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Novel asymmetrical azines appending 1,3,4-thiadiazole sulfonamide: synthesis, molecular structure analyses, *in silico* ADME, and cytotoxic effect†

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Toward finding potential and novel anticancer agents, we designed and prepared novel differently substituted unsymmetrical azine-modified thiadiazole sulfonamide derivatives using the "combi-targeting approach". An efficient procedure for synthesizing the designed compounds starts with 5-acetyl-3-*N*-(4-sulfamoylphenyl)-2-imino-1,3,4-thiadi-azoline **4**. The *E/Z* configuration for compound **5** was investigated based on spectral analysis combined with quantum mechanical calculation applying the DFT-B3LYP method and 6-31G(d) basis set. The computational results found that the *E* isomer was energetically more favorable than the *Z* isomer by 2.21 kcal mol⁻¹. Moreover, ¹H and ¹³C chemical shifts for the *E* and *Z* isomers in DMSO were predicted using the GIAO-B3LYP/6-31G(d) computations and IEF-PCM solvation model. The computed chemical shifts for both isomers are consistent with those observed experimentally, indicating that they exist in the solution phase. Moreover, the *E/Z* configuration for the synthesized azines **7a–c**, **9**, **11**, **13**, **15a** and **15b** was also studied theoretically using the DFT-B3LYP/6-31G(d) calculations. *In silico* prediction for the biological activities was reported regarding the HOMO–LUMO energy gaps and molecular reactivity descriptors besides the ADMET/drug-likeness properties. The cytotoxic effect of the synthesized compounds has been assayed *via* the determination of their IC₅₀.

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1. Introduction

Current obstacles to treating cancer include the emergence of drug resistance and unfavorable off-target effects of anticancer drugs, which energizes medicinal chemists to continuously produce novel anticancer medications with high efficacy and low toxicity.^{1,2}

One of the most promising candidates in the field of synthetic drugs is sulphonamides (Fig. 1a).^{3–6} The thiadiazole platform constitutes intriguing and rapidly expanding sulfonamide derivative systems. Several medicines on the market are related to this system, making it a flexible tool for drug design (Fig. 1b).^{7,8} According to a literature survey, azines and derivatives serve as crucial structural components in various versatile scaffolds with a wide range of drug applications (Fig. 1c).⁹

Azines have recently gained attention for configurations, and tautomers that profoundly affect biochemical processes.¹⁰

Recently, a vital approach/strategy in drug discovery is the amalgamation of two or more complete medications into a single molecular structure, known as a combi-molecule, which may be a good solution to achieve bioactive molecules, with high potency and different mechanisms of action, due to the synergistic effect.¹¹ Hence, cohesive systems incorporating 1,3,4-thiadiazole sulfonamide with azines may help design new anticancer hybrids to improve biological properties.

To investigate the application of a combi-molecule strategy, we designed and prepared novel compounds **7a–c**, **9**, **11**, **13** and **15a–b** in this work by linking azines fragments and thiadiazole sulfonamides. These compounds were then subjected to cytotoxic assays on three cancerous cell lines. Their cytotoxic assay was comparable to the positive control staurosporine in the low micromolar region.

Moreover, we employ quantum mechanical (QM) computations to provide theoretical analyses for compound **5**, including the *E/Z* configurations and conformational study toward free rotatable single bonds. Thus, QM calculations were reported using the density functional theory (DFT) at the level of B3LYP¹² and 6-31G(d) basis set. Owing to its reliable accuracy and reduced computation time, the B3LYP/6-31G(d) calculation was

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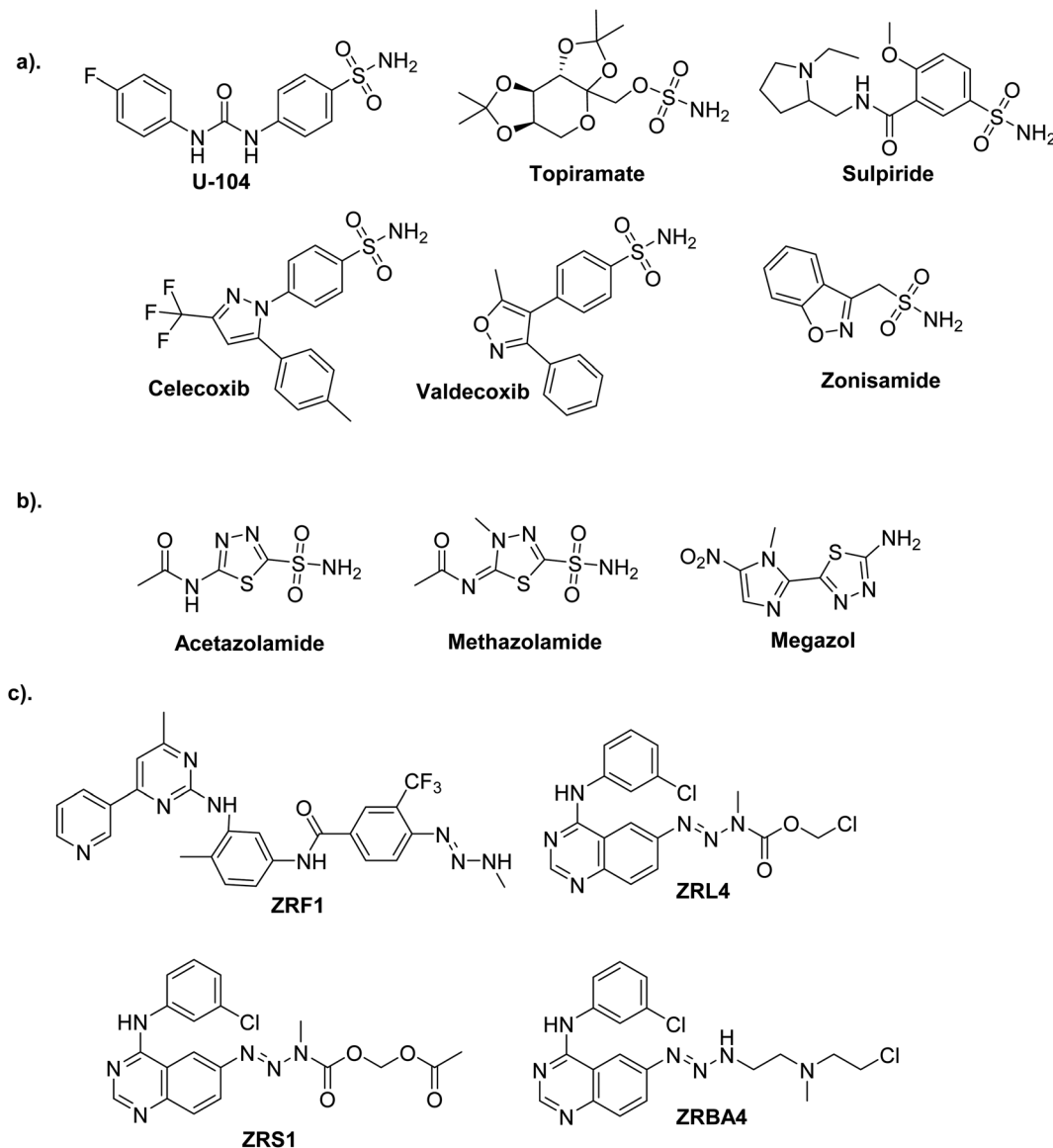



Fig. 1 Commercial drugs on the market contain bioactive cores. (a) Sulfonamides. (b) Thiadiazole sulfonamides. (c) Asymmetrical azines.

widely recognized and applied for theoretical studies of organic molecules of medium-large size.^{13,14} The ^1H and ^{13}C NMR chemical shifts (δ , ppm) were also computed by applying the approach of gauge-invariant atomic orbitals, GIAOs,¹⁵ to investigate the *E/Z* configuration of **5** in the solution phase. *In silico* techniques have been widely applied to drug screening.^{16–19} Various computational tools and methods may be used to identify the candidate drug from other compounds depending on multiple features such as physicochemical/pharmacokinetic parameters and drug-likeness. Herein, QM calculations were carried out for the synthesized compounds to correlate their structures with biological activities *via* analysis of the FMOs and quantum chemical descriptors. To assess the synthesized compounds **7a–c**, **9**, **11**, **13**, **15a** and **15b** as drug candidates, the SwissADME²⁰ and pkCSM²¹ servers were used to predict

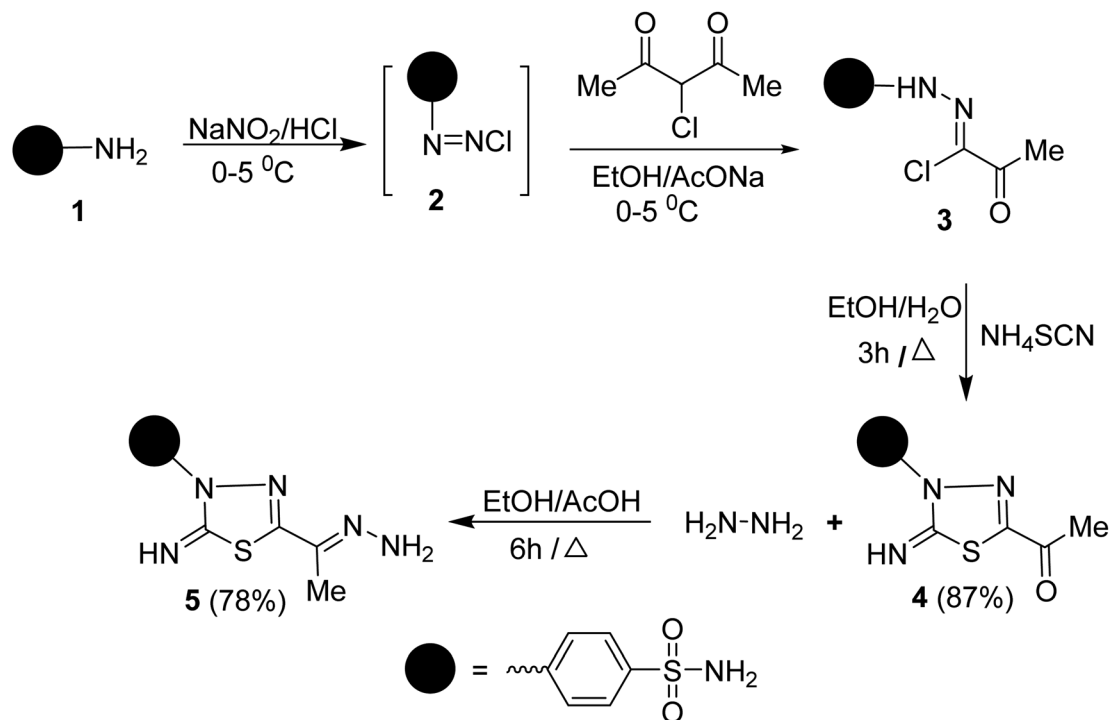
physicochemical characteristics, drug-likeness, and ADMET properties.

2. Results and discussion

2.1. Chemistry

To synthesize the target compounds **7a–c**, **9**, **11**, **13**, **15a** and **15b** the synthetic sequence starts with the preparation of 5-acetyl-3-*N*-(4-sulfamoylphenyl)-2-imino-1,3,4-thiadiazoline **4** as commencing material. 1,3,4-Thiadiazoline **4** was efficiently prepared *via* a cyclization reaction of freshly synthesized 2-oxo-*N*-(4-sulfamoylphenyl)propanehydrazonoyl chloride **3**, obtained from the Japp–Klingemann reaction of 3-chloro-2,4-pentanedione with diazonium chloride of sulfanilamide **2** in a buffered ethanolic solution, with an aqueous ethanolic solution of ammonium thiocyanate under reflux (Scheme 1). Compound **4**'s structure was determined *via* microanalysis and





Scheme 1 Synthesis of compound 5.

spectral data. In the IR spectrum, three bands were observed at 3352, 3289, and 3277 cm^{-1} , indicating the existence of primary and secondary N-H stretching vibration, respectively. The bands at 1692, 1330, and 1298 cm^{-1} indicated the existence of a carbonyl group ($\text{C}=\text{O}$) and sulfonamide group (SO_2NH_2), respectively. The ^1H -NMR spectrum displayed three singlet signals at δ 2.50, 7.45, and 9.62 ppm, characteristic for CH_3 , SONH_2 , and NH protons, respectively, as well as two doublet signals, resonating at δ 7.92 and 8.18 ppm with the identical coupling constant value ($J = 9.35$ Hz) and integrating for four protons indicating the existence of 4-disubstituted benzene. Its ^{13}C -NMR spectrum showed the presence of eight signals which agrees with its molecular structure. The signals of CH_3 and a carbonyl carbon resonate at 24.88 and 189.83 ppm, respectively. In the mass spectrum (MS), the molecular ion peak (M^+) for 4 was found at $m/z = 298$, which is compatible with its molecular formula ($\text{C}_{10}\text{H}_{10}\text{N}_4\text{O}_3\text{S}_2$).

