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Thermoresponsive properties of polyacrylamides in physiological solutions†

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Polymer solutions with a lower critical solution temperature (LCST) undergo reversible phase separation when heated above their cloud point temperature (T_{CP} or CPT). As such, they have been proposed for a wide range of biomedical applications, from injectable drug depots to switchable coatings for cell adhesion. However, in systematic studies, the T_{CP} of these thermoresponsive polymers has been mostly measured in non-physiological solutions, thereby hindering the development of their medicinal applications. Here, we analysed the thermoresponsive properties of four acrylamide-based polymers with LCST, namely poly[(*N*-2,2-difluoroethyl)acrylamide] (**pDFEA**), poly[(*N*-isopropyl)acrylamide] (**pNIPAM**), poly[(*N,N*-diethyl)acrylamide] (**pDEA**), and poly[(*N*-acryloyl)pyrrolidine] (**pAP**). As shown by turbidimetry, their T_{CP} in phosphate saline buffer (PBS) and foetal bovine serum (FBS) were consistently lower than those reported in the literature, typically assessed in pure water, even when using the same setup. In addition, these physiological solutions affected the variation of T_{CP} as a function of polymer concentration (1.25 to 10.0 mg mL⁻¹) and molar mass (20 to 50 kg mol⁻¹). As shown by isothermal calorimetry, interactions between proteins in FBS and polymer aggregates were predominantly exothermic, which indicates that protein–polymer complexes are formed through enthalpically driven processes. In conclusion, the T_{CP} of thermoresponsive polymers strongly depends on solvent composition and therefore should be measured under physiological conditions for future medicinal applications.

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Introduction

Many hydrophilic polymers with a lower critical solution temperature (LCST) form homogeneous solutions below a threshold temperature (cloud point temperature, T_{CP} or CPT), but they separate into two phases with high and low concentrations when heating above this T_{CP} .¹ Numerous applications for such “smart polymers” have been proposed, across various fields, including switchable substrates for cell/tissue culture,^{2–5} drug delivery systems,^{6,7} *in situ* depot formation, controlled drug release (therapy/theranostics),^{7–11} gene delivery/therapy,^{12–14} tissue engineering,^{4,15–17} wound dressing,^{18,19} biosensors,²⁰ vaccines/immunotherapy,²¹ and injectable brachytherapy,^{22,23} among others.^{24,25} Therefore, developing such applications requires understanding the properties of these polymers, particularly their T_{CP} .

T_{CP} depends on polymer molar mass (dispersity)^{1,26–29} and concentration,^{1,26,30} on solution pH^{9,31–33} and on the concentration and type of ions^{33–41} (due to the Hofmeister effect³⁴), proteins and other macromolecules^{33,42–45} in solution (due to the excluded volume/molecular crowding effect,⁴⁶ competition for solvent, or non-covalent protein–polymer interactions⁴⁷), among

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Fig. 1 Structures of the study polymers: poly[(*N*-2,2-difluoroethyl)acrylamide] (pDFEA), poly[(*N*-isopropyl)acrylamide] (pNIPAM), poly[(*N,N*-diethyl)acrylamide] (pDEA), and poly[(*N*-acryloyl)pyrrolidine] (pAP).



Fig. 2 Polymer synthesis and subsequent modifications.

Table 1 Polymer characteristics

Polymer		M_w^a (kg mol ⁻¹)	M_n^a (kg mol ⁻¹)	D_M^a	Yield ^b (%)	dn/dc^c (mL g ⁻¹)
pDFEA	F1	26.2	24.2	1.08	68.3	0.088 ± 0.003
	F2	36.2	35.1	1.03	79.7	0.092 ± 0.005
	F3	49.6	46.9	1.06	79.5	0.095 ± 0.004
pNIPAM	I1	20.2	19.6	1.03	50.6	0.167 ± 0.018
	I2	31.6	30.8	1.03	58.1	0.151 ± 0.012
	I3	48.4	45.2	1.07	55.6	0.143 ± 0.013
pDEA	E1	22.3	21.2	1.06	60.5	0.171 ± 0.004
	E2	34.7	31.7	1.09	61.3	0.176 ± 0.011
	E3	41.3	37.5	1.10	57.5	0.145 ± 0.010
pAP	P1	19.6	17.6	1.11	86.0	0.192 ± 0.007
	P2	36.0	32.9	1.09	77.7	0.180 ± 0.015
	P3	51.2	46.3	1.11	79.1	0.164 ± 0.007

