 1 Representative bioactive C-glycosides and N-(glycosyloxy) acetamide-based glycosylations.

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 $(Butyl)_2BOTf(2.0)$

 $ZrCl_4$ (2.0)

SnCl₄ (2.0)

 $SnBr_{4}(2.0)$

SnBr₄ (1.5)

SnBr₄ (1.2)

SnBr₄ (1.0)

 $Sn(OTf)_2$ (2.0)

Table 1 Optimization of the reaction conditions^a

BnO-BnO-	OBn OBnONHAc +	TMS Promoter, Solvent Additive, T, 4Å MS	BnO BnO 3	OBn OBn
Entry	Promoter (equiv.)	Solvent	T (°C)	Yield ^b (%)
1	Cu(OTf) ₂ (4.0)	CH ₂ Cl ₂	30	10
2	$Cu(OTf)_2(4.0)$	CH_3CN	30	0
3	$Cu(OTf)_2$ (4.0)	Ether	30	10
4	$Cu(OTf)_2$ (4.0)	$CHCl_3$	30	35
5	$Cu(OTf)_2$ (4.0)	CH_3NO_2	30	40
6	$Cu(OTf)_2$ (4.0)	CHCl ₃	50	45
7	$Cu(OTf)_2$ (4.0)	$CHCl_3/CH_3NO_2 = 1/1$	50	75
8	$Cu(OTf)_2$ (4.0)	$CHCl_3/CH_3NO_2 = 2/1$	50	37
9	$Cu(OTf)_2$ (4.0)	$CHCl_3/CH_3NO_2 = 1/2$	50	50
10	$BF_3 \cdot Et_2O(2.0)$	$CHCl_3/CH_3NO_2 = 1/1$	rt	78
11	BCl_3 (2.0)	$CHCl_3/CH_3NO_2 = 1/1$	Rt	Trace
12	$BBr_3(2.0)$	$CHCl_3/CH_3NO_2 = 1/1$	rt	Trace
13	$\mathrm{HBF_4 \cdot Et_2O} \ (2.0)$	$CHCl_3/CH_3NO_2 = 1/1$	rt	72

 a Reaction conditions: 1a (30.0 mg, 0.05 mmol), 2a (15.9 μ L 0.1 mmol), 4 Å MS (100 mg) and solvent (1.0 mL). b Isolated yield.

 $CHCl_3/CH_3NO_2 = 1/1$

 $CHCl_3/CH_3NO_2 = 1/1$

 $\mathrm{CHCl_3}/\mathrm{CH_3NO_2} = 1/1$

 $CHCl_3/CH_3NO_2 = 1/1$

55

61

82

93

95

91

87

rt

rt

rt

Trace

CH₃CN, ether, CHCl₃, and CH₃NO₂, were tested and the yield of 3a seemed to improve on changing the solvent (Table 1, entries 3-5; Table S1†). In CHCl3 and CH3NO2 at 30 °C, 35% yield and 40% yield, respectively, were achieved (Table 1, entries 4-5). On increasing the reaction temperature to 50 °C, the yield of the coupled product 3a improved to 45% in CHCl₃ (Table 1, entry 6). When a mixed solvent of CHCl3 and CH₃NO₂ was used, the yield of 3a increased to 75% (Table 1, entry 7). A ratio of 1:1 was found to be the optimal solvent ratio of CHCl₃ to CH₃NO₂ in terms of the yield (Table 1, entries 7-9). The additives did not increase the yield of 3a (Table S2†). Further screening of the Lewis acids revealed that BF₃·Et₂O, HBF₄·Et₂O, SnCl₄, and SnBr₄ can be employed as efficient activators (Table 1, entries 10-18). SnBr₄ gave the best yield (Table 1, entry 18, 93% yield). It was found that decreasing the amount of SnBr4 to 1.0 equiv. or 1.2 equiv. leads to reduced yield of the target product (Table 1, entries 20-21). The optimal loading of SnBr₄ is 1.5 equiv. (Table 1, entry 19). It is worth noting that the reaction can be completed within 0.5 h. The α/β ratio of product 3a is about 5/1. Therefore, the optimal conditions were identified to be as follows: SnBr₄ (1.5 equiv.) as the activator and CHCl₃/CH₃NO₂ (v/v = 1/1) as the solvent in the presence of 4 Å molecular sieves at room temperature for 0.5 h.