Condensation of 5-acetyl-3-*N*-(4-sulfamoylphenyl)-2-imino-1,3,4-thiadiazoline 4 with hydrazine afford the respective 4-(5-(1-hydrazonoethyl)-2-imino-1,3,4-thiadiazol-3(2*H*)-yl)benzenesulfonamide 5 (Scheme 1). The structure of later hydrazone 5 was confirmed through spectroscopic analyses. An examination of its IR spectrum revealed the lack of a carbonyl absorption band and the existence of azomethine ($\text{C}=\text{N}$) and amino groups at wavenumbers 1645, 3416, and 3285 cm^{-1} . Interestingly, the ^1H -NMR spectrum of hydrazone 5 showed two sets of resonances that supported the presence of 5 in two isomeric forms. The separation of signals of aromatic, $-\text{NH}_2$, and CH_3 protons in the two isomers is well resolved. The population ratio of the major and minor isomers is (52 : 48). Based on previous studies,

the most stable and the major isomer is assigned to the *E*-isomer around the $\text{C}=\text{N}$ bond.²² That is supported by the observed chemical shift value of the hydrazone- NH_2 protons in *E*- and *Z*-isomers. In *E*-isomer, the hydrazone- NH_2 protons appear as a singlet signal at δ 5.66 ppm. In comparison, *Z*-isomer resonates as two separate singlet signals at 7.33 and 7.32 ppm due to the possible formation of intramolecular H-bonds between N-H proton and $\text{C}=\text{N}$ group in the thiadiazole ring as shown in Fig. 2. According to MS, the M^+ at m/z is the molecular weight.

To synthesize unsymmetrical azines, we investigated the reactivity of the terminal $-\text{NH}_2$ group of hydrazone 5 towards aromatic aldehydes, heterocyclic aldehydes, and cyclic ketones (Scheme 2). Hence, condensation of compound 5 with aromatic aldehydes, namely, 4-methoxy benzaldehyde 6a, 4-(*N,N*-dimethylamino) benzaldehyde 6b, 4-nitrobenzaldehyde 6c and 2,4,6-trimethoxybenzaldehyde 8 in refluxing ethanol yielded unsymmetrical azines, 1-(4-substituted benzylidene)-2-(1-(2-imino-3-(4-sulfamoylphenyl)-1,3,4-thiadiazol-5-yl)ethylidene) hydrazine 7a-c and 1-(2,4,6-trimethoxy benzylidene)-2-(1-(2-imino-3-(4-sulfamoyl phenyl)-1,3,4-thiadiazol-5-yl)ethylidene)hydrazine 9, respectively (Scheme 2). The molecular structures of the later azines were secured from microanalyses and spectral data. As a representative example, the IR spectrum of azine 7a displayed a strong absorption band at 3360, 3255, 3110, and 1604 cm^{-1} due to NH_2 , NH, and $\text{C}=\text{N}$ functions, respectively. The ^1H -NMR spectrum of 7a exhibited five characteristic singlet signals at δ 2.49, 3.84, 7.49, 8.63, 9.42 ppm corresponding to CH_3 , CH_3O , NH_2 , azomethine proton ($\text{CH}=\text{N}$), and imine ($\text{C}=\text{NH}$) protons, respectively. Moreover, the ^{13}C -NMR spectrum of 7a displayed



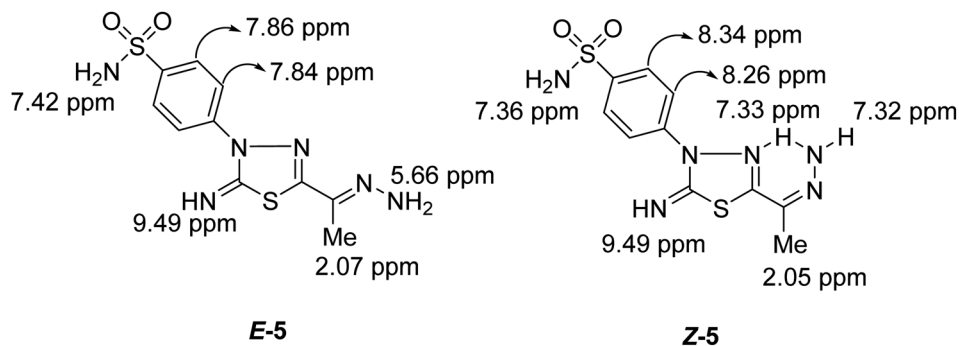


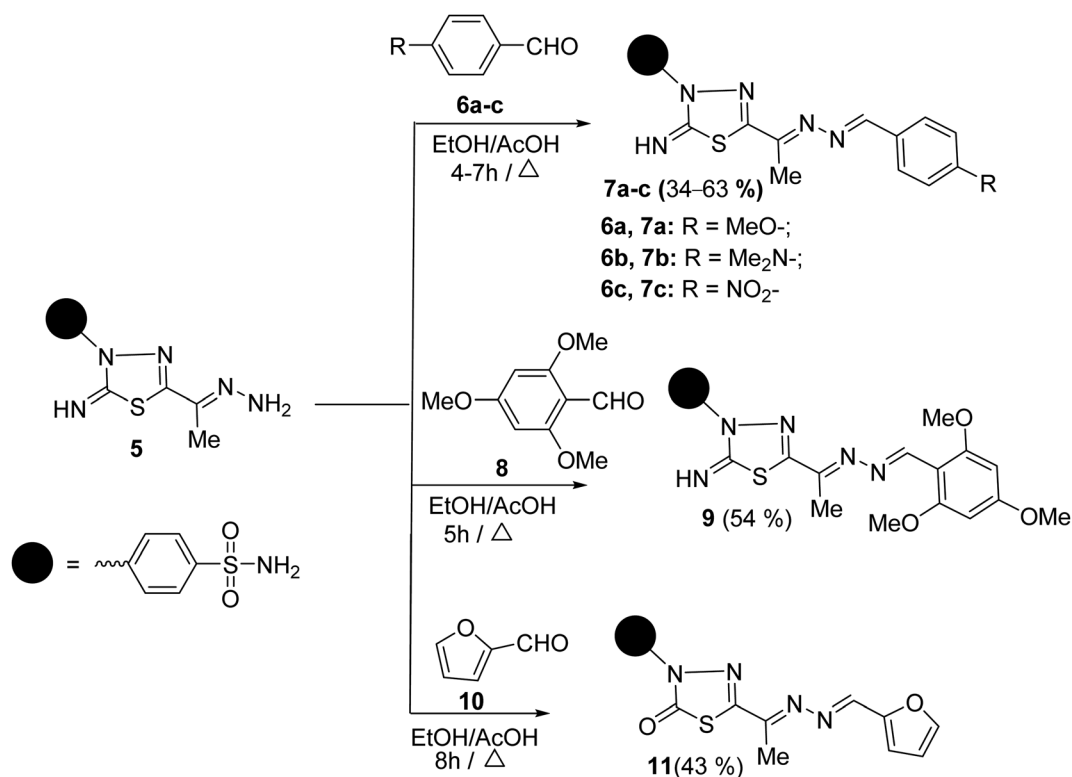
Fig. 2 Protons chemical shift values of *E*- & *Z*-isomers of compound 5.

fourteen carbon signals. Four characteristic peaks for four azomethine carbons showed at δ 161.91 (CH=N), 162.74 (C=N), 162.89 (thiadiazole-C₅), and 168.75 (thiadiazole-C₂) ppm. The signals at δ 13.46 and 55.85 ppm were attributed to methyl and methoxy carbons, respectively.

A M^+ at m/z 430 is also revealed in the mass spectra of **7a**, which corresponds to its molecular formula (C₁₈H₁₈N₆O₃S₂). The mass fragmentation pattern of compound **7a** (Scheme 1 at ESI[†]). The M^+ undergoes N–N bond cleavages to give a cation ion peak at m/z = 134 a.m.u. followed by loss of hydrogen atom to afford ion peak at m/z = 133 a.m.u. Due to the formation of *p*-methoxybenzonitrile. Then, due to phenyl cation, it loses further methoxy and nitrile groups to afford the base peak at m/z = 77. The parent ion peak can also undergo denitrogenation

with the formation of ion peaks at m/z = 402 and 403 a.m.u. assignable to 4-(2-imino-5-(1-(4-methoxyphenyl)prop-1-en-2-yl)-1,3,4-thiadiazol-3(2*H*)-yl)benzenesulfonamide and its protonated form, respectively (Scheme 1 at ESI[†]).

Similarly, the treatment of aminohydrazone **5** with furfural **10**, as heterocyclic aldehyde, in ethanol afforded 1-(furan-2-ylmethylene)-2-(1-(2-oxo-3-(4-sulfamoylphenyl)prop-1-en-2-yl)-1,3,4-thiadiazol-5-yl) ethylidene)hydrazine **11** (Scheme 2). The IR spectrum of **11** exhibited bands at 3338–3245, 1685, 1625, 1325, and 1291 cm^{−1}, which were attributed to NH₂, C=O, C=N, and SO₂ functions, respectively. Furthermore, the MS of **11** showed a M^+ at m/z = 391 and an ion peak at m/z = 393 (M^+ + 2). The ¹H-NMR spectrum of **11** revealed the absence of imine proton at about 9.50 ppm. It disclosed the existence of three singlet signals at



Scheme 2 The synthesis of unsymmetrical azines **7a–c**, **9**, **11**.



δ 2.44, 7.47, 8.48 ppm assigned to CH_3 , NH_2 , and $\text{CH}=\text{N}$ protons, along with the expected signals of furan and benzene residue. Its ^{13}C -NMR spectrum displayed important signals resonating at δ 13.68, 159.01, and 168.29 ppm characteristics for CH_3 , $\text{CH}=\text{N}$, and $\text{C}=\text{O}$ carbons, respectively.