^a Determined by SEC. ^b Polymerisation yield after the purification procedure based on monomer weight. ^c Determined by differential refractometry in PBS at 29 °C and 620 nm in PBS.

merisation⁷⁶ (Fig. 2) using 4-cyano-4-[(dodecylsulfanylthiocarbonyl)sulfanyl]pentanoic acid as the CTA and 4,4'-azobis(4-cyanovaleric acid) (ACVA) as the initiator (see Table S1† for the initial quantities of the reagents). Subsequently, we purified and mixed the polymers first with propylamine (aminolysis of the terminal CTA)⁷⁶ and then with an excess of methyl acrylate (to mask the reactive thiol moiety by Michael addition),⁷⁶ as shown in Fig. 2 and Table S2.† Lastly, we purified and characterised these polymers by size exclusion chromatography and

NMR spectroscopy to determine their purity and to confirm CTA removal (see chapter S10.3†). The properties of these polymers are outlined in Table 1.

PBS and FBS affect the T_{CP} of thermo-responsive polyacrylamide solutions

The T_{CP} s of our thermo-responsive polymers are either known (for pNIPAM^{33–37,43,44,67,69} and pAP³⁷) or expected (for pDFEA and pDEA) to be lower in physiologically relevant solutions



(FBS and PBS) than in pure water because these buffered solutions have a higher pH and ion concentration (as discussed in the Introduction). However, these polymers also have terminal carboxylic moieties ($pK_a \approx 4.8$), and while they are almost exclusively dissociated at neutral pH, unbuffered water dissolves atmospheric carbon dioxide,⁵² lowering the pH to values near the pK_a of carboxylic acid (Table S7†). As a result, the carboxylic groups will no longer be fully deprotonated, decreasing the hydrophilicity^{32,52} and T_{CP} of these polymers.⁵² In contrast, both PBS and FBS (140 mM; pH = 7.4) reliably maintain the pH and osmotic pressure at physiological values.^{70,72}

Considering these differences, we have compared the effect of different solvents (water, Dulbecco's PBS⁷⁰ and FBS) on T_{CP} ($c = 10.0 \text{ mg mL}^{-1}$; Fig. 3). In line with previous studies, most T_{CP} values of our polymers were higher in pure water than in PBS^{33–37,40,43,44,67} and FBS^{33,43} because the ions and proteins of buffered solutions lower the T_{CP} of these polymers. Since differences in T_{CP} s between solvents may be significant for many applications, polymers should be tested in an environment as similar as possible to that of the intended application.

Thermoresponsive polymers may require DLS for accurate T_{CP} determination

As shown in Fig. 3B, the T_{CP} of pDFFEA with the highest molar mass (F3) was much higher in PBS than in water. To understand the unexpected T_{CP} of this polymer, as measured by turbidimetry, we assessed polymer aggregation as a function of temperature (10 to 50 °C) by DLS, in PBS, at a concentration of 10.0 mg mL^{-1} (Fig. S45–S56†). Unlike most samples, pDFFEA polymers showed two major changes in population size with the increase in temperature (Fig. S45–S47†). The first change (at 26 to 30 °C) can be ascribed to unimer aggregation (radius 20 to 100 nm), and the second (from 40 to 50 °C) to aggregate coalescence into even larger polymer assemblies (radius

1000 nm or larger). Thus, turbidimetry detects the first thermal change in most samples, but only the second in F3.

Long pDFFEA (co)polymers, such as F3, may form non-typical nanogel-like aggregates with low polymer concentrations, as shown in our previous studies.^{63,64,79} In these particles, the aggregation causes only a minor local increase in polymer concentration. When combined with the low dn/dc of the solute (Table 1), this increase in polymer concentration accounts for the small difference in refractive indices between the phase-separated polymer and the bulk solution, which may prevent an accurate determination of T_{CP} by turbidimetry.⁵⁹ Upon further heating, these nanogel-like particles aggregate/coalesce, increasing the turbidity. Therefore, discrepancies in turbidimetric measurements may be explained by differences in the architecture of polymer aggregates and by the low dn/dc of pDFFEA.

