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Cu(I) stabilizing crosslinked polyethyleneimine

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Polyethyleneimine (PEI) is known for its metal-coordinating properties and in its crosslinked form has applications in different areas ranging from drug delivery to waste water treatment and recovery of trace metals. With the aim to regulate the coordination environment of Cu(I) and Cu(II) ions in marine coatings, we have prepared a triazole cross-linking agent with 'soft' coordination that can crosslink PEI via indirect reductive amination. We have shown that this triazole crosslinker not only increases the Cu(II) absorption capacity of the crosslinked PEI in comparison to the traditionally used glutaraldehyde-crosslinked PEI (PEI-GA), but also allows the crosslinked polymer network to stabilize the Cu(I) oxidation state more effectively. The Cu(II) uptake and Cu(I) stabilization of the polymer have been determined by elemental analysis and UV-vis spectroscopy. It was found that the triazole-crosslinked polymer (PEI-TA) could coordinate up to 12 wt% of Cu(II) before and 6 wt% Cu(II) after imine reduction.

1 Introduction

Trace metals such as Cu, Fe, and Zn are essential elements for most living organisms.¹ At the same time, these metals are also hazardous to the environment and human health in higher concentrations.¹⁻³ Due to their toxicity to marine species, Cu and Zn have been used in coatings to combat marine foulers that settle and grow on ship hulls.^{4,5} These foulers are a big challenge for marine transportation and increase fuel consumption and maintenance. However, the release of toxic substances into seawater is a major problem especially in harbours and other areas with limited water circulation. This has led to regulatory restrictions on the use and release of copper in some countries^{6,7} and there is a need to find environmentally friendly solutions to the problem of marine fouling.

Both Cu(I) and Cu(II) are efficient anti-fouling agents.^{8,9} However, Cu(I) is unstable under aerobic conditions and rapidly oxidizes to Cu(II) in seawater.^{10,11} It is widely accepted that it is Cu(I) that passes through the biological membrane of organisms,^{12,13} and that the coordination environment of most biological cells favours Cu(I).¹⁴ Our working hypothesis is that by changing the coordinating characteristics of a coating, the redox potential for the Cu(I)/Cu(II) pair is attuned so that the concentration of Cu(I) increases, leading to a better antibiofouling efficiency.¹⁵ Maintaining such a window where the redox cycle of Cu(I)/Cu(II) occurs also allows the Cu ions to impose the most toxic effect.¹⁶ This work is one step towards developing a new type of 'green' coating that requires no toxic biocide additives, but instead makes use of the copper already

present in seawater. The aim with this type of coating is to create a flux of copper at the coating interface; in effect, a permanent cyclic mechanism of copper uptake and release into and out of the coating. Of particular interest for this purpose are coordinating polymers that can bind to both Cu(II) and Cu(I) ions.

Tris[(benzyltriazolyl)methyl]amine (TBTA) (Figure 1) is a tripodal ligand that has found applications in Cu(I)-catalysed reactions since its first introduction in 2004 by Fokin et al.¹⁷ Both the presence of triazole groups and the tripodal structure of this ligand, which can adapt to the different geometries required for Cu(I) and Cu(II) binding, have made it suitable for applications where coordination to both these ions is of importance. The calculated optimized structures of TBTA and its complexes with Cu(I) and Cu(II) are shown in Figure 1, where the $[\text{Cu}^{\text{I}}_2(\text{TBTA})_2]$ complex forms a distorted tetrahedral while the Cu(II) complex forms a distorted trigonal bipyramidal structure with the formula of $[\text{Cu}^{\text{II}}\text{Cl}(\text{TBTA})]$. The calculated structures are also in line with the reported crystal structures for these complexes.¹⁸ We have previously reported the synthesis and characterization of coordinating polymers functionalized by TBTA (poly(TBTA)) via a one-pot click reaction procedure.¹⁹ We demonstrated that a thin poly(TBTA) film can absorb up to 76 mg g⁻¹ (7.1 wt%) of Cu(II) from an aqueous solution of copper. However, despite its performance, the multi-step synthesis process of poly(TBTA) can be a drawback for large-scale applications. Therefore, we here present alternative amine-based polymeric structures via a simpler synthetic route which possess a similar 'soft' coordinating character.

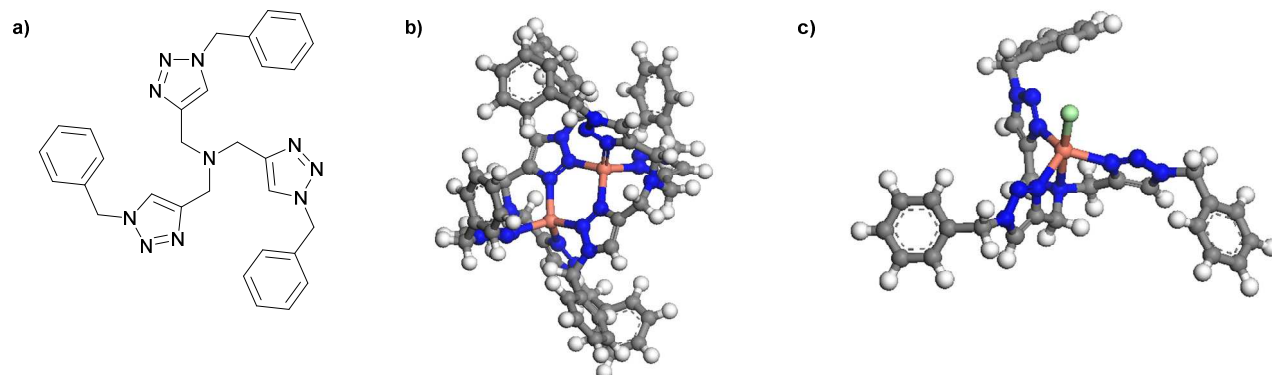
