

Organic & Biomolecular Chemistry

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

Preparation of Luminescent Chemosensors by Post-Functionalization of Vesicle Surfaces

Andreas Müller and Burkhard König*

Institut für Organische Chemie, Universität Regensburg, 93040 Regensburg, Germany

Fax: +49 943 1717; Tel: +49 943 4575; E-mail: burkhard.koenig@ur.de

Abstract:

Surface-reactive luminescent vesicles were prepared by self-assembly of phospholipids, amphiphilic maleimides and fluorophors in aqueous solution. Those preformed liposomes were functionalized with various thiolated receptor units using a thiol-click reaction. As recognition elements, a bis-Zn²⁺-cyclen derivative for the detection of phosphate moieties or a DNA aptamer for the specific binding of the antibiotic ampicillin were utilized. A FRET-based assay revealed the close spatial proximity of the membrane-embedded dansyl molecules with the subsequently immobilized thiols, which is the origin for the signaling mechanism of the obtained vesicular sensors. Those receptor-functionalized liposomes indicate the binding of the targets to their surface by changes of the fluorescence emission properties of the membrane co-embedded carboxyfluorescein dyes. The post-functionalization concept can also be used for molecular imprinting on vesicle surfaces. The template-guided patterning of receptors based on bis-Zn²⁺-cyclen resulted in fluorescent sensors suitable for the specific recognition of a bivalent peptide.

1. Introduction

Molecular recognition at biomembranes is of fundamental importance for biological processes like cell recognition, adhesion, fusion or membrane transport phenomena. Since phospholipid vesicles can be considered as simple cell mimics, non-covalent interactions at the surface of receptor-functionalized synthetic liposomes resemble recognition events at membranes as they appear in nature.¹ Hence, the knowledge obtained from the investigation of artificial vesicular systems can help to gain improved insights into the correspondent processes in biology. Conversely, a better understanding of the natural prototypes can be exploited for the development of powerful devices for analytical applications. Various strategies have been established to transform the binding event at the vesicle membrane into

an observable signal.² Fluorescent assays provide high sensitivities allowing the detection of analytes present in trace amounts, whereas colorimetric formats enable the analytic readout by the naked eye.³ Typically, the signaling unit is in direct contact with the recognition element by either covalent attachment^{4,5} or by non-covalent coordination as in indicator-displacement assays.^{4,6} However, membranes are highly dynamic due to their supramolecular composition.¹ Based on this, our group recently reported on a novel strategy for the modular construction of chemosensors by means of unilamellar vesicular membranes, which serve as self-assembled supporting matrix for amphiphilic metal complex receptors and fluorescent reporter dyes.⁷ Due to lateral phase separation, the co-embedded amphiphilic fluorophores and binding sites are expected to cluster on the surface resulting in fluorescence quenching of the amphiphilic dyes (Figure 1). Binding of analyte molecules to the surface-accessible receptors alters the physico-chemical properties in their microenvironment and thus induces a reorganization of the membrane components. These processes influence the optical properties of the co-embedded dyes, which in turn respond with a change of their emission intensity.

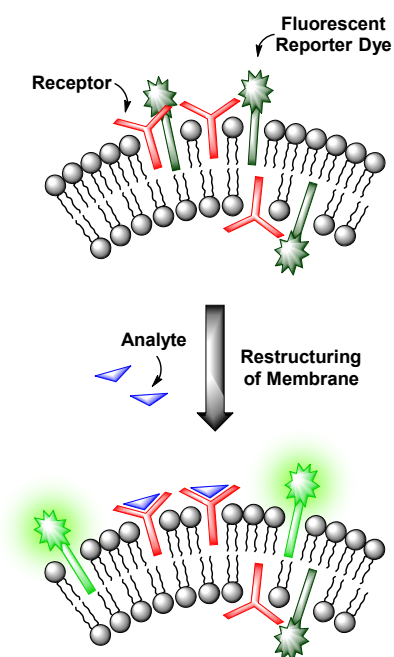


Figure 1: Sensing mechanism of receptor-functionalized fluorescent vesicles.

This co-embedding strategy allows an effortless preparation of sensor devices by simple mixing of different functional amphiphiles in aqueous solution. By careful variation of the binding and signaling units, their ratios and concentrations on the surface and by the choice of suitable phospholipids determining the physical properties of the membrane, vesicular sensors with specific characteristics can be designed. However, as the synthesis of

the amphiphilic receptors can be quite tedious, the availability of suitable binding sites and hence the variety of addressable analytes is limited. Moreover, control over the two-dimensional receptor assembly for multipoint interactions on the surface is still a challenge.⁸ Its accomplishment is an essential requirement for the highly selective recognition of multivalent analytes. In order to approach these issues, we developed a more universal method, which allows the facile preparation of vesicular chemosensors in a modular fashion by means of surface “post-functionalization” *via* click chemistry. The surface modification of preformed liposomes is an established method for the conjugation of various ligands ranging from sugars to peptides, proteins and antibodies and has found applications in, *e.g.*, the investigation of drug delivery systems.⁹ We now combine this concept of vesicle post-modification with our co-embedding strategy of functional amphiphiles into phospholipid membranes. Only groups of the vesicle membranes which are exposed to the outer side are functionalized. The resulting asymmetric membrane composition represents a much better mimic of the bilayers appearing in nature. In view of more complex receptors of biological origin, we utilized a thiol–ene click reaction¹⁰ due to the frequent occurrence of thiol groups in biomolecules. The reaction is fully biocompatible and does not, in contrast to the copper(I)-catalyzed azide–alkyne cycloaddition,¹¹ require a metal catalyst.

2. Results and Discussion

We first tested the applicability of the UV light-induced radical addition of thiols to electron-rich alkene moieties.¹² This type of reaction would implicate the possibility to externally trigger the functionalization process by controlled exposure to UV light. However, the envisaged photoreaction is at micromolar concentrations in aqueous solution not efficient. Thus, we focused on the widely used Michael addition of thiols to maleimides.¹³ This reaction performs well over a large concentration range in aqueous media under benign conditions without the formation of byproducts and is commonly applied in bioconjugation.¹⁰ We recently employed this method for the construction of an aptamer-based biosensor by attachment of thiol-modified aptamers to membrane-embedded amphiphilic maleimide **Mal-C₁₆** (Figure 2).¹⁴

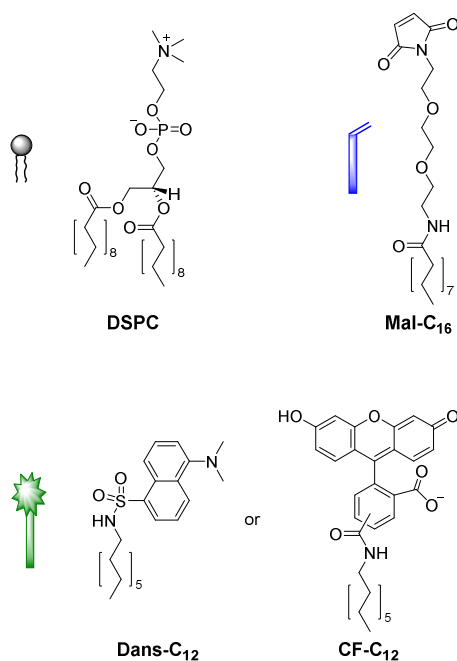


Figure 2: Structures of the functional amphiphiles used for vesicle preparation.

