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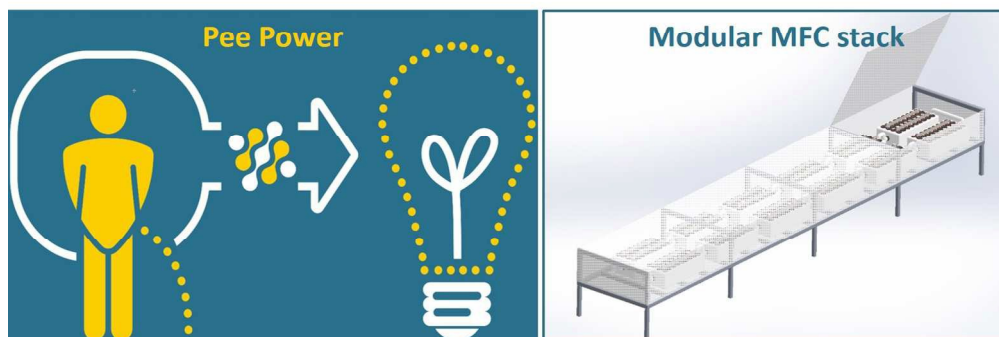
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Water Impact Statement: In general, 28% of UK clean water consumption is attributed to commercial urinals, with similar consumption rates across the Developed World. MFCs host a sustainable microbial community that transforms urine into power, water and minerals. Urine is an abundant resource and a highly effective feedstock for MFCs, deployment of which will reduce dependency on fossil fuel, decarbonisation of water management processes and elemental recycling.

Graphical abstract



275x113mm (150 x 150 DPI)

Pee Power Urinal – Microbial Fuel Cell Technology Field Trials In The Context Of Sanitation

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Abstract

This paper reports on the Pee Power urinal field trials, which are using Microbial Fuel Cells for internal lighting. The first trial was conducted on Frenchay Campus (UWE, Bristol) from February-May 2015 and demonstrated the feasibility of modular MFCs for lighting, with University staff and students as the users; the next phase of this trial is ongoing. The second trial was carried out during the Glastonbury Music Festival at Worthy Farm, Pilton in June 2015, and demonstrated the capability of the MFCs to reliably generate power for internal lighting, from a large festival audience (~1000 users/day). The power output recorded for individual MFCs is 1-2mW, and the power output of one 36-MFC-module, was commensurate of this level of power. Similarly, the real-time electrical output of both the Pee Power urinals was proportional to the number of MFCs used, subject to temperature and flow rate: the campus urinal consisted of 288 MFCs, generating 75mW (mean), 160mW (max) with 400mW when the lights were connected directly (no supercapacitors); the Glastonbury urinal consisted of 432 MFCs, generating 300mW (mean), 400mW (max) with 800mW when the lights were connected directly (no supercapacitors). The COD removal was >95% for the campus urinal and on average 30% for the Glastonbury urinal. The variance in both power and urine treatment was due to environmental conditions such as temperature and number of users. This is the first time that urinal field trials have demonstrated the feasibility of MFCs for both electricity generation and direct urine treatment. In the context of sanitation and public health, an independent power source utilising waste is essential in terms of both Developing and Developed World.

Keywords: microbial fuel cells; modular design; pee power urinal; sanitation; ceramic materials; fluidic isolation

Introduction

Microbial fuel cells (MFCs) have been receiving increased attention from the scientific community, even though the technology has been viewed with scepticism, at different levels of society. MFCs generate electrical energy directly from the break-down of organic matter via the metabolism of inhabitant microbes, with the rates of reaction being dictated by the microbial metabolic state ¹. Electrical output is therefore thermodynamically limited by the carbon-energy metabolism of the constituent cells of the biofilm community (mono- or mixed-culture) colonising the electrode ²⁻⁴. Microbial reactions are inherently lower than chemical or even purely enzymatic reactions, and therefore the magnitude of the absolute power output at any given time, is typically orders of magnitude lower than those generated from conventional chemical fuel cells ^{5,6}. Be that as it may, electricity generated in a MFC comes directly from waste or wastewater material, which in the break-down/utilisation

51 process is rendered cleaner and potentially suitable for direct discharge to the environment
52 ⁷⁻⁹. This is a competitive advantage that largely compensates for the lower levels of power.

53

54 Over the years, a wide range of organic substrates has been shown to work as fuels in MFCs.
55 Without being exhaustive, these are: types of food waste such as rotten fruit and prawn
56 shells ¹⁰; various types of wastewater from the paper industry, agriculture, dairy farms,
57 municipal treatment plants, oil industry, wine distilleries and breweries, and tanning
58 industry ¹¹; more recently, biodegradable materials ^{12,13} as well as human urine and septic
59 tank content, have also been shown to work very well as fuels for electricity generation ¹⁴⁻¹⁷.
60 The application of low cost ceramic membranes allowed to decrease the cost of structural
61 material, which is separating the anode and the cathode, to as low as 4.14 GBP/m² ¹⁸.
62 Utilising human waste directly and decreasing the cost of MFCs by the use of ceramic
63 membranes has allowed the technology to be exploited in the context of sanitation,
64 especially in countries of the Developing World, which lack the basic infrastructure for clean
65 water and sewerage ¹⁹. More than 2.5 billion people lack access to an improved sanitation
66 facility while 1 billion practice open defecation ²⁰. Inadequate drinking water, sanitation and
67 hygiene (WASH) are important risk factors where diarrhoeal disease burden relies on access
68 to water and sanitation facilities rather than water quality. The importance of improving
69 water and sanitation is the key for the prevention of diarrhoeal diseases ²¹. In addition to the
70 philanthropic dimension that the MFC approach has, sanitation in the Developing World
71 offers the ground for step-wise scale up field trials, to evaluate the technology in the real
72 world environment and thus assess its feasibility.

