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Citrus flavanones prevent systemic inflammation and ameliorate oxidative stress in C57BL/6J mice fed high-fat diet

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Abbreviation list:

- IL-6: interleukin-6
- IL-10: interleukin-10
- TNF- α : tumor necrosis factor- α
- MCP-1: macrophage chemoattractant protein-1
- hs-CRP: high-sensitivity C-reactive protein
- TBARS: thiobarbituric acid reactive substance
- CVD: cardiovascular disease
- NF-κB: nuclear factor kappa-B
- COX-2: cycloxigenase-2
- ALT: alanine transaminase
- AST: aspartate transaminase
- ABTS*+: 2,2'-azino-di-(3-ethylbenzothiazoline)-6-sulfonic acid
- TEAC: Trolox equivalent antioxidant capacity
- MDA: malondialdehyde
- LDLR: LDL receptor

1 ABSTRACT

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2 The flavanones hesperidin, eriocitrin and eriodictyol were investigated for their prevention of the oxidative stress and systemic inflammation caused by high-fat diet in C57BL/6J mice. The mice 3 received a standard diet (9.5% kcal from fat), high-fat diet (45% kcal from fat) or high-fat diet 4 supplemented with hesperidin, eriocitrin or eriodictyol for a period of four weeks. Hesperidin, 5 eriocitrin and eriodictyol increased the serum total antioxidant capacity, and restrained the 6 elevation of interleukin-6 (IL-6), macrophage chemoattractant protein-1 (MCP-1), and C-7 8 reactive protein (hs-CRP). In addition, the liver TBARS levels and spleen mass (g/kg body 9 weight) were lower for the flavanone-treated mice than in the unsupplemented mice. Eriocitrin and eriodictyol reduced TBARS levels in the blood serum, and hesperidin and eriodictyol also 10 11 reduced fat accumulation and liver damage. The results showed that hesperidin, eriocitrin and eriodictyol had protective effects against inflammation and oxidative stress caused by high-fat 12 13 diet in mice, and may therefore prevent metabolic alterations associated with the development of cardiovascular diseases in other animals. 14 Keywords: Citrus flavonoids; high-fat diet; obesity; inflammation; oxidative stress; C57BL/6J 15

16 mice

17 Introduction

Prospective studies show that dietary patterns can modify risks for coronary disease. 18 High-fat diets combined with excess body weight lead to adverse metabolic outcomes, and 19 according to the Framingham Heart Study,¹ the risk for cardiovascular disease (CVD) is 20 21 particularly increased when abdominal obesity is present. Furthermore, obesity is a major modifiable risk factor that can lead to dyslipidemia, type 2 diabetes and hypertension.²In 22 23 contrast, the consumption of diets rich in fruits, vegetables and unsaturated fatty acids are associated with a lowering of CVD risk, due to their antioxidant nutrients and bioactive 24 25 compounds.

Adipose tissue accumulation in mice fed high-fat diets increases migration of 26 macrophages, leading to the stimulation of free radical production and secretion of inflammatory 27 cytokines.^{3,4}This leads to lipotoxicity in non-fat tissues, causing structural and functional 28 changes in cells, and stimulation of further release of cytokines and chemokines, ultimately 29 resulting in chronic systemic low-grade inflammation.⁵High levels of fat in obese rodents 30 31 increase systemic levels of interleukin-6 (IL-6), tumor necrosis factor- α (TNF- α), and monocyte chemoattractant protein-1 (MCP-1), which are related to the development of liver steatosis and 32 insulin resistance.^{6,7} 33

Hesperidin and eriocitrin are flavanone glycosides from oranges and limes, and are 34 deglycosylated by intestinal bacteria to hesperetin and eriodictvol, respectively.⁸Subsequently 35 36 they are conjugated to glucuronides and sulfates in enterocytes and hepatocytes, both yielding homoeriodictyol and hesperetin conjugated metabolites.⁹Thesemetabolites have antioxidant and 37 anti-inflammatory activities capable of scavenging free radicals and inhibiting in vitro 38 inflammation.¹⁰⁻¹⁵Moreover, in diabetic rats eriodictyol attenuated inflammation and decreased 39 nitric oxide, proinflammatory cytokines and plasma lipid peroxidation.¹⁶Hesperetin blocked the 40 activation of nuclear factor kappa-B (NF- κ B) by TNF- α in mice adipocytes, reduced oxidative 41