Here, we report on the development of the post-functionalization strategy of luminescent vesicle surfaces with receptor sites and demonstrate its facile and versatile applicability for the preparation of various sensors. As signaling units, either amphiphilic dansyl dye **Dans-C₁₂**¹⁵ or carboxyfluorescein **CF-C₁₂** were incorporated into membranes of small unilamellar DSPC vesicles. Figure 3 depicts the different thiols which were used for our investigations. The syntheses of compounds **Mal-C₁₆**¹⁴ and **CF-C₁₂**^{7a} were previously reported by us, the preparation of **Cys-Trp** and **Zn-Cyclen₂** is described in the ESI.

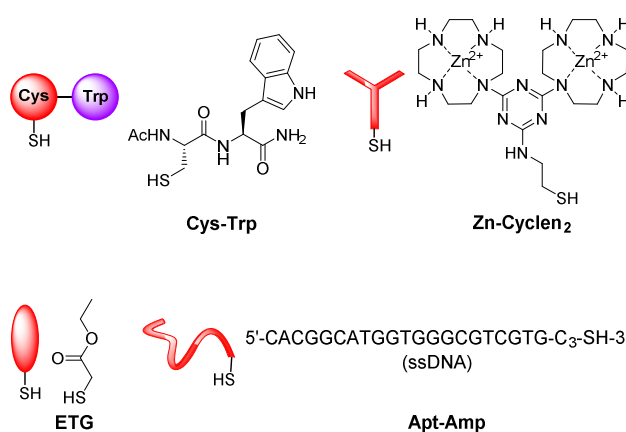


Figure 3: Thiols used for the post-functionalization of vesicle surfaces.

2.1 Optimization of the Post-Functionalization of Preformed Vesicles

The sensing mechanism of our vesicular sensors is based on the close spatial proximity of the binding sites to the membrane-embedded dyes. To test whether the subsequently immobilized receptor molecules fulfill this condition, we developed an assay based on the distance-dependent Förster resonance energy transfer (FRET, Figure 4). As simple model system, we synthesized dipeptide **Cys-Trp** (Figure 3). The FRET partner of tryptophan, the amphiphilic dansyl dye **Dans-C₁₂** (Förster distance: 2.1 nm),¹⁶ was co-embedded (5.0 mol-%) into the membrane of DSPC vesicles together with the attachment site for thiols **Mal-C₁₆** (5.0 mol-%). The preparation of the vesicles was performed in 25 mM HEPES buffer at a pH of 7.4 according to previously reported procedures (see ESI for details). Extrusion through 100 nm polycarbonate membranes yielded vesicles **V-Dans** with a size distribution of around 130–150 nm (see Figure S1, ESI). The non-functionalized vesicles were stable and did not show significant aggregation after one day (see Figure S2, ESI).

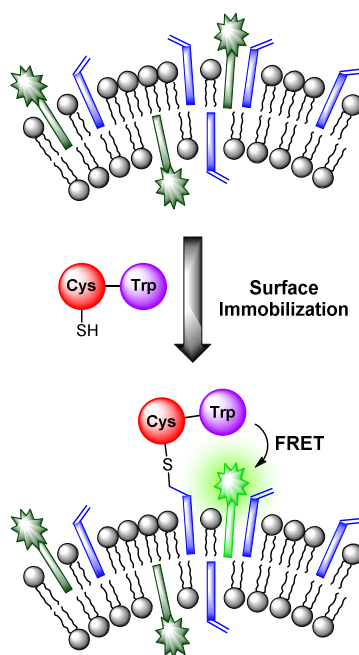


Figure 4: FRET assay proving the close spatial proximity of immobilized thiols to the membrane-embedded fluorophors.

Addition of **Cys-Trp** to **V-Dans** showed a time-dependent decrease of the tryptophan emission evidencing the fluorescence energy transfer and a considerable increase of the dansyl signal (Figure 5).

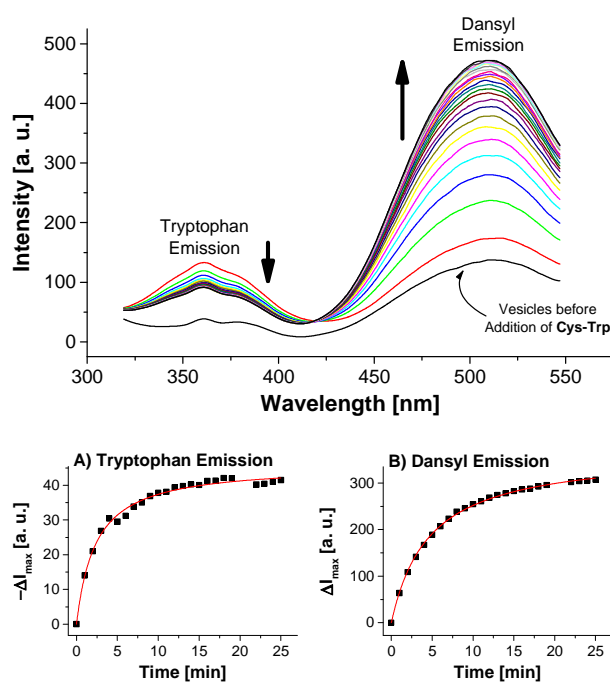


Figure 5: Top: Time-dependent fluorescence spectra of vesicles **V-Dans** after addition of 3.0 mol-% of peptide **Cys-Trp** ($\lambda_{\text{ex}} = 280$ nm). Bottom: Course of surface reaction as evidenced by the emission signals of either the tryptophan or dansyl dye.

Apart from the contribution of the resonance energy transfer from the tryptophan moiety, the extraordinarily strong increase of the dansyl signal must be attributed to two additional independent effects: (i) It is well documented that the fluorescence emission of dansyl dyes is quenched by adjacent maleimides *via* photoinduced electron transfer (PET), which proceeds from the excited state of the fluorophor to the maleimide.¹⁷ After conjugation of thiols to the electron-deficient double bond of the maleimide π -system of **Mal-C₁₆**, this PET process is interrupted resulting in a dansyl fluorescence quantum yield enhancement. (ii) Furthermore, by the altered physico-chemical properties of the local environment upon surface functionalization, the optical properties of the membrane-embedded fluorophors change causing an increase in emission intensity. A fluorescence increase of **V-Dans** during the surface reaction with a non-tryptophan-containing thiol like **ETG** (ethyl thioglycolate, Figure 3) confirms this interpretation (see Figure S3, ESI). The observations clearly show that the thiol is covalently attached to the membrane-embedded maleimide **Mal-C₁₆** and the membrane-bound thiols in turn are in close proximity to the co-embedded fluorophors. Membranes without **Mal-C₁₆** show a much higher initial dansyl emission intensity and no time-dependent change of the emission signal of either fluorophors, the tryptophan or dansyl dye, occurred after addition of **Cys-Trp** (see Figure S5, ESI). The same observation was

made for vesicles **V-Dans** whose surface-accessible maleimides were saturated with **ETG** prior to the addition of **Cys-Trp** (see Figure S4, ESI). The experiments demonstrate that non-functionalized, maleimide-containing vesicles **V-Dans** can also be regarded as simple self-assembled sensors for thiols.¹⁸