73

74 The efficient utilization of urine through MFCs incorporated in stacks/modules would no
75 longer require conventional energy intensive treatment by the wastewater companies and
76 also result in better balanced fertiliser ¹⁴. Efficient energy harvesting electronics for direct
77 MFC usage, is also a major challenge for scale up and implementation ²². So far, there have
78 been a few MFC field trials, like for example: a ceramic cascade temporarily installed in a
79 municipal wastewater treatment plant (Winfield et al 2012), a multi electrode MFC system
80 for contaminant removal ²³ as well as for winery wastewater treatment ²⁴, wireless sensors
81 ²⁵ and more recently, floating MFCs at the Nosedo, Milan wastewater treatment plant ²⁶.

82

83 The present study is based on previously reported novel ceramic designs developed as single
84 MFCs, showing high power performance with catholyte production and an ability to operate
85 practical applications, including direct LED lighting ²⁷ and recharging a mobile phone via a
86 single MFC unit ²⁸. The design gives the advantage of simplicity and functionality by utilising
87 multiple MFCs submerged in the same feedstock tank. The multiplication of units in parallel
88 would form a module, which could then be connected in series or parallel with other
89 modules, to scale-up into a flexible and robust stack. The study reported herewith, funded by
90 the Bill & Melinda Gates Foundation and Oxfam, had the following aims: (i) evaluate the
91 modular approach of stacking MFCs in a pilot scale trial for energy generation; (ii) integrate
92 the technology with the toilets that Oxfam uses in refugee camps and disaster areas to
93 demonstrate utility in terms of indoor lighting; (iii) scale-up of the urinal, at a systems level,
94 for testing during the Glastonbury Music Festival 2015 and (iv) assess the efficacy of urine
95 treatment.

96

97 **Methods**

98

99 MFC construction

100 MFC units were constructed using closed at one end terracotta caves (Weston Mill Pottery,
101 UK) as previously described ²⁷. The dimensions of the ceramic cylinders used in this work

102 were 150mm long, 48mm outside and 42mm inside diameter. Anode electrodes were made
103 of 30g/m² carbon veil fibre (PRF Composites, UK) of the dimensions: 600 x 260mm and
104 folded in half along its length. The carbon veil was then wrapped around the ceramic
105 cylinder and tied with a 50mm diameter stainless steel wire to secure the anode in place and
106 to provide a connection to the external circuit. Cathode electrodes were prepared using
107 activated carbon and PTFE mixture as previously described and inserted inside the ceramic
108 cylinder²⁷ as a single sheet of 130 x 140mm dimensions. Stainless steel crocodile clips were
109 then used to connect the cathode to the electrical circuit.

110

111 MFC module design & Inoculation

112 Thirty six MFCs were fitted into a plastic container of dimensions 70 (length) x 30 (width) x
113 16cm (depth). The anodes and cathodes were connected in a parallel electrical configuration
114 using aluminium bus bars and stainless steel wire, nuts and washers. The container was
115 inoculated with a mixture of activated sewage sludge (Cam Valley, Saltford, UK) and fresh
116 urine and operated in batch mode for the preliminary test. The total liquid capacity was 25
117 litres. Urine was collected from healthy individuals with no known previous medical
118 conditions, and pooled together before using as a feedstock. The pH would be on average 6.4.
119 No pH control was applied to the MFC stacks in both urinals.

120

121 Pee Power Oxfam urinal- UWE campus

122 Eight modules (288 MFCs in total) as described above were fluidically connected using
123 plastic elbow connectors and pipes to create a series loop and air gaps between the boxes.
124 This was to allow the 8 modules to be connected in a series electrical configuration. They
125 were inserted under the men's urinal unit installed at the Frenchay Campus, University of
126 the West of England, as shown in Figure 1. The structure was built to accommodate two
127 urinal bowls directly feeding the MFC modules fitted underneath the structure. The urinal on
128 the University campus resembles toilets produced by Oxfam and used in refugee camps to
129 make the trial as realistic as possible. Inside the cubicle, LED light modules were fitted to be
130 energised by the MFC stack via a capacitor bank consisting of 4x3000F capacitors in a series
131 parallel configuration (BCAP3000 p270, Maxwell Technologies). The lighting consisted of 4x
132 4.5W modified LED modules (Dial MR16-3H-WH-A1 12V-50Hz 530mA 4.5W 14W20). The
133 purpose of modification was to reduce the LED forward voltage from ~12V to ~3V and so
134 better suit the requirements of the MFC system. With this modification, the 4 LED modules
135 were consuming approximately 1.2W. The switching of the LED lights was controlled using a
136 low power passive-infra-red (PIR) sensor and a low power microcontroller (Microchip
137 PIC24F16KA102). This also allowed for a 3V backup power supply in the case of MFC system
138 failure. The holding tank was fitted as the initial (inlet/buffer) tank at the beginning of the
139 stack, providing feedstock for the MFC modules. There was also a collection tank fitted at the
140 outlet of the MFC stack. The operational time was 3 months starting in 05/03/15 – 31/05/15
141 and the analytical data presented herein were collected over the period of 5 weeks to assess
142 power and nutrient removal. Urine was donated voluntarily by the campus student and staff
143 population.