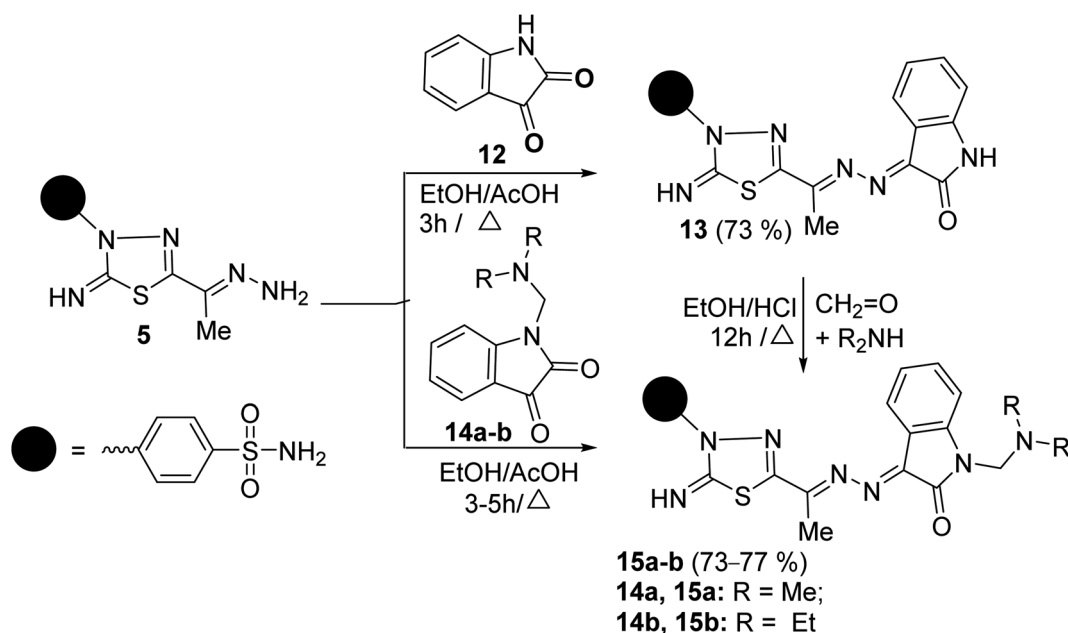
The scope of these condensation reactions was extended *via* the treatment of aminohydrazone **5** with heterocyclic ketones, namely, isatin **12**, 1-(*N,N*-dimethylaminomethyl)isatin **14a**,²³ and 1-(*N,N*-diethylaminomethyl)isatin **14b** (ref. 23) to afford the respective unsymmetrical azines **13**, **15a** and **15b** (Scheme 3). The MS of **13**, **15a**, and **15b** showed parent ion peaks at m/z = 441, 498, and 526 a.m.u. which is consistent with their molecular formulas ($\text{C}_{18}\text{H}_{15}\text{N}_7\text{O}_3\text{S}_2$), ($\text{C}_{21}\text{H}_{22}\text{N}_8\text{O}_3\text{S}_2$) and ($\text{C}_{23}\text{H}_{26}\text{N}_8\text{O}_3\text{S}_2$), respectively. The IR spectrum of **13** showed bands at 3303–3257, 3115, 1717, 1607, and 1587 characteristics for NH_2 , NH , $\text{C}=\text{O}$, $\text{C}=\text{N}$, and $\text{C}=\text{C}$ functions, respectively. The ^1H -NMR spectrum of **13** showed three D_2O -exchangeable signals at δ 10.92, 9.54, 7.41 ppm assignable to indoline-NH, imine-NH, NH_2 proton, and a singlet signal at 2.32 ppm assignable to the CH_3 protons. Its ^{13}C -NMR spectrum revealed signals at δ 163.91, 158.09, 154.71, and 13.49 ppm specific for carbonyl, thiadiazole- C_2 , $\text{C}=\text{N}$, and CH_3 carbons, respectively. Additionally, the structures of **15a** and **15b** were chemically indicated *via* Mannich reactions of **13** with each formalin and dimethylamine or formalin and diethylamine.

2.2. Quantum mechanical study

2.2.1. Molecular structure analyses. Initially, the *Z/E* configurations were theoretically investigated for **5** according to the orientation of amino N and thiadiazol C attached to the $\text{C}=\text{N}$ bond where they are in the same (*Z* isomer) or opposite sides (*E* isomer), see Fig. 3. The computational outcomes using B3LYP functional combined with the 6-31G(d) basis set reveal

that the *E* configuration is more stable than the *Z* form by 769 cm^{-1} ($2.21\text{ kcal mol}^{-1}$). While adopting the *E/Z* configurations, we need to explore the exact conformation of $-\text{C}(\text{CH}_3)\text{NNH}_2$, phenyl sulfonamide, sulfonamide, and methyl moieties which result from free rotation about C_2-C_6 , N_4-C_{15} , $\text{C}_{18}-\text{S}_{25}$ and C_6-C_7 single bonds, respectively. For this purpose, a relaxed scan of the PES was performed throughout the rotation of the dihedral angles $\tau_{\text{N}_8\text{C}_6\text{C}_2\text{N}_3}$, $\tau_{\text{C}_{16}\text{C}_{15}\text{N}_4\text{N}_3}$, $\tau_{\text{N}_{28}\text{S}_{25}\text{C}_{18}\text{C}_{19}}$, and $\tau_{\text{H}_{12}\text{C}_7\text{C}_6\text{C}_2}$ from 0° to 360° in steps of 10° proceeded by optimization process after every scan point. The obtained curves of the PES scan using B3LYP/6-31G(d) calculations for *E* and *Z* isomers are given at ESI (Fig. S1 and S2†). For the *E* isomer, the predicted PES curve from the rotation of $-\text{C}(\text{CH}_3)\text{NNH}_2$ moiety shows a global minimum when $\tau_{\text{N}_8\text{C}_6\text{C}_2\text{N}_3}$ reaches 180.0° in which the imine bonds ($\text{C}=\text{N}$) are *trans* to each other.

In contrast, minimum energy for *Z*-isomer was obtained at $\tau_{\text{N}_8\text{C}_6\text{C}_2\text{N}_3}$ equal to 0.0° where both $\text{C}=\text{N}$ bonds are *cis* to each other where stabilization could be attributed to intra-molecular H-bonding interaction between amino hydrogen and thiadiazol nitrogen. Moreover, the energetically favored conformation of benzenesulfonamide moiety ($-\text{C}_6\text{H}_4\text{SO}_2\text{NH}_2$) concerning the thiadiazol ring was assigned at $\tau_{\text{C}_{16}\text{C}_{15}\text{N}_4\text{N}_3}$ of 10.0° where the phenyl ring is almost planar towards the thiadiazol ring. The predicted curve for the scan of PES throughout the rotation of sulfonamide moiety for both *E* and *Z* isomer exhibit a minimum conformation at $\tau_{\text{N}_{28}\text{S}_{25}\text{C}_{18}\text{C}_{19}}$ equal to 260.0° in which the amino group is perpendicular to the phenyl ring. The internal rotation of the methyl group gives rise to a minimum at $\tau_{\text{H}_{12}\text{C}_7\text{C}_6\text{C}_2}$ equal to 60.0° and 0.0° for *Z* and *E* configuration, where the methyl group is orientated in *eclipsed* and *staggered* conformation to the adjacent $\text{C}=\text{N}$ bond, respectively. In conclusion, opposite conformations of $\text{C}(\text{CH}_3)\text{NNH}_2$ and methyl moieties were obtained for *E* and *Z* isomers. In contrast



Scheme 3 Reaction of aminohydrazone **5** with heterocyclic ketones **12**, **14a–b**.

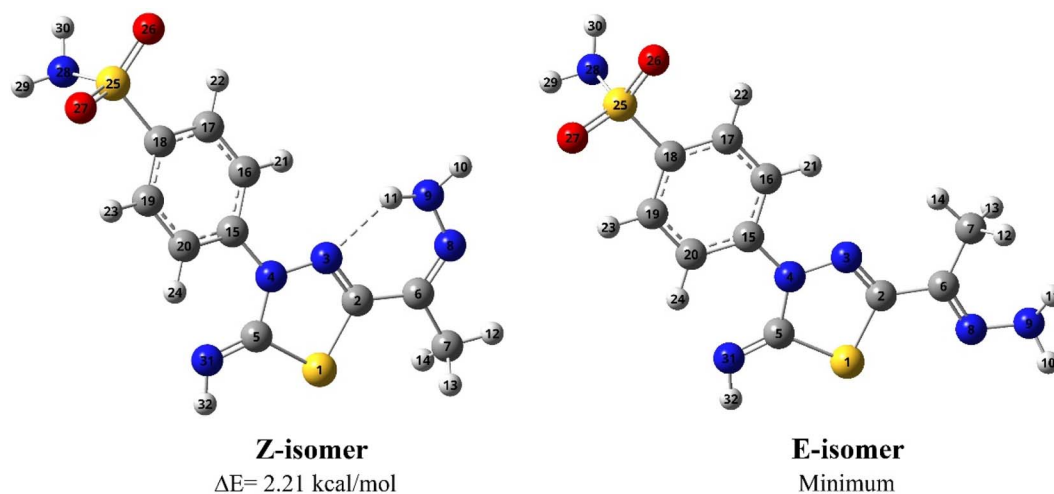


Fig. 3 Optimized geometries and atom numbering for *Z* and *E* isomers of **5** predicted using the B3LYP/6-31G(d) calculations.

to the *E* isomer, the global minimum of the *Z* isomer has the C=N bond in *cis* orientation to the thiadiazol C=N bond with the *eclipsed* conformation of the methyl group towards the C=N bond (Fig. 3).

The measured ^{13}C NMR spectrum for **5** displays 20 signals, double the number of carbon atoms, and reveals the presence of both *Z* and *E* isomers in the solution phase. Thus, the ^1H and ^{13}C NMR chemical shifts (δ in ppm) were calculated for both isomers using B3LYP/6-31G(d) calculations and compared to those observed experimentally. The computed ^1H and ^{13}C chemical shifts for *Z* and *E* isomers have equivalent values in the experimental spectra, as Table 1 shows that both isomers exist in the solution phase. In contrast to the *E* isomer, one proton of amino-hydrazone moiety (H_{11}) for the *Z* isomer was predicted to resonate at 7.31 ppm and matches the observed signal at 7.33 ppm, confirming the presence of the *Z* isomer in the solution phase. The downfield shift for H_{11} accounts for the intra-molecular H bonding with adjacent thiadiazol nitrogen. The ^1H NMR spectrum shows three signals at 7.36, 7.36, and 7.43 ppm corresponding to *amino* protons of sulfonamide moiety (H_{29} and H_{30}) in excellent agreement with those calculated for both *E* and *Z* isomers, 7.34–7.41 ppm. It's worth noting that, compared to tetramethylsilane (TMS), the prediction of chemical shifts for amino protons was improved when a multi-standard technique^{24,25} was applied using a comparable skeleton as a reference.

The predicted chemical shift for the *Z* isomer at 126.49 ppm belongs to the hydrazone carbon atom (C_6) and better matches the ^{13}C signal observed at 126.80 ppm. The counterpart value for the *E* isomer was computed at 138.02 ppm, which is consistent with the signal observed at 139.07 ppm. The experimental ^{13}C NMR spectrum shows signals at 151.26, 141.75, 122.43 and 121.33 ppm are better correlated to the *Z* isomer's computed values of 149.14 (C_2), 138.60 (C_{18}), 121.93 (C_{16}) and 120.83 (C_{20}) ppm, respectively. Regarding the *E* isomers, these atoms are predicted to resonate at 155.67, 136.97, 120.39, and 118.09 ppm, respectively, which match the signals displayed at 153.01, 134.63, 120.24, and 118.60 ppm, respectively. Compared

to the experimental value, the calculated chemical shifts of thiadiazol C_5 (for *E* isomer) and methyl C_7 (for *Z* isomer) were noticeably overestimated by 5.30 and 8.78 ppm, respectively. In

Table 1 Calculated^a $^1\text{H}/^{13}\text{C}$ NMR chemical shift (δ , ppm) for *Z* and *E* isomers of **5** compared to experimental values

Atom ^b	δ_{calc}			Atom ^b	δ_{calc}		
	<i>Z</i>	<i>E</i>	δ_{exp}^c		<i>Z</i>	<i>E</i>	δ_{exp}^c
C_2	149.14		151.26	H_{10}	5.52		(5.67)
		155.67	153.01			5.16	(5.67)
C_5	160.48		(159.23)	H_{11}	7.31		7.33
		164.53	(159.23)			5.63	5.67
C_6	126.49		126.80	H_{12}	1.95		(2.08)
		138.02	139.07			1.76	(2.08)
C_7	20.14		11.36	H_{13}	2.19		(2.08)
		7.64	10.27			1.68	(2.08)
C_{15}	144.59		142.89	H_{14}	2.21		(2.08)
		145.49	145.89			2.71	(2.08)
C_{16}	121.93		122.43	H_{21}	7.66		7.87
		120.39	120.24			7.79	8.28
C_{17}	128.93		127.15	H_{22}	7.73		8.27
		129.00	(131.35)			7.70	7.88
C_{18}	138.60		141.75	H_{23}	7.65		7.86
		136.97	134.63			7.60	7.84
C_{19}	129.02		(131.35)	H_{24}	8.57		8.35
		129.49	(131.35)			8.97	8.36
C_{20}	120.83		121.33	H_{29}	7.41		7.43
		118.09	118.60			7.36	(7.36)
				H_{30}	7.38		(7.36)
						7.34	7.34
				H_{32}	10.71		9.15
						10.82	9.31
rms ^d	3.24	2.41			0.49	0.57	
R^{2e}	0.9988	0.9988			0.9838	0.9804	

^a Calculations were carried out in DMSO solution by B3LYP/6-31G(d) method utilizing the IEF-PCM solvation model. ^b For the structures and atom numbering of *Z* and *E* isomers (Fig. 3). ^c The values of chemical shifts between brackets are assigned to more than one atom. ^d rms refers to the root mean square deviations of the theoretical values of chemical shifts from their corresponding experimental values. ^e The correlation coefficient (R^2) between the computed and observed values of chemical shifts.



sum, the computed chemical shifts of hydrogen/carbon nuclei for both *Z* and *E* isomers are well compared to those observed in the experimental $^1\text{H}/^{13}\text{C}$ NMR spectra, with high correlation coefficients (R^2) of 0.9838/0.9988 and 0.9804/0.9988, respectively.

