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Comparing organic and metallo-organic hydrazone molecular cages as potential carriers for doxorubicin delivery

Our work describes a rare comparative study between a metallo-organic cage and a fully organic analogous system, both obtained by hydrazone bond formation self-assembly. The organic cage shows better properties, including stability and affinity towards the anticancer drug doxorubicin. Additionally, the organic cage shows minimal cell toxicity, whilst the doxorubicin-cage complex shows *in vitro* anti-cancer activity. Our results show that the properties of the organic cage are suitable for the future challenges of *in vivo* drug delivery using molecular cages. Cancer cell image created with BioRender.com.

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Comparing organic and metallo-organic hydrazone molecular cages as potential carriers for doxorubicin delivery†

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Molecular cages are three-dimensional supramolecular structures that completely wrap guest molecules by encapsulation. We describe a rare comparative study between a metallo-organic cage and a fully organic analogous system, obtained by hydrazone bond formation self-assembly. Both cages are able to encapsulate the anticancer drug doxorubicin, with the organic cage forming a 1:1 inclusion complex with μM affinity, whereas the metallo-organic host experiences disassembly by interaction with the drug. Stability experiments reveal that the ligands of the metallo-organic cage are displaced in buffer at neutral, acidic, and basic pH, while the organic cage only disassembles under acidic conditions. Notably, the organic cage also shows minimal cell toxicity, even at high doses, whilst the doxorubicin-cage complex shows *in vitro* anti-cancer activity. Collectively, these results show that the attributes of the pure organic molecular cage are suitable for the future challenges of *in vivo* drug delivery using molecular cages.

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Introduction

Supramolecular chemistry has been largely driven by the design and study of host molecules that use non-covalent interactions for efficient guest binding.^{1,2} For over 60 years, the structures of synthetic hosts have changed enormously, evolving from acyclic, macrocyclic, to fully three-dimensional cage structures that completely wrap guest molecules.^{3,4} Despite the challenges of synthesis, the development of efficient self-assembly methods has enabled the synthesis of cages with almost any

possible geometry.^{5–7} Self-assembly allowed preparing molecular cages by the design of ligands with a specific shape to self-assemble into metallo-cages and purely organic cages.^{5,6} For this, it is possible to use either reversible metal–ligand interactions, to give coordination cages, or dynamic covalent chemistry to yield purely organic cages.^{8,9}

Cage synthesis is mainly based on the self-assembly of building blocks with complementary connectivity and geometry under thermodynamic control.¹⁰ Many studies on molecular cages have focused on establishing efficient synthetic protocols, allowing multigram-scale production of cages in some cases, facilitating large-scale synthesis for different applications.^{11,12} Moreover, the careful design of the building blocks allows a precise control of the cavity size, shape, and inward-facing functionalisation for selective and efficient encapsulation.^{13–15} Main uses of molecular cages include catalysis,^{16–19} stabilisation of species,^{20,21} molecular recognition,^{5,6} release of encapsulated guests,^{5,6,22} sensing,²³ separation processes,^{13,24,25} among many others.^{26,27} However, the exploration of molecular cages in biological and biomedical contexts remains nascent.^{5,6,28–32}

Preparing cages for bio-medical applications presents a unique set of challenges, most obviously developing systems that are compatible with biological conditions, such as the prerequisite for water solubility.^{5,6,33} In general, fully organic cages have low solubility in water due to their lack of charge and hydrophobic nature, requiring water solubilizing groups in some instances or the use of organic cosolvents.⁵ In contrast, metallo-cages are typically charged species that can result in

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nicotinaldehyde–Pd complex **2**, then Pd₂L₄ assembly **C1** is obtained. The solubility of **C1** can be controlled using different counteranions; the BARF cage **C1**·BARF, prepared *via* anion metathesis of **C1**·BF₄, is soluble in organic solvents,¹⁶ whereas the nitrate cage **C1**·NO₃, obtained directly from the reaction of **1** and **2**·NO₃, can dissolve in aqueous mixtures containing DMSO. Organic cage **C2** was obtained by the reaction of **1** and calixarene starting material **3**. This cage is soluble in both organic solvents and in aqueous mixtures containing DMSO. For both cages it is required typically 1–5% DMSO in water to achieve μM concentration. All cages were fully characterised by ¹H, ¹³C, DOSY NMR, and HRMS (see ESI†).