In contrast to dansyl dyes, carboxyfluorescein derivatives do not show PET processes in presence of maleimides in the membrane. To further study the effect (ii), we prepared vesicles **V-CF** with embedded carboxyfluorescein **CF-C₁₂**, which showed a time-dependent increase of the fluorescence signal after addition of thiol **ETG** indicating membrane restructuring processes upon surface functionalization (see Figure S6, ESI). By non-linear curve fitting, the second-order rate constants of thiol–maleimide conjugations on the vesicle membranes were determined (Table 1). The rate constants are all in the same order of magnitude. Small deviations along with different combinations of employed thiol and membrane composition can be explained by the course of the thiol–maleimide addition *via* thiolate anions. Their concentration is directly related to the pH of the reaction medium and to the pK_a value of the corresponding thiol. These pK_a values in turn are influenced by both the chemical nature of the thiol and the physico-chemical properties in their environment, *i.e.*, by the individual properties of the particular vesicle–water interface.¹⁹ Noteworthy, considerably basic conditions drastically enhance the nucleophilicity, however, also accelerate the oxidation of thiols to disulfides. Therefore, the pH value of 7.4 in our vesicular systems poses a good compromise between reaction rate and thiol stability. Compared to literature known rate constants from homogeneous systems ($600\text{--}8850\text{ M}^{-1}\text{ s}^{-1}$ at pH = 7.0),²⁰ the lower rate constants of about one order of magnitude at the liposomal surfaces are probably due to the lower diffusion rates of vesicles.²¹

Table 1: Rate constants for vesicle functionalization reactions based on the signal changes of the co-embedded fluorophors.

Entry	Vesicles	Thiol	k_{obs} [$\text{M}^{-1}\text{ s}^{-1}$]
1	V-Dans	Cys-Trp	315
2	V-Dans	ETG	108
3	V-CF	ETG	455

2.2 Preparation of Chemosensors for Phosphates by Post-Functionalization of Vesicles with Zn-Cyclen₂

Next, we utilized the established conditions for the membrane attachment of thiolated species for the preparation of a simple chemosensor. As phosphates are ubiquitous in nature, they pose important targets for analytical devices. Recently, our group reported on the preparation of vesicular chemosensors for phosphates based on amphiphilic derivatives of bis-Zn²⁺-cyclen complexes.^{4,7a} Those receptors are known to bind to phosphates with nanomolar affinities.²² To demonstrate the proof of principle of our post-functionalization approach and to show its applicability for sensor preparation in analogy to our previous investigations, we synthesized the thiolated bis-Zn²⁺-cyclen receptor **Zn-Cyclen₂** (Figure 3) and immobilized it on maleimide-modified vesicles **V-CF**. For that purpose, we added a sub-stoichiometric amount of the thiol (0.8 equivalents)²³ with respect to the surface-accessible concentration of **Mal-C₁₆** in order to avoid the separation of excess **Zn-Cyclen₂** after vesicle functionalization. Fluorescence measurements revealed a remarkable initial drop of the emission signal (see Figure S7, ESI) upon addition of **Zn-Cyclen₂** followed by a further time-dependent decrease of the fluorescence intensity indicating the progress of the surface reaction (Figure 6).

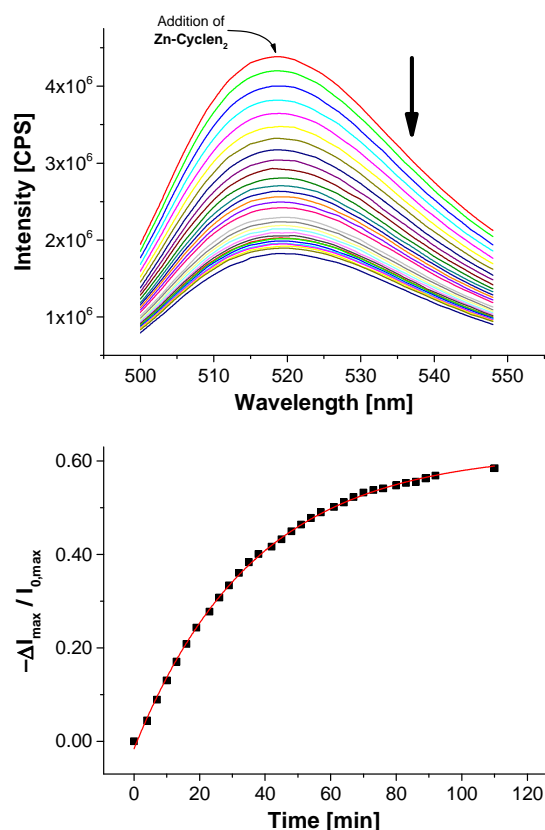


Figure 6: Time-dependent fluorescence response showing the functionalization progress of vesicles **V-CF** with 2.0 mol-% of **Zn-Cyclen₂**.

Compared to the surface reaction of vesicles **V-CF** with **ETG**, the functionalization with **Zn-Cyclen₂** was considerably slower (Table 2, entry 1). We explain this with the occurrence of multiple equilibria diminishing the concentration of free **Zn-Cyclen₂**: (i) The strong complexation abilities of thiol groups towards metal centers like zinc cations are supposed to result in intermolecular $\text{Zn}^{2+}\text{-SH}$ coordination.²⁴ (ii) Furthermore, the considerable instant quenching of the fluorescence signal after addition of the receptor indicates electrostatic interactions of the bis- Zn^{2+} -cyclen head groups with the negatively charged membrane-embedded carboxyfluorescein derivatives. To further verify these assumptions, we varied the amount of receptor **Zn-Cyclen₂** since the proposed equilibria are expected to be shifted towards the coordinated species at higher concentrations of **Zn-Cyclen₂** (entries 2 and 4). Additionally, we performed the surface reaction in the presence of pyrophosphate (PP_i , entries 3 and 5) due to its ability to compete for the Zn^{2+} -centered coordination site of the receptor and thus to increase the concentration of free **Zn-Cyclen₂** thiol functions. The results presented in Table 2 show two obvious trends which confirm our hypothesis: An enhancement of the reaction rate in the presence of PP_i and a deceleration of the reaction along with higher thiol concentrations.

Table 2: Rate constants for the functionalization of vesicles **V-CF** with different amounts of **Zn-Cyclen**₂ in absence or presence of **PP_i** (for fluorescence emission data, see Figures S7 and S8, ESI).

Entry	Zn-Cyclen ₂ ^a	PP_i ^b	<i>k_{obs}</i> [M ⁻¹ s ⁻¹]
1	0.8 eq (2.0 mol-%)	–	82
2	1.0 eq (2.5 mol-%)	–	78
3	1.0 eq (2.5 mol-%)	2.0 eq	322
4	2.0 eq (5.0 mol-%)	–	31
5	2.0 eq (5.0 mol-%)	2.0 eq	202

^a Equivalents with respect to surface-accessible amount of **Mal-C₁₆**.