The *E/Z* configuration for the synthesized products **7a–c**, **9**, **11**, **13** and **15a–b** were theoretically explored based on the orientation around the hydrazonyl $\text{C}=\text{N}$ bonds, which results in four possible configurations, **1** (*EE*), **2** (*ZZ*), **3** (*EZ*) and **4** (*ZE*). Therefore, a full geometry optimization was carried out for each configuration followed by frequency applying B3LYP method at 6-31G(d) basis set. The ESI† provides the equilibrium geometries and computed energy difference for the suggested configurations (Fig. S3–S10/Table S1†). For all synthesized compounds, the computational outcomes revealed *EE* (**1**) isomer to be the favored configuration with the lowest energy. Fig. 4 shows the predicted equilibrium geometries for the most stable configuration (**1**, *EE*) for **7a–c**, **9**, **11**, **13**, **15a** and **15b**.

The impact of FMOs and their associated molecular reactivity descriptors on molecule biological reactivity has recently

been considered.^{26,27} Herein, The FMOs were predicted for the optimized geometries of the synthesized molecules (**5**, **7a–c**, **9**, **11**, **13** and **15a–b**) to evaluate the reactivity and to correlate their biological activities. Subsequently, the computed energies for HOMO and LUMO were used to calculate a set of quantum chemical descriptors (Table 2), which are useful in assessing the molecule's overall reactivity.^{28,29}

The distribution for electron density in HOMOs and LUMOs for the investigated compounds is shown in Fig. 5, along with their energy gaps predicted using the B3LYP/6-31G(d) computations. The HOMO is largely localized on the thiadiazol ring and phenyl ring of the benzenesulfonamide moiety except for **7b**, where the HOMO is mostly distributed over the dimethylphenylamine moiety. The LUMO localized over the whole molecule for all synthesized molecules except the region containing the benzenesulfonamide moiety.

The energy gap ($E_{\text{HOMO}} - E_{\text{LUMO}}$) is a valuable sign of a molecule's chemical reactivity and kinetic stability. A molecule with a small energy gap is more polarized and has a higher chemical reactivity and lower kinetic stability.³⁰ Furthermore, Small

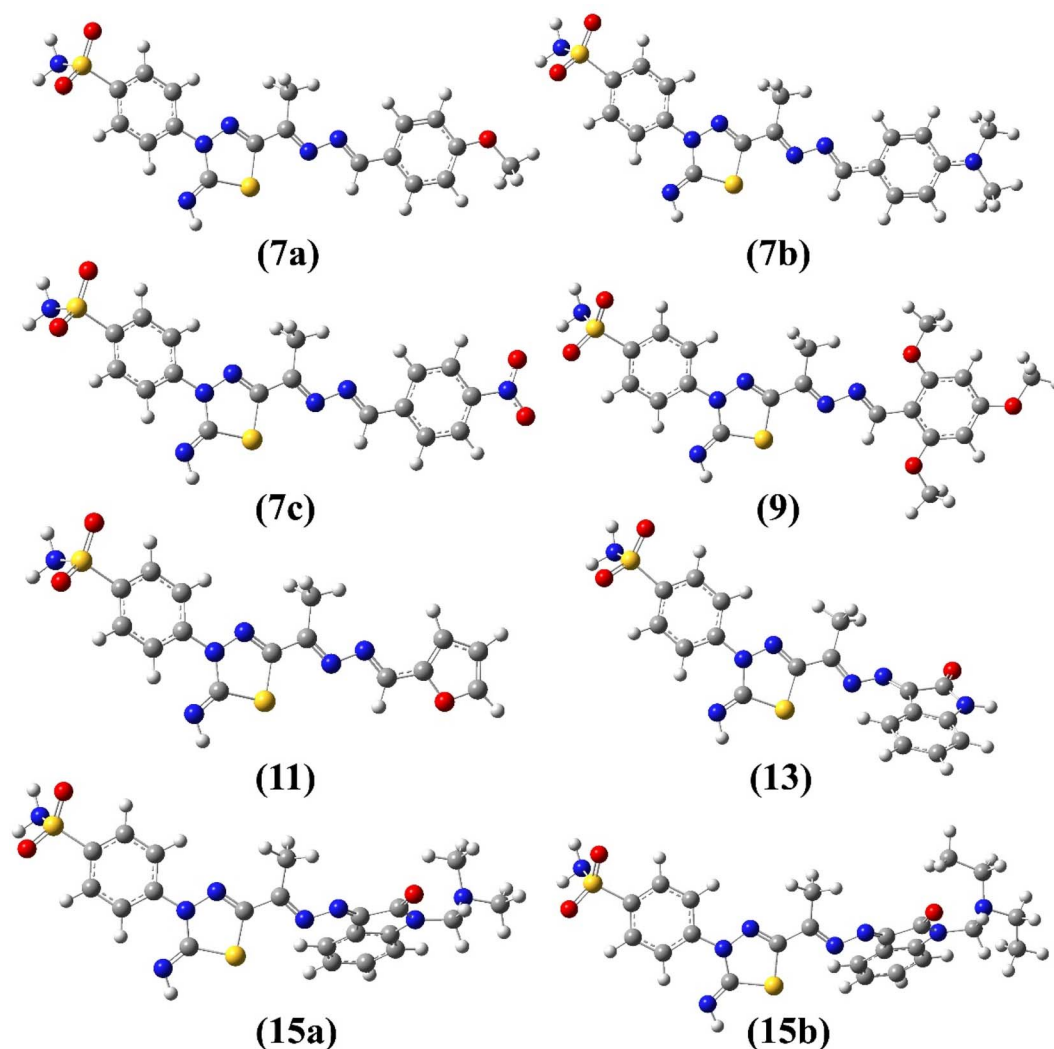


Fig. 4 Optimized geometries of **7a–c**, **9**, **11**, **13** and **15a–b** obtained from B3LYP/6-31G(d) calculations.



Table 2 DFT-B3LYP based^a quantum chemical descriptors^b for the synthesized compounds

Cpd	<i>E</i>	<i>I</i>	<i>A</i>	<i>E_g</i>	η	μ	ω	μ_{tot}	α
5	−45 266.30	5.75	1.64	4.11	2.06	−3.69	3.32	7.01	210.19
7a	−55 706.90	5.76	2.31	3.45	1.73	−4.03	4.71	9.53	367.89
7b	−56 236.08	5.36	2.11	3.24	1.62	−3.74	4.30	12.07	407.38
7c	−58 155.23	6.17	3.21	2.96	1.48	−4.69	7.43	3.69	368.79
9	−61 939.41	5.48	2.00	3.49	1.74	−3.74	4.01	11.91	402.22
11	−52 529.97	5.83	2.48	3.35	1.68	−4.15	5.15	7.73	321.64
13	−57 148.21	6.08	2.85	3.23	1.61	−4.46	6.17	5.25	339.36
15a	−61 863.38	6.02	2.76	3.25	1.63	−4.39	5.93	8.37	386.01
15b	−64 002.88	6.00	2.76	3.24	1.62	−4.38	5.92	8.77	409.20
Staurosporine	−41 561.54	4.90	0.79	4.12	2.06	−2.84	1.96	7.91	342.39
Valdecocib	−36 773.65	6.60	1.34	5.25	2.63	−3.97	3.00	3.07	203.20
Methazolamide	−39 083.89	6.58	1.78	4.80	2.40	−4.18	3.64	1.50	126.35
ZSL4	−55 935.10	6.06	2.12	3.94	1.97	−4.09	4.25	2.18	279.79

^a Calculations were carried out using 6-31G(d) basis set. ^b *E*, total energy; *I*, ionization potential = $-E_{\text{HOMO}}$; *A*, electron affinity = $-E_{\text{LUMO}}$; *E_g*, energy gap = $E_{\text{LUMO}} - E_{\text{HOMO}}$; η , hardness = $(I - A)/2$; μ , chemical potential = $-(I + A)/2$; ω , electrophilicity index = $\mu^2/2\eta$ (all in eV); μ_{tot} , total dipole moment in Debye; α , polarizability in a.u.

energy gaps suggest that the molecule undergoes a large intra-molecular charge transfer, which might affect the molecule's biological activity.³¹ The energy gaps calculated for the synthesized compounds ranged from 2.96 to 4.11 eV, comparable to the reported values for bioactive molecules^{32–34} and that computed for the standard reference drug, staurosporine (4.12 eV). For all compounds, the calculated energy gap *E_g* (Table 2) decreases in the order **5** (4.11 eV) > **9** > **7a** > **11** > **15a** > **15b** = **7b** > **13** > **7c** (2.96 eV). These results are compatible with the observed high activities for **7c**, **13**, **15b**, and **15a** against HepG-2 and Caco2 (Table 3).

Chemical potential (μ) is the inverse of electronegativity (χ), and it defines how much energy a molecule absorbs or releases during a chemical process. Both descriptors significantly impact a molecule's inhibitory effectiveness.³⁵ Compound **7c**, with a high χ value (of 4.69 eV), is more active than **7a** and **7b** with low values (4.03 and 3.74 eV), which is attributed to the presence of the nitro group in **7c** and validates the observed higher activity against MCF-7 (Table 3). Also, the hardness value for **7c** (1.48 eV) is lower than that calculated for **7a** and **7b**. It shows that it is a softer and more reactive molecule compared to **7a** and **7b**, in agreement with the estimated energy gap, see Table 2. Owing to the values of the electrophilicity index (ω), organic compounds were categorized as strong ($\omega > 1.5$ eV), moderate ($0.8 < \omega < 1.5$ eV), or weak ($\omega < 0.8$ eV) electrophiles.³⁶ Compounds **7c** and **13** are powerful electrophiles in this study, with ω values of 7.43 and 6.17, respectively. Compound **9** has the lowest biological activity against Caco2, which might be explained by its high energy gap (3.49 eV) and poor electrophilicity (4.01 eV) between all the synthesized compounds. To support the combi-targeting technique in this study, QM descriptors were computed for three commercial drugs with similar cores, valdecocib, methazolamide, and ZSL4 (Fig. 1, Table 2). As a result, the computed *E_g* and η values for the ZSL4 drug matched those predicted for compound **5** and staurosporine. Also, compounds **7a** and **11** have μ values of −4.03

and −3.97 eV, respectively, and are similar to those obtained for valdecocib and ZSL4 drugs.

The dipole moment is also considered when correlating the molecule's biological activity, which might influence the degree of interaction between drugs and the active sites of protein.³⁷ Compounds **15a–b**, for example, are more active against Caco2 and MCF-7 than **13**, which may be explained by the fact that **15a–b** has a higher total dipole moment (8.37–8.77 Debye) than **13** (5.25). As shown in Table 2, compounds **7b** and **9** have high total dipole moments (12.07 and 11.91 Debye, respectively) and exhibit high polarizabilities (407.38 and 402.22 a.u.), indicating that they are a good candidate for non-linear optical (NLO) materials.³⁸

2.2.2. ADMET prediction. Besides the high potency, the drug candidate's success involves favorable ADMET (absorption, distribution, metabolism, excretion, and toxicity) properties.³⁹ The ADMET prediction models have been introduced as an additional tool to aid drug discovery.¹⁶ This study predicted the *in silico* ADMET characteristics of all synthesized compounds **7a–c**, **9**, **11**, **13**, **15a** and **15b** using the SwissADME²⁰ and pkCSM²¹ servers, as shown in Table 4. The synthesized compounds show percent absorption ranged from 73.44 to 78.33%, suggesting they are well absorbed *via* the human intestine. Table 4 shows that three compounds **7a–c** have no violations of Lipinski's rule of five⁴⁰ for drug-likeness features and are considered orally active drugs. Other compounds have just one violation. All the synthesized compounds **7a–c**, **9**, **11**, **13**, **15a** and **15b** were found to be AMES nontoxic in nature and exhibit lethal doses (LD₅₀) ranging from 2.02 to 3.32 mol kg^{−1}, indicating these compounds seem to be suitably safe.