Additionally, the formation of the molecular cage **C1**·NO₃ was unambiguously confirmed by X-ray crystallography (Fig. 2, see ESI† for details). The crystal structure of the **C1**·NO₃ cage shows a quasi-spherical geometry, with a Pd–Pd distance of 18.6 Å and a *trans* ether O–O separation of 19.9 Å. In this structure, all the hydrazone NH bonds are pointing into the cavity of the cage, providing potential sites for polar interactions with an encapsulated guest. While we have been unable to grow XRD quality crystals of **C2**, a molecular model (X-ray structure shown in Fig. 2, additional details are available in the ESI†) indicates that the size and shape of the organic cage is similar to **C1**; the distance between the centroids of the two calixarene motifs is 18.8 Å while the *trans* ether O atoms are separated by 19.7 Å.

Host–guest chemistry

The affinity of the cages towards DOXO was assessed by means of titration experiments.⁶¹ The photophysical properties of DOXO, which has a fluorescence emission maximum at 590 nm ($\lambda_{\text{exc}} = 470$ nm), enabled the use of emission change to quantify encapsulation of the guest, which was also confirmed by ¹H NMR studies. Both **C1**·NO₃ and **C2** are soluble in the range 20–100 μM in phosphate buffer (buffer concentration 100 μM) with 1–5% v/v DMSO. As the absorption spectra of the molecular cages and DOXO do not overlap (Fig. S32 and S33†), quantitative fluorescence titrations of DOXO with increasing amounts of molecular cages were performed. The titrations with both **C1**·NO₃ and **C2** show a DOXO fluorescence emission decrease (Fig. S28 and S29†). This is attributed to the presence of electron-poor Pd[(pyridine)₄]²⁺ moieties in cage **C1**·NO₃ and electron-rich resorcinol rings in cage **C2**, that quench the DOXO emission. The quenching can take place by intermolecular photo-induced electron transfer (PET) process as we estimated using the Rehm–Weller equation (see ESI†).^{62,63}

The fluorescence quenching experiments reveal a significant difference between cages **C1**·NO₃ and **C2**. The decrease in fluorescence emission was much more noticeable for **C2** compared to **C1**·NO₃, showing more efficient fluorescence quenching. Fitting the emission intensity *versus* the concentration of added cage to a 1 : 1 binding isotherm⁶⁴ using R and RStudio,^{65,66} we determined an association constant of 3.2×10^6 M⁻¹ for the organic cage **C2** (Fig. 3). For cage **C1**·NO₃, we estimated an association constant in the order of magnitude of 10⁴ M⁻¹ with low reproducibility, likely associated to cage disassembly by complexation of Pd²⁺ by the NH₂ moiety of DOXO as