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| 42 | stress, cycloxigenase-2 (COX-2) expression and production of IL-6 in mice with colon |
|----|--|
| 43 | carcinogenesis. ^{17,18} |
| 44 | These studies suggest, therefore, that such citrus flavanones may lessen chronic low- |
| 45 | grade systemic inflammation in animals fed high-fat diets, and as a result may reduce the |
| 46 | occurrence of diabetes and CVD. To test this, we analyzed the effects of the citrus flavanones |
| 47 | hesperidin, eriocitrin and eriodictyol on the antioxidant capacities in liver and blood serum and |
| 48 | on the systemic inflammation in C57BL/6J mice fed a high-fat diet. |
| 49 | |
| 50 | Materials and methods |
| 51 | Animals and dietary treatment |
| 52 | Sixty nine-week-old male C57BL/6J mice (São Paulo State University, SP, Brazil) were |
| 53 | maintained in an isolated macro-environment system, with a 12-h light/12-h dark cycle, and 22 \pm |
| 54 | 2 °C, receiving food and water ad libitum. The animals was maintained individually in |
| 55 | conventional housing inside of the ventilated storage cabinets (Tecniplast, S.A., Buguggiatte, |
| 56 | VA, Italy). They were randomly divided into six groups, with similar body weight distributions |
| 57 | in each group, ten mice per group, and were fed either a standard diet, a high-fat diet, or a high- |
| 58 | fat diet supplemented with ibuprofen, hesperidin, eriocitrin, or eriodictyol. Mice were allocated |
| 59 | one per cage, and the body weight was used for the dosage of the supplements, offered in mg/kg |
| 60 | body weight. All supplements were mixed into the diet, based on the food intake of the previous |
| 61 | day (grams of food ingested/day) with an additional of 10% to ensure the intake of the daily |
| 62 | dose. The food intake was monitored at regular intervals of 24 hours, and the supplements were |
| 63 | adjusted accordingly. Body weights were recorded weekly and food consumption daily. The |
| 64 | high-fat diet contained 21, 34 and 45% energy from protein, carbohydrates and fat, respectively |
| 65 | (Rhoster Industry and Trade LTD, SP, Brazil). The control diet contained 15, 76 and 10% energy |
| 66 | from protein, carbohydrates, and lipids, respectively (Table 1), based on the AIN93 M |
| 67 | diet. ¹⁹ After four weeks of treatment, mice were anesthetized with xylazine/ketamine (16/60 mg/g |

of body weight) via i.p. injection following a 10-h fast, and blood samples were drawn by 68 cardiac puncture. Organs and serum samples were stored at -80°C until analysis. One lobe of 69 each liver sample was fixed in 10% buffered formalin for 48h, rinsed with distilled water and 70 71 soak in 70% alcohol solution before perform histological analysis by a pathologist. All the animals were handled according to the guidelines of the Brazilian College of Animal 72 73 Experimentation (COBEA) and the experimental animal protocol was approved by the Animal Use Committee of the Pharmaceutical Sciences Faculty, São Paulo State University, SP, Brazil 74 $(n^{\circ} 18/2013).$ 75

76

77 Supplementation

78 Hesperidin supplements were given at a dose of 100 mg/kg body weight, eriocitrin and eriodictyol at 200 mg/kg body weight, and ibuprofen at 20 mg/kg body weight (included as a 79 80 positive control for anti-inflammatory activity). Doses were selected on the basis of scientific 81 literature that have shown the hesperidin effectiveness to reduce inflammation and oxidative 82 stress in the rodent models.20, 21, 22 Furthermore, supplements were added to the regular diet of 83 the animals to favor the normal physiological pathway of intake and, were offered regularly to maintain a constant level of the compound in the body. Hesperedin purity was > 98 %, and was 84 extracted from the fruit peel of Citrus sinensis (Rutaceae). Eriocitrin purity was >85% and eri-85 odictyol was > 95%, and both were extracted from the fruit peel of *Citrus limon (Rutaceae)*. 86

