

Environmental Science Water Research & Technology

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

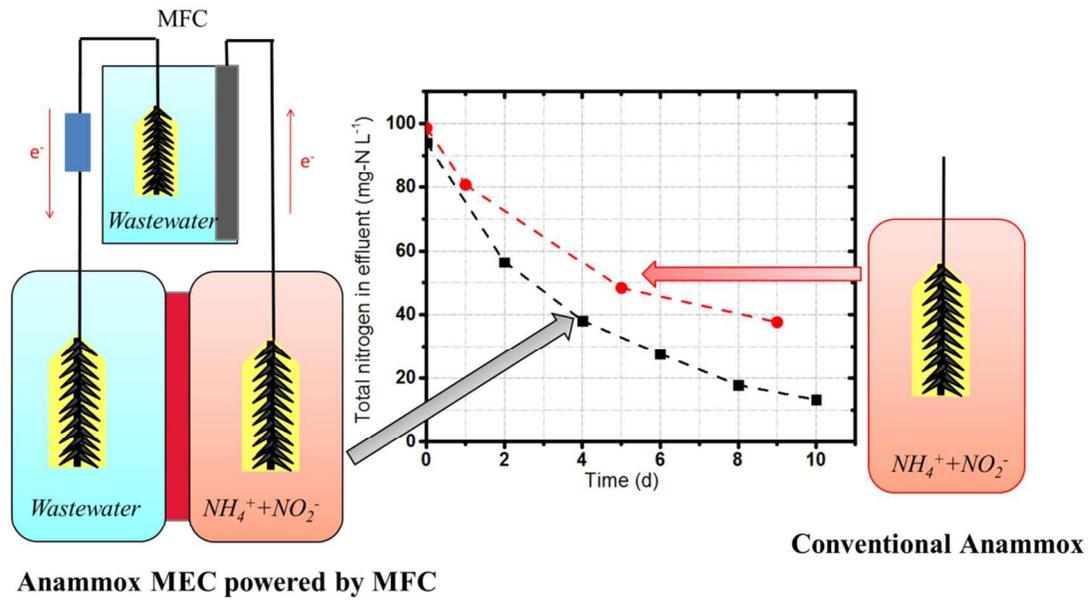
Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

This pioneer study provides a practical solution to the imminent problem of anammox: slow growing anammox bacteria. Unique integration of MECs and MFCs alleviates the dependence on anammox bacteria, and accelerates total nitrogen (TN) removal efficiency 30% higher than conventional anammox. A novel bioelectrochemical anammox kinetic model is developed. This study greatly improves the scientific understanding and engineering application of energy-positive high rate biological nutrient removal processes.

Graphic abstract:



1 Self-sustained high-rate Anammox: from biological to bioelectrochemical process

2 Yan Li Zhiheng Xu Dingyi Cai Brandon Holland Baikun Li *

3 Department of Civil and Environmental Engineering, University of Connecticut, 261 Glenbrook
4 Road, Unit 2037, CT 06269, USA

5 * Corresponding author. Tel.: +1 860 486 2339

6 Email address: baikun@enr.uconn.edu

8 Abstract

9 Slow growth rate of anammox bacteria is the imminent problem for system efficiency and
10 stability. An innovative solution was explored in this study by accelerating anammox in
11 microbial electrolysis cells (MECs) and alleviating the dependence on anammox bacteria. The
12 batch tests showed that 85 % of total nitrogen (TN) was removed in the MEC system, while only
13 62 % of TN was removed in conventional anammox. Simulation of the modified Nernst-Monod
14 model revealed that the maximum specific utilization rate ($0.30\text{-}0.38 \text{ mmol g}^{-1}\text{VSS h}^{-1}$) in the
15 anammox MEC was 60 % higher than in conventional anammox ($0.18\text{-}0.20 \text{ mmol g}^{-1}\text{VSS h}^{-1}$).
16 Harvesting the power generated in microbial fuel cells (MFCs) to support MECs substantially
17 saved energy consumption and effectively utilized the low power output of MFCs. Simulation of
18 power management system (PMS) interface demonstrated the charge/discharge cycles for power
19 supply by MFCs and power consumption by MECs. The integrated MEC-MFC system
20 accelerated anammox, avoided external carbon requirement, effectively utilized wastewater
21 energy, and thus achieving self-sustained nitrogen removal.

22 **Keywords:** microbial electrolysis cell, anammox, nitrite, Nernst-Monod model, microbial fuel
23 cell, power management system.

24

25 Introduction

26 Nitrogen in wastewater is normally removed using biological nitrogen removal (BNR) processes
27 consisting of aerobic nitrification and anoxic denitrification. However, conventional BNR is
28 energy-negative and carbon-intensive due to the requirements of high aeration in nitrification,
29 pH adjustment, long sludge retention time (SRT), and exogenous organic carbon in
30 denitrification.¹ In the past decade, anaerobic ammonium oxidation (ANAMMOX) has been
31 developed in which ammonium (NH_4^+) and nitrite (NO_2^-) are directly reacted to form nitrogen
32 gas (N_2) by anoxic anammox bacteria, and thus possessing unique advantages over conventional
33 BNRs including no need for aeration and external carbon sources.² Nevertheless, anammox
34 suffers from long growth time (doubling time of 11 days) of anammox bacteria and difficult
35 cultivation, making it barely operated in full-scale wastewater treatment plants.³

36 Bioelectrochemical systems (BES) have drawn global attention for converting the chemical
37 energy stored in wastewater to electrical energy.⁴⁻¹⁰ Microbial fuel cells (MFCs) utilize the
38 difference between anode and cathode potentials as the driving force for electricity generation in
39 spontaneous reactions (e.g. oxygen reduction, metal (Cr^{6+} , Cu^{2+}) reduction) and have been
40 studied to reduce the energy cost of denitrification with nitrite and nitrate as the electron
41 acceptors in the cathode chamber.¹¹⁻¹⁶ On the other hand, microbial electrolysis cells (MECs)
42 that the difference between anode and cathode potentials is deficient can be supported with an
43 external power supply (voltage < 1.0 V) to make non-spontaneous reaction proceed and
44 accelerate spontaneous reactions. For instance, chromium (VI) reduction was accelerated in
45 MECs by changing cathode potential.¹⁷ Nitrogen removal with nitrification on anode and
46 denitrification on cathode was achieved in MECs.¹⁸ Previous study found that ammonium served
47 as the anodic fuel in MFCs and current generation was promoted during the process of electron

48 transport from ammonium to nitrite.¹⁹ Therefore, installing power supply to anammox in MECs
49 could accelerate ammonium oxidation.

50 The limited electric power output (less than 5 W m^{-2}) of MFCs has posed a problem for using
51 MFCs as the sole power source.^{20,21} Thus, an efficient utilization of low MFC power is critical
52 for its real-world application. Past several years have seen MFCs as the power source for low
53 power subsea devices, water quality sensors, and MECs of hydrogen production and metal
54 treatment.^{13,22-25} Especially, the low voltage requirement of MECs ($<0.5 \text{ V}$) makes the MFC a
55 proper power supply. But the interface between power production in MFCs and power
56 consumption in MECs has not been established, making it difficult to predict the power supply
57 and consumption between MFCs for MECs.

58 Reliable estimation of kinetic parameters in anammox modeling is critical to better understand
59 anammox mechanisms and enhance its performance. Monod kinetics, pseudo first order model
60 and Haldane-type model were used for anammox simulation.²⁶⁻²⁹ A wide range of half-velocity
61 constants ($0.003\text{-}13.7 \text{ mM}$) and maximum specific utilization rate ($0.09\text{-}3.74 \text{ mmol N g}^{-1}\text{VSS h}^{-1}$)
62 has been obtained.²⁹ With two reactants (ammonium and nitrite) being involved in anammox
63 reactions, Monod model with multiple and/or dual substrates should be used, which could reflect
64 the dependence of each substrate in reactions. In the meantime, the Nernst-Monod model has
65 been developed for electron donation and acceptance between substrates and biofilm electrodes
66 in MFCs and MECs, and modified for the reduction of nitrate and nitrite in cathode.^{30,31} Until
67 now, there has been no model for anammox in MECs or MFCs. Due to the complexity of redox
68 reactions and multiple substrates involved in anammox, a novel Nernst-Monod model coupling
69 with multi-substrates Monod model should be developed to simulate the anammox in BES
70 systems and the dependence of each substrates.