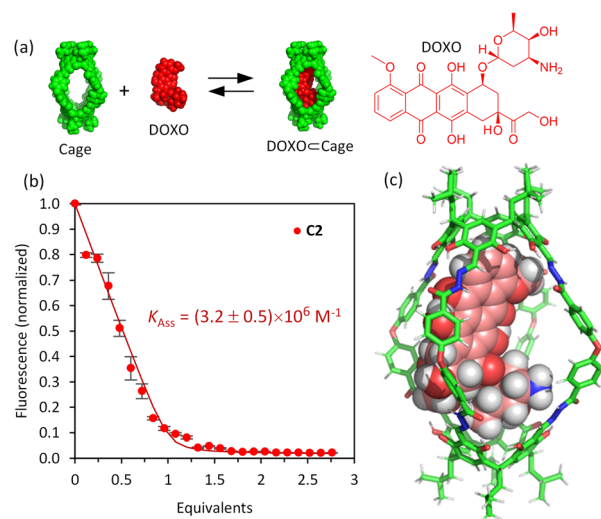


Fig. 3 Fluorescence titration experiments. (a) Schematic representation of drug encapsulation; (b) binding data for the encapsulation of DOXO by cage **C2** obtained from the fluorescence spectra for the addition of **C2** to DOXO (50 μM) in 100 μM phosphate buffer, pH 7.2. The solid points are experimental data, and the continuous lines is the fitted binding isotherm for a 1 : 1 host–guest model, obtained as the average of 4 independent titrations (vertical bars show the standard error for each point). Fluorescence measured at 590 nm with $\lambda_{\text{exc}} = 470$ nm (phosphate buffer 100 μM/DMSO 0.25–2.0%, pH 7.2); (c) MMFF molecular model of the supramolecular complex [DOXO ⊂ **C2**] obtained with the Spartan'20 software.

observed by NMR titrations (Fig. S24 and S28†). The increase of affinity is likely associated with the more hydrophobic nature of uncharged cage **C2** in contrast to the tetracationic **C1**·NO₃.⁶⁷ The inward facing NH groups of **C1**·NO₃, as determined from the X-ray crystal structure, could also lead to an energetically more favourable hydration of the cavity, leading to poorer DOXO binding. The high association constant of DOXO with **C2** permitted a Job's plot analysis to confirm the 1 : 1 stoichiometry of [DOXO ⊂ **C2**] (see ESI Fig. S30†), which is also supported by molecular modelling, suggesting that DOXO fits into the cavity of cage **C2** (Fig. 3c, see ESI† for molecular modelling details).

To check cavity occupancy by the guest, the cavity volume of both cages was determined using the CageCavityCalc Python script. The calculations gave 1350 Å³ for cage **C1** and 1500 Å³ for cage **C2**, showing that both cages have a similar cavity space.⁶⁸ As the size of DOXO is 498 Å³, the single guest occupancy for **C1** and **C2** would be 37% and 33%, respectively, significantly less than the optimal 55% predicted by Rebek for closed-shell organic hosts.⁶⁹ Therefore, the binding observed does not follow the 55%-rule as cages **C1** and **C2** have very large portals, differing from the closed-shell hosts used to develop the 55%-rule.¹⁵

It is also possible to trigger the decomplexation of [DOXO ⊂ **C2**] by adding increasing amounts of DMSO (Fig. 4a). This causes the fluorescence intensity to increase, presumably because the exclusion of DOXO reduces quenching. From this data, we estimated that *ca.* 70% DOXO is released when the solution reaches 55% v/v DMSO (Fig. 4b). The decrease in