87

88 Blood serum analyses

Serum was obtained by centrifugation and levels of alanine transaminase (ALT), aspartate
transaminase (AST), glucose, triglycerides, total cholesterol and HDL-C were evaluated by
enzymatic colorimetric assay using commercial kits (Labtest, MG, Brazil). LDL-C was
calculated using the Friedewald formula.²³ High-sensitivity C-reactive protein (hs-CRP) was
measured by immunoturbidimetric assay with commercial kits (Labtest, MG, Brazil), and

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| 94 | inflammatory cytokines assays (IL-6, IL-10, TNF- α , and MCP-1) were performed by ELISA |
|-----|--|
| 95 | assays using the Multiplex LuminexMAP detection method (Genese Diagnostics Products Ltd, |
| 96 | SP, Brazil). |
| 97 | |
| 98 | Organs and liver histology |
| 99 | Organ weights (visceral adipose tissue, liver, spleen and heart) were normalized against |
| 100 | body weight of the respective animal (organ weight/body weight) to give a relative percentage. |
| 101 | Liver tissues fixed in formalin were embedded in paraffin and sectioned to 4–6 mm of thickness. |
| 102 | Deparaffinized liver tissue sections were stained with hematoxylin-eosin and Masson's |
| 103 | trichrome, using standardized protocols (Pathology Department of Odontology Faculty from Sao |
| 104 | Paulo State University, SP, Brazil). A pathologist analyzed all treated mice hepatic cells by |
| 105 | optical microscopy to recognize any morphologic alteration in comparison to control group. |
| 106 | |
| 107 | Oxidative stress parameters |
| 108 | Liver and serum oxidative stress were measured by thiobarbituric acid-reactive |
| 109 | substances (TBARS). Liver TBARS values were given in μ M MDA/mg protein, quantified by |
| 110 | Lowry method, $^{24}\!and$ serum TBARS in μM MDA. Total antioxidant activity was measure by the |
| 111 | ABTS assay. ²⁵ ABTS ^{*+} radical formation was measured at 734 nm, using Trolox (Sigma) as a |
| 112 | standard and given as mM of Trolox equivalent antioxidant capacity (TEAC). All analyzes were |
| 113 | performed in triplicate. |
| 114 | |
| 115 | Statistical analysis |
| 116 | All results are presented as means ± standard deviation. The data distributions were tested |
| 117 | for normality, and subsequently, the intergroup variation was measured by one-way ANOVA |
| 118 | followed by post hoc analysis (Student Newman Keuls test) to evaluate the effects of diets and/or |

supplement, with a significance level of p < 0.05 (Sigma Stat Software, USA).

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120

121 Results

122 Diet- and supplement-induced changes in body weight and organs

123 The high-fat diet with or without the flavanone supplements showed good acceptance by 124 the animals without harmful effects, and the supplement daily doses reached 97% on average of 125 the calculated dose. Groups fed the high-fat diet with or without supplements showed higher 126 body weight gain, with exception of the hesperidin group, and higher visceral fat compared to 127 the mice fed the standard diet (Table 2). Animals fed the high-fat diet supplemented with 128 hesperidin, eriocitrin or eriodictyol showed lower spleen mass (g/g of body weight) than the 129 unsupplemented high-fat diet group; the spleen mass of the ibuprofen and standard diet groups 130 was intermediate; and the treatment with the unsupplemented high-fat diet increased the heart mass, while eriocitrin showed an intermediate value and hesperidin, eriodictyol and ibuprofen 131 132 showed the lowest heart masses (Table 2).

133

134 **Biochemical profile**

135 After four weeks with the high-fat diet, mice blood glucose levels were elevated 20% with the unsupplemented high-fat diet, while the increases were only approximately 8% for the groups 136 137 fed the high-fat diet supplemented with hesperidin, eriocitrin, eriodictyol and ibuprofen, however 138 this effect was not statistically significant. Furthermore, no effect was observed from the high-fat 139 diet with ibuprofen or citrus flavanones on bloodserum triglycerides compared to the non-140 supplemented high-fat diet. Total cholesterol, HDL-C and LDL-C were similarly increased by 141 the high-fat diet even with the supplements, where only hesperidin was able to lower the LDL-C 142 levels by 28%. Serum hepatic enzymes ALT and AST were not altered by the high-fat diet alone 143 or with flavanones or ibuprofen (Table 3).

