

Emerging investigator series: Modeling of Wastewater Treatment Bioprocesses: Current Development and Future Opportunities

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Wastewater treatment bioprocesses are conventionally modeled using mechanistic, data-driven, or hybrid models. Herein, we identify the knowledge gaps in those models. We also propose potential modeling strategies to incorporate genomic data for handling a large amount of the physical, biochemical, and microbiological data collected from biological wastewater treatment systems, with the overarching goal to achieve real-time monitoring and optimize system performance.

1	Emerging investigator series: Modeling of Wastewater Treatment Bioprocesses: Current
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15 Abstract

16 For more than a half-century, modelers have developed various modeling strategies to facilitate 17 the transition of wastewater treatment bioprocesses from lab-scale demonstration to full-scale 18 applications. This review presents the mathematical fundamentals of mechanistic models, 19 machine learning algorithms of data-driven models, and hybrid modeling strategies for different 20 biological wastewater treatment systems including activated sludge processes, anaerobic 21 digesters, anammox processes, and bioelectrochemical systems. The discussion is focused on the 22 biological principles in those modeling strategies. The conventional Monod expressions are a 23 prevailing tool to describe the mathematical connection between microbial kinetics and state 24 variables in mechanistic models. Stoichiometric equations and steady-state conditions are also 25 required for the mechanistic modeling approach to predict system performance such as the 26 removal of carbon, nitrogen, and phosphorus. On the other hand, data-driven models statistically 27 link the inputs and outputs for the prediction and optimization of system performance with a 28 minimum requirement of a priori knowledge. Although this strategy shows outstanding learning 29 ability of data interpolation, the predictions are often uninterpretable due to the black-box nature. 30 Hybrid modeling strategies have the potential to dress the inherent limitations of standalone 31 models. Currently, the mechanistic and data-driven components in hybrid models are still 32 structured based on microbial kinetics and trained with physical and biochemical data, 33 respectively. This problem can be potentially solved by incorporating genomics data into model 34 construction to link microbial kinetic to microbial population and functional dynamics. We 35 discuss the perspectives of incorporating genomic data into model construction and propose 36 genomics-enabled hybrid modeling strategies for future research.

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- 38 Keywords: hybrid model, wastewater bioprocess, data-driven model, mechanistic model,
- 39 genomics-enabled model

40 **1. Introduction**

41 A variety of wastewater treatment systems are developed to harness microorganisms to remove 42 organic contaminants and nutrients.¹ These include activated sludge processes, anaerobic 43 digesters, and membrane bioreactors that have been extensively applied in full-scale treatment 44 plants, as well as emerging technologies such as anaerobic ammonium oxidation (anammox) and 45 bioelectrochemical systems. By taking advantage of the metabolic versatility of microorganisms, 46 some of those systems can be engineered to achieve sustainable treatment of complex waste 47 streams. For example, anammox as an anaerobic and autotrophic process converts ammonia to nitrogen gas with little energy input,² and bioelectrochemical systems can recover energy and 48 49 resource from waste streams.³

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51 Biological wastewater treatment systems are commonly monitored using real-time biochemical 52 data such as biological oxygen demand (BOD), chemical oxygen demand (COD), and mixed liquor volatile suspended solids (MLVSS) as an indication of biomass concentration.^{4–6} This is 53 54 an operationally simple method that leads to a quick assessment of the microbiological activity 55 and system performance. However, those macroscopic state variables sometimes do not help 56 explain the inconsistency in treatment performance caused by environmental and operational 57 perturbations, primarily because they are unable to reflect the complex microbial physiology, 58 community structure, metabolism, and interspecies interactions in the systems.⁴ As conventional 59 bioprocesses face challenges from emerging pollutants and more stringent discharge limits over 60 the past decades,⁷ real-time monitoring and experimental trials are laborious and frequently fall 61 short to provide insights into system optimization.

63 The high complexity of bioprocesses and the pressing need to develop more sustainable systems 64 drive environmental researchers and engineers to build computational models to gain mechanistic and predictive understandings of the dynamic behaviors in the systems.⁸ A 65 66 prevailing strategy is to build mechanistic models, also known as white-box or first-principle 67 models, based on the mathematical expressions of the physical/chemical/biological principles of the processes involved.⁸ This modeling strategy starts with defining the model purpose followed 68 69 by model structure selection, input identification, data collection/reconciliation, and model 70 calibration/correction.⁹ To improve the robustness of mechanistic models, it is important to 71 understand the fundamental microbial kinetics. Monod expressions are the most well-established 72 tool for modeling microbial growth and substrate utilization kinetics and have been extensively applied to simulate many bioprocesses.^{10–15} The kinetic parameters in Monod equations are 73 commonly derived from biochemical measurements.¹⁶ For ill-defined systems that are driven by 74 75 uncharacterized functional populations, it becomes challenging to determine which biochemical 76 indicators to measure to reliably reflect their growth and substrate utilization kinetics.

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78 Data-driven models seek to establish statistical connections between the inputs and outputs and 79 require little knowledge about the fundamental principles of the processes involved. This is of 80 great interest to the modeling of bioprocesses, in which the lifestyles of many functional unknown.¹⁷ Data-driven modeling strategies 81 populations remain started to attract 82 environmental/biological engineers' attention in the early 90s with the practical implementation of artificial neural networks (ANNs) and major advances in machine learning.¹⁸⁻²⁰ Using 83 84 appropriate training datasets and network architecture, neural networks were trained to directly predict wastewater treatment performance,^{20,21} as well as the effects of sludge volume index and 85

total nitrogen concentration on the effluent quality.²² Recent studies demonstrated the 86 applicability of several machine learning algorithms, including support vector machine, random 87 forest, extreme gradient boosting, and k-nearest neighbors, to full-scale digesters.²³ Those 88 89 algorithms have also been used to train models with biochemical data collected from literature. 90 presenting a powerful tool for data mining.²⁴ Despite the outstanding learning performance, datadriven models are black boxes that often generate uninterpretable predictions.²⁵ This is 91 92 particularly problematic for bioprocesses when the models are trained solely with physical and biochemical data.^{21-23,26} 93

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95 Hybrid models are built through the integration of mechanistic and data-driven sub-models to 96 improve the shortcomings of those individual modeling strategies.²⁷ Hybrid models are ideal for 97 ill-defined systems in which only part of the process can be mechanistically described (e.g., the 98 mass balance of measurable biochemical variables), while another part of the process is too 99 complicated to be derived from first principles (e.g. microbial interactions).²⁸ Depending on the 100 significance of the known and unknown processes in a system, the mechanistic and data-driven sub-models can be integrated through parallel, in series, or a mix of both structures.²⁹ Such 101 102 integration allows the designer to structure the model more flexibly based on the availability of 103 the a priori knowledge.²⁸ Recent studies demonstrate that hybrid models are more accurate and flexible in predicting the dynamic behaviors of wastewater treatment bioprocesses.^{30,31} 104

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This review discusses the applications and drawbacks of major modeling approaches that have been developed for simulating wastewater treatment bioprocesses over the past 60 years in chronological order (Figure 1). We first review the mechanistic modeling of several systems

109 including activated sludge processes, anaerobic digesters, anammox, and bioelectrochemical 110 systems. The discussion of this modeling strategy is focused on the biological principles, in 111 particular microbial growth and substrate utilization kinetics. In the following sections, we 112 review data-driven models capable of capturing the dynamic behaviors of bioprocesses, as well 113 as hybrid models that take advantage of both modeling strategies (Figure 2). We also discuss the 114 potential of incorporating high-throughput sequencing data into model construction to unfold the 115 underlying interactions among microbial kinetics, community structure, and functional dynamics. 116 Finally, we propose a conceptual framework to build hybrid models with omics-based results for 117 robust and interpretable prediction of microbial interactions and wastewater treatment 118 performance.

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120 2. Mechanistic Modeling of Wastewater Treatment Bioprocesses

121 **2.1 Early development of microbial kinetic model**

Microbial growth is impacted by several factors such as cell metabolic activity, substrate availability, oxygen concentration, temperature, etc. Among them, the substrate is arguably the most important one as it is directly involved in cell metabolism.³² To model the kinetic behaviors of microbial growth, a greater understanding of the effects of substrate concentration on metabolic quotients, specific growth rate, and yield is required. Between the 1920s and 1960s, many hypotheses and models were proposed to explain the biological mechanisms of substrate consumption for cellular metabolism and storage.³³

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130 The Monod equation is a microbial kinetic model that describes the hyperbolic growth behavior

131 of microbes in batch systems at exponential and steady-state phases.¹⁶ In the Monod equation,

the specific growth rate of a culture is a function of the concentration of a given substrate,³⁴ with the substrate saturation constant (i.e., the substrate concentration when the specific growth rate is half of the maximum specific growth rate) indicating the affinity of the substrate to cell growth.¹⁶ The Monod equation and its modified expressions were demonstrated to be robust for modeling different pure cultures under varied conditions.³⁵ Since then, a variety of mechanistic models have been built on Monod expressions.^{10,13,36–38}

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139 Several studies pointed out the inadequacy of the original Monod expression in dealing with substrate inhibition, cell decay, diffusional limitation, etc.^{36,39} Substrate inhibition as a common 140 141 issue arises when the complex composition in wastewater shows different affinity to microbial 142 cells, leading to competition among functional populations and/or inhibition to cell growth. The 143 Haldane-Andrews equation included an inhibition constant in the Monod expression to reflect substrate inhibition.³⁶ However, because the constant is inferred through the generalized 144 145 substrate inhibition model, a normal distribution method with an error range, such a modification 146 does not reveal the actual impacts of substrate inhibition on microbial growth.⁴⁰

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148 **2.2 Modeling of activated sludge processes**

The activated sludge models have been developed for more than 70 years since the early 60s.¹¹ Early mechanistic models were derived based on steady-state applications in which the cell growth rate remained constant.³⁴ The biochemical parameters used to quantify cell growth were total organic carbon (TOC), total oxygen demand (TOD), and COD along with 5-day BOD.⁴¹ They are sufficient for modeling steady-state conditions but have severe limitations in real-time situations where cell growth behaves dynamically under the influence of substrate variation. Oxygen uptake rate and mixed liquor volatile suspended solid were later included to address the limitation.^{36,41} Mathematical techniques such as the feedforward-feedback strategy were also applied to control the flowrate disturbance. These modifications allowed mechanistic models to respond to dynamic situations with real-time biochemical data.⁴¹

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In 1982, the International Association on Water Pollution Research and Control published a preliminary model on activated sludge systems. Later, Dold and Marais incorporated the preliminary model to a final version called the Activated Sludge Model No. 1 (ASM1). ^{12,42} ASM1 is considered a reference model and is generally accepted as a fundamental component for wastewater treatment modeling.⁴³ In particular, the Monod equation in ASM1 was proven reasonably appropriate to describe the microbial growth and substrate utilization behaviors in wastewater.⁴⁴