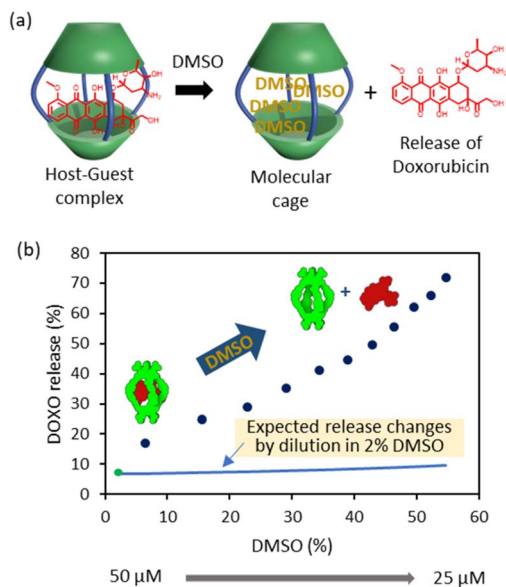


Fig. 4 DOXO release experiments. (a) Schematic representation of DOXO release by competition with DMSO molecules. (b) Plot of recovery of non-bound DOXO versus percentage of added DMSO (6–55%) to the solution containing C2 ($C_{\text{initial}} = 50 \mu\text{M}$, $C_{\text{final}} = 25 \mu\text{M}$) and DOXO ($C_{\text{initial}} = 50 \mu\text{M}$, $C_{\text{final}} = 25 \mu\text{M}$) in phosphate buffer (1 mM, pH 7.2). The green dot indicates the release at 2% DMSO as determined in the binding experiments of Fig. 3. The blue line indicates the expected release changes by dilution from $50 \mu\text{M}$ to $25 \mu\text{M}$ in a solution containing 2% DMSO.

affinity is likely attributed to the weakening of the hydrophobic effect and competition of DMSO molecules with DOXO for the cavity of the cage.

^1H NMR titration experiments have also been undertaken to further corroborate the formation of the host-guest [DOXO \subset cage] complexes. At the cage concentration needed for NMR experiments ($\approx 0.5\text{--}1 \text{ mM}$), it was found that both C1 and C2 are not soluble enough in D_2O to make a meaningful comparison to the fluorescence titrations. Also considering that the use of organic solvents is much less relevant to binding under biological conditions, no attempt was made to quantify encapsulation by NMR. Nonetheless, the changes in individual chemical shifts provide localised information that is not provided from emission data. From these experiments we observed small chemical shift changes consistent with binding (see ESI Section 4 and Fig. S24–S26[†]). Remarkably, it was observed an upfield shift of the inward-facing protons of the cages evidencing the encapsulation of DOXO inside the cavity of both C1 and C2 cages. Similar observations of small chemical shift changes and large fluorescence changes upon anticancer drug encapsulation have been described in similar works.⁵¹

Cage disassembly experiments in mixed aqueous solution

Disassembly experiments of cages C1 and C2 were performed at different pH values (pH 5.6, 7.2, and 7.8) in DMSO/phosphate buffer. This process was initially studied by UV-visible spectroscopy by monitoring the decrease in the absorption band

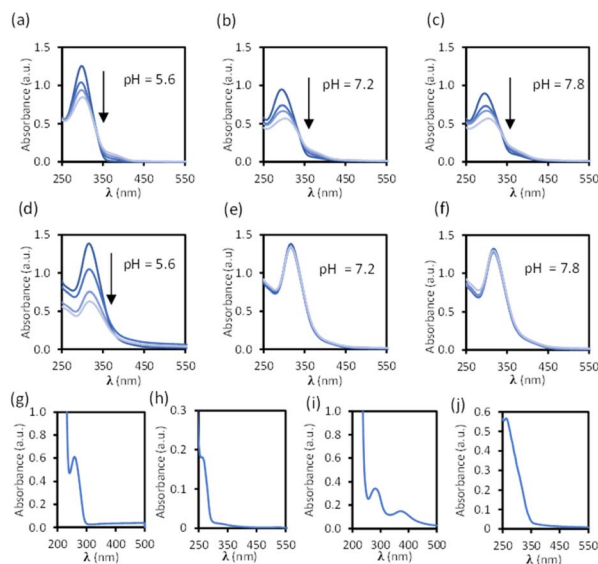


Fig. 5 Stability of cages C1·NO₃ ($10 \mu\text{M}$, (a–c) 0, 1, 2, 4 days) and C2 ($10 \mu\text{M}$, (d–f) 0, 1, 2, and 3 days) at different pH values over time in a buffered solution with phosphate ($100 \mu\text{M}$) and 1% DMSO. Absorbance spectra of building block components (pH 7.1 buffered solution with phosphate ($100 \mu\text{M}$) with 5% DMSO): (g) linker 1 ($25 \mu\text{M}$). (h) Pd(II) complex 2·NO₃ ($50 \mu\text{M}$). (i) Cavitand 3 ($25 \mu\text{M}$). (j) Ligand S2 ($20 \mu\text{M}$).