144



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146

147 **Figure 1.** (a) Pee Power field trial funded by Oxfam at the UWE campus in February –May
 148 2015; (b) 3D representation of the MFC stack with the inlet and outlet tanks underneath the
 149 urinal.

150

151 Pee Power field trial – Glastonbury

152 A field trial was performed at the Glastonbury Music Festival, England, between 22/06/15 –
 153 30/06/15. A specially adapted urinal (Dunster House, UK) was installed in the “Sacred
 154 Space” (aka “Stone Circle”) field. The urinal structure was fitted with 3 troughs which
 155 collected the urine from festival-goers and was used to ‘feed’ the MFC modules. Next to the
 156 urinal, an educational information point was used to interact with the public explaining the
 157 ideas and the technology behind the project. The men’s urinal was installed as shown in
 158 Figure 2 where 12 MFC modules (8 from the Oxfam Pee Power urinal + 4 new ones) were
 159 installed giving in total 432 MFCs in the stack and 300 litres of working volume. Similar to
 160 the previous trial, supercapacitors (10x3000F in a series parallel configuration giving
 161 7500F) were used as an energy store. The same LEDs as the ones used in the Oxfam system,
 162 but a higher number of 6, were used, due to the larger urinal facility. The total power
 163 consumption of the 6 LED modules was 1.8W. Due to the high number of users (between
 164 825-1000 per day) the estimated flow rate was approximately 330 l/day and the hydraulic
 165 retention time was 0.9 days.

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170 **Figure 2.** (a) Pee Power field trial in Glastonbury Music Festival, June 2015; (b) Urinal
 171 assembly and MFC stack arranged in 12 modules.

172

173 Analysis

174 Power performance was monitored with a multi-channel Agilent 34972A, LXI Data
175 Acquisition Unit (Farnell, UK) and were then processed using the Microsoft Excel and
176 GraphPad Prism software packages. Parameters such as pH and conductivity were measured
177 with a Hanna 8424 pH meter (Hanna, UK) and 470 Jenway conductivity meter (Camlab, UK)
178 respectively. Dry weight was determined by drying 1 mL of catholyte over 72 h in ambient
179 temperature and weighing the dry mass.

180

181 COD was analysed using the potassium dichromate oxidation method (COD HR test vials,
182 Camlab, UK) with an MD 200 photometer (Lovibond, UK) where 0.2 mL samples were taken
183 before and during MFC treatment and filter-sterilised prior to analysis. Total Nitrogen (TN)
184 was measured using MD 500 colorimeter (Lovibond, UK) and Vario Tube Test (0.5-25 mg/L)
185 on diluted samples. The concentration of anions in the anolyte (inlet, outlet) and catholyte
186 samples was determined by ion chromatography using a 930 Compact IC Flex (Metrohm,
187 UK). The samples were diluted with ultrapure water before they were collected by the 858
188 professional sample processor and introduced into the ion chromatograph.

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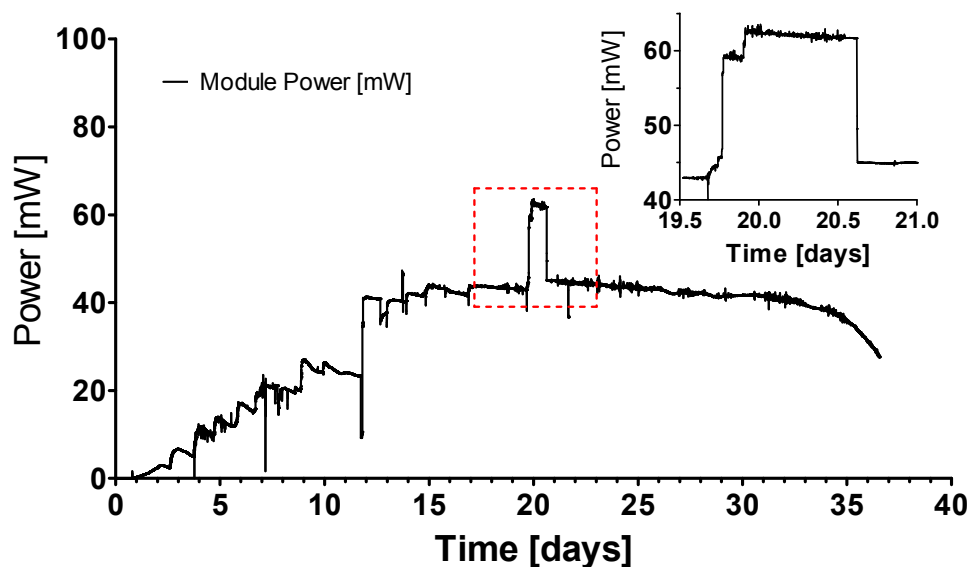
190 **Results and Discussion**