71 The breakthrough of this study was to accelerate anammox in MECs without solely counting
72 on anammox bacteria and to explore the feasibility of the integrated MFC-MEC in wastewater
73 treatment plants. MECs were powered by MFCs treating wastewater to achieve self-sustained
74 nitrogen removal without extra energy input and efficiently utilize the low power output of
75 MFCs, which none of existing BNR and BES has accomplished. There were four tasks in this
76 study. First, ammonium and nitrite were fed in MECs to examine their degradation rates. The
77 accelerated anammox mechanisms in MECs were verified by conducting three control tests
78 (conventional anammox, ammonium/nitrite alone, and abiotic MEC test). Second, the kinetic
79 model of accelerated anammox in MECs was developed to fundamentally understand the self-
80 sustained anammox and the dependency of ammonium and nitrite in anammox reactions. Third,
81 the variation of important parameters (e.g. pH, redox potential) was examined to determine the
82 occurrence of anammox in MECs, and relate with nitrogen removal rate. Finally, the interface of
83 the integrated MFC-MEC system was modeled to predict the power supply of MFCs to
84 anammox MECs. Energy saving of the integrated system was calculated and compared with
85 traditional BNR and anammox processes to confirm the novel self-sustained anammox MFC-
86 MEC with high nitrogen removal rate and minimal energy consumption.

87

88 **Materials and Method**

89 **The anammox MEC setup**

90 The batch-mode anammox MEC consisted of an anode chamber (volume: 140 mL) and a
91 cathode chamber (volume: 140 mL) separated by proton exchange membrane (N117, DuPont
92 Fuel Cells, DE) (Fig. 1 Anammox MEC). During the acclimation period, the anammox MEC
93 was powered by a programmable power supply (model 3645A; Circuit Specialists, Inc.) with the

94 stable voltage output of 0.5 V. The power supply was later replaced by two single-chamber
95 microbial fuel cells (SCMFCs) (each volume: 140 mL) connected in series during the
96 experimental period. The anode of the anammox MEC was connected with the cathode of
97 SCMFCs and the cathode of the anammox MEC was connected with the external resistance (R_{ext}
98 5Ω) in the circuit connecting the anode of SCMFCs. The voltages across the R_{ext} were
99 continuously recorded every 120 s using a Keithley 2700 data logging system. Carbon brushes
100 (4cm long by 4cm diameter as projected area, Mill-Rose Carbon Fiber Brush) were used as the
101 anode and cathode of anammox MEC, and the anode of the SCMFCs. Carbon cloth ($4 \times 4 \text{ cm}^2$,
102 Fuel Cell Earth LLC, MA) loaded with platinum (Pt) (0.5 mg cm^{-2}) was used as cathode of
103 SCMFCs.

104 Please add Fig. 1 here
105

106 **Inoculation and operation of anammox MEC and control experiments**

107 The anode chamber of the anammox MEC was inoculated with the mixture of aerobic and
108 anoxic sludge (volume ratio: 1:1) from the University of Connecticut Wastewater Treatment
109 Plant (UConn-WWTP). The ammonium chloride (NH_4Cl , $\sim 50 \text{ mg-N L}^{-1}$), sodium nitrite
110 (NaNO_2 , $\sim 40 \text{ mg-N L}^{-1}$), and the medium solution (0.2 mM Na_2HPO_4 , 0.1 mM NaH_2PO_4 , 0.2
111 mM KCl, 0.2 mM $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, and 6mM NaHCO_3) were fed to stimulate the anammox
112 bacteria growth in the anode. The inoculation lasted five weeks by changing half of the anodic
113 solution (around 70 ml) weekly and refilling with fresh ammonium, nitrite and nutrient solution.
114 When the experiment started, the initial volatile suspended solids (VSS) concentration was about
115 0.4 g L^{-1} in the anammox MECs. The cathode was fed with the influent wastewater from UConn-
116 WWTP. Both the anode and cathode were sealed with plastic caps to secure anaerobic conditions

117 and prevent the occurrence of nitrification. The SCMFCs were filled with the mixture of the
118 influent wastewater from UConn-WWTP and sodium acetate (15 mM) with the chemical oxygen
119 demand (1500 mg L⁻¹) and operated more than one month to provide stable voltage (0.5 V) for
120 anammox MECs.

121 Along with the experiment of anammox MECs, three control experiments were operated side
122 by side for comparison. Control 1 (conventional anammox) was conducted in a sealed plexiglass
123 bottle (volume: 140 mL) with a carbon brush for biofilm growth (Fig. 1 Control 1). No power
124 supply or SCMFCs was connected. Control 2 had the similar setup to the anammox MEC, except
125 feeding with either ammonium or nitrite solution (Fig. 1 Control 2) to examine the nitrogen
126 removal under single nitrogen species, and the dependence of ammonium and nitrite for
127 anammox and denitrification. Control 3 (abiotic test) was the similar setup to the anammox MFC,
128 expect no sludge was inoculated in the anode (Fig. 1 Control 3) to compare with Control 1
129 (biotic anammox tests) and determine the role of bacteria in nitrogen removal in MECs. All the
130 tests of anammox MECs and Controls were carried out at 30°C in duplicate.

131

132 **Analysis of anammox MEC and control experiments**

133 Ammonium concentration, nitrite, nitrate, chemical oxygen demand (COD), alkalinity were
134 measured with a spectrophotometer (DR 2800, HACH company, CO) and TNTplusTM series
135 (HACH company, CO). The pH in anammox MEC and controls were measured with a portable
136 pH meter (Thermo Fisher Scientific Orion 3-star). The redox potential (ORP) of the anammox
137 MEC and control experiments were measured with ORP meter (Thermo Fisher Scientific Orion
138 3-star). The open circuit potentials (OCPs) of anodes and cathodes were measured using an
139 Ag/AgCl (+197 mV vs SHE) as the reference.

140

141 **Modeling anammox MEC and conventional anammox**

142 In order to predict the variation of ammonium and nitrite concentrations in the anammox MEC
 143 and estimate the kinetic parameters, Nernst-Monod Equation (Eq.1 and Eq.2) modified from
 144 previous studies was used,^{30,31} assuming that ammonium and nitrite (nitrate was negligible in this
 145 study) were the electron donors and anode electrode was the electron acceptor. In the meantime,
 146 Monod kinetics of double substrates was used to simulate nitrogen removal in conventional
 147 anammox without power supply (Control 1) (Eq.3 and Eq.4).

148

$$149 \quad \frac{d[\text{NH}_4^+]}{dt} = -\mu_{m\text{NH}_4^+\text{MEC}} \left(\frac{[\text{NH}_4^+]}{[\text{NH}_4^+] + K_{\text{SNH}_4^+\text{MEC}}} \right) \left(\frac{[\text{NO}_2^-]}{([\text{NO}_2^-] + K_{\text{SNO}_2^-\text{MEC}})} \right) \left(\frac{1}{1 + \exp\left[-\frac{F}{RT}\eta\right]} \right) \quad (1)$$

$$150 \quad \frac{d[\text{NO}_2^-]}{dt} = -\mu_{m\text{NO}_2^-\text{MEC}} \left(\frac{[\text{NO}_2^-]}{([\text{NO}_2^-] + K_{\text{SNO}_2^-\text{MEC}})} \right) \left(\frac{[\text{NH}_4^+]}{[\text{NH}_4^+] + K_{\text{SNH}_4^+\text{MEC}}} \right) \left(\frac{1}{1 + \exp\left[-\frac{F}{RT}\eta\right]} \right) \quad (2)$$

$$151 \quad \frac{d[\text{NH}_4^+]}{dt} = -\mu_{m\text{NH}_4^+\text{ana}} \left(\frac{[\text{NH}_4^+]}{[\text{NH}_4^+] + K_{\text{SNH}_4^+\text{ana}}} \right) \left(\frac{[\text{NO}_2^-]}{([\text{NO}_2^-] + K_{\text{SNO}_2^-\text{ana}})} \right) \quad (3)$$

$$152 \quad \frac{d[\text{NO}_2^-]}{dt} = -\mu_{m\text{NO}_2^-\text{ana}} \left(\frac{[\text{NO}_2^-]}{([\text{NO}_2^-] + K_{\text{SNO}_2^-\text{ana}})} \right) \left(\frac{[\text{NH}_4^+]}{[\text{NH}_4^+] + K_{\text{SNH}_4^+\text{ana}}} \right) \quad (4)$$