centred at 300–320 nm characteristic of each cage, and not present in the building block components (see ESI[†]) as a function of time (Fig. 6). The UV-visible spectra of molecular cage C1·NO₃ shows a decrease in absorbance over several days at all of the three different pH values 5.6, 7.2, and 7.8 (Fig. 5a–c). In contrast, the UV-visible spectra of molecular cage C2 remained unchanged over time (0–3 days) under neutral and basic conditions (Fig. 5e and f) but showed a decrease in absorbance at pH 5.6 (Fig. 5d). These results would suggest that C1·NO₃ and C2 disassemble *via* different mechanisms; the metallo-organic cage is likely degraded by coordination of the phosphate buffer anion to the palladium (see ^1H NMR experiments below), resulting in ligand displacement, while the pure organic cage dissociates through acid-catalysed hydrolysis of the hydrazone bonds. The time scale of the hydrolysis is relevant in a biological context as the sustained delivery of drugs over several days is a desirable property in anticancer drug delivery.⁷⁹

Further evidence for a ligand displacement process was obtained by recording the ^1H NMR spectra of C1·NO₃ in DMSO- d_6 before and after the addition of phosphate buffer after *ca.* 5 min (Fig. 6). Addition of buffer results in upfield shifting of the pyridyl moiety protons (H_f , H_i , H_j , H_k), which is in agreement with the loss of the inductive effect associated with coordination to Pd(II). Furthermore, a comparison to the ^1H NMR spectra of free ligand clearly indicates that this species is released intact without hydrolysis of the hydrazone bond. The fast disassembly observed in the NMR experiments (in *ca.* 5 min) is associated to the larger concentration of buffer with respect to cage, as we observed a slower kinetics reducing the buffer concentration (see ESI[†]). Unfortunately, the low solubility of cage C2 at the required concentrations for NMR, as well as the broad features



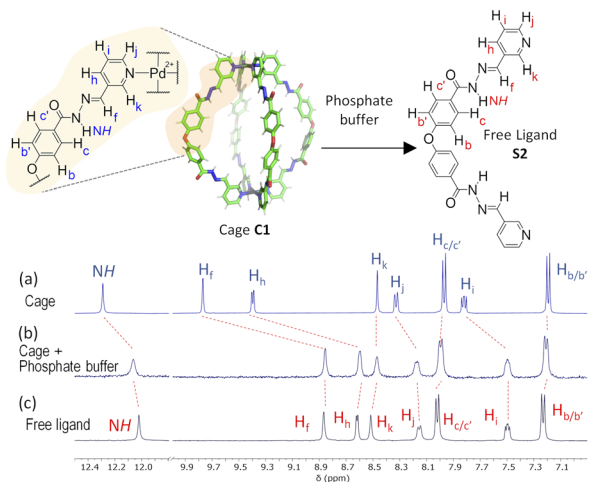


Fig. 6 ^1H NMR disassembly experiment of $\text{C1}\cdot\text{NO}_3$ in $\text{DMSO-}d_6$ (0.5 mM, 550 μL) with phosphate buffer (pH 7.2, 100 mM, 50 μL). (a) ^1H NMR spectra of cage $\text{C1}\cdot\text{NO}_3$. (b) ^1H NMR spectrum after the addition of phosphate buffer (ca. 5 min), causing cage disassembly. (c) ^1H NMR spectra of free ligand (see structure S2 in ESI†).

of these spectra in phosphate buffer/DMSO have hindered a similar study.

Overall, these stability experiments highlight some of the challenges in using metallo-organic cages for bio-medical applications.⁶ They also show that pure organic cages may be better suited to this purpose. Moreover, the selective hydrolysis of the hydrazone bonds in C2 at slightly acidic pH presents an ideal scenario for stimuli responsive drug delivery at the acidic microenvironment of a tumor.