144

145 Anti-oxidative stress and anti-inflammatory effects

9

| 146 | Unsupplemented high-fat diet increased TBARS levels by 75% in the liver and 25% in the blood |
|-----|--|
| 147 | serum in comparison to standard diet. In contrast, all supplements (ibuprofen, hesperidin, |
| 148 | eriocitrin and eriodictyol) were able to maintain liver TBARS at the levels of the standard diet |
| 149 | group. Blood serum TBARS, after ibuprofen and hesperidin, were not different to the values |
| 150 | observed with the standard diet, but they were lowered 34% by the eriocitrin and eriodictyol |
| 151 | supplements. Furthermore, total antioxidant capacity measured by the reduction of $ABTS^{*+}$ |
| 152 | radical in the blood serum was significantly lowered by the high-fat diet, and ibuprofen was able |
| 153 | to maintain the same level as with the standard diet. Hesperidin, eriocitrin and eriodictyol |
| 154 | slightly increased an average of 3.5% the blood serum antioxidant capacity (Table 4). |
| 155 | After four weeks with high-fat diet, IL-6 and MCP-1 increased by 7.6 and 3.2 foldin the |
| 156 | blood serum of mice, showing the effect of the chronic dietary treatment over these |
| 157 | inflammatory markers. Ibuprofen and all flavanones were able to decreaseIL-6to the standard |
| 158 | diet level, without any significant differences among them.MCP-1production was similarly |
| 159 | inhibited by these compounds in comparison to the high-fat diet group. No changes were |
| 160 | observed in serum levels of TNF- α and IL-10 for any of the studied groups. Finally, hs-CRP |
| 161 | levels were increased by 13% with the high-fat diet, while hesperidin, eriocitrin and eriodictyol |
| 162 | supplemented groups were able to lower the hs-CRP by 25%, 25%, and 13%, respectively |
| 163 | compared to the standard and unsupplemented high-fat diet groups(Table 4). |
| 164 | |
| 165 | Liver histology |
| 166 | Liver sections from mice fed the standard diet showed typical morphology with normal |
| 167 | microvesicular depots of fat (Figure 1, A), unlike the mice fed the high-fat diet that showed |
| 168 | largely macrovesicular fat depots (Figure 1, B). Hepatocytes from these mice also showed |
| | |

Liver sections from mice fed the standard diet showed typical morphology with normal microvesicular depots of fat (Figure 1, A), unlike the mice fed the high-fat diet that showed largely macrovesicular fat depots (Figure 1, B). Hepatocytes from these mice also showed morphologic alterations, as peripheral nuclei (Figure 1, B-1), undefined contours or even fragmented cells (Figure 1, B-2) characteristic of cellular necrosis, and some cells evidenced cellular ballooning (Figure 1, B-3). Although the high-fat diet had high potential to induce

steatosis, the ibuprofen and the citrus flavanone hesperidin did not show cellular alterations and
reduced fat depots (Figure 1, C and D). No evidence of cellular ballooning or cell degradation in
the liver parenchyma was observed for the mice fed the high-fat diet supplemented with the
flavanones and ibuprofen. However, liver sections of mice supplemented with eriocitrin (Figure
1, E) showed histological features similar to the unsupplemented high-fat diet fed mice.

177

178 Discussion

The citrus flavanones hesperidin, eriocitrin and eriodictyol were able to prevent key 179 metabolic changes induced by high-fat diet consumption, specifically: (1) increased blood serum 180 levels of IL-6 and MCP-1; (2) elevated TBARS levels in liver and blood serum; (3) liver lipid 181 182 accumulation and liver damage; (4) decreased antioxidant capacity in the blood serum; and (5) higher mass of heart and spleen. Among the flavanones, only hesperidin showed LDL-C 183 lowering activity, decreasing its level by 28%, an effect that has been reported by others.^{26,27}In 184 previous studies this ability was related to the reduction of Apo B secretion and synthesis of 185 VLDL-C, and increased expression of the LDL receptor (LDLR) gene, thereby decreasing the 186 levels of LDL-C circulating.^{28, 29} In this present study, all animals that received the high-fat diet 187 with 4.7 times more calories from fat compared with the standard diet, and supplemented with 188 flavanones or not, showed higher body weight and visceral fat, and higher levels of serum lipids 189 190 and glucose. These deleterious effects of the high-fat diet subsequently led to elevation of key 191 inflammatory markers and increased hepatic lipid deposition.