153 Where $[\text{NH}_4^+]$ and $[\text{NO}_2^-]$ are the effluent concentrations of ammonium and nitrite (mg-N
 154 L^{-1}); μ_m is the maximum specific utilization rate (mg-N $\text{g}^{-1}\text{VSS h}^{-1}$); K_s is the half-velocity
 155 constant (mg-N L^{-1}); F is Faraday's constant ($9.64853 \times 10^4 \text{ C mol}^{-1}$); R is gas constant (8.314 J K^{-1}
 156 mol^{-1}); T is operation temperature (K); η is the local potential (V), it was estimated as 0.2 V in
 157 this study.²⁷

158 The results of ammonium and nitrite concentrations in the anammox MEC and Control 1 over
 159 the experimental period were fit into the models (Eqs.1-4) to calculate critical kinetic parameters
 160 –maximum specific utilization rate and half-velocity constant. Integration of differential
 161 equations was conducted using the Matlab software (R2014a).

162

163 **Modeling power support and consumption of the integrated MFC-MEC**

164 The power support and consumption between anammox MEC and SCMFCs was simulated.
 165 Power output from MFCs was evaluated based on total coulombs (Q), and the current produced
 166 during the operational period (t) was determined based on the organics consumption (ΔCOD) in
 167 wastewater (Eq.5). Coulombs from SCMFCs (Q) were consumed in anammox MEC through the
 168 anammox pathways, so that the concentration of nitrogen removed (ΔC_N) could be simulated
 169 with the amount of electrons transferred per mole of nitrogen (NH_4^+) (Eq.6). By establishing the
 170 interface of power generation in SCMFCs and power consumption in anammox MEC, ΔCOD in
 171 SCMFCs and ΔC_N in anammox MEC could be simulated.

$$172 \quad Q = \int_0^t I dt = \frac{FV_{MFC} \Delta COD}{8} \times CE_{MFC} \quad (5)$$

$$173 \quad \Delta C_N = \frac{QM}{nFV_{MEC}} \quad (6)$$

174 Where Q is the total coulombs harvested from MFCs (C); V_{MFC} and V_{MEC} are the SCMFC
 175 volume and MEC cathode volume (L); ΔCOD is the total COD consumed in SCMFCs, (g L^{-1});
 176 CE_{MFC} is the coulombic efficiency of SCMFC (10-27 %),³² with 20% being used in Eq. 5; M is
 177 the molecular weight of nitrogen (14 g mol^{-1}); n is the electron transfer per mole of nitrogen; F is
 178 the Faraday's constant ($9.64853 \times 10^4 \text{ C mol}^{-1}$).

179

180 Results and discussion

181 Variation of ammonium and nitrite concentrations in MECs

182 Please add Fig. 2 here

183 Ammonium (NH_4^+) and nitrite (NO_2^-) concentrations gradually decreased in the MEC with the
184 power supply of 0.5 V (Fig. 2). The initial concentrations of ammonium and nitrite were set at 50
185 mg L^{-1} and 40 mg L^{-1} , respectively, to simulate domestic wastewater and the effluent after short-
186 cut nitrification (ammonium to nitrite).¹⁶ It took 10 days to remove 90 % of ammonium (average
187 concentration below 5 mg L^{-1} in the effluent) and remove 85 % of nitrite (average concentration
188 below 6 mg L^{-1} in the effluent). Nitrite decreased fast in the first 4 days and then slowed down.
189 Nitrate (NO_3^-) in the MEC was low (less than 5 mg L^{-1}) and originated from the acclimated
190 sludge residue taken from the anoxic tank of UConn WWTP. The continuous nitrogen removal
191 at the anaerobic condition ($\text{DO} < 0.1 \text{ mg L}^{-1}$, with both anode and cathode being sealed)
192 indicated the biotic process of this experiment especially the function of anammox bacteria, since
193 ammonium and nitrite could hardly be volatilized in the sealed system, nitrification could not
194 occur under anaerobic condition, and the anammox reaction could only take place with bacteria.
195 The average nitrogen removal rate was 8.2 $\text{mg-N L}^{-1} \text{ d}^{-1}$, which was in the range of the reported
196 values in BES (3-51 $\text{mg-N L}^{-1} \text{ d}^{-1}$), but was much lower than BNR (50-400 $\text{mg-N L}^{-1} \text{ d}^{-1}$).^{14,33-35}
197 The main reason was the difference in substrate transfer in these systems. Conventional BNR
198 was mainly operated in the continuous flow mode, but the lab-scale BES (e.g. MEC, MFC) was
199 normally operated in the batch mode, which limited substrate diffusion from wastewater to
200 biomass.³⁶

201

202 Control tests to verify the occurrence of anammox and denitrification in MECs

203 Please add Fig. 3 here

204 Three controls were conducted in order to further elucidate the removal pathways of ammonium
205 and nitrite in MECs. All the control tests were operated for 10 days (the same duration as the
206 MEC tests). Control test 1 (ammonia+nitrite, without power supply) simulated the anammox
207 process (Fig. 1 Control 1), in which the average ammonium, nitrite and nitrate removal rates
208 were $2.80 \text{ mg-N L}^{-1} \text{ d}^{-1}$, $3.24 \text{ mg-N L}^{-1} \text{ d}^{-1}$, $0.44 \text{ mg-N L}^{-1} \text{ d}^{-1}$ respectively (Fig. 3). In contrast,
209 the rates for the anammox MEC were $4.38 \text{ mg-N L}^{-1} \text{ d}^{-1}$, $3.38 \text{ mg-N L}^{-1} \text{ d}^{-1}$, and $0.33 \text{ mg-N L}^{-1} \text{ d}^{-1}$,
210 respectively (Fig. 3). The reason for the lower ammonium removal rate in Control 1 than
211 MEC was that power supply in the MEC expedited the ammonium removal, while ammonium
212 removal in Control 1 was solely dependent on anaerobic ammonium oxidation by nitrite. This
213 indicated that ammonium could work as the anodic fuel in MECs and transferred electrons to the
214 anode other than nitrite. In this study, the high potential provided by the power supply intrigued
215 more ammonium oxidation in MECs than in Control 1, implying that MECs had other electron
216 acceptor for ammonium oxidation compared with traditional anammox process. This finding was
217 in accordance with previous study that ammonium oxidation possessed the negative Gibbs free
218 energy to generate electricity.¹⁹ In addition, ferric ion was found to work as electron acceptor by
219 anammox bacteria without nitrite³⁷ and nitrate-dependent ferrous iron was oxidized by anammox
220 bacteria,³⁸ indicating that anammox bacteria could utilize different types of electron donors and
221 acceptors, and anammox involves numerous complicated processes. In contrast with ammonium
222 removal difference in MEC and Control 1, nitrite removal rates were similar between Control 1
223 and MEC (Fig. 3), implying that nitrite removal were not related with power supply.

224 Control 2 was the test of single nitrogen species (ammonium or nitrite) with power supply in
225 order to elucidate whether ammonium and nitrite should be co-present in wastewater for the
226 enhanced anammox-like process in MECs (Fig. 1 Control 2). The ammonium removal rate
227 without nitrite input was $1.12 \text{ mg-N L}^{-1} \text{ d}^{-1}$ (Fig. 3), which was much lower than those of MEC
228 and Control 1, while the nitrite removal rate without ammonia input was $3.28 \text{ mg-N L}^{-1} \text{ d}^{-1}$ (Fig.
229 3), which was similar with those of MEC test and Control 1. Furthermore, the slopes of linear
230 regression of ammonium, nitrite and nitrate removal rates were analyzed for the MEC, Control 1,
231 and Control 2, showing an obvious change of ammonium removal rate compared with those of
232 nitrite and nitrate (Fig. 3). The slope of ammonium data from MEC to Control 2 was -1.64,
233 meaning MEC was effective at ammonium removal, while the slope of nitrite was only -0.025,
234 meaning there was literally no change of nitrite removal among these three systems. These
235 implied that co-presence of nitrite facilitated ammonium oxidation and nitrite was the main
236 oxidant of ammonium. In contrast, nitrite removal was not affected by ammonium absence,
237 indicating that nitrite could be reduced by anammox bacteria as well as by denitrifying bacteria.
238 Denitrification process was dominant when without ammonium. Control 2 results implied that
239 ammonium oxidation and nitrite reduction were carried out by anammox bacteria and/or
240 anaerobic ammonium oxidation bacteria and denitrification bacteria. This synergic correlation
241 accelerated the electron transfer from ammonium to nitrite and ultimately to anode.