^b Equivalents with respect to total amount of **Zn-Cyclen**₂.

The addition of disulfide precursor of **Zn-Cyclen**₂ (compound **7** in Scheme S2, ESI) to **V-CF** leads to a strong decrease of the emission signal supporting the proposed coordinative interaction of the Zn²⁺-complex with **CF-C₁₂**. However, as expected no further time-dependent decrease was observed due to the absence of free thiol groups required for the surface attachment (see Figure S9, ESI).

After completion of the functionalization process of **V-CF** with **Zn-Cyclen**₂, the obtained vesicles **V-CF-Zn** were ready for use as luminescent sensors for phosphate species. Titration of the analyte **PP_i** resulted in a strong increase of their fluorescence emission signal (Figure 7).

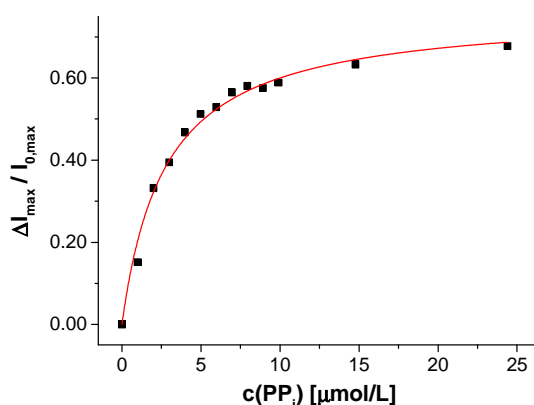


Figure 7: Fluorescence response upon titration of **PP_i** to functionalized vesicles **V-CF-Zn**.

For control experiments, non-functionalized vesicles were prepared by either omitting **Zn-Cyclen**₂ or by addition of **Zn-Cyclen**₂ to vesicles lacking the membrane-embedded attachment site **Mal-C₁₆**. Titration of **PP_i** did not show a significant response of the

fluorescence signal in those cases (for titration data, see Figure S10, ESI). These results proved that the membrane attachment of the receptor molecules is an essential requirement for the sensing mechanism of the PP_i analyte molecules.

We examined different phosphate-containing analytes in their binding affinities to vesicles **V-CF-Zn** and found binding constants which are in good agreement with literature values demonstrating the proof of concept (Table 3). The absence of a fluorescence emission change upon titration of sulfate as a dianionic control analyte demonstrated the specificity of **V-CF-Zn** towards phosphate anions.

Table 3: Binding affinities of **Zn-Cyclen**₂-functionalized vesicles **V-CF-Zn** to different analytes (for titration data, see Figure S11, ESI).

Entry	Analyte	$\lg K_a$	Reference ^a
1	PP_i	6.3	6.2 ^{7a}
2	D-Fructose 1,6 bisphosphate	6.0	6.1 ⁴
3	Phosphorylated hexapeptide Pep-P-His (Figure 8)	5.6	5.9 ^{26a}
4	SO_4^{2-}	no response	–

^a Refers to vesicular sensor systems obtained by co-embedding of amphiphilic bis- Zn^{2+} -cyclen receptors into the membrane.

Next, we investigated the dependency of the fluorescence response on the concentration of membrane-embedded fluorophors and immobilized receptor molecules (see Figure S12, ESI). Best signal to noise ratios were obtained with increasing amounts of both dye and receptor. These results are consistent with our previous observations and support our mechanistic hypothesis of receptor–dye patches on the vesicle surface, which reorganize upon formation of receptor–analyte complexes.^{7a}

2.3 Molecular Imprinting of Vesicle Surfaces

Our investigations with the **Zn-Cyclen**₂ receptor revealed that the post-functionalization technique is well suited for the preparation of luminescent chemosensors in a modular fashion. Table 3 shows that the obtained binding constants of the different phosphates are almost equal. However, for the development of powerful analytical devices

being able to discriminate between structurally related analytes, well-defined multipoint interactions between hosts and guests are essential. That means, the arrangement of the recognition elements on the vesicle surface must be complementary to the position of the binding units at the envisaged targets. Molecular imprinting is the classic technique in polymer science to achieve this aim.²⁵ In this connection, a molecule is utilized as a stamp to create its negative print in a suitable matrix. Recently, our group reported on two different strategies towards the preparation of highly specific, multi-receptor chemosensors. Both approaches are based on dynamic, template-guided recruitment of amphiphilic receptors on fluid vesicle membranes followed by fixation of the assemblies either by cooling the membrane down below the vesicular phase transition temperature^{26a,b} or by photopolymerization of the membrane-embedded receptor arrays.^{26c} A post-functionalization strategy can also be used for the molecular imprinting of defined two-dimensional receptor assemblies since the maleimide-decorated vesicles can be regarded as reactive matrices for template-assisted covalent functionalization. A fundamental prerequisite for that approach is the rigidity of the vesicle membrane preventing the lateral redistribution of the receptors after surface attachment. Hence, DSPC membranes with a phase transition temperature of 54 °C²⁷ represent an ideal scaffold for that purpose. Figure 8 illustrates the principle of molecular imprinting by vesicle post-functionalization. In detail, membrane-attached receptor–template complexes direct the surface fixation of additional receptor molecules by coordinative interactions in spatially defined arrangements. Subsequent separation of the templates leads to an imprinted vesicle surface with patterns of binding sites.