191

192 Initial MFC module testing

193 A single box assembled with 36 MFCs was initially tested under laboratory conditions. After
194 inoculation the MFC module was connected to a fixed resistor load and it was supplemented
195 with fresh and/or old urine daily. The resistor load was adjusted between 2.3 and 3.3 Ω with
196 stable performance achieved under a 3.3 Ω load. The module reached steady state
197 performance at 40 mW, however, when the resistor was adjusted to 2.3 and with some
198 modification and improvement of the cathode current collector, the peak power reached 62
199 mW (Figure 3), which is consistent with the individual ceramic MFC performance of up to
200 2.58 mW under controlled conditions. Multiple individual MFC units were also previously
201 tested in series and parallel configurations. These preliminary experiments constituted the
202 first tests of multiple MFC units in the same anodic feedstock, which simplified the
203 realisation of an MFC collective.

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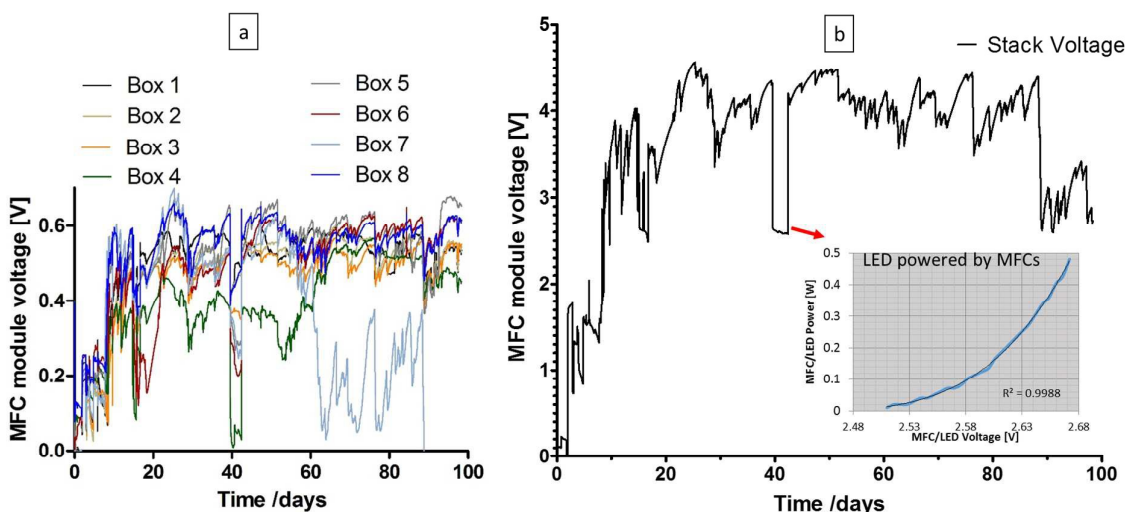


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206 **Figure 3.** Real time power output from a single module of 36 MFCs connected in parallel.
 207 Inset graph is a magnification of the marked area, where the load was changed to 2.3Ω for
 208 approximately 24 hours.
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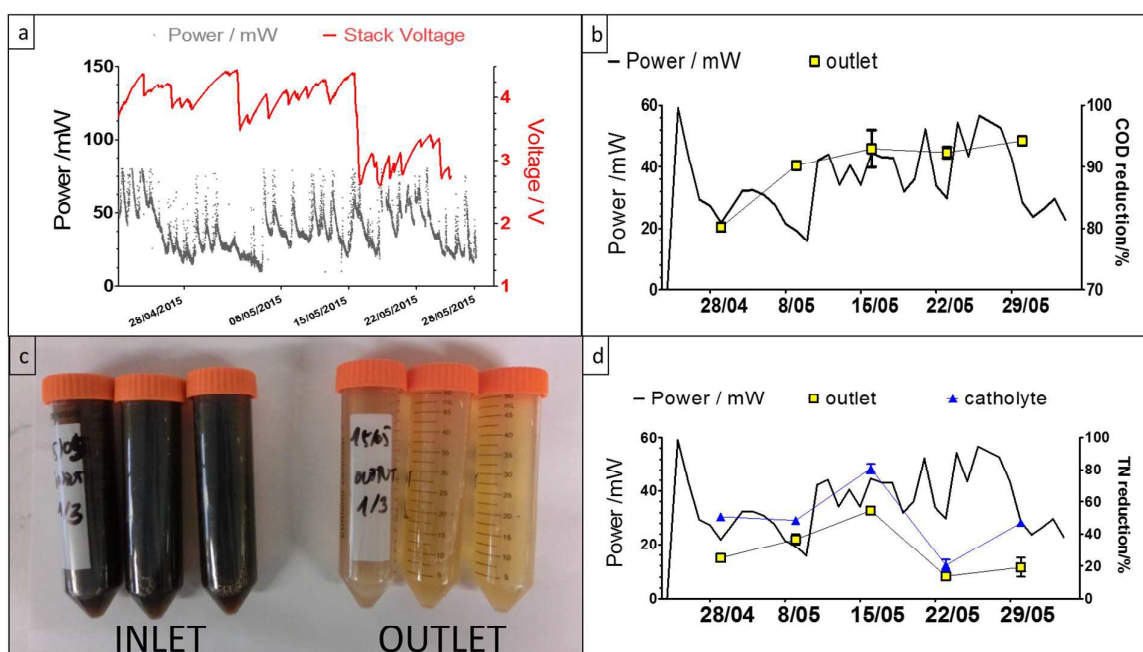
210 Campus (Oxfam) Trial