The effect of FBS proteins on T_{CP} varies with polymer concentration

Proteins (in FBS) can affect the T_{CP} of polymers indirectly ('non-specifically', *i.e.*, by competing with polymers for its solvation as well as by excluded volume/crowding effect^{34,46}) or directly ('specifically', *i.e.*, by forming complexes with the polymers^{34,47}).³⁴ In turn, inorganic salts (in PBS and FBS) can also indirectly interfere with polymers by interacting with and destabilizing their solvation shell, thus decreasing their solubility and T_{CP} (Hoffmeister effect), regardless of polymer concentration.^{33,40} Unlike inorganic salts, however, proteins affect T_{CP} as a function of polymer concentration, as shown by our results (Fig. 4).

At low polymer concentrations (1.25 to 2.50 mg mL^{-1}), polymers form soluble protein–polymer complexes (protein binding) in FBS, which prevent them from aggregating, thereby increasing their T_{CP} above that of PBS solutions, as observed in pDEA and pNIPAM (at both low and high pAP concentrations). Because the polymer-binding capacity of proteins



Fig. 3 (A) Comparison of 6 turbidimetric measurements of I2 (pNIPAM) in water, PBS and FBS ($c = 10.0 \text{ mg mL}^{-1}$); (B) T_{CP} of all polymers in water, PBS and FBS ($c = 10.0 \text{ mg mL}^{-1}$) expressed as a mean of 6 measurements \pm standard deviation. The results indicate a shift in T_{CP} .



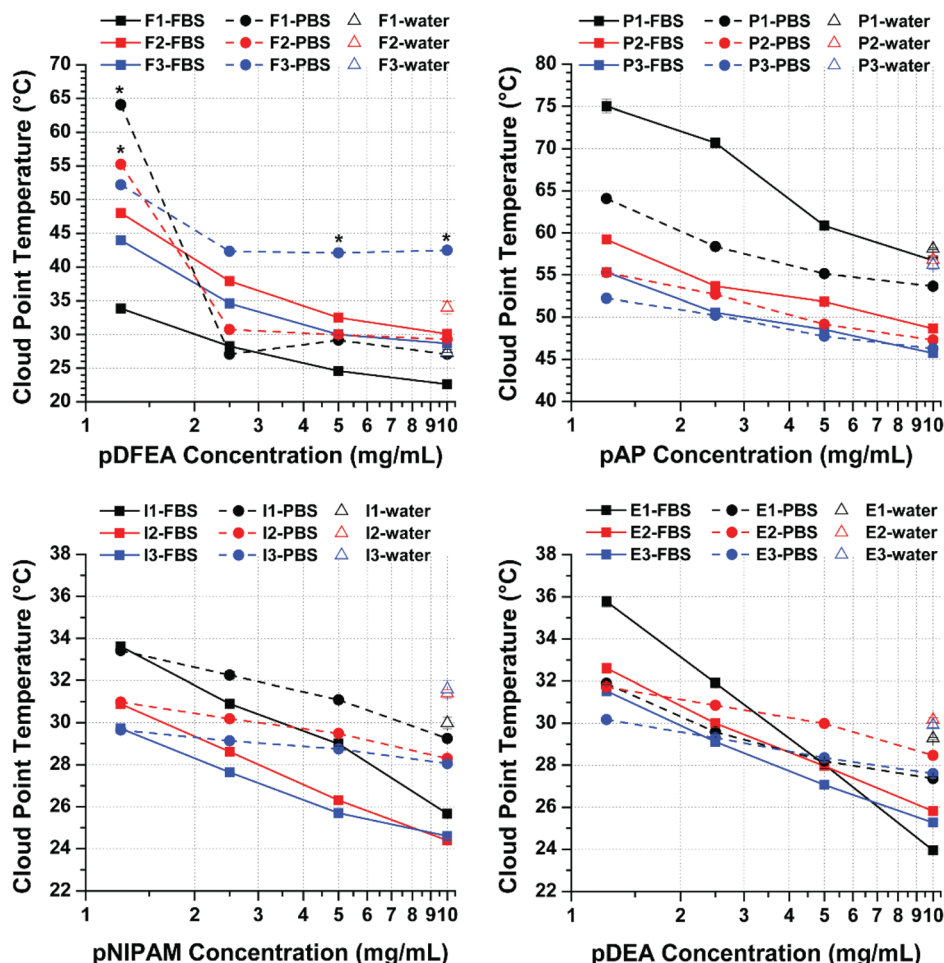


Fig. 4 Plot of T_{CPS} of the polymers pDFEA (top-left), pAP (top-right), pNIPAM (bottom-left) and pDEA (bottom-right) in FBS and PBS as a function of polymer concentrations. All T_{CPS} are expressed as the mean of 6 measurement cycles \pm standard deviation. Asterisks (*) indicate potential outliers.