242 Control 3 (ammonium+nitrite, with power supply, abiotic test) was conducted to verify the
243 role of bacteria in anammox MECs (Fig. 1 Control 3). Both ammonium and nitrite removal rates
244 ($0.25 \text{ mg-N L}^{-1} \text{ d}^{-1}$ and $0.41 \text{ mg-N L}^{-1} \text{ d}^{-1}$) were tremendously lower than those of MEC, Control
245 1 and Control 2 (Fig. 3), indicating that the removal of ammonium and nitrite was the biological
246 process and bacteria carried out ammonium oxidation and nitrite reduction. All the MEC and

247 Control tests had low nitrate concentration ($<5 \text{ mg L}^{-1}$, mainly coming from raw wastewater) and
248 low nitrate removal rate (less than $0.49 \text{ mg-N L}^{-1} \text{ d}^{-1}$) (Fig. 3), indicating that nitrification hardly
249 occurred in anoxic conditions. Compared with complete nitrification (ammonium to nitrite and
250 then to nitrate), short-cut nitrification (ammonium to nitrite) saved substantial reaction time,
251 provided the essential electron acceptors (nitrite) in the following anammox, and saved carbon
252 sources for partial denitrification (nitrite reduction).

253 The simultaneous removal of ammonium and nitrite in the anoxic condition in 10 days (Fig. 2)
254 ascertained the occurrence of anammox in the MEC system. Nitrification was completely
255 inhibited under anoxic condition ($\text{DO} < 0.1 \text{ mg L}^{-1}$, with both anode and cathode being sealed),
256 which was verified by the stable decrease of nitrite (no accumulation) over the 10-day period.
257 This was a clear evidence that ammonium oxidation was carried out by anammox bacteria with
258 nitrite as the electron acceptor. Several studies of anammox MFCs treating wastewater had the
259 similar configurations (two chamber system), inoculation protocols (wastewater and nutrients
260 ingredients), operational duration (5-14 days), and the same trend of decrease in ammonium and
261 nitrite as this study³⁹⁻⁴², among which some validated the anammox reactions through
262 engineering tests^{39,42}, and some confirmed the existence^{39,42} of anammox bacteria by microbial
263 community analysis (e.g. anammox 16S rRNA gene as a molecular marker and functional
264 biomarkers (e.g. nitrite reductase gene) involved in the anammox metabolisms)^{40,41}. Furthermore,
265 simultaneous removal of ammonium and nitrite in anaerobic and anoxic units had been observed
266 in traditional anammox systems (not BES) treating various types of wastewater (e.g. municipal
267 wastewater, swine wastewater, landfill leachate, salinity wastewater) in bioreactors (e.g.
268 sequencing batch reactor, rotating biological contactor, upflow anaerobic sludge blanket (UASB),
269 moving bed biofilm reactor)⁴³⁻⁵¹, among which some confirmed the occurrence of anammox

270 reactions through biochemical measurement and engineering operation⁴³⁻⁴⁸ and some detected
271 anammox bacteria using microbial analysis (e.g. cloning with anammox gene primers and gene
272 sequencing, fluorescence in situ hybridization (FISH) with specific anammox bacterial probes)
273 ^{36,49-51}. By side-side comparing the experimental results in this work and previous anammox
274 studies, the existence of anammox bacteria in the MFCs was ascertained.

275 **Kinetic modeling of the accelerated anammox in MECs**

276 Please add Fig. 4 here

277 Please add Table 1 here

278 The experimental data of ammonium and nitrite concentrations in anammox MEC and Control 1
279 (conventional anammox) were fitted in Models (Eq. 1-4) to determine critical kinetic parameters.
280 Specifically, the modified Nernst-Monod equation was used for anammox MEC (Fig. 4a), even
281 though not all of the electrons were accepted by the electrode but consumed by other processes
282 (e.g. anammox and denitrification). Total nitrogen concentration change was measured over time
283 (Fig. 4b). Multiplicative Monod modeling with dual substrates was used for Control 1 (Fig. 4c).
284 The fittings of the model simulated data and the experimental results were examined using
285 statistical analysis. The coefficient of determination (R^2) of the experimental results and the
286 model simulated data was all higher than 97%, with ammonium in MECs 99.12%, nitrite in
287 MECs 98.68%, ammonium in Control 1 97.24%, and nitrite in Control 1 99.48%, respectively.
288 In addition, the models were examined by the difference (the subtraction of predicted data from
289 the observed or actual data) between the ammonium and nitrite experimental data in the MEC
290 and Control 1 (MEC experiment duplicated 4 times as shown in Fig 4a, Control 1 duplicated
291 twice as shown in Fig 4c) to validate the model fitness (the inserted figures in Fig. 4). The

292 distribution of the difference over the independent variable (time) indicated that they were
293 randomly distributed and tended to cluster towards the middle of the plots ($y=0$) in both MEC
294 and Control 1. In addition, the maximum difference was single digits (MEC maximum difference:
295 6.0 and -7.3, and Control 1 maximum difference: 5.2 and -6.0) without high bias, demonstrating
296 that the models well fitted to the experimental data.

297 The calculated maximum specific utilization rate of ammonium in the MEC was much higher
298 ($0.38 \text{ mmol-N g}^{-1}\text{VSS h}^{-1}$) than that of Control 1 ($0.18 \text{ mmol-N g}^{-1}\text{VSS h}^{-1}$) (Table 1). The
299 maximum specific utilization rate of nitrite in the MEC was also higher ($0.3 \text{ mmol-N g}^{-1}\text{VSS h}^{-1}$)
300 than that of Control 1 ($0.2 \text{ mmol-N g}^{-1}\text{VSS h}^{-1}$), implying MEC accelerated anammox reactions.
301 A wide range ($0.09\text{-}3.74 \text{ mmol-N g}^{-1}\text{VSS h}^{-1}$) of the maximum specific utilization rate had been
302 reported for conventional anammox, and associated with anammox bacteria enrichment and
303 operation modes.²⁹ In this study, anoxic bacteria (e.g. denitrifying bacteria, electrogenic bacteria)
304 in the MEC co-present with anammox bacteria, which might cause the maximum specific
305 utilization rate ($0.3\text{-}0.38 \text{ mmol-N g}^{-1}\text{VSS h}^{-1}$) not higher than the reported values. Besides, high
306 specific utilization rate was reported in the continuous flow mode with high N-loading
307 (concentration: 230 mg-N L^{-1}),²⁸ while the MEC was conducted in the batch mode in this study
308 with total nitrogen concentration of 90 mg-N L^{-1} . Nevertheless, the ammonium utilization rate in
309 the MEC (anammox with power supply) was twice as in Control 1 (conventional anammox) in
310 this study, indicating the substantial acceleration of anammox in MECs.

311 As for the half-velocity constant (K_S), a wide range ($0.003\text{-}13.7 \text{ mM}$) had been obtained due to
312 various model simulations.²⁶⁻²⁹ K_S for ammonium (3.47 mM) and nitrite (3.40 mM) were almost
313 same in Control 1 (conventional anammox) (Table1), which corresponded with the equal
314 consumption of ammonium and nitrite in anammox reaction ($\text{NH}_4^+ + \text{NO}_2^- \rightarrow \text{N}_2 + 2\text{H}_2\text{O}$). In

315 contrast, nitrite constant (2.33 mM) was lower than ammonium constant (4.11 mM) in anammox
316 MEC (Table1), indicating that ammonium was more easily utilized than nitrite, and thus the
317 power supply of MECs promote the ammonium utilization than nitrite. These results
318 corresponded well with the results of the removal rates of ammonium and nitrite in anammox
319 MEC and Control 1 (Fig. 3), implying that power supply of MEC assisted the ammonium uptake
320 more than nitrite.