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168 The microbiological principles of ASM1 include the growth of aerobic and anoxic heterotrophic 169 organisms, the growth of aerobic autotrophic organisms, the decay of heterotrophs and 170 autotrophs, hydrolysis of slowly biodegradable substrate, ammonification, and hydrolysis of organic nitrogen.¹⁰ In addition, the model also describes the following dynamic mass balances 171 172 that have impacts on biomass concentration: 1) readily biodegradable substrates, 2) slowly 173 biodegradable substrates, 3) inert particulate substances, 4) particulate organic nitrogen, 5) soluble organic nitrogen, 6) ammonia, 7) nitrate, and 8) oxygen.¹⁰ These state variables serve as 174 175 explicit indicators of the nutrient removal processes. Among all parameters used in ASM1, the 176 growth and decay rates are of key importance as they control the biomass concentration as a 177 function of the influent substrate concentration. The full description of ASM1 and the

178 comprehensive review of model development can be found elsewhere.^{8,10} The robustness of 179 ASM1 has been demonstrated by numerous studies. Forty years after its first implementation, 180 ASM1 is still playing a central role in the mechanistic modeling of bioprocesses and has been 181 incorporated in commercial software as a core structure for the simulation of full-scale 182 wastewater treatment plants.⁴⁵

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184 Advancing from ASM1, ASM2 incorporates polyphosphate-accumulating organisms (PAO), a 185 functional population enriched during enhanced biological phosphorus removal in activated 186 sludge systems. In addition to common microbial kinetics, the model structure for PAO also 187 includes the storage of glycogen, polyhydroxyalkanoate, and polyphosphate, which are expressed as a function of oxygen availability according to their physiological traits.⁴⁶ In the 188 189 1990s, ASM2d was developed based on ASM2 by including the ability of PAOs to use internal cellular materials for denitrification,¹⁰ thus linking the metabolism of nitrate and phosphorous 190 191 under anoxic conditions. In the absence of oxygen, PAOs can use nitrate as a terminal electron 192 acceptor for phosphorous uptake.^{10,47} In ASM2d, a fraction of the maximum growth rate of PAO 193 is assigned to complete denitrification. The fraction varies depending on PAOs' activity 194 including growth, denitrification, and anoxic phosphorus uptake.⁴⁸ ASM2 and ASM2d are 195 comprehensively reviewed by Henze et al.¹⁰

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ASM3 was modified to improve the prediction of oxygen consumption, sludge production, nitrification, and denitrification. Key modifications include cellular storage of organic substrates and the consumption of dead cells through endogenous respiration (instead of the decay and recycling processes described by ASM1).³⁷ With these modifications, ASM3 is more accurate in

describing the substrate uptake and storage behaviors, but the enhanced prediction may not be relevant for most treatment plants where ASM1 is sufficient for simulation of general performance.⁴⁶ Therefore, ASM3 is needed only when specific metabolic activities are modeled.

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205 There are several commercial software packages available to simulate the activated sludge 206 processes following the development of ASM1/2/3. Olsson and Newell provided a detailed 207 overview of the simulator environments for the bioprocesses in wastewater treatment plants.⁴⁹ 208 Some of the simulators, such as MATLAB-based Simulink, serve a general purpose with high 209 flexibility to complete simulation. Other simulators contain a library of predetermined models 210 for specific bioprocesses, and the process configuration is a unit-based simulation environment. 211 Examples of this type of simulator are AQUASIM, BioWin, EFOR, GPS-X, SIMBA, STOAT, 212 and WEST. The computation package can solve mechanistic models with multiple mathematical equations simultaneously.⁵⁰ BioWin, for example, is a flexible software tool that includes 213 multiple microbial processes presented in ASMs and anaerobic digestor models (ADMs).⁵¹⁻⁵⁵ It 214 215 has been used to model the biological systems in full-scale wastewater treatment plants, 52-55 including activated sludge,⁵⁶ anaerobic digestion,^{57,58} and anammox processes.⁵¹ In some of the 216 217 studies, BioWin provided a good match between the measured and predicted data (difference 218 <10%) for both small-scale batch reactors and full-scale systems,^{56–58} and it can perform 219 optimization of sludge retention time and nitrogen removal under different DO and return activated sludge flows.⁵³ In most of the studies, however, model calibration is required to 220 221 improve the prediction performance. Improper calibration, for example, with nitrogen, 222 phosphorus, and other microbial inhibitory substances has been reported to cause inaccurate prediction of methane production in anaerobic digestion.⁵⁸ Because commercial software still 223

requires both in-depth knowledge of the bioprocess and expertise in modeling, it is not very user-friendly to treatment plant staff.

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227 **2.3 Modeling of anaerobic digesters**

228 Anaerobic digesters have a highly complex microbial community composed of fermentative bacteria, syntrophs, acidogens/acetogens, and methanogens.⁵⁹ The model structure in early 229 230 models was improved with a greater understanding of the microbial kinetics of those functional 231 populations and the associated biochemical reactions. For example, Andrews applied the 232 Haldane function on the Monod expression to modify the substrate uptake function with inhibition under rate-limiting conditions,60 Andrews and Graef later proposed to include the 233 234 effects of pH change and buffering through liquid-gas phase interaction and carbonate 235 equilibrium, leading to more accurate modeling of microbial kinetics.⁶¹ Based on those studies, 236 Hill and Barth further included a function that described the inhibition effects of volatile fatty 237 acids (VFAs) and ammonia on methanogens, as well as charge balance to correct temperature-238 dependent pH.62

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After decades of study of the microbial ecology in anaerobic environments, a task group from the International Water Association consolidated the up-to-date knowledge and formulated Anaerobic Digester Model No. 1 (ADM1) as a common platform model for anaerobic processes.¹³ This model involves 4 typical digestion processes (hydrolysis, acidogenesis, acetogenesis, and methanogenesis) and several physicochemical steps including gas-liquid diffusion, ion association, and dissociation. ADM1 describes in total 29 processes and 32 variables at dynamic state, 24 of which are based on the Monod equation and first-order kinetics.

247 The model also allows modifications for specific applications such as sulfate reduction, 248 phosphorous conversion, and mineral precipitation.¹³ ADM1 has been successfully implemented 249 to simulate effluent characteristics with various types of substrates and operating conditions. The 250 most common function of this comprehensive model is to predict and optimize biogas production. 251 Satpathy et al. applied ADM1 to simulate biogas production from rare substrates such as chicken 252 manure.⁶³ Other applications of ADM1 include the prediction of effluent quality from systems fed with winery wastewater⁶⁴ or treating phenol from olive mill waste.⁶⁵ For example, Ozkan-253 254 Yucel and Gokcay applied ADM1 to a full-scale anaerobic digester under varying organic loading rates to predict total VFAs and COD in the effluent.⁶⁶ Modified ADM1 can also help 255 256 troubleshoot operational problems caused by inhibition effects,⁴⁸ in particular, the accumulation of VFAs.68 257

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259 Despite successful implementation for specific applications, the extensions of sulfate reduction 260 and mineral precipitation can cause a significant computational burden. This is because the 261 precipitation of multiple components (CaHPO₄, struvite, and other unknown compounds) and 262 their release mechanisms involve a large number of processes, in which the fundamental knowledge is not available for the model to perform *ab initio* prediction.⁶⁹ Another critical issue 263 264 is that ADM1 is still not able to fully recapitulate the actual functional populations due to the 265 lack of an in-depth understanding of the microbial community. Specifically, ADM1 is structured 266 without considering the production of different short-chain VFAs, alcohols, and hydrogen. The 267 model is thus not able to meet the growing interest in those value-added products.⁶⁴ Shi et al. 268 attempted to solve this problem by redefining the pseudo-stoichiometric dynamic parameters of VFAs and alcohols corresponding to the hydrogen partial pressure.⁷⁰ The modified model 269

270 successfully predicted the concentrations of acetate, propionate, butyrate, ethanol, and hydrogen 271 with standard errors < 0.04. However, there remains some discrepancy between the predicted and 272 observed hydrogen levels and effluent COD when the system was fed with high-strength streams.⁷⁰ In terms of methane production, the model did not include the methanogenic 273 274 population via direct interspecies electron transfer,⁷¹ which was recently found to be a ubiquitous electron transfer mechanism in many engineered and natural environments.⁷² A reaction-275 276 diffusion-electrochemistry model composed of activation and ohmic losses predicted that 277 methanogenesis could be an order of magnitude faster via direct interspecies electron transfer 278 than via the classic route of interspecies hydrogen transfer.⁷³ Modified ADM1 further predicted 279 over one-third of the CH₄ produced via this novel electron transfer mechanism,⁷⁴ underpinning 280 its critical role in methane production.

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282 **2.4 Modeling of emerging bioprocesses**

283 Discovered in the 90s, anammox has been applied as an alternative technology for biological nitrogen removal.⁷⁵ Anammox can be engineered as a single-step process, which is efficient in 284 285 of with conventional terms energy, space, and cost compared two-step nitrification/denitrification.⁷⁶ In anammox systems, part of the ammonium is oxidized by 286 287 ammonium oxidizing bacteria through partial nitrification to nitrite, which serves as the electron acceptor for anammox bacteria to oxidize the remaining ammonium to nitrogen gas.⁷⁷ Due to the 288 289 unique bioprocesses, a previous study built an anammox model from fundamental processes 290 including diffusion, hydrolysis, and microbial kinetics of anammox bacteria for simulation of 291 long-term nitrogen and COD removal by a granular up-flow anaerobic sludge blanket reactor.¹⁵ 292 Because the model only assumed cell growth under optimum conditions, the predicted and

293 observed nitrogen removal did not correlate well. The anammox process was later modeled using 294 modified ASM1. Through experimental calibration, the model yielded satisfactory prediction of 295 nitrogen removal efficiency of a laboratory-scale bioreactor.⁷⁸ Interestingly, anammox is better simulated when coupled to other bioprocesses, e.g., sulfur-driven denitrification,¹⁴ as the kinetic 296 297 parameters in the model can be more accurately estimated by varying both sulfur (sulfite, sulfur, 298 and sulfate) and nitrogen (ammonium, nitrogen, nitrite, and nitrate).¹⁴ Although the models have 299 demonstrated the potential of anammox systems, the modeling of anammox is still challenging 300 because the system performance is highly dependent on the dynamic interactions between 301 ammonium/nitrite oxidizing bacteria and anammox bacteria.¹⁴ A reliable method is to calibrate 302 the maximum growth rates of ammonium oxidizing bacteria, nitrite oxidizing bacteria, and 303 anammox bacteria using experimental data from the full-scale bioreactors and validate the 304 calibration based on the sensitivity analysis.⁷⁹ In addition, the substrate affinity coefficients 305 should be adjusted based on reported literature to fit the microbial substrate utilization under the 306 possible effects of mass transfer in flocs.⁷⁹

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308 In addition to predicting and optimizing the performance of well-developed bioprocesses such as 309 anammox, mechanistic models can also be implemented for emerging biotechnology such as 310 bioelectrochemical systems to gain an in-depth understanding of their potential and facilitate 311 practical applications. A typical bioelectrochemical system is composed of a cathode and an 312 anode in which the electrochemical reactions are catalyzed by microorganisms. It is challenging 313 to model bioelectrochemical systems primarily because of the close yet uncharacterized 314 connections among microbial kinetics, extracellular electron transfer mechanisms, and electrochemical factors (e.g., internal/external resistance).⁸⁰ A few mathematical models for this 315

novel system have been developed and reviewed elsewhere.^{17,81–83} Noticeably, the growth and substrate utilization rates of the functional groups are still based on the Monod expression and corrected with electron mediator concentration, which in turn is expressed as a function of the substrate utilization rate of electroactive bacterial.^{18,31,84–86} Meanwhile, the Nernst-Michaelis-Menten equation was used to calculate the electron transfer rate in the system⁸⁷ and the Nernst-Planck equation was introduced to represent ion diffusion through the membrane between anode and cathode.^{88,89}