is limited (albeit very high for pAP), this effect is only detected at low polymer concentrations and decreases with the increase in polymer concentration (until being offset by ‘non-specific’ effects, which decrease the T_{CP}).

At high polymer concentrations (5.00 to 10.0 mg mL⁻¹), in contrast, proteins predominantly have a ‘non-specific’ effect by competing with polymer chains for solvation with the polymer^{33,34,40,41,80} (similarly to inorganic ions), thereby facilitating aggregation and lowering the T_{CPS} . Proteins may also stabilize polymer aggregates *via* hydrophobic interactions, further lowering the T_{CPS} . Under such conditions, most polymers in FBS have the lowest T_{CPS} of all three media tested in this study (except for F3 in PBS, as discussed above).

For the purpose of this analysis (Fig. 4), we disregarded the transition temperatures of the highest polymer concentrations (20.0 and 40.0 mg mL⁻¹) because they differed considerably between independent measurements (low reproducibility), not only in FBS but also in PBS. Nevertheless, the complete dataset is provided in Tables S3–S6.† Furthermore, in the range used in this analysis (1.25 to 10.0 mg mL⁻¹), except for a few outliers, all three molar masses of each polymer showed similar

T_{CP} trends, that is, T_{CP} decreased with the increase in polymer concentration, in line with previous studies.^{1,34–37}

PBS and FBS differentially affect LCST polymer aggregation

LCST polymer aggregation is an entropy-driven endothermic process^{1,26,59} affected by surrounding ions/proteins. Adding these ions/proteins can induce conformational changes in the polymer (aggregation) or affect its solvation shell, both of which can be detected as heat effects.^{1,82–84} A decrease in enthalpy after adding ions/proteins indicates an enthalpic effect, *i.e.*, polymers interact with ions/proteins (or new strong polymer–polymer interactions are formed). Conversely, an increase in enthalpy after adding these ions/proteins indicates an entropic effect, *i.e.*, the loss of specific interactions, thereby increasing the entropy, *e.g.*, due to the loss of the solvation shell.⁸⁴

Considering the above, we assessed the enthalpy of interaction of pDFEA, pNIPAM, pDEA, and pAP with PBS and FBS by isothermal titration calorimetry (ITC), which revealed a complex titration isotherm⁸¹ (Fig. 5B), with three phases: I, II and III (Fig. 5A). In phase I, the heat flux can be ascribed to





Fig. 5 ITC of 10.0 mg mL⁻¹ F3 (pDFEA solution in ultrapure water with PBS: heat flux as a function of time, titration of PBS to water (blank, red line), titration of PBS to polymer (black line) (A); integrated heat, normalised to 1 mol of F3 injectant (scatter), fit to model (line) (B). Enthalpies of the titration of 10.0 mg mL⁻¹ polymer solutions with PBS (C) and FBS (D), normalised to 1 mol of monomer units at three different temperatures. The values of enthalpies are outlined in Table S7.†

the neutralization of the terminal carboxylic acid because adding 3 to 5 μL of PBS (pH = 7.41) or FBS (pH = 7.46) to the solution of benzoic acid had a similar effect on the heat flux (Fig. S41†). For this reason, the corresponding data points were excluded from the titration isotherms. Subsequent adding of titrant induced exo- or endothermic processes, which strongly depended on the titrant concentration (phase II on the titration isotherms). After the critical concentration of titrant (Fig. 5B), a non-zero heat flux, weakly dependent on the titrant, was still detected (phase III). Based on these results, we focused on the heat flux from phase II to analyse the solvent-polymer interactions.