321

322 **Variation of ORP and pH throughout nitrogen removal processes**

323

Please add Fig. 5 here

324 ORP (redox potential) and pH are the critical indicators for BNR processes.^{1,52} ORP values
325 increased over time in the anammox MEC, Control 1 and Control 2 (fed with nitrite), but
326 decreased greatly in Control 2 (fed with ammonium) and remained stable in Control 3 (Fig. 5b).
327 For Control 2 with ammonium input, there was no oxidizer in the system, so that ORP dropped
328 as anoxic bacteria consumed the residue dissolved oxygen and fermentation took place, which
329 also explained its low ammonium removal rate (Fig. 3). For Control 3 (abiotic test) without
330 anoxic bacteria, ORP did not change, indicating that reactions barely happen without anoxic
331 bacteria, and further proved the function of bacteria in the anammox MECs. For MEC test,
332 Control 1 and Control 2 (fed with nitrite), ammonium was oxidized with nitrite as the electron
333 acceptor and ORPs increased over time.

334 pH values increased in the MEC, Control 1, and Control 2 (fed with nitrite) (Fig. 5b),
335 indicating the occurrence of denitrification, since pH did not change in anammox ($\text{NH}_4^+ + \text{NO}_2^-$
336 $\rightarrow \text{N}_2 + 2\text{H}_2\text{O}$). In contrast, pH dropped from 7.61 to 6.87 in Control 2 (fed with ammonium),

337 which verified the occurrence of fermentation and corresponded to the ORP drop (Fig. 5a). pH
338 remained constant in Control 3 (abiotic test without bacteria), indicating the absence of
339 biochemical reactions that corresponded to low nitrogen removal rate (Fig. 3).

340

341 **The interface between SCMFCs and anammox MEC**

342 The power required by anammox MEC was provided from SCMFCs treating wastewater. The
343 variation of COD concentration (ΔCOD) in SCMFCs was correlated with the targeted nitrogen
344 concentration (ΔC_N) in anammox MEC (Eq.5 and Eq. 6). For anammox MEC, the amount of
345 electrons transferred per mole of nitrogen to the anode (n in Eq.6) was determined based on the
346 direct donation species of nitrogen. With ammonium as the electron donor for the anode (Fig. 4),
347 the value of n was 3 in Eq.6 since 3 moles of electrons were transferred per mole of N in NH_4^+ to
348 form nitrogen gas. The simulation of Eq.5 and Eq. 6 revealed the linear relationship between the
349 COD change in SCMFCs and nitrogen concentration change in MEC (Fig. 6a).

350

Please add Fig.6 here

351 COD consumption in MFCs was selected in the range of 0-4500 mg L⁻¹ based on diverse types
352 of wastewater (municipal wastewater COD: 100-500 mg L⁻¹, food wastewater COD: ~8000 mg
353 L⁻¹).^{53,54} Nitrogen consumption range of 0-1000 mg L⁻¹ in MECs was selected as ammonium
354 concentration could reach ~1000 mg L⁻¹ in certain types of wastewater (e.g. reject water).⁵⁵ The
355 positive linear correlation of nitrogen and COD concentrations in the integrated MFC-MEC was
356 different from that of conventional BNR where nitrogen removal deteriorated at high COD
357 concentration due to the dominance of heterotrophic bacteria over autotrophic nitrifying
358 bacteria.¹ In the meantime, sufficient carbon source is needed for denitrification in conventional

359 BNR.¹ Thereby, BNR has a strict requirement for COD concentration. The integrated MFC-
360 MEC substantially overcame this requirement, since COD was consumed in MFCs to provide
361 electric power for MECs, and nitrogen removal in MECs do not solely rely on slow growing
362 anammox bacteria, which greatly reduced the dependence on COD concentrations. With more
363 COD being consumed in MFCs, higher electric power was supplied to MECs for accelerated
364 anammox and autotrophic denitrification.

365 A reliable interface between MFC and MEC should be built using power management system
366 (PMS) in order to provide the stable voltage of MFCs to MECs.^{22,56} In a PMS, the current flew
367 through the MFC to charge a capacitor to a given charge potential value (V_c : 0.5 V in this study),
368 and then the system started discharging the stored energy to MEC until the capacitor reached a
369 discharge potential value (V_d : 0 V in this study). Through these charging/discharging cycles, the
370 PMS could provide more stable voltage and power from MFCs to MEC than the direct
371 connection of MFCs to MEC. The charge and discharge curve of the PMS capacitor was
372 simulated (Fig. 6b). With the initial COD concentration of 2000 mg L⁻¹ in the batch-mode
373 SCMFCs, stable charging/discharging cycles lasted for 4 days with a frequency of 0.167 mHz
374 (estimated based on previous research⁵⁶). As the COD was consumed in the batch-mode SCMFC
375 over time, the voltage provided from MFC started to drop after 4 days when COD was below
376 500 mg L⁻¹. The frequency slowed down to 0.030 mHz at the end of 5th day. With the PMS to
377 connect MFCs and MECs, the drop of power output will be alleviated by adjusting the charge/
378 discharge frequency.

379

380 **Energy savings of high-rate anammox in the integrated MFC-MEC system**

381 The operational costs of the integrated MFC-MEC system and conventional BNR were
382 compared in terms of energy and carbon source. The electric power consumption in conventional
383 BNRs was around 0.505 kWh m⁻³ with aeration in complete nitrification as the main electricity
384 cost.⁵⁷ In contrast, anammox in the integrated MFC-MEC system only needs short-cut
385 nitrification for nitrite production, so that one third of electric power (~0.189 kWh m⁻³) was
386 consumed compared with traditional BNR (Table 2). Furthermore, nitrite could be produced
387 without energy consumption. For example, nitrite can be produced in MFCs with oxygen as the
388 electron acceptor in the cathode ($2\text{NH}_4^+ + 3\text{O}_2 \rightarrow 2\text{NO}_2^- + 2\text{H}_2\text{O} + 4\text{H}^+$).¹⁵ Nitrite accumulated in
389 autotrophic denitrification MFCs without aeration ($\text{NO}_3^- + 2\text{H}^+ + 2\text{e}^- \rightarrow \text{NO}_2^- + \text{H}_2\text{O}$, $2\text{NO}_2^- + 8\text{H}^+ + 6\text{e}^-$
390 $\rightarrow \text{N}_2 + 4\text{H}_2\text{O}$) can be used for anammox.³¹

391 Another saving of anammox compared with conventional denitrification is the carbon source,
392 since anammox microorganisms were autotrophic.² Nitrogen removal in MECs did not require
393 organic carbon, while in the conventional BNR, carbon source was required for heterotrophic
394 denitrification, which was about 3.5-5.0 of COD NO₃-N⁻¹ (Table 2). Moreover, anammox was
395 expedited in MECs powered by two series-connected single-chamber MFCs (SCMFCs) in this
396 study (Fig. 1). One MFC produced the maximum power density of 417 mW m⁻² with the
397 maximum voltage output of 0.3 V at the external resistance (R_{ext.}) of 800 Ω. The series
398 connection produced the steady voltage of ~0.50 V for the MEC and provided the energy of 0.28
399 kWh m⁻³ for nitrogen removal over the 10-day period. Therefore, the integrated MFC-MEC
400 system is capable of accelerating anammox with the power produced from wastewater, and
401 achieving self-sustained high-rate nitrogen removal at low energy and carbon cost.