Biological studies

Toxicity studies were first performed to determine the biocompatibility of the empty cages. Tumoral 4T1 (murine triple-negative breast cancer) and human melanoma SK-Mel-103 cell lines were treated with increasing concentrations of $\text{C1}\cdot\text{NO}_3$ and C2 and cell viability was measured at 48 h (Fig. 7). The $\text{C1}\cdot\text{NO}_3$ palladium cage produced toxicity at low concentrations (viability lower than 80% was found at concentration of the cage of 3.12 μM in agreement with previous works on metallo-organic cages),^{40,71} whereas the organic cage C2 was shown to be non-toxic at concentrations as high as 25–50 μM . We also examined that the building blocks components of cage C2 (ligand 1 and calixarene 3) are non-toxic (Fig. S37a in ESI†). Selecting the less toxic organic cage C2 for encapsulation studies, when cancer cells were treated with the $[\text{DOXO}\subset\text{C2}]$ cage, an antitumoral effect was achieved in 4T1 and SK-Mel-103 cells attributed to cage internalisation and cage disassembly in the lysosomes. This is supported by comparing cells treated with the building blocks components of cage C2 (Ligand 1 and calixarene 3) and DOXO (1 + 3 + DOXO) with cells treated with $[\text{DOXO}\subset\text{C2}]$. In both cases, $[\text{DOXO}\subset\text{C2}]$, and 1 + 3 + DOXO, a comparable antitumoral effect is achieved indicating that the efficacy of DOXO remains unaffected by the C2 framework or $[\text{DOXO}\subset\text{C2}]$ formation (Fig. 7c, d and S37b in ESI†).

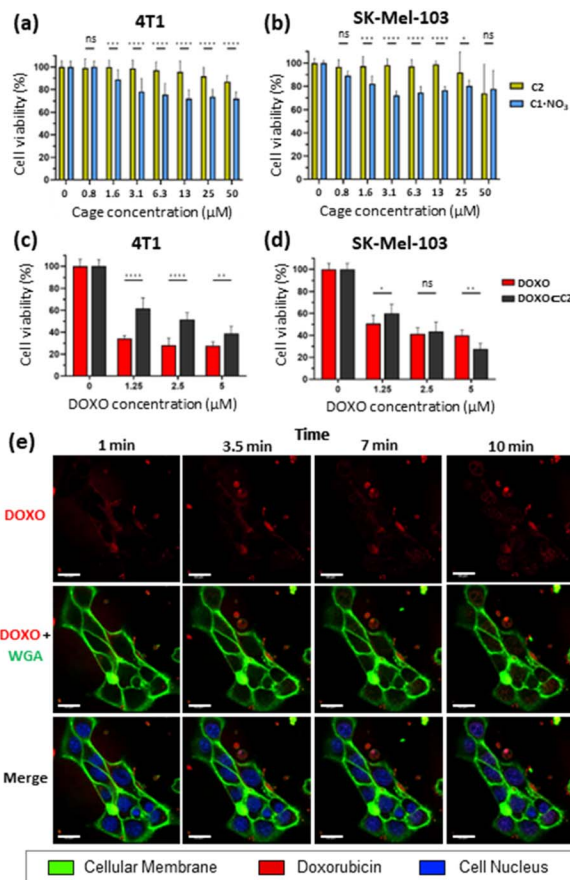


Fig. 7 Cell viability measured by WST-1. Data represented as mean \pm SEM ($n = 3$). Toxicity assay with organic cage C2 (yellow) and palladium metallo-organic cage $\text{C1}\cdot\text{NO}_3$ (blue) on: (a) 4T1 cells and (b) SK-Mel-103 cells. Treatment with DOXO \subset C2 complex (black) and free doxorubicin (red) on: (c) 4T1 cells and (d) SK-Mel-103 cells. Increasing concentrations of DOXO and a fixed concentration of organic cage C2 (25 μM) was used to obtain 95% encapsulation of DOXO. (e) Time-lapse confocal images of SK-Mel-103 cells incubated with Hoechst (blue nuclei marker), WGA (green membrane marker) and treated with the DOXO \subset C2 complex (fluorescent red) at 5 μM up to 10 min. Scale bar represents 20 μm .