With the optimized conditions in hand, we first examined the substrate scope of alkyl *C*-nucleophiles such as allyltrimethylsilane and silyl enol ethers (Table 2). When donor **1a** and (2,2-dimethyl-1-methylenepropoxy)trimethylsilane (**2b**)

Table 2 C-Glycosylation of allyltrimethylsilane or silyl enol ethers (2a–2c) with N-(glycosyloxy)acetamides (1a–1d) a

PGC	ONHAc +		rt, 4Å MS / CH ₃ NO ₂ PGO 3b-3h
Entry	Donor	C-Nucleophile	Product (yield, α/β ratio) ^{b,c}
1	BnO O O O O O O O O O O O O O O O O O O	TMSO	BnO OBn OBn 3b, 83%, α/β= 3: 1
2	BnO ONHAc	Zb TMS Za	BnO O O O O O O O O O O O O O O O O O O
3	1b	2b	BnO O O O O O O O O O O O O O O O O O O
4	1b	TMSO 2c	BnO O O O O O O O O O O O O O O O O O O
5	BnO OBn BnO ONHAc	2a	BnO OBn BnO 3 f , 97%, α only
6	1c	2b	BnO OBn BnO O 3g, 95%, α only
7	OBn O-WONHAC BnO	2a	BnO OBn BnO 3h, 92%, α/β= 1: 1

 a Conditions: Donor (0.05 mmol), $\it C$ -nucleophile (0.10 mmol), $\rm SnBr_4$ (0.075 mmol), 4 Å MS (100 mg), $\rm CHCl_3$ (0.5 mL) and $\rm CH_3NO_2$ (0.5 mL), 0 °C to rt. b Isolated yield. c Ratio determined using 1H NMR spectra.

were treated under the standard conditions, the desired product 3b was obtained in 83% yield with a moderate α-stereoselectivity (entry 1). The glycosylation of allyltrimethylsilane (2a) with 2-deoxy-glucopyranosyl donor 1b furnished the corresponding C-glycoside 3c in excellent yield and with good stereoselectivity (entry 2). Silyl enol ethers 2b and 2c also underwent this reaction to give the products 3d and 3e in good yields (entries 3 and 4). Both 2-deoxy-galactopyranosyl donor 1c and arabinofuranosyl donor 1d were amenable for this C-glycosylation to deliver the alkyl C-glycosides in excellent yields (entries 5-7). The exclusive α -stereoselectivity was observed with compound 1c as the glycosyl donor for the formation of C-glycosides (entries 5-6),18 whereas no stereoselectivity was observed in the case of arabinofuranosyl donor 1d (entry 7). Unfortunately, the protocol could not be applied to the disarmed donors. All reactions were finished within 30 min at room temperature. The anomeric configuration of all alkyl C-glycosides was unambiguously identified by their ¹H and ¹³C NMR analyses as described in the literature. ¹⁹

Next, the glycosylation of phenols with donor **1a** was performed to examine whether *C*-glycosides would be produced

Scheme 2 The coupling reactions of 1a and phenols 4a-4c. Reaction conditions: 1a (0.05 mmol), phenol (0.10 mmol), SnBr₄ (0.075 mmol), 4 Å MS (100 mg), CHCl₃ (0.5 mL) and CH₃NO₂ (0.5 mL), 0 °C to rt.

via an O-C rearrangement²⁰ (Scheme 2). The coupling reactions of donor 1a with 3-methoxyphenol (4a) and 4-methoxyphenol (4b) were tried, and only the corresponding O-glycosides (5a and 5b) were isolated as the major products under the standard conditions. The glycosylation of more electron-rich 3,4-dimethoxyphenol (4c) gave the same results. Interestingly, phenolic glycoside 5b was isolated as a single α -isomer. The addition of a co-promoter such as Sc(OTf)₃, ²¹ TMSOTf, 22 and BF3:Et2O23 was unable to facilitate the O-C rearrangement, either at the beginning of the reaction or after the generation of O-glycosides. We speculate that the fully oxygenated pyranosyl donors might be unsuitable for the construction of aryl C-glycosides.