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324 2.5 Inherent drawbacks of mechanistic modeling

325 The model structure in response to intracellular biochemical reactions is oftentimes inadequate, 326 which represents a major limitation of mechanistic models. Structural inadequacy stems from the 327 generalization of microbial growth and substrate utilization kinetics with Monod expressions.⁸ 328 Ideally, the Monod expression for the growth rate of a functional population should consider all 329 degradable and inhibitory compounds and the corresponding factors. The substrates in practical 330 applications (e.g., wastewater) contain various degradable and inhibitory compounds that result 331 in highly complex microbial communities, in which many of the functional populations are uncharacterized.⁹⁰⁻⁹² Therefore, it is not clear which substrates should be experimentally 332 333 measured to reflect their growth and substrate utilization kinetics. The absence of expressions for 334 novel methanogenesis mechanism via direct interspecies electron transfer is an example of 335 inadequate model structure.⁵⁷ Such an issue is more problematic for emerging bioprocesses. The 336 models for bioelectrochemical systems were typically structured with fermentative, electroactive, 337 and methanogenic populations. Although the microbial community in bioelectrochemical systems is much more diverse than those in anaerobic digestors,^{93,94} uncharacterized populations 338

cannot be incorporated into the model structure because their physiological traits related to
 growth and substrate utilization are poorly understood.^{95–97}

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342 Another challenge lies in the unmeasurable kinetic parameters in Monod expressions. Most of 343 the kinetic parameters such as inhibition constant, maximum growth rates, half-saturation 344 constants, and substrate utilization rates can only be derived from biochemical measurements (e.g., substrate concentration).98 From an experimental perspective, when microbial kinetics 345 346 responds to changing influent characteristics in real-time, it is unrealistic to derive the kinetic 347 parameters throughout the variation. As the operating condition changes, biochemical 348 measurements can vary significantly, and the estimation of kinetic parameters becomes 349 conditional.²⁷ The identifiability of unmeasurable kinetic parameters, i.e., the possibility to 350 derive a unique set of values for the parameters from experimental data, is then of great concern. 351 Firstly, uncertainty issues arise because the derivation of the kinetic parameters is obtained through rate-controlling experiments with specific temperatures, pH, BOD, and COD, etc.,³⁴ 352 353 while in wastewater treatment processes, microbial cells are exposed to varied substrates. Those 354 parameters need constant calibration. An uncalibrated model is not likely to yield accurate 355 predictions. As a result, it is not certain to what extent the predictions can be used to explain the 356 observed physical, chemical, and biological mechanisms..⁹⁹ Secondly, to identify the kinetics 357 parameters, the identification of the biochemical and physical parameters responsible for 358 biomass concentrations must be achieved first. Flotats et al. applied the Taylor Series Expansion 359 to four state variables (acetate, propionate, valerate, and methane) of ADM1 to identify the 360 parameters related to valerate consumption and biomass concentration.¹⁰⁰ Such a method has only been reported to solve simple models with a few unidentifiable parameters.^{101,102} For 361

362 complex models such as second-order models, parameter identification was handled with the 363 asymptotic behavior of the maximum likelihood estimator and multiple shooting approach 364 described in Muller et al.¹⁰³ Those previous studies collectively show that the identification and 365 computation processes of kinetic parameters are cumbersome and uncertain in reality.

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367 3. Data-Driven Modeling of Wastewater Treatment Bioprocesses

368 **3.1 Neural networks**

369 Neural networks, first reported in 1943, are arguably the most prevailing data-driven models 370 across various research fields.^{104,105} A neural network is composed of multiple layers of 371 interconnected nodes (neurons), through which the inputs are propagated to the final output 372 layer.¹⁰⁶ Each input to a neuron has a weight factor that determines the interconnection strength 373 to the next neuron. By adjusting the weight factors, a neural network can be properly trained to 374 perform problem-solving. The training algorithm can be divided into three types: supervised, 375 unsupervised, and hybrid training. In supervised training, neural networks are trained with a 376 labeled dataset that provides feedback about the prediction accuracy.¹⁰⁶ Unsupervised training 377 allows networks to be trained with unlabeled data, and the algorithm extracts features and 378 patterns on its own.¹⁰⁶ The hybrid training strategy uses unsupervised training for the hidden 379 neurons and supervised training for the output neurons.¹⁰⁷

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Neural networks were first implemented for continuously stirred bioreactors to predict the fermentation products and pH in the effluent.¹⁰⁸ Boger applied the modeling strategy to full-scale wastewater treatment plants and showed that neural networks could be a solution for simulating expert rules, a set of boundary values that confined the neural network prediction, from historical

operating data.¹⁹ Traditionally, neural networks were trained in a feed-forward fashion, meaning 385 386 that the feed was directed forward-only through layers of training. Yang and Linkenst found that 387 the back-propagation method, which fed outputs from a random layer back to a previous layer, could help the network lower the error rate of prediction.¹⁰⁹ Backpropagation thus became a 388 389 prevailing algorithm of neural network modeling. Several studies used this method to predict 390 effluent COD, biogas production, and NH4+-N removal in different biological wastewater 391 treatment systems including activated sludge processes.²¹ up-flow anaerobic sludge blanket reactors,²⁶ sequencing batch reactors,¹¹⁰ and anaerobic digesters.¹¹¹ 392

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394 Previous studies have combined neural networks with other types of data-driven models to 395 improve the simulation of ill-defined systems. One of the strategies is to use the genetic 396 algorithm to select the initial dataset for downstream neural network training, thereby identifying 397 the optimal training parameters and reducing the computational burden.¹¹² Bagheri et al. have 398 successfully applied the genetic algorithm to optimize the weights and thresholds of neural 399 networks to accurately predict sludge volume index.²² Neural networks can also be coupled to 400 particle swarm optimization, a population-based optimization technique that searches for optimal 401 weights and biases through multiple iterations of particle positions in a given search space. This 402 coupling approach was intended to lower the training time and computational cost for finding an optimal neural network structure.¹¹³ Compared to the genetic algorithm, neural networks coupled 403 404 to particle swarm optimization is more memory-efficient in searching optimal weight parameters but is less practical because it has no crossover and mutation in its operator.¹¹⁴ 405

406

407 Neural networks have been proven feasible for interpolation within the training data range.⁹¹
408 Insufficient training data can result in low prediction accuracy, particularly when prediction is
409 performed with a dataset of fewer than 10 samples.⁹³ Another issue is that the model structure is
410 often determined based on a trial-and-error approach, leading to significant time consumption
411 and computational cost.⁹² The fact that neural networks are black box built based on data fitting
412 rather than the mechanistic understanding of the processes suggests that their outputs cannot be
413 used to explain the mechanisms where the inputs are sourced.²⁵

414

415 **3.2 Random forest**

Random forest is initially developed as a stochastic discrimination approach for classification purposes in the 90s.¹¹⁵ Later, the approach was extended to combine bagging and random selection features to construct a collection of decision-making trees with control variance.¹¹⁶ To use the random forest algorithm, the input data are classified through layers of tree branches consisting of a variety of features and classes, and multiple trees composed of the same number of features and classes are collectively used for prediction.¹¹⁶

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In the wastewater treatment field, the random forest algorithm has been implemented for activated sludge processes, anaerobic digesters, membrane bioreactors, and anammox processes.^{117–120} The main use of random forest-based models includes the prediction of system performance, fault finding, big data handling, model comparisons, and exploration of datasets with applicable reservations and constraints.¹²¹ Although random forest-based models, similar to other data-driven models, are not able to integrate biological principles, these models allow for the identification of key features and conditions that are most influential on the process. The

430 inference can shed light on the underlying biochemical mechanisms. Song et al. implemented 431 this modeling approach with wastewater treatment inputs as multivariate datasets to predict N₂O 432 emission from the aerated zones of activated sludge processes.¹¹⁸ Based on the model inference, 433 they identified inorganic carbon concentration and specific ammonia oxidation activity as two of 434 the dominant factors that determined treatment performance.¹¹⁸ The model was further used to 435 identify the different mechanisms of N₂O generation in oxic and anoxic environments and 436 demonstrated the key role of N₂O in those zones in promoting niche-specific biochemical reactions.¹²² 437

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439 Random forest can be combined with other algorithms such as principal component analysis 440 (PCA) and neural networks to improve the prediction of effluent quality. Preprocessing of data 441 using PCA could enhance the robustness of random forest-based models, leading to a more 442 accurate prediction of membrane flux in membrane bioreactors as compared with neural networks.¹¹⁷ When coupled to neural networks, random forest-based models could be trained to 443 predict the settleability in the biological reactor chamber,¹¹⁹ as well as to evaluate the effects of 444 key operating factors on treatment performance.¹²³ The results suggested that such a combined 445 446 strategy could help achieve real-time monitoring and optimize operating conditions.¹²⁴

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448 **3.3 Fuzzy logic**

Fuzzy logic is an if-then algorithm that can be used to develop a set of flexible rules for diagnosis and control. A fuzzy logic-based system has four robust components: a fuzzifier, a fuzzy rule-base, an inference engine, and a defuzzifier.¹²⁵ The fuzzifier is responsible for converting crisp inputs into fuzzy sets, which are mapped by the inference engine to produce

another fuzzy set as the output. The fuzzy rule base is a collection of rules that guide the fuzzy
engine to produce the outputs. Finally, the defuzzifier transfers the output fuzzy sets back to
crisp values.¹²⁵

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Fuzzy logic can be applied to numerous scenarios in wastewater treatment bioprocesses such as diagnosis and control of sequencing batch reactor processes,¹²⁶ simulations and prediction of phosphorus removal,¹²⁷ as well as design, evaluation, and decision optimization of activated sludge processes.¹²⁸ Robles et al. developed a fuzzy logic-based controller to optimize biogas production and VFA concentration at varied influent flow rates.¹²⁹ The designed controller was able to help prevent acidification in a closed-loop setting.¹²⁹

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464 A combination of fuzzy logic and neural networks could bring together the learning powers of both algorithms, enabling fault tolerance during the modeling of complex systems.^{107,108} For 465 466 example, the adaptive neuro-fuzzy inference system that pairs neural networks with fuzzy logic 467 allows modelers to insert a priori knowledge into the neural network structure as rules for 468 training. The combined modeling strategy was used to predict suspended solids, COD, pH and 469 DO levels in activated sludge systems.^{108–110} Using the adaptive neuro-fuzzy inference system, 470 Essienubong et al. obtained a strong correlation between the experimental and predicted biogas 471 production with temperature, pH, substrate/water ratio, and hydraulic retention time as the 472 inputs.¹³⁴ The work by Hosseinzadeh et al. also demonstrated a higher sensitivity of the adaptive 473 neuro-fuzzy inference system when predicting water flux in an osmotic membrane bioreactor.¹¹² 474 In addition to neural networks, fuzzy logic was coupled to genetic algorithms and particle swarm

475 optimization, which outperformed the adaptive neuro-fuzzy inference system during the
476 prediction of BOD, ammonium, and suspended solids in specific bioprocesses.¹³⁶

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478 Although fuzzy logic models are powerful tools for predicting system outputs using observable 479 environmental data and human-like logic, these data-driven models cannot capture the behaviors 480 of the complex kinetic reactions in engineered biological bioprocesses, which presents the major 481 criticism among other concerns of implementation.¹³⁷ From an engineering point of view, 482 detailed descriptions of the chemical, physical, and microbiological principles in bioprocesses 483 and computing-based predictive methodology are equally important.¹³⁸ Unfortunately, fuzzy 484 logic systems, like most of the data-driven models, are incapable of providing mechanistic 485 insight into troubleshooting and system optimization due to their black-box nature.

