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Chloride anion transporters inhibit growth of methicillin-resistant *Staphylococcus aureus* (MRSA) *in vitro*†

Andrew I. Share,^a Khushali Patel,^a Cristina Nativi,^b Eun J. Cho,^c Oscar Francesconi,^{*b} Nathalie Busschaert,^{‡d} Philip A. Gale,^{*d} Stefano Roelens^{*b} and Jonathan L. Sessler^{*a}

A series of aminopyrrolic receptors were tested as anion transporters using POPC liposome model membranes. Many were found to be effective Cl⁻ transporters and to inhibit clinical strains of *Staphylococcus aureus* growth *in vitro*. The best transporters proved effective against the methicillin-resistant *Staphylococcus aureus* (MRSA) strains, Mu50 and HP1173. Tris-thiourea tren-based chloride transporters were also shown to inhibit the growth of *S. aureus in vitro*.

There is a tremendous interest in novel antibiotics that can combat highly resistant bacterial strains such as MRSA.¹ Recent studies of compounds that can mediate the transport of anions demonstrated significant biological activity. Several have shown antiproliferative activity in cancer cell lines.^{2,3} For instance, the strapped calixpyrrole (**1**) displays modest NaCl transport in liposome models and is able to inhibit growth of cancer cell lines *in vitro*.⁴ This effect was ascribed to a combination of receptor mediated chloride transport and sodium transport involving endogenous ion channels. This and other recent developments in the anion transport field^{5,6} have led us to consider that other biologically active species may be mediating their effect in whole or in part by an ability to transport chloride anions⁷ into cells. §¶

As a first test of this hypothesis, two known antibiotic agents, thiocarlidate and trichlorocarbanalidate were tested for their ability to transport chloride anions through a standard 1-palmitoyl-2-oleoyl-*sn*-glycero-3-phosphocholine (POPC) liposomal model membrane. They were found to be modest NaCl (or HCl||) co-transporters and

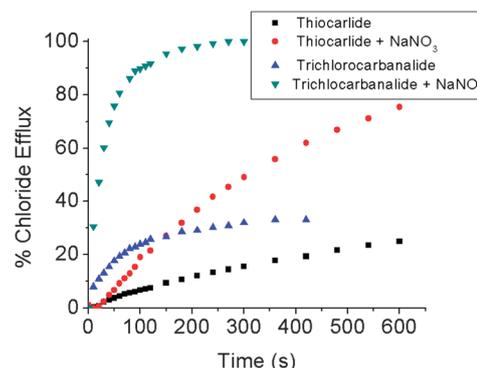
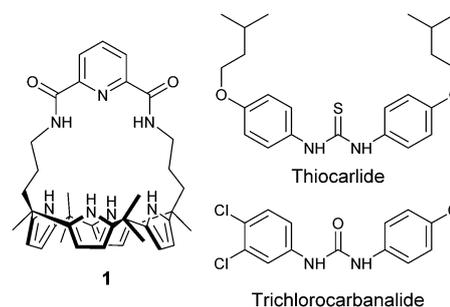


Fig. 1 Chloride efflux caused by the addition of the antibiotics thiocarlidate or trichlorocarbanalidate (0.675 mol%), with and without NaNO₃ added to the exterior of POPC vesicles loaded with 0.5 M NaCl, 0.1 M Na₂SO₄, and 20 mM phosphate buffer, pH = 7.2. 100% chloride efflux was determined by lysing the liposomes with Triton X.

effective Cl⁻/NO₃⁻ antiporters (Fig. 1). This finding, as well as the ability of several cation ionophores to act as antibiotics,⁸ has led us to consider that anion transporters might show antibiotic activity.



With this view in mind, we selected a family of aminopyrrolic compounds, namely **2–15** (Scheme 1), and investigated their chloride anion transport properties. The antibiotic activity of these compounds, as well as the known Cl⁻/HCO₃⁻ anion antiport agents **16–18** (Scheme 2), were also tested. The present study

^a Department of Chemistry, The University of Texas, Austin, TX 78712-1224, USA.
E-mail: sessler@cm.utexas.edu

^b Department of Chemistry and INSTM, University of Florence, Polo Scientifico e Tecnologico, 50019 Sesto Fiorentino, Firenze, Italy.
E-mail: oscar.francesconi@unifi.it, stefano.roelens@unifi.it