Subsequently, we turn our attention to 2-deoxy-N-(glycosyloxy)acetamide donors (Table 3). Gratifyingly, the reaction of 2-deoxy-N-(glucosyloxy)acetamide 1b and 4a provided the corresponding C-glycoside 5d in 71% yield and with full β-stereoselectivity (entry 1). Phenols including 3,5-dimethoxyphenol (4d), 3,4,5-trimethoxyphenol (4e), 1-naphthol (4f), 2-naphthol (4g), and 7-methoxy-2-naphthol (4h) were well tolerated, affording the desired C-glycosides 5e-5i in good to excellent yields (entries 2-6). When 2-deoxy-N-(galactosyloxy) acetamide 1c was used as the glycosyl donor, the reaction also proceeded well, providing the corresponding aryl C-glycosides 5j-5m in high yields and with excellent β-selectivity (entries 7-10). Surprisingly, the reaction of fully oxygenated arabinofuranosyl donor 1d with phenol 4d smoothly afforded 5n in 70% yield in the form of a single β -configuration (entry 11). Complete β-selectivity was observed regardless of the nature of the phenols and the donors employed, which might be attributed to the formation of thermodynamically more stable β-C-glycosides at room temperature via the O-C rearrangement.²⁴

In summary, we have developed a facile and mild C-glycosylation of C-nucleophiles with N-(glycosyloxy)acetamides as donors. This reaction proceeds by the use of SnBr₄ as the activator in the mixed solvent of CHCl₃-CH₃NO₂ at room temperature for 0.5 h. A variety of alkyl C-glycosides and aryl C-glycosides were obtained in good to excellent yields. In par-

Table 3 β -Stereoselective C-glycosylation of phenols with N-(glycosyloxy)acetamides^a

ОН

PGO	ONHAc +	$R = \frac{\text{SnBr}_4, \text{ rt, 4Å}}{\text{CHCl}_3/\text{ CH}_3\text{N}}$	MS PGO PGO
	1b-1d 4a,	4d-4j	5d-5n
Entry	Donor	Phenol	Product (yield, α/β ratio) ^b
1	BnO OBn ONHAc	HO————————————————————————————————————	BnO HO OMe BnO 5d, 71%, β only
2	1b	HO OMe OMe 4d	BnO HO OMe BnO Se, 80%, β only OMe
3	1b	HO OMe OMe OMe	BnO HO OMe BnO OMe 5f, 82%, β only OMe
4	1b	OH 4f	BnO O OH Sg, 72%, β only OH
5	1b	OH 4g	BnO HO BnO BnO 5h, 90%, β only
6	1b	HO 4h OMe	BnO HO BnO Si, 91%, β only OMe
7	BnO OBn BnO 1c ONHAc	OH OBn	BnO OBn BnO 5j, 78%, β only OH
8	1c	4e	BnO OMe OMe OMe St, 81%, β only HO
9	1c	4g	BnO OBn BnO OH 51, 85%, β only OH
10	1c	MeO OH	OMe BnO OBn BnO OH 5m, 88%, β only OH
11	BnO OBn ONHAc	4d	OMe HO OMe BnO BnO

^a Conditions: Donor (0.05 mmol), phenol (0.10 mmol), SnBr₄ (0.075 mmol), 4 Å MS (100 mg), CHCl₃ (0.5 mL) and CH₃NO₂ (0.5 mL), 0 °C to rt. b Isolated yield.

ticular, the reaction of phenols with 2-deoxy-N-(glycosyloxy) acetamides gave an exclusive β-stereoselectivity via an O-C rearrangement. The disclosed method provides an alternative approach for the synthesis of C-glycosides with biological significance.

70%, β only

Conflicts of interest

There are no conflicts to declare.

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