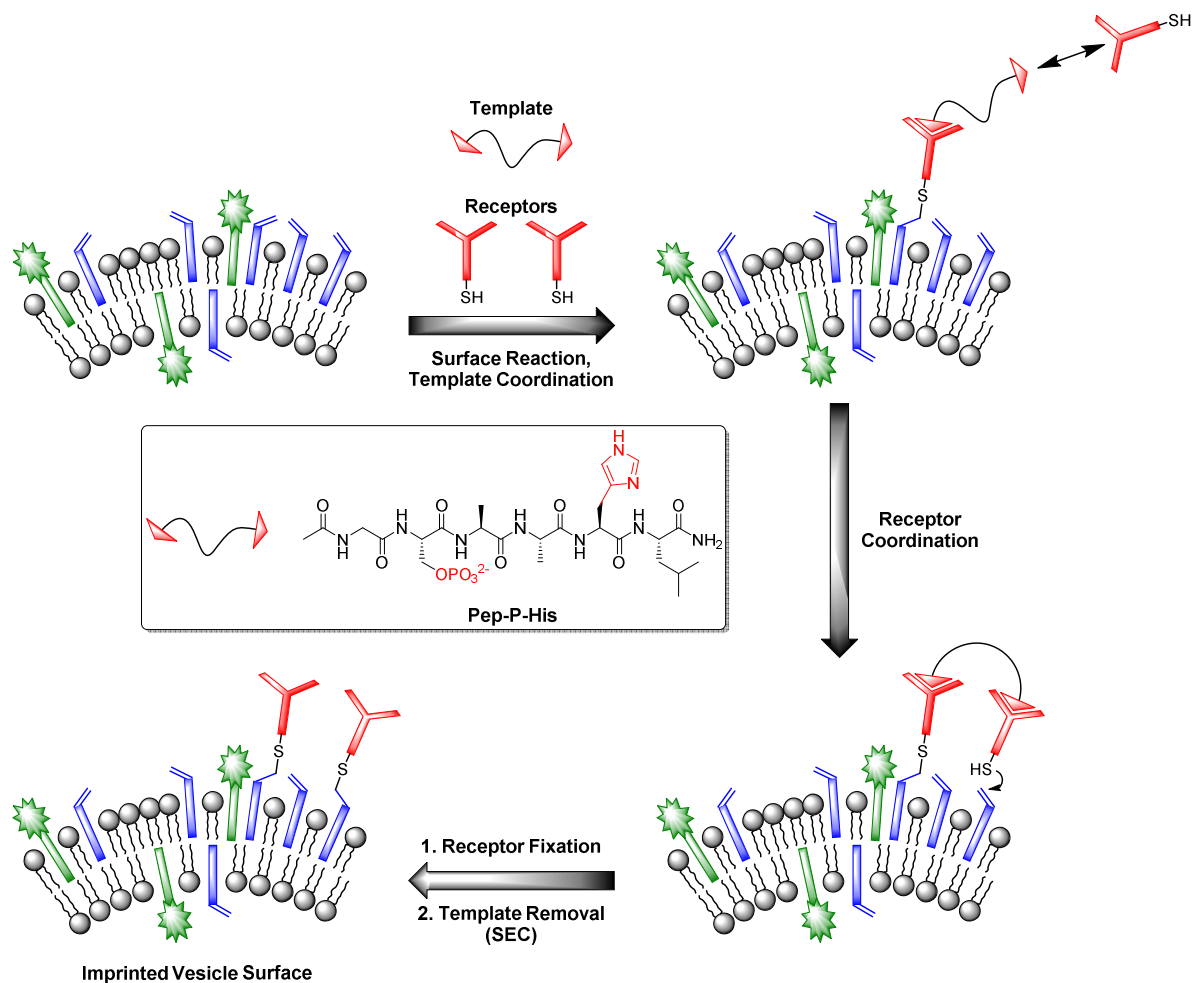


Figure 8: Principle of vesicle surface imprinting by template-assisted post-functionalization and bivalent peptide **Pep-P-His** used as model template.

Complex **Zn-Cyclen₂** is known to bind besides phosphates also to histidine moieties.²⁸ As template for our imprinting studies, we therefore utilized bivalent hexapeptide **Pep-P-His** (Figure 8) being able to form a ternary complex with two receptor molecules. We incubated a 1:2 mixture of the peptide and the thiolated receptor **Zn-Cyclen₂** with vesicles **V-CF** (0.4 equivalents of receptor with respect to surface-accessible maleimide) and removed the template molecules by size-exclusion chromatography (SEC) after completion of the surface functionalization. For comparison, vesicle surfaces were post-functionalized with **Zn-Cyclen₂** in absence of the bivalent peptide, but treated identically apart from that. Rebinding studies of the two obtained vesicle samples revealed a significantly different course of the fluorescence emission responses (Figure 9). While the imprinted vesicles displayed a fluorescence decrease upon titration of up to 0.5 equivalents of peptide **Pep-P-His**, the non-imprinted samples showed a steady increase. After further peptide addition, the sign of the fluorescence signal

change reversed in case of the imprinted vesicles and the curve progression approximated that of the non-imprinted ones.

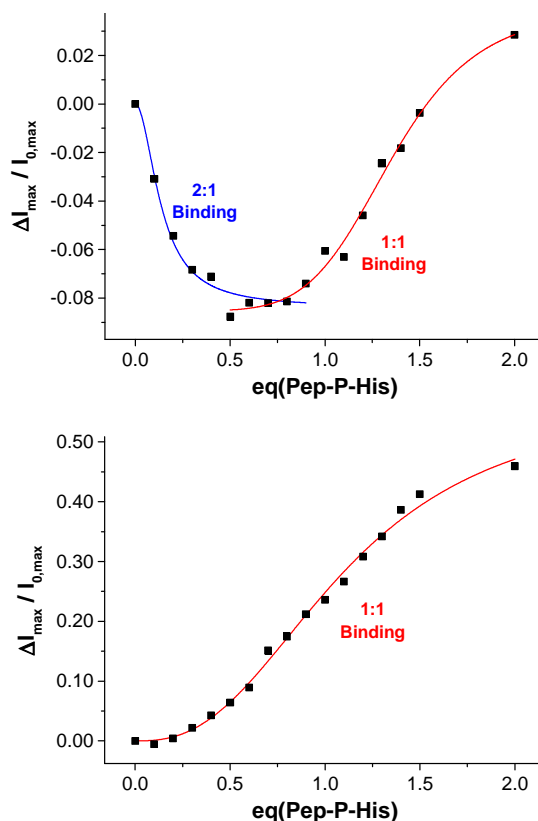


Figure 9: Rebinding isotherms of imprinted (top) and non-imprinted (bottom) vesicles **V-CF-Zn** upon addition of **Pep-P-His**.

This divergent behavior can be rationalized by different binding modes in the two cases. We hypothesize that the geometric confinement of the functional amphiphiles on the vesicle surface upon the formation of 2:1 complexes results in a further quenching of the co-embedded fluorophors. Gradual transition into 1:1 binding mode after addition of more than 0.5 equivalents of the peptide is assumed to provide a higher translational freedom of the membrane-embedded compounds leading to an emission intensity increase of the dye molecules.²⁹ Due to the superposition of two opposing effects in the case of imprinted vesicles, the overall fluorescence response is much lower compared to that of the non-imprinted samples. The macroscopic binding constants in the two cases obtained by Hill fitting of the titration curves are $\lg K_a = 7.1$ for the imprinted vesicles (based on the addition of up to 0.8 equivalents of **Pep-P-His**) *versus* 5.1 for the control experiment.³⁰ However, due to the superposition of two diverging signal responses, the apparent affinity constant for the multipoint binding is likely to be underestimated.

Variation of the receptor concentration indicated that the success of vesicle surface patterning is dependent on the proportion of added receptor molecules and membrane-embedded maleimides. At an almost equimolar ratio (≈ 0.8 equivalents of receptor with respect to surface-accessible maleimide), the different binding behavior of imprinted and non-imprinted samples is much less pronounced (see Figure S13, ESI). We explain this result with the fact that here, approximately all surface-accessible maleimide molecules bear a receptor unit, *i.e.*, the receptor assembly simply reflects the random assembly of the membrane-embedded **Mal-C**₁₆. Thus, no external control over their spatial distribution is possible and the receptors cannot attain an appropriate geometric arrangement for multipoint binding to the analyte. Conversely, when immobilizing the receptor in sub-stoichiometric amounts, only membrane-embedded maleimides whose mutual distances are compatible to the formation of ternary receptor–template complexes are functionalized. The other maleimides remain unmodified and thereby do not impede the 2:1 binding on the vesicle surface.