402 Please add Table 2 here

403

404 **Significance of accelerating anammox in the integrated MFC-MEC system for wastewater**
405 **treatment plants**

406 Low growth rate of anammox bacteria has posed a major obstacle for the acceptance of
407 anammox in wastewater treatment plants. An innovative solution as explored in this study by
408 expediting anammox in the integrated MFC-MEC system without solely relying on traditional
409 anammox process, and thus expanding anammox applications. The simultaneous removal of
410 ammonium and nitrite under anoxic condition clearly indicated the occurrence of anammox in
411 MECs, given the previous anammox MFCs studies shared the similar inoculation/operational
412 protocols, had the same ammonium/nitrite trend as this study and confirmed the existence of
413 anammox bacteria.³⁹⁻⁴² By supporting MECs with the electric power generated in MFCs treating
414 wastewater, the integrated system well utilized the unique feature of MFCs (long-term power
415 production stability but low power output) and achieved the self-sustained anammox with higher
416 nitrogen removal rate ($8.2 \text{ mg-N L}^{-1} \text{ d}^{-1}$) than conventional anammox ($6.8 \text{ mg-N L}^{-1} \text{ d}^{-1}$).
417 Although the reaction rate was limited in the batch-mode system compared with conventional
418 continuous flow BNRs (μ_m value, Table 2), the integrated MFC-MEC system was more energy
419 efficient and required much less carbon sources than conventional BNRs.

420 **Conclusion**

421 Novel integrated MEC-MFC was developed to accelerate anammox without external power
422 consumption and alleviate the dependence on slow growing anammox bacteria. Batch-mode tests
423 clearly showed more than 85 % of nitrogen was removed in the anammox MEC within 10 days
424 while only 62 % of nitrogen was removed in conventional anammox. The simulation of the

425 modified Nernst-Monod model revealed the enhancement of half-velocity constant (K_s) and the
426 maximum specific utilization rate (μ_m) in the anammox MFC over conventional anammox. The
427 power supply of MECs intrigued the electron transfer from ammonium to anode, leading to a
428 higher anammox rate. MFCs as the power supply to MECs achieved the self-sustained
429 wastewater treatment, and well applied the low power output of MFCs. By saving energy
430 consumption, avoiding carbon requirement, and accelerating anammox in a single unit, the
431 integrated MFC-MEC possess a great potential for self-sustained advanced nitrogen removal in
432 wastewater treatment plants.

433

434 **Acknowledgement**

435 The graduate student (Yan Li) is supported by the China Scholarship Council (CSC) Doctorate
436 Program.

437

438 Reference

- 439 1 G. Tchobanoglous, F.L. Burton and H.D. Stensel, *Wastewater engineering, treatment and reuse*,
440 4th ed, Metcalf&Eddy, New York, 2003.
- 441 2 M. Strous, E.V. Gerven, P. Zheng, J.G. Kuenen and M.S.M. Jetten, *Water Res.*, 1997, 31, 1955-
442 1962.
- 443 3 S.Q. Ni and J. Zhang, *BioMed. Res. Int.*, 2013, Article ID 469360, 10 p, doi:
444 10.1155/2013/469360.
- 445 4 R.A. Rozendal, H.V. Hamelers and C.J. Buisman, *Environ. Sci. Technol.*, 2006, 40, 5206-5211.
- 446 5 D. Pant, G. V. Bogaert, L. Diels and K. Vanbroekhoven, *Bioresource Tech.*, 2010, 101, 1533-
447 1543.
- 448 6 U. Schröder, F. Harnisch and L.T. Angenent, *Energy Environ. Sci.*, 2015, 8, 513-519.
- 449 7 M.R. Arredondo, P. Kuntke, A.W. Jeremiasse, T.H.J.A. Sleutels, C.J.N. Buisman and A. ter
450 Heijne, *Environ. Sci.: Water Res. Technol.*, 2015, 1, 22-33.
- 451 8 B. Xu, Z. Ge and Z. He, *Environ. Sci.: Water Res. Technol.*, 2015, 1, 279-284.
- 452 9 C. Santoro, I. Ieropoulos, J. Greenman, P. Cristiani, T. Vadas, A. Mackay and B. Li, *J. Power*
453 *Sources*, 2013, 238, 190-196.
- 454 10 C. Santoro, A. Stadlhofer, V. Hacker, G. Squadrito, U. Schröder and B. Li, *J. Power Sources*,
455 2013, 243, 499-507.
- 456 11 Y. Li, Y. Wu, S. Puranik, Y. Lei, T. Vadas and B. Li, *J. Power Sources*, 2014, 269, 430-439.
- 457 12 B.E. Logan, B. Hamelers, R. Rozendal, U. Schröder, J. Keller, S. Freguia, P. Aelterman, W.
458 Verstraete and K. Rabaey, *Environ. Sci. Technol.*, 2006, 40, 5181-5192.
- 459 13 Y. Li, Y. Wu, B. Liu, H. Luan, T. Vadas, W. Guo, J. Ding and B. Li, *Bioresource Technol.*, 2015,
460 192, 238-246.
- 461 14 F. Zhang and Z. He, *J. Chem. Technol. Biotechnol.*, 2012, 87, 153-159.
- 462 15 B. Viridis, K. Rabaey, R.A. Rozendal, Z. Yuan and J. Keller, *Water Res.*, 2010, 44, 2970-2980.
- 463 16 Y. Li, I. Williams, Z. Xu, B. Li and B. Li, *Appl. Energy*, 2016, 163, 352-360.
- 464 17 G. Zhan, L. Zhang, D. Li, W. Su, Y. Tao and J. Qian, *Bioresource Technol.*, 2012, 116, 271-277.
- 465 18 L. Hang, X. Chai, G. Chen, B.E. Logan, *Environ. Sci. Technol.*, 2011, 45, 5025-5031.
- 466 19 Z. He, J. Kan, Y. Wang, Y. Huang, F. Mansfield and K.H. Neilson, *Environ. Sci. Technol.*, 2009,
467 43, 3391-3397.
- 468 20 X. Xie, M. Ye, L. Hu, N. Liu, J.R. McDonough, W. Chen, H.N. Alshareef, C.S. Criddle and Y.
469 Cui, *Energy Environ. Sci.*, 2012, 5, 5265-5270.
- 470 21 Y. Chen, Z. Lv, J. Xu, D. Peng, Y. Liu, J. Chen, X. Sun, C. Feng and C. Wei, *J. Power Sources*
471 2012, 201, 136-141.
- 472 22 B. Liu, A. Weinstein, M. Kolln, C. Garrett, L. Wang, A. Bagtzoglou, U. Karra, Y. Li and B. Li, *J.*
473 *Power Sources*, 2015, 286, 210-216
- 474 23 Z. Xu, B. Liu, Q. Dong, Y. Lei, Y. Li, J. Ren, J. McCutcheon and B. Li, *Bioresource Technol.*,
475 2015, 197, 244-251.
- 476 24 M.C. Hatzell, Y. Kim and B.E. Logan BE, *J. Power Sources*, 2013, 229, 198-202.
- 477 25 M.E. Nielsen, C.E Reimers, H.K. White, S. Sharma and P.R. Girguis, *Energy Environ. Sci.*, 2008,
478 1, 584-593.
- 479 26 S.Q. Ni, S. Sung, Q.Y. Yue and B.Y. Gao, *Ecol. Eng.*, 2012, 38, 30-36.
- 480 27 M. Strous, J.J. Heijnen, J.G. Kuenen and M.S.M Jetten, *Appl. Microbiol. Biotechnol.*, 1998, 50,
481 589-596.
- 482 28 T. Chen, P. Zheng, L. Shen, S. Ding and Q. Mahmood, *J. Hazard Mater.*, 2011, 190, 28-35.
- 483 29 D. Puyol, J.M. Carvajal-Arroyo, B. Garcia, R.S. Alvarez and J.A. Field, *Bioresource Technol.*,
484 2013, 139, 94-100.
- 485 30 A.K. Marcus, C.I. Torres and B.E. Rittmann, *Biotechnol. Bioeng.*, 2007, 98, 1171-1182.
- 486 31 V. Srinivasan, J. Weinrich and C. Butler, *Environ. Sci.: Water Res. Technol.*, 2016, 2, 344-352.
- 487 32 H. Liu, S. Cheng and B.E. Logan, *Environ. Sci. Technol.*, 2005, 39, 658-662.