211 The pilot field trial ran for a total of three months. After inoculation with 1:1 activated sludge
 212 and urine mix, MFC modules were installed in the urinal and fed with neat fresh urine. The
 213 MFC voltage output of all eight modules is shown in Figure 4 (a) and the capacitor voltage in
 214 Figure 4 (b), where the inset presents the calibration curve for the LED lights directly
 215 connected to the MFC stack (i.e. by-passing the supercapacitors); during this time, the
 216 maximum power generated by the MFCs to power the lights was 0.4W for 75 hours. As can
 217 be seen from the graph on the left, 7 of the 8 MFC boxes were more consistent, in terms of
 218 performance, for the majority of the time (with the exception of when the LEDs were
 219 powered directly by the MFCs) and even with 1 of the boxes underperforming, the system
 220 was still operational without any polarity reversal; this demonstrated the robustness of the
 221 collective MFC modules under adverse conditions. As a joint project with Oxfam, it was
 222 hoped that electricity generated by MFCs will provide light for cubicles in refugee camps.
 223 The successful trial demonstrated that MFCs have got this capability (the campus Pee Power
 224 urinal has been re-started and it was still running successfully at the point of submitting this
 225 paper). Over the three month period, there was an estimated 5-10 users/day, which resulted
 226 in the processing of 2.5L-5L of urine daily. The hydraulic retention time for the whole stack
 227 was estimated to range between 2-3 weeks. The total catholyte synthesised²⁷ during the 3
 228 month period for the 288 MFCs of 100mL internal volume was ca. 34L.
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Figure 4. (a) Individual MFC module voltage performance of the 8 Oxfam stack boxes and (b) voltage output of the connected capacitor. The decreases in the voltage data curves are from when volunteers were visiting the urinal, and hence activating the lights to switch ON. The magnitude and length of decrease is an indication of the length of time the lights were ON. Figure 4B (inset) shows a calibration curve for the power consumed by the LED lighting at a given voltage. As can be seen the MFC stack gave a maximum power of approximately 0.4W for 75hrs.



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Figure 5 a) Power and Voltage performance over the 5 week analysis period; b) COD reduction in the system with reference to the power of the stack; c) Samples collected from the inlet and the outlet tanks on the 15/05/2015 (week three); d) Reduction of the total nitrogen from the outlet and catholyte samples compared to the inlet, with reference to the power performance.

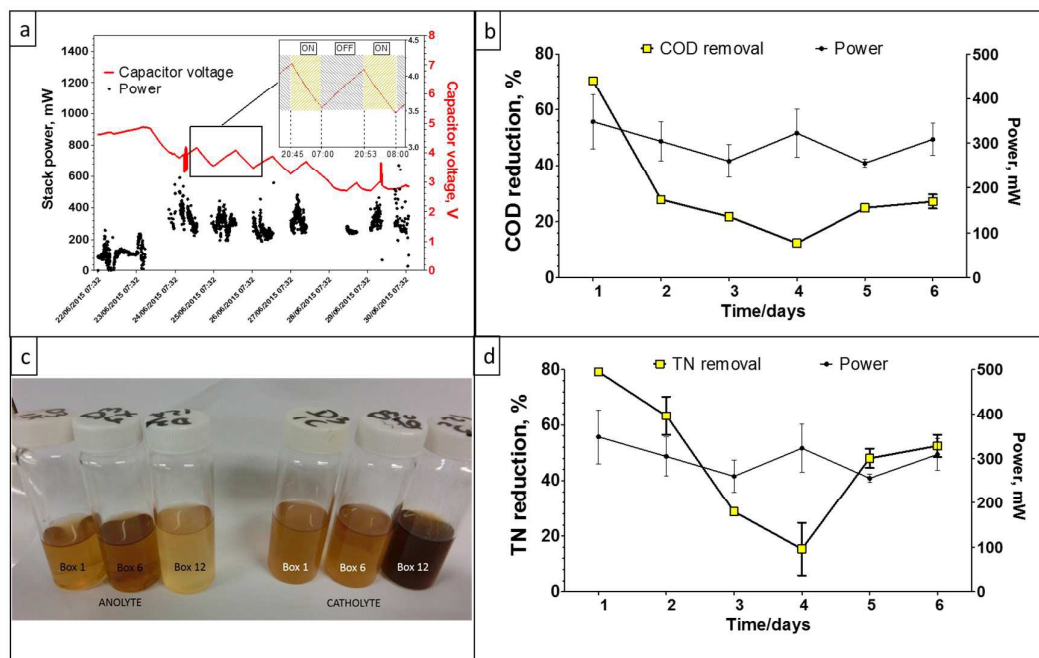
249 The power behaviour in Figure 5a demonstrates the dynamic nature of the system, and the response to the lights switching ON when people were entering the urinal, and OFF after a period of 3 minutes (based on a pIR sensor, also powered by the MFC stack). The peak values during this 5 week period were reaching, on average 75mW, and the highest value recorded was ca. 160mW; the variation in performance was due to natural temperature conditions as well as frequency of uses. The COD reduction is shown to be >90% reaching a maximum of 98%, which was mainly due to the long HRT, and this high reduction level was maintained throughout the trial. This is also evident from the colour of the inlet and outlet samples collected (Fig.5c), which clearly showed that the urine was being treated during the process. The highest total nitrogen reduction was >50% for the anolyte and the lowest was <20%, which was the same for the synthesised catholyte (Fig.5d). For the catholyte, the maximum total nitrogen reduction was approximately 80%. Increased TN reduction in the catholyte was probably due to the MFC electrochemical operation stimulating pH increase in the cathodic chamber (Supporting Information, Figure S1a) that allowed ammonia stripping¹⁵ and more efficient nitrogen removal.