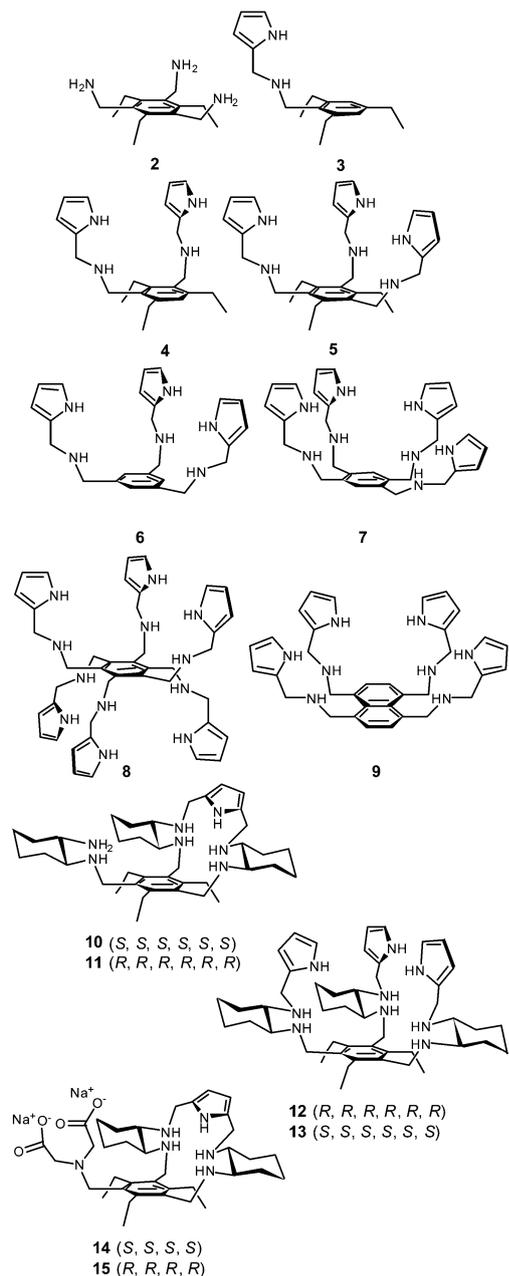
^c Targeted Therapeutic Drug Discovery and Development (TTDDDP), College of Pharmacy, University of Texas-Austin, Austin TX 78723, USA

^d Chemistry, University of Southampton, Southampton, SO17 1BJ, UK.
E-mail: philip.gale@soton.ac.uk

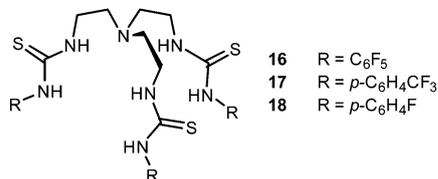
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‡ Present address: Silver Center for Arts and Science, New York University, Department of Chemistry, 100 Washington Square E, New York, NY 10003, USA.





Scheme 1 Structures of compounds 2–15.



Scheme 2 Structures of compounds 16–18.

was prompted by previous work, showing that several aminopyrrolic compounds were biologically active vs. pathogenic yeasts,¹⁰ exhibited antiviral activity,¹¹ induced apoptosis in HeLa and PLC/PRF/5 cells,¹² and were found to recognize the chloride anion well

in polar organic solvents.^{13,14} Here, we report the finding that for the aminopyrrolic compounds 2–15 chloride anion transport in liposomal model correlates well with *in vitro* antibiotic activity against the *S. aureus* strains UAMS1, HP1173, and Mu50.

Compounds 2–15 were examined for their ability to transport chloride anions across a phospholipid bilayer. In a typical experiment, 160 nm POPC vesicles that contained NaCl (500 mM), Na₂SO₄ (100 mM), and HEPES buffer (5 mM) were suspended in a solution of Na₂SO₄ (100 mM) and HEPES buffer (5 mM). The compound subject to study (0.675 mol%, compared to POPC concentration) was added to the solution and the rate of chloride efflux from the vesicles was monitored using a Cl[−] ion selective electrode (ISE). After 9 minutes, the liposomes were lysed with polyethylene glycol *tert*-octylphenyl ether (Triton X-114), and a final reading from the ISE corresponding to 100% chloride release was taken. High levels of transport activity were seen in descending order for compounds 12, 13, 10, 8, and 11 (Fig. 2). In these studies the solutions external to the liposomes contained only SO₄^{2−} and HEPES. These are hydrophilic species¹⁵ that are typically not transported through POPC membranes.¹⁶ Therefore, we infer that under these experimental conditions, anion antiport should not contribute to the efflux of chloride from the liposome. Studies involving compound 5 were also carried out when the external sulphate anion was replaced by either NO₃[−] or HCO₃[−] (as the sodium salts). The nitrate and bicarbonate anions are considerably more lipophilic than the sulphate anion and can facilitate the transport of chloride through the liposomal membrane by promoting Cl[−]/NO₃[−] or Cl[−]/HCO₃[−] antiport processes. In these experiments a higher concentration of NaCl (1 M) inside the liposome was used to give a larger response with the ISE. The rate of chloride efflux is slightly raised on the addition of the more lipophilic anions nitrate and bicarbonate to the external solution (Fig. 3). Nevertheless, this difference only accounts for a small percentage of the total efflux of chloride from the liposomes. We thus conclude that anion antiport only plays a minor role in chloride efflux and that the receptors of the present study are functioning primarily as cation–anion cotransporters. ||

