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A comprehensive review on antimicrobial face masks: an emerging weapon in fighting pandemics

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The world has witnessed several incidents of epidemics and pandemics since the beginning of human existence. The gruesome effects of microbial threats create considerable repercussions on the healthcare systems. The continually evolving nature of causative viruses due to mutation or re-assortment sometimes makes existing medicines and vaccines inactive. As a rapid response to such outbreaks, much emphasis has been placed on personal protective equipment (PPE), especially face mask, to prevent infectious diseases from airborne pathogens. Wearing face masks in public reduce disease transmission and creates a sense of community solidarity in collectively fighting the pandemic. However, excessive use of single-use polymer-based face masks can pose a significant challenge to the environment and is increasingly evident in the ongoing COVID-19 pandemic. On the contrary, face masks with inherent antimicrobial properties can help in real-time deactivation of microorganisms enabling multiple-use and reduces secondary infections. Given the advantages, several efforts are made incorporating natural and synthetic antimicrobial agents (AMA) to produce face mask with enhanced safety, and the literature about such efforts are summarised. The review also discusses the literature concerning the current and future market potential and environmental impacts of face masks. Among the AMA tested, metal and metal-oxide based materials are more popular and relatively matured technology. However, the repeated use of such a face mask may pose a danger to the user and environment due to leaching/detachment of nanoparticles. So careful consideration is required to select

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AMA and their incorporation methods to reduce their leaching and environmental impacts. Also, systematic studies are required to establish short-term and long-term benefits.

1. Introduction

Outbreaks of communicable diseases are not uncommon and have existed since humankind's hunter-gatherer days. However, they became more prominent when the world shifted towards settled community life. Pathogenic organisms such as viruses, bacteria, protozoa, helminths, and fungi are the major causes of infectious diseases in humans and animals. It is estimated that one-fourth of annual global deaths are due to these pathogenic organisms.¹ Mainly, flu viruses and non-flu viruses like *Flavivirus*, *Filoviridae*, and human coronavirus are responsible for major epidemics and pandemics starting from Russian flu-1889 to the present Coronavirus disease (COVID-19). The four communicable diseases that rattled the world are Influenza, Ebola, Acquired immunodeficiency syndrome (AIDS), and COVID-19. According to the World Health Organisation (WHO), the respiratory diseases from annual seasonal influenza alone

results in 290 000–650 000 deaths each year.^{2,3} Also, the rapidly evolving influenza viruses cause annual epidemics and occasional pandemics (3 to 4 times per century).⁴ The fatality rate in Ebola is estimated to be as high as 50%.⁵ AIDS, with no complete cure, continues to be a significant global public health issue,⁶ and the present COVID-19 pandemic has already resulted in more than 95 million cases worldwide (till 17th January 2021).⁷

The emergence of more resistant strains with high infection potential happens because of mutation or re-assortment of pre-existing microbial strains, which sometimes makes the vaccines and antibiotics incompetent. Therefore, to control or delay the spread of the infection, WHO and the Centers for Disease Control and Prevention (CDC) suggests various non-pharmaceutical interventions such as hand washing, personal protective equipment (PPE), isolation, quarantine, personal hygiene, use of disinfectants, and social distancing. For instance, the use of respiratory protective devices (RPDs) has gained massive impetus during COVID-19. This intervention is highly beneficial in pandemic situations to reduce exposure to infectious respiratory aerosols and droplets and slow down the spread of the disease.^{8–12} Realising the fact, most of the countries have enforced or advised its citizen to wear face masks in public places.¹³

However, there are several concerns associated with the use of conventional face masks such as the survivability of microorganisms on the face mask surface,^{14,15} re-aerosolisation of settled particles,¹⁶ safe management and disposal of used face masks,¹⁷ and fomite transmission.¹⁸ Some of these concerns have led to the development of face masks with inherent antimicrobial properties to impart surface contact killing/deactivation property in addition to particle filtration.



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Antimicrobial face masks pave the way for reusability and considerably reduces the surging demand for single-use face masks, especially during outbreaks in highly populated, developing, and middle-income countries. Moreover, the ongoing pandemic has further stressed the significance of such development. Thorough knowledge of suitable antimicrobial agents (AMA) and incorporation methods are required to develop potential antimicrobial face masks. This is one of the prospective areas evolving through research and gaining much interest among scientists and researchers. The lucrative market steers the entrepreneurs and industrialists to focus on improvising face masks to meet consumers' safety and quality demands, especially in the COVID-19 and post COVID-19 scenarios. More clarity on various aspects of the antimicrobial mask is imminent at this juncture, realising the importance of a review on antimicrobial face masks.

The paper presents a comprehensive review of the face mask with a particular emphasis on antimicrobial face masks as an emerging weapon in containing the spread of infectious diseases. The topics such as AMA used in face masks and their methods of incorporation, filtration efficacy, and current and future market potential are discussed. A detailed description of the evolution of face masks and their role in preventing the spread of infectious disease, potential issues with the conventional face masks, and possible environmental challenges associated with the usage of face masks are also included. The review also gives a brief insight into the history of pandemics, epidemics, and their socio-economic impacts. The authors feel that this review is vital in the present scenario of COVID-19 and post COVID-19 due to the recurring nature of infectious diseases, to acquaint the readers with the relevant aspects of face mask in general and antimicrobial face masks in particular.

2. Major epidemics and pandemics in history

The occurrence of pandemics and epidemics have been reported throughout history. It is accepted that pandemics occur at unpredictable intervals with no specific occurrence pattern.¹⁴ The years (in A.D.) 1510, 1557, 1580, 1593, 1732–1733, 1767, 1781–1782, 1802–1803, 1830–1833, 1836–1837, 1847–1848, 1850–1851, 1855, 1857–1858 and 1874–1875 have been recorded as pandemic years. The reports on epidemics and pandemics outbreak before the 12th century are defective, unreliable, or does not warrant definite conclusions.¹⁹ Among the pandemics, the Bubonic plague or Black death (AD 1347 to 1351) was the deadliest in terms of total death (75 to 200 million) and fatality rate (30 to 100%).²⁰ In the last 20 decades, the diseases due to flu viruses (Influenza), primarily transmitted through respiratory aerosols, are prevailing worldwide. Influenza takes a notable place among various infectious diseases because of its pronounced pandemic character than others.¹⁹

Spanish flu-1918 pandemic was the most fatal among various flu diseases, with 10 and 26.8% fatality rate (total number of death \times 100/total number of infected individuals) and infection rate (total number of infected individuals \times 100/population), respectively. Asian flu-1957 and Hong Kong flu-1968 pandemics had a fatality rate of 0.2 to 0.8%. The infection rate of Asian flu-1957 and Hong Kong flu-1968 was 17.4 and 14.1%, respectively. The fatality rate (0.01 to 0.04%) of Swine flu-2009 pandemic was low, but the infection rate was as high as 21%.^{21,22} The COVID-19 has also resulted in a catastrophic fatality rate of approximately 2.2% and an infection rate of 1.2%.^{7,23} This data reveals that COVID-19 is the worst pandemic since the Spanish flu-1918 in terms of fatality rate.

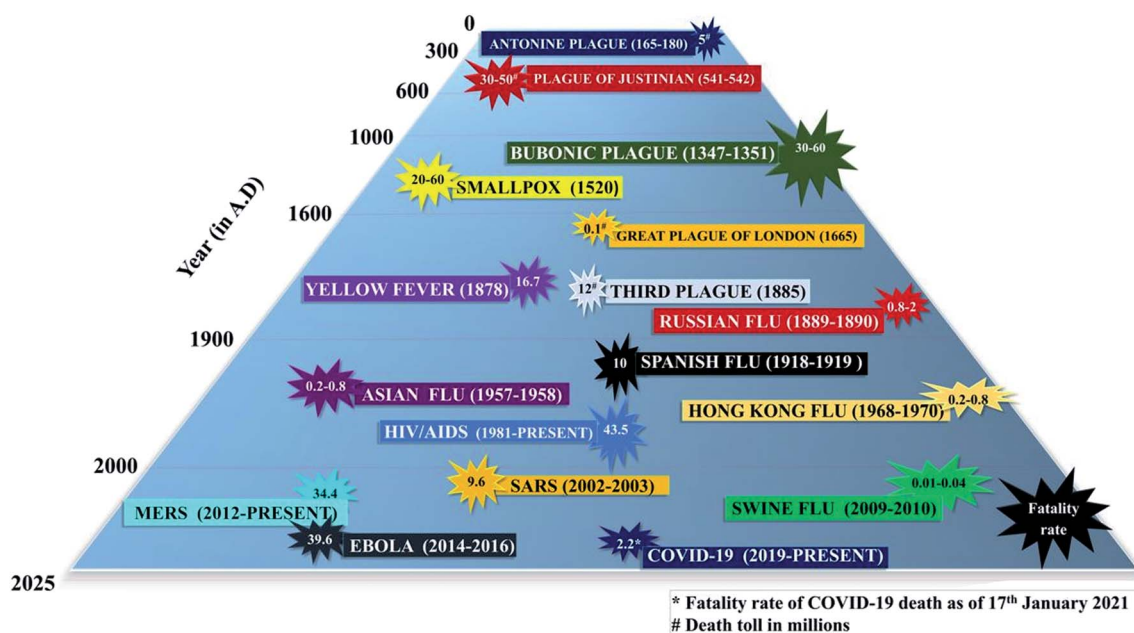


Fig. 1 Timeline of various deadly pandemics in history. This figure is adapted from the ref. 29 with modifications and the data is from the ref. 7, 20, 21, 24–26, 28 and 29.



Among the non-flu diseases, AIDS, Ebola, Severe acute respiratory syndrome (SARS, 2003) and the Middle East respiratory syndrome (MERS, 2012) were of much concern to the entire world. AIDS is given a special term 'global pandemic' with almost 76 million people infected, and 33 million deaths since its first reported occurrence in 1981.²⁴ AIDS is one of the significant infectious viral diseases in terms of fatality rate (43.5%), followed by Ebola (39.6%).^{21,22,24} SARS-2003 and MERS-2012 with a fatality rate of 9.6% and 34.4%, respectively are epidemics that target the human respiratory system.^{25–27} Even though the fatality rate is higher for SARS-2003 and MERS-2012, the infected rate is 3 or 4 orders lesser than COVID-19. COVID-19 has resulted in 95 016 073 cases, 2 032 342 deaths affecting 219 countries and territories worldwide, plus two international conveyances (till 17th January 2021).⁷ A summary of the well-documented deadliest pandemics in history is presented in Fig. 1.

3. Socio-economic impacts of epidemics and pandemics

The impacts of the pandemic are far-reaching, disrupting the social, economic, and political conditions of the affected nations. During the Spanish flu-1918 wave, cities worldwide ran out of space to bury the dead, and several villages perished.⁴ The vast difference in fatality rate between the high and low-income countries was also noticed during the Spanish flu times. Social inequality is an unnoticed factor associated with pandemics, as evident from the fact that fatality rates are highest for individuals with the lowest socio-economic status.^{30,31} The recent COVID-19 pandemic has resulted in massive and simultaneous shutdowns/lockdowns worldwide with the suspension of social gatherings and drastic changes in the work and education culture. More than half of the world population were under complete lockdown by the first week of April 2020. Drastic behavioural changes and long-term social stigma arising out of

the isolation of the infected and suspected individuals were prevalent during Spanish flu-1918. Similar manifestations are observed in the present COVID-19 as well. Mood changes, insomnia, psychiatric issues, and fatigue were also reported for 40% of the survivors of SARS-2003 in Hong Kong, and maybe the case with all other pandemics.³²

Pandemics can also increment the anxieties of nations with a weak political framework.³⁰ For example, during the Cholera pandemic-1832, deaths of people in a particular community were linked to some conspiracy theories, resulting in communal violence.³³ In light of COVID-19, publishing in the newspaper was banned in some countries, and political instability surged in some places.^{34,35} These are examples of the influence of pandemic in arenas like sovereignty, civil rights, and democracy.

Apart from social and political repercussions, pandemic and its aftermath affect all major sectors of the economy – trade, agriculture, tourism, and industrial activities. These can be through multiple channels, including short-term fiscal shocks and long-term impact on economic growth. The major incursion of the direct and indirect costs include setting up proper diagnostic and treatment facilities, imparting the preventive steps, strengthening core public-health infrastructure, disinfecting the affected regions, promoting research on developing vaccines, and most essentially, providing essential amenities to the unemployed poor people.³⁰ Economic impacts can scale up to an increased cost of living and a wider gap between the rich and the poor. Pandemics can reduce the capacity of the infected and recovering population to undertake income-generating activities, eroding them financially. It is estimated that a pandemic with 1.4 million deaths could result in 0.8% gross domestic product (GDP) loss.³⁶ Fig. 2 shows the financial losses in terms of GDP loss (%) associated with some of the major pandemics and epidemics.^{36–42}

Spanish flu-1918 has impacted the European region the most. The GDP losses in Great Britain, Canada and the United States (US) were 16.91, 14.68 and 11.32%, respectively. Asian flu-



Fig. 2 Economic impact expressed as GDP loss (%).



1957 has incurred a GDP loss of 2.6, 3.1, 3.3 and 3.5% in Japan, the US, Great Britain and Canada, respectively.³⁶ Most cases of SARS-2003 were reported in regions of China, Hong Kong, Taiwan, Vietnam, Singapore, Canada, and the US.⁴³ SARS-2003 displayed a negative impact on China's economy, with a total loss of 25.3 billion USD and 1 to 2 percentage point reduction in GDP rate than expected.⁴⁴ After the Spanish flu-1918, the worst impact on the economy is due to the ongoing COVID-19. The GDP losses (%) are in two digits range for the major economies of the world with the US, India, United Kingdom (UK), France, Italy, Canada, and Germany incurring 32.9, 23.9, 20.4, 13.8, 12.4, 12, and 10.6%, respectively. In contrast, the Chinese economy grew by 3.2% in the April to June quarter of 2020.⁴¹ One can see that China's economy has shown a V-shaped recovery after the first quarter. The fast recovery is attributed to increased medical exports, manufacturing, and industrial production.^{45,46}

Individual behavioural changes such as aversion for work environments hamper the financial progress of the country. The disrupted domestic and international distribution channels and consumer hoarding during pandemics can heighten the price of agricultural and other commodities, thereby reducing the household purchasing power. The International Labour Organisation has predicted that unemployment could rise by 5.3 to 24.7 million due to COVID-19.³⁷ When young people in the age group of 15 to 54 are affected by pandemics, it can significantly decline the GDP growth rate. The world economic outlook report by the International Monetary Fund predicted that the global economy would contract by 3% in 2020, which is much worse than the 2008–09 financial crisis. Moreover, a resurgence of COVID-19 in 2021 could leave the economies to wrestle for posterity.^{47,48} The World Bank estimates that the occurrence of any pandemic in the future with high severity, similar to Spanish flu-1918, might decline the GDP by 5%.³⁶ If the severe effects continue, even food security would be at threat, especially in low-income countries.³⁷ Considering various socio-economic impacts of the pandemic, a trans-disciplinary approach should be adopted to study them. Such a study in which several factors are considered to prepare the pandemic preparedness plan would reduce the burden of disease.³¹

4. Role of face mask in preventing the spread of infectious diseases

The spread of infectious diseases from person to person and the degree of transmission vary based on the aetiology and mode of

transmission. Usually, the degree of transmission is estimated by a mathematical number known as reproduction number. It determines the number of infections caused by a single infected individual. It combines factors such as duration of infectiousness, the speed of contact to vulnerable individuals per unit time, and the probability of transmission per contact.⁴⁹ The transmission ways are mainly classified into the following three: (i) direct transmission – infected droplet/aerosol transmission, (ii) airborne route – inhaling infected respiratory aerosols, and (iii) contact transmission through secretions on fomites or directly such as through physical touch resulting in hand-to-mouth, hand-to-eye or hand-to-nose transmission.^{50,51} The information regarding the causative agent, transmitting host, transmission route, and reproduction number help in implementing a robust and effective protocol to control infectious diseases. Table 1 summarises major viral diseases declared by WHO as a public health emergency in the affected countries with their causative agents, hosts, and mode of transmission.⁵²

A face mask as an RPD helps prevent airborne, droplet and aerosol transmission where the primary control mechanism lies in reducing exposure. Face mask help in filtering out particles of different sizes through various mechanisms:^{53,54} (i) inertial impaction by which particles with more mass and inertia are diverted from the streamline around the filter fibre, (ii) interception by filter fibres of larger particles, (iii) diffusion which results from the collision with air molecules or fibres resulting in deviation from streamline on account of the Brownian movement of fine particles, and (iv) electrostatic attraction between oppositely charged particles and filter media, without any size distinction of particles. Diffusion mechanism mainly comes into play when the particle sizes are lesser than 0.1 μm , and a combination of diffusion and interception applies for particle size between 0.1 to 0.5 μm . Particles greater than 0.5 μm are removed by interception and inertial impaction.⁵³ The diameter of bacteria is typically 0.4 μm .⁵⁵ Diameter of viruses usually ranges between 0.01 to 0.20 μm .⁵⁶ Diagrammatic representation of filtration mechanisms and filter efficiency *versus* particle diameter are shown in Fig. 3(A) and (B), respectively. In case of antimicrobial face masks, in addition to the physical entrapment, the AMA deactivates/kills the microorganisms that are deposited or captured. Herein, deactivation or killing is mainly based on the mode of action of AMA and is discussed in Section 6.

A recent study in Hong Kong has revealed 96.6% general public complying with face mask use during COVID-19. The incidence rates reported in Hong Kong were significantly lower than Spain, Italy, Germany, France, the US, the UK, Singapore,

Table 1 Causative agents, hosts, mode of transmission, and reproductive number for the major viral diseases

| Causative virus | Transmitting hosts | Mode of transmission | Reproductive number |
|-----------------------------------------------|--------------------|-----------------------------|---------------------|
| SARS coronavirus (SARS-CoV) ⁹ | Bats, Civets | Respiratory droplets | 4 |
| MERS coronavirus (MERS-CoV) ⁹ | Bats, Camels | Respiratory droplets, cough | 1 |
| Ebola ⁹ | Nonhuman primates | Contact with body fluids | 1.5 to 2.5 |
| Seasonal Influenza (H3N2, H1N1) ¹⁰ | Pig | Respiratory droplets | 2 |
| SARS-CoV-2 (ref. 11 and 12) | Bats | Respiratory droplets | 1.5 to 3.5 |



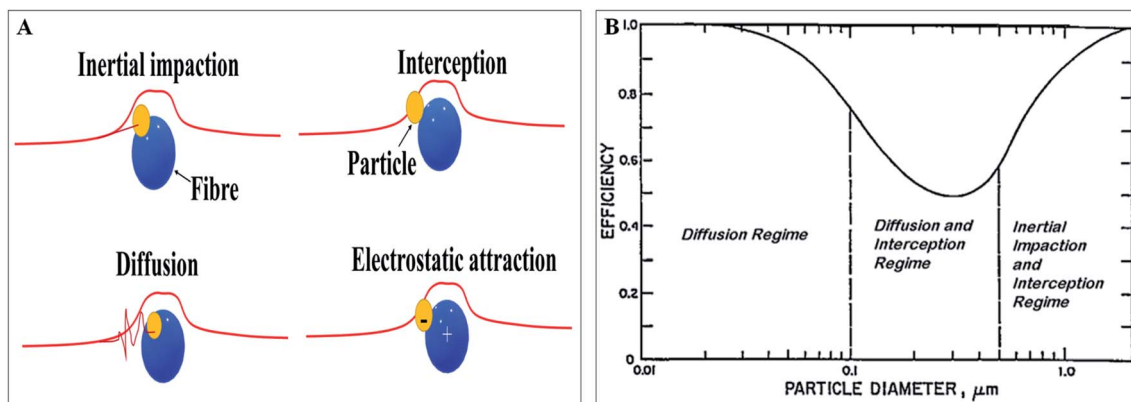


Fig. 3 (A) Particles filtration mechanisms in face masks; (B) face mask filter efficiency versus particle diameter. (B) is reproduced from the ref. 53 (open source subjected to creative common attribution 3.0 license).

and South Korea,⁵⁷ emphasising the importance of face mask in reducing community-wide transmission. The use of respiratory protection in healthcare first started gaining prominence during the 1990 tuberculosis outbreak.^{58,59} Extensive use of surgical face masks was recorded during the SARS-2003, Avian bird flu-2007 in Japan, and Swine flu-2009 in the US and Mexico.⁶⁰ It was reported that none of the health care workers in hospitals in Hong Kong and China was infected during SARS-2003, who followed appropriate and consistent use of face masks, gloves, and gowns.³⁰

Additionally, virus isolation rates were considerably reduced for 76% of the general public who wore face masks during SARS-2003.⁶¹ The air pollution incident in China in 2012 has led to the widespread manufacturing and use of N95 and KN90 respirators to filter out fine particulates.⁶² Eventually, face masks became an integral part of PPE for health workers to protect themselves from the particles.⁵³ It is now globally accepted that

usage of surgical face masks or respirators produces a lesser incidence of virus infections.⁶³ The face mask is inevitably an emerging weapon for the frontline healthcare workers in the COVID-19 war. A diagrammatic representation of the importance of face mask in reducing transmission in healthcare settings is shown in Fig. 4.

5. Evolution of face mask

The earliest records of the use of face mask-like cloth coverings are visible on doors of Persian tombs. A 13th-century travelogue describes the use of silk scarves by the servants of the Emperor of the Yuan dynasty, possibly to prevent their breath from affecting the food being prepared.⁶² The development of functional face mask-like objects was noticed during the 16th century when the Great plague hit the city of London. The mask was popularly known as beak masks and was believed to be filled with herbs such as clove, cinnamon, or camphor to mask unpleasant odours and was used by doctors.^{64,65} During the same period, it is reported that Leonardo Da Vinci promoted the use of wet clothes as respiratory filters.⁶² Before the 18th century, loose animal bladder skins and wet clothes were also known to be used to filter dust and harmful agents.^{58,66} In 1877, the Nealy Smoke face mask was patented,⁶⁷ which used a series of water-saturated sponges and a bag of water attached to the neck strap to filter out smoke. In 1899, a French doctor created a face mask out of six-layer gauze, which is considered the starting point of modern time tie or string face masks.^{62,68} These face masks were also combined with additional impervious layers and held on metal frames.⁶⁹

With the onset of chemical warfare associated with World War I, the need for better respiratory protection like an emergency escape breathing apparatus was observed.^{67,69} The spread of Manchurian plague-1910 and Spanish flu-1918 boosted the widespread use of face mask-like objects among healthcare workers and the general public.⁶⁹ The respirator certification program was initiated in 1919 by the US Bureau of Mines. In 1920, the agency certified the first respirator known as Gibbs Respirator, mainly used by miners, soldiers, and firefighters.⁶⁷ During the 1930s, single-use paper face masks were prevalent,

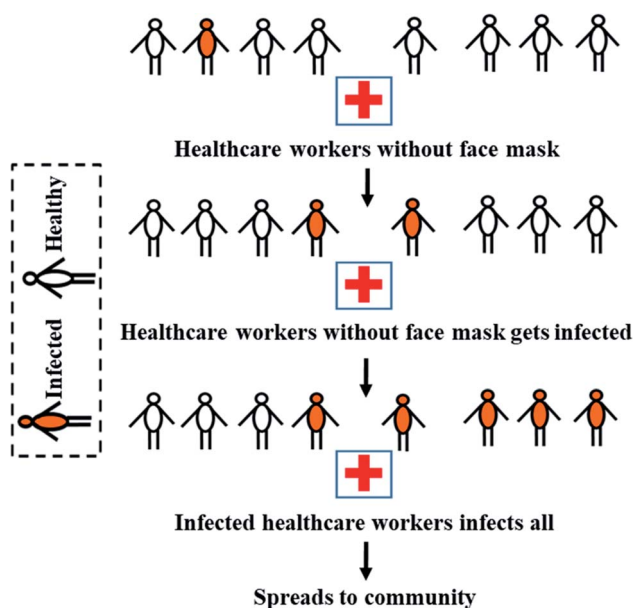


Fig. 4 Importance of wearing a face mask in healthcare settings.