The oral bioavailability was estimated using the SwissADME's bioavailability radar, which considers six physicochemical parameters: size, solubility, lipophilicity, polarity, flexibility, and saturation. The bioavailable radars for the investigated compounds in this work are given (Fig. S11 at ESI†). Accordingly, the pink zone for all examined compounds has four parameters that provide physicochemical space



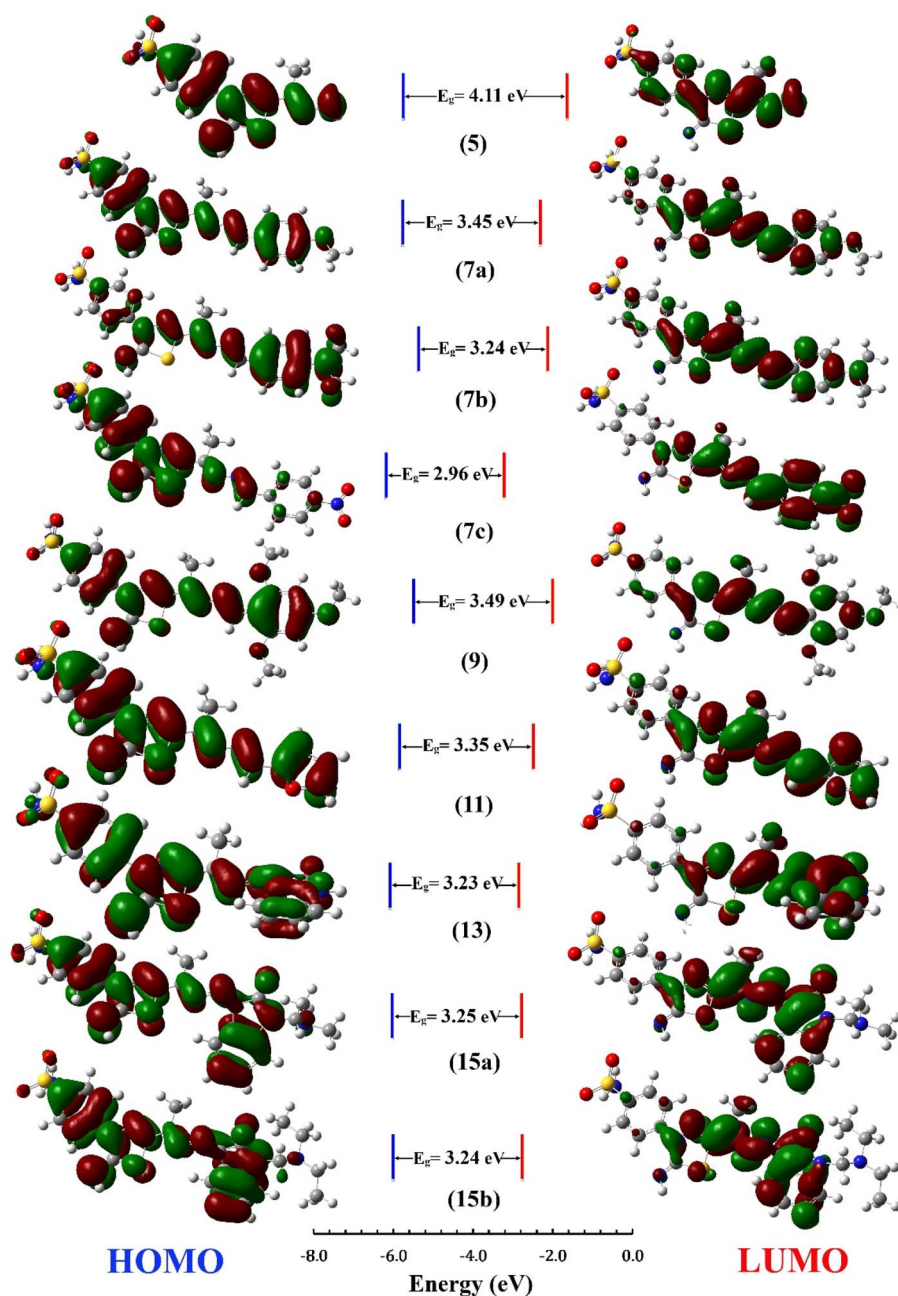


Fig. 5 HOMO and LUMO Frontier MOs and energy gaps (E_g) for the synthesized compounds predicted using B3LYP/6-31G(d) calculations.

acceptable for oral bioavailability. However, saturation and polarity were found beyond the bioavailability radar's pink zone due to a low saturation (fraction of $C_{sp}^3 < 0.25$) and high polarity ($TPSA > 130 \text{ \AA}^2$).

2.3. Bio-evaluation

2.3.1. Cytotoxicity evaluation. Screening of the cytotoxic effects of the synthesized compounds compared with staurosporine as a positive control against three human cancer cell lines: hepatocellular carcinoma (HepG-2), colon cancer (Caco-2), breast cancer (MCF-7) and one normal lung fibroblast (WI-38) (*vide* Table 3). The activity was determined using the

standard MTT colorimetric assay.^{41,42} Fig. 6 summarizes the results of the *in vitro* cytotoxic evaluation of synthesized compounds. As seen from Table 3, compounds 7a, 7c, 9, 13, 15a, and 15b displayed more activity against the HepG-2 cell line than staurosporine ($IC_{50} = 13.60 \text{ \mu M}$). However, compound 11 ($IC_{50} = 15.00 \text{ \mu M}$) showed nearly equipotent activity to staurosporine, and compound 7b showed the highest activity among the synthesized compound ($IC_{50} = 26.80 \text{ \mu M}$). As for activity against Caco2, compounds 7a, 7c, 15a, and 15b ($IC_{50} = 5.28, 3.16, 3.37, 2.33, \text{ and } 0.78 \text{ \mu M}$, respectively) were more active than staurosporine ($IC_{50} = 8.18 \text{ \mu M}$). As well as, compounds 11 and 13 ($IC_{50} = 8.55 \text{ and } 8.99 \text{ \mu M}$, respectively) exhibited almost

Table 3 Cytotoxicity (IC₅₀) of tested compounds **7a–c**, **9**, **11**, **13**, **15a** and **15b** on different cell lines

Compound No.	IC ₅₀ (μM)			
	HepG-2	Caco2	MCF-7	WI 38
7a	4.17 ± 0.20	3.16 ± 0.20	31.50 ± 1.4	28.20 ± 1.50
7b	26.80 ± 1.50	33.60 ± 1.70	35.20 ± 1.6	49.80 ± 2.60
7c	3.04 ± 0.20	3.37 ± 0.20	9.59 ± 0.4	25.40 ± 1.90
9	12.20 ± 0.70	47.20 ± 2.30	7.21 ± 0.3	25.40 ± 1.30
11	15.00 ± 0.80	8.55 ± 0.40	3.34 ± 0.2	24.50 ± 1.80
13	1.76 ± 0.10	8.99 ± 0.40	17.70 ± 0.8	11.40 ± 0.60
15a	6.54 ± 0.40	2.33 ± 0.10	4.34 ± 0.2	13.40 ± 0.70
15b	1.62 ± 0.10	0.78 ± 0.04	4.43 ± 0.2	16.80 ± 0.90
Staurosporine	13.60 ± 0.80	8.18 ± 0.70	6.19 ± 0.3	25.20 ± 1.30

similar activity to staurosporine. Moreover, compounds **7b** and **9** (IC₅₀ = 33.60 and 47.20 μM) showed moderate activity against the same cell line. Concerning the activity against the MCF-7 cell line, compounds **11** and **15a** (IC₅₀ = 3.34, 4.34, and 4.43 μM) were more active than staurosporine (IC₅₀ = 6.90 μM). On the other hand, compounds **7c**, **9**, and **13** (IC₅₀ = 9.59, 7.21, and 17.70 μM, respectively) showed high growth-inhibitory activity against MCF-7 cell line compared with the moderate activity of compounds **7a** and **7b** (IC₅₀ = 31.50 and 35.20 μM, respectively). Compounds **7a** and **7b** (IC₅₀ = 28.20, and 49.80 μM, respectively) displayed less toxicity against the normal fibroblast cell line than staurosporine (IC₅₀ = 25.20 μM).

3. Experimental section

3.1. Synthesis and spectroscopic characterization

3.1.1. Synthesis of 5-acetyl-3-*N*-(4-sulfamoylphenyl)-2-imino-1,3,4-thiadiazoline (4). A solution of propanehydrazonoyl chloride **1** (1.37 g, 0.005 mol) and ammonium thiocyanate (0.76 g, 0.005 mol) in EtOH (30 mL) was refluxed for 3 h. The reaction mixture was cooled and poured onto ice-cold water (40 mL). The formed precipitate was filtered and

recrystallized from EtOH to give compound **4** as orange crystals (1.30 g, 87%); M.P. 181–182 °C. IR $\nu_{\max}/\text{cm}^{-1}$ = 3352, 3289 (NH₂), 3277 (NH), 1692 (C=O), 1599 (C=N), 1579 (C=C), 1330, 1298 (SO₂); ¹H-NMR (DMSO-*d*₆): δ_{ppm} = 2.50 (s, 3H, CH₃), 7.45 (s, 2H, NH₂), 7.92 (d, *J* = 9.35 Hz, 2H, Ar-H_{2,6}), 8.18 (d, *J* = 9.35 Hz, 2H, Ar-H_{3,5}), 9.62 (s, 1H, NH); ¹³C-NMR (DMSO-*d*₆): δ_{ppm} = 24.88 (CH₃), 122.00 (2CH, Ar-C_{2,6}), 126.54 (2CH, Ar-C_{3,5}), 141.28 (Ar-C₄), 141.48 (Ar-C₁), 148.04 (thiadiazole-C₅), 158.30 (thiadiazole-C₂), 189.83 (C=O); MS *m/z* (%): 298 (M⁺, 55.9), 297 (43.7), 241 (0.9), 218 (1.0), 198 (10.2), 182 (0.8), 181 (4.6), 176 (1.3), 172 (3.5), 169 (1.8), 148 (1.0), 134 (2.2), 120 (1.1), 106 (100), 93 (2.3), 78 (10.0), 54 (4.9). Anal. calc. For C₁₀H₁₀N₄O₃S₂ (298.34): C, 40.26; H, 3.38; N, 18.78%; found: C, 40.24; H, 3.39; N, 18.77%.

3.1.2. Synthesis of (E)-4-(5-(1-hydrazineylideneethyl)-2-imino-1,3,4-thiadiazol-3(2*H*)-yl) benzenesulfonamide (5). A solution of **4** (2.98 g, 0.01 mol) and N₂H₄ (1.5 mL, 0.03 mol) in EtOH (40 mL) containing AcOH (3 drops) under reflux for 6 h. The precipitate was filtered and recrystallized from EtOH to get product **5** as a brown powder (2.43 g, 78%); M.P. 207–210 °C. IR $\nu_{\max}/\text{cm}^{-1}$ = 3416, 3285, 3233, 3119 (2NH₂), 3065 (NH), 1645 (C=N), 1585 (C=C), 1323, 1305 (SO₂); ¹H-NMR (DMSO-*d*₆):

Table 4 Physicochemical and pharmacokinetic properties for the synthesized compounds **7a–c**, **9**, **11**, **13** and **15a–b**

Parameter ^a	7a	7b	7c	9	11	13	15a	15b	Staurosporine
MW (g mol ^{−1})	430.50	443.55	445.48	490.56	390.44	441.49	498.58	526.63	466.53
<i>n</i> _{rot}	6	6	6	8	5	4	6	8	2
HBA	8	7	9	10	8	8	9	9	4
HBD	2	2	2	2	2	3	2	2	2
TPSA (Å ²)	172.40	166.41	208.99	190.86	176.31	192.27	186.72	186.72	69.45
%Abs	77.97	76.85	75.87	74.37	78.33	78.25	74.61	76.12	95.79
log <i>P</i> _{o/w}	2.42	2.40	2.40	2.33	2.33	2.33	1.83	1.83	3.03
log <i>S</i>	−4.10	−4.26	−4.26	−4.25	−4.25	−4.25	−4.26	−4.26	−5.06
GI abs	Low	Low	Low	Low	Low	Low	Low	Low	High
log <i>K</i> _p (cm s ^{−1})	−7.17	−7.15	−7.15	−7.58	−7.58	−7.58	−7.81	−7.81	−6.85
Lipinski, violation	Yes; 0	Yes; 0	Yes; 0	Yes; 1	Yes; 1	Yes; 1	Yes; 1	Yes; 1	Yes; 0
AMES toxicity	No	No	Yes	No	No	No	No	No	Yes
LD ₅₀ (mol kg ^{−1})	2.14	2.24	3.32	2.20	2.18	2.02	2.13	2.21	2.46
Skin sensitization	No	No	No	No	No	No	No	No	No

^a MW, molecular weight (g mol^{−1}); *n*_{rot}, number of rotatable bonds; HBA, number of H-bond acceptors; HBD, number of H-bond donors; TPSA, topological polar surface area; %Abs, intestinal absorption (human); log *P*_{o/w}, lipophilicity; log *S*, water solubility; GI abs, gastrointestinal absorption; log *K*_p, skin permeation, Lipinski, rule of five for drug likeness; LD₅₀, oral rat acute toxicity.