486

487 4. Hybrid Models That Address the Limitations of Conventional Modeling Strategies

The concept of combining mechanistic and data-driven sub-models for hybrid modeling was first proposed in the early 90s and immediately implemented for a fermentation bioprocess to reduce the dependence on microbial kinetics.²⁹ Hybrid modeling strategies were further examined with activated sludge processes and anaerobic digesters through parallel or serial combinations of the mechanistic and data-driven components.^{139,140}

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In a parallel structure, the outputs from the mechanistic and data-driven components are combined primarily through pure superposition (i.e., summation of the outputs).^{106,140–142} Weighing functions can be introduced to adjust the weight of the outputs, thereby improving the overall prediction accuracy.¹⁴³ It should be noted that the prediction performance of a parallel-

structured hybrid model is highly dependent on the robustness of the individual sub-models.¹⁴¹ In cases when a biological system is too dynamics/nonlinear and some of the biochemical data are too expensive to collect in real-time (e.g., H_2 in anaerobic digesters), neither the mechanistic nor the data-driven components could predict accurately, leading to poor prediction performance of the hybrid model. It is therefore argued that the sub-models in a parallel structure are not well integrated due to the lack of interactions (e.g., cross-feeding of the outputs as done in a serial structure).

505

506 In a serial structure, the data-driven component acts as a parameter simulator and estimates 507 kinetic parameters for the mechanistic component to complete simulation.^{29,140} Serial coupling of 508 the sub-models leverages the prediction power of the data-driven component and is sometimes 509 capable of extrapolating system outputs outside of the observation range. A mixture of both 510 structures has been implemented for chemical systems but not bioprocesses.¹⁰⁶ As the selection of the structures depends on the availability of the mechanistic information,¹⁴² we argue that with 511 more functional populations being characterized in wastewater biological systems,¹⁴⁴ a serial or 512 513 mixed structure may better reflect the underlying biological mechanisms whilst accurately 514 predicting the system performance.

515

516 One of the advantages of hybrid modeling is that unmeasurable mechanistic parameters, in 517 particular microbial growth and substrate utilization rates, can be determined by the data-driven 518 component (Figure 2).^{29,145} This was done in the first study of hybrid modeling of engineered 519 bioprocesses, in which a neural network was trained to estimate the specific growth rate of the 520 overall microbial community, and the outputs were used to establish a biomass balance, resulting

in more accurate predictions than those from a standalone neural network.²⁹ Compared to 521 522 conventional mechanistic models that perform one-time estimation of the kinetic parameters, the 523 data-driven component in hybrid models allows the microbial kinetics to be updated in a timely 524 manner based on the collected data, thereby making the hybrid model more robust under varied 525 conditions.²⁹ Another benefit is that the data-driven component can capture more dynamic data 526 to compensate for the prediction of the mechanistic component. For instance, a neural network 527 was used to estimate the operational data at time t+1 based on the influent and operational data at 528 time t, and the outputs were subsequently used to correct the mechanistic predictions at time t+1.¹⁴⁰ In this way, the hybrid model could capture the disturbance caused by shock loadings of 529 530 toxic compounds and deliver more accurate prediction of the effluent composition.¹⁴⁰

531

532 Similar to standalone mechanistic models, hybrid models still contain a Monod expression-based 533 model structure. As previously discussed, Monod expressions approximate the physiological 534 traits of functional populations (i.e., microbial growth and substrate utilization) by assuming a 535 homogeneous culture.^{16,35} Real wastewater and sewage sludge are highly heterogeneous with 536 various degradable and inhibitory compounds that result in diverse microbial populations.^{43,55} 537 For well-characterized populations with known functions, unmeasurable kinetic parameters are 538 predominantly derived from biochemical measurements.^{14,16,34,35,40} Although the data-driven 539 component can help correct the estimates and improve the prediction performance, the estimated 540 kinetic parameters do not necessarily reflect the actual activity of those populations. This is even 541 more problematic for uncharacterized functional populations, whose microbial activity 542 information can be inferred with the data-driven component but does not help interpret the final 543 prediction because those populations are frequently overlooked in the mechanistic component.

544 Therefore, conventional hybrid models constructed with biochemical and physical data still 545 suffer from interpretation issues. A potential solution is to incorporate microbial population and 546 functional dynamics directly into the mechanistic and data-driven components.

547

548 5. Genomics-Enabled Modeling Strategies for Accurate and Interpretable Prediction

549 5.1 Genomics-enabled data-driven modeling

550 The rapid development of high throughput sequencing techniques and bioinformatics has led to a 551 greater understanding of the microbiomes in wastewater treatment bioprocesses.^{146–149} 552 Recovering the 16s rRNA sequences allows us to unfold the taxonomy and phylogeny of the 553 core populations in activated sludge, anaerobic digester, and many other systems.¹⁴⁶ Meanwhile, 554 metagenomic and meta-transcriptomic data have greatly advanced our knowledge about the 555 genetic potential and functional dynamics of uncharacterized populations.^{147,148} The findings 556 gained with those powerful tools have validated the mechanistic structure of existing models formulated with known functional populations.¹⁴⁹ The million-dollar question now is how to 557 558 incorporate those genomic data into modeling in a more direct manner (Figure 3).

559

Two pioneering studies integrated 16S rRNA amplicon sequencing data with machine learning algorithms (neural networks and Bayesian networks) to reconstruct the microbial communities in natural ecosystems.^{150,151} Following similar strategies, several machine learning-based models were trained with microbial taxon abundance and generated semi-interpretable predictions of system performance and stability.^{152,153} The inferences suggested that for specific systems, classifying taxonomic data at the family level could enhance the prediction accuracy, whilst abundances of specific genera could act as better predictors, highlighting the potential to improve 567 the prediction interpretability with proper data preparation. Before training Bayesian networks, 568 Yuan et al. prepared the genomic data collected from a bioelectrochemical system by selecting 569 dominant taxa at the phylum, genus (Figure 4A), and operational taxonomic unit levels.¹⁵⁴ The 570 genomics-enabled data-driven modeling approach was rigorously cross-validated using three 571 validation strategies.¹⁵⁴ Firstly, the difference between the predicted and observed relative 572 abundances of the selected populations remained within an acceptable range as indicated by a 573 relative root-mean-square error (RMSE) of ~20%. Secondly, the microbial communities 574 reconstructed with the predicted abundances of the selected populations shared high Bray-Curtis 575 similarity with the observed communities at all taxonomic levels. Finally, the predicted system 576 outputs agreed well with the experimental data. For example, current production as the most 577 important system performance for bioelectrochemical systems was predicted with high accuracy 578 at the order level ($R^2 = 0.97$ for prediction vs. observation, Figure 4B). After validation, the 579 model was used to predict current production as a function of operating conditions (e.g., 580 substrate salinity, Figure 4C) and provided practical insights into system optimization.

581

582 Functional genomic data as the training input can improve prediction interpretability. A previous 583 study trained ANNs with gene expression levels to infer metabolic behaviors, resulting in a 584 plausible explanation of microbes' stress adaptation behaviors under environmental perturbations.¹⁵⁵ Using a similar but more dynamic modeling strategy, Yuan et al. trained 585 586 Bayesian networks with meta-transcriptomic data to explain the contribution of interspecies hydrogen transfer and direct interspecies electron transfer to methanogenesis.³⁰ It is highly 587 588 desired to develop a predictive understanding of the involvement of the two mechanisms due to the lack of measurement techniques.^{73,74} To prepare data for model training, the genes for alcohol 589

590 metabolism, hydrogen metabolism, extracellular electron transfer, and methanogenesis were 591 extracted from the metagenome-assembled genomes of the dominant microbes. A Bayesian 592 network trained with those genes is composed of two components (Figure 5A): upstream gene-593 gene interactions that predict the expression level of the relevant genes in methanogens, and a 594 downstream sub-network that links the genes encoding methanogenesis to methane. A complete 595 network could accurately predict methane production ($R^2 = 0.96$ for prediction vs. observation, 596 Figure 5B). To statistically infer the contribution of the electron transfer mechanisms, relevant 597 genes were manually silenced. When the simulation was performed without the genes for 598 hydrogen metabolism, the prediction accuracy was significantly compromised as evidenced by 599 the noticeable difference between the predicted and observed methane production and high 600 RMSE (AIHT in Figure 5B). In contrast, the prediction remained accurate with *in silico* knockout 601 of the genes for direct interspecies electron transfer ($\Delta DIET$ in Figure 5B). The results thus 602 implied a more critical role of hydrogen-mediated electron transfer in methane production.

603

604 5.2 Genomics-enabled hybrid modeling

Thus far, genomic data have only been used to train data-driven models for semi-interpretable prediction. There is a growing interest to incorporate it into hybrid modeling to predict the underlying mechanisms for system design and optimization. One potential strategy is to infer unidentifiable kinetic parameters from microbial population and gene dynamics. The mechanistic component can be formulated following conventional modeling procedures to estimate kinetic parameters, and the estimates can be combined with microbial taxon abundance and operating conditions to train the data-driven component. The resulting hybrid model is therefore a kinetics 612 simulator that statistically infers kinetic parameters, which can then be fed back to the613 mechanistic component for prediction of system performance.

614

615 The concept has been proven valid by a recent study,³¹ in which a genomics-enabled hybrid 616 model was implemented for bioelectrochemical systems based on 77 samples collected from 13 617 publications. The mechanistic component of the hybrid model was built to estimate the 618 maximum growth and substrate utilization rates of three functional populations: fermentative, 619 electroactive, and methanogenic microbes, which were subsequently combined with the relative 620 abundances of 38 core taxa at the genus level to train a hybrid Bayesian network (Figure 6A). 621 When examined with six new samples that were not included in network training, the hybrid 622 model achieved the most accurate prediction of current production (Hybrid + Mechanistic in 623 Figure 6B) compared with standalone data-driven models. The enhanced prediction performance 624 of the hybrid model likely results from the close connection between population dynamics and 625 microbial kinetics.