- 488 33 B. Virdis, K. Rabaey and J. Yuan, *Water Res.*, 2008, 42, 3013-3024.
489 34 S. Xie, P. Liang, Y. Chen, X. Xia and X. Huang, *Bioresource Technol.*, 2011, 102, 348-354.
490 35 L.K. Wang, N.C. Pereira, Y.T. Huang and N.K. Shammas, *Biological treatment processes*, vol.8.,
491 Humana Press, New York, 2009.
492 36 M.S.M Jetten, M. Strous, K.T. van de Pas-Schoonen, J. Schalk, U.G.J.M. van Dongen, A.A. van
493 de Graaf, S. Logemann, G. Muyzer, M.C.M van Loosdrecht and J.G. Kuenen, *FEMS Microbiol.*
494 *Rev.*, 1999, 22, 421-437.
495 37 R. Zhao, H. Zhang, Y. Li, T. Jiang and F. Yang, *Curr. Microbiol.*, 2014, 69, 880-887.
496 38 M. Oshiki, I.K. Yoshida, N. Fujii, M. Ishiguro, H. Satoh and S. Okabe, *Appl. Environ. Microbiol.*,
497 2013, 79, 4087-4093.
498 39 Y. Lee, L. Martin, P. Grasel, K. Tawfiq and G. Chen, *Environ Technol.*, 2013, 34, 2727-2736.
499 40 E.G. Di Domenico, G. Petroni, D. Mancini, A. Geri, L. Di Palma and F. Ascenzioni, *BioMed. Res.*
500 *Int.*, 2015, Article ID 351014, 10 p, doi: 10.1155/2015/351014.
501 41 C. Li, H. Ren, M. Xu and J. Cao, *Bioresource Technol.*, 2015, 175, 454-552.
502 42 D.A. Jadhav and M.M. Ghangrekar, *Environ Technol.*, 2015, 36, 767-775.
503 43 M.K.H. Winkler, R. Kleerebezem and M.C.M. van Loosdrecht, *Water Res.*, 2012, 46, 136-144.
504 44 T. Yamamoto, K. Takaki, T. Koyama and K. Furukawa, *Bioresource Technol.*, 2008, 99, 6419-
505 6425.
506 45 X. Xu, Y. Xue, D. Wang, G. Wang and F. Yang, *Bioresource Technol.*, 2014, 155, 427-431.
507 46 E.M. Gilbert, S. Agrawal, S. M. Karst, H. Horn, P.H. Nelsen and S. Lackner, *Environ. Sci.*
508 *Technol.*, 2014, 48, 8784-8792.
509 47 C. Tang, P. Zheng, C. Wang, Q. Mahmood, J. Zhang, X. Chen, L. Zhang and J. Chen, *Water Res.*,
510 2011, 45, 135-144.
511 48 W.R.L. van der Star, W.R. Abma, D. Blommers, J. Mulder, T. Tokutomi, M. Strous, C.
512 Picioreanu and M.C.M. van Loosdrecht, *Water Res.*, 2007, 41, 4149-4163.
513 49 M. Waki, T. Yasuda, K. Suzuki, T. Sakai, N. Suzuki, R. Suzuki, K. Matsuba, H. Yokoyama, A.
514 Ogino, Y. Tanaka, S. Ueda, M. Takeuchi, T. Yamagishi and Y. Suwa, *Bioresource Technol.*,
515 2010, 101, 2685-2690.
516 50 B. Kartal, M. Koleva, R. Arsov, W. van der Star, M.S.M. Jetten and M. Strous, *J. Biotechnol.*,
517 2006, 126, 546-553.
518 51 K. Egli, U. Fanger, P.J.J. Alvarez, H. Siegrist, J.R. van der Meer and A.J.B. Zehnder, *Arch*
519 *Microbiol.*, 2001, 175, 198-207.
520 52 B. Li and S. Irvin, *Biochem. Eng. J.*, 2007, 34, 248-255.
521 53 J. Ditzig, H. Liu and B.E. Logan, *Water Res.*, 2007, 32, 2296-2304.
522 54 S.E. Oh and B.E. Logan, *Water Res.*, 2005, 39, 4673-4682.
523 55 P.T. Kelly and Z. He, *Bioresource Technol.*, 2014, 153, 351-360.
524 56 A. Dewan, C. Donovan, D. Heo and H. Beyenal, *J. Power Sources*, 2010, 195, 90-96.
525 57 R. Goldstein and W. Smith, *Water & sustainability (volume 4): U.S. electricity consumption for*
526 *water supply & treatment-the next half century*, Electric power research institute, Inc., Palo Alto,
527 2002.
528
529
530

531 **Caption list of tables and figures.**

532

533 **Table 1.** Values of kinetic parameters derived from kinetics models (Eqs.1-4).

534 **Table 2.** Comparison of anammox MEC, conventional anammox and BNR regarding energy
535 consumption and saving.

536 **Fig. 1** Diagram of the integrated MFC-MES system and Control setup.

537

538 **Fig. 2** Variation of nitrogen concentration in the MEC anode over time.

539 **Fig. 3** Nitrogen removal rates in anammox MEC and Control tests.

540 **Fig. 4** Variation of total nitrogen, ammonium and nitrite concentrations in anammox MEC and
541 conventional anammox (Control 1) over time (**a**: ammonium and nitrite concentrations in anammox MEC
542 based on model simulation. **b**: experimental results of total nitrogen concentration. **c**: ammonium and
543 nitrite concentrations in conventional anammox (Control 1) based on model simulation. The inserted
544 figure is the difference between ammonium and nitrite observed data over time).

545 **Fig. 5** ORP (a) and pH (b) changes in anammox MEC and Controls before and after of experiments.

546 **Fig. 6** Interface simulation between MFC and MEC (a: Correlation between COD consumption in MFCs
547 and nitrogen consumption in MECs. b: the charging/discharging cycles of PMS).

548

549

550 **Table 1.** Values of kinetic parameters derived from kinetics models (Eqs.1-4).

Symbol	Description	Value	Unit
$K_{SNH4+ MEC}$	half-velocity constant of ammonium in anammox MEC	4.11	mM
$K_{SNO2- MEC}$	half-velocity constant of nitrite in anammox MEC	2.33	mM
$K_{SNH4+ ana}$	half-velocity constant of ammonium in Control 1	3.47	mM
$K_{SNO2- ana}$	half-velocity constant of nitrite in Control 1	3.4	mM
$\mu_{mNH4+ MEC}$	maximum specific utilization rate of ammonium in anammox MEC	0.38	mmol g ⁻¹ VSS h ⁻¹
$\mu_{mNO2- MEC}$	maximum specific utilization rate of nitrite in anammox MEC	0.3	mmol g ⁻¹ VSS h ⁻¹
$\mu_{mNH4+ ana}$	maximum specific utilization rate of ammonium in Control 1	0.18	mmol g ⁻¹ VSS h ⁻¹
$\mu_{mNO2- ana}$	maximum specific utilization rate of nitrite in Control 1	0.2	mmol g ⁻¹ VSS h ⁻¹

551

552

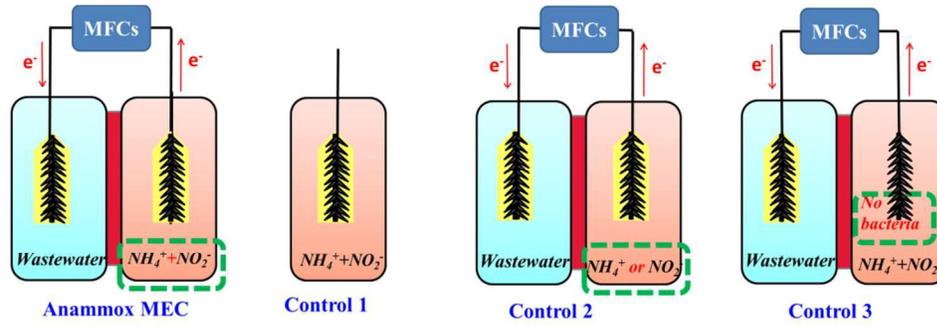
553 **Table 2.** Comparison of anammox MEC, conventional anammox and BNR regarding energy
 554 consumption and saving.