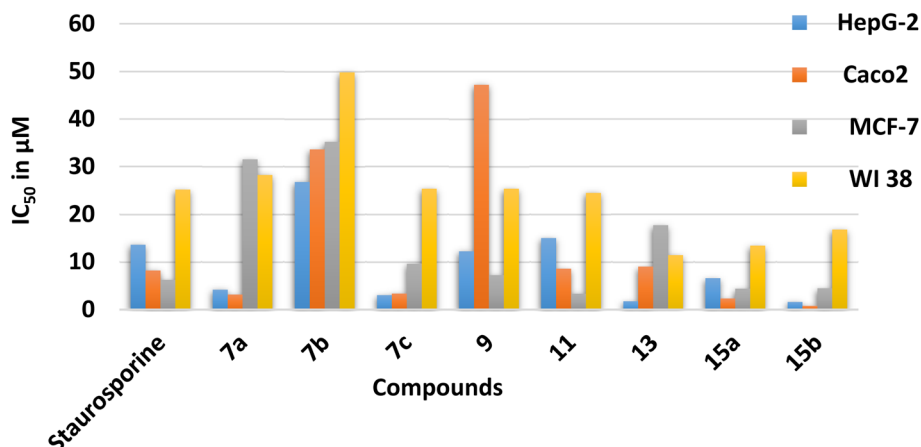


Fig. 6 The *in vitro* cytotoxic evaluation of derivatives 7a–c, 9, 11, 13, 15a and 15b presented as IC₅₀ in μM.

$\delta_{\text{ppm}} = 2.07$ (s, 3H, CH₃), 5.66 (s, 2H, NH₂), 7.42 (s, 2H, NH₂), 7.84 (d, $J = 8.5$ Hz, 2H, Ar–H_{2,6}), 7.86 (d, $J = 8.5$ Hz, 2H, Ar–H_{3,5}), 9.49 (s, 1H, NH); 2.05 (s, 3H, CH₃), 7.32 (s, 1H, ²N–H^b), 7.33 (s, 1H, ²N–H^a), 7.36 (s, 2H, NH₂), 8.26 (d, $J = 8.5$ Hz, 2H, Ar–H_{2,6}), 8.34 (d, $J = 8.5$ Hz, 2H, Ar–H_{3,5}), 9.49 (s, 1H, NH); ¹³C-NMR (DMSO-d₆): $\delta_{\text{ppm}} = 11.55$ (CH₃), 120.24 (2CHAr–C_{2,6}), 126.80 (2CHAr–C_{3,5}), 131.35 (Ar–C₄), 139.75 (Ar–C₁), 145.89 (C=N), 151.28 (thiadiazole–C₅), 159.23 (thiadiazole–C₂); MS m/z (%): 312 (M^+ , 10.3), 270 (1.3), 251 (2.5), 227 (2.2), 205 (1.4), 193 (1.5), 176 (9.6), 158 (2.4), 145 (2.2), 129 (7.1), 111 (8.9), 88 (2.2), 65 (11.7), 57 (100); anal. calc. For C₁₀H₁₂N₆O₂S₂ (312.37): C, 38.45; H, 3.87; N, 26.90%; found: C, 38.46; H, 3.86; N, 26.91%.

3.1.3. General synthetic procedure for unsymmetrical azines: 1-arylidene (hetarylalidene)-2-(1-(2-imino-3-(4-sulfamoylphenyl)-1,3,4-thiadiazole-5-yl)ethylidene) hydrazines (7a–c, 9, 11). A mixture of 5 (1 g, 0.003 mol), and 0.003 mol of an aromatic aldehydes 6a–c, 8 or heterocyclic aldehyde 10, was refluxed in EtOH (40 mL) containing AcOH (3 drops) under reflux for 3–8 h. The precipitate was filtered and recrystallized from EtOH to afford the desired compounds (7a–c, 9, 11, respectively).

3.1.3.1. 4-[2-Imino-5-(1-((4-methoxybenzylidene)hydrazineylidene)ethyl)-1,3,4-thiadiazol-3(2H)-yl]benzenesulfonamide (7a). The compound was obtained by heating a solution of 4-methoxybenzaldehyde 6a (0.43 g, 0.003 mol) for 5 h under reflux. Brown powder (0.70 g, 54%); M.P. 249–252 °C; IR $\nu_{\text{max}}/\text{cm}^{-1} = 3360, 3255$ (NH₂), 3110 (NH), 1604 (C=N), 1550 (C=C), 1332, 1305 (SO₂); ¹H-NMR (DMSO-d₆): $\delta_{\text{ppm}} = 2.49$ (s, 3H, CH₃), 3.84 (s, 3H, OCH₃), 7.07 (d, $J = 9.3$ Hz, 2CH, Ar–H_{3,5}), 7.49 (s, 2H, NH₂), 7.88 (d, $J = 9.3$ Hz, 2CHAr–H_{2,6}), 7.97 (d, $J = 2.5$ Hz, 2CH, Ar–H_{2,6}), 8.03 (d, $J = 2.5$ Hz, 2CHAr–H_{3,5}), 8.63 (s, 1H, CH=N), 9.42 (s, 1H, C=N); ¹³C-NMR (DMSO-d₆): $\delta_{\text{ppm}} = 13.46$ (CH₃), 55.85 (OCH₃), 114.87 (2CHAr–C_{3,5}), 121.32 (2CHAr–C_{2,6}), 126.88 (2CHAr–C_{2,6}), 127.34 (Ar–C₁), 131.16 (2CHAr–C_{3,5}), 131.33 (Ar–C₄), 142.91 (Ar–C₁), 149.40 (Ar–C₄), 161.91 (CH=N), 162.71 (C=N), 162.89 (thiadiazole–C₅), 168.75 (thiadiazole–C₂); MS m/z (%): 430 (M^+ , 4.2), 429 (0.4), 417 (3.9), 404 (11.2), 386 (0.09), 357 (0.1), 333 (0.1), 318 (0.1), 299 (0.1), 279 (0.1), 264 (0.4), 247 (0.2), 225 (0.7), 200 (1.5), 177 (6.4), 160 (2.1), 147 (4.3), 115

(3.0), 77 (100), 63 (50.4), 57 (31.6); anal. calc. For C₁₈H₁₈N₆O₃S₂ (430.50): C, 50.22; H, 4.21; N, 19.52%; found: C, 50.21; H, 4.22; N, 19.51%.

3.1.3.2. 4-[5-(1-((4-(Dimethylamino)benzylidene)hydrazineylidene)ethyl)-2-imino-1,3,4-thiadiazol-3(2H)-yl]benzenesulfonamide (7b). The compound was obtained by heating a solution of 4-(dimethylamino)benzaldehyde 6b (0.47 g, 0.003 mol) for 4 h under reflux. Brown powder (0.45 g, 34%); M.P. 235–237 °C; IR $\nu_{\text{max}}/\text{cm}^{-1} = 3310, 3225$ (NH₂), 3116 (NH), 1601 (C=N), 1524 (C=C), 1324, 1230 (SO₂); ¹H-NMR (DMSO-d₆): $\delta_{\text{ppm}} = 2.46$ (s, 3H, CH₃), 3.02 (s, 6H, 2CH₃), 7.39 (s, 2H, NH₂), 6.77 (d, $J = 9.3$ Hz, 2CH, Ar–H_{2,6}), 7.73 (d, $J = 9.3$ Hz, 2CH, Ar–H_{3,5}), 7.90 (d, $J = 8.5$ Hz, 2CH, Ar–H_{2,6}), 8.04 (d, $J = 8.5$ Hz, 2CH, Ar–H_{3,5}), 8.43 (s, 1H, CH), 9.38 (s, 1H, NH); ¹³C-NMR (DMSO-d₆): $\delta_{\text{ppm}} = 13.33$ (CH₃), 40.30 (2CH₃), 112.08 (2CHAr–C_{3,5}), 121.19 (2CHAr–C_{2,6}), 122.02 (Ar–C₁), 126.87 (2CHAr–C_{3,5}), 127.33 (Ar–C₄), 131.31 (2CHAr–C_{2,6}), 142.80 (Ar–C₁), 153.32 (Ar–C₄), 158.83 (CH=N), 162.66 (C=N), 163.69 (thiadiazole–C₅), 168.80 (thiadiazole–C₂); MS m/z (%): 443 (M^+ , 12.9), 442 (1.4), 428 (1.1), 414 (3.0), 383 (1.3), 353 (0.9), 342 (1.1), 299 (1.0), 263 (1.0), 251 (1.9), 223 (1.0), 195 (1.0), 181 (1.3), 149 (3.6), 116 (6.2), 90 (17.8), 63 (18.4), 57 (100); anal. calc. For C₁₉H₂₁N₇O₂S₂ (443.54): C, 51.45; H, 4.77; N, 22.11%; found: C, 51.46; H, 4.78; N, 22.12%.

3.1.3.3. 4-[2-Imino-5-(1-((4-nitrobenzylidene)hydrazineylidene)ethyl)-1,3,4-thiadiazol-3(2H)-yl]benzenesulfonamide (7c). The compound was obtained by heating a solution of 4-nitrobenzaldehyde 6c (0.48 g, 0.003 mol) for 7 h under reflux. Brown powder (0.84 g, 63%); M.P. 177–179 °C; IR $\nu_{\text{max}}/\text{cm}^{-1} = 3305, 3258$ (NH₂), 3112 (NH), 1623 (C=N), 1516 (C=C), 1307, 1291 (SO₂); ¹H-NMR (DMSO-d₆): $\delta_{\text{ppm}} = 2.47$ (s, 3H, CH₃), 7.49 (s, 2H, NH₂), 7.90 (d, $J = 1.7$ Hz, 2CH, Ar–H_{2,6}), 8.04 (d, $J = 1.7$ Hz, 2CHAr–H_{3,5}), 8.16 (d, $J = 3.4$ Hz, 2CH, Ar–H_{2,6}), 8.36 (d, $J = 3.4$ Hz, 2CHAr–H_{3,5}), 8.67 (s, 1H, N=CH), 9.49 (s, 1H, NH); ¹³C-NMR (DMSO-d₆): $\delta_{\text{ppm}} = 14.31$ (CH₃), 121.53 (2CHAr–C_{2,6}), 124.73 (2CHAr–C_{3,5}), 127.04 (2CHAr–C_{2,6}), 130.12 (2CHAr–C_{3,5}), 130.23 (Ar–C₄), 140.05 (Ar–C₁), 142.16 (Ar–C₁), 149.40 (N=CH), 151.55 (Ar–C₄), 158.41 (C=N), 159.60 (thiadiazole–C₅), 168.70 (thiadiazole–C₂); MS m/z (%): 445 (M^+ , 7.9), 444 (8.3), 429 (2.7), 416 (2.8), 393 (0.6), 368 (1.9), 354 (1.9), 335 (0.6), 308 (0.8),



269 (0.7), 237 (0.8), 223 (0.6), 205 (1.5), 190 (19.8), 174 (2.0), 141 (3.0), 119 (20.3), 95 (16.1), 73 (16.7), 57 (100); anal. calc. For $C_{17}H_{15}N_7O_4S_2$ (445.47): C, 45.84; H, 3.39; N, 22.01%; found: C, 45.83; H, 3.40; N, 22.02%.