2.4 Vesicle Functionalization with an Aptamer for Ampicillin

In living systems, multipoint interactions dominate the molecular recognition phenomena and provide for remarkably high specificities. Thus, to explore the full potential of our technique, we next extended the vesicular post-functionalization strategy to more complex binding units of biological origin. Among those, oligonucleotide-based aptamers represent one of the most exciting and promising class of receptors. Since they are developed by an evolutionary process of repeated selection and amplification steps, aptamers exhibit very high binding affinities and specificities to their targets ranging from small organics to proteins and pathogens.³¹ In many cases, structurally related analytes can be discriminated on basis of subtle differences such as the presence or absence of methyl or hydroxyl groups. Compared to other biological recognition elements like antibodies, aptamers display some remarkable advantages including low immunogenicity, small size, easy production and modification by established solid-phase synthesis methods and therefore inexpensive commercial availability. Moreover, they are characterized by a high stability and convenient handling. Fluorescence-based methods for the detection of aptamer–target binding typically rely on covalent modification of the oligonucleotides with luminescent reporter moieties.³² Another established technique involves hybridization of the aptamers with accordingly labeled complementary oligonucleotide strands and their subsequent displacement by the

analyte molecules. This requirement of covalent attachment of fluorophors can make such assays laborious and costly. Label-free strategies on the other hand, *e.g.*, based on intercalation of a fluorescent dye into a double helix domain, are often limited in their applicability as they depend on particular properties of the aptamers such as specific folding modes upon complex formation with the target. In contrast to these analytical methods, the signaling mechanism of our vesicular sensors is generally supposed to originate rather from an analyte-induced redistribution of charges at the liposome–water interface than on defined conformational changes of the recognition sites. Therefore, aptamer-functionalized vesicles with co-embedded reporter dyes potentially represent versatile systems for the detection of aptamer–target interactions. The modular nature of such aptasensors based on self-assembly and subsequent membrane modification allows for their easy preparation and facile optimization of their composition. Recently, we reported on the post-functionalization of luminescent vesicles with the thrombin-binding aptamer resulting in liposomal sensors, which signal the binding of the protein analyte to their surface.¹⁴ To demonstrate the universal applicability of this approach for the preparation of various aptasensors, we utilized a DNA-based aptamer developed by Song *et al.* which specifically binds to the antibiotic ampicillin (Figure 10) with an affinity constant of $\lg K_a = 8.0$.³³

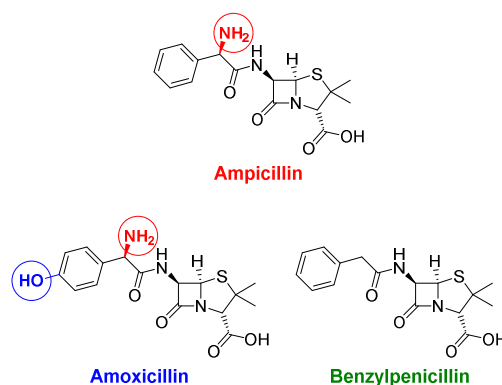


Figure 10: Structures of the antibiotic ampicillin and of related control antibiotics amoxicillin and benzylpenicillin; structural differences are marked with colored circles.

For the selection of the ampicillin aptamer, the authors used TRIS buffer with added salts (50 mM NaCl, 5 mM KCl, 5 mM MgCl₂). The presence of cations such as K⁺ or Mg²⁺ is important to stabilize the folded analyte binding conformations of some aptamers. On the other hand, it is well explored that in particular bivalent cations can induce aggregation or even fusion of negatively charged vesicles (as it is the case for **V-CF**) by neutralizing their surface charge and thus by decreasing their electrostatic repulsion barrier.³⁴ In our case, it

turned out that non-functionalized vesicles **V-CF** are indeed unstable and aggregate under the original buffer conditions. For that reason, we screened different buffer systems with respect to both vesicle stability and emission response of aptamer-functionalized vesicles after addition of ampicillin (see ESI for details). Buffer systems without added salts did not result in significant fluorescence responses, whereas very high salt concentrations induced vesicle aggregation as determined by dynamic light scattering. HEPES buffer with added KCl (5 mM) and MgCl₂ (5 mM) represented the best compromise. The surface functionalization of maleimide-containing vesicles **V-CF** was performed with thiol-modified oligonucleotide **Apt-Amp** (Figure 3). A time-dependent change of the emission signal again displayed the progress of the aptamer-immobilization on the vesicular membrane (Figure 11).

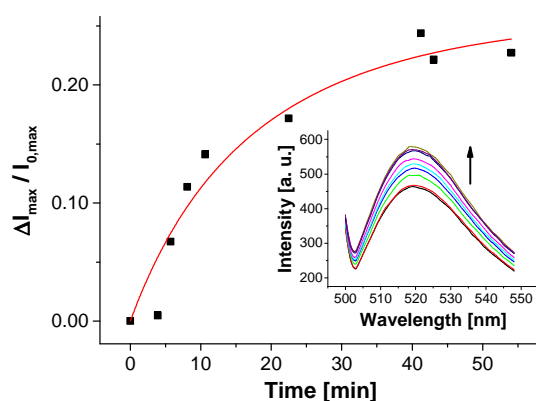


Figure 11: Fluorescence response showing the immobilization progress of vesicles **V-CF** with aptamer **Apt-Amp** (1.0 mol-%).

In contrast to the post-functionalization of **V-CF** with receptor **Zn-Cyclen₂**, the fluorescence signal increased during the surface reaction. We believe that the presence of the highly negatively charged oligonucleotides at the vesicle surface partially displaces the likewise negatively charged carboxyfluorescein molecules from the clusters of the functional amphiphiles. As a consequence, a reduction of the fluorophore self-quenching occurs. Binding of ampicillin to the surface-immobilized aptamers is thought to induce a more compact conformation of the aptamer strands. Therefore, a higher negative charge density at the vesicle membrane is generated, which in turn enforces more dye molecules to be squeezed out of their membrane domains. This process gives rise to a significant further increase of the emission signal (Figure 12).³⁵

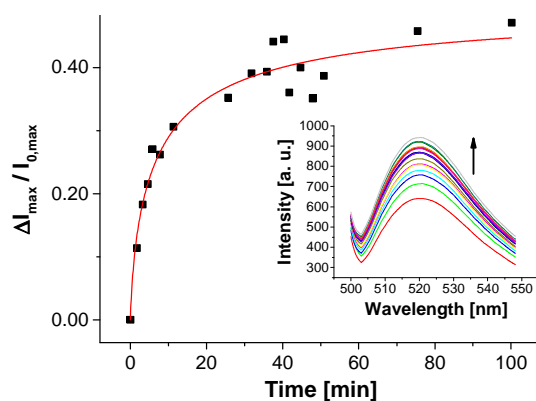


Figure 12: Time-dependent change of fluorescence emission of aptamer-functionalized vesicles **V-CF-Amp** (2.0 mol-% of **Apt-Amp**) after addition of ampicillin (0.40 μM).

The time-dependent fluorescence spectra depicted in Figure 12 show a rather slow emission response of **V-CF-Amp** upon addition of ampicillin. We assume that the aptamer-functionalized vesicles are more prone to salt-induced aggregation than the non-functionalized ones due to their higher surface charge.³⁴ This assumption is supported by investigations of the vesicular size distribution, which revealed a significant cross linking of the vesicles after functionalization with the oligonucleotide (see Figure S15, ESI).³⁶ This process in turn reduces the accessibility of the surface-bound aptamers for ampicillin. Binding of the analyte molecules to their membrane receptors, however, seems to partially reverse the aggregation as indicated by a subsequent decrease of the size distribution maximum of the vesicles.³⁷

Variation of the aptamer concentration and addition of ampicillin to the functionalized vesicles gave best emission responses of almost 50 % with 2.0 mol-% of oligonucleotides immobilized on the surface (Figure 13). These conditions having approximately equimolar amounts of the receptor molecules with respect to the membrane-embedded fluorophors on the outer vesicle provide highest sensitivities, which is in accordance with our results from **Zn-Cyclen**₂-functionalized vesicles.