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Glastonbury Trial

267 The data presented in Figure 6a shows the stack power performance over the period of 8 days. The first two days of data show the system in charge mode only in preparation for the festival proper. On the 23rd of June the light in the cubicle was switched on (Further controlled by a PIR sensor) and it is represented by the supercapacitor voltage drop. From this point the lighting system was disabled during daylight hours (Figure 6a, inset) to allow the system to recharge and then re-enabled during the night. Stack power had increased from the system start date until the 24th of June (first day of the festival) reaching up to 400

274 mW total power output and showed good and stable power performance through the time of
 275 the festival due to high usage and constant fast flow of fresh feedstock. The long decrease in
 276 capacitor voltage between the 27th-28th June is when the lights were intentionally left ON
 277 for 24 hours. As can be seen, at the end of this 24hr period, the system had almost reached
 278 equilibrium (MFC power-in = LED power-out). The spike between the 29th-30th June is
 279 when the supercapacitors were disconnected and the data following this (going into the 30th
 280 June) are from when the MFCs were directly powering the lights. The peak power output was
 281 up to 800mW, which is equivalent to 19.2Wh over 24hrs, and showed a mean output of
 282 1.85mW/MFC, which is consistent with the laboratory data. Power output is significantly
 283 higher than the power levels recorded for the campus Pee Power Oxfam trial, which might be
 284 due to the increased number of modules (from 8 to 12), very high flow rate at the festival
 285 and elevated temperature due to the direct sunlight exposure. The catholyte generated
 286 during this trial was approximately 43 litres as each of the MFC units produced 0.1L of
 287 catholyte. The very high flow rate and reduced HRT, did however affect the COD reduction
 288 capability of the system as a whole, recording a maximum of ca. 70%, a minimum of 15% and
 289 mean for the majority of the time of 25%. The same effect was observed for the total
 290 nitrogen reduction of the system, where the maximum recorded was 79% and the minimum
 291 was 6%. The high usage did however result in struvite accumulation inside the connecting
 292 pipes, as a result of the inlet tank not holding urine for the required amount of time. The
 293 colour of the collected (anolyte) inlet and outlet samples, also confirmed the reduced
 294 treatment performance.
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299 **Figure 6.** a) Real time performance of the urinal stack with voltage. The inset represents a 2-
 300 day charge-discharge cycle of the capacitors when the LED lights were turned OFF (day) and
 301 ON (night); b) COD reduction and power performance over the 6 days of the field trial; c)
 302 anolyte and catholyte samples collected on day 3; d) Total Nitrogen (TN) reduction rates in
 303 Box 1, 6 and 12 in relation to stack power performance.
 304

305

306 MFC electrochemical treatment is actively changing the chemistry of the treated substrate
 favouring pH and ion separation (see Supporting information), which leads to the recovery

307 of slow-release fertiliser and electricity from urine (Zang et al., 2012). Porous terracotta has
308 been reported to allow ionic movement from the anodic chamber to the cathode surface ²⁹,
309 however when the cathode is exposed as part of an outside surface (as opposed to a secluded
310 inner surface used in the Pee Power examples), might also lead to chemical scaling (salt
311 deposition) and consequently biofouling ³⁰ of the outer cathode.

312
313 Ceramic is a cost effective replacement for the cation exchange membrane ^{31,32} and it proved
314 once again to be a valid building block for MFCs. Moreover, it is a functional medium for the
315 electroosmotic flow of ions induced by the MFC electric field ²⁷. It promotes extraction and
316 ion separation essential for elemental recovery and recycling. This electro-kinetic function of
317 MFCs depends on power performance ³³, which implies that the more efficiently the systems
318 perform, the more improved the elemental extraction with the added advantage of direct
319 monitoring of effluent quality ³⁴. Environmental sustainability is an integral part of the
320 design, maintenance and operation of a urinal facility, promoting the awareness of
321 environmental issues, whilst providing the tools and incentives to address them. Such field
322 trials are essential so that the technology can be further advanced and applied at larger scale.