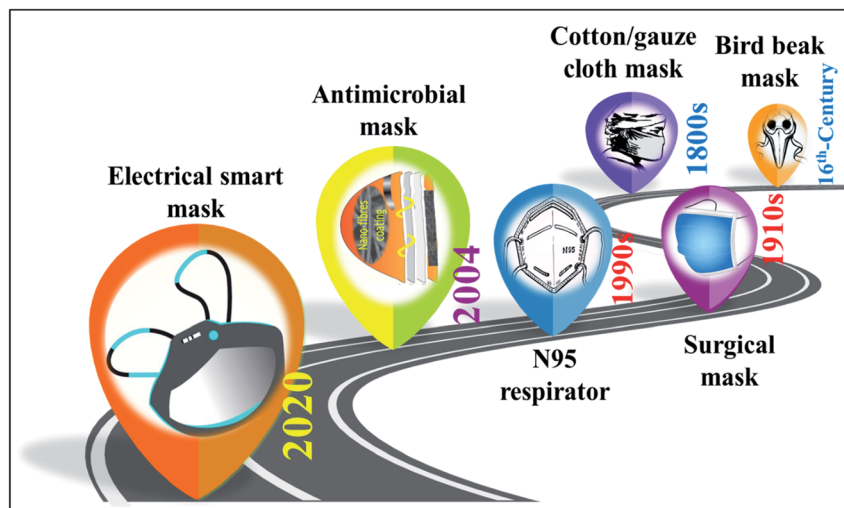


Fig. 5 Milestones in the evolution of face masks.

and towards the 1960s, disposable face masks made of fleece⁷⁰ or nonwoven synthetic fibre materials⁶⁹ came into use. This new type of face mask material facilitated massive transformation by filtering incoming and outgoing air and preventing droplet spread along with better fit and comfort.⁶⁹ Some other face masks present in the period included a layered gauze face mask with a six-inch rubber piece, a Jel face mask with gauze and filter, and a cellulose-based face mask incorporating plastacele and cotton pledgets.⁷¹

Eventually, several factors, like adequate protection, comfort, and physical convenience in wearing, service-life, and materials used, were evaluated for regulation or standardising higher quality respiratory protection.⁵⁸ Occupational Safety and Health Administration (OSHA) and National Institute of Occupational Safety and Health (NIOSH) were promulgated in the 1970s to assist in safety recommendations and regulation in the workplace, respiratory protection, and other areas of PPE. The factors such as fit and tightness of the face masks also started gaining importance during these times.

Modern face masks are typically made of nonwoven polymeric fibres of different thickness and porosity. The major types of face masks used in the market are respirators and surgical face masks. Respirators are used primarily by healthcare workers to reduce exposure to aerosols and provide leak-proof protection. Respirators should comply with the standards of NIOSH or any other internationally accepted organisations. Respiratory filters are typically made of polypropylene (PP) wool felt fabric or fibreglass paper.⁶⁶ Surgical face masks are not necessarily designed to provide proper fit or seal to the face; however, its primary purpose is to intercept larger droplets from being expelled by the wearer into the environment. They are designed to be fluid resistant and usually discarded after single-use.⁵³ Surgical face masks are typically made from PP filter layers placed between nonwoven fabrics.⁷² Polyester, fibreglass, polycarbonate, polyethylene (PE) are also employed in the manufacture.^{72–74}

The ever increasing safety concerns and need for adequate microbial protection have led to the development of

antimicrobial compound(s) infused face masks and smart electrical masks. After SARS-2003, much research and development activities have been reported in the area of antimicrobial face mask. According to the report, the Li and team have published the first article on the antimicrobial effect of nanoparticle (NP) coated face masks.⁷⁵ However, a patent filed in 2004 claims to have developed a face mask based on carbon material having anti-virus and anti-germ effect.⁷⁶ Since the incidence of COVID-19, the need for the antimicrobial face mask has increased and reached new heights. For instance, recently introduced *Guardian G-Volt* face mask exploits antimicrobial properties of graphene and electric charge to repel and deactivate microorganisms coming in contact and provide sterilisation and reuse potential.⁷⁷ The face mask technology has now advanced to include artificial intelligence and facial recognition facilities to create next-generation smart face masks. Fig. 5 summarises the significant milestones in the evolution of face masks.

6. Antimicrobial face masks

The use of a face mask as an RPD is instrumental in preventing airborne, droplet, and aerosol transmission, where the primary control mechanism lies in reducing exposure. However, there are serious concerns about the usage of conventional disposable face masks. When infected individual coughs or sneezes, the pathogen laden droplets can splash out and adheres to the immediate contact surfaces, and remain viable for several days.⁷⁸ Many studies have proven the presence of SARS-CoV-2 on various material surfaces, including inner and outer layers of face masks for a period of 4 and 7 days, respectively.^{79,80} These contaminated face mask surfaces, in turn, becomes a source of fomite transmission.⁸¹ Besides, the warm and humid conditions inside the face mask due to breathing and saliva creates a favourable environment for the intercepted microorganisms to grow and flourish. The moist conditions will induce a capillary action due to which the intercepted microorganisms transfer further into the inner layers *via* suction, thus



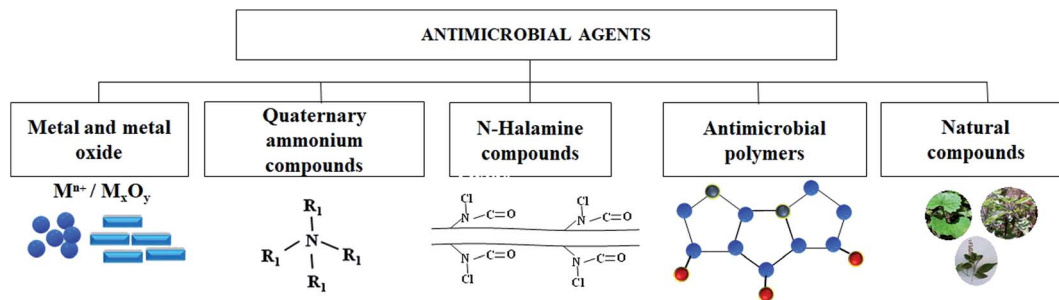


Fig. 6 Various antimicrobial agents on face masks.

endangering the health of the wearer.⁷⁵ Eventually, these microorganisms will aggregate as an extracellular polymeric matrix containing polysaccharides, proteins, and deoxyribonucleic acid (DNA) resulting in the formation of biofilms. In these situations, an unexpected danger arises due to the re-aerosolisation of the settled particles during intense sneezing or coughing by the wearer.¹⁶

Microbial survival and re-growth on conventional face masks after usage and improper storage can also lead to secondary infections in humans.^{14,15} So, face masks are typically discarded after a single-use to avoid the inoculation and spread of highly infectious pathogens. This type of single-use and discard culture can lead to its massive shortage and the generation of a large quantum of hazardous waste, especially during pandemic times.^{82,83} The gap in supply and demand coupled with unaffordability, disposal challenges, and possible adverse impact on the environment calls for reusable face masks.

Antimicrobial face masks look attractive over conventional face mask and can address some of the concerns associated with single-use face masks by providing *in situ* real-time antimicrobial protection. Several AMA are developed over the years and are available in different forms, including films, coatings, beads, and NPs. The surfaces coated AMA are reported to be effective in deactivating/killing microorganisms and preventing the formation of biofilms. The active moieties which impart the biocidal qualities include metal and metal oxides,^{75,84–95} quaternary ammonium or phosphonium groups,^{96–100} antimicrobial polymers,^{101–107} N-halamine compounds,^{108–112} antimicrobial peptides,¹⁰⁵ and natural compounds,^{82,113–121} as depicted in Fig. 6.

Given the wide choice of the AMA, a comprehensive understanding of AMA would help in the selection of right material-technology combination for the development of efficient, sustainable, and reusable face masks. The materials presented below include the AMA that are tested on the face masks and other promising materials studied for similar applications.

6.1. Metal and metal oxides

Metals such as silver (Ag),^{122–126} copper (Cu),^{84,85,87,127,128} zinc oxide (ZnO),^{95,128} tungsten oxide/carbide,^{94,129,130} magnesium peroxides,⁹¹ and their combinations are employed as AMA in fabrics or face masks.

Since ancient times, Ag is a widely used metal for antimicrobial applications. Ag, especially in nano-forms, and its derivatives are known to exhibit antibacterial, antiviral, and

antifungal activities.^{122–126} The interaction of Ag and its compounds with microorganisms is a complex process, and they are known to exhibit a multifaceted mechanism of deactivation. The prominent deactivation pathways include penetration into the cell, interaction with the thiol groups, the formation of free radicals, anchoring with soft bases such as sulphur and phosphorus compounds, and modulation of microbial signal transduction pathways.^{131,132} Various Ag and Ag-based compounds used in face masks and their potential ability in deactivating microorganisms are reviewed below.

Aloufy and El-Messiry (2013) demonstrated a simple method of soaking nylon-6 nanofibres in silver nitrate (AgNO₃) solution to yield Ag incorporated nylon-6 nanofibres. The nanofibres used in the study were fabricated by electrospinning, followed by loading with silver nanoparticles (AgNPs). A 24% (w/w) polymer yielded smooth and uniform nanofibres of diameter 150 to 250 nm. It was reported that after 1 to 2 h of incubation, *Escherichia coli* (*E. coli*) colonies were vanished from the material surface due to the antibacterial effect of Ag, as determined from the spread plate method.⁸⁹ However, Ag-loaded fabrics generally wear off with time due to the detachment of active Ag from the material or often result in agglomeration of the NPs.^{133–135} The processes such as electrospinning^{133,136} and melt-blowing^{133,137} can be employed to strengthen the attachment of biocidal components in the nanofibres. In a study, Ag has been used as an antimicrobial agent to yield an eco-friendly electrospun composite of *Acacia* lignin with polyvinyl alcohol (PVA).¹³⁸ The electrospun PVA–lignin–AgNP hybrids nanofibres were tested for antimicrobial activity using agar well diffusion assay. The nanofibre mat of 1 × 1 cm in size showed inhibition zones of 1.1 ± 0.05 and 1.3 ± 0.08 cm against *E. coli* and *Bacillus circulans* after 24 h of incubation. The study reveals that the antimicrobial activity is mainly due to AgNPs, which inhibit cell permeability and respiration. Also, the lignin, which is a phenolic compound, results in cell damage and leak of cellular components.

In another study, *in situ* AgNP incorporated polyacrylonitrile (PAN) was electrospun onto a melt-blown PP layer to develop an antimicrobial bi-layer fabric.⁹⁰ Dimethylformamide (DMF) was used as a solvent for the polymer and reducing agent for the formation of AgNPs. The developed antimicrobial bi-layer composite filter demonstrated biocidal activity against *E. coli* and *Staphylococcus aureus* (*S. aureus*). The PAN–Ag-15% fabric showed 66 and 78% reduction against *S. aureus* and *E. coli* after



incubation for 6 h, as determined from the spread plate technique. Bacterial filtration efficiency (BFE) was conducted according to American Society for Testing and Materials (ASTM) F2101.01 (ref. 139) using *S. aureus* as the biological aerosol. It was observed that the spun-bonded PP nonwoven fabric of 50 to 55 GSM (g m^{-2}), electrospinning time of 2 to 2.5 h, and 10 to 12.5% (w/w) of PAN–Ag was sufficient to achieve a BFE of 99%. A similar course of steps was followed to create a bi-layer micro-nano fibrous membrane.¹³³ *In situ* AgNP incorporated polyurethane was electrospun onto a melt-blown PP layer. DMF was used as a solvent for electrospinning polyurethane and as a reducing agent for the formation of AgNPs. Disk diffusion assays performed on the material showed large zones of inhibition against *E. coli* and *S. aureus*. The study reveals that AgNPs release Ag^+ ions, which damages the cell membrane resulting in leakage of cellular components and disruption of DNA. According to a report, electrospun or melt-blown nanofibres alone form a very soft and fragile substrate, but combining it with a nonwoven polymer substrate layer enhances the mechanical property and applicability in developing face masks.⁹⁰ Also, such a bi-layer formation helps to achieve adequate filtration performance without significant pressure drop.¹³³

In a study, the antimicrobial activity of melt-blown nonwoven PP was evaluated by incorporating Microban® (Ag-based) and Viroksan (magnesium monoperoxyphthalate), separately.¹³⁷ Microban® and Viroksan were added to the fibre forming head part of the melt-blowing machine to obtain good mixing and even distribution of the compound in the fabric during the fibre formation process. It was observed that 1% (w/w) of Microban® produced complete *E. coli* reduction in 4 h. In contrast, Viroksan 0.86% (w/w) produced complete killing immediately after the deposition on test fabric. The magnesium monoperoxyphthalate has profound storage stability, low toxicity, and is approved by the Environmental Protection Agency (EPA) for use in consumer products.¹⁴⁰ However, assessment should be carried out for determining BFE, antiviral activity, and skin compatibility to confirm the practical utility of such surface-modified face mask.

In addition to Ag immobilised on fabrics, a few studies are reported on Ag immobilised face masks. A commercially available face mask was treated with a starch capped colloidal Ag solution prepared by the chemical reduction method in a study.⁹² The face mask was soaked for 5 to 7 h in 50 and 100 mg L^{-1} colloidal Ag solution followed by drying. From the well-diffusion assay against the *E. coli* and *S. aureus*, the zone of inhibition was found to be >650 and 1000 mm^2 for 50 and 100 mg L^{-1} of Ag colloidal solution, respectively. The study demonstrated that the mask treated with colloidal AgNPs solution inhibits the growth and multiplication of Gram-positive and Gram-negative bacteria. However, the presence of broad peaks in ultraviolet-visible (UV-Vis) absorption spectra for colloidal Ag and lumps in scanning electron microscope (SEM) images revealed an uneven distribution and aggregation of Ag.

Compared to physical attachment through a dip (soak) coating and spray coating, the surface modification mediated coating renders long-term attachment of AgNPs.^{16,134,135,137} In

a study, the surface of an N95 respirator was treated with a surfactant, sodium oleate, which imparted carboxylic functionalities to the surface.¹⁶ The carboxylic sites are reported to be responsible for improved hydrophilicity and served as reaction sites for attachment for AgNPs. Microscopic observations of the fabric using Field Emission SEM after the spread plate tests revealed that the material effectively inhibited the growth of two model airborne bacterial strains, *S. aureus* and *Pseudomonas aeruginosa* (*P. aeruginosa*). However, this study mandates further works to assess the antimicrobial performance of the material and stability of the coating to evaluate the effectiveness of such surface-mediated coatings.

Metallic Cu, copper nanoparticles (CuNPs), copper(II) oxide (CuO), copper(I) oxide (Cu_2O), copper iodide (CuI) are found to be effective in inactivating human immunodeficiency virus, influenza virus, bacteriophages, H9N2 virus, poliovirus, bronchitis virus, and herpes simplex virus.^{60,87,127} Cu ions interact with the negatively charged bacterial cell membrane through electrostatic interaction. The inactivation is through redox cycling between Cu^{2+} and Cu^+ at the cell surface, which catalyses the hydroxyl radicals and superoxide anions. The generation of free radicals damages the lipids, protein, and DNA of bacteria and viruses.⁸³ The antiviral activity of Cu is because of the interaction of Cu^{2+} with DNA, where the nucleotide and the base pair of the DNA offer the binding sites.¹⁴¹ The ROS formed during redox recycling also results in the degradation fragment of viral proteins, hemagglutinin and neuraminidase.⁸⁷ It is said that the virucidal activity of Cu is enhanced in the presence of peroxide. Peroxide opens the protein coat of the virus and facilitates the entry of Cu ions to denature nucleic acids.^{85,87} Cu ions also interfere with the cell permeability resulting in the loss of potassium ions and cell death.⁸³

Cu oxo-hydroxide NPs is reported to be better virucidal agents than other Cu combinations.⁹³ The stability of CuNPs could be improved by functionalisation using carboxylate ligands. CuO and Cu oxo-hydroxide can be coated on face masks, surgical gowns and gloves, respirator filters, polymeric materials, and other medical equipment fabrics without losing its antiviral property.^{84,87,93,142} A four-layered NIOSH approved N95 respirator was modified with CuO particles to impart antimicrobial property.⁶⁰ The first and fourth-layers were prepared using spun-bonded PP impregnated *in situ* with 2% (w/w) CuO particles. The second-layer was made up of a melt-blown PP with 2.2% (w/w) of CuO particles, and the third-layer was fabricated with polyester. The bioaerosol filtration tests revealed that 99.85% of H1N1 and H9N2 viruses were filtered under simulated breathing conditions with no infectious viral titers recovery within 30 min, by employing a method customised from ASTM F2101.01 for the virus.¹³⁹ Moreover, the CuO coated face mask successfully passed BFE as per EN 14683:2005 and ASTM F2101 standards.^{139,143} The modified face mask also passed differential pressure (EN 14683:2005),¹⁴³ latex particle challenge (ASTM F2299),¹³⁹ and resistant to penetration by synthetic blood tests (ASTM F1862)¹⁴⁴ and NIOSH N95 standards. Furthermore, it was claimed that no skin sensitisation or irritation was observed for the outer layers of the face mask as determined from animal studies.



In yet another study, Hashmi and coworkers (2019) reported a polymer solution made from PAN (8% w/w) in DMF and CuO. The composite solution was electrospun to develop an antimicrobial face mask. Disk diffusion assay showed that CuO (1% w/w) incorporated face mask was effective against both *Bacillus subtilis* (*B. subtilis*) and *E. coli*. Based on the breathability test (ASTM E96),¹⁴⁶ air permeability test, and antimicrobial activity test, the authors claimed that the face mask is useful for breath mask applications. The nanofibre structure is intact even after contact with water for 72 h, and a concentration beyond 1% (w/w) of CuO is toxic.¹⁴⁵

Compared to individual metal and metal oxides, combinations with other metal and metal oxides have shown complete inactivation in lesser contact time.^{75,88} A combination of different AMA also reduces the chances of resistant strains of microorganisms.⁷⁵ In 2012, Davison developed BioFriend™ and BioMask™ masks, which comprised four layers of standard filtration materials approved for facemasks.⁸⁸ The first and fourth-layer were made up of spun-bonded PP. The first-layer was treated with citric acid to create a low-pH hydrophilic coating, which can induce structural rearrangements in the lipid layer of viruses. The second-layer was fabricated with cellulose/polyester infused with Cu and zinc ions, which can bind the virus through sulfhydryl and carboxyl groups, destroying it. The third-layer was made up of melt-blown PP. The outer surface of both the mask was challenged with bacterial (initial 10^5 to 10^7 CFU mL⁻¹) and viral (initial 5.35 to 7.79 log₁₀ TCID₅₀) suspensions for 5 to 180 min. The samples were then tested for the remaining virus titers using the Spearman–Kärber method. Bacterial colonies were tested according to the American Association of Textile Chemists and Colorists (AATCC)-100 method.¹⁴⁷ These masks showed reductions of 99.93% @ 30 min, 89.22% @ 60 min, 90.15% @ 60 min, 88.97% @ 60 min, 99.99% @ 8 h against methicillin-resistant *S. aureus*, *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Mycobacterium terrae*, and *Staphylococcus epidermis* (*S. epidermis*), respectively. These masks showed ≥ 4 log₁₀ reductions within 5 min of contact for 19 different subtypes of viruses, including influenza A and B viruses, paramyxovirus, SARS-CoV, and herpes simplex virus. Both the face masks conformed to ASTM2100:11/EN14683:2005 (ref. 143 and 148) and EN 149:2000+A1:2009 (ref. 149) standards. Additionally, both the face masks demonstrated no dermal irritation or sensitisation and allergenic potential under simulated breathing conditions.

In another study, the first-layer of a surgical face mask fabric was coated with 0.4 mg cm⁻² NP emulsion consisting of a mixture of 2 g L⁻¹ aluminium oxide, 2 g L⁻¹ AgNO₃, 1.5 g L⁻¹ titanium dioxide, 2 g L⁻¹ Ca²⁺, 2 g L⁻¹ Mg²⁺, and 60 g L⁻¹ oleophobol C employing a textile finishing machine.⁷⁵ Oleophobol C was added to ensure homogenous dispersion of the NP emulsion on the face mask surface. Minimum inhibitory concentration (MIC) and minimum bactericidal concentrations (MBC) for *E. coli* were found to be 1/128. The MIC and MBC for *S. aureus* were found to be 1/512 and 1/64 dilutions of the NP emulsion, respectively. The antibacterial tests revealed a complete reduction of *E. coli* and *S. aureus*, and no skin

irritation was observed for volunteers who wore the face mask. The higher MBC for *S. aureus* can be attributed to the thick peptidoglycan layer for the Gram-positive bacteria. It was concluded that the biocidal effect arises upon damage to the plasma membrane and leakage of cytoplasmic contents. This study claims to be the first report on the antimicrobial effect of NP coated face mask.

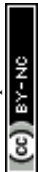
Apart from the above-discussed metal and metal oxides or their combinations, tungsten oxide,⁹⁴ magnesium peroxide,⁹¹ and ZnO⁹⁵ are also antimicrobial. According to a report, magnesium peroxide coated cotton exhibited antimicrobial activity up to 70 washing cycles.⁹¹ It is reported that tungsten oxide and tungsten oxide composite microparticles are also used on face masks, filters, gloves, and other medical appliances.⁹⁴ Studies have proved excellent antiviral property of zinc oxide nanoparticles (ZnONPs) and polyethylene glycol (PEG)-coated ZnONPs against influenza virus strains.⁹⁵ However, the stability and effectiveness of these compounds need to be evaluated to establish its practical utility.

6.2. Antimicrobial polymer

Antimicrobial polymers are a versatile class of material with tunable surface chemistries, size, and multiple scale lengths; that can be combined with different matrices or surfaces and tailored to yield contact-killing abilities.¹⁵⁰ There are two broad classes of antimicrobial polymers *viz.*, bio-passive and bio-active. Bio-passive polymers are used for repelling surfaces to prevent bacterial adhesion. The polymers functionalised with bio-active agents are called bio-active polymers, and these are capable of killing the microorganisms adhered to the polymer surface.^{150,151} The active moieties include metals, metal oxides, N-halamines, quaternary ammonium or phosphonium compounds, antimicrobial peptides, and antibiotics.¹⁵¹

6.2.1. Polymers with N-halamines. N-halamine is one of the most effective contact-active biocide tested in polymers. The halamine bond (*i.e.* the nitrogen–halogen covalent bond) can effectively inactivate a broad spectrum of microorganisms in a short time (5 to 30 min) on direct contact.^{110–112,152} The oxidative transfer of the halogen to the specific biological receptors like amine and thiol groups in the protein interferes with the enzymatic and metabolic process, resulting in cell lysis.^{110,153,154} The oxidative transfer of halogen can also result in electrophilic substitution of halamine bond with hydrogen.¹⁵⁵ Once the halogen is depleted, the antimicrobial activity can be regenerated by treating the material with halogen donor compounds such as hypochlorite, sodium hypobromite, trichloroisocyanuric acid, and sodium dichlorocyanurate.^{153,156} The polymers such as polyvinyl acetate, PP, nylon, polystyrene (PS), PAN, polyethylene terephthalate (PET), and polyamide are compatible with N-halamine.