PBS promotes aggregation in thermoresponsive polymers by decreasing their solvation shell

In non-aggregated polymers (at low temperatures), adding PBS to the solution had a positive enthalpic effect on pNIPAM, pAP and pDEA, as expected based on the Hoffmeister effect, but surprisingly had a significantly negative enthalpic effect on pDFEA. On the one hand, PBS decreases the solvation shell of all polymers (positive enthalpic effect). On the other hand, PBS increases polymer-polymer bonds in the aggregates (negative enthalpic effect). Since PBS had a net negative enthalpic effect on pDFEA, pDFEA must have strong intramolecular interactions (possibly due to hydrogen bonding between CF₂H moieties⁷⁴ – further investigated in chapter S7.4†).

In aggregated polymers (at high temperatures), PBS had a negative enthalpic effect on pDFEA, pDEA and pAP, indicating that this buffer promotes the formation of enthalpically favourable polymer-polymer. However, adding PBS to pNIPAM had a positive enthalpic effect, suggesting further dehydration result-

ing from salting out. Overall, PBS disrupts the solvation shell of thermoresponsive polymers, thereby promoting aggregation.

FBS proteins stabilize LCST polymer aggregates

Adding FBS to both non-aggregated and aggregated polymers (at both low and high temperatures) had a more negative enthalpic effect than adding PBS due to the additional strong polymers-proteins interactions (Fig. 5). The amphiphilic proteins in FBS may bind to polymers and polymeric aggregates^{5,33} via hydrophobic interactions^{3,5} and thus stabilize them. In all polymers, adding FBS to polymer aggregates had a stronger negative enthalpic effect than adding FBS to non-aggregating polymers, possibly because polymer aggregates are more prone to interact with proteins through hydrophobic interactions. Consequently, FBS stabilizes thermoresponsive polymer aggregates more strongly than PBS.

Conclusions

The cloud point temperature (T_{CP}) of thermoresponsive polyacrylamides is considerably lower in physiologically relevant solvents (PBS and FBS) than in water. In particular, FBS proteins stabilize LCST polymer aggregates, but the effect of FBS proteins on T_{CP} depends on polymer concentration. At high polymer concentrations, proteins decrease the T_{CP} by competing for solvation. At low polymer concentrations, by contrast, proteins form complexes with the polymers, thus increasing aggregation. However, proteins have limited polymer-binding capacities, so their effect decreases with the increase in con-



centration. In turn, PBS promotes aggregation in thermo-responsive polymers by decreasing their solvation shell. Overall, our results suggest that thermo-responsive polymers with a high potential for biomedical applications should be characterised (i) by DLS for accurate T_{CP} determination and (ii) in physiologically relevant solutions rather than in pure water because they may be otherwise discarded merely for their unsuitable LCST behaviour in water. Moreover, our findings may enable us to better predict the biological properties of thermo-responsive polymers *in vivo* based on their T_{CP} . Therefore, these results may be used to optimise polymeric drug delivery systems for *in vivo* applications through *in vitro* studies.

Conflicts of interest

The authors have no conflicts of interests to declare.