3.1.3.4. 4-[2-Imino-5-(1-((2,4,6-trimethoxybenzylidene)hydrazineylidene)ethyl)-1,3,4-thiadiazol-3(2H)-yl]benzenesulfonamide (9). The compound was obtained by heating a solution of 2,4,6-trimethoxybenzaldehyde **8** (0.62 g, 0.003 mol) for 5 h under reflux. Brown powder; (0.79 g, 54%); M.P. 189–192 °C; IR $\nu_{\max}/\text{cm}^{-1}$ = 3305, 3247 (NH₂), 3115 (NH), 1620 (C=N), 1592 (C=C), 1325, 1294 (SO₂); ¹H-NMR (DMSO-d₆): δ_{ppm} = 2.50 (s, 3H, CH₃), 3.82 (s, 3H, OCH₃), 3.86 (s, 6H, 2OCH₃), 6.33 (s, 2H, Ar-H), 7.42 (s, 2H, NH₂), 7.97 (d, J = 8.5 Hz, 2H, Ar-H_{2,6}), 8.01 (d, J = 8.5 Hz, 2H, Ar-H_{3,5}), 8.68 (s, 1H, =CH), 10.3 (s, 1H, NH); ¹³C-NMR (DMSO-d₆): δ_{ppm} = 13.55 (CH₃), 56.50 (CH₃), 56.64 (CH₃), 56.76 (CH₃), 91.64 (2CHAr-C_{3,5}), 103.76 (Ar-C₁), 122.13 (2CHAr-C_{2,6}), 122.29 (2CHAr-C_{3,5}), 127.34 (Ar-C₄), 127.38 (Ar-C₁), 142.85 (thiadiazole-C₅), 143.07 (C=N), 149.4 (CH=N), 159.2 (Ar-C₄), 161.98 (Ar-C-O), 168.89 (thiadiazole-C₂); MS m/z (%): 490 (M⁺, 6.9), 489 (8.3), 472 (1.9), 461 (1.9), 443 (2.3), 408 (3.5), 378 (2.7), 354 (2.2), 322 (2.6), 280 (3.2), 266 (3.3), 229 (3.0), 217 (4.6), 196 (3.6), 179 (100), 142 (4.4), 121 (32.4), 76 (26.6), 57 (41.5); anal. calc. For $C_{20}H_{22}N_6O_5S_2$ (490.55): C, 48.98; H, 4.53; N, 17.12%; found: C, 48.99; H, 4.52; N, 17.13%.

3.1.3.5. 4-[5-(1-((Furan-2-ylmethylene)hydrazineylidene)ethyl)-2-oxo-1,3,4-thiadiazol-3(2H)-yl]benzene sulfonamide (11). The compound was obtained by heating a solution of furan-2-carbaldehyde **10** (0.30 g, 0.003 mol) for 8 h under reflux. Brown powder; (0.50 g, 43%); M.P. 222–223 °C; IR $\nu_{\max}/\text{cm}^{-1}$ = 3338, 3245 (NH₂), 1685 (C=O), 1625 (C=N), 1593 (C=C), 1325, 1291 (SO₂); ¹H-NMR (DMSO-d₆): δ_{ppm} = 2.44 (s, 3H, CH₃), 6.72 (d, J = 3.4 Hz, 1H, furan-H₃), 6.92 (t, J = 3.4 Hz, 1H, furan-H₄), 7.21 (d, J = 3.4 Hz, 1H, furan-H₂), 7.47 (s, 2H, NH₂), 7.98 (d, J = 5.1 Hz, 2CH, Ar-H_{2,6}), 8.01 (d, J = 8.5 Hz, 2CH, Ar-H_{3,5}), 8.48 (s, 1H, N=CH); ¹³C-NMR (DMSO-d₆): δ_{ppm} = 13.68 (CH₃), 113.48 (furan-C₃), 119.59 (2CHAr-C_{2,6}), 121.68 (furan-C₄), 126.91 (2CHAr-C_{3,5}), 139.67 (Ar-C₄), 146.64 (Ar-C₁), 148.87 (furan-C₂), 150.35 (thiadiazole-C₅), 151.53 (furan-C₅), 152.54 (C=N), 159.01 (C=NH), 168.29 (C=O); MS m/z (%): 391 (M⁺, 7.5), 390 (0.6), 378 (0.1), 365 (3.0), 341 (0.1), 311 (0.3), 295 (0.1), 280 (0.1), 262 (0.1), 239 (0.7), 221 (0.1), 205 (0.1), 198 (0.2), 188 (0.1), 176 (0.1), 154 (0.2), 128 (0.4), 116 (1.1), 94 (100), 78 (4.7), 65 (6.0); anal. calc. For $C_{15}H_{13}N_5O_4S_2$ (391.42): C, 46.03; H, 3.35; N, 17.89%; found: C, 46.02; H, 3.36; N, 17.90%.

3.1.4. General synthetic procedure for unsymmetrical azines: 1-arylidene (hetarylidene)-2-(1-(2-imino-3-(4-sulfamoyl-phenyl)-1,3,4-thiadiazole-5-yl)ethylidene) hydrazines (13, 15a–b). A solution of **5** (1 g, 0.003 mol) and 0.001 mol of cyclic heterocyclic ketones (**12** or **14a–b**) was refluxed in absolute EtOH (40 mL) containing AcOH (3 drops) for 3–5 h. The precipitate was filtered and recrystallized from EtOH to yield the desired compounds (**13** or **15a–b**, respectively).

3.1.4.1. 4-[2-Imino-5-(1-((2-oxoindolin-3-ylidene)hydrazineylidene)ethyl)-1,3,4-thiadiazol-3(2H)-yl]benzenesulfonamide (13). The compound was obtained by heating a solution of indoline-2,3-dione **12** (0.47 g, 0.003 mol) for 4 h under reflux. Green powder; (0.96 g, 73%); M.P. > 300 °C; IR $\nu_{\max}/\text{cm}^{-1}$ = 3303, 3257

(NH₂), 3115, 3019 (2NH), 1717 (C=O), 1607 (C=N), 1587 (C=C), 1322, 1292 (SO₂); ¹H-NMR (DMSO-d₆): δ_{ppm} = 2.32 (s, 3H, CH₃), 6.90–7.39 (m, 4H, Ar-H), 7.41 (s, 2H, NH₂), 7.89 (d, J = 8.5 Hz, 2H, Ar-H_{2,6}), 8.23 (d, J = 8.5 Hz, 2H, Ar-H_{3,5}), 9.54 (s, 1H, NH), 10.92 (s, 1H, indole-NH); ¹³C-NMR (DMSO-d₆): δ_{ppm} = 13.49 (CH₃), 111.12 (2CHAr-C_{2,6}), 116.13 (indole-C₄), 121.22 (indole-C₇), 122.47 (indole-C₅), 126.48 (indole-C₄), 126.94 (2CHAr-C_{3,5}), 128.19 (Ar-C₄), 134.34 (indole-C₆), 140.80 (indole-C₃), 141.70 (indole-C₅), 145.40 (Ar-C₁), 147.30 (thiadiazole-C₅), 154.71 (C=N), 158.09 (thiadiazole-C₂), 163.91 (C=O); MS m/z (%): 441 (M⁺, 11.5), 440 (2.3), 417 (0.7), 381 (0.7), 367 (1.3), 335 (0.7), 300 (1.7), 292 (0.7), 266 (0.9), 232 (0.6), 206 (2.3), 171 (2.5), 144 (9.3), 104 (100), 65 (41.8), 50 (34.8); anal. calc. For $C_{18}H_{15}N_7O_3S_2$ (441.48): C, 48.97; H, 3.42; N, 22.21%; found: C, 48.96; H, 3.41; N, 22.22%.

3.1.4.2. 4-[5-(1-((1-((Dimethylamino)methyl)-2-oxoindolin-3-ylidene)hydrazineylidene)ethyl)-2-imino-1,3,4-thiadiazol-3(2H)-yl] benzenesulfonamide (15a). The compound was obtained by heating a solution of 1-((dimethylamino)methyl)indoline-2,3-dione **14a** (0.65 g, 0.003 mol) for 3 h under reflux. Brown powder; (1.09 g, 54%); 73%; M.P. > 300 °C; IR $\nu_{\max}/\text{cm}^{-1}$ = 3303, 3269 (NH₂), 3115 (NH), 1721 (C=O), 1609 (C=N), 1590 (C=C), 1320, 1304 (SO₂); ¹H-NMR (DMSO-d₆): δ_{ppm} = 2.42 (s, 3H, CH₃), 2.50 (s, 6H, 2CH₃), 4.36 (s, 2H, CH₂-N), 6.92 (d, J = 7.6 Hz, 1CH, indole-H₇), 7.06 (t, J = 7.6 Hz, 1CH, indole-H₅), 7.44 (t, J = 7.6 Hz, 1CH, indole-H₆), 7.49 (s, 2H, NH₂), 7.61 (d, J = 7.6 Hz, 1CH, indole-H₄), 7.90 (d, J = 8.5 Hz, 2CH, Ar-H_{2,6}), 8.01 (d, J = 8.5 Hz, 2CH, Ar-H_{3,5}), 10.94 (s, 1H, NH); ¹³C-NMR (DMSO-d₆): δ_{ppm} = 14.18 (CH₃), 40.39 (CH₃)₂, 65.03 (N-CH₂-N), 111.55 (2CHAr-C_{2,6}), 116.49 (indole-C₇), 121.17 (indole-C₅), 123.06 (indole-C₄), 127.85 (2CHAr-C_{3,5}), 128.80 (Ar-C₄), 142.00 (Ar-C₁), 147.61 (thiadiazole-C₅), 153.27 (C=N), 154.63 (thiadiazole-C₂), 164.35 (C=O); MS m/z (%): 498 (M⁺, 2.6), 497 (3.1), 462 (2.9), 402 (2.6), 336 (3.0), 321 (2.5), 304 (2.6), 270 (3.6), 256 (3.1), 220 (3.8), 202 (2.5), 188 (6.1), 164 (4.0), 128 (7.0), 111 (16.0), 90 (40.7), 67 (31.5), 55 (100); anal. calc. For $C_{21}H_{22}N_8O_3S_2$ (498.58): C, 50.59; H, 4.45; N, 22.48%; found: C, 50.58; H, 4.44; N, 22.47%.

3.1.4.3. 4-[5-(1-((1-((Diethylamino)methyl)-2-oxoindolin-3-ylidene)hydrazineylidene)ethyl)-2-imino-1,3,4-thiadiazol-3(2H)-yl] benzenesulfonamide (15b). The compound was obtained by heating a solution of 1-((diethylamino)methyl)indoline-2,3-dione **14b** (0.74 g, 0.003 mol) for 5 h under reflux. Brown powder; (1.22 g, 77%); M.P. > 300 °C; IR $\nu_{\max}/\text{cm}^{-1}$ = 3305, 3264 (NH₂), 3115 (NH), 1728 (C=O), 1609 (C=N), 1589 (C=C), 1380, 1304 (SO₂); ¹H-NMR (DMSO-d₆): δ_{ppm} = 2.03 (t, J = 7.6 Hz, 3H, CH₃), 2.34 (s, 3H, CH₃), 2.92 (q, J = 7.6 Hz, 2H, CH₂), 4.36 (s, 2H, CH₂), 6.92 (d, J = 7.6 Hz, 1CH, indole-H₇), 7.06 (t, J = 7.6 Hz, 1CH, indole-H₅), 7.41 (s, 2H, NH₂), 7.45 (t, J = 7.6 Hz, 1CH, indole-H₆), 7.90 (d, J = 8.5 Hz, 2CH, Ar-H_{2,6}), 8.01 (d, J = 8.5 Hz, 2CH, Ar-H_{3,5}), 9.56 (s, 1H, NH); ¹³C-NMR (DMSO-d₆): δ_{ppm} = 11.52 (CH₃), 14.18 (CH₃), 41.61 (CH₂N), 65.03 (N-CH₂-N), 111.58 (2CHAr-C_{2,6}), 116.49 (indole-C₄), 121.17 (indole-C₇), 123.05 (indole-C₅), 126.95 (indole-C₄), 127.34 (2CHAr-C_{3,5}), 127.85 (Ar-C₄), 128.74 (1CHAr-C₆), 134.83 (C=N-N), 142.00 (Ar-C₁), 145.85 (1H-indole-C₅), 147.61 (thiadiazole-C₅), 148.07 (C=N), 154.60 (thiadiazole-C₂), 164.29 (C=O); MS m/z (%): 526 (M⁺, 1.4), 451 (1.9), 409 (1.7), 383 (2.5), 369 (4.7), 325 (2.0), 299



(3.5), 265 (2.9), 241 (1.4), 208 (1.8), 175 (1.9), 166 (2.2), 145 (3.0), 123 (12.7), 110 (10.3), 91 (8.5), 69 (58.8), 57 (100); anal. calc. For $C_{23}H_{26}N_8O_3S_2$ (526.63): C, 52.46; H, 4.98; N, 21.28%; found: C, 52.45; H, 4.97; N, 21.29%.