In a study, a simple soaking method was employed to modify the PP surface with N-halamine.¹¹⁰ The melt-blown PP nonwoven-fabrics (22 and 50 GSM) were soaked in N-halamine 1-chloro-2,2,5,5-tetramethyl-4-imidazolidinone (MC), for 10 min and then padded and dried. The antimicrobial PP fabric with 0.47% (w/w) oxidative chlorine loading showed complete



inactivation ($6 \log_{10}$) of *S. aureus* and *E. coli* in 5 and 10 min of contact time for 50 and 22 GSM fabric, respectively as determined from the spread plate technique. The higher GSM fabric provided greater surface area, higher chlorine availability, and rapid antimicrobial activity. Further, the bioaerosol filtration studies conducted as per ASTM F2101.01 (ref. 139) revealed that no viable cells were recovered up to 3 h exposure against *S. aureus* and *E. coli* aerosol. However, the N-Cl bond is prone to photo-dissociation. Hence, all the N-halamine coated fabrics are recommended for dark storage. A similar soaking, padding, and drying procedure were followed to coat PET fabric with 5% (w/w) terpolymer, poly(hydantoinyl acrylamide-co-glycidyl methacrylate-co-2-(methacryloyloxy)ethyl trimethylammonium chloride). After soaking, the fabric was halogenated using 10% (w/w) Clorox solution (bleach), followed by washing and drying. The material demonstrated a complete reduction of *S. aureus* and *E. coli* cells ($6 \log_{10}$ reduction) within 2 min of contact time for swatches having 0.19% (w/w) oxidative chlorine concentration, as determined from the spread plate technique.¹⁴¹ The rapid inactivation was partly due to the hydrophilicity imparted by the cetyltrimethylammonium chloride groups.

The incorporation of N-halamine precursors through graft copolymerisation resulted in better and permanent coating compared to physical coating through blending, soaking or spraying.^{109,152} In a study, N-halamine precursors such as acrylamide, and methacrylamide were attached separately to the PP through graft copolymerisation.¹⁰⁹ The modified PP was processed into fibres through an extrusion process. The fibres were then halogenated by soaking in chlorine bleach ($\sim 1500 \text{ mg L}^{-1}$ of available chlorine) followed by washing and drying. The material demonstrated a complete reduction of *E. coli* within 30 min, tested as per AATCC-100 method.¹⁴⁷ Also, both antimicrobial fabrics demonstrated $>1.48 \log_{10}$ reduction against *S. aureus*.

In a study by Tan and Obendorf (2007), N-halamine additives [chlorinated 5,5-dimethyl hydantoin (CDMH), chlorinated 2,2,5,5-tetramethyl-imidozalidin-4-one (CTMIO) and chlorinated 3-dodecyl-5,5-dimethyl hydantoin (CDDMH)] were separately incorporated into nylon-6 dope and electrospun to develop nanofibrous antimicrobial fabrics.¹⁰⁸ Nylon 6-CDMH, nylon 6-CTMIO, and nylon 6-CDDMH fabrics with an active chlorine content of 1125 mg L^{-1} achieved a total reduction of *E. coli* and *S. aureus* within 10, 30, and 30 min, respectively (tested as per AATCC-100 method).¹⁴⁷ The difference in the inactivation is due to the N-Cl bond of the three compounds. The dissociation rate is in the order of imide $>$ amide $>$ amine type halamine. In another study, N-halamine compound MC was introduced into PAN polymer dissolved in DMF solvent and electrospun to yield nanofibres (PAN/MC-5%).¹¹² From the antibacterial assays, it was found that PAN/MC-5% with 0.29% oxidative chlorine loading inactivated all *S. aureus* and *E. coli* cells within 1 and 10 min of contact, respectively, tested as per AATCC-100 method.¹⁴⁷ Also, PAN/MC-5% achieved $>6 \log_{10}$ reduction against *S. aureus* aerosols after 3 h of exposure, tested as per ASTM F2101.01.¹³⁹ Moreover, the breathability of these membranes was reported to be 27.3 mm s^{-1} , which is higher than the commercial nano-N95 and nano-surgical face mask.

In contrast to physical coating methods, covalent attachment and *in situ* incorporation *via* electrospinning eliminate the chances of leaching of N-halamines from the fabric surface.^{108,109,152,157} Even though few studies are reported on the leaching of N-halamine modified fabrics in water,¹⁰⁸ the direct release of N-halamine from fabric or face mask surface is yet to be evaluated to assess its effect on the skin. Since the AMA generally coated on middle filtration layers of face mask, direct contact with skin is unlikely. Though studies reveal the excellent antibacterial activity of N-halamine incorporated fabrics, there is no literature on application of this compound in developing antimicrobial face mask. Performing antibacterial and antiviral assays, BFE and VFE would establish the practical utility of N-halamine incorporated face mask.

6.2.2. Polymers with quaternary ammonium compounds.

QACs belong to the class of cationic surfactants with good biocidal ability. The antimicrobial activity of QACs is a function of the N-alkyl chain length, and QACs with 8 to 18 carbon atoms possess good germicidal activity.¹⁵³ The positive charge on QACs facilitates the penetration of these cationic biocides into the microbial cells. The antiviral property of QACs is due to the high affinity of QACs to the proteinaceous coat and lipid nature of the viral envelope.^{153,158,159} The mechanism of action is as follows: (i) adsorption and penetration of the compound into the cell membrane, (ii) interaction with the phospholipids present and affecting cell permeability, (iii) leakage of intracellular components like potassium, phosphates and then large molecular weight molecules, (iv) damage of protein and DNA, and (v) cell lysis by the autolytic enzymes.

Studies have reported the immobilisation of QACs on glass surfaces,¹⁶⁰ polymer surfaces,¹⁶¹ surgical face masks,¹⁰⁰ and nonwoven fabrics^{96,98} through surface-mediated coatings, grafting, melt-blowing and spray coating to kill/deactivate airborne and aerosol borne microorganisms. QACs such as cetyltrimethylammonium bromide (CTAB) can form reactive oxygen species (ROS) and increases the superoxide mediated stress resulting in biocidal activity.¹⁶² Superoxide anions, hydrogen peroxide, and hydroxyl radicals are produced by penetrating oxygen into the cells during the intracellular enzymatic reactions. Montazer *et al.* (2010) developed antimicrobial polyester, PP, and viscose nonwoven fabrics coated with CTAB and fluorochemical using a padding procedure (bath coating).⁹⁹ PP and polyester exhibited $6 \log_{10}$ reductions of *E. coli* and *S. aureus* and $>2 \log_{10}$ reductions of *P. aeruginosa* with 1% CTAB solution, as determined from the spread plate technique. In contrast, nonwoven viscose fabric was unable to achieve the expected results due to the negative influence of adhesive used in fabric production. It is reported that fluorochemical was added to reduce the hydrophilicity of the surface to achieve good water or blood repellent property.

In another work, Gutarowska and coworkers (2014) studied the antimicrobial effect of bath (soak) and spray coating of Sanitised® T 99-19 (commercially available QAC) on needled nonwoven polyester, and melt-blown nonwoven PP.⁹⁶ Sanitised® T 99-19 was sprayed and soaked to obtain 0.7 to 2% deposition on polymer fibres. Better antimicrobial activity is obtained for needled nonwoven polyester due to higher



migration of active compound to fibre surface when compared to melt-blown nonwoven PP. It is attributed that the compact surface of a melt-blown nonwoven PP facilitates the only surface distribution of the active compound. The bathing method offered better antimicrobial activity than spray coating since it wets the entire volume of the material. It is reported that needled nonwoven polyester subjected to bath coating with 2% of Sanitized® T 99-19 was biocidal against *E. coli*, *S. aureus*, *Candida albicans* (*C. albicans*), and *Aspergillus niger* (*A. niger*), tested as per AATCC-100 method.¹⁴⁷ Whereas, for melt-blown nonwoven PP, the biocidal values were not much significant. Further, paraffin oil mist and airflow resistance of the material was tested according to the EN 149:2001+A1:2009 standards.¹⁴⁹ However, bioaerosol filtration studies have to be performed to understand the efficacy of the material for its applicability in preparing an RPD.

Studies have shown that perlite (volcanic glass) carriers could enhance the hydrophilicity, antimicrobial activity, and flexibility of PP to bond with various chemical agents.^{98,163} Bio-perlite and bio-bentonite refer to alkylammonium microbicides (QAC compound) on perlite and bentonite (aluminosilicate base) carriers, respectively. These compounds were used in a study to impart biocidal quality to PP fibres.¹⁶³ The compounds were added in powder form to the fibre forming head to obtain thorough mixing with PP during the fibre formation process. Subsequent melt-blowing resulted in uniform distribution of the biocidal agent on the nonwoven PP. It is reported that melt-blown PP with 15% bio-perlite (0.69% of microbicide) is biocidal and achieved 96.8% and 74.1% BFE against *E. coli* and *S. aureus*, respectively. The study utilised the electrostatic interaction between the positively charged microbicide and the negatively charged mineral carriers to create bio-perlite and bio-bentonite. The mineral carriers remained attached to the fibres, while the microbicide remained free and capable of antimicrobial activity. The study was further extended to explore the antimicrobial respiratory protection using a three-layer half-mask model by Gutarowska and coworkers in 2010.⁹⁸ The first-layer was made from needling nonwoven fabric consisting of Bico biocomponent (Inatex, Poland) and an acrylic fibre (Unia, Poland). The second-layer was an antimicrobial filter layer made of nonwoven PP with bio-perlite. The third-layer was made from needled nonwoven polyester and Bico biocomponent. The filtering half-mask with a bio-active nonwoven PP with 0.37% of the active compound (8% bio-perlite) achieved a complete reduction of *E. coli* and *S. aureus*, tested as per AATCC-100 method.¹⁴⁷ The bio-active nonwoven PP fabrics were also effective against *P. aeruginosa* and *K. pneumoniae*, *Mariniluteicoccus flavus*, and *B. subtilis*. Further, it was established that the filtration efficiency of the material was excellent against liquid particles (>95% against paraffin oil mist aerosol), solid particles (>98% against 0.6 µm NaCl aerosol), and microorganisms (99.9% for *E. coli* and *S. aureus* suspended in 0.85% NaCl solution).

Majchrzycka *et al.* (2017) developed a biocidal agent, namely Gemini surfactant (QAC-based salt) on a halloysite carrier that exhibited temporal biocidal activity triggered by humidity.¹⁶⁴ Varying amounts of 1,2-propanediol was added to regulate the

humidity, resulting in temporal activation of the biocidal agent. The biocidal agent was added to the semi-liquid polymer during the melt-blowing process of PP, resulting in a permanent bonding. The material with 0.336% of the Gemini surfactant exhibited 97.46, 87.85, 94.53, and 46.78% reduction against bacteria *E. coli*, *Pseudomonas fluorescens*, moulds *A. niger* and *Penicillium chrysogenum*, respectively, tested as per AATCC-100 method.¹⁴⁷ This filtering nonwoven PP is a potential material in producing reusable RPDs. Further, the temporal release of the particular biocide is impressive from safety aspects as it would reduce the skin contact upon continuous use of RPDs.

Tseng *et al.* (2016) modified a US Food and Drug Administration (US FDA) approved three layers surgical mask by spraying QAC based Goldshield-5 solution (GS5, 1%), which gave 0.1 mL cm⁻² coating.¹⁰⁰ The GS5 coated face mask was challenged with settled aerosol through an aerosol test system (tested with an aerosol test system modified from ASTM 2721-10¹⁶⁵), which generated 10⁸ CFU mL⁻¹ of bacterial strains. The test results showed a 2.32 to 5.02 log₁₀ reduction of colonies in 60 min contact with GS5 coated mask for *Acinetobacter baumannii* (*A. baumannii*), *Enterococcus faecalis* (*E. faecalis*), and *S. aureus*. Further bioaerosol filtration study revealed >98% efficiency against all the three bacterial aerosols for 60 min contact time. However, in the study, simplified spraying method is used for coating GS5. Spray coating would not ensure complete penetration of antimicrobial compound into the crevices or shadowed regions, thus warranting proper investigation into coating efficiency and stability of the coating. Also, various QAC compounds are known to be corrosive and pose health issues such as work-related asthma and skin dermatitis.^{166,167} Therefore, a detailed evaluation of the biocompatibility is recommended.

6.2.3. Miscellaneous groups on polymers. Polyethyleneimine (PEI) is a synthetic, non-biodegradable, cationic polymer containing primary, secondary, and tertiary amine functional groups. Microbial cell rupture results from the electrostatic interaction between cationic PEI and the negatively charged bacterial outer cell membranes. In a study, cotton, wool, nylon and polyester were modified by derivatives of N-peralkylated PEI to render them antimicrobial.¹⁰³ Aerosolised suspension of *S. aureus*, *S. epidermis*, *E. coli*, *P. aeruginosa*, *C. albicans*, and *Saccharomyces cerevisiae* of 10⁵ cells per mL concentration were sprayed onto the modified textiles and placed in agar and autoclaved to detect any growth of cells. It was reported that the antimicrobial fabric showed at least 1 to 2 log₁₀ reduction of the deposited aerosolised bacteria. In another study, the replaceable cellulosic filter layer of a commercial face mask was modified by dip coating with PEI to induce antiviral activity.¹⁰⁷ Filtration tests on the aerial suspension of the T4D bacteriophage virus of *E. coli* B showed 5 log₁₀ reductions within 1 h contact time, using a laboratory aerosol filtration setup with an effective filtration area of 15.9 cm². Further, to determine the filtration efficiency against the H5N2 virus, a separate filtration setup was created, which was authorised. Two layers of these PEI modified nonwoven cellulosic fibre filters were reported to block the droplet-borne/airborne influenza A (H5N2) virus. However, the study can be extended to determine whether the



captured viruses are deactivated by the presence of PEI using antiviral assays. Air diffusion study showed that PEI was still intact on this filter layer after 5 h use under normal breathing conditions.

In another study, polyester fabrics were incorporated with polypyrrole nanoparticles and investigated for antimicrobial activity.¹⁰⁶ The antimicrobial activity of polypyrrole is due to the positive charges introduced along its backbone during the polymerisation process.¹⁶⁸ Dispersions of stable polypyrrole nanoparticles, produced by chemical oxidative polymerisation, were coated on polyester using an ultrasound-assisted coating process. A polypyrrole loading of $4.33 \pm 0.13 \text{ g m}^{-2}$ on polyester showed a complete reduction of *S. aureus* and *E. coli* cells, as determined from the spread plate technique.

Hong and Sun (2009) probed the incorporation of photosensitisers such as benzophenone (BP) into polymers and textiles to render them antimicrobial. In the study, polymer solutions of PP, PE, PVA, and PS were thoroughly mixed with BP solution and melted on glass slides and cast into films separately.¹⁶⁹ Initially, the microbial suspension was placed on the film and subjected to irradiation with UV light for 1 h. Bacterial concentration remaining was determined by the conventional spread plate technique. Complete reduction of *E. coli* and *S. aureus* cells was achieved for PVA/BP (0.25% w/w), PE/BP (1% w/w), and PS/BP (1% w/w). Among various polymers tested, PVA showed the highest antimicrobial activity. Polymers such as PVA with BP groups are potential self-decontaminating material with proven application in hygiene gloves, protective face masks, and surgical gowns. When BP is photo-irradiated, it forms BP ketyl radicals by accepting hydrogen atoms due to the interaction of amines and alcohols groups present in the supporting polymer (*i.e.*, PVA). Upon exposure to oxygen, the BP ketyl radicals immediately reoxidised to form BP and H_2O_2 . The formed reactive radicals and H_2O_2 play a significant role in the antimicrobial action.¹⁶⁹ However, the production of reactive radicals and H_2O_2 is of concern. Hence, biocompatibility and skin sensitisation tests have to be conducted to ensure the safety of the wearer.

Natural host defence peptides (HDPs) are part of our innate immune system, which deactivates a broad spectrum of pathogens, playing a vital role in the host response system. Biocidal cationic polymers inspired by HDPs are biocompatible,¹⁰⁵ do not possess cytotoxicity and ecotoxicity,¹⁷⁰ and has excellent applicability in antimicrobial surfaces.¹⁷¹ An antimicrobial nanofibrous substrate was prepared by grafting L-cysteine onto PP, followed by electrospinning to form fibres and cross-linked by Cys-LC-LL-37.¹⁰⁵ Antibacterial assays on the fabric were performed according to ASTM E 2180-01 standard test method¹⁷² and showed 1.6 and 6 \log_{10} reduction against *P. aeruginosa* and *S. aureus* after 24 h contact time. The material was reported to be effective against nosocomial pathogens and their spread to the community. Due to their biocompatibility, antimicrobial peptides could be potential candidates for developing novel antimicrobial face masks with least environmental concern.

Apart from the aforementioned groups, natural polymers, polymers with aromatic or heterocyclic groups (imidazole derivatives), polyacrylamides, polyacrylates, poly-siloxanes,

poly-ionenes, poly-oxazolines, hyperbranched and dendritic polymers (poly(amidoamine) and poly(propylene imine) dendrimers) and polymers with guanidine groups, possess intrinsic antimicrobial qualities.¹⁵³ The antimicrobial abilities of these groups after incorporating on face mask are yet to be evaluated.

6.3. Natural compounds

Several antimicrobial compounds extracted from medicinal plants are found to be effective against bacteria and viruses. Unlike chemical polymeric or nano-based materials, the natural compounds do not produce toxic effects. Extracts of *Vitex trifolia*,^{173,174} *Punica granatum*,^{175–177} *Allium sativum*,^{178–180} *Acacia nilotica*,¹⁸¹ *Andrographis paniculata*,^{182,183} *Sphaeranthus indicus*,¹⁸⁴ *Strobilanthes cusia*,^{185,186} *Chromolaena odorata*,¹⁸⁷ *Aloe barbadensis*,^{188–190} and *Azadirachta indica*^{113,191} have antimicrobial properties. Also, the combination of *eugenol*, *eugenol acetate*, *carvacrol*, *thymol*, and *vanillin*,¹¹⁵ *Melaleuca alternifolia*,¹¹⁶ ginkgo leaf extract,¹¹⁷ and ginkgo extract in combination with sumac (*Anacardiaceae* family)¹¹⁸ have been studied for their antiviral properties. These compounds are found to be potential agents that could be applied to face mask, air filters with plastic and nonwoven polymer fabrics.

Mangosteen extracts (α -mangostin (46.36%) and γ -mangostin (5.45%)) were spray-coated on commercially available melt-blown PP layers to develop a three-layer antimicrobial mask with improved hydrophilic character.¹¹⁹ BFE was conducted as per ASTM F2101.01 (ref. 139) and was reported to be $97.9 \pm 0.2\%$ for the face mask containing 5% (w/v) mangosteen extract. Antimicrobial studies as per AATCC-100 method¹⁴⁷ revealed a 5 and 2 \log_{10} reductions in *S. aureus* and *Mycobacterium tuberculosis*, respectively, after 24 h exposure. The 97.2% antimicrobial activity of the extract was retained even after 21 days of storage and did not compromise the mechanical attributes of the filter. The coated face mask did not exhibit any toxicity as revealed from indirect cytotoxicity studies based on the viability of L929 mouse fibroblasts. In mangosteen extract, α -mangostin and γ -mangostin are the bioactive components, which inhibits the bacterial activity by inducing rupture of the cytoplasmic cell membrane resulting in leakage of intracellular components.¹¹⁹

A recent study reports the development of a three-layer antibacterial face mask consisting of a nonwoven PP outer layer, antibacterial filter layer and an inner medical cotton yarn layer.¹²⁰ Microcapsule made from the extract of *Scutellaria baicalensis* was used as an antimicrobial agent in face mask. The microcapsules were incorporated to filter clothes such as cotton, polyester and nonwoven PP by soaking followed by plasma mediated surface modification. Antibacterial studies as per AATCC-100 method¹⁴⁷ revealed that all filter clothes exhibited >96% and >95% reduction against *E. coli* and *S. aureus*, respectively. Further, nonwoven PP, cotton, and polyester exhibited BFE of 97.31, 95.44, and 91.7%, respectively. It was observed that oxygen plasma surface treatment helped in the long-term attachment of microcapsules (up to 100 cycles) on filter clothes when compared to the untreated surface. It is reported that the antibacterial activity of *Scutellaria baicalensis* is



due to the bioactive components present like wogonin, skullcapflavone, baicalein, and baicalin.¹⁹² However, its mechanism of action is not studied in detail.¹⁹³

In another study, oil of a Vietnamese medicinal plant, *Folium Plectranthii amboinicii*, and sustainable filter paper was used to develop a seven-layer antimicrobial face mask.⁸² The first, second, sixth, and seventh-layer were made from filter paper; third and fifth-layer with blotter paper; and fourth-layer with waterproof paper. The third and fifth-layers contained activated carbon, 5% standardised BSI (Benzoic acid, Salicylic, and Iode) solution and 0.5% *Plectranthii amboinicii* plant oil extract. This face mask is reported to be qualified by the Directorate for Standards, Metrology, and Quality of Vietnam criteria. The fractional concentration of nitric oxide in exhaled breath (FE_{NO}), a relevant inflammatory biomarker of the respiratory system, was found to be significantly lower ($p = 0.01$) after the usage of the face mask. Also, the wearing of the mask resulted in clearer nasal passages ($p < 0.0001$), fewer throat symptoms ($p < 0.020$), and fewer respiratory symptoms ($p < 0.020$). The antibacterial effect of *Plectranthii amboinicii* is accounted due to terpinen-4-ol and carvacrol, which inhibits bacterial proliferation.¹⁹⁴

Chen *et al.* (2020) explored the suitability of biocompatible amino acid-grafted enzymatic hydrolysis lignin derivatives with both cationic/anionic groups as AMA for face masks. Antimicrobial tests on the middle layer of commercial N95 face mask soaked in arginine modified enzymatic hydrolysis lignin (33% w/w) revealed that the number of viable *E. coli* cells decreases to a non-measurable level from an initial load of 10^7 CFU mL⁻¹, as determined by a method of the National Committee for Clinical Laboratory Standards Institute.¹²¹ The enzymatic hydrolysis lignin has plenty of phenolic and carboxylic groups which disturbs the bacterial cell membrane. Besides, the grafting of amino acid groups enhances the positive charges and causes cell membrane rupture.¹²¹

Catel-Ferreira *et al.* (2015) developed a novel bio-based antiviral face mask through the laccase enzyme-mediated modification of the nonwoven cellulosic fibre filter with catechin polyphenols. The modified filter targets the viruses and produced 5 log₁₀ reductions against T4D bacteriophage in 1 h, determined using a laboratory aerosol filtration setup with an effective filtration area of 15.9 cm². According to the report, the best virus capture factor was obtained when two numbers of this modified cellulosic filter are placed inside the medical face mask. This bio-based antiviral face mask also showed excellent antiviral activity against the T4D bacteriophage virus of *E. coli* B.¹⁹⁵

6.4. Miscellaneous agents

Quan *et al.* (2017) found that saline coatings can enhance the hydrophilic character of face mask surfaces, which promotes microbial adhesion.¹⁹⁶ NaCl was incorporated as an active moiety in the middle PP layer of a three-ply surgical face mask in the study. Saline loading of 18.6 mg cm⁻² showed 85% filtration efficiency against H1N1 influenza virus aerosol (2.5 to 4 μm), using a laboratory aerosol filtration setup. Saline coated filters demonstrated complete hemagglutinin activity (HA) loss of adsorbed virus (physical destruction of the virus) within

5 min contact time. *In vivo* experiments on mice exposed to viral aerosol derived from saline coated filters showed higher survival against the H1N1 influenza virus and other strains, including A/Puerto Rico/08/1934 (PR/34 H1N1) and A/Vietnam/1203/2004 (VN/04 H5N1). Further, it was observed that the saline remains coated on the filters retaining its performance even after storing for 15 days at 37 °C and 70% relative humidity. Even though a change in grain orientation was observed, the stability of coating was ensured even under harsh environmental conditions. When bioaerosols come in contact with salt, it will cause the salts to dissolve locally and the osmotic pressure imbalance happens to the bioaerosol. Due to evaporative drying, the salt concentration increases and attains super-saturation, resulting in recrystallisation of salt. The physical damage due to membrane perturbation and protein denaturation during recrystallisation and osmotic imbalance results in inactivation of the bioaerosol. The physical damage is linked to the HA activity loss of the virus in the above study.^{196,197}

Iodine, along with an element in period 4 to period 6 and group 8 to group 15 (Cu, Ag, antimony, iridium, germanium, tin, titanium, palladium, bismuth, gold, iron, zinc cobalt, nickel, indium and mercury) are active antiviral agents. It is reported that these antiviral agents can be immobilised on fibres of face masks, filters, clothes, and polymers such as polyester, PE, PET, PAN, PP, PVA, polyvinyl chloride (PVC), polytetramethylene terephthalate, polybutylene terephthalate polyamide, polyacrylic acid, polytetrafluoroethylene, kevlar, poly(methyl methacrylate), cupra, polynosic, tinsel, rayon, wool, cotton, silk, hemp, and bamboo to yield potential antimicrobial fabrics.¹⁹⁸ Heimbuch *et al.* (2006) incorporated a thin coating of iodine releasing polymer on P95 face masks and found 4.3 log₁₀ reduction in MS2 coliphage.¹⁹⁹ Iodine based antimicrobial masks such as iodine-based flat-panel N95, iodine-based cup-shaped P95, and NIOSH approved *SafeLife* N95 filtering face-piece respirator (FFR) with Triosyn (iodine) agent have been studied for their antimicrobial efficacy.^{9,10} In a study, it was observed that both the antimicrobial FFR, *i.e.* iodine-based flat-panel N95 (AM-N95) and iodine-based cup-shaped P95 with exhalation valve (AM-P95) demonstrates no significant difference in physical and viable penetration tests. This implies that the performance is based on physical effects rather than antimicrobial qualities.⁹ Both the iodine-based antimicrobial FFR did not show enhanced performance over conventional N95 and P100 FFRs.