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References

- V. Aseyev, H. Tenhu and F. M. Winnik, in *Self Organized Nanostructures of Amphiphilic Block Copolymers II*, ed. A. H. E. Müller and O. Borisov, Springer Berlin Heidelberg, Berlin, Heidelberg, 2010, vol. 242, pp. 29–89.
- L. Yang, X. Fan, J. Zhang and J. Ju, *Polymers*, 2020, **12**, 389.
- N. Yamada, T. Okano, H. Sakai, F. Karikusa, Y. Sawasaki and Y. Sakurai, *Makromol. Chem., Rapid Commun.*, 1990, **11**, 571–576.
- B. L. Ekerdt, C. M. Fuentes, Y. Lei, M. M. Adil, A. Ramasubramanian, R. A. Segalman and D. V. Schaffer, *Adv. Healthcare Mater.*, 2018, **7**, 1800225.
- T. Okano, N. Yamada, M. Okuhara, H. Sakai and Y. Sakurai, *Biomaterials*, 1995, **16**, 7.
- K. Kolouchova, O. Groborz, P. Svec, Z. Starcuk, M. Slouf and M. Hruby, *Biomacromolecules*, 2021, **22**, 2325–2337.
- W. B. Liechty, D. R. Kryscio, B. V. Slaughter and N. A. Peppas, *Annu. Rev. Chem. Biomol. Eng.*, 2010, **1**, 149–173.
- A. Hatefi and B. Amsden, *J. Controlled Release*, 2002, **80**, 9–28.
- K. Kolouchova, D. Jirak, O. Groborz, O. Sedlacek, N. Ziolkowska, M. Vit, E. Sticova, A. Galisova, P. Svec, J. Trousil, M. Hajek and M. Hruby, *J. Controlled Release*, 2020, **327**, 50–60.
- M. A. Ward and T. K. Georgiou, *Polymers*, 2011, **3**, 1215–1242.
- E. Ruel-Gariépy and J.-C. Leroux, *Eur. J. Pharm. Biopharm.*, 2004, **58**, 409–426.
- A. M. Alhoranta, J. K. Lehtinen, A. O. Urtili, S. J. Butcher, V. O. Aseyev and H. J. Tenhu, *Biomacromolecules*, 2011, **12**, 3213–3222.
- M. D. Lavigne, S. S. Pennadam, J. Ellis, L. L. Yates, C. Alexander and D. C. Górecki, *J. Gene Med.*, 2007, **9**, 44–54.
- S. Ganta, H. Devalapally, A. Shahiwala and M. Amiji, *J. Controlled Release*, 2008, **126**, 187–204.
- F. Doberenz, K. Zeng, C. Willems, K. Zhang and T. Groth, *J. Mater. Chem. B*, 2020, **8**, 607–628.
- K. Y. Lee and D. J. Mooney, *Chem. Rev.*, 2001, **101**, 1869–1880.
- S. Van Vlierberghe, P. Dubruel and E. Schacht, *Biomacromolecules*, 2011, **12**, 1387–1408.
- D.-Q. Wu, J. Zhu, H. Han, J.-Z. Zhang, F.-F. Wu, X.-H. Qin and J.-Y. Yu, *Acta Biomater.*, 2018, **65**, 305–316.
- I. Elloumi-Hannachi, M. Yamato and T. Okano, *J. Intern. Med.*, 2010, **267**, 54–70.
- Y. Wei, Q. Zeng, Q. Hu, M. Wang, J. Tao and L. Wang, *Biosens. Bioelectron.*, 2018, **99**, 136–141.
- N. Pacifici, A. Bolandparvaz and J. S. Lewis, *Adv. Ther.*, 2020, **3**, 2000129.
- O. Sedláček, P. Černoch, J. Kučka, R. Konefal, P. Štěpánek, M. Vetrík, T. P. Lodge and M. Hrubý, *Langmuir*, 2016, **32**, 6115–6122.
- M. Hrubý, J. Kucka, O. Lebeda, H. Mackova, M. Babič, Č. Koňák, M. Studenovský, A. Síkora, J. Kozempel and K. Ulbrich, *J. Controlled Release*, 2007, **119**, 25–33.
- E. Ayano and H. Kanazawa, *J. Sep. Sci.*, 2006, **29**, 738–749.
- D. E. Bergbreiter, *Chem. Rev.*, 2002, **102**, 3345–3384.
- A. Halperin, M. Kröger and F. M. Winnik, *Angew. Chem., Int. Ed.*, 2015, **54**, 15342–15367.
- N. S. Jeong, M. Hasan, D. J. Phillips, Y. Saaka, R. K. O'Reilly and M. I. Gibson, *Polym. Chem.*, 2012, **3**, 794–799.
- D. G. Lessard, M. Ousaleem and X. X. Zhu, *Can. J. Chem.*, 2001, **79**, 1870–1874.
- Q. Li, A. P. Constantinou and T. K. Georgiou, *J. Polym. Sci.*, 2021, **59**, 230–239.
- S. Fujishige, K. Kubota and I. Ando, *J. Phys. Chem.*, 1989, **93**, 3311–3313.
- Y. Pei, J. Chen, L. Yang, L. Shi, Q. Tao, B. Hui and J. Li, *J. Biomater. Sci., Polym. Ed.*, 2004, **15**, 585–594.
- C. Pietsch, R. Hoogenboom and U. S. Schubert, *Angew. Chem., Int. Ed.*, 2009, **48**, 5653–5656.
- Y. Hiruta, Y. Nagumo, Y. Suzuki, T. Funatsu, Y. Ishikawa and H. Kanazawa, *Colloids Surf., B*, 2015, **132**, 299–304.
- Y. Zhang, S. Furyk, L. B. Sagle, Y. Cho, D. E. Bergbreiter and P. S. Cremer, *J. Phys. Chem. C*, 2007, **111**, 8916–8924.
- R. Freitag and F. Garret-Flaudy, *Langmuir*, 2002, **18**, 3434–3440.