626

627 An alternative strategy to incorporate genomic data into hybrid modeling is to replace Monod 628 expression-based model structures in conventional mechanistic models by simulating microbial 629 population dynamics. This can be achieved with an iterative strategy, in which the data-driven 630 component trained with processed genomic data infers instantaneous biochemical and microbial 631 intermediates, and the intermediates are fed into the mechanistic component to predict steady-632 state biochemical outputs. This novel strategy mimics microbial community assembly driven by 633 environmental perturbations in engineered systems:^{156–158} operating conditions and biochemical 634 inputs impose selective pressure and together shape the microbial community structure, and the 635 enriched functional populations produce biochemical intermediates that are rapidly mixed with 636 the inputs to form a new steady-state, causing the community structure to further shift until the 637 biochemical outputs reach equilibrium. The proposed strategy thus frees models from microbial 638 kinetics-based structures, while the abiotic and microbial processes and their interplay are 639 revealed in each iteration. Successful modeling with this strategy relies on the ability of hybrid 640 models to act as a community simulator to predict microbial taxon abundance and reconstruct 641 microbial communities in silico. For example, Bayesian networks could infer the relative 642 abundances of dominant taxa, resulting in Bray-Curtis similarity of over 90% between the simulated and observed microbial communities at the phylum level.¹⁵⁶ However, the similarity 643 644 dropped to 83% at the order level and 69% at the OTU level, likely because of the presence of 645 functionally redundant taxa in the small data pool. The potential of the proposed strategy 646 warrants investigation with big data collected from global databases.

647

648 6. Conclusions

649 This review focuses on three major types of models: mechanistic, data-driven, and hybrid models. 650 Mechanistic models can provide fundamental insights but need laborious calibration because the 651 Monod-based model structure is inadequate to reflect the biological principles whilst the 652 microbial kinetic parameters are largely unidentifiable. As a result, a mechanistic model built for 653 a specific system frequently falls short when applied to other bioprocesses. Data-driven models 654 can provide predictive insights but yield uninterpretable predictions due to their black-box nature. 655 Hybrid models are believed to overcome the issues of structural inadequacy, parametric 656 unidentifiability, and uninterpretable prediction of the standalone models. Recent 657 biotechnological development such as high throughput sequencing data and omics-based analysis

658 can further enable the incorporation of microbial population and functional dynamics into the 659 model to directly reflect the biological principles. Genomics-enabled hybrid modeling strategies 660 require the mechanistic and data-driven components to be integrated interactively. Two strategies 661 are proposed: kinetics simulator and community simulator, and their applicability warrant further 662 studies. Although hybrid models can potentially overcome the drawbacks of standalone models, 663 the main rationale of modeling selection and design is largely dependent on its intended use. 664 Additionally, the availability of omics-based data and computational cost require more effort in 665 preparation, collection, process, and analysis, which demands technical labor, time, and financial 666 investment. All these factors need to be taken into consideration when modelers design and 667 modify existing models.

668

669 **Conflicts of Interest**

- 670 There are no conflicts of interest to declare.
- 671

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675 **Reference**

- USEPA, Emerging Technologies for Wastewater Treatment and In-Plant Wet Weather
 Management, U.S. Environmental Protection Agency, Fairfax, 2013.
- J. S. Seelam, S. A. Maesara, G. Mohanakrishna, S. A. Patil, A. Ter Heijne and D. Pant, in *Waste Biorefinery: Potential and Perspectives*, Elsevier, 2018, pp. 535–570.
- *Module 16: the Activated sludge process part II*, Pennsylvania Department of Environmental
 Protection, Harrisburg, 2014.
- 682 5 Biological Treatment: suspended growth processes study guide, Wisconsin Department of
 683 Natural Resouces, Madison, WI, 2015.
- 6 R. Snyder and D. Wyant, *Activated sludge process control: training manual for wastewater treatment plant operators*, State of Michigan Department of Environmental Quality.
- A. Xu, Y.-H. Wu, Z. Chen, G. Wu, Q. Wu, F. Ling, W. E. Huang and H.-Y. Hu, Towards the
 new era of wastewater treatment of China: Development history, current status, and future
 directions, *Water Cycle*, 2020, 1, 80–87.
- 689 8 U. Jeppsson, Modelling Aspects of Wastewater Treatment Processes, 1996.
- K. V. Gernaey, M. C. M. Van Loosdrecht, M. Henze, M. Lind and S. B. Jørgensen,
 Activated sludge wastewater treatment plant modelling and simulation: State of the art, 2004,
 763-783.
- M. Henze, W. Gujer, T. Mino, T. Matsuo, M. Wentzel, G. Marais and M. Loosdrecht,
 Activated sludge model No.2D, ASM2D, *Water Sci. Technol.*, 1999, **39**, 165–182.
- M. C. M. Van Loosdrecht, C. M. Lopez-Vazquez, S. C. F. Meijer, C. M. Hooijmans and D.
 Brdjanovic, Twenty-five years of ASM1: Past, present and future of wastewater treatment
 modelling, *J. Hydroinformatics*, 2015, 17, 697–718.
- M. Henze, W. Gujer, T. Mino and M. Van Loosedrecht, Activated sludge models ASM1,
 ASM2, ASM2d and ASM3, *Water Intell. Online*, 2006, 5, 47–77.
- D. J. Batstone, J. Keller, I. Angelidaki, S. V Kalyuzhnyi, S. G. Pavlostathis, A. Rozzi, W. T.
 M. Sanders, H. Siegrist and V. A. Vavilin, *The IWA Anaerobic Digestion Model No 1*(ADM1), 2002.
- 14 Y.-F. Deng, W.-T. Tang, H. Huang, J. Qian, D. Wu and G.-H. Chen, Development of a
 kinetic model to evaluate thiosulfate-driven denitrification and anammox (TDDA) process,
 Water Res., 2021, **198**, 117155.
- B. J. Ni, Y. P. Chen, S. Y. Liu, F. Fang, W. M. Xie and H. Q. Yu, Modeling a granule-based
 anaerobic ammonium oxidizing (ANAMMOX) process, *Biotechnol. Bioeng.*, 2009, 103,
 490–499.
- 16 J. Monod, The Growth of Bacterial Cultures, Annu. Rev. Microbiol., 1949, 3, 371–394.
- S. Luo, H. Sun, Q. Ping, R. Jin and Z. He, A Review of Modeling Bioelectrochemical
 Systems: Engineering and Statistical Aspects, *Energies*, 2016, 9, 1–27.
- Y. Liu, M. Qin, S. Luo, Z. He and R. Qiao, Understanding Ammonium Transport in
 Bioelectrochemical Systems towards its Recovery, *Sci. Rep.*, 2016, 6, 1–10.
- 714 19 Z. Boger, Application of neural networks to water and wastewater treatment plant operation,
 715 *ISA Trans.*, 1992, **31**, 25–33.
- N. Bhat and T. J. McAvoy, Use of neural nets for dynamic modeling and control of chemical
 process systems, *Comput. Chem. Eng.*, 1990, 14, 573–582.

- 718 21 H. Moral, A. Aksoy and C. F. Gokcay, Modeling of the activated sludge process by using
 719 artificial neural networks with automated architecture screening, *Comput. Chem. Eng.*, 2008,
 720 32, 2471–2478.
- M. Bagheri, S. A. Mirbagheri, Z. Bagheri and A. M. Kamarkhani, Modeling and
 optimization of activated sludge bulking for a real wastewater treatment plant using hybrid
 artificial neural networks-genetic algorithm approach, *Process Saf. Environ. Prot.*, 2015, 95,
 12–25.
- D. De Clercq, D. Jalota, R. Shang, K. Ni, Z. Zhang, A. Khan, Z. Wen, L. Caicedo and K.
 Yuan, Machine learning powered software for accurate prediction of biogas production: A
 case study on industrial-scale Chinese production data, *J. Clean. Prod.*, 2019, 218, 390–399.
- 24 L. Wang, F. Long, W. Liao and H. Liu, Prediction of anaerobic digestion performance and
 identification of critical operational parameters using machine learning algorithms,
 Bioresour. Technol., 2020, 298, 1–9.
- C. Rudin, Stop explaining black box machine learning models for high stakes decisions and use interpretable models instead, *Nat. Mach. Intell.*, 2019, 1, 206–215.
- C. Mendes, R. da Silva Magalhes, K. Esquerre and L. M. Queiroz, Artificial neural network
 modeling for predicting organic matter in a full-scale up-flow anaerobic sludge blanket
 (UASB) reactor, *Environ. Model. Assess.*, 2015, 20, 625–635.
- 736 27 M. J. Wade, Not just numbers: Mathematical modelling and its contribution to anaerobic
 737 digestion processes, *Processes*, 2020, 8, 1-31, DOI:10.3390/PR8080888.
- 738 28 T. Bohlin, *Practical grey-box process identification: theory and applications*, Springer,
 739 London, 2006.
- D. C. Psichogios and L. H. Ungar, A hybrid neural network-first principles approach to
 process modeling, *AIChE J.*, 1992, **38**, 1499–1511.
- 30 H. Yuan, X. Wang, T.-Y. Lin, J. Kim and W.-T. Liu, Disentangling Syntrophic Electron
 Transfer Mechanisms in Methanogenesis Through Electrochemical Stimulation, Omics, and
 Machine Learning, 2021, 11, 1-22, DOI:10.21203/rs.3.rs-288821/v1.
- 745 31 Z. Cheng, S. Yao and H. Yuan, Linking Population Dynamics to Microbial Kinetics for
 746 Hybrid Modeling of Engineered Bioprocesses, *bioRxiv*, 2021, 2021.04.15.440059.
- 32 B. E. Rittmann and P. L. McCarty, *Environmental biotechnology: Principles and applications*, McGraw-Hill Education, 2001.
- J. E. McKee, G. M. Fair and L. S. Kraus, Load Distribution in the Activated Sludge Process,
 Sew. Works J., 1942, 14, 121–146.
- 34 K. Kovárová-Kovar and T. Egli, Growth kinetics of suspended microbial cells: from singlesubstrate-controlled growth to mixed-substrate kinetics, *Microbiol. Mol. Biol. Rev.*, 1998, 62,
 646–666.
- J. Heijnen and B. Romein, Derivation of Kinetic Equations for Growth on Single Substrates
 Based on General Properties of a Simple Metabolic Network, 1995, 11, 712–716.
- J. F. Andrews, A mathematical model for the continuous culture of microorganisms utilizing
 inhibitory substrates, *Biotechnol. Bioeng.*, 1968, 10, 707–723.
- 37 W. Gujer, M. Henze, T. Mino and M. Van Loosdrecht, Activated sludge model no. 3, *Water Sci. Technol.*, 1999, **39**, 183–193.
- 760 38 J. Alex, L. Benedetti, J. Copp, K. V. Gernaey, U. Jeppsson, I. Nopens, M. N. Pons, L. Rieger,
- C. Rosen, J. P. Steyer, P. Vanrolleghem and S. Winkler, Benchmark Simulation Model no. 1(BSM1), 2008, 62.

39 M. Mohammadi, A. R. Mohamed, G. D. Najafpour, H. Younesi and M. H. Uzir, Kinetic
Studies on Fermentative Production of Biofuel from Synthesis Gas Using Clostridium
ljungdahlii, *Sci. World J.*, 2014, 2014, 1–9.