Since the discovery of graphene in 2004, it has been a widely used nanomaterial for several applications, including antimicrobial application. The first report on the antimicrobial activity of graphene was published in 2010. According to data, 85 μg mL⁻¹ of graphene oxide has shown a 98.5% reduction in *E. coli* load from an initial load of 10^7 CFU mL⁻¹.²⁰⁰ The antimicrobial activity of graphene is attributed to the physical destruction of the cell membrane upon contact with the sharp edges of graphene. According to Kumar *et al.* (2019), the generation of ROS and the consequent exertion of oxidative stress that damages the proteins and lipids of the cell membrane is the major pathway of antimicrobial action.²⁰¹ Among various nanoscale carbon derivatives, graphene,²⁰² graphene oxide,^{203,204} and



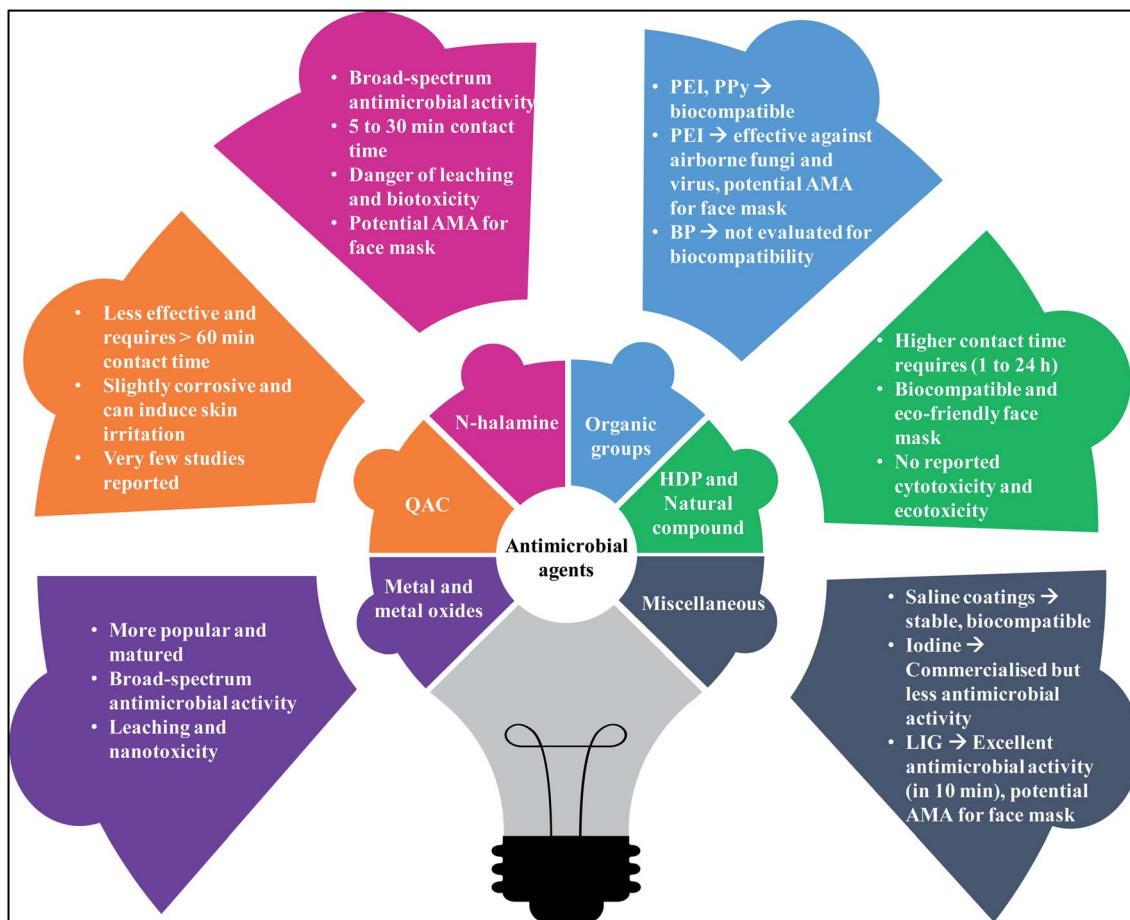


Fig. 7 Overview of various antimicrobial agents employed in antimicrobial face masks.

reduced graphene oxide²⁰⁵ are employed to make antimicrobial fabrics effective against bacteria and viruses.^{201,206,207}

Laser-Induced Graphene (LIG) was discovered in 2014. The method uses an infrared laser scribe to convert carbonaceous materials into porous 3D graphene under an ambient environment. It is a surface graphitising technique used for patterning graphene in various shapes on desirable substrates.^{208,209} Recently, LIG technology is being explored to develop self-decontaminating face masks. Zhong *et al.* (2020) employed LIG technology to deposit graphene over a commercial surgical face mask.²¹⁰ Contact angle measurements revealed graphene-coated mask provides a superhydrophobic surface (contact angle > 140°), which prevents the attachment of virus-laden droplets onto the face mask surface. From photothermal studies, it was revealed that on exposure to sunlight, the temperature of the graphene-coated face mask spikes to 70 °C in 40 s, with potential to eliminate any virus deposited on the face mask surface (any virus is expected to be inactivated at 70 °C). However, a thorough evaluation of the antimicrobial property and bioaerosol filtration efficiency need to be carried out to implement it in the real field applications. In a similar study, Huang and co-workers (2020) observed that LIG deposited commercial surgical face mask exhibited 81.57% reduction against *E. coli* load from an initial from an initial load of 10⁷ to

10⁸ CFU mL⁻¹. The face mask demonstrated 100% inhibition against *E. coli* when irradiated at 0.75 kW m⁻² sunlight for 10 min.²¹¹ The present environmental problems faced due to single-use face masks can be overcome by developing similar self-cleaning or self-sterilising mask, which assures the reusability of face masks. An overview of Section 6.1 to 6.4 regarding various AMA is given in Fig. 7.

6.5. Methods of incorporation of antimicrobial agents

AMA can be incorporated into a face mask through various methods such as dip coating, spray coating, melt-blowing, and electrospinning. The diagrammatic representations of these techniques are illustrated in Fig. 8.

Dip coating is a method of depositing a thin homogenous layer of a solution containing particles of ceramic, metallic, fibrous materials, biomolecules, and polymer films to form a coating on the substrate. The process is done in 5 stages: immersion, start-up, and deposition, followed by drainage and evaporation. The thickness of the coating depends on the solution properties such as surface tension, viscosity, density, and processing parameters such as withdrawal speed, evaporation rate, and duration of immersion, start-up, and deposition.²¹⁴ This technique is restricted to flat or smooth surfaces, and it is not suitable for selective coating a single side of the



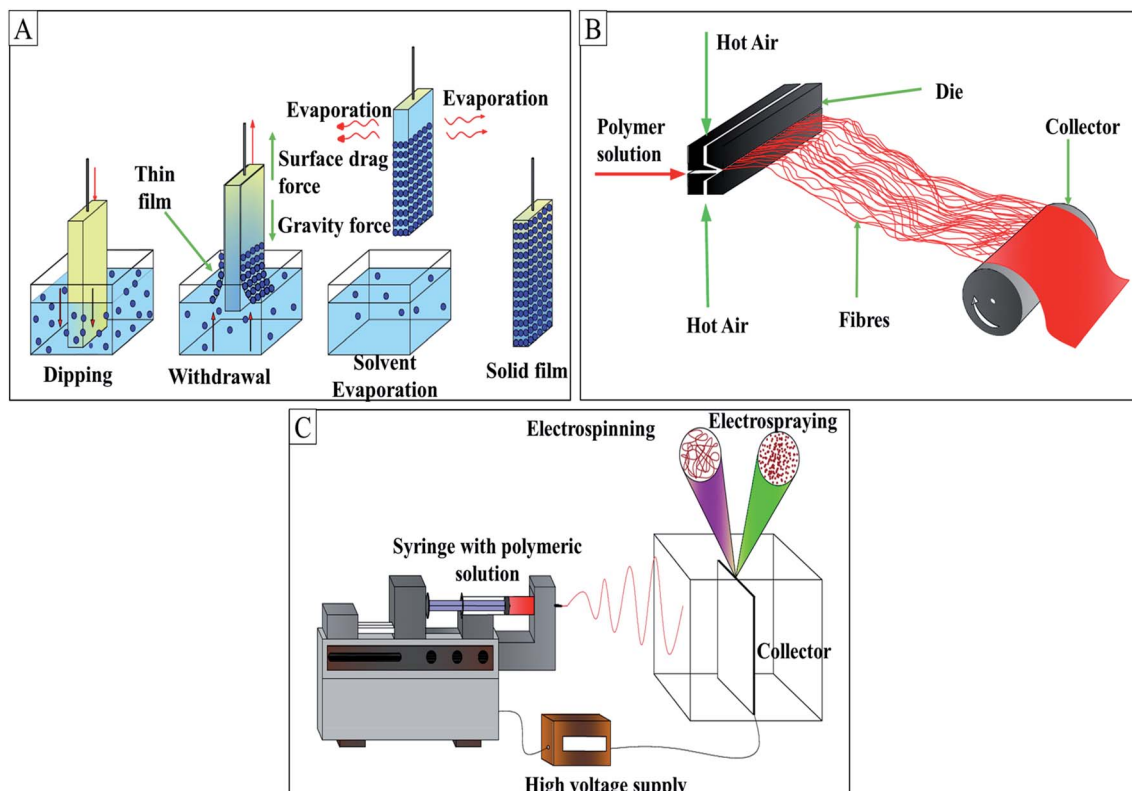


Fig. 8 (A) Dip coating; (B) melt-blowing; (C) electrospinning; this figure is adapted from the ref. 212 and 213 with modifications.

object. The instability of the precursor in case of fast condensation, large requirement of the coating solution, and difficulty in handling are the critical issues. Fig. 9(A)–(C) depicts SEM image of cotton, polyester and nonwoven fabric soaked with a microcapsule based on extractant of *Scutellaria baicalensis*. However, it is observed that the dip (soak) method has resulted in aggregation of microcapsules resulting in non-uniform distribution. Fig. 9C also shows that nonwoven fabric has randomly fashioned fibres with disordered and unsystematic features. Spin coating can address these issue and is a preferred method over dip coating for the said purpose.²¹⁵ Spin coating is practised to incorporate compounds such as QACs, antimicrobial peptides, N-halamine, metal NPs, and phosphonium salts onto fabrics.²¹⁶

Spray coating employs liquid spray to coat the material surfaces. Cold spray and thermal spray are the two types of spray coating. Cold spray is a suitable method for ceramics, metals, nanostructured materials, polymers, and non-metallic substrates. The solution is accelerated over a velocity range of 300–1200 m s⁻¹ via De-Laval nozzle.²¹⁷ Since the particles are passed at high velocity, the plastic deformation on the surface disrupts the thin film. Metals like Cu cannot be used in this technique since the high impact energy disrupts the surface. Spray technique is used to incorporate antimicrobial agents like QACs, fluorochemicals, natural compounds onto the surface of face masks.^{100,116,119,218} Fig. 10 shows the SEM image of a commercial three-layer mask spray-coated with QAC-based compound GS5. The white and opaque coating over the fibres



Fig. 9 SEM images of fabrics incorporated with Chinese herb microcapsule: (A) cotton; (B) polyester; (C) nonwoven; this figure is reproduced from the ref. 120 (Gigvvy science open access publishing platform, subject to CC BY-SA 4.0 creative common attribution license).



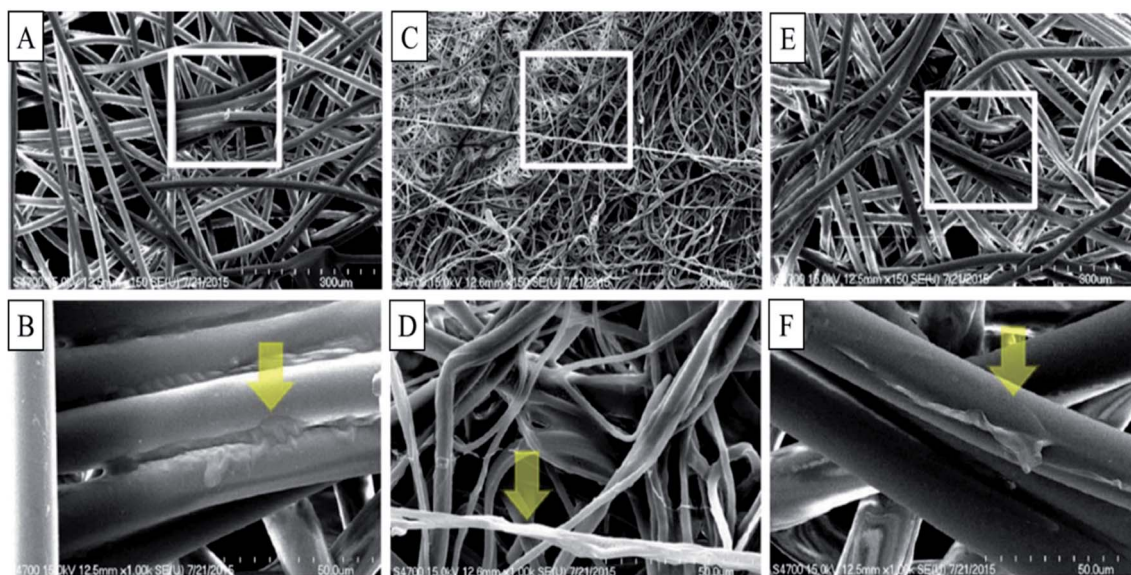


Fig. 10 SEM images: (A and B) first-layer; (C and D) second-layer; (E and F) third-layer of a commercial three-layered surgical mask coated with 1% GS5. (A), (C) and (E) have a magnification of 150 \times . White boxes in (A), (C), and (E) is magnified by 1000 \times , as shown in (B), (D), and (F). This figure is reproduced from ref. 100 with permission from Taylor & Francis, copyright 2016.

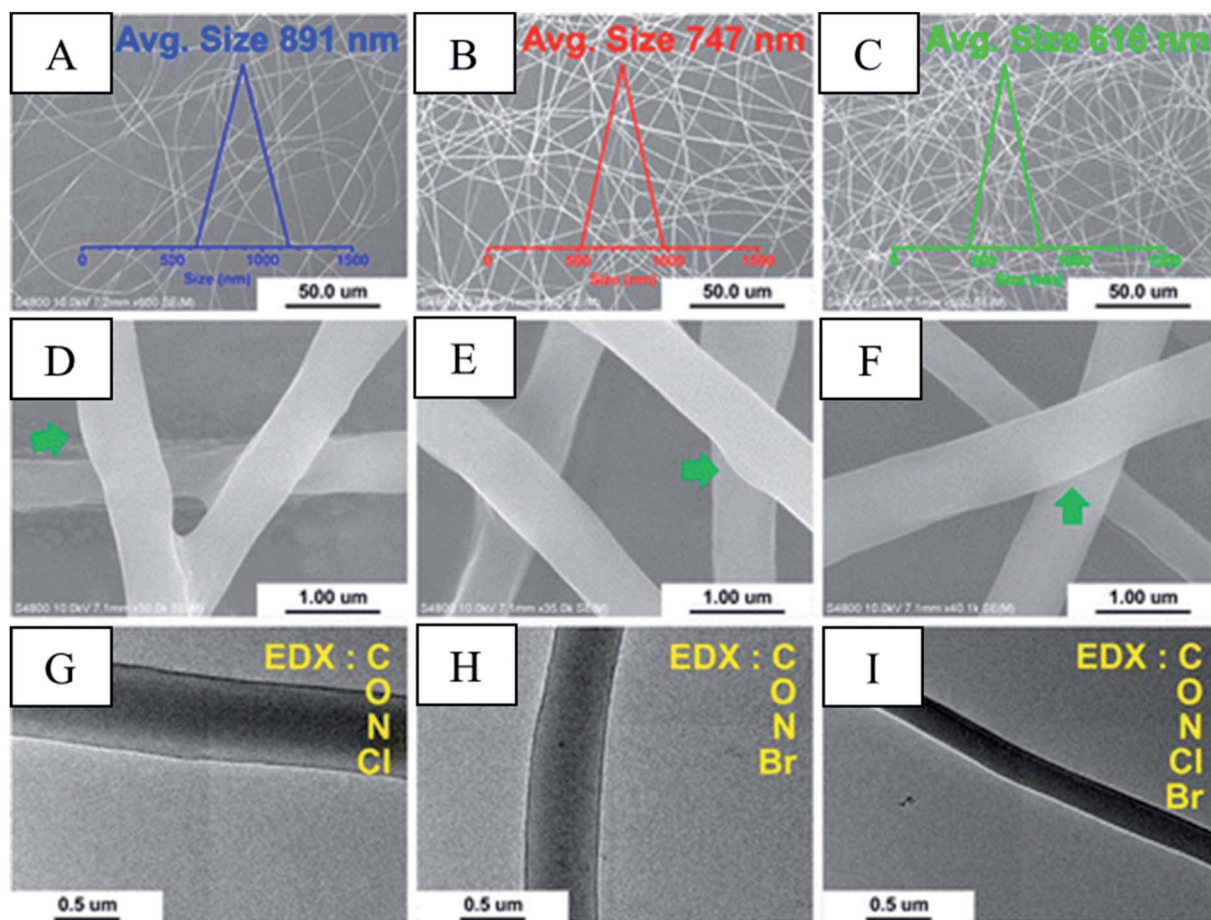


Fig. 11 SEM image: (A and D) PMMA–DCDMH; (B and E) PMMA–DBDMH; (C and F) PMMA–DCDMH/DBDMH; TEM image: (G) PMMA–DCDMH; (H) PMMA–DBDMH; (I) PMMA–DCDMH/DBDMH nanofibres developed through electrospinning. This figure is reproduced from the ref. 229 with permission from American Chemical Society, copyright 2016.



Table 2 Commercially available antimicrobial face masks

| Sl no. | Name of the product | Developed by | Antimicrobial agent/ technology | Performance efficacy | Remarks |
|--------|--------------------------------------------------------------|----------------------------------------------------------------------|-------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| 1 | <i>SonoFace</i> ^{230,231} | Sonovia (Israel) | Nano metal-oxide such as ZnO and CuO | <ul style="list-style-type: none"> • 1 log₁₀ reduction of SARS-CoV-2 • 6 log₁₀ reduction of viruses similar to SARS-CoV-2 | <ul style="list-style-type: none"> • >98% particle filtration (5 µm) • Reusable and potent up to 100 wash cycles at 70 °C • Less toxic to human and the environment • Pack of three costs 159 USD • Pose no hazard upon disposal • Conformance Européenne (CE) and FDA certified • Costs less than a conventional N95 face mask • This sticker can be attached to regular surgical face masks for the usage |
| 2 | <i>MedCu's CuO face mask</i> ²³⁰ | MedCu Technologies (Israel) | CuO | 4 log ₁₀ reduction of viruses within 30 min | <ul style="list-style-type: none"> • Pose no hazard upon disposal • Conformance Européenne (CE) and FDA certified • Costs less than a conventional N95 face mask • This sticker can be attached to regular surgical face masks for the usage |
| 3 | <i>Maya sticker</i> ²³⁰ | National Emergency Team of Israel and Israel Institute of Technology | Nanofibres coated with antiseptics | Blocks and neutralise viral particles | <ul style="list-style-type: none"> • This sticker can be attached to regular surgical face masks for the usage |
| 4 | <i>ViriFace</i> ²³⁰ | Israel | Data not available | Viruses including SARS-CoV-2 | <ul style="list-style-type: none"> • Filters must be replaced every 12 h • Wearable part can be washed and reused • Two filters and disposable envelopes cost 69 USD |
| 5 | <i>GS-3D N95</i> (ref. 232) | Gold Shield (China) | Organosilane based antimicrobial compound | 3 log ₁₀ reduction against bacteria and viruses like H1N1, methicillin-resistant <i>S. aureus</i> (MRSA), and SARS-CoV | <ul style="list-style-type: none"> • Five-ply 3D N95 face mask • Effective against particulates, pollutants, and pollens • US FDA approved • Reusable up to 14 days • Reusable cotton face mask |
| 7 | <i>Silver Shield Ag⁺</i> TM (ref. 233) | Good Research Solutions LLC (US) | Silver ions | 2 log ₁₀ reduction against pathogens | <ul style="list-style-type: none"> • 5-Layer activated carbon filter present • Potent up to 50 wash cycles |
| 8 | <i>SpectraShield 9500</i> (ref. 234) | Nexera medical Inc. (Canada) | <i>Sciessent's Ag-Cu</i> based technology | <ul style="list-style-type: none"> • 4 log₁₀ reduction of bacteria in 1 h • 5 log₁₀ reduction of influenza viruses in 5 min | <ul style="list-style-type: none"> • Ag-Cu impounded to polyester fibres • Used up to 8 h continuously • NIOSH approved |
| 9 | <i>NanoHack</i> ²³⁵ | Copper Company (Chile) | Nano Cu | Antiviral activity against influenza viruses | <ul style="list-style-type: none"> • Three-layer nonwoven PP filter to embed nano-Cu • Recyclable face mask made up of biocompatible polymer • Blocks 96.4% (1 µm) and 89.5% (0.02 µm) microorganisms • Four-layer washable and reusable face mask made up of cotton fabrics • Durable up to 50 wash cycles • Costs 29.56 USD per piece • Complies with EN149:2001+A1:2009 standard • BFE 99.8% • VFE 99.7% |
| 10 | <i>Copper KN99</i> (ref. 236) | Copper Compression (US) | Cu | Filters 99% of airborne particles equivalent to KN99 | <ul style="list-style-type: none"> • Recyclable face mask made up of biocompatible polymer • Blocks 96.4% (1 µm) and 89.5% (0.02 µm) microorganisms • Four-layer washable and reusable face mask made up of cotton fabrics • Durable up to 50 wash cycles • Costs 29.56 USD per piece • Complies with EN149:2001+A1:2009 standard • BFE 99.8% • VFE 99.7% |
| 11 | <i>NovekoTM RD2 respirator</i> ²³⁷ | Noveko International Inc (Canada) | Data not available | H1N1, H5N1, <i>S. aureus</i> , <i>K. pneumoniae</i> , MRSA, and endospores of <i>Bacillus</i> , <i>Enterococcus</i> , <i>Acinetobacter</i> | <ul style="list-style-type: none"> • Fluid resistance provides protection from blood, liquid splashes, and wet environmental contaminants • Activated carbon layer combats organic vapours and odours |
| 12 | <i>Safe LifeTM</i> (ref. 238) | Fisher scientific (US) | Triosyn iodine-polymeric resin compound | 99.99% effective against airborne and droplet borne viruses | <ul style="list-style-type: none"> • Fluid resistance provides protection from blood, liquid splashes, and wet environmental contaminants • Activated carbon layer combats organic vapours and odours |