3.1.5. Alternative method for synthesis of unsymmetrical azines: 1-arylidene (hetaryali dene)-2-(1-(2-imino-3-(4-sulfamoylphenyl)-1,3,4-thiadiazole-5-yl)ethylidene)hydrazines (15a–b). A solution of **13** (1 g, 0.002 mol), 0.002 mol of formalin, and dimethylamine or formalin and diethylamine in EtOH (30 mL) containing concentrated HCl (three drops) under reflux for 12 h. The precipitate was filtered and recrystallized from EtOH to afford the compounds **15a–b**, respectively.

3.2. Computational details

Gaussian 09 package software⁴³ was used to carry out all quantum chemical calculations using the method of DFT-B3LYP^{12,13} combined with a standard 6-31G(d) basis set. The suggested geometries for the synthesized compounds were initially optimized using Pulay's gradient approach.⁴⁴ Frequency calculations were performed to verify that the optimized geometries are actual minimums with real wavenumbers. The geometry of the *E* and *Z* forms of **5** were reoptimized in DMSO solution using the IEF-PCM solvation model,⁴⁵ followed by ¹H and ¹³C NMR chemical shift calculations applying the GIAO approach.^{14,15} The multi-standard approach^{24,25} was applied to get the theoretical chemical shifts for *E* and *Z* isomers using the isotropic magnetic shielding values (σ_i , ppm) for H and C atoms acquired from the Gaussian output file. In this approach, methanol and benzene were used as references to predict sp^3 and sp^2 hybridized Cs/C–Hs, respectively, whereas comparable skeletons were used in the case of N–H protons. Thus, benzenesulfonamide,⁴⁶ acetophenonehydrazone,⁴⁷ and 5-iso-propyl-3-methyl-2-imino-1,3,4-thiadiazole⁴⁸ were used as references to calculate the N–H chemical shifts in the benzenesulfonamide, hydrazone and iminothiadiazoles moieties, respectively. Furthermore, the HOMO/LUMO (FMOs) and energy gaps were predicted for the optimized geometries of the synthesized compounds **7a–c**, **9**, **11**, **13**, **15a** and **15b**. Accordingly, molecular reactivity descriptors such as ionization potential, electron affinity, hardness, chemical potential, and electrophilicity index were calculated using the values of energies for FMOs.^{28,29} The synthesized compounds' pharmacokinetics, drug-like characteristics, and toxicities were evaluated using the SwissADME²⁰ and pkCSM²¹ online servers.

3.3. Biological evaluation

3.3.1. Cytotoxicity evaluation. HepG2, MCF-7, WI 38, Caco-2 cancer cell lines were cultured in complete media of RPMI and DMEM, respectively, at 5% carbon dioxide and 37 °C following standard tissue culture work. The cells were grown in “10% fetal bovine serum (FBS) and 1% penicillin–streptomycin” in 96-multiwell plate. All the synthesized compounds were screened for their cytotoxicity using 20 μ L of MTT solution (Promega, USA) for 48 h using untreated and treated cells with concentrations of (0.01, 0.1, 1, 10, and 100 μ M) for 48 h.^{41,42} The plate was cultured for 3 hours. The percentage of cell viability was

calculated following this equation: $(100 - (A_{\text{sample}})/(A_{\text{control}})) \times 100$. An ELISA microplate reader was used to measure the absorbance at 690 nm to calculate the viability *versus* concentration, and the IC₅₀ value using GraphPad prism software.

4. Conclusions

Herein, the combi-molecule strategy was used to synthesize a bundle of differently hybrid 1,3,4-thiadiazole sulfonamide derivatives and unsymmetrical azines **7a–c**, **9**, **11**, **13**, **15a** and **15b** started with 5-acetyl-3-*N*-(4-sulfamoylphenyl)-2-imino-1,3,4-thiadiazoline **4**. In the solution phase, compound **5** exists as a mixture of *E* and *Z* configurations, according to the results of computed/observed ¹H and ¹³C NMR chemical shifts. The DFT-B3LYP calculations favor the final synthesized products in the *EE* configuration with reference to the azomethine C=N bonds. For the newly synthesized compounds, quantum chemical descriptors and drug-likeness properties were predicted and correlated to their *in vitro* bioactivities. Experimentally, the synthesized compounds were investigated for their anticancer effects *in vitro* against cancer cell lines: HepG-2, Caco-2, MCF-7, and WI-38, where they exhibited promising results.

Conflicts of interest

The authors declare that there are no conflicts of interest.

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References

- 1 M. Wu, M. Wang, H. Jia and P. Wu, *Drug Delivery*, 2022, **29**, 2513–2538.
- 2 M. Szumilak, A. Wiktorowska-Owczarek and A. Stanczak, *Molecules*, 2021, **26**, 2601.
- 3 Y. Wan, G. Fang, H. Chen, X. Deng and Z. Tang, *Eur. J. Med. Chem.*, 2021, **226**, 113837.
- 4 A. Oving and J. Bhattacharyya, *Biophys. Rev.*, 2021, **13**, 259–272.
- 5 J. E. Lesch, *The first miracle drugs: how the sulfa drugs transformed medicine*, Oxford University Press, USA, 2007.
- 6 M. Mustafa and J. Y. Winum, *Expert Opin. Drug Discovery*, 2022, **17**, 501–512.
- 7 A. K. Jain, S. Sharma, A. Vaidya, V. Ravichandran and R. K. Agrawal, *Chem. Biol. Drug Des.*, 2013, **81**, 557–576.
- 8 M. F. Khan, G. Verma, W. Akhtar, M. Shaquiquzzaman, M. Akhter, M. A. Rizvi and M. M. Alam, *Arabian J. Chem.*, 2019, **12**, 5000–5018.
- 9 G. H. Elgemeie, R. A. Azzam and R. E. Elsayed, *Med. Chem. Res.*, 2019, **28**, 1099–1131.
- 10 G. Wang, Q. Han and R. Zhu, *J. Mol. Model.*, 2022, **28**, 393.
- 11 S. Fortin and G. Bérubé, *Expert Opin. Drug Discovery*, 2013, **8**, 1029–1047.



- 12 A. D. Becke, *Phys. Rev. A*, 1988, **38**, 3098–3100.
- 13 S. G. Patel, R. M. Vala, P. J. Patel, D. B. Upadhyay, V. Ramkumar, R. L. Gardas and H. M. Patel, *RSC Adv.*, 2022, **12**, 18806–18820.
- 14 E. Eroglu and H. Türkmen, *J. Mol. Graphics Modell.*, 2007, **26**, 701–708.
- 15 D. B. Chesnut and C. G. Phung, *J. Chem. Phys.*, 1989, **91**, 6238–6245.
- 16 S. Bharadwaj, A. Dubey, U. Yadava, S. K. Mishra, S. G. Kang and V. D. Dwivedi, *Briefings Bioinf.*, 2021, **22**, 1361–1377.
- 17 A. Sheikh Ali, D. Khan, A. Naqvi, F. F. Al-Blewi, N. Rezki, M. R. Aouad and M. Hagar, *ACS Omega*, 2020, **6**, 301–316.
- 18 P. K. Singh, J. Singh, T. Medhi and A. Kumar, *ACS Omega*, 2022, **7**, 33067–33078.
- 19 İ. Çapan, S. Servi, İ. Yıldırım and Y. Sert, *ChemistrySelect*, 2021, **6**, 5838–5846.
- 20 A. Daina, O. Michielin and V. Zoete, *Sci. Rep.*, 2017, **7**, 42717.
- 21 D. E. V. Pires, T. L. Blundell and D. B. Ascher, *J. Med. Chem.*, 2015, **58**, 4066–4072.
- 22 V. T. Angelova, V. Valcheva, N. G. Vassilev, R. Buyukliev, G. Momkov, I. Dimitrov, L. Saso, M. Djukic and B. Shivachev, *Bioorg. Med. Chem. Lett.*, 2017, **27**, 223–227.
- 23 H. Hellmann and I. Löschmann, *Chem. Ber.*, 1954, **87**, 1684–1690.
- 24 A. M. Sarotti and S. C. Pellegrinet, *J. Org. Chem.*, 2009, **74**, 7254–7260.
- 25 A. M. Sarotti and S. C. Pellegrinet, *J. Org. Chem.*, 2012, **77**, 6059–6065.
- 26 S. Tighadouini, S. Radi, F. Abridgach, R. Benabbes, D. Eddike and M. Tillard, *J. Chem. Inf. Model.*, 2019, **59**, 1398–1409.
- 27 A. I. Khodair, M. K. Awad, J.-P. Gesson and Y. A. M. M. Elshaier, *Carbohydr. Res.*, 2020, **487**, 107894.
- 28 T. Koopmans, *Physica*, 1934, **1**, 104–113.
- 29 R. G. Parr, L. v. Szentpály and S. Liu, *J. Am. Chem. Soc.*, 1999, **121**, 1922–1924.
- 30 J. I. Aihara, *J. Phys. Chem. A*, 1999, **103**, 7487–7495.
- 31 Y. N. Mabkhot, F. D. Aldawsari, S. S. Al-Showiman, A. Barakat, S. M. Soliman, M. I. Choudhary, S. Yousuf, M. S. Mubarak and T. B. Hadda, *Chem. Cent. J.*, 2015, **9**, 24.
- 32 R. Srivastava, *ACS Omega*, 2021, **6**, 24891–24901.
- 33 A. R. Bhat, R. S. Dongre, F. A. Almalki, M. Berredjem, M. Aissaoui, R. Touzani, T. B. Hadda and M. S. Akhter, *Bioorg. Chem.*, 2021, **106**, 104480.
- 34 S. K. Nagarajan, S. Babu, H. Sohn and T. Madhavan, *ACS Omega*, 2020, **5**, 21145–21161.
- 35 C.-G. Zhan, J. A. Nichols and D. A. Dixon, *J. Phys. Chem. A*, 2003, **107**, 4184–4195.
- 36 L. R. Domingo and P. Pérez, *Org. Biomol. Chem.*, 2011, **9**, 7168–7175.
- 37 M. Kouza, A. Banerji, A. Kolinski, I. Buhimschi and A. Kloczkowski, *Molecules*, 2018, **23**, 1995.
- 38 M. Marinescu, A. Emandi, G. Marton, L. O. Cinteza and C. Constantinescu, *Nanosci. Nanotechnol. Lett.*, 2015, **7**, 846–854.
- 39 J. G. Cumming, A. M. Davis, S. Muresan, M. Haeblerlein and H. Chen, *Nat. Rev. Drug Discovery*, 2013, **12**, 948–962.
- 40 C. A. Lipinski, F. Lombardo, B. W. Dominy and P. J. Feeney, *Adv. Drug Delivery Rev.*, 1997, **23**, 3–25.
- 41 T. Mosmann, *J. Immunol. Methods*, 1983, **65**, 55–63.
- 42 M. M. Hammouda, A. A. Elmaaty, M. S. Nafie, M. Abdel-Motaal, N. S. Mohamed, M. A. Tantawy, A. Belal, R. Alnajjar, W. M. Eldehna and A. A. Al-Karmalawy, *Bioorg. Chem.*, 2022, **127**, 105995.
- 43 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery Jr, J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, Ö. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski, and D. J. Fox, *Gaussian 09, (Revision B.01)*, Gaussian Inc., Wallingford CT, 2009.
- 44 P. Pulay, *Mol. Phys.*, 1969, **17**, 197–204.
- 45 J. Tomasi, B. Mennucci and E. Cancès, *J. Mol. Struct.: THEOCHEM*, 1999, **464**, 211–226.
- 46 S. L. Graham and T. H. Scholz, *Synthesis*, 1986, **1986**, 1031–1032.
- 47 T. Kleine, R. Fröhlich, B. Wibbeling and E.-U. Würthwein, *J. Org. Chem.*, 2011, **76**, 4591–4599.
- 48 D. Leppard and H. Sauter, *J. Heterocycl. Chem.*, 1980, **17**, 1469–1472.