323
324 The decreased treatment performance of the Glastonbury Pee Power urinal can easily be
325 rectified with the appropriate fluidic arrangements, to allow the same number of users, but
326 elongate the hydraulic retention time of the collective MFCs, so that higher COD and TN
327 reduction efficiencies can be achieved. This was not done in this case, due to the timing and
328 location constraints of the MFC stack, which was mainly for demonstration purposes. The
329 high number of users, resulting in a high throughput, implies that a further scaled-up Pee
330 Power urinal will be more efficient in treating higher urine volumes and at the same time
331 maintain a high level of power performance.

332

333 **Conclusions**

334 The provision of safe water, adequate sanitation and hygiene, is critically important for
335 promoting individual and community-level health in the Developing World. MFC based
336 technologies prove to be a sustainable solution even in remote locations, improving
337 sanitation and hygiene, and opening the way to elemental recycling. The Pee Power urinals
338 are perhaps one example of how these can be achieved.

339

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351

352 **References**

353

- 354 (1) Ledezma, P.; Greenman, J.; Ieropoulos, I. Maximising electricity production by
355 controlling the biofilm specific growth rate in microbial fuel cells. *Bioresour. Technol.*
356 **2012**, *118*, 615–618.

357

- 358 (2) Kim, N.; Choi, Y.; Jung, S.; Kim, S. Effect of initial carbon sources on the performance of
359 microbial fuel cells containing *Proteus vulgaris*. *Biotechnol. Bioeng.* **2000**, *70* (1), 109–
360 114.
361
- 362 (3) Jong, B. C.; Kim, B. H.; Chang, I. S.; Liew, P. W. Y.; Choo, Y. F.; Kang, G. S. Enrichment,
363 Performance, and Microbial Diversity of a Thermophilic Mediatorless Microbial Fuel
364 Cell. *Environ. Sci. Technol.* **2006**, *40* (20), 6449–6454.
365
- 366 (4) Ieropoulos, I.; Melhuish, C.; Greenman, J. Artificial gills for robots: MFC behaviour in
367 water. *Bioinspir. Biomim.* **2007**, *2* (3), S83–S93.
368
- 369 (5) Kirubakaran, A.; Jain, S.; Nema, R. K. A review on fuel cell technologies and power
370 electronic interface. *Renew. Sustain. Energy Rev.* **2009**, *13* (9), 2430–2440.
371
- 372 (6) Mekhilef, S.; Saidur, R.; Safari, A. Comparative study of different fuel cell technologies.
373 *Renew. Sustain. Energy Rev.* **2012**, *16* (1), 981–989.
374
- 375 (7) Winfield, J.; Ieropoulos, I.; Greenman, J. Investigating a cascade of seven hydraulically
376 connected microbial fuel cells. *Bioresour. Technol.* **2012**, *110*, 245–250.
377
- 378 (8) Ledezma, P.; Greenman, J.; Ieropoulos, I. MFC-cascade stacks maximise COD reduction
379 and avoid voltage reversal under adverse conditions. *Bioresour. Technol.* **2013**, *134*,
380 158–165.
381
- 382 (9) Habermann, W.; Pommer, E. Biological fuel cells with sulphide storage capacity. *Appl.*
383 *Microbiol. Biotechnol.* **1991**, *35* (1), 128–133.
384
- 385 (10) Ieropoulos, I.; Melhuish, C. EcoBot-II: An artificial agent with a natural metabolism. *J.*
386 *Adv. Robot. Syst.* **2005**, *2* (4), 295–300.
387
- 388 (11) Pant, D.; Van Bogaert, G.; Diels, L.; Vanbroekhoven, K. A review of the substrates used
389 in microbial fuel cells (MFCs) for sustainable energy production. *Bioresour. Technol.*
390 **2010**, *101* (6), 1533–1543.
391
- 392 (12) Winfield, J.; Ieropoulos, I.; Rossiter, J.; Greenman, J.; Patton, D. Biodegradation and
393 proton exchange using natural rubber in microbial fuel cells. *Biodegradation* **2013**, *24*
394 (6), 733–739.
395
- 396 (13) Winfield, J.; Chambers, L. D.; Rossiter, J.; Greenman, J.; Ieropoulos, I. Urine-activated
397 origami microbial fuel cells to signal proof of life. *J. Mater. Chem. A* **2015**, *3* (13), 7058–
398 7065.
399