Table 2 (Contd.)

| Sl no. | Name of the product | Developed by | Antimicrobial agent/ technology | Performance efficacy | Remarks |
|--------|--------------------------------------------|---------------------------------------------------------------------|-----------------------------------------|---------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| 13 | A400 series N95 (ref. 239) | Safe Life Corp (US) | Iodine | Data not available | <ul style="list-style-type: none"> • Compact, flat-fold design • Was widely distributed by healthcare during Swine flu-2009 |
| 14 | Nsafe ⁺ ²⁴⁰ | Nanosafe solutions and Indian Institute of Technology Delhi (India) | Data not available | Data not available | <ul style="list-style-type: none"> • Triple-layer filtration technology with antimicrobial layer • BFE 99.2% • Complies with ASTM standard • Reuse potential up to 50 times • Patented and EPA authorised antimicrobial agent |
| 15 | MyClyns® (ref. 241) | ViralClyns (US) | Data not available | 95% effective against bacteria, viruses, spores, and protozoa | <ul style="list-style-type: none"> • Electric charge repels particles deposited • 99% effective against 0.3 µm particles |
| 16 | Guardian G-Volt ⁷⁷ | LIGC Applications (US) | Graphene and electrical charge | Data not available | <ul style="list-style-type: none"> • Reusable cotton face mask potent up to 10 cycles • Repels the PM 2.5 dust |
| 17 | Graphene-enhanced face mask ²⁴² | PlanarTECH & IDEATI's 2AM (UK and Thailand) | Graphene and other carbon nanomaterials | 99.95% effective against <i>S. aureus</i> within 24 h | <ul style="list-style-type: none"> • Costs 24.95 USD for a pack of three |
| 18 | Leaf ^{243,244} | Redcliffe medical Devices Inc. (US) | Ultraviolet C (UVC) | 99.99% reduction of influenza A virus | <ul style="list-style-type: none"> • FDA cleared UVC N99 face mask • HEPA and active carbon filters • 99.9997% filtration of 0.3 µm size |

represents the GS5 coating. However, it is reported that spray coating remains mostly on outer surfaces without much penetration into crevices or shadowed region where microorganisms can get stuck.¹⁰⁰

In the melt-blowing technique, nanofibres (unbonded and self-bonded webs) are formed by blowing molten thermoplastic resin at high velocity from an extruder die tip to the collector.²¹⁹ The diameter of the nanofibre (<0.1 µm, 0.5 to 10 µm, ≥10 µm) can be optimised based on the airflow velocity.²²⁰ The factors such as polymeric types, airflow velocity, die to collector distance, angle of approach, polymer throughput rate, and temperature play a role in forming fibres. Usually, PP, polybutylene terephthalate, PET, and polylactic acid-based nonwoven materials can be formed using this process.²²¹ Melt-blowing helps in developing breathable fibres, which can act as efficient filters. Double quaternary ammonium salts,¹⁶⁴ alkylammonium microbicides on perlite,⁹⁸ Ag,²²² natural compounds,¹¹⁹ and CuO⁶⁰ have been incorporated into fabrics using this technique.

Electrospinning is a facile methodology that aids in the preparation of nonwoven nanofibres. The method is applicable for both natural and synthetic polymers, metals, ceramics, and polymer alloys. The polymers like PS, PVA, PAN, and polyvinyl pyrrolidone can be electrospun to yield nanofibres to prepare disposable face masks.²²³ In the electrospinning process, the polymeric solution in the syringe is passed through single or multiple needles at a constant rate with the help of a syringe pump.²²⁴ The droplets get charged under a high electric field, and the electrostatic repulsion overcomes the surface tension.²²⁵ The charged jet gets ejected and is directed towards the collector as thin nanofibres. The collector can be a flat plate, parallel plates/strips, or a rotating disc or drum. The diameter of the nanofibres is controlled by solution and process parameters.²²⁶ Most organic polymers, including natural polymers like chitosan, chitin, silk fibroin, dextran, collagen, alginate, gelatin, and synthetic polymers such as PS, PVC, PAN, PET, polyurethane, polyamide, are suitable for electrospinning.²²⁷ Biocompatible and biodegradable polymers such as polylactic acid, poly lactic-co-glycolic acid, conductive polymers such as polyaniline, polypyrrole, and functional polymers as polyvinylidene fluoride can also be electrospun.²²³ Electrospinning technique can be employed to develop air filtration membranes and protective clothing materials with antimicrobial substances like metal NP, antibiotics, biopolymers, carbon nanomaterials, and natural compounds.^{227,228}

It has been observed that when compared to dip (soak) and spray coating, incorporation of AMA during electrospinning yield even distribution without aggregation. Fig. 11(A)–(C) shows randomly oriented, straight and continuous features of electrospun poly(methyl methacrylate) (PMMA) with 1,3-dichloro-5,5-dimethylhydantoin (DCDMH), 1,3-dibromo-5,5-dimethylhydantoin (DBDMH) and DCDMH/DBDMH. Here, DCDMH and DBDMH are N-halamine based antimicrobial compounds. From Fig. 11(D)–(F), it is clear that electrospinning PMMA along with DCDMH and DBDMH shows coarse features of the nanofibres. The transmission electron microscope (TEM) images of PMMA with DCDMH, DBDMH and DCDMH/DBDMH

as shown in Fig. 11(G)–(I) reveals the uniform distribution of DCDMH and DBDMH without any noticeable aggregation, obtained through electrospinning.

6.6. Antimicrobial face masks in the commercial market

Recent face masks launched in the market reveals that research and development are progressing towards creating novel antimicrobial face masks. Worldwide many start-ups are actively involved in the production of antimicrobial face masks/respirators. Table 2 provides the details of various commercially available antimicrobial face masks.

Worldwide companies are competing to develop novel antimicrobial face masks claiming to be safer, effective, and environmentally friendly than existing ones. The same can be comprehended from the current market potential of face masks discussed in Section 6 of the review.

7. Market potential of face masks

The face mask market depends on various factors such as decreasing air quality linked to air pollution, customer base, the number of manufacturing units for the production of face masks,²⁴⁵ the prevalence of respiratory diseases along with consumer health awareness, investment in the healthcare sector, and new launches in the market and its availability. The market is categorised based on usage (reusable, disposable), end-use (public, healthcare workers), and region (North America, Latin America, Europe, Asia Pacific, Middle East & Africa). A market research-based company, Zion, has analysed the global scenario on the PPE market, especially on face masks.²⁴⁶ It reports that the worldwide market for the N95 face mask reached around 822.6 million USD in 2019, and it can generate an income of about 1890.3 million USD by the end of the year 2026, representing an impressive compound annual growth rate (CAGR) of over 12.8% during the forecast period. COVID-19 and 2009 H1N1 outbreaks have surged the demand for face masks and respirators among the frontline workers and the general public. The Asia-Pacific region has the highest disposable market share of 33.9% in the year 2019, with a rising trend due to increasing demand for N95 face masks in highly populated countries like China and India.²⁴⁷

China is the producer of half the world's face mask, even before COVID-19. Other countries are highly dependent on them to procure nonwoven PP fabrics.²⁴⁸ PP is of high demand as the nonwoven PP face masks are free from latex, PVC, and di (2-ethylhexyl) phthalate material. PP resists allergic reactions for consumers with sensitive skin.²⁴⁷ It is reported that during January to May 2020 in China, more than 70 000 companies have registered for face mask production or trade, bringing a 13.56 fold increase compared to 2019.²⁴⁹ More than 7000 new companies have registered to make or trade the melt-blown fabric incurring 23.77 times surge compared to 2019. The data support that COVID-19 has resulted in a rise in the investment, research, and development in the N95 face mask market. However, the high cost and disparity in the performance can restrict the global growth of the N95 market in the future.²⁴⁶

The demand for disposable face masks is rising due to its widespread availability even in online shopping platforms like Amazon.com, Inc., and eBay Inc., and this trend is expected to continue in the near future.²⁴⁷ Besides, retail stores like supermarkets, hypermarkets, drug stores, and department stores also contribute to the global face mask market shares.²⁵⁰ However, fake brands are playing in the market where the customers are highly price-sensitive, which affects the trade of genuine products.²⁵¹ Currently, the global key players dominating in respirators and face masks market are 3M, Hakugen, Vogface mask, Prestige Ameritech, Kimberly-Clark, Ansell, Sinotextiles, Dasheng, DACH Schutzbekleidung, Uvex group, Honeywell International Inc, Moldex-Metric, Inc., Kowa Company, Ltd., the Gerson Company, and SAS Safety Corp.²⁵²

The unprecedented situations brought by COVID-19 have increased the markets for antimicrobial face masks as well. It is reported that consumer awareness and health concerns due to escalating fungi and bacteria prone diseases drives the demand for breathable antimicrobial coatings. Incorporation of antimicrobials on face masks can enhance the reusability of face masks. It is reported that the reusable N95 respirators recorded the highest market share in 2019 due to its superior advantages over the disposable N95 respirators.²⁴⁶ In contrast, the UK and US promote single-use or disposable face masks, as they abolish the need for sterilisation after use and avoids cross-contamination with other reusable products.²⁵² North America and Europe take up a significant share in the breathable antimicrobial coating, including antimicrobial face masks. The market potential in the Asia-Pacific region is expected to improve as they will exhibit a considerable consumption rise in the forecast period from 2019 to 2026.²⁵³

Major retailers like Amazon and Etsy have taken up the large customer base of antimicrobial face masks. The escalating trend also has boosted the activity of biotechnology companies. Studies demonstrating the quick deactivation of COVID-19 on Cu surfaces are definite to enhance the market potential of antimicrobial face masks.²⁵⁴ The superiority of antimicrobial face masks, with self-sanitising properties that can prevent unpleasant odours and cross-contamination, increases the likelihood of usage of antimicrobial face masks, as reported by the Chief Executive Officer of Noble Biomaterials, a company specialising in antimicrobials for soft surfaces.

8. Possible environmental impacts associated with the usage of face masks

In response to the control of infectious diseases, single-use PPEs, including face masks and respirators with secondary infection potential, opens up a serious question of their safe and effective disposal. Improper handling and disposal of face masks and respirators can lead to various environmental challenges soon. Medical face masks, which are rated for single-use, pose a challenging problem of managing a large volume of hazardous waste. Prata and team estimated that approximately 129 billion masks and 65 billion plastic gloves are used every



month worldwide in the COVID-19 scenario.²⁵⁵ The increasing demand for face masks made of synthetic polymers such as PE, PP, polyurethane, PAN, polycarbonate, PS, and polyester adds to the plastic waste, which is harder to recycle^{256,257} and causing a significant burden on the environment.⁸² It is reported that improper disposal of even 1% of face masks can add 10 million face masks to the solid-waste stream and pose a significant threat to the environment.²⁵⁷

New York Times reports that 80% of Americans wear face mask regularly when going out or in close contact with other people.²⁵⁸ Data reveal that 60 to 90% of the population wears face masks in public spaces in Spain, Italy, Germany, and France since July 2020.²⁵⁹ A report estimates that if every individual in the UK wears a single-use mask per day, it will mound to 66 000 tonnes of contaminated waste plus 57 000 tonnes of plastic packaging waste annually.²⁶⁰ Even in a normal situation, an adult in Japan uses 43 numbers of face masks per year on average.²⁶¹ In highly populated countries like China and India, the volume of use is expected to be significantly high. According to Environment Pollution (Prevention and Control) Authority, New Delhi is presently producing more than 300 tonnes per day of biomedical waste, which is substantially higher than the installed combined capacity of 74 tonnes per day of the two Biomedical Waste Treatment Centres (BMWTCs) in the city.^{262,263} A recent report shows that COVID-19 pandemic has doubled the generation of biomedical waste from approximately 12 to 24 tonnes per day in Mumbai.^{262,264} On average, India produced 600 tonnes per day of biomedical waste, and the installed capacity of 198 BMWTCs is even inadequate to handle the waste generated during pre-COVID-19 times.²⁶² Hospitals in Wuhan were producing 240 tonnes of biomedical waste per day at the peak of pandemic in February 2020, *i.e.* six times the no-COVID situation,^{265–267} necessitating the establishment of mobile waste treatment facilities for the mass influx of waste.²⁶⁸ Another study conducted in a few Asian cities during COVID-19 reported a 5-time daily increase in biomedical waste compared to pre-COVID-19 time.²⁶⁹

Even though there are stringent regulations and disposal mechanisms for used PPEs in clinical settings,⁸² most countries worldwide are not equipped to handle the unprecedented volume of such wastes generated in a short period. Face masks littering creates a new form of environmental challenge for both terrestrial and aquatic ecosystems.¹⁷ As per a United Nations report, approximately 75% of the discarded face masks find their way to dump yards or water bodies.²⁷⁰ In the COVID-19 scenario, there have been alarming reports of washing up of face masks on beaches resulting in maritime littering,²⁵⁷ which reduces the aesthetic and recreational values of coastal areas impacting the tourism industry.^{271,272} The dispersed face masks in the aquatic system are a source of riverine and marine pollution with microplastics and microfibres.²⁷³ The microplastics in face masks often contain phthalate, polybrominated biphenyl ether, nonylphenol, organotin, and triclosan, which are toxic and can cause high risks when released during their disintegration.²⁷¹ The microplastics leached from the face masks get ingested in marine animals like fishes and sharks.^{271,274} Through the fishes and other aquatic animals, the

microplastics enter the food chain and pose health risks to living beings at higher trophic levels.¹⁷

The ongoing COVID-19 pandemics has also generated an unusually large quantum of household medical wastes, and their disposal is a mammoth task, especially for developing and populated countries like India. Data reveals that 3 billion people lack access to safe disposal mechanisms worldwide and relies on open, uncontrolled dumping and burning of used face masks and gloves. When mixed with municipal solid wastes, these wastes lead to a large mass of hazardous waste if not correctly segregated.²⁷³ Improper waste collection and management systems, together with ignorance and irresponsible behaviour of ordinary people, can further burden the environment. The only means of disposal of face mask for them remains dumping or burning. Piling up of used face masks on roadside and dump yards can inflict health threat to individuals, sanitation workers, animals, birds, and rodents coming in contact with contaminated face masks. In contrast, open burning or uncontrolled incineration practices of the petrochemical-derived face masks can release harmful gases, including dioxins and furans, into the environment.^{257,270}

The conventional waste management systems like landfilling and incineration have limitations. The poorly maintained incinerators and overloaded landfills are not equipped to handle hazardous nature of face masks, causing emissions and leaching of toxins.²⁵⁷ On the other hand, the land disposal of face masks can result in the release of biohazards, microplastics, and AMA and contaminate the soil and water bodies.²⁷⁵ The carbon footprint exerted from the polymer resin production, conversion, and disposal stage is critically high.²⁷⁶ It is reported that 2.7 tonnes of greenhouse gas could be potentially released into the environment per tonne of face mask from the polymer resin production stage alone.²⁷⁶ Studies reveal that one tonne of PP, PE, PS, and PVC produces 1.5, 2.4, 3.1, and 2.2 tonnes of CO₂ during the production stage and 1.7, 1.6, 2.1, and 0.9 tonnes of CO₂ during incineration with energy recovery, respectively.^{277,278}

Recently, efforts are made to improve the life and reusability of the face mask by incorporating AMA. Reusable antimicrobial face masks have been found useful for multiple cycles and can reduce the waste generation significantly. Though such face masks are promising in reducing the volume of waste generated and associated environmental impacts, experts are somehow worried about the fate of AMA at the service life of face masks. Since most of the AMA incorporated onto face masks are in nano-forms, the unpredictable interaction of these materials with the surrounding environment raises the issue of nanotoxicity.²⁷⁹ Waste treatment options such as landfilling, incineration, or even recycling do not nullify the harmful effects posed by the NPs.²⁸⁰ A portion of the NPs gets released through emissions and leaching and impacts the microenvironment. Moreover, there are chances that metal-based antimicrobial materials detach from the polymer matrix leaching into water bodies and soil matrix.¹³⁴ For instance, during washing and reuse of face mask, the NPs leaching from the surface reduces the antimicrobial efficacy



of face masks and finds their way into aqueous media.^{16,281} The fate of the AMA in water, soil, and air media has to be looked into to get a full picture of the environmental compatibility of antimicrobial face masks.

Apart from the environmental concerns, health impacts from the use of such novel AMA have to be studied. The health impacts arising out of face mask are of a high priority since the antimicrobial compounds will be in very close contact with the skin. Few studies express concern over the use of NPs in consumer products due to exposure to occupational and environmental hazards.^{282,283} The principal mechanism by which metal NPs induce toxicity is through the formation of ROS, followed by oxidative stress in cells and tissue.^{279,284} ROS in contact with skin is known to develop dermatological disorders.²⁸⁵ So it is highly recommended to conduct tests to evaluate skin irritation and user comfort for any combination of different classes of antimicrobials. While implementing novel technology, the evaluation of biocompatibility and cytotoxicity should be of prime concern. The data warrant an integrated approach for effective PPE waste management to reduce the microbial transmission with the least health and ecological implications.

9. Conclusion

The large-scale outbreaks of infectious diseases and associated public health crisis are not new to human history. Flu viruses, non-flu viruses, and human coronavirus are the significant causes of pandemics starting from Russian flu-1889 to COVID-19. These great pandemics of the past and the present have posed significant public health issues and negatively impacted society, economy, global security, and the environment to a great extent. With the onset of the COVID-19 pandemic, economies worldwide have weakened significantly due to prolonged lockdown. The pandemic has killed several thousands of people worldwide directly and indirectly. The pandemic has also taken away the opportunities and livelihoods and pushed several families to hunger and poverty, and caused irreversible damage to society.

Countries across the world have taken several control measures to prevent the spread of COVID-19 and save the lives of their citizens. However, the advisories given to use face masks in public seems to be the most effective and practical short-term measures taken to prevent the spread of the disease. Face masks are proven effective in preventing the airborne, droplet, and aerosol transmission of pathogenic microorganisms. Its role in preventing the spread of infectious diseases are well recognised and time tested. The same is apparent in the ongoing COVID-19 pandemic as well. Given the role of face masks in pandemic preparedness, face masks have evolved in material and design to meet the increasing safety and comfort requirements from the cotton coverings of the middle ages to the recent hi-tech smart face masks. However, most of the face mask available in the commercial market are designed and developed for single-use. The widespread use of disposable face mask can pose a significant challenge to the environment due to the high generation of hazardous waste.

The risk of secondary transmission, cross-contamination, and the difficulty in managing a large quantum of plastic waste laden with biohazards has heightened the development of reusable face mask with inherent antimicrobial properties. The reusable antimicrobial face masks can also reduce the surge in demand and the stress in the supply chain. It is established that antimicrobial face masks exhibit enhanced performance over conventional face masks by providing *in situ* real-time antimicrobial control. However, choosing the right AMA is crucial for optimal performance. Now there are different classes of AMA that can be appropriately used in various combinations to yield a novel and effective antimicrobial face mask. Among various AMA studied, Ag and Cu infused antimicrobial face masks are more popular owing to their high reactivity and excellent antimicrobial efficacy in nano-forms. However, the detachment of the nanoparticles from the face mask and associated nanotoxicity are the concerns. Metal and metal oxide nanoparticles incorporated through electro-spinning and melt-blowing techniques have shown higher stability and less leaching.

New classes of AMA such as antimicrobial polymers (with active moieties like antimicrobial peptides, QACs, N-halamine compounds), iodine, NaCl, and natural compounds such as mangosteen extracts, extracts of *Scutellaria baicalensis* extracts, *Folium Plectranthii amboinicii* oil, *Vitex trifolia*, *Punica granatum*, *Allium sativum* are identified as effective against various microorganisms including viruses. Polymers, which acts as the primary substrate for face masks, can be fine-tuned to impart bio-active and bio-passive properties. The active moieties like N-halamines, QACs, PEI, BP, polypyrrole, and inorganic groups (mostly metals) have been incorporated to yield various antimicrobial polymers suitable for making a reusable face mask. Among these, N-halamine and QACs are proven powerful against a broad spectrum of microorganisms. Bath coating, spray coating, and immobilisation *via* carriers have been employed to yield QACs modified antimicrobial fabrics. Direct polymerisation of monomers and covalent attachment of QACs may enhance the stability of the coating and the performance. All the studies pertaining to N-halamines reports high effectiveness in short contact times in antimicrobial fabrics. However, the real field applicability of N-halamine on face masks has not been explored yet. Natural compounds and antimicrobial peptides are promising AMA due to less ecotoxicity and proven antimicrobial properties. Besides these compounds, saline coatings and iodine (employed in NIOSH approved commercial face masks) have been investigated for their antimicrobial properties after incorporation on face masks. Lately, graphene-coated face masks are gaining much prominence owing to their self-decontaminating property.

Research on antimicrobial face masks started gaining momentum after the SARS-2003 epidemic. Further with the onset of COVID-19, the spike in demand for antimicrobial face masks has escalated its market share. The CAGR growth of 12.8% of the face mask market between 2020 and 2026 project the potential opportunities in investment, innovation, research, and development in the area. However, the



mushrooming sales of antimicrobial masks, without adequate quality control, in online and offline shopping platforms is a cause of concern.

Though antimicrobial face masks seem superior over conventional face masks on multiple facets, a detailed study on various aspects associated with their usage and disposal should be performed to understand their short and long-term benefits and drawbacks. Besides, the methods reported for testing antimicrobial activities are different for different microorganisms studied, assays used, and experimental setup. The lack of a standardised methodology makes it difficult to compare their performance. In this context, extensive research is also needed to develop a robust protocol and validating standards about the use of AMA, antimicrobial loading, and testing the antibacterial and antiviral activity of antimicrobial face mask. The performance of antimicrobial face masks should be evaluated for both antibacterial and antiviral activities, to establish the claim of “antimicrobial face mask” on more substantial grounds for developing a protective face mask. There should be a thorough evaluation of the biotoxicity and ecotoxicity associated with the AMA and the antimicrobial face masks. The risk of unknown toxicity calls for proper assessment of, skin compatibility, and stability of the antimicrobial coatings. Therefore, proper usage, reuse, and disposal protocol should be formulated to avail the full benefits. In short, there are ample opportunities for various stakeholders dealing with the antimicrobial mask to develop affordable, safe, and efficient antimicrobial face mask that can overcome the challenges present with the single-use face masks.