- 40 G. C. Okpokwasili and C. O. Nweke, Microbial growth and substrate utilization kinetics, *Afr. Journal Biotechnol.*, 2005, 5, 305–317.
- 41 M. K. Stenstrom, W. Kido, R. F. Shanks and M. Mulkerin, Estimating Oxygen Transfer
 Capacity of a Full-Scale Pure Oxygen Activated Sludge Plant, *J. Water Pollut. Control Fed.*,
 1989, **61**, 208–220.
- 42 P. L. Dold, G. A. Ekama, G. Marais and G. R. Marais Van, A general model for the activated sludge process, *Prog. Water Technol.*, 1980, 12, 47–77.
- 43 K. V. Gernaey, M. C. M. Van Loosdrecht, M. Henze, M. Lind and S. B. Jørgensen,
 Activated sludge wastewater treatment plant modelling and simulation: State of the art, *Environ. Model. Softw.*, 2004, **19**, 763–783.
- 44 J. H. Sherrard, R. O. Mines, J. E. Alleman and M. S. Kennedy, Activated Sludge, *J. Water Pollut. Control Fed.*, 1983, 55, 615–622.
- P. J. Roeleveld and M. C. M. Van Loosdrecht, Experience with guidelines for wastewater
 characterisation in The Netherlands | Water Science & Technology, *Water Sci. Technol.*,
 2002, 45, 77–87.
- 46 M. C. M. Van Loosdrecht, C. M. Lopez-Vazquez, S. C. F. Meijer, C. M. Hooijmans and D.
 Brdjanovic, Twenty-five years of ASM1: Past, present and future of wastewater treatment
 modelling, *J. Hydroinformatics*, 2015, 17, 697–718.
- 784 47 N. Boontian, Cranfield University, 2012.
- 48 Z. R. Hu, M. C. Wentzel and G. A. Ekama, Modelling biological nutrient removal activated
 sludge systems A review, *Water Res.*, 2003, 37, 3430–3444.
- 49 G. Olsson and B. Newell, Wastewater Treatment Systems: Modelling, Diagnosis and Control.
 2005, 4.
- A. Serdarevic and A. Dzubur, Wastewater process modeling, *Coupled Syst. Mech.*, 2016, 5, 21–39.
- 51 A. G. Dorofeev, Yu. A. Nikolaev, M. N. Kozlov, M. V. Kevbrina, A. M. Agarev, A. Yu.
 Kallistova and N. V. Pimenov, Modeling of anammox process with the biowin software suite, *Appl. Biochem. Microbiol.*, 2017, 53, 88–95.
- 52 O. Oleyiblo, J. Cao, Q. Feng, G. Wang, X. Zhaoxia and F. Fang, Evaluation and
 improvement of wastewater treatment plant performance using BioWin, *Chin. J. Oceanol. Limnol.*, 2014, 33, 468–476.
- A. Elawwad, M. Matta, M. Abo-Zaid and H. Abdel-Halim, Plant-wide modeling and
 optimization of a large-scale WWTP using BioWin's ASDM model, *J. Water Process Eng.*,
 2019, **31**, 100819.
- 800 54 K. Rathore, University of South Floida, 2018.
- 55 C. Moragaspitiya, J. Rajapakse, W. Senadeera and I. Ali, Simulation of Dynamic Behaviour
 of a Biological Wastewater Treatment Plant in South East Queensland, Australia using BioWin Software, *Eng. J.*, 2017, 21, 1–22.
- 804 56 R. Vitanza, I. Colussi, A. Cortesi and V. Gallo, Implementing a respirometry-based model
 805 into BioWin software to simulate wastewater treatment plant operations, *J. Water Process* 806 *Eng.*, 2016, **9**, 267–275.
- 57 I. Hamawand and C. Baillie, Anaerobic Digestion and Biogas Potential: Simulation of Lab
 and Industrial-Scale Processes, *Energies*, 2015, 8, 454–474.

- 809 58 J. L. Callahan, Text, Colorado School of Mines, 2018.
- 59 K. Venkiteshwaran, B. Bocher, J. Maki and D. Zitomer, Relating Anaerobic Digestion
 Microbial Community and Process Function : Supplementary Issue: Water Microbiology,
 Microbiol. Insights, 2015, 8s2, MBI.S33593.
- 60 J. F. Andrews, A mathematical model for the continuous culture of microorganisms utilizing
 inhibitory substrates, *Biotechnol. Bioeng.*, 1968, 10, 707–723.
- 815 61 J. F. Andrews and S. P. Graef, in *Anaerobic Biological Treatment Processes*, American
 816 Chemical Society, 1971, pp. 126–162.
- 62 D. T. Hill and C. L. Barth, A Dynamic Model for Simulation of Animal Waste Digestion on
 JSTOR, *Water Pollut. Control Fed.*, 1977, 49, 2129–2143.
- 819 63 P. Satpathy, S. Steinigeweg, F. Uhienhut and E. Siefert, Application of Anaerobic Digestion
 820 Model 1 (ADM1) for Prediction of Biogas Production, *Int. J. Sci. Eng. Res.*, 2013, 4, 86–89.
- 64 D. J. Batstone, M. Torrijos, C. Ruiz and J. E. Schmidt, Use of an anaerobic sequencing batch
 reactor for parameter estimation in modelling of anaerobic digestion, *Water Sci. Technol.*,
 2004, **50**, 295–303.
- B. Fezzani and R. Ben Cheikh, Extension of the anaerobic digestion model No. 1 (ADM1) to
 include phenolic compounds biodegradation processes for the simulation of anaerobic codigestion of olive mill wastes at thermophilic temperature, *J. Hazard. Mater.*, 2009, 162,
 1563–1570.
- 66 U. G. Ozkan-Yucel and C. F. Gökçay, Application of ADM1 model to a full-scale anaerobic
 digester under dynamic organic loading conditions, *Environ. Technol.*, 2010, **31**, 633–640.
- A. Ramachandran, R. Rustum and A. J. Adeloye, Review of anaerobic digestion modeling
 and optimization using nature-inspired techniques, *Processes*, 2019, 7, 1–12.
- 68 J. Palatsi, J. Illa, F. X. Prenafeta-Boldú, M. Laureni, B. Fernandez, I. Angelidaki and X.
 Flotats, Long-chain fatty acids inhibition and adaptation process in anaerobic thermophilic
 digestion: Batch tests, microbial community structure and mathematical modelling, *Bioresour. Technol.*, 2010, **101**, 2243–2251.
- Batstone, M. Torrijos, C. Ruiz and J. E. Schmidt, Use of an anaerobic sequencing batch
 reactor for parameter estimation in modelling of anaerobic digestion, *Water Sci. Technol.*,
 2004, **50**, 295–303.
- 839 70 E. Shi, J. Li and M. Zhang, Application of IWA Anaerobic Digestion Model No. 1 to
 840 simulate butyric acid, propionic acid, mixed acid, and ethanol type fermentative systems
 841 using a variable acidogenic stoichiometric approach, *Water Res.*, 2019, 161, 242–250.
- A. E. Rotaru, P. M. Shrestha, F. Liu, M. Shrestha, D. Shrestha, M. Embree, K. Zengler, C.
 Wardman, K. P. Nevin and D. R. Lovley, A new model for electron flow during anaerobic
 digestion: Direct interspecies electron transfer to Methanosaeta for the reduction of carbon
 dioxide to methane, *Energy Environ. Sci.*, 2014, 7, 408–415.
- 72 D. R. Lovley, Syntrophy Goes Electric: Direct Interspecies Electron Transfer, *Annu. Rev. Microbiol.*, 2017, **71**, 643–664.
- 73 T. Storck, B. Virdis and D. J. Batstone, Modelling extracellular limitations for mediated
 versus direct interspecies electron transfer, *ISME J.*, 2016, 10, 621–631.
- 74 Y. Liu, Y. Zhang, Z. Zhao, H. H. Ngo, W. Guo, J. Zhou, L. Peng and B. J. Ni, A modeling
 approach to direct interspecies electron transfer process in anaerobic transformation of
 ethanol to methane, *Environ. Sci. Pollut. Res.*, 2017, 24, 855–863.
- 75 A. Terada, S. Zhou and M. Hosomi, Presence and detection of anaerobic ammonium oxidizing (anammox) bacteria and appraisal of anammox process for high-strength

855		nitrogenous wastewater treatment: a review, <i>Clean Technol. Environ. Policy</i> , 2011, 13 , 759–
856 857 858	76	X. Chang, D. Li, Y. Liang, Z. Yang, S. Cui, T. Liu, H. Zeng and J. Zhang, Performance of a completely autotrophic nitrogen removal over nitrite process for treating wastewater with
859		different substrates at ambient temperature <i>J Environ Sci</i> 2013 25 688–697
860	77	K. A. Third, A. O. Sliekers, J. G. Kuenen and M. S. M. Jetten, The CANON system
861		(completely autotrophic nitrogen-removal over nitrite) under ammonium limitation:
862		Interaction and competition between three groups of bacteria, Syst. Appl. Microbiol., 2001,
863		24 , 588–596.
864	78	G. Cema, A. Sochacki, J. Kubiatowicz, P. Gutwiński and J. Surmacz-Górska, Start-up,
865		modelling and simulation of the anammox process in a membrane bioreactor, Chem. Process
866		Eng Inzynieria Chem. Proces., 2012, 33 , 639–650.
867	79	BJ. Ni, A. Joss and Z. Yuan, Modeling nitrogen removal with partial nitritation and
868		anammox in one floc-based sequencing batch reactor, 2014, 67 , 321–329.
869	80	D. Pant, A. Singh, G. Van Bogaert, S. Irving Olsen, P. Singh Nigam, L. Diels and K.
870		Vanbroekhoven, <i>RSC Adv.</i> , 2012, 2 , 1248–1263.
871	81	C. Xia, D. Zhang, W. Pedrycz, Y. Zhu and Y. Guo, J. Power Sources, 2018, 373, 119–131.
8/2	82	V. B. Oliveira, M. Simoes, L. F. Melo and A. M. F. R. Pinto, Overview on the developments
8/3	07	of microbial fuel cells, <i>Biochem. Eng. J.</i> , 2013, 7 3 , 53–64.
8/4	83	V. M. Ortiz-Martineza, M. J. Salar-Garciaa, A. P. de los Rioso, F. J. Hernandez-Fernandeza,
8/3 876		J. A. Egeac and L. J. Lozanoa, Developments in microbial fuel cell modeling, Chem. Eng. J., 2015 271 50 60
870 877	81	2013, 271, 30-00. B D Pinto B Srinivasan A Escana and B Tartakovsky Multi population model of a
878	04	microbial electrolysis cell <i>Environ Sci Tachnol</i> 2011 45 5039–5046
879	85	R P Pinto B Srinivasan M F Manuel and B Tartakovsky A two-population bio-
880	05	electrochemical model of a microbial fuel cell <i>Bioresour Technol</i> 2010 101 5256–5265
881	86	O Ping C Zhang X Chen B Zhang Z Huang and Z He Mathematical Model of
882	00	Dynamic Behavior of Microbial Desalination Cells for Simultaneous Wastewater Treatment
883		and Water Desalination, Environ. Sci. Technol., 2014, 48, 13010–13019.
884	87	A. K. Marcus, C. I. Torres and B. E. Rittmann, Conduction-based modeling of the biofilm
885		anode of a microbial fuel cell, <i>Biotechnol. Bioeng.</i> , 2007, 98, 1171–1182.
886	88	F. Harnisch, R. Warmbier, R. Schneider and U. Schröder, Modeling the ion transfer and
887		polarization of ion exchange membranes in bioelectrochemical systems, Bioelectrochemistry,
888		2009, 75 , 136–141.
889	89	J. Kim, H. Kim, B. Kim and J. Yu, Computational fluid dynamics analysis in microbial fuel
890		cells with different anode configurations, <i>Water Sci. Technol.</i> , 2014, 69 , 1447–1452.
891	90	D. C. Vuono, J. Regnery, D. Li, Z. L. Jones, R. W. Holloway and J. E. Drewes, rRNA Gene
892		Expression of Abundant and Rare Activated-Sludge Microorganisms and Growth Rate
893	0.1	Induced Micropollutant Removal, <i>Environ. Sci. Technol.</i> , 2016, 50 , 6299–6309.
894	91	L. Wu, D. Ning, B. Zhang, Y. Li, P. Zhang, X. Shan, Q. Zhang, M. Brown, Z. Li, J. D. Van
895		Nostrand, F. Ling, N. Xiao, Y. Zhang, J. Vierneilig, G. F. Wells, Y. Yang, Y. Deng, Q. Iu,
890 807		A. wang, I. Zhang, Z. He, J. Keher, P. H. Nielsen, P. J. J. Alvarez, C. S. Uridale, M. Wagner, I. M. Tiodia, O. Ha, T. P. Curtis, D. A. Stahl, J. Alvarez, Cohen, P. E. Dittmann, V.
07/ 800		Wan I Zhou D. Acavado M. Agullo Paroolo C. I. Anderson J. C. do Aroujo V. Dochrite
070 800		P Bond C B Bott P Boyin R K Brewster F Buy A Cabezas I Cabrol S Chap C
900 900		Fichebehere A Ford D Frigon I S $G\tilde{A}^{3}$ mez I S Griffin A 7 Gu M Habagil I Hala
200		Eunovenier, A. Polu, D. Prigui, J. S. OA mcz, J. S. Offinni, A. Z. Ou, W. Hauagii, L. Hale,