- 400 (14) Ieropoulos, I.; Gajda, I.; You, J.; Greenman, J. Urine—Waste or Resource? The Economic
401 and Social Aspects. *Rev. Adv. Sci. Eng.* **2013**, *2* (3), 192–199.
402
- 403 (15) Kuntke, P.; Smiech, K. M.; Bruning, H.; Zeeman, G.; Saakes, M.; Sleutels, T. H. J. a;
404 Hamelers, H. V. M.; Buisman, C. J. N. Ammonium recovery and energy production from
405 urine by a microbial fuel cell. *Water Res.* **2012**, *46* (8), 2627–2636.
406
- 407 (16) Ieropoulos, I.; Greenman, J.; Melhuish, C. Urine utilisation by microbial fuel cells;
408 energy fuel for the future. *Phys. Chem. Chem. Phys.* **2012**, *14* (1), 94–98.
409
- 410 (17) Yazdi, H.; Alzate-Gaviria, L.; Ren, Z. J. Pluggable microbial fuel cell stacks for septic
411 wastewater treatment and electricity production. *Bioresour. Technol.* **2015**, *180*, 258–
412 263.
413
- 414 (18) Pasternak, G.; Greenman, J.; Ieropoulos, I. Comprehensive study on ceramic
415 membranes for low cost microbial fuel cells. *ChemSusChem* **2015**.
416
- 417 (19) UNESCO. “Water in a Changing World” *The Third edition of the United Nations World*
418 *Water Development Report (WWDR3)*; 2009.
419
- 420 (20) World Health Organisation. *WHO | UN-water global analysis and assessment of*
421 *sanitation and drinking-water (GLAAS) 2014 - report*; World Health Organization, 2014.
422
- 423 (21) Prüss-Ustün, A.; Bartram, J.; Clasen, T.; Colford, J. M.; Cumming, O.; Curtis, V.; Bonjour,
424 S.; Dangour, A. D.; De France, J.; Fewtrell, L.; et al. Burden of disease from inadequate
425 water, sanitation and hygiene in low- and middle-income settings: a retrospective
426 analysis of data from 145 countries. *Trop. Med. Int. Health* **2014**, *19* (8), 894–905.
427
- 428 (22) Wang, H.; Park, J.; Ren, Z. J. Practical Energy Harvesting for Microbial Fuel Cells: A
429 Review. *Environ. Sci. Technol.* **2015**, *49* (6), 3267–3277.
430
- 431 (23) Heidrich, E. S.; Edwards, S. R.; Dolfing, J.; Cotterill, S. E.; Curtis, T. P. Performance of a
432 pilot scale microbial electrolysis cell fed on domestic wastewater at ambient
433 temperatures for a 12 month period. *Bioresour. Technol.* **2014**, *173*, 87–95.
434
- 435 (24) Cusick, R. D.; Bryan, B.; Parker, D. S.; Merrill, M. D.; Mehanna, M.; Kiely, P. D.; Liu, G.;
436 Logan, B. E. Performance of a pilot-scale continuous flow microbial electrolysis cell fed
437 winery wastewater. *Appl. Microbiol. Biotechnol.* **2011**, *89* (6), 2053–2063.
438
- 439 (25) Donovan, C.; Dewan, A.; Heo, D.; Beyenal, H. Batteryless, wireless sensor powered by a
440 sediment microbial fuel cell. *Environ. Sci. Technol.* **2008**, *42* (22), 8591–8596.
441
- 442 (26) Martinucci, E.; Pizza, F.; Perrino, D.; Colombo, A.; Trasatti, S. P. M.; Lazzarini Barnabei,

- 443 A.; Liberale, A.; Cristiani, P. Energy balance and microbial fuel cells experimentation at
444 wastewater treatment plant Milano-Nosedo. *Int. J. Hydrogen Energy* **2015**, *40* (42),
445 14683–14689.
446
- 447 (27) Gajda, I.; Greenman, J.; Melhuish, C.; Ieropoulos, I. Simultaneous electricity generation
448 and microbially-assisted electrosynthesis in ceramic MFCs. *Bioelectrochemistry* **2015**,
449 *104*, 58–64.
450
- 451 (28) Gajda, I.; Stinchcombe, A.; Greenman, J.; Melhuish, C.; Ieropoulos, I. Ceramic MFCs with
452 internal cathode producing sufficient power for practical applications. *Int. J. Hydrogen*
453 *Energy* **2015**, *40* (42), 14627–14631.
454
- 455 (29) Ghadge, A. N.; Ghangrekar, M. M. Development of low cost ceramic separator using
456 mineral cation exchanger to enhance performance of microbial fuel cells. *Electrochim.*
457 *Acta* **2015**, *166*, 320–328.
458
- 459 (30) Ghadge, A. N.; Ghangrekar, M. M. Performance of low cost scalable air-cathode
460 microbial fuel cell made from clayware separator using multiple electrodes. *Bioresour.*
461 *Technol.* **2015**, *182*, 373–377.
462
- 463 (31) Winfield, J.; Greenman, J.; Huson, D.; Ieropoulos, I. Comparing terracotta and
464 earthenware for multiple functionalities in microbial fuel cells. *Bioprocess Biosyst. Eng.*
465 **2013**, *36* (12), 1913–1921.
466
- 467 (32) Behera, M.; Jana, P. S.; Ghangrekar, M. M. Performance evaluation of low cost microbial
468 fuel cell fabricated using earthen pot with biotic and abiotic cathode. *Bioresour.*
469 *Technol.* **2010**, *101* (4), 1183–1189.
470
- 471 (33) Gajda, I.; Greenman, J.; Melhuish, C.; Santoro, C.; Li, B.; Cristiani, P.; Ieropoulos, I.
472 Electro-osmotic-based catholyte production by Microbial Fuel Cells for carbon
473 capture. *Water Res.* **2015**.
474
- 475 (34) Chouler, J.; Di Lorenzo, M. Water Quality Monitoring in Developing Countries; Can
476 Microbial Fuel Cells be the Answer? *Biosensors* **2015**, *5* (3), 450–470.
477