Abbreviations

| | |
|---------------------|----------------------------------------------------|
| <i>A. baumannii</i> | <i>Acinetobacter baumannii</i> |
| <i>A. niger</i> | <i>Aspergillus niger</i> |
| AgNP | Silver nanoparticle |
| AIDS | Acquired immunodeficiency syndrome |
| AMA | Antimicrobial agents |
| ASTM | American Society for Testing and Materials |
| <i>B. subtilis</i> | <i>Bacillus subtilis</i> |
| BFE | Bacterial filtration efficiency |
| BMWTC | Biomedical waste treatment centre |
| BP | Benzophenone |
| BSI | Benzoic acid, Salicylic, and Iode |
| <i>C. albicans</i> | <i>Candida albicans</i> |
| CAGR | Compound annual growth rate |
| CDC | Centers for Disease Control and Prevention |
| CDDMH | Chlorinated 3-dodecyl-5,5-dimethyl hydantoin |
| CDMH | Chlorinated 5,5-dimethyl hydantoin |
| CE | Conformité Européenne |
| CFU | Colony-forming unit |
| COVID | Coronavirus disease |
| CTAB | Cetyltrimethylammonium bromide |
| CTMIO | Chlorinated 2,2,5,5-tetramethyl-imidazolidin-4-one |
| CuNP | Copper nanoparticle |
| DBDMH | 1,3-Dibromo-5,5-dimethylhydantoin |
| DCDMH | 1,3-Dichloro-5,5-dimethylhydantoin |

| | |
|----------------------|------------------------------------------------------------|
| DMF | Dimethylformamide |
| DNA | Deoxyribonucleic acid |
| <i>E. coli</i> | <i>Escherichia coli</i> |
| <i>E. faecalis</i> | <i>Enterococcus faecalis</i> |
| EN | European Nation |
| EPA | Environmental Protection Agency |
| FDA | Food and Drug Administration |
| FE _{NO} | Fractional concentration of nitric oxide in exhaled breath |
| FFR | Filtering facepiece respirator |
| GDP | Gross domestic product |
| GS5 | Goldshield 5 |
| GSM | Gram per metre square |
| HA | Hemagglutinin activity |
| HDP | Host defence peptide |
| <i>K. pneumoniae</i> | <i>Klebsiella pneumoniae</i> |
| LIG | Laser-induced graphene |
| MBC | Minimum bactericidal concentration |
| MC | 1-Chloro-2,2,5,5-tetramethyl-4-imidazolidinone |
| MERS | Middle East respiratory syndrome |
| MERS-CoV | Middle East respiratory syndrome coronavirus |
| MIC | Minimum inhibitory concentration |
| NIOSH | National Institute for Occupational Safety and Health |
| NP | Nanoparticle |
| OSHA | Occupational Safety and Health Administration |
| <i>P. aeruginosa</i> | <i>Pseudomonas aeruginosa</i> |
| PAN | Polyacrylonitrile |
| PE | Polyethylene |
| PEG | Polyethylene glycol |
| PEI | Polyethyleneimine |
| PET | Polyethylene terephthalate |
| PMMA | Poly(methyl methacrylate) |
| PPE | Personal protective equipment |
| PP | Polypropylene |
| PS | Polystyrene |
| PVA | Polyvinyl alcohol |
| PVC | Polyvinyl chloride |
| QAC | Quaternary ammonium compound |
| ROS | Reactive oxygen species |
| RPD | Respiratory protective device |
| <i>S. aureus</i> | <i>Staphylococcus aureus</i> |
| <i>S. epidermis</i> | <i>Staphylococcus epidermis</i> |
| SARS | Severe acute respiratory syndrome |
| SARS-CoV | Severe acute respiratory syndrome coronavirus |
| SEM | Scanning electron microscope |
| TEM | Transmission electron microscope |
| UK | United Kingdom |
| US | United States |
| USD | United States Dollar |
| UVC | Ultraviolet C |
| UV-Vis | Ultraviolet-visible |
| VFE | Viral filtration efficiency |
| w/v | Weight/volume |
| w/w | Weight/weight |
| WHO | World Health Organisation |
| ZnONP | Zinc oxide nanoparticle |



Credit authorship contribution statement

G. P., U. K., G. S., and S. M. M. designed and wrote the original draft, S. M. M. and D. V. K. reviewed and edited the paper, S. M. M. and D. V. K. secured the fund.

Conflicts of interest

There are no conflicts of interest to declare.

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References

- 1 Y. Xue, H. Xiao and Y. Zhang, Antimicrobial Polymeric Materials with Quaternary Ammonium and Phosphonium Salts, *Int. J. Mol. Sci.*, 2015, **16**, 3626–3655.
- 2 WHO, *Up to 650 000 people die of respiratory diseases linked to seasonal flu each year*, 2017, <https://www.who.int/news-room/detail/14-12-2017-up-to-650-000-people-die-of-respiratory-diseases-linked-to-seasonal-flu-each-year>.
- 3 A. D. Iuliano, K. M. Roguski, H. H. Chang, D. J. Muscatello, R. Palekar, S. Tempia, *et al.*, Estimates of global seasonal influenza-associated respiratory mortality: a modelling study, *Lancet*, 2018, **391**, 1285–1300.
- 4 M. S. Smolinski, M. A. Hamburg and J. Lederberg. *Microbial Threats to Health: Emergence, Detection, and Response*, National Academies Press, Washington, D.C., 2003, <http://www.nap.edu/catalog/10636>.
- 5 WHO, *Ebola virus disease*, 2020, <https://www.who.int/news-room/fact-sheets/detail/ebola-virus-disease>.
- 6 WHO, *HIV/AIDS*, 2020. <https://www.who.int/news-room/fact-sheets/detail/hiv-aids>.
- 7 Worldometer, *Coronavirus Update (Live): 95,016,073 Cases and 2,032,342 Deaths from COVID-19 Virus Pandemic - Worldometer*, 2021, <https://www.worldometers.info/coronavirus/>.
- 8 A. C. K. Lai, C. K. M. Poon and A. C. T. Cheung, Effectiveness of facemasks to reduce exposure hazards for airborne infections among general populations, *J. R. Soc. Interface*, 2012, **9**, 938–948.
- 9 M. B. Lore, J. M. Sebastian, T. L. Brown, A. S. Viner, N. V. McCullough and S. H. Hinrichs, Performance of conventional and antimicrobial-treated filtering facepiece respirators challenged with biological aerosols, *J. Occup. Environ. Hyg.*, 2012, **9**, 69–80.
- 10 D. A. Harnish, B. K. Heimbuch, M. Husband, A. E. Lumley, K. Kinney, R. E. Shaffer, *et al.*, Challenge of N95 filtering facepiece respirators with viable H1N1 influenza aerosols, *Infect. Control Hosp. Epidemiol.*, 2013, **34**, 494–499.
- 11 N. H. L. Leung, D. K. W. Chu, E. Y. C. Shiu, K.-H. Chan, J. J. McDevitt, B. J. P. Hau, *et al.*, Respiratory virus shedding in exhaled breath and efficacy of face masks, *Nat. Med.*, 2020, **26**, 676–680.
- 12 C. R. MacIntyre and A. A. Chughtai, A rapid systematic review of the efficacy of face masks and respirators against coronaviruses and other respiratory transmissible viruses for the community, healthcare workers and sick patients, *Int. J. Nurs. Stud.*, 2020, **108**, 103629.
- 13 S. Feng, C. Shen, N. Xia, W. Song, M. Fan and B. J. Cowling, Rational use of face masks in the COVID-19 pandemic, *Lancet Respir. Med.*, 2020, **8**, 434–436.
- 14 A. L. Pasanen, J. Keinanen, P. Kalliokoski, P. I. Martikainen and J. Ruuskanen, Microbial growth on respirator filters from improper storage, *Scand. J. Work, Environ. Health*, 1993, **19**, 421–425.
- 15 L. M. Brosseau, N. V. McCullough and D. Vesley, Bacterial survival on respirator filters and surgical masks, *J. Am. Biol. Saf. Assoc.*, 1997, **2**, 32–43.
- 16 C. R. Zheng, S. Li, C. Ye, X. Li, C. Zhang and X. Yu, Particulate respirators functionalized with silver nanoparticles showed excellent real-time antimicrobial effects against pathogens, *Environ. Sci. Technol.*, 2016, **50**, 7144–7151.
- 17 O. O. Fadare and E. D. Okoffo, Covid-19 face masks: A potential source of microplastic fibers in the environment, *Sci. Total Environ.*, 2020, **737**, 140279.
- 18 S. Rengasamy, E. Fisher and R. E. Shaffer, Evaluation of the survivability of MS2 viral aerosols deposited on filtering face piece respirator samples incorporating antimicrobial technologies, *Am. J. Infect. Control*, 2010, **38**, 9–17.
- 19 A. Hirsch, *Handbook of Geographical and Historical Pathology*, New Sydenham Society, 1883, p. 740.
- 20 WHO, *Plague*, 2020, <https://www.who.int/westernpacific/health-topics/plague>.
- 21 M. Rosenwald, *History's deadliest pandemics, from ancient Rome to modern America*, Washington Post, 2020, <https://www.washingtonpost.com/graphics/2020/local/retropolis/coronavirus-deadliest-pandemics/>.
- 22 Worldometer, *World Population by Year - Worldometer*, 2020, <https://www.worldometers.info/world-population/world-population-by-year/>.
- 23 Worldometer, *World Population Clock: 7.8 Billion People (2021) - Worldometer*, 2021, <https://www.worldometers.info/world-population/>.
- 24 WHO. *WHO/HIV/AIDS*, WHO, World Health Organization, 2020, <http://www.who.int/gho/hiv/en/>.
- 25 CDC, *SARS/Basic Factsheet/CDC*, 2019, <https://www.cdc.gov/sars/about/fs-sars.html>.



- 26 WHO, *WHO/Middle East respiratory syndrome coronavirus (MERS-CoV)*, WHO, World Health Organization, 2020, <http://www.who.int/emergencies/mers-cov/en/>.
- 27 S. Hamid, M. Y. Mir and G. K. Rohela, Novel coronavirus disease (COVID-19): a pandemic (epidemiology, pathogenesis and potential therapeutics), *New Microbes New Infect.*, 2020, **35**, 100679.
- 28 E. T. Ewing, La Grippe or Russian influenza: Mortality statistics during the 1890 Epidemic in Indiana, *Influenza Other Respir. Viruses*, 2019, **13**, 279–287.
- 29 N. LePan, *Visualizing the History of Pandemics*, Visual Capitalist, 2020, <https://www.visualcapitalist.com/history-of-pandemics-deadliest/>.
- 30 N. Madhav, B. Oppenheim, M. Gallivan, P. Mulembakani, E. Rubin and N. Wolfe, Pandemics: Risks, Impacts, and Mitigation, in *Disease Control Priorities: Improving Health and Reducing Poverty*, ed. D. T. Jamison, H. Gelband, S. Horton, P. Jha, R. Laxminarayan, C. N. Mock, *et al.*, The International Bank for Reconstruction and Development/The World Bank, Washington, DC, 3rd edn, 2017, <http://www.ncbi.nlm.nih.gov/books/NBK525302/>.
- 31 S.-E. Mamelund, *Social inequality – a forgotten factor in pandemic influenza preparedness*, Tidsskr Den Nor Legeforening, 2017, <https://tidsskriftet.no/2017/05/global-helse/social-inequality-forgotten-factor-pandemic-influenza-preparedness>.
- 32 Q. Fottrell, *Another warning from the 1918 flu for COVID-19: 'Survival does not mean that individuals fully recovered'*, MarketWatch, 2020, <https://www.marketwatch.com/story/another-warning-from-1918-spanish-flu-for-covid-19-survival-does-not-mean-that-individuals-fully-recovered-2020-08-18>.
- 33 L. Spinney, How pandemics shape social evolution, *Nature*, 2019, **574**, 324–326.
- 34 CPJ, *Iran bans printing of all newspapers, citing spread of coronavirus*, Committee to Protect Journalists, 2020, <https://cpj.org/2020/03/iran-bans-printing-of-all-newspapers-citing-spread/>.
- 35 I. Berman, *Coronavirus & Iran: Regime's End?* *National Review*, 2020, <https://www.nationalreview.com/2020/03/will-irans-regime-survive-coronavirus/>.
- 36 W. McKibbin and A. Sidorenko, *Global Macroeconomic Consequences of Pandemic Influenza*, Centre for Applied Macroeconomic Analysis, Crawford School of Public Policy, The Australian National University, 2006, <https://econpapers.repec.org/paper/eencamaaa/2006-26.htm>.
- 37 African Development Bank Group, *African Economic Outlook 2020 - Supplement*, African Development Bank - Building today, a better Africa tomorrow. African Development Bank Group, 2020, <https://www.afdb.org/en/documents/african-economic-outlook-2020-supplement>.
- 38 EFE, *MERS outbreak to reduce South Korean GDP by 0.3 percent in 2015*, www.efe.com, 2015, <https://www.efe.com/efe/english/business/mers-outbreak-to-reduce-south-korean-gdp-by-0-3-percent-in-2015/50000265-2640814>.
- 39 IMF, *World Economic Outlook, April 2020: The Great Lockdown*, IMF, 2020, <https://www.imf.org/en/Publications/WEO/Issues/2020/04/14/weo-april-2020>.
- 40 UNDRR, *The known and unknown economic and social consequences of pandemics*, 2020, <https://www.preventionweb.net/go/72070>.
- 41 Business Today, *Which top economies have suffered worst GDP fall due to COVID-19?*, 2020, <https://www.businesstoday.in/current/economy-politics/which-top-economies-have-suffered-worst-gdp-fall-due-to-covid-19/story/414683.html>.
- 42 WHO, *WHO/Worldwide reduction in MERS cases and deaths since 2016*, WHO, World Health Organization, 2020, <http://www.who.int/emergencies/mers-cov/worldwide-reduction-in-mers-cases-and-deaths-since-2016/en/>.
- 43 WHO, *WHO/SARS (Severe Acute Respiratory Syndrome)*, WHO, World Health Organization, 2020, <https://www.who.int/ith/diseases/sars/en/>.
- 44 Z. Zhao, W. Hai, J. Wang and Z.-G. Hou, *The Short-term Impact of SARS on the Chinese Economy*, ResearchGate, 2004, https://www.researchgate.net/publication/24089622_The_Short-term_Impact_of_SARS_on_the_Chinese_Economy.
- 45 BBC, *Chinese economy bounces back into growth*, BBC News, 2020, <https://www.bbc.com/news/business-53399999>.
- 46 K. Yao and G. Crossley, *China's economy has rebounded after a steep slump - but challenges lie ahead*, World Economic Forum, 2020, <https://www.weforum.org/agenda/2020/07/chinas-economy-rebounds-after-steep-slump-u-s-tensions-weak-consumption-raise-challenges/>.
- 47 IMF, *World Economic Outlook Update, June 2020: A Crisis Like No Other, An Uncertain Recovery*, IMF, 2020, <https://www.imf.org/en/Publications/WEO/Issues/2020/06/24/WEOUpdateJune2020>.
- 48 B. Winck, *The IMF says its forecast for the COVID-19 recession might now be too optimistic*, World Economic Forum, 2020, <https://www.weforum.org/agenda/2020/04/imf-economy-coronavirus-covid-19-recession/>.
- 49 M. M. Farley, 2009 H1N1 Influenza: A Twenty-first century pandemic with roots in the early twentieth century, *Am. J. Med. Sci.*, 2010, **340**, 202–208.
- 50 N. J. Cox and K. Subbarao, Global epidemiology of influenza: Past and present, *Annu. Rev. Med.*, 2000, **51**, 407–421.
- 51 T. P. Weber and N. I. Stilianakis, Inactivation of influenza A viruses in the environment and modes of transmission: A critical review, *J. Infect.*, 2008, **57**, 361–373.
- 52 C.-C. Lai, T.-P. Shih, W.-C. Ko, H.-J. Tang and P.-R. Hsueh, Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and coronavirus disease-2019 (COVID-19): The epidemic and the challenges, *Int. J. Antimicrob. Agents*, 2020, **55**, 105924.
- 53 L. Brosseau, *ScD, Ann RB. N95 Respirators and Surgical Masks/Blogs/CDC*, 2009, <https://blogs.cdc.gov/niosh-science-blog/2009/10/14/n95/>.
- 54 A. Tcharkhtchi, N. Abbasnezhad, M. Zarbini Seydani, N. Zirak, S. Farzaneh and M. Shirinbayan, An overview of filtration efficiency through the masks: Mechanisms of the aerosols penetration, *Bioact. Mater.*, 2021, **6**, 106–122.
- 55 P. A. Levin and E. R. Angert, Small but mighty: cell size and bacteria, *Cold Spring Harbor Perspect. Biol.*, 2015, **7**, 1–11.



- 56 A. Lošdorfer Božič, A. Šiber and R. Podgornik, Statistical analysis of sizes and shapes of virus capsids and their resulting elastic properties, *J. Biol. Phys.*, 2013, **39**, 215–228.
- 57 V. C.-C. Cheng, S.-C. Wong, V. W.-M. Chuang, S. Y.-C. So, J. H.-K. Chen, S. Sridhar, *et al.*, The role of community-wide wearing of face mask for control of coronavirus disease 2019 (COVID-19) epidemic due to SARS-CoV-2, *J. Infect.*, 2020, **81**, 107–114.
- 58 CDC, *100 Years of Respiratory Protection History/NPPTL/NIOSH/CDC*, 2019, <https://www.cdc.gov/niosh/npptl/Respiratory-Protection-history.html>.
- 59 Tuberculosis I of M (US) C on ROE to, M. J. Field, Respiratory Protection and Control of Tuberculosis in Health Care and Other Facilities, *Tuberculosis in the Workplace*. National Academies Press (US), 2001, <https://www.ncbi.nlm.nih.gov/books/NBK222460/>.
- 60 G. Borkow, S. S. Zhou, T. Page and J. Gabbay, A Novel anti-influenza copper oxide containing respiratory face mask, *PLoS One*, 2010, **6**, 1–8.
- 61 WHO, Nonpharmaceutical interventions for pandemic influenza, national and community measures, *Emerging Infect. Dis.*, 2006, **12**, 88–94.
- 62 Global Times, *The evolution of face masks - Global Times*, 2020, <https://www.globaltimes.cn/content/1179358.shtml>.
- 63 R. MacIntyre and A. A. Chughtai, Facemasks for the prevention of infection in healthcare and community settings, *BMJ*, 2015, **350**(h694), 1–12.
- 64 E. Blakemore, *Why plague doctors wore those strange beaked masks*, 2020, <https://www.nationalgeographic.com/history/reference/european-history/plague-doctors-beaked-masks-coronavirus/>.
- 65 V. Postrel, *Pandemics come and go but medical masks are eternal*, Bloomberg.com, 2020, <https://www.bloomberg.com/opinion/articles/2020-04-10/medical-face-masks-an-illustrated-history>.
- 66 M. M. Weiss, P. D. Weiss, D. E. Weiss and J. B. Weiss, Disrupting the transmission of influenza A: Face masks and ultraviolet light as control measures, *Am. J. Public Health*, 2007, **97**, S32–S37.
- 67 D. Spelce, T. R. Rehak, R. W. Metzler and J. S. Johnson, Pre-World War I firefighter respirators and the U.S. Bureau of Mines involvement in WWI, *J. Int. Soc. Respir. Prot.*, 2017, **34**, 128–135.
- 68 S. Durn, *A brief history of medical face masks*, Gizmodo, 2020, <https://gizmodo.com/a-brief-history-of-medical-face-masks-1843698852>.
- 69 B. J. Strasser and T. Schlich, A history of the medical mask and the rise of throwaway culture, *Lancet*, 2020, **396**, 19–20.
- 70 C. Matuschek, F. Moll, H. Fangerau, J. C. Fischer, K. Zänker, M. van Griensven, *et al.*, The history and value of face masks, *Eur. J. Med. Res.*, 2020, **25**, 1–6.
- 71 J. L. Spooner, History of Surgical Face Masks: The myths, the masks, and the men and women behind them, *AORN J.*, 1967, **5**, 76–80.
- 72 B. Henneberry, *How surgical masks are made, tested and used*, 2020, <https://www.thomasnet.com/articles/other/how-surgical-masks-are-made>.
- 73 N. L. Belkin, The evolution of the surgical mask: Filtering efficiency versus effectiveness, *Infect. Control Hosp. Epidemiol.*, 1997, **18**, 49–57.
- 74 B. Hogan and L. P. Samaranyake, The surgical mask unmasked: A review, *Oral Surg. Oral Med. Oral Pathol.*, 1990, **70**, 34–36.
- 75 Y. Li, P. Leung, L. Yao, Q. W. Song and E. Newton, Antimicrobial effect of surgical masks coated with nanoparticles, *J. Hosp. Infect.*, 2006, **62**, 58–63.
- 76 S. Chou, Mask having anti-virus and anti-germ effect, US20050183727A1, 2005, <https://patents.google.com/patent/US20050183727/en>.
- 77 I. Block, *Guardian G-Volt masks use graphene and electrical charge to repel viruses*, Dezeen, 2020, <https://www.dezeen.com/2020/03/06/guardian-g-volt-face-mask-graphene-coronavirus-bacteria/>.
- 78 N. van Doremalen, T. Bushmaker, D. H. Morris, M. G. Holbrook, A. Gamble and B. N. Williamson, Aerosol and surface stability of SARS-CoV-2 as compared with SARS-CoV-1, *N. Engl. J. Med.*, 2020, 1–3.
- 79 H. A. Aboubakr, T. A. Sharafeldin and S. M. Goyal, Stability of SARS-CoV-2 and other coronaviruses in the environment and on common touch surfaces and the influence of climatic conditions: A review, *Transboundary Emerging Dis.*, 2020, 1–17.
- 80 A. W. H. Chin, J. T. S. Chu, M. R. A. Perera, K. P. Y. Hui, H.-L. Yen, M. C. W. Chan, *et al.*, Stability of SARS-CoV-2 in different environmental conditions, *Lancet Microbe*, 2020, **1**, e10.
- 81 WHO, *Transmission of SARS-CoV-2: implications for infection prevention precautions*, 2020, <https://www.who.int/news-room/commentaries/detail/transmission-of-sars-cov-2-implications-for-infection-prevention-precautions>.
- 82 S. Duong-Quy, X. Ngo-Minh, T. Tang-Le-Quynh, T. Tang-Thi-Thao, B. Nguyen-Quoc, K. Le-Quang, *et al.*, The use of exhaled nitric oxide and peak expiratory flow to demonstrate improved breathability and antimicrobial properties of novel face mask made with sustainable filter paper and Folium Plectranthii amboinicii oil: additional option for mask shortage during COVID-19 pandemic, *Multidiscip. Respir. Med.*, 2020, **15**, 664.
- 83 H. Wu, J. Huang, C. J. P. Zhang, Z. He and W.-K. Ming, Facemask shortage and the novel coronavirus disease (COVID-19) outbreak: Reflections on public health measures, *EclinicalMedicine*, 2020, **21**, 100329.
- 84 G. Borkow and J. Gabbay, Putting copper into action: copper-impregnated products with potent biocidal activities, *FASEB J.*, 2004, **18**, 1728–1730.
- 85 G. Borkow and J. Gabbay, Copper as a biocidal tool, *Curr. Med. Chem.*, 2005, **12**, 2163–2175.
- 86 A. Smith and J. Taylor, Antiviral metal impregnated activated carbon cloth components, US20110114095A1, 2011, <https://patents.google.com/patent/US20110114095A1/en>.
- 87 Y. Fujimori, T. Sato, T. Hayata, T. Nagao, M. Nakayama, T. Nakayama, *et al.*, Novel antiviral characteristics of nanosized copper(I) iodide particles showing inactivation