	Nitrification and denitrification in conventional BNR⁵⁷	Conventional anammox²⁶⁻²⁹	Anammox MEC powered by MFC
Oxygen consumption (mole per mole of NH ₄ ⁺)	2	0.75	0.75
Energy production (kWh m ⁻³)	–	–	0.28
Energy consumption (kWh m ⁻³)	0.505	0.19	0.19
Energy saving (kWh m ⁻³)	-0.505	-0.19	+0.09
COD requirement (mg L ⁻¹ per mg-N L ⁻¹)	3.5-5	–	–
μ_m (mmol g ⁻¹ VSS h ⁻¹)	1.21-9.86	0.09-3.74 (0.18-0.2 in Control 1, this study)	0.3-0.38

555

556

557



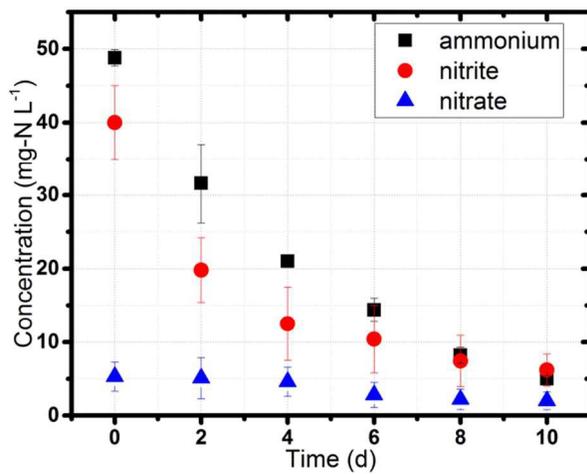
558

559

Fig. 1 Diagram of the integrated MFC-MES system and Control setup.

560

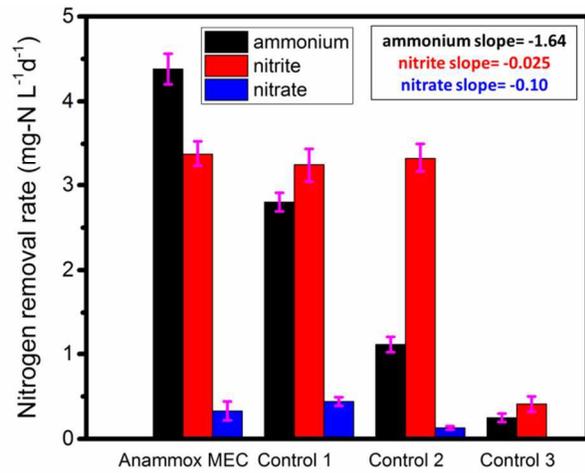
561



562

563 **Fig. 2** Variation of nitrogen concentration in the MEC anode over time.

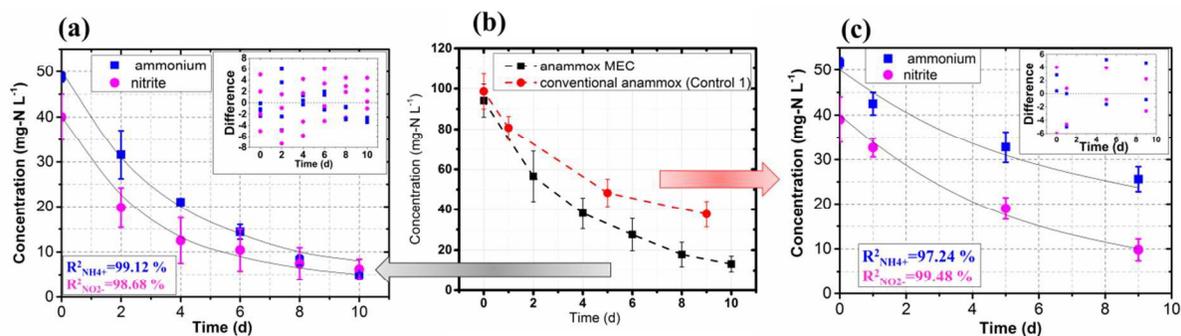
564



565

566 **Fig. 3** Nitrogen removal rates in anammox MEC and Control tests (Inserted values are the slopes of the
567 linear regression of average removal rates).

568

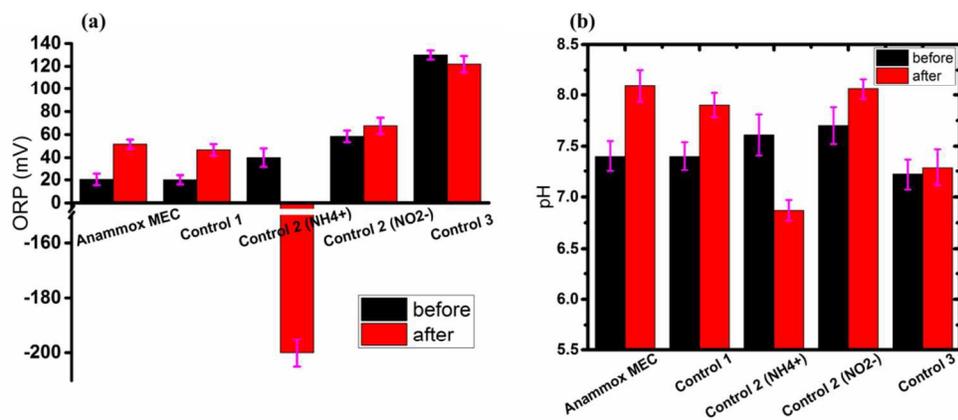


569
 570 **Fig. 4** Variation of total nitrogen, ammonium and nitrite concentrations in anammox MEC and
 571 conventional anammox (Control 1) over time (**a**: ammonium and nitrite concentrations in anammox MEC
 572 based on model simulation. **b**: experimental results of total nitrogen concentration. **c**: ammonium and
 573 nitrite concentrations in conventional anammox (Control 1) based on model simulation. The inserted
 574 figure is the difference between ammonium and nitrite observed data over time).

575

576

577

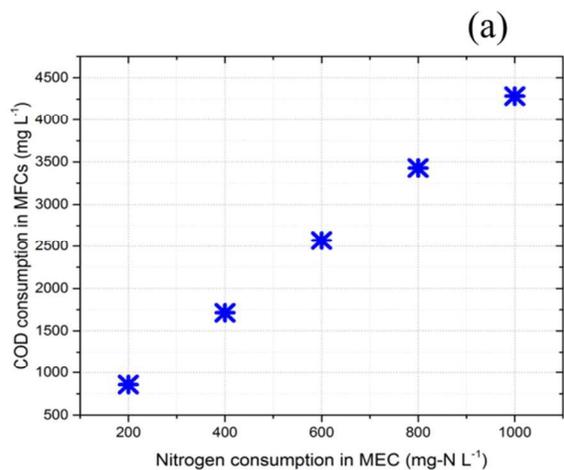


578

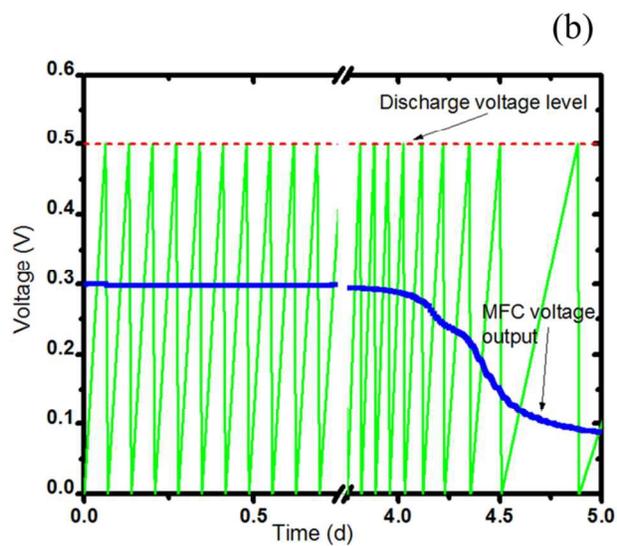
579 **Fig. 5** ORP (a) and pH (b) changes in anammox MEC and Controls before and after of experiments.

580

581



582



583

584 **Fig. 6** Interface simulation between MFC and MEC (a: Correlation between COD consumption in MFCs
585 and nitrogen consumption in MECs. b: the charging/discharging cycles of PMS).