901	S. D. Hardeman, M. Harmon, H. Horn, Z. Hu, S. Jauffur, D. R. Johnson, A. Keucken, S.
902	Kumari, C. D. Leal, L. A. Lebrun, J. Lee, M. Lee, Z. M. P. Lee, M. Li, X. Li, Y. Liu, R. G.
903	Luthy, L. C. MendonÃSa-Hagler, F. G. R. de Menezes, A. J. Meyers, A. Mohebbi, A.
904	Oehmen, A. Palmer, P. Parameswaran, J. Park, D. Patsch, V. Reginatto, F. L. de los Reyes,
905	A. Noyola, S. Rossetti, J. Sidhu, W. T. Sloan, K. Smith, O. V. de Sousa, K. Stephens, R.
906	Tian, N. B. Tooker, D. De los Cobos Vasconcelos, S. Wakelin, B. Wang, J. E. Weaver, S.
907	West, P. Wilmes, S. G. Woo, J. H. Wu, L. Wu, C. Xi, M. Xu, T. Yan, M. Yang, M. Young,
908	H. Yue, Q. Zhang, W. Zhang, Y. Zhang and H. Zhou, Global diversity and biogeography of
909	bacterial communities in wastewater treatment plants, Nat. Microbiol., 2019, 4, 1183–1195.
910	92 A. M. Saunders, M. Albertsen, J. Vollertsen and P. H. Nielsen, The activated sludge
911	ecosystem contains a core community of abundant organisms, <i>ISME J.</i> , 2016, 10 , 11–20.
912	93 S. Jung and J. M. Regan, Comparison of anode bacterial communities and performance in
913	microbial fuel cells with different electron donors, Appl. Microbiol. Biotechnol., 2007, 77,
914	393–402.
915	94 D. Pant, G. Van Bogaert, L. Diels and K. Vanbroekhoven, A review of the substrates used in
916	microbial fuel cells (MFCs) for sustainable energy production, Bioresour. Technol., 2010,
917	101 , 1533–1543.
918	95 S. Ishii, S. Suzuki, T. M. Norden-Krichmar, A. Tenney, P. S. G. Chain, M. B. Scholz, K. H.
919	Nealson and O. Bretschger, A novel metatranscriptomic approach to identify gene
920	expression dynamics during extracellular electron transfer, Nat. Commun., 2013, 4, 1-10.
921	96 B. E. Logan, Exoelectrogenic bacteria that power microbial fuel cells, Nat. Rev. Microbiol.,
922	2009, 7, 375–381.
923	97 Y. Xiao, Y. Zheng, S. Wu, EH. Zhang, Z. Chen, P. Liang, X. Huang, ZH. Yang, IS. Ng,
924	BY. Chen and F. Zhao, Pyrosequencing Reveals a Core Community of Anodic Bacterial
925	Biofilms in Bioelectrochemical Systems from China, Front. Microbiol., 2015, 6, 1410.
926	98 X. Zhao, L. Li, D. Wu, T. Xiao, Y. Ma and X. Peng, Modified Anaerobic Digestion Model
927	No. 1 for modeling methane production from food waste in batch and semi-continuous
928	anaerobic digestions, Bioresour. Technol., 2019, 271, 109-117.
929	99 P. Reichert and N. Schuwirth, Linking statistical bias description to multiobjective model
930	calibration, <i>Water Resour. Res.</i> , 2012, 48 , 2011WR011391.
931	100X. Flotats, B. K. Ahring and I. Angelidaki, Parameter identification of thermophilic
932	anaerobic degradation of valerate, <i>Appl. Biochem. Biotechnol.</i> , 2003, 109 , 47–62.
933	101 N. Noykova, I. Müller, M. Gyllenberg and J. Timmer, Quantitative analyses of anaerobic
934	wastewater treatment processes: identifiability and parameter estimation., <i>Biotechnol.</i>
935	Bloeng., 2002, 78, 89–103.
930	102A. Donoso-Bravo, J. Maller, C. Martin, J. Rouriguez, C. A. Aceves-Lara and A. V. Wouwer,
937	2011 AF 5247 5264
938	2011, 45, 5547–5504. 102T. G. Müller, N. Nevkeye, M. Gullenberg and I. Timmer, Decemeter identification in
939	dynamical models of anarchic waste water treatment Math Biosci 2002 177 178 147
940 0/1	160
947	1047 Boger Application of neural networks to water and wastewater treatment plant operation
943	ISA Trans 1992 31 25–33
944	105 J. Jawad, A. H. Hawari and S. Javaid Zaidi, Artificial neural network modeling of
945	wastewater treatment and desalination using membrane processes: A review, <i>Chem. Eng. J.</i>
946	2021, 419 , DOI:10.1016/j.cej.2021.129540.

- 947 106S. Zendehboudi, N. Rezaei and A. Lohi, Appl. Energy, 2018, 228, 2539–2566.
- 948 107 M. Ben Nasr and M. Chtourou, A hybrid training algorithm for feedforward neural networks,
 949 *Neural Process. Lett.*, 2006, 24, 107–117.
- 108J. Thibault, V. Van Breusegem and A. Chéruy, On-line Prediction of Fermentation Variables
 Using Neural Networks, *Biotechnol. Bioeng.*, 1990, **36**, 1041–1048.
- 109Y. Y. Yang and D. A. Linkenst, Modelling of continuous bioreactors via neural networks,
 Trans. Inst. Meas. Control, 1993, 15, 158–169.
- 110P. Kundu, A. Debsarkar and S. Mukherjee, Artificial Neural Network Modeling for
 Biological Removal of Organic Carbon and Nitrogen from Slaughterhouse Wastewater in a
 Sequencing Batch Reactor, *Adv. Artif. Neural Syst.*, 2013, 2013, 1-15,
 DOI:10.1155/2013/268064.
- 111B. Mahanty, M. Zafar and H. S. Park, Characterization of co-digestion of industrial sludges
 for biogas production by artificial neural network and statistical regression models, *Environ. Technol. U. K.*, 2013, 34, 2145–2153.
- 961 112P. Koehn, University of Tennessee, 1994.
- 113M. Shariati, M. S. Mafipour, P. Mehrabi, A. Bahadori, Y. Zandi, M. N. A. Salih, H. Nguyen,
 J. Dou, X. Song and S. Poi-Ngian, Application of a hybrid artificial neural network-particle
 swarm optimization (ANN-PSO) model in behavior prediction of channel shear connectors
 embedded in normal and high-strength concrete, *Appl. Sci. Switz.*, 2019, 9,
 DOI:10.3390/app9245534.
- 114H. Lu, J. Chen and L. Guo, in *Comprehensive Energy Systems*, Elsevier, 2018, vol. 5–5, pp.
 258–314.
- 115H. Tyralis, G. Papacharalampous and A. Langousis, A brief review of random forests for
 water scientists and practitioners and their recent history in water resources, *Water*, 2019, 11,
 910.
- 972 116L. Breiman, RANDOM FORESTS-RANDOM FEATURES, 1999.
- 117W. Li, C. Li and T. Wang, Application of machine learning algorithms in mbr simulation
 under big data platform, *Water Pract. Technol.*, 2020, 15, 1238–1247.
- 118M. J. Song, S. Choi, W. Bin Bae, J. Lee, H. Han, D. D. Kim, M. Kwon, J. Myung, Y. M.
 Kim and S. Yoon, Identification of primary effecters of N2O emissions from full-scale
 biological nitrogen removal systems using random forest approach, *Water Res.*, 2020, 184,
 116114, DOI:10.1016/j.watres.2020.116144.
- 119B. Szeląg, A. Gawdzik and A. Gawdzik, Application of selected methods of black box for
 modeling the settlability process in wastewater treatment plant, 2017, 24, 119-127,
 DOI:10.1515/eces-2017-0009.
- 120B. Szeląg, J. Drewnowski, G. Łagód, D. Majerek, E. Dacewicz and F. Fatone, Soft sensor
 application in identification of the activated sludge bulking considering the technological and
 economical aspects of smart systems functioning, *Sens. Switz.*, 2020, 20, 1941.
- 121 H. Tyralis, G. Papacharalampous and A. Langousis, A Brief Review of Random Forests for
 Water Scientists and Practitioners and Their Recent History in Water Resources, *Water*,
 2019, 11, 910.
- 988 122 M. J. Song, S. Choi, W. B. Bae, J. Lee, H. Han, D. D. Kim, M. Kwon, J. Myung, Y. M. Kim
- and S. Yoon, Identification of primary effecters of N2O emissions from full-scale biological
- nitrogen removal systems using random forest approach, *Water Res.*, 2020, **184**, 116114.