The presence of hepatic steatosis and a high heart mass, observed in the group fed the unsupplemented high-fat diet, should be a result of the loss of ability to regulate the synthesis and storage of triglycerides in adipose tissue, caused by inflammation.^{30,31} Also, the observed increased spleen mass has been previously shown to be related to the inflammatory stimulus of a high-fat diet.³⁰ However, hesperidin, eriodictyol and ibuprofen efficiently suppressed the heart mass increase, and all of the flavanones prevented spleen mass increase, but ibuprofen did not.

The lipotoxicity in heart tissue can induce inflammation and contribute to its remodeling, as shown previously with transgenic mice, where pro-inflammatory cytokine and macrophage infiltration preceded the onset of myocardial dysfunction.³¹The citrus flavanones and ibuprofen were able to prevent the increased mass of heart and spleen possibly by lowering the oxidative stress and inflammation in these organs, as shown by the suppression of systemic levels of IL-6, MCP-1 and TBARS.

204 With an excessive fat supply, the liver is the organ most susceptible to lipotoxicity, 205 because of the large amounts of fat directly received through the portal vein along with circulating lipids and lipoproteins.³²In the present study, four weeks of the unsupplemented high-206 207 fat diet consumption caused severe variations in hepatic structure, showing morphological 208 changes as ballooning, cellular and nuclear degradation characterizing a process of cell death in the presence of hepatic steatosis, presented as macro and microvesicles of fat. A recent study 209 210 showed that consumption of a high-fat diet by mice could lead to hepatic steatosis in three weeks or less.³³ In comparison, it was previously shown³⁴ that the consumption of a fast-food diet, rich 211 212 in both fat and sugar, revealed the presence of hepatic steatosis with pronounced ballooning and progressive fibrosis, while the consumption of a high-fat diet caused hepatic steatosis without 213 214 other alterations.

Supplementation with hesperidin, eriodictyol and ibuprofen decreased lipid droplets in 215 216 the liver and prevented morphological alterations in hepatocytes of mice fed a high-fat diet, 217 showing the anti-inflammatory action against liver damage. In addition, flavanones significantly reduce the serum levels of C-reactive protein, which is an indicator of liver damage.^{35,36}Along 218 219 with the liver, accumulation of visceral adipose tissue (epididymal, retroperitoneal and perirenal) has been widely used to measure inflammation in humans and rodents obese.³⁷⁻⁴² In 220 general, those metabolic changes induce the elevation of systemic levels of MCP-1, which has 221 been linked to the infiltration of macrophage and inflammatory cells in adipocytes, 222 proportionally to the increase of the adipose tissue.^{4,43,44} 223