- 36 Y. Zhang, S. Furyk, D. E. Bergbreiter and P. S. Cremer, *J. Am. Chem. Soc.*, 2005, **127**, 14505–14510.
- 37 K. Skrabania, J. Kristen, A. Laschewsky, O. Akdemir, A. Hoth and J.-F. Lutz, *Langmuir*, 2007, **23**, 84–93.
- 38 H. L. Judah, P. Liu, A. Zarbakhsh and M. Resmini, *Polymers*, 2020, **12**, 2590.
- 39 Ł. Otulakowski, M. Kasprów, A. Strzelecka, A. Dworak and B. Trzebicka, *Polymers*, 2021, **13**, 90.
- 40 M. M. Bloksma, D. J. Bakker, C. Weber, R. Hoogenboom and U. S. Schubert, *Macromol. Rapid Commun.*, 2010, **31**, 724–728.
- 41 J. P. Magnusson, A. Khan, G. Pasparakis, A. O. Saeed, W. Wang and C. Alexander, *J. Am. Chem. Soc.*, 2008, **130**, 10852–10853.
- 42 X. Zhu, X. Gu, L. Zhang and X.-Z. Kong, *Nanoscale Res. Lett.*, 2012, **7**, 519.
- 43 T. Trongsatitkul and B. M. Budhlall, *Colloids Surf., B*, 2013, **103**, 244–252.
- 44 K. Bebis, M. W. Jones, D. M. Haddleton and M. I. Gibson, *Polym. Chem.*, 2011, **2**, 975–982.
- 45 Y. Okada and F. Tanaka, *Macromolecules*, 2005, **38**, 4465–4471.
- 46 R. J. Ellis, *Trends Biochem. Sci.*, 2001, **26**, 597–604.
- 47 S. Suzuki, T. Sawada, T. Ishizone and T. Serizawa, *Chem. Commun.*, 2016, **52**, 5670–5673.
- 48 F. D. Jochum and P. Theato, *Chem. Soc. Rev.*, 2013, **42**, 7468–7483.
- 49 B. Ray, Y. Okamoto, M. Kamigaito, M. Sawamoto, K. Seno, S. Kanaoka and S. Aoshima, *Polym. J.*, 2005, **37**, 234–237.
- 50 M. Hrubý, V. Šubr, J. Kučka, J. Kozempel, O. Lebeda and A. Sikora, *Appl. Radiat. Isot.*, 2005, **63**, 423–431.
- 51 Y. Xia, N. A. D. Burke and H. D. H. Stöver, *Macromolecules*, 2006, **39**, 2275–2283.
- 52 G. Cao, G. Li, Q. Yang, Z. Liu, Z. Liu and J. Jiang, *Macromol. Rapid Commun.*, 2018, **39**, 1700684.
- 53 C. Panayiotou and I. C. Sanchez, *J. Phys. Chem.*, 1991, **95**, 10090–10097.
- 54 F. Afroze, E. Nies and H. Berghmans, *J. Mol. Struct.*, 2000, **554**, 55–68.
- 55 L. P. N. Rebelo, Z. P. Visak, H. C. de Sousa, J. Szydłowski, R. Gomes de Azevedo, A. M. Ramos, V. Najdanovic-Visak, M. Nunes da Ponte and J. Klein, *Macromolecules*, 2002, **35**, 1887–1895.
- 56 C. Zhao, Z. Ma and X. X. Zhu, *Prog. Polym. Sci.*, 2019, **90**, 269–291.
- 57 J. Heyda and J. Dzubiella, *J. Phys. Chem. B*, 2014, **118**, 10979–10988.
- 58 B. H. Chang and Y. C. Bae, *Polymer*, 1998, **39**, 6449–6454.
- 59 Q. Zhang, C. Weber, U. S. Schubert and R. Hoogenboom, *Mater. Horiz.*, 2017, **4**, 109–116.
- 60 D. G. Lessard, M. Ousalem, X. X. Zhu, A. Eisenberg and P. J. Carreau, *J. Polym. Sci., Part B: Polym. Phys.*, 2003, **41**, 1627–1637.
- 61 M. Philipp, R. Aleksandrova, U. Müller, M. Ostermeyer, R. Sanctuary, P. Müller-Buschbaum and J. K. Krüger, *Soft Matter*, 2014, **10**, 7297–7305.