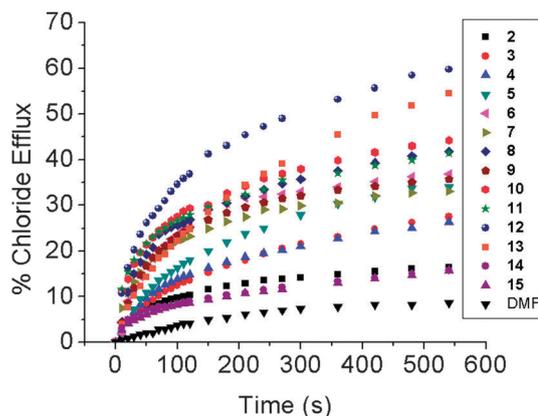


Fig. 2 Chloride efflux was initiated by the addition of 2–15 (0.675 mol%) to a solution of POPC vesicles (1 mM POPC, internal solution: 0.5 M NaCl, 0.1 M Na₂SO₄, 5 mM HEPES, pH = 7.2, external solution: 0.1 M Na₂SO₄, 5 mM HEPES pH = 7.2).



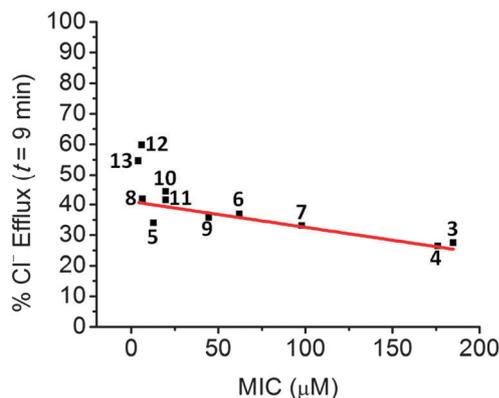


Fig. 5 Plot of chloride efflux promoted by 2–13 in liposomal transport models as detailed in Fig. 2 against their effectiveness in reducing the growth of the Mu50 (resistant) strain of *S. aureus*.

transport that are responsible for the antibacterial activity in *S. aureus* or that there are differences between the properties of the model membranes used in this study and the bacterial membrane.¹⁸ Further study of these and other anion transporters are on-going in an effort to elucidate more fully the underlying mechanisms of action and to develop more active receptor-based antibiotic agents.

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Notes and references

§ Simple cation transporters often function as antibiotics.⁸

¶ During the course of this work, Schmitzer and coworkers reported that certain imidazolium-based anion transporters likewise displayed strong antibiotic effects.⁹

|| The counter cation accompanying chloride anion egress was not monitored directly. The observation of a cation dependence on the rate when the inner solution was changed from NaCl to KCl so as to provide a less hydrated cation¹⁴ is consistent with a Na⁺/Cl⁻ cotransport mechanism (see Supporting Information). However, the presence of readily

protonated amino functionality in receptors 2–15 makes it difficult to discount an alternative mechanism involving H⁺/Cl⁻ cotransport. Through-membrane proton transport is expected to be most important at early times before a proton gradient is built up across the liposomal membrane. An effort to distinguish between limiting NaCl vs. HCl cotransport mechanisms has not been made in the case of thiocarlide and trichlorocarbanalide.

** Although further study is needed, it is likely that artificial receptors such as those of the present study will prove more effective as both ion carriers and antibiotics in the case of Gram positive bacteria than Gram negative bacteria since Gram positive bacteria have a single cellular membrane, whereas Gram negative bacteria have 2 cell membranes.