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| 224 | In liver and adipose tissue, high fat levels are associated with increased NF-KB activity, |
|-----|---|
| 225 | systemic levels of pro-inflammatory cytokines and acute phase proteins. ⁴⁵ In addition, liver |
| 226 | production of C-reactive protein, stimulated by increased levels of IL-6 and TNF- α , lead to a |
| 227 | systemic inflammatory response. ⁴⁶ As observed in this study, mice fed the unsupplemented high- |
| 228 | fat diet showed an increase in serum levels of both IL-6 and MCP-1, but no changes in serum |
| 229 | levels of IL-10 and TNF- α . In sharp contrast, the supplements hesperidin, eriocitrin, eriodictyol |
| 230 | and ibuprofen dramatically suppressed the high-fat diet stimulated elevation of the |
| 231 | proinflammatory IL-6 and MCP-1 in the serum, leading ultimately to significantly suppress |
| 232 | systemic inflammation in the supplement-treated mice, efficiently defeating pro-inflammatory |
| 233 | response. |
| 234 | The metabolites of the flavanones produced in the mice were not analyzed in this study, |
| 235 | but other previous studies have shown that mammalian metabolism of these compounds initially |
| 236 | produce a series of intact flavanone glucuronides, sulfates, and glucurono-sulfate conjugates |
| 237 | mainly at the 3', 4', and 7 positions of the flavanone B and A rings, respectively. ^{47, 48} Very little, if |
| 238 | any, of the unmodified flavanone aglycones are detected as metabolites. Subsequent to the |
| 239 | production of the glucuronide and sulfate metabolites, further metabolism produces advanced |
| 240 | ring-fission products, including compounds like <i>m</i> -coumaric acid, ferulic acid, 3,4- |
| 241 | dihydroxyphenylpropionic acid, 3-methoxy-4-hydroxyphenylpropionic acid, m-hydroxyhippuric |
| 242 | acid, <i>m</i> -hydroxybenzoic acid, 3-methoxy-4-hydroxybenzoic acid, and others. ⁴⁹ |
| 243 | Research in this field has not advanced to the point of knowing the relative impacts of |
| 244 | these diverse metabolites on the sum-total biological effects observed in the test animals |
| 245 | following the flavanone doses. Yet, it is known that the effects of glucuronide metabolites of |
| 246 | hesperidin are not equal in their effects on cytokine production during inflammation, ⁵⁰ but rather |
| 247 | strong structure/function relationships play roles in biological responses. Also, while |
| 248 | assumptions are made that flavonoids, in general are anti-inflammatory, it is also known that in |
| 249 | some cases flavonoids promote the production of pro-inflammatory cytokines. ⁵¹ Hence, the most |
| | |

| 250 | conclusive evidence of anti-inflammatory actions is that obtained by whole animal studies, while | | | | | | |
|-----|---|--|--|--|--|--|--|
| 251 | in vitro studies point to many possible pathways and outcomes, the selections of which are | | | | | | |
| 252 | difficult to predict. However, the benefits of the supplementation observed in this experimental | | | | | | |
| 253 | model are consistent with the fact that eriodictyol can exert high antioxidant activity due to its o- | | | | | | |
| 254 | dyhydroxyl structure in the B ring, ⁹ and metabolites of eriocitrin and hesperidin increase the | | | | | | |
| 255 | concentration of antioxidant enzymes in vivo. ¹⁰ Eriodictyol and hesperetin can inhibit cellular | | | | | | |
| 256 | oxidative stress and inflammatory damage via regulating antioxidant responses. ^{52, 53} All these | | | | | | |
| 257 | effects are related to lower degree of inflammation and liver damage. | | | | | | |
| 258 | | | | | | | |
| 259 | Conclusion | | | | | | |
| 260 | In conclusion, the present study showed that supplementation with hesperidin, eriocitrin | | | | | | |
| 261 | and eriodictyol was efficiently able to suppress the systemic state of inflammation induced by a | | | | | | |
| 262 | high-fat diet, and to prevent damage to organs such as liver, heart and spleen. Further tests are | | | | | | |
| 263 | planned to test whether these flavanones are also useful in preventing metabolic disorders and | | | | | | |
| 264 | chronic disease, as CVD and diabetes mellitus. | | | | | | |
| 265 | | | | | | | |
| 266 | Conflict of interest | | | | | | |
| 267 | None of the authors have any conflicts of the interest. | | | | | | |
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Figure Legend

Figure 1. Histological sections of liver tissue (40 x magnification) of mice fed standard diet (A), high-fat diet (B), high-fat diet supplemented with ibuprofen (C), hesperidin (D), eriocitrin (E), and eriodictyol (F). Numbers in section B: (1) peripheral nuclei, (2) absence of nucleus and (3) cellular ballooning.