- activity against 2009 pandemic H1N1 influenza virus, *Appl. Environ. Microbiol.*, 2012, **78**, 951–955.
- 88 A. M. Davison, Pathogen inactivation and filtration efficacy of a new anti-microbial and anti-viral surgical facemask and N95 against dentistry-associated microorganisms, *International dentistry Australasian edition*, 2012, **7**, 6.
- 89 A. K. Aloufy and M. A.-M. El-Messiry, in *Proceedings of World congress on Advances in Nano, Biomechanics, Robotics, and Energy Research*, 2013, vol. 18, pp. 811–824.
- 90 A. K. Selvam and G. Nallathambi, Polyacrylonitrile/silver nanoparticle electrospun nanocomposite matrix for bacterial filtration, *Fibers Polym.*, 2015, **16**, 1327–1335.
- 91 R. Navik, L. Thirugnanasampathan, H. Venkatesan, M. Kamruzzaman, F. Shafiq and Y. Cai, Synthesis and application of magnesium peroxide on cotton fabric for antibacterial properties, *Cellulose*, 2017, **24**, 3573–3587.
- 92 C. B. Hiragond, A. S. Kshirsagar, V. V. Dhapte, T. Khanna, P. Joshi and P. V. More, Enhanced anti-microbial response of commercial face mask using colloidal silver nanoparticles, *Vacuum*, 2018, **156**, 475–482.
- 93 C. A. P. Bastos, S. F. A. Bruggraber, N. J. R. Faria and J. J. Powell, Antibacterial compositions comprising copper oxo-hydroxide nanoparticles and their uses as biocidal agents, *US Pat.*, US20180147177A1, 2018, <https://patents.google.com/patent/US20180147177A1/en>.
- 94 K. Nakano, A. Sato, T. Kusaka, S. Kasamatsu, A. Sasaki and D. Fukushi, Antiviral material, antiviral film, antiviral fiber, and antiviral product, US10327445B2, 2019, <https://patents.google.com/patent/US10327445B2/en>.
- 95 H. Ghaffari, A. Tavakoli, A. Moradi, A. Tabarraei, F. Bokharaei-Salim, M. Zahmatkeshan, *et al.*, Inhibition of H1N1 influenza virus infection by zinc oxide nanoparticles: another emerging application of nanomedicine, *J. Biomed. Sci.*, 2019, **26**, 70.
- 96 B. Gutarowska, J. Skora, E. Nowak and I. Lysiak, Antimicrobial activity and filtration effectiveness of nonwovens with sanitized for respiratory protective equipment, *Fibres & Textiles in Eastern Europe*, 2014, **22**(3(105)), 120–125, https://www.researchgate.net/publication/262143415_Antimicrobial_Activity_and_Filtration_Effectiveness_of_Nonwovens_with_Sanitized_for_Respiratory_Protective_Equipment.
- 97 L. Chang, X. Zhang, X. Shi, L. Zhao and X. Liu, Preparation and characterization of a novel antibacterial fiber modified by quaternary phosphonium salt on the surface of polyacrylonitrile fiber, *Fibers Polym.*, 2014, **15**, 2026–2031.
- 98 B. Gutarowska, B. Brycki, K. Majchrzycka and A. Brochocka, New bioactive polymer filtering material composed of nonwoven polypropylene containing alkylammonium microbicides on a perlite carrier, *Polimery*, 2010, **55**, 568–574.
- 99 M. Montazer, F. Rangchi and F. Siavoshi, *Preparation of protective disposable hygiene fabrics for medical applications*, in *Medical and Healthcare Textiles*, Elsevier, 2010, pp. 164–170, <https://linkinghub.elsevier.com/retrieve/pii/B9781845692247500212>.
- 100 C.-C. Tseng, Z.-M. Pan and C.-H. Chang, Application of a quaternary ammonium agent on surgical face masks before use for pre-decontamination of nosocomial infection-related bioaerosols, *Aerosol Sci. Technol.*, 2016, **50**, 199–210.
- 101 L. Cen, K. G. Neoh and E. T. Kang, Antibacterial activity of cloth functionalized with N-alkylated poly(4-vinylpyridine), *J. Biomed. Mater. Res., Part A*, 2004, **71**, 70–80.
- 102 M. Ignatova, N. Manolova and I. Rashkov, Novel antibacterial fibers of quaternized chitosan and poly(vinyl pyrrolidone) prepared by electrospinning, *Eur. Polym. J.*, 2007, **43**, 1112–1122.
- 103 J. Lin, S. Qiu, K. Lewis and A. M. Klibanov, Mechanism of bactericidal and fungicidal activities of textiles covalently modified with alkylated polyethylenimine, *Biotechnol. Bioeng.*, 2003, **83**, 168–172.
- 104 K. Majchrzycka, M. Okrasa, A. Jachowicz, J. Szulc, B. Brycki and B. Gutarowska, Application of biocides and super-absorbing polymers to enhance the efficiency of filtering materials, *Molecules*, 2019, **24**, 3339.
- 105 F. Nogueira, P. Teixeira and I. C. Gouveia, Electrospinning polypropylene with an amino acid as a strategy to bind the antimicrobial peptide Cys-LC-LL-37, *J. Mater. Sci.*, 2018, **53**, 4655–4664.
- 106 D. O. Sanchez Ramirez, A. Varesano, R. A. Carletto, C. Vineis, I. Perelshtein, M. Natan, *et al.*, Antibacterial properties of polypyrrole-treated fabrics by ultrasound deposition, *Mater. Sci. Eng., C*, 2019, **102**, 164–170.
- 107 G. Tiliket, D. L. Sage, V. Moules, M. Rosa-Calatrava, B. Lina, J. M. Valleton, *et al.*, A new material for airborne virus filtration, *Chem. Eng. J.*, 2011, **173**, 341–351.
- 108 K. Tan and S. K. Obendorf, Fabrication and evaluation of electrospun nanofibrous antimicrobial nylon 6 membranes, *J. Membr. Sci.*, 2007, **305**, 287–298.
- 109 M. R. Badrossamay and G. Sun, Acyclic halamine polypropylene polymer: Effect of monomer structure on grafting efficiency, stability and biocidal activities, *React. Funct. Polym.*, 2008, **68**, 1636–1645.
- 110 B. Demir, I. Cerkez, S. D. Worley, R. M. Broughton and T.-S. Huang, N-Halamine-modified antimicrobial polypropylene nonwoven fabrics for use against airborne bacteria, *ACS Appl. Mater. Interfaces*, 2015, **7**, 1752–1757.
- 111 I. Cerkez, H. B. Kocer, S. D. Worley, R. M. Broughton and T. S. Huang, Antimicrobial functionalization of poly(ethylene terephthalate) fabrics with waterborne N-halamine epoxides, *J. Appl. Polym. Sci.*, 2016, **133**(43088), 1–6.
- 112 C. Huang, Y. Liu, Z. Li, R. Li, X. Ren and T.-S. Huang, N-halamine antibacterial nanofibrous mats based on polyacrylonitrile and N-halamine for protective face masks, *J. Eng. Fibers Fabr.*, 2019, **14**, 155892501984322.
- 113 G. Thilagavathi and T. Kannaian, Dual antimicrobial and blood repellent finishes for cotton hospital fabrics, *Indian J. Fibre Text. Res.*, 2008, **23**–29 <http://nopr.niscair.res.in/handle/123456789/364>.
- 114 V. Balachandar, I. Mahalaxmi, J. Kaavya, G. Vivekanandhan, S. Ajithkumar, N. Arul, *et al.*, COVID-19: emerging protective measures, *Eur. Rev. Med. Pharmacol. Sci.*, 2020, **24**, 3422–3425.



Review

- 115 H. Belbachir and J. Angelidis, Decontaminating composition having simultaneously bactericidal, fungicidal and virocidal properties, methods for obtaining and using said composition, *US Pat.*, US8529968B2, 2013.
- 116 M. Reynolds, Antimicrobial compositions and applications therefore, US20100092398A1, 2010, <https://patents.google.com/patent/US20100092398A1/en>.
- 117 H. J. Kim and C. J. Park, Composition for preventing infection of new influenza a (H1N1) virus comprising ginkgo extract, air filter comprising the same, and air cleaning device comprising the filter, US20120148507A1, 2012, <https://patents.google.com/patent/US20120148507A1/en>.
- 118 H. J. Kim and C. J. Park, Composition for prevention of influenza viral infection comprising sumac extract, air filter comprising the same and air cleaning device comprising the filter, US9119814B2, 2015, <https://patents.google.com/patent/US9119814B2/en>.
- 119 P. Ekabutr, P. Chuysinuan, S. Suksamrarn, W. Sukhumsirichart, P. Hongmanee and P. Supaphol, Development of antituberculosis melt-blown polypropylene filters coated with mangosteen extracts for medical face mask applications, *Polym. Bull.*, 2019, **76**, 1985–2004.
- 120 Y.-F. Wang, F. Kang, S.-J. You, C.-H. Tsai and G.-L. Lin, Preparation and characteristic of antibacterial facemasks with chinese herbal microcapsules, *Aerosol Air Qual. Res.*, 2017, **17**, 2120–2129.
- 121 K. Chen, X. Qiu, D. Yang and Y. Qian, Amino acid-functionalized polyampholytes as natural broad-spectrum antimicrobial agents for high-efficient personal protection, *Green Chem.*, 2020, **19**, 6357–6371.
- 122 J. R. Morones, J. L. Elechiguerra, A. Camacho, K. Holt, J. B. Kouri, J. T. Ramirez, *et al.*, The bactericidal effect of silver nanoparticles, *Nanotechnology*, 2005, **16**, 2346–2353.
- 123 K.-J. Kim, W. S. Sung, B. K. Suh, S.-K. Moon, J.-S. Choi, J. G. Kim, *et al.*, Antifungal activity and mode of action of silver nano-particles on *Candida albicans*, *BioMetals*, 2009, **22**, 235–242.
- 124 L. Lu, R. W.-Y. Sun, R. Chen, C.-K. Hui, C.-M. Ho, J. M. Luk, *et al.*, Silver nanoparticles inhibit hepatitis B virus replication, *Antiviral Ther.*, 2008, **13**(2), 253.
- 125 S. Park, H. H. Park, S. Y. Kim, S. J. Kim, K. Woo and G. Ko, Antiviral properties of silver nanoparticles on a magnetic hybrid colloid, *Appl. Environ. Microbiol.*, 2014, **80**, 2343–2350.
- 126 G. Franci, A. Falanga, S. Galdiero, L. Palomba, M. Rai, G. Morelli, *et al.*, Silver nanoparticles as potential antibacterial agents, *Molecules*, 2015, **20**, 8856–8874.
- 127 M. Horie, H. Ogawa, Y. Yoshida, K. Yamada, A. Hara, K. Ozawa, *et al.*, Inactivation and morphological changes of avian influenza virus by copper ions, *Arch. Virol.*, 2008, **153**, 1467–1472.
- 128 A. Brandelli, A. C. Ritter, F. F. Veras, *Antimicrobial Activities of Metal Nanoparticles*, in *Metal Nanoparticles in Pharma*, ed. M. Rai and R. Shegokar, Springer, Cham, 2017, pp. 337–363.
- 129 R. K. Matharu, L. Ciric, G. Ren and M. Edirisinghe, Comparative study of the antimicrobial effects of tungsten nanoparticles and tungsten nanocomposite fibres on hospital acquired bacterial and viral pathogens, *Nanomaterials*, 2020, **10**, 1–16.
- 130 U. Baig, M. A. Gondal, S. Rehman and S. Akhtar, Facile synthesis, characterization of nano-tungsten trioxide decorated with silver nanoparticles and their antibacterial activity against water-borne gram-negative pathogens, *Appl. Nanosci.*, 2020, **10**, 851–860.
- 131 S. Galdiero, A. Falanga, M. Vitiello, M. Cantisani, V. Marra and M. Galdiero, Silver nanoparticles as potential antiviral agents, *Molecules*, 2011, **16**, 8894–8918.
- 132 T. Tashi, N. V. Gupta and V. B. Mbuya, Silver nanoparticles: Synthesis, mechanism of antimicrobial action, characterization, medical applications, and toxicity effects, *J. Chem. Pharmaceut. Res.*, 2016, **8**(2), 526–537.
- 133 H. J. Kim, S. W. Han, M. K. Joshi and C. S. Kim, Fabrication and characterization of silver nanoparticle-incorporated bilayer electrospun-melt-blown micro/nanofibrous membrane, *Int. J. Polym. Mater. Polym. Biomater.*, 2017, **66**, 514–520.
- 134 A. Pasricha, S. L. Jangra, N. Singh, N. Dilbaghi, K. N. Sood, K. Arora, *et al.*, Comparative study of leaching of silver nanoparticles from fabric and effective effluent treatment, *J. Environ. Sci.*, 2012, **24**, 852–859.
- 135 X. Zhu, L. Tang, K.-H. Wee, Y.-H. Zhao and R. Bai, Immobilization of silver in polypropylene membrane for anti-biofouling performance, *Biofouling*, 2011, **27**, 773–786.
- 136 S. Agarwal, A. Greiner and J. H. Wendorff, Functional materials by electrospinning of polymers, *Prog. Polym. Sci.*, 2013, **38**, 963–991.
- 137 A. Brochocka and K. Majchrzycka, Technology for the production of bioactive melt-blown filtration materials applied to respiratory protective devices, *Fibres Text. East. Eur.*, 2009, **17**, 92–98.
- 138 K. R. Aadil, S. I. Mussatto and H. Jha, Synthesis and characterization of silver nanoparticles loaded poly(vinyl alcohol)-lignin electrospun nanofibers and their antimicrobial activity, *Int. J. Biol. Macromol.*, 2018, **120**, 763–767.
- 139 ASTM, ASTM F2101 - 01 Standard Test Method for Evaluating the Bacterial Filtration Efficiency (BFE) of Medical Face Mask Materials, Using a Biological Aerosol of *Staphylococcus aureus*, 2019, <https://compass.astm.org/Standards/HISTORICAL/F2101-01.htm>.
- 140 M. G. Baldry, The antimicrobial properties of magnesium monoperoxyphthalate hexahydrate, *J. Appl. Bacteriol.*, 1984, **57**, 499–503.
- 141 J. L. Sagripanti, P. L. Goering and A. Lamanna, Interaction of copper with DNA and antagonism by other metals, *Toxicol. Appl. Pharmacol.*, 1991, **110**, 477–485.
- 142 N. P. Klochko, K. S. Klepikova, D. O. Zhadan, V. R. Kopach, S. M. Chernyavskaya, S. I. Petrusenko, *et al.*, Thermoelectric textile with fibers coated by copper iodide thin films, *Thin Solid Films*, 2020, **704**, 138026.
- 143 EN 14683:2005, *Surgical masks - Requirements and test methods*, European Nation standards, 2005<https://>



- standards.iteh.ai/catalog/standards/cen/12acf007-918b-4825-9217-4b4a96938dcf/en-14683-2005.
- 144 ASTM F1862/F1862M - 17, *Standard test method for resistance of medical face masks to penetration by synthetic blood (Horizontal Projection of Fixed Volume at a Known Velocity)*, 2017 <https://www.astm.org/Standards/F1862.htm>.
- 145 M. Hashmi, S. Ullah and I. S. Kim, Copper oxide (CuO) loaded polyacrylonitrile (PAN) nanofiber membranes for antimicrobial breath mask applications, *Curr. Res. Biotechnol.*, 2019, **1**, 1–10.
- 146 ASTM E96/E96M - 16, *Standard test methods for water vapor transmission of materials*, <https://www.astm.org/Standards/E96>.
- 147 AATCC, <https://members.aatcc.org/store/tm100/513/>.
- 148 ASTM F2100 - 20, Standard specification for performance of materials used in medical face masks, <https://www.astm.org/Standards/F2100.htm>.
- 149 European Committee for Standardization, *CEN - Technical Bodies*, https://standards.cen.eu/dyn/www/?p=204:110:0:::FSP_PROJECT,FSP_ORG_ID:32928,6062&cs=1FC98AD34A5EE26A0CB5A6155ED4D6E5E.
- 150 M. Charnley, M. Textor and C. Acikgoz, Designed polymer structures with antifouling–antimicrobial properties, *React. Funct. Polym.*, 2011, **71**(3), 329–334 <https://linkinghub.elsevier.com/retrieve/pii/S1381514810001872>.
- 151 K.-S. Huang, C.-H. Yang, S.-L. Huang, C.-Y. Chen, Y.-Y. Lu and Y.-S. Lin, Recent advances in antimicrobial polymers: A mini-review, *Int. J. Mol. Sci.*, 2016, **17**, 1578.
- 152 M. R. Badrossamay and G. Sun, Preparation of rechargeable biocidal polypropylene by reactive extrusion with diallylamino triazine, *Eur. Polym. J.*, 2008, **44**, 733–742.
- 153 A. Jain, L. S. Duvvuri, S. Farah, N. Beyth, A. J. Domb and W. Khan, Antimicrobial polymers, *Adv. Healthcare Mater.*, 2014, **3**, 1969–1985.
- 154 S. D. Worley, D. E. Williams and R. A. Crawford, Halamine water disinfectants, *Crit. Rev. Environ. Control*, 1988, **18**, 133–175.
- 155 R. Nayak and R. Padhye, 12 - Antimicrobial finishes for textiles, in *Functional Finishes for Textiles*, ed. R. Paul, Woodhead Publishing, 2015, pp. 361–85, Woodhead Publishing Series in Textiles, <http://www.sciencedirect.com/science/article/pii/B9780857098399500121>.
- 156 L.-C. Xu and C. A. Siedlecki, Antibacterial polyurethanes, in *Advances in Polyurethane Biomaterials*, ed. S. L. Cooper and J. Guan, Woodhead Publishing, 2016, pp. 247–284, <http://www.sciencedirect.com/science/article/pii/B9780081006146000093>.
- 157 C. Yao, X. Li, K. G. Neoh, Z. Shi and E. T. Kang, Antibacterial activities of surface modified electrospun poly(vinylidene fluoride-co-hexafluoropropylene) (PVDF-HFP) fibrous membranes, *Appl. Surf. Sci.*, 2009, **255**, 3854–3858.
- 158 A. Carmona-Ribeiro and L. de Melo Carrasco, Cationic antimicrobial polymers and their assemblies, *Int. J. Mol. Sci.*, 2013, **14**, 9906–9946.
- 159 S. P. Denyer, Mechanisms of action of antibacterial biocides, *Int. Biodeterior. Biodegrad.*, 1995, **36**, 227–245.
- 160 J. C. Tiller, C.-J. Liao, K. Lewis and A. M. Klibanov, Designing surfaces that kill bacteria on contact, *Proc. Natl. Acad. Sci. U. S. A.*, 2001, **98**, 5981–5985.
- 161 J. C. Tiller, S. B. Lee, K. Lewis and A. M. Klibanov, Polymer surfaces derivatized with poly(vinyl-N-hexylpyridinium) kill airborne and waterborne bacteria, *Biotechnol. Bioeng.*, 2002, **79**, 465–471.
- 162 K. Nakata, T. Tsuchido and Y. Matsumura, Antimicrobial cationic surfactant, cetyltrimethylammonium bromide, induces superoxide stress in Escherichia coli cells, *J. Appl. Microbiol.*, 2011, **110**, 568–579.
- 163 K. Majchrzycka, B. Gutarowska, A. Brochocka and B. Brycki, New filtering antimicrobial nonwovens with various carriers for biocides as respiratory protective materials against bioaerosol, *Int. J. Occup. Saf. Ergon.*, 2012, **18**, 375–385.
- 164 K. Majchrzycka, M. Okrasa, J. Szulc, B. Brycki and B. Gutarowska, Time-Dependent Antimicrobial Activity of Filtering Nonwovens with Gemini Surfactant-Based Biocides, *Molecules*, 2017, **22**, 1620.
- 165 ASTM E2721 - 10, Standard test method for evaluation of effectiveness of decontamination procedures for surfaces when challenged with droplets containing human pathogenic viruses, 2010 <https://www.astm.org/DATABASE.CART/HISTORICAL/E2721-10.htm>.
- 166 A. Purohit, M. C. Kopferschmitt-Kubler, C. Moreau, E. Popin, M. Blaumeiser and G. Pauli, Quaternary ammonium compounds and occupational asthma, *Int. Arch. Occup. Environ. Health*, 2000, **73**, 423–427.
- 167 S. E. Anderson, H. Shane, C. Long, E. Lukomska, B. J. Meade and N. B. Marshall, Evaluation of the irritancy and hypersensitivity potential following topical application of didecyltrimethylammonium chloride, *J. Immunotoxicol.*, 2016, **13**, 557–566.
- 168 A. Varesano, C. Vineis, A. Aluigi, F. Rombaldoni, C. Tonetti and G. Mazzuchetti, Antibacterial efficacy of polypyrrole in textile applications, *Fibers Polym.*, 2013, **14**, 36–42.
- 169 K. H. Hong and G. Sun, Photoinduced antimicrobial polymer blends with benzophenone as a functional additive, *J. Appl. Polym. Sci.*, 2009, **112**, 2019–2026.
- 170 J. L. Fox, Antimicrobial peptides stage a comeback, *Nat. Biotechnol.*, 2013, **31**, 379–382.
- 171 G. J. Gabriel, A. Som, A. E. Madkour, T. Eren and G. N. Tew, Infectious disease: Connecting innate immunity to biocidal polymers, *Mater. Sci. Eng., R*, 2007, **57**, 28.
- 172 ASTM Compass, https://compass.astm.org/EDIT/html_annot.cgi?E2180+18.
- 173 V. Geetha, A. Doss and A. P. A. Doss, Antimicrobial potential of Vitex trifolia Linn, *Anc. Sci. Life*, 2004, **23**, 30–32.
- 174 S. Esath Natheer, Evaluation of antibacterial activity of Morinda citrifolia, Vitex trifolia and Chromolaena odorata, *Afr. J. Pharm. Pharmacol.*, 2012, **6**(29), 10–14.
- 175 P. S. Negi and G. K. Jayaprakash, Antioxidant and antibacterial activities of punica granatum peel extracts, *J. Food Sci.*, 2003, **68**, 1473–1477.
- 176 M. Haidari, M. Ali, S. Ward Casscells and M. Madjid, Pomegranate (Punica granatum) purified polyphenol