Additionally, the treatment with free DOXO resulted in a slightly enhanced antitumoral effect. This can be attributed to the efficient entrapment of the free drug, leading to differential internalisation and processing of $[\text{DOXO}\subset\text{C2}]$ by cancer cells (*vide infra* and Fig. 7c and d). Nevertheless, the results indicate that the activity of DOXO is fully recovered upon its release from $[\text{DOXO}\subset\text{C2}]$. Overall, these results highlights the suitability of the organic cage for drug delivery and the low toxicity of the unloaded organic cage compared to the metallo-organic cage.

To assess the internalisation of the molecular cages, cellular uptake studies of the doxorubicin-encapsulated organic cage were focused on SK-Mel-103 cells. Time-lapse confocal images revealed an increase in DOXO fluorescence (in red) within the cytoplasm of cells (limited by membrane marker in green) that finally reaches the nuclei (in blue) of the cells (Fig. 7). The rapid internalisation, within minutes, suggests passive diffusion as a mechanism of cellular uptake. Although, membrane



permeability is typically limited to molecules with sizes of 1500 Å³,^{72–75} (cage C2 has a molecular volume of 2600 Å³ just slightly over the limit) several works have demonstrated that cages of various sizes can cross cell membranes and deliver anticancer drugs in cells.^{37,41,42,45,48,53,76,77} Analogous time-dependent confocal images with free DOXO confirmed the different drug-internalisation ratio when compared with [DOXO⊂C2] (Fig. S38 in ESI†). The images showed a faster diffusion of DOXO from cellular medium to the cells, with a large amount accumulated in the nucleus after 10 minutes compared to [DOXO⊂C1]. Overall, our studies indicate the proper formation of [DOXO⊂C2] cage and ability to enter in cells and deliver DOXO.

Conclusions

In this work we have compared a similarly shaped metallo-organic cage and a fully organic cage. We found that the organic cage C2 is a promising drug delivery system, with clear advantages over the analogue the palladium cage C1. Both cages are water-soluble requiring only a 1–5% of DMSO to achieve micromolar concentrations, which is compatible with *in vitro* cell culture experiments. The organic cage C2 remains intact in neutral pH phosphate buffer, unlike the palladium cage C1, which breaks down under similar conditions. Cage C2 has a high binding affinity towards the anticancer drug doxorubicin (DOXO), allowing efficient encapsulation at the micromolar concentration required for biological drug delivery. In contrast, C1·NO₃ exhibits weaker binding and disassembly by interaction with DOXO, lacking sufficient binding affinity and stability. In addition, cage disassembly takes place when cage C2 is placed in a slightly acidic pH, making the cage an ideal pH-responsive drug delivery system. In addition to the excellent chemical properties, cage C2 exhibits a remarkable cellular compatibility. Cage C2 does not show any cellular toxicity even at high doses (25–50 μM), in contrast to the significant toxicity observed in C1. Additionally, cage C2 effectively delivers DOXO to cells from the DOXO⊂C2 complex, preserving the cytotoxic activity of this anticancer drug. Overall, the organic cage C2 is a proof-of-concept low toxicity drug delivery system that shows an excellent performance in cells. These results highlight the possibility of using pure organic molecular cages as suitable drug-delivery systems due to their absence of toxicity compared to metallo-organic cages.

Data availability

Data associated to the manuscript is available as ESI† in PDF format and the crystal structure data in CIF format.

Author contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

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