- 62 F. Meersman, J. Wang, Y. Wu and K. Heremans, *Macromolecules*, 2005, **38**, 8923–8928.
- 63 D. Babuka, K. Kolouchová, O. Groborz, Z. Tošner, A. Zhigunov, P. Štěpánek and M. Hrubý, *Nanomaterials*, 2020, **10**, 2231.
- 64 D. Babuka, K. Kolouchova, M. Hruby, O. Groborz, Z. Tosner, A. Zhigunov and P. Stepanek, *Eur. Polym. J.*, 2019, **121**, 109306.
- 65 P. Štěpánek, in *Dynamic Light Scattering*, ed. W. Brown, Clarendon Press, Oxford, 1993, p. 760.
- 66 C. Boutris, E. G. Chatzi and C. Kiparissides, *Polymer*, 1997, **38**, 2567–2570.
- 67 H. G. Schild and D. A. Tirrell, *J. Phys. Chem.*, 1990, **94**, 4352–4356.
- 68 K. Van Durme, G. Van Assche and B. Van Mele, *Macromolecules*, 2004, **37**, 9596–9605.
- 69 K. Otake, H. Inomata, M. Konno and S. Saito, *Macromolecules*, 1990, **23**, 283–289.
- 70 R. Dulbecco and M. Vogt, *J. Exp. Med.*, 1954, **99**, 167–182.
- 71 T. Yao and Y. Asayama, *Reprod. Med. Biol.*, 2017, **16**, 99–117.
- 72 J. van der Valk, K. Bieback, C. Buta, B. Cochrane, W. G. Dirks, J. Fu, J. J. Hickman, C. Hohensee, R. Kolar, M. Liebsch, F. Pistollato, M. Schulz, D. Thieme, T. Weber, J. Wiest, S. Winkler and G. Gstraunthaler, *ALTEX – Altern. Anim. Exp.*, 2018, **35**, 99–118.
- 73 J. M. Bak and H. Lee, *J. Polym. Sci., Part A: Polym. Chem.*, 2013, **51**, 1976–1982.
- 74 Y. Zafrani, D. Yeffet, G. Sod-Moriah, A. Berliner, D. Amir, D. Marciano, E. Gershonov and S. Saphier, *J. Med. Chem.*, 2017, **60**, 797–804.
- 75 B. D. Monnery and R. Hoogenboom, *Polym. Chem.*, 2019, **10**, 3480–3487.
- 76 C. Zhou, M. A. Hillmyer and T. P. Lodge, *Macromolecules*, 2011, **44**, 1635–1641.
- 77 N. Bertrand, J. G. Fleischer, K. M. Wasan and J.-C. Leroux, *Biomaterials*, 2009, **30**, 2598–2605.
- 78 L. Wyffels, T. Verbruggen, B. D. Monnery, M. Glassner, S. Stroobants, R. Hoogenboom and S. Staelens, *J. Controlled Release*, 2016, **235**, 63–71.
- 79 K. Kolouchova, O. Sedlacek, D. Jirak, D. Babuka, J. Blahut, J. Kotek, M. Vit, J. Trousil, R. Konefał, O. Janouskova, B. Podhorska, M. Slouf and M. Hruby, *Biomacromolecules*, 2018, **19**, 3515–3524.
- 80 H. I. Okur, J. Hladílková, K. B. Rembert, Y. Cho, J. Heyda, J. Dzubiella, P. S. Cremer and P. Jungwirth, *J. Phys. Chem. B*, 2017, **121**, 1997–2014.
- 81 T. Wiseman, S. Williston, J. F. Brandts and L.-N. Lin, *Anal. Biochem.*, 1989, **179**, 131–137.
- 82 P. D. Ross and M. V. Rekharsky, *Biophys. J.*, 1996, **71**, 2144–2154.
- 83 J. M. Pestman, J. Kevelam, M. J. Blandamer, H. A. van Doren, R. M. Kellogg and J. B. F. N. Engberts, *Langmuir*, 1999, **15**, 2009–2014.
- 84 G. Némethy, *Angew. Chem., Int. Ed. Engl.*, 1967, **6**, 195–206.