| Composition | G/kg | | | |
|---------------------------|----------|----------|--|--|
| Diet | High-fat | Standard | | |
| Protein (%kcal) | 20.8 | 14.7 | | |
| Carbohydrate (%kcal) | 33.8 | 75.8 | | |
| Fat (% kcal) | 45.4 | 9.5 | | |
| Energy (kcal/g) | 5.4 | 4.2 | | |
| Corn starch | 78 | 466 | | |
| Maltodextrin | 117 | 155 | | |
| Sucrose | 201 | 100 | | |
| Casein | 240 | 140 | | |
| L-cistein | 3.5 | 1.8 | | |
| Soybean oil | 29 | 40 | | |
| Lard | 207 | 0 | | |
| Fiber | 58 | 50 | | |
| Vitamin mix | 12 | 10 | | |
| Mineral mix | 12 | 35 | | |
| Dibasic calcium phosphate | 15 | 0 | | |
| Calcium carbonate | 6.4 | 0 | | |
| Potassium citrate | 19 | 0 | | |
| Choline bitartrate | 2.3 | 2.5 | | |
| Tert-butylhydroquinone | 0.1 | 8.0 | | |
| | | | | |

Table 1.Composition of high-fat diet and standard diet fed to male C57BL/6J mice for 4 weeks

 Table 2.Body weight and organ percentage of mice fed high-fat diet supplemented with flavanones and ibuprofen for 4weeks

| Diet | Standard | | | High-fat | | |
|-----------------|--------------|----------------|----------------|---------------|----------------|----------------|
| Supplement | None | None | Ibuprofen | Hesperidin | Eriocitrin | Eriodictyol |
| Weight gain (g) | $4.5\pm0.9a$ | $7.2 \pm 3.6b$ | $6.8 \pm 3.2b$ | $5.6\pm0.9ab$ | $7.0 \pm 1.4b$ | $7.5 \pm 2.5b$ |
| Visceralfat(%) | 1.0±0.3a | 4.5±1.9b | 4.6±1.9b | 3.8±1.4b | 3.8±1.3b | 4.5±1.3b |
| Liver (%) | 4.8±0.4b | 4.2±0.3a | 4.0±0.4a | 3.9±0.4a | 3.9±0.4a | 3.9±0.4a |
| Spleen(%) | 2.6±0.3ab | 3.0±0.8b | 2.5±0.6ab | 2.2±0.5a | 2.4±0.4a | 2.3±0.4a |
| Heart(%) | 0.47±0.05ab | 0.50±0.08b | 0.41±0.06a | 0.43±0.06a | 0.45±0.05ab | 0.42±0.07a |
| | | | | | | |

Values are mean \pm SD. Data analyzed by 1-factor ANOVA, followed by the Student Newman Keuls multiple comparison test. Means in a row followed by different letters differ significantly (p>0.05).

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Table 3. Biochemical profiles of mice fed high-fat diet supplemented with flavanones and ibu

 profen for 4weeks¹

| Diet | Standard | | | High-fat | | |
|---------------------------|--------------|---------------|----------------|----------------|----------------|---------------|
| Supplement | None | None | Ibuprofen | Hesperidin | Eriocitrin | Eriodictyol |
| Glucose(mg/dL) | $302 \pm 4a$ | $375 \pm 59b$ | $321 \pm 47ab$ | $337 \pm 85ab$ | $332 \pm 44ab$ | 316 ± 50ab |
| Triglycerides (mg/dL) | 81 ± 8 | 71 ± 14 | 88 ± 17 | 84 ± 20 | 90 ± 18 | 88 ± 20 |
| Total cholesterol (mg/dL) | 89 ± 14a | $138 \pm 2bc$ | $150 \pm 27c$ | $122 \pm 20b$ | $147 \pm 34c$ | $157 \pm 22c$ |
| HDL-C (mg/dL) | $55 \pm 9a$ | $83 \pm 1bc$ | $88 \pm 13c$ | 77 ± 15b | $94 \pm 22c$ | $99 \pm 14c$ |
| LDL-C(mg/dL) | $18 \pm 9a$ | $40 \pm 12c$ | $44 \pm 13c$ | $29\pm9b$ | $36 \pm 12bc$ | $40 \pm 8c$ |
| ALT ² (U/L) | 52 ± 13 | 44 ± 23 | 55 ± 9 | 41 ± 16 | 39 ± 14 | 41 ± 11 |
| AST ³ (U/L) | 152 ± 36 | 153 ± 52 | 161 ± 36 | 146 ± 45 | 166 ± 55 | 133 ± 50 |

¹Values are mean \pm SD. Data analyzed by 1-factor ANOVA, followed by the Student Newman Keuls multiple comparison test. Means in a row followed by different letters differ significantly (p>0.05).