- 1 M. E. Stryjewski and G. R. Corey, *Clin. Infect. Dis.*, 2014, **58**, S10–S19.
- 2 N. Busschaert, M. Wenzel, M. E. Light, P. Iglesias-Hernández, R. Pérez-Tomás and P. A. Gale, *J. Am. Chem. Soc.*, 2011, **133**, 14136–14148.
- 3 (a) S. J. Moore, M. Wenzel, M. E. Light, R. Morley, S. J. Bradberry, P. Gómez-Iglesias, V. Soto-Cerrato, R. Pérez-Tomás and P. A. Gale, *Chem. Sci.*, 2012, **3**, 2501–2509; (b) S. J. Moore, C. J. E. Haynes, J. González, J. L. Sutton, S. J. Brooks, M. E. Light, J. Herniman, G. J. Langley, V. S. Cerrato, R. Pérez-Tomás, I. Marques, P. J. Costa, V. Félix and P. A. Gale, *Chem. Sci.*, 2013, **4**, 103–117.
- 4 S.-K. Ko, S. K. Kim, A. Share, V. M. Lynch, J. Park, W. Namkung, W. V. Rossom, N. Busschaert, P. A. Gale, J. L. Sessler and I. Shin, *Nat. Chem.*, 2014, **6**, 885–892.
- 5 N. Busschaert and P. A. Gale, *Angew. Chem., Int. Ed.*, 2013, **52**, 1374–1382.
- 6 A. V. Jentzsch, A. Henning, J. Mareda and S. Matile, *Acc. Chem. Res.*, 2013, **46**, 2791–2800.
- 7 (a) H. Li, H. Valkenier, L. W. Judd, P. R. Brotherhood, S. Hussain, J. A. Cooper, O. Jurček, H. A. Sparkes, D. N. Sheppard and A. P. Davis, *Nat. Chem.*, 2014, **8**, 24–32; (b) J. T. Davis, O. Okunola and R. Quesada, *Chem. Soc. Rev.*, 2010, **39**, 3843–3862; (c) G. W. Gokel and S. Negin, *Acc. Chem. Res.*, 2013, **46**, 2824–2833.
- 8 D. A. Kevin II, D. A. F. Meujo and M. T. Hamann, *Expert Opin. Drug Discovery*, 2009, **4**, 109–146.
- 9 C. R. Elie, G. David and A. R. Schmitzer, *J. Med. Chem.*, 2015, **58**, 2358–2366.
- 10 C. Nativi, O. Francesconi, G. Gabrielli, I. De Simone, B. Turchetti, T. Mello, L. D. C. Mannelli, C. Ghelardini, P. Buzzini and S. Roelens, *Chem. – Eur. J.*, 2012, **18**, 5064–5072.
- 11 O. Francesconi, C. Nativi, G. Gabrielli, I. De Simone, S. Noppen, J. Balzarini, S. Liekens and S. Roelens, *Chem. – Eur. J.*, 2015, **21**, 10089–10093.
- 12 S.-H. Park, Y. P. Choi, J. Park, A. Share, O. Francesconi, C. Nativi, W. Namkung, J. L. Sessler, S. Roelens and I. Shin, *Chem. Sci.*, 2015, **6**, 7284–7292.
- 13 S. Roelens, A. Vacca and C. Venturi, *Chem. – Eur. J.*, 2009, **15**, 2635–2644.
- 14 S. Roelens, A. Vacca, O. Francesconi and C. Venturi, *Chem. – Eur. J.*, 2009, **15**, 8296–8302.
- 15 Y. Marcus, *J. Chem. Soc., Faraday Trans.*, 1991, **87**, 2995–2999.
- 16 N. Busschaert, L. E. Karagiannidis, M. Wenzel, C. J. E. Haynes, N. J. Wells, P. G. Young, D. Makuc, J. Plavec, K. A. Jolliffe and P. A. Gale, *Chem. Sci.*, 2014, **5**, 1118–1127.
- 17 X. Wu, L. W. Judd, E. N. W. Howe, A. M. Withecombe, V. Soto-Cerrato, H. Li, N. Busschaert, H. Valkenier, R. Pérez-Tomás, D. N. Sheppard, Y. Jiang, A. P. Davis and P. A. Gale, *Chem*, 2016, DOI: 10.1016/j.chempr.2016.04.002.
- 18 M. J. Spooner and P. A. Gale, *Chem. Commun.*, 2015, **51**, 4883–4886.