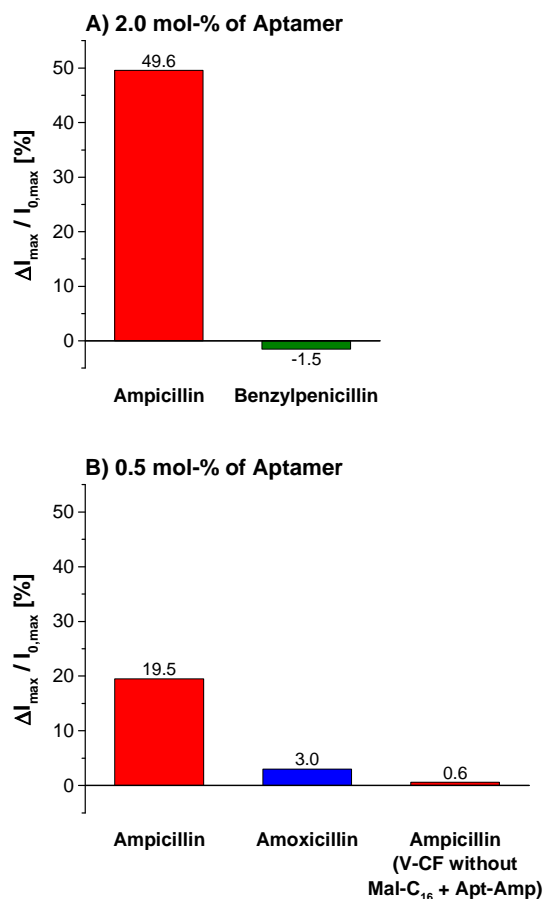


Figure 13: Comparison of relative fluorescence emission responses of **V-CF-Amp**, functionalized with different amounts of the aptamer, and of a mixture of maleimide-lacking vesicles **V-CF** and **Apt-Amp** after addition of ampicillin, benzylpenicillin or amoxicillin (0.40 μM in each case).

The specificity of the aptasensors **V-CF-Amp** towards ampicillin was illustrated by addition of the structurally related antibiotics benzylpenicillin or amoxicillin, which showed no significant fluorescence responses of the membrane-embedded carboxyfluorescein dyes (Figure 13). Vesicles lacking the amphiphilic attachment site for thiols **Mal-C₁₆** were prepared and incubated with the aptamer. Addition of ampicillin to that sample did not lead to a change of the emission. This observation proves that the analyte signaling depends on the interactions with the membrane-conjugated aptamers.

3. Conclusion

In summary, we have demonstrated that preformed, maleimide-decorated luminescent vesicles can be easily functionalized at the surface with different receptors by nucleophilic thiol addition. It was shown that the immobilized compounds may range from small organic molecules to peptides and metal complexes, such as bis-Zn²⁺-cyclen, and to even more

complex structures like oligonucleotides. The membrane-embedded fluorophores respond to the presence of the receptor units and to the formation of host–guest complexes on the vesicle surface with changing emission properties. We demonstrated potential applications of the concept including the detection of simple phosphates with surface-attached **Zn-Cyclen**₂ or the specific sensing of ampicillin by an aptamer. Moreover, we showed that the post-functionalization strategy allows external control over the two-dimensional distribution of the receptors by assistance of multivalent templates. Here, significant differences between imprinted and non-imprinted samples were obtained in the fluorescence output of rebinding titrations. Due to the highly modular approach, our presented strategy allows an easy, fast and inexpensive access to a broad variety of supramolecular indicators with tailor-made properties. The non-functionalized vesicles can be prepared in advance. Since many heterobifunctional reagents for the *in situ* introduction of thiol groups to amines are commercially available (*e.g.*, Traut's reagent^{38a} or SATA^{38b}), our method is not only limited to thiol-containing host molecules, but can be readily extended to recognition elements based on other peptides, oligonucleotides or complex macromolecules. Hence, the development of novel customized bioanalytical devices may be facilitated.

Acknowledgements

A.M. thanks the Bayerische Eliteförderung for a research fellowship.

4. Notes and References

† Electronic Supplementary Information (ESI) available: Experimental details, synthesis of compounds, preparation of functionalized vesicles, emission spectra, binding curves and optimization studies.

- 1 (a) J. Voskuhl and B. J. Ravoo, *Chem. Soc. Rev.*, 2009, **38**, 495–505; (b) K. Ariga, H. Ito, J. P. Hill and H. Tsukube, *Chem. Soc. Rev.*, 2012, **41**, 5800–5835.
- 2 (a) K. A. Edwards and A. J. Baeumner, *Talanta*, 2006, **68**, 1421–1431; (b) Q. Liu and B. J. Boyd, *Analyst*, 2013, **138**, 391–409.
- 3 R. Martínez-Mañez and F. Sancenón, *Chem. Rev.*, 2003, **103**, 4419–4476.
- 4 B. Gruber, S. Stadlbauer, K. Woinaroschy and B. König, *Org. Biomol. Chem.*, 2010, **8**, 3704–3714.
- 5 T. W. Bell and N. M. Hext, *Chem. Soc. Rev.*, 2004, **33**, 589–598.
- 6 B. T. Nguyen and E. V. Anslyn, *Coord. Chem. Rev.*, 2006, **250**, 3118–3127.
- 7 (a) B. Gruber, S. Stadlbauer, A. Späth, S. Weiss, M. Kalinina and B. König, *Angew. Chem. Int. Ed.*, 2010, **49**, 7125–7128; *Angew. Chem.*, 2010, **122**, 7280–7284; (b) S. Banerjee, M. Bhuyan and B. König, *Chem. Commun.*, 2013, **49**, 5681–5683.
- 8 (a) M. Mammen, S.-K. Choi and G. M. Whitesides, *Angew. Chem. Int. Ed.*, 1998, **37**, 2754–2794; *Angew. Chem.*, 1998, **110**, 2908–2953; (b) H. Jung, A. D. Robison and P. S. Cremer, *J. Struct. Biol.*, 2009, **168**, 90–94; (c) A. Grochmal, E. Ferrero, L. Milanesi and S. Tomas, *J. Am. Chem. Soc.*, 2013, **135**, 10172–10177.
- 9 (a) L. Nobs, F. Buchegger, R. Gurny and E. Allémann, *J. Pharm. Sci.*, 2004, **93**, 1980–1992; (b) V. P. Torchilin, *Nat. Rev. Drug Discovery*, 2005, **4**, 145–160; (c) F. S. Hassane, B. Frisch and F. Schubert, *Bioconjugate Chem.*, 2006, **17**, 849–854; (d) R. I. Jølcck, L. N. Feldborg, S. Andersen, S. M. Moghimi and T. L. Andresen, *Adv. Biochem. Engin./Biotechnol.*, 2011, **125**, 251–280.
- 10 (a) C. E. Hoyle and C. N. Bowman, *Angew. Chem. Int. Ed.*, 2010, **49**, 1540–1573; *Angew. Chem.*, 2010, **122**, 1584–1617; (b) C. E. Hoyle, A. B. Lowe and C. N. Bowman, *Chem. Soc. Rev.*, 2010, **39**, 1355–1387.
- 11 V. D. Bock, H. Hiemstra and J. H. van Maarseveen, *Eur. J. Org. Chem.*, 2006, 51–68.
- 12 C. E. Hoyle, T. Y. Lee and T. Roper, *J. Polym. Sci., Part A: Polym. Chem.*, 2004, **42**, 5301–5338.
- 13 (a) B. Frisch, C. Boeckler and F. Schubert, *Bioconjugate Chem.*, 1996, **7**, 180–186.; (b) P. Schelte, C. Boeckler, B. Frisch and F. Schubert, *Bioconjugate Chem.*, 2000, **11**, 118–123; (c) R. J. Pounder, M. J. Stanford, P. Brooks, S. P. Richards and A. P. Dove, *Chem. Commun.*, 2008, 5158–5160.
- 14 A. Müller and B. König, *Chem. Commun.*, 2014, **50**, 12665–12668.
- 15 Y. Kitano, Y. Nogata, K. Matsumura, E. Yoshimura, K. Chiba, M. Tada and I. Sakaguchi, *Tetrahedron*, 2005, **61**, 9969–9973.
- 16 P. Wu and L. Brand, *Anal. Biochem.*, 1994, **218**, 1–13.
- 17 J. Guy, K. Caron, S. Dufresne, S. W. Michnick, W. G. Skene and J. W. Keillor, *J. Am. Chem. Soc.*, 2007, **129**, 11969–11977.