- extract inhibits influenza virus and has a synergistic effect with oseltamivir, *Phytomedicine*, 2009, **16**, 1127–1136.
- 177 P. Jadhav, N. Kapoor, B. Thomas, H. Lal and N. Kshirsagar, Antiviral potential of selected Indian medicinal (ayurvedic) plants against herpes simplex virus 1 and 2, *N. Am. J. Med. Sci.*, 2012, **4**, 641.
- 178 N. D. Weber, D. O. Andersen, J. A. North, B. K. Murray, L. D. Lawson and B. G. Hughes, In vitro virucidal effects of *Allium sativum* (Garlic) extract and compounds, *Planta Med.*, 1992, **58**, 417–423.
- 179 L. P. Rees, S. F. Minney, N. T. Plummer, J. H. Slater and D. A. Skyrme, A quantitative assessment of the antimicrobial activity of garlic (*Allium sativum*), *World J. Microbiol. Biotechnol.*, 1993, **9**, 303–307.
- 180 N. Benkeblia, Antimicrobial activity of essential oil extracts of various onions (*Allium cepa*) and garlic (*Allium sativum*), *LWT-Food Sci. Technol.*, 2004, **37**, 263–268.
- 181 A. Banso, *Phytochemical and antibacterial investigation of bark extracts of Acacia nilotica*, 2009, <https://paper/Phytochemical-and-antibacterial-investigation-of-of-Banso/5ecd8a95cc2402e2c0bc3e53f0ae5eb3c291e46b>.
- 182 C. Wiart, K. Kumar, M. Y. Yusof, H. Hamimah, Z. M. Fauzi and M. Sulaiman, Antiviral properties of ent-labdene diterpenes of *Andrographis paniculata* nees, inhibitors of herpes simplex virus type 1, *Phytother. Res.*, 2005, **19**, 1069–1070.
- 183 S. Roy, K. Rao, C. Bhuvanewari, A. Giri and L. N. Mangamoori, Phytochemical analysis of *Andrographis paniculata* extract and its antimicrobial activity, *World J. Microbiol. Biotechnol.*, 2009, **26**, 85.
- 184 V. J. Galani, B. G. Patel and D. G. Rana, *Sphaeranthus indicus* Linn.: A phytopharmacological review, *Int. J. Res. Ayurveda Pharm.*, 2010, **1**, 247–253.
- 185 W. Gu, W. Wang, X. Li, Y. Zhang, L. Wang, C. Yuan, *et al.*, A novel isocoumarin with anti-influenza virus activity from *Strobilanthes cusia*, *Fitoterapia*, 2015, **107**, 60–62.
- 186 Y.-C. Tsai, C.-L. Lee, H.-R. Yen, Y.-S. Chang, Y.-P. Lin, S.-H. Huang, *et al.*, Antiviral action of tryptanthrin isolated from *Strobilanthes cusia* leaf against human coronavirus NL63, *Biomolecules*, 2020, **10**, 366.
- 187 D. V. Zige, E. I. Ohimain and M. B. Nodu, Antibacterial activity of ethanol, crude and water extract of *Chromolaena odorata* leaves on *S. typhi* and *E. coli*, *Greener J. Microbiol. Antimicrob.*, 2013, **1**, 016–019.
- 188 K. Saoo, H. Miki, M. Ohmori and W. D. Winters, Antiviral activity of Aloe extracts against Cytomegalovirus, *Phytother. Res.*, 1996, **10**, 348–350.
- 189 K. Zandi, M. A. Zadeh, K. Sartavi and Z. Rastian, Antiviral activity of Aloe vera against herpes simplex virus type 2: An *in vitro* study, *Afr. J. Biotechnol.*, 2007, **6**(15), 1770–1773.
- 190 R. Pandey and A. Mishra, Antibacterial activities of crude extract of *Aloe barbadensis* to clinically isolated bacterial pathogens, *Appl. Biochem. Biotechnol.*, 2010, **160**, 1356–1361.
- 191 M. Joshi, S. W. Ali and S. Rajendran, Antibacterial finishing of polyester/cotton blend fabrics using neem (*Azadirachta indica*): A natural bioactive agent, *J. Appl. Polym. Sci.*, 2007, **106**, 793–800.
- 192 M. Kubo, Y. Kimura, T. Odani, T. Tani and K. Namba, Studies on *Scutellariae Radix*, *Planta Med.*, 1981, **43**, 194–201.
- 193 C. Duan, S. Matsumura, N. Kariya, M. Nishimura and T. Shimono, In vitro antibacterial activities of *Scutellaria baicalensis* Georgi against cariogenic bacterial, *Pediatr. Dent. J.*, 2007, **17**, 58–64.
- 194 S. Duong-Quy, Study of active compounds of *Plectranthii amboinicii* leaves growing in Dalat city, Vietnam and its antibacterial and antioxidant activities, *World J. Pharm. Pharmaceut. Sci.*, 2017, **6**(4), 77–94.
- 195 M. Catel-Ferreira, H. Thani, C. Hellio, P. Cosette and L. Lebrun, Antiviral effects of polyphenols: Development of bio-based cleaning wipes and filters, *J. Virol. Methods*, 2015, **212**, 1–7.
- 196 F.-S. Quan, I. Rubino, S.-H. Lee, B. Koch and H.-J. Choi, Universal and reusable virus deactivation system for respiratory protection, *Sci. Rep.*, 2017, **7**, 39956.
- 197 H.-J. Choi, D.-G. Yoo, B. J. Bondy, F.-S. Quan, R. W. Compans, S.-M. Kang, *et al.*, Stability of influenza vaccine coated onto microneedles, *Biomaterials*, 2012, **33**, 3756–3769.
- 198 Y. Fujimori, T. Nakayama and T. Sato, Antiviral agent, US20170086463A1, 2017, <https://patents.google.com/patent/US20170086463A1/en>.
- 199 B. K. Heimbuch and J. D. Wander, *Bioaerosol challenges to antimicrobial surface treatments: Enhanced Efficacy Against MS2 Coli Phage of Air Filter Media Coated With Polystyrene-4-Methyltrimethylammonium Triiodide*, Appl. Res. Assoc. Inc., Tyndall AFB, FL, 2006, vol. 10.
- 200 W. Hu, C. Peng, W. Luo, M. Lv, X. Li, D. Li, *et al.*, Graphene-based antibacterial paper, *ACS Nano*, 2010, **4**, 4317–4323.
- 201 P. Kumar, P. Huo, R. Zhang and B. Liu, Antibacterial properties of graphene-based nanomaterials, *Nanomaterials*, 2019, **9**(5:737), 1–32.
- 202 N. Noor, S. Mutalik, M. W. Younas, C. Y. Chan, S. Thakur, F. Wang, *et al.*, Durable antimicrobial behaviour from silver-graphene coated medical textile composites, *Polymers*, 2019, **11**, 2000.
- 203 J. Zhao, B. Deng, M. Lv, J. Li, Y. Zhang, H. Jiang, *et al.*, Graphene oxide-based antibacterial cotton fabrics, *Adv. Healthcare Mater.*, 2013, **2**, 1259–1266.
- 204 F. Yaghoubidoust and E. Salimi, A Simple method for the preparation of antibacterial cotton fabrics by coating graphene oxide nanosheets, *Fibers Polym.*, 2019, **20**, 1155–1160.
- 205 M. S. Stan, I. C. Nica, M. Popa, M. C. Chifiriuc, O. Iordache, I. Dumitrescu, *et al.*, Reduced graphene oxide/TiO₂ nanocomposites coating of cotton fabrics with antibacterial and self-cleaning properties, *J. Ind. Text.*, 2019, **49**, 277–293.
- 206 A. Kumar, K. Sharma and A. R. Dixit, Role of graphene in biosensor and protective textile against viruses, *Med. Hypotheses*, 2020, **144**, 110253.



- 207 J. Molina, Graphene-based fabrics and their applications: a review, *RSC Adv.*, 2016, **6**, 68261–68291.
- 208 J. Lin, Z. Peng, Y. Liu, F. Ruiz-Zepeda, R. Ye, E. L. G. Samuel, *et al.*, Laser-induced porous graphene films from commercial polymers, *Nat. Commun.*, 2014, **5**, 5714.
- 209 R. Ye, Y. Chyan, J. Zhang, Y. Li, X. Han, C. Kittrell, *et al.*, Laser-induced graphene formation on wood, *Adv. Mater.*, 2017, **29**, 1702211.
- 210 H. Zhong, Z. Zhu, J. Lin, C. F. Cheung, V. L. Lu, F. Yan, *et al.*, Reusable and recyclable graphene masks with outstanding superhydrophobic and photothermal performances, *ACS Nano*, 2020, **14**(5), 6213–6221.
- 211 L. Huang, S. Xu, Z. Wang, K. Xue, J. Su, Y. Song, *et al.*, Self-reporting and photothermally enhanced rapid bacterial Killing on a laser-induced graphene mask, *ACS Nano*, 2020, **14**, 12045–12053.
- 212 R. R. Bresee and U. A. Qureshi, Influence of processing conditions on melt blown web structure: Part 1 – DCD, *Int. Nonwovens J.*, 2004, **os-13**, 1558925004os–13.
- 213 H. Kaoud, *Tissue Regeneration*, IntechOpen publisher, 2018, ISBN 978-1-78923-261-5.
- 214 L. E. Scriven, Physics and applications of dip coating and spin coating, *MRS Proc.*, 1988, **121**, 717.
- 215 D. R. Ceratti, B. Louis, X. Paquez, M. Faustini and D. Grosso, A new dip coating method to obtain large-surface coatings with a minimum of solution, *Adv. Mater.*, 2015, **27**, 4958–4962.
- 216 D. Park, J. Wang and A. M. Klivanov, One-step, painting-like coating procedures to make surfaces highly and permanently bactericidal, *Biotechnol. Prog.*, 2006, **22**, 584–589.
- 217 H. Singh, T. S. Sidhu and S. B. S. Kalsi, Cold spray technology: future of coating deposition processes, *Frat. ed Integrita Strutt.*, 2012, **6**, 69–84.
- 218 W. JIC and P. P. Tsai, Cotton-comfortable face masks with protective finishes and electret filter media for safety from microbial threats, *Res. J. Text. Apparel*, 2006, **10**, 33–43.
- 219 I. M. Hutten, Processes for nonwoven filter media, in *Handbook of Nonwoven Filter Media*, ed. I. M. Hutten, Butterworth-Heinemann, Oxford, 2007, pp. 195–244, <http://www.sciencedirect.com/science/article/pii/B9781856174411500202>.
- 220 J. G. McCulloch, The history of the development of melt blowing technology, *Int. Nonwovens J.*, 1999, **os-8**, 1558925099OS–80.
- 221 N. Hiremath and G. Bhat, Melt blown polymeric nanofibers for medical applications-an overview, *Nanosci. Nanotechnol.*, 2015, **2**, 1–9.
- 222 B. Gutarowska and A. Michalski, Antimicrobial activity of filtrating meltblown nonwoven with the addition of silver ions, *Fibres Text. East. Eur.*, 2009, **3**(74), 23–28.
- 223 J. Xue, T. Wu, Y. Dai and Y. Xia, Electrospinning and electrospun nanofibers: Methods, materials, and applications, *Chem. Rev.*, 2019, **119**, 5298–5415.
- 224 M. Zhu, J. Han, F. Wang, W. Shao, R. Xiong, Q. Zhang, *et al.*, Electrospun nanofibers membranes for effective air filtration, *Macromol. Mater. Eng.*, 2017, **302**, 1600353.
- 225 N. Bhardwaj and S. C. Kundu, Electrospinning: A fascinating fiber fabrication technique, *Biotechnol. Adv.*, 2010, **28**, 325–347.
- 226 *Electrospun Nanofibers for Energy and Environmental Applications*, ed. B. Ding and J. Yu, Springer Berlin Heidelberg, Berlin, Heidelberg, 2014, Nanostructure Science and Technology, <http://link.springer.com/10.1007/978-3-642-54160-5>.
- 227 M. Zahmatkeshan, M. Adel, S. Bahrami, F. Esmaeili, S. M. Rezayat, Y. Saeedi, *et al.*, Polymer-Based Nanofibers: Preparation, Fabrication, and Applications, in *Handbook of Nanofibers*, A. Barhoum, M. Bechelany and A. S. H. Makhlof, Springer International Publishing, Cham, 2019, pp. 215–261, https://doi.org/10.1007/978-3-319-53655-2_29.
- 228 R. Purwar, K. Sai Goutham and C. M. Srivastava, Electrospun Sericin/PVA/Clay nanofibrous mats for antimicrobial air filtration mask, *Fibers Polym.*, 2016, **17**, 1206–1216.
- 229 R. Bai, Q. Zhang, L. Li, P. Li, Y.-J. Wang, O. Simalou, *et al.*, N-Halamine-containing electrospun fibers kill bacteria via a contact/release co-determined antibacterial pathway, *ACS Appl. Mater. Interfaces*, 2016, **8**, 31530–31540.
- 230 A. K. Leichman, *7 Israeli mask and face shield solutions for coronavirus*, ISRAEL21c, 2020, <https://www.israel21c.org/7-israeli-mask-and-face-shield-solutions-for-coronavirus/>.
- 231 Sonovia, *Innovative active protection*, 2020, <https://sonoviatech.com/>.
- 232 tradeKorea, *Goldshield 3DN95 medical mask/tradeKorea*, tradeKorea, Global B2B Trade Website - Offer Global Business, Buyer Matching Services, 2020, <https://www.tradeKorea.com/product/detail/P756854/Goldshield-3DN95-medical-mask.html>.
- 233 Good research solutions LLC. silver shield Ag+, silver shield masks, 2020, <https://www.silvershieldmasks.com/products/face-mask-black>.
- 234 Nonwovens Industry, *Nexera launches first FDA-approved antimicrobial surgical respiratory mask*, Nonwovens Industry Magazine- News, Markets & Analysis for the Nonwovens Industry, 2016 https://www.nonwovens-industry.com/issues/2016-06-06/view_breaking-news/nexera-launches-first-fda-approved-antimicrobial-surgical-respiratory-mask/.
- 235 Copper3d. *Hack the Pandemic – Copper 3D/Antibacterial 3D Printing*, 2020, <https://copper3d.com/hackthepandemic/>.
- 236 Copper and Copper Clothing, *Copper clothing copper infused face mask*, 2020, <https://www.copperclothing.com/product/copper-infused-face-mask/>.
- 237 Noveko International Inc, *Noveko receives the green light to market its new antimicrobial respirator model in Europe* n.d., 2020, <https://www.newswire.ca/news-releases/noveko-receives-the-green-light-to-market-its-new-antimicrobial-respirator-model-in-europe-544931932.html>.
- 238 Scientific. FischerSafe, *Life™ Disposable respirators with Triosyn™ antimicrobial protection*, 2020, <https://www.fishersci.com/shop/products/safe-life-disposable-respirators-triosyn-antimicrobial-protection-universal/19171016>.



- 239 Infection Control Today, *Safe life introduces N95 respirator with antimicrobial protection*, Infection Control Today, 2009 <https://www.infectioncontroltoday.com/view/safe-life-introduces-n95-respirator-antimicrobial-protection>.
- 240 Nanosafe Solutions P. Ltd, *Nanosafe Solutions P. Ltd./India*, 2020, <https://nanosafesolutions.com/fight-against-covid-19.php>.
- 241 Erlanger, *Union springs pharmaceuticals launches new disposable respirator with antimicrobial coating*, n.d. EMS1, 2020, <https://www.ems1.com/ems-products/ambulance-disposable-supplies/press-releases/union-springs-pharmaceuticals-launches-new-disposable-respirator-with-antimicrobial-coating-61XAN4zzmncYQZj/>.
- 242 R. Peleg, *planarTECH and IDEATI launch graphene-enhanced antibacterial face masks*/Graphene-Info, 2020, <https://www.graphene-info.com/planartech-and-ideati-launch-graphene-enhanced-antibacterial-face-masks>.
- 243 Yanko Design, *World's first transparent FDA registered, N99+ smart mask with UV-C sterilizing that shows-off your smile*/Yanko Design, 2020, <https://www.yankodesign.com/2020/06/27/worlds-first-transparent-fda-registered-n99-smart-mask-with-uv-c-sterilizing-that-shows-off-your-smile/>.
- 244 T N, *COVID-[Update], 19: First FDA-registered transparent N99+ mask with UV-C sterilizing now available*/Tech Times, 2020 <https://www.techtimes.com/articles/250624/20200626/covid-19-update-first-fda-approved-transparent-n99-mask-with-uv-c-sterilizing-now-available.htm>.
- 245 Research and Markets, *Global disposable face mask market (2020 to 2025) - Growth, trends and forecasts*, GlobeNewswire News Room, 2020 <http://www.globenewswire.com/news-release/2020/06/15/2047842/0/en/Global-Disposable-Face-Mask-Market-2020-to-2025-Growth-Trends-and-Forecasts.html>.
- 246 Zion Market Research, *Global N95 mask market set for rapid growth, to reach Value USD 1,890.3 Million by 2026*, 2020 <https://www.zionmarketresearch.com/news/n95-mask-market>.
- 247 Grand View Research, *Disposable face mask market size*/Industry Report, 2020-2027, 2020, <https://www.grandviewresearch.com/industry-analysis/disposable-face-masks-market>.
- 248 K. Bradsher and L. Alderman, *The world needs masks. China makes them, but has been hoarding them*, The New York Times, 2020, <https://www.nytimes.com/2020/03/13/business/masks-china-coronavirus.html>.
- 249 C. Zhou, *Coronavirus bubble bursts for China's low-end mask makers*, South China Morning Post. 2020, <https://www.scmp.com/economy/china-economy/article/3088810/coronavirus-wheels-come-chinas-mask-making-gravy-train-low>.
- 250 CISION, *Global Face Mask Market Overview 2020-2025: Growth Analysis of N95, KN95, FFP2 & FFP3, P95 and R95 Masks Amid COVID-19*, 2020, <https://www.prnewswire.com/news-releases/global-face-mask-market-overview-2020-2025-growth-analysis-of-n95-kn95-ffp2-ffp3-p95-and-r95-masks-amid-covid-19-301101156.html>.
- 251 Research and Markets, *Face mask market by type, material, nature, and distribution channel: Global Opportunity Analysis and Industry Forecast 2021-2027*, 2020, <https://www.researchandmarkets.com/reports/5125521/face-mask-market-by-type-material-nature-and>.
- 252 La Grande Traversee, *Mask Blog*, 2020, <https://www.lagrandetraversee.ca/>.
- 253 Fortune Business Insights, *Breathable antimicrobial coatings market size, share, Industry Forecast 2026*, 2020, <https://www.fortunebusinessinsights.com/breathable-antimicrobial-coatings-market-102764>.
- 254 H. Horvath, *Buying antimicrobial face masks? Here's what experts recommend*, NBC News, 2020, <https://www.nbcnews.com/shopping/apparel/best-antimicrobial-face-masks-n1231803>.
- 255 J. C. Prata, A. L. P. Silva, T. R. Walker, A. C. Duarte and T. Rocha-Santos, *COVID-19 Pandemic repercussions on the use and management of plastics*, *Environ. Sci. Technol.*, 2020, **54**, 7760–7765.
- 256 L. Tenenbaum, *The amount of plastic waste is surging because of the coronavirus pandemic*, Forbes, n.d. 2020, <https://www.forbes.com/sites/lauratenenbaum/2020/04/25/plastic-waste-during-the-time-of-covid-19/>.
- 257 K. R. Vanapalli, H. B. Sharma, V. P. Ranjan, B. Samal, J. Bhattacharya, B. K. Dubey, *et al.*, *Challenges and strategies for effective plastic waste management during and post COVID-19 pandemic*, *Sci. Total Environ.*, 2020, **750**, 141514.
- 258 J. Katz, M. Sanger-Katz and K. Quealy, *A detailed map of who is wearing masks in the U.S.*, The New York Times, 2020, <https://www.nytimes.com/interactive/2020/07/17/upshot/coronavirus-face-mask-map.html>.
- 259 E. Beswick, *Coronavirus: How the wearing of masks has exposed a divided Europe*, Euronews, 2020, <https://www.euronews.com/2020/07/14/coronavirus-how-the-wearing-of-face-masks-has-exposed-a-divided-europe>.
- 260 A. L. Allison, E. Ambrose-Dempster, T. Domenech Apars, M. Bawn, M. Casas Arredondo, C. Chau, *et al.*, *The environmental dangers of employing single-use face masks as part of a COVID-19 exit strategy*, *UCL Open Environ. Prepr.*, 2020.
- 261 S. Jones, *The Great Mask Divide: Lessons from Asia*, Asia Pacific Foundation of Canada, 2020, <https://www.asiapacific.ca/publication/great-mask-divide-lessons-asia>.
- 262 A. Reddy, *Delhi to Vijayawada, India has started dumping Covid-19 infected waste in public places*, ThePrint, 2020, <https://theprint.in/opinion/delhi-vijayawada-india-dumping-covid-19-infected-waste-public-places/496396/>.
- 263 K. Samir Purkayastha, *COVID-19: Rise in biomedical waste generation poses fresh challenge*, The Federal, 2020, <https://thefederal.com/news/covid-19-rise-in-biomedical-waste-generation-poses-fresh-challenge/>.
- 264 B. Chatterjee, *Mumbai's Covid-19 waste doubles in 3 months, treatment facility runs out of storage space*, Hindustan Times, 2020, <https://www.hindustantimes.com/india-news/city-s-covid-19-waste-doubles-in-3-months-treatment-facility>.



- runs-out-of-storage-space/story-RB9z2efys8dlctIXXuXL0H.html.
- 265 S. Ramteke and B. L. Sahu, Novel coronavirus disease 2019 (COVID-19) pandemic: considerations for the biomedical waste sector in India, *Case Stud. Chem. Environ. Eng.*, 2020, 100029.
- 266 M. Varmani, *Bio-medical waste management during COVID-19*, 2020, <https://www.investindia.gov.in/team-india-blogs/bio-medical-waste-management-during-covid-19>.
- 267 M. A. Zambrano-Monserrate, M. A. Ruano and L. Sanchez-Alcalde, Indirect effects of COVID-19 on the environment, *Sci. Total Environ.*, 2020, 728, 138813.
- 268 J. Calma, *The COVID-19 pandemic is generating tons of medical waste*, The Verge, 2020, <https://www.theverge.com/2020/3/26/21194647/the-covid-19-pandemic-is-generating-tons-of-medical-waste>.
- 269 I. Tiseo, *COVID-19: Healthcare waste generation in Asia 2020*, Statista, 2020, <https://www.statista.com/statistics/1167512/healthcare-waste-generation-before-during-covid-19-asia-by-city/>.
- 270 United Nations, *Five things you should know about disposable masks and plastic pollution*, UN News, 2020, <https://news.un.org/en/story/2020/07/1069151>.
- 271 T. A. Aragaw, Surgical face masks as a potential source for microplastic pollution in the COVID-19 scenario, *Mar. Pollut. Bull.*, 2020, 159, 111517.
- 272 J. Lynch, *The Plastic That Washes up on Hawaii's beaches affects me personally*, NIST, 2019, <https://www.nist.gov/blogs/taking-measure/plastic-washes-hawaiis-beaches-affects-me-personally>.
- 273 United Nation Environment Programme, *COVID-19 Waste management factsheets*, UNEP - UN Environment Programme, 2020, <http://www.unenvironment.org/resources/factsheet/covid-19-waste-management-factsheets>.
- 274 J. Chadwick, *Face mask remnants and other microplastics found in the guts of sharks*, Mail Online, 2020, <https://www.dailymail.co.uk/sciencetech/article-8548139/Face-masks-remnants-guts-sharks.html>.
- 275 P. C. Ray, H. Yu and P. P. Fu, Toxicity and environmental risks of nanomaterials: Challenges and future needs, *J. Environ. Sci. Health, Part C: Environ. Carcinog. Ecotoxicol. Rev.*, 2009, 27, 1–35.
- 276 M. H. Chua, W. Cheng, S. S. Goh, J. Kong, B. Li, J. Y. C. Lim, *et al.*, Face masks in the New COVID-19 normal: Materials, testing, and perspectives, *Research*, 2020, 2020, 1–40.
- 277 I. D. Posen, P. Jaramillo, A. E. Landis and W. M. Griffin, Greenhouse gas mitigation for U.S. plastics production: energy first, feedstocks later, *Environ. Res. Lett.*, 2017, 12, 034024.
- 278 I. D. Posen, P. Jaramillo and W. M. Griffin, Uncertainty in the life cycle greenhouse gas emissions from U.S. production of three biobased polymer families, *Environ. Sci. Technol.*, 2016, 50, 2846–2858.
- 279 M. N. Moore, Do nanoparticles present ecotoxicological risks for the health of the aquatic environment?, *Environ. Int.*, 2006, 32, 967–976.
- 280 D. T. Donia and M. Carbone, Fate of the nanoparticles in environmental cycles, *Int. J. Environ. Sci. Technol.*, 2019, 16, 583–600.
- 281 T. Benn, B. Cavanagh, K. Hristovski, J. D. Posner and P. Westerhoff, The release of nanosilver from consumer products used in the home, *J. Environ. Qual.*, 2010, 39, 1875–1882.
- 282 I. J. Yu, M. Gulumian, S. Shin, T. H. Yoon and V. Murashov, Occupational and environmental health effects of nanomaterials, *BioMed Res. Int.*, 2015, 2015, e789312.
- 283 K. Hegde, R. Goswami, S. J. Sarma, V. D. Veeranki, S. K. Brar and R. Y. Surampalli, *Environmental Hazards and Risks of Nanomaterials*, ResearchGate, 2015, https://www.researchgate.net/publication/283788431_Environmental_Hazards_and_Risks_of_Nanomaterials.
- 284 P. P. Fu, Q. Xia, H.-M. Hwang, P. C. Ray and H. Yu, Mechanisms of nanotoxicity: Generation of reactive oxygen species, *J. Food Drug Anal.*, 2014, 22, 64–75.
- 285 D. R. Bickers and M. Athar, Oxidative stress in the pathogenesis of skin disease, *J. Invest. Dermatol.*, 2006, 126, 2565–2575.