²Alanine transaminase.

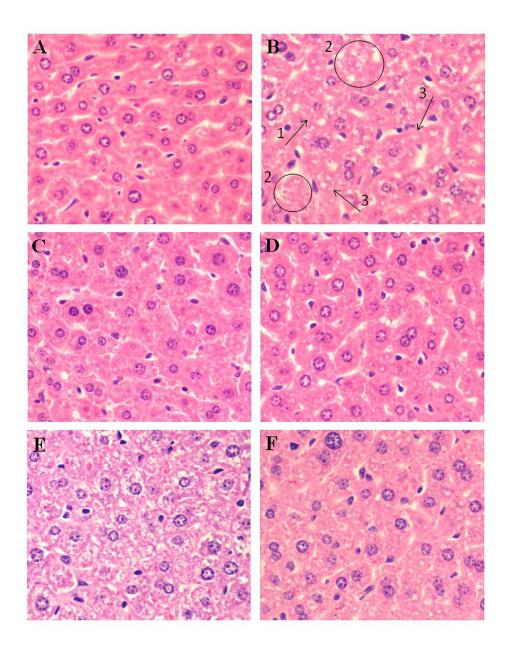
³Aspartate transaminase.

Table 4. Oxidative stress and inflammatory markers of mice fed high-fat diet supplemented with

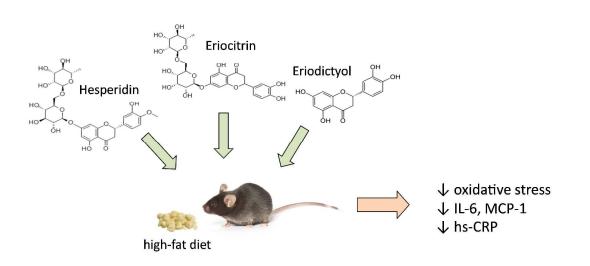
 flavanones and ibuprofen for 4weeks

| Diet | Standard | | | High-fat | | |
|---------------------|-------------|------------|--------------|-------------|------------|-------------|
| Supplement | None | None | Ibuprofen | Hesperidin | Eriocitrin | Eriodictyol |
| Serum TBARS (µm) | 0.95±0.21ab | 1.19±0.64b | 1.04±0.22ab | 0.81±0.17ab | 0.62±0.05a | 0.63±0.10a |
| Liver TBARS (µm/mg) | 0.08±0.03a | 0.14±0.08b | 0.03±0.01a | 0.07±0.01a | 0.06±0.02a | 0.05±0.01a |
| Serum ABTS (mm) | 1.50±0.06ab | 1.46±0.07a | 1.51±0.08abc | 1.56±0.08bc | 1.59±0.06c | 1.55±0.04bc |
| IL-6 (pg/ml) | 8.44±7.2a | 64.4±44.3b | 21.9±16.7a | 5.76±4.32a | 3.09±1.54a | 5.71±2.53a |
| IL-10 (pg/ml) | 4.97±2.16 | 3.22±1.77 | 6.02±4.20 | 5.27±2.23 | 3.95±2.03 | 4.71±1.57 |
| TNF-α (pg/ml) | 2.44±0.72 | 2.54±0.99 | 2.10±1.69 | 2.68±0.30 | 2.05±0.85 | 2.04±0.18 |
| MCP-1 (pg/ml) | 22.8±6.9a | 73.3±43.6b | 43.4±29.2a | 27.6±8.5a | 19.3±10.4a | 14.1±5.9a |
| Hs-CRP (mg/L) | 0.08±0.0bc | 0.09±0.0c | 0.08±0.01b | 0.06±0.02a | 0.06±0.02a | 0.07±0.02ab |

Values are mean \pm SD. Data analyzed by 1-factor ANOVA, followed by the Student Newman Keuls multiple comparison test. Means in a row followed by different letters differ significantly (p>0.05).



222x287mm (300 x 300 DPI)



In vivo antioxidant and anti-inflammatory effects of citrus flavanones