- 18 For examples of optical probes for thiol detection, see: X. Chen, Y. Zhou, X. Peng and J. Yoon, *Chem. Soc. Rev.*, 2010, **39**, 2120–2135.
- 19 M. P. Lutolf, N. Tirelli, S. Cerritelli, L. Cavalli and J. A. Hubbell, *Bioconjugate Chem.*, 2001, **12**, 1051–1056.
- 20 J. Li, Q. Xu, D. M. Cortes, E. Perozo, A. Laskey and A. Karlin, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 11605–11610.
- 21 P. Schelté, C. Boeckler, B. Frisch and F. Schuber, *Bioconjugate Chem.*, 2000, **11**, 118–123.
- 22 (a) T. Koike, S. Kajitani, I. Nakamura, E. Kimura and M. Shiro, *J. Am. Chem. Soc.*, 1995, **117**, 1210–1219; (b) E. Kimura, S. Aoki, T. Koike and M. Shiro, *J. Am. Chem. Soc.*, 1997, **119**, 3068–3076; (c) S. Aoki, M. Zulkefeli, M. Shiro, M. Kohsako, K. Takeda and E. Kimura, *J. Am. Chem. Soc.*, 2005, **127**, 9129–9139.
- 23 With respect to a total concentration of **Mal-C₁₆** of 5.0 mol-% and therefore a surface-accessible concentration of approximately 2.5 mol-%, 0.8 equivalents of **Zn-Cyclen₂** equate to 2.0 mol-% relating to the total amphiphile concentration.
- 24 S. Lacerda, M. P. Campello, I. C. Santos, I. Santos and R. Delgado, *Polyhedron*, 2007, **26**, 3763–3773.
- 25 G. Wulff, *Angew. Chem., Int. Ed. Engl.*, 1995, **34**, 1812–1832; *Angew. Chem.*, 1995, **107**, 1958–1979.
- 26 (a) B. Gruber, S. Balk, S. Stadlbauer and B. König, *Angew. Chem. Int. Ed.*, 2012, **51**, 10060–10063; *Angew. Chem.*, 2012, **124**, 10207–10210; (b) S. Balk and B. König, *J. Inclusion Phenom. Macrocyclic Chem.*, 2014, DOI: 10.1007/s10847-014-0442-2; (c) S. Banerjee and B. König, *J. Am. Chem. Soc.*, 2013, **135**, 2967–2970.
- 27 M. Ueno, S. Katoh, S. Kobayashi, E. Tomoyama, S. Ohsawa, N. Koyama and Y. Morita, *J. Colloid Interface Sci.*, 1990, **134**, 589–592.
- 28 A. Grauer, A. Riechers, S. Ritter and B. König, *Chem. Eur. J.*, 2008, **14**, 8922–8927.
- 29 In our previously reported dynamic imprinting approaches,²⁶ we observed a steady increase of the signal in both cases, however, with the different binding affinities being reflected in different slopes of the binding isotherms. We explain this with the dynamic nature of the receptor recruitment process in that case providing a diverse arrangement of the fluorophor molecules relative to the receptor assemblies.
- 30 The lower value of $\lg K_a$ in case of non-imprinted vesicles compared to that from Table 3 might be attributed to an incomplete separation of the template peptide by the SEC in that case. However, as the exactly identical conditions for SEC were applied for both the imprinted vesicles and the non-imprinted control solutions, the relative change of the binding affinities has to be considered.
- 31 (a) S. D. Jayasena, *Clin. Chem.*, 1999, **45**, 1628–1650; (b) C. K. O’Sullivan, *Anal. Bioanal. Chem.*, 2002, **372**, 44–48.
- 32 (a) E. J. Cho, J.-W. Lee and A. D. Ellington, *Annu. Rev. Anal. Chem.*, 2009, **2**, 241–264; (b) R. E. Wang, Y. Zhang, J. Cai, W. Cai and T. Gao, *Curr. Med. Chem.*, 2011, **18**, 4175–4184.
- 33 K.-M. Song, E. Jeong, W. Jeon, M. Cho and C. Ban, *Anal. Bioanal. Chem.*, 2012, **402**, 2153–2161.
- 34 S. Nir, J. Bentz, J. Wilschut and N. Düzgünes, *Prog. Surf. Sci.*, 1983, **13**, 1–124.

- 35 The surface-immobilized **Zn-Cyclen**₂ complexes, on the other hand, are supposed to electrostaticly interact with the membrane-embedded carboxyfluorescein molecules resulting in a partial quenching of their fluorescence emission.
- 36 Vesicle aggregation based on specific intermolecular interactions between surface-attached oligonucleotides is unlikely due to the absence of complementary sequences in the immobilized aptamer required for hybridization.
- 37 We did not face that aggregation behavior in our previously reported studies regarding the vesicular thrombin-aptasensor.¹⁴ There, the presence of Mg²⁺ was not required for the conformational stabilization of the oligonucleotides. As a result, a dynamic and quick fluorescence response over a wide concentration range was observed.
- 38 (a) R. Jue, J. M. Lambert, L. R. Pierce and R. R. Traut, *Biochemistry*, 1978, **17**, 5399–5406; (b) R. Julian, S. Duncan, P. D. Weston and R. Wrigglesworth, *Anal. Biochem.*, 1983, **132**, 68–73.