- 123 D. Wang, S. Thunéll, U. Lindberg, L. Jiang, J. Trygg, M. Tysklind and N. Souihi, A machine
 learning framework to improve effluent quality control in wastewater treatment plants, *Sci. Total Environ.*, 2021, **784**, 147138.
- 994 124G. Louppe, University of Liège, 2014.
- 125J. M. Mendel and G. C. Mouzouris, Designing Fuzzy Logic Systems, *IEEE Trans. Circuits Syst.*, 1997, 44, 885.
- 126L. Fan and K. Boshnakov, in *Proceedings of the World Congress on Intelligent Control and Automation (WCICA)*, 2010, pp. 4142–4146.
- 127H. Xu and R. Vilanova, in 2015 23rd Mediterranean Conference on Control and Automation, MED 2015 - Conference Proceedings, Institute of Electrical and Electronics Engineers Inc., 2015, pp. 545–550.
- 1002 128T.J.J.Kalker, C.P.van Goor, P.J.Roeleveld, M.F.Ruland and R.Babuška, Fuzzy control of
 aeration in an activated sludge wastewater treatment plant: design, simulation and evaluation,
 Water Sci. Technol., 1999, 39, 61–69.
- 1005 129 A. Robles, E. Latrille, M. V. Ruano and J. P. Steyer, A fuzzy-logic-based controller for
 1006 methane production in anaerobic fixed-film reactors, *Public Health Titles*, 2017, 38, 42–52.
- 1007 130J. S. R. Jang, ANFIS: Adaptive-Network-Based Fuzzy Inference System, *IEEE Trans. Syst.* 1008 *Man Cybern.*, 1993, 23, 665–685.
- 1009 131R. Rustum, Modelling Activated Sludge Wastewater Treatment Plants Using Artificial
 1010 Intelligence Techniques (Fuzzy Logic and Neural Networks), 2009.
- 1011 132T. Y. Pai, P. Y. Yang, S. C. Wang, M. H. Lo, C. F. Chiang, J. L. Kuo, H. H. Chu, H. C. Su, L.
 1012 F. Yu, H. C. Hu and Y. H. Chang, Predicting effluent from the wastewater treatment plant of
 1013 industrial park based on fuzzy network and influent quality, *Appl. Math. Model.*, 2011, 35,
 1014 3674–3684.
- 1015 133M. Huang, J. Wan, K. Hu, Y. Ma and Y. Wang, Enhancing dissolved oxygen control using
 1016 an on-line hybrid fuzzy-neural soft-sensing model-based control system in an
 1017 anaerobic/anoxic/oxic process, *J Ind Microbiol Biotechnol*, 2013, 40, 1393–1401.
- 1018 134I. A. Essienubong, A.-I. Effiong Ndon and J. Etim, Fuzzy modeling and optimization of
 anaerobic co-digestion process parameters for effective biogas yield from bio-wastes, *Int. J.* 1020 *Energy Eng. Sci.*, 2020, 43–61.
- 1021 135A. Hosseinzadeh, J. L. Zhou, A. Altaee, M. Baziar and X. Li, Modeling water flux in osmotic membrane bioreactor by adaptive network-based fuzzy inference system and artificial neural network, *Bioresour. Technol.*, 2020, **310**, 123391,
- 1024 DOI:10.1016/j.biortech.2020.123391.
- 1025 136M. Ansari, F. Othman and A. El-Shafie, Optimized fuzzy inference system to enhance
 prediction accuracy for influent characteristics of a sewage treatment plant, *Sci. Total Environ.*, 2020, **722**, 137878.
- 1028 137K. Yetilmezsoy, in *Handbook of Environmental Materials Management*, Springer
 1029 International Publishing, 2019, pp. 2001–2046.
- 1030 138C. González-Figueredo, R. Alejandro Flores-Estrella and O. A. Rojas-Rejón, in *Current* 1031 *Topics in Biochemical Engineering*, IntechOpen, 2019.
- 1032 139Y. D. Zhao, E. N. Yamoah and P. G. Gillespie, Regeneration of broken tip links and
 1033 restoration of mechanical transduction in hair cells, *Proc. Natl. Acad. Sci. U. S. A.*, 1996, 93,
 1034 15469–15474.
- 1035 140D. S. Lee, C. O. Jeon, J. M. Park and K. S. Chang, Hybrid neural network modeling of a full-1036 scale industrial wastewater treatment process, *Biotechnol. Bioeng.*, 2002, **78**, 670–682.

- 1037 141D. S. Lee, P. A. Vanrolleghem and M. P. Jong, Parallel hybrid modeling methods for a full 1038 scale cokes wastewater treatment plant, *J. Biotechnol.*, 2005, 115, 317–328.
- 1039 142M. von Stosch, R. Oliveira, J. Peres and S. Feyo de Azevedo, Hybrid semi-parametric
 1040 modeling in process systems engineering: Past, present and future, *Comput. Chem. Eng.*,
 1041 2014, **60**, 86–101.
- 143 S. Banihashemi, G. Ding and J. Wang, Developing a Hybrid Model of Prediction and
 Classification Algorithms for Building Energy Consumption, *Energy Procedia*, 2017, **110**,
 371–376.
- 1045 144S. J. McIlroy, A. M. Saunders, M. Albertsen, M. Nierychlo, B. McIlroy, A. A. Hansen, S. M.
 1046 Karst, J. L. Nielsen and P. H. Nielsen, MiDAS: the field guide to the microbes of activated
 1047 sludge, *Database*.
- 1048 145N. Hvala and J. Kocijan, Design of a hybrid mechanistic/Gaussian process model to predict
 1049 full-scale wastewater treatment plant effluent, *Comput. Chem. Eng.*, 2020, 140, 106914,
 1050 DOI:10.1016/j.compchemeng.2020.106934.
- 1051 146T. Větrovský and P. Baldrian, The Variability of the 16S rRNA Gene in Bacterial Genomes
 1052 and Its Consequences for Bacterial Community Analyses, *PLoS ONE*, 2013, 8, 1-10,
 1053 DOI:10.1371/journal.pone.0057923.
- 1054 147V. Aguiar-Pulido, W. Huang, V. Suarez-Ulloa, T. Cickovski, K. Mathee and G. Narasimhan,
 1055 *Evol. Bioinforma.*, 2016, 12, 5–16.
- 1056 148K. Yu and T. Zhang, Metagenomic and metatranscriptomic analysis of microbial community
 1057 structure and gene expression of activated sludge, *PLoS ONE*, 2012, 7, 38183.
- 1058 149L. Ye, R. Mei, W. T. Liu, H. Ren and X. X. Zhang, Machine learning-aided analyses of 1059 thousands of draft genomes reveal specific features of activated sludge processes, 1060 *Microbiome*, 2020, 8, 1–13.
- 1061 150P. E. Larsen, D. Field and J. A. Gilbert, Predicting bacterial community assemblages using
 an artificial neural network approach, *Nat. Methods*, 2012, 9, 621–625.
- 1063 151 J. L. Metcalf, Z. Z. Xu, S. Weiss, S. Lax, W. Van Treuren, E. R. Hyde, S. J. Song, A. Amir,
 1064 P. Larsen, N. Sangwan, D. Haarmann, G. C. Humphrey, G. Ackermann, L. R. Thompson, C.
 1065 Lauber, A. Bibat, C. Nicholas, M. J. Gebert, J. F. Petrosino, S. C. Reed, J. A. Gilbert, A. M.
 1066 Lynne, S. R. Bucheli, D. O. Carter and R. Knight, Microbial community assembly and
 1067 metabolic function during mammalian corpse decomposition, *Science*, 2016, 351, 158–162.
- 1068 152K. L. Lesnik and H. Liu, Predicting Microbial Fuel Cell Biofilm Communities and
 1069 Bioreactor Performance using Artificial Neural Networks, *Environ. Sci. Technol.*, 2017, 51,
 1070 10881–10892.
- 1071 153K. L. Lesnik, W. Cai and H. Liu, Microbial Community Predicts Functional Stability of
 1072 Microbial Fuel Cells, *Environ. Sci. Technol.*, DOI:10.1021/acs.est.9b03667.
- 1073 154H. Yuan, S. Sun, I. M. Abu-Reesh, B. D. Badgley and Z. He, Unravelling and
 1074 Reconstructing the Nexus of Salinity, Electricity, and Microbial Ecology for
 1075 Bioelectrochemical Desalination, *Environ. Sci. Technol.*, 2017, **51**, 12672–12682.
- 1076 155J. Kuang, L. Huang, Z. He, L. Chen, Z. Hua, P. Jia, S. Li, J. Liu, J. Li, J. Zhou and W. Shu,
 1077 Predicting taxonomic and functional structure of microbial communities in acid mine
 1078 drainage, *ISME J.*, 2016, **10**, 1527–1539.
- 1079 156H. Yuan, R. Mei, J. Liao and W. T. Liu, Nexus of stochastic and deterministic processes on
- microbial community assembly in biological systems, *Front. Microbiol.*, 2019, 10, 1-12,
 DOI:10.3389/fmicb.2019.01536.

- 1082 157I. Vanwonterghem, P. D. Jensen, P. G. Dennis, P. Hugenholtz, K. Rabaey and G. W. Tyson,
 1083 Deterministic processes guide long-term synchronised population dynamics in replicate
 1084 anaerobic digesters, *ISME J.*, 2014, **8**, 2015–2028.
- 1085 158J. S. Griffin and G. F. Wells, Regional synchrony in full-scale activated sludge bioreactors 1086 due to deterministic microbial community assembly, *ISME J.*, 2017, **11**, 500–511.
- 1087 159L. Zhang, P. Zheng, C. Tang and R. Jin, Anaerobic ammonium oxidation for treatment of
- ammonium-rich wastewaters., J. Zhejiang Univ. Sci. B, 2008, 9, 416–426.
- 1089

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1092 Figure 1. Timeline of model concepts (red), model strategies (blue), control and monitoring

1093 development (green), computational infrastructure (orange), and development of molecular

1094 biology and bioinformatics (black).

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1097 Figure 2. The characteristics and advantages of mechanistic, data-driven, and hybrid models



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1100 Figure 3. Incorporation of genomic data into model construction. The diagrams of S-, N-, and C-

- 1101 cycle are originated from the study of Wu and Yin⁷. The figures of phylogenic trees, PCoA, and
- 1102 data-driven modeling analysis on functional expression are based on the study of Cheng et al.³¹



Figure 4. (A) A Bayesian network trained with the microbial population dynamics at the order
level in a bioelectrochemical system. (B) Predicted vs. observed current production. (C)
Prediction of current production as a function of substrate salinity. Figures adapted from Yuan et
al.¹⁵⁴



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Figure 5. (A) A Bayesian network trained with the genes for alcohol metabolism, hydrogen
metabolism, direct interspecies electron transfer, and methanogenesis from dominant microbes.
(B) Prediction of methane production with a complete Bayesian network and in-silico knockout
of relevant genes. Figures adapted from Yuan et al.³⁰



1114 Figure 6. (A) A Bayesian network as the data-driven component of the hybrid model was trained

- 1115 with microbial population dynamics and microbial kinetic parameters estimated from the
- 1116 mechanistic component (green oval nodes). (B) Predicted vs. observed current production.
- 1117 Figures adapted from Cheng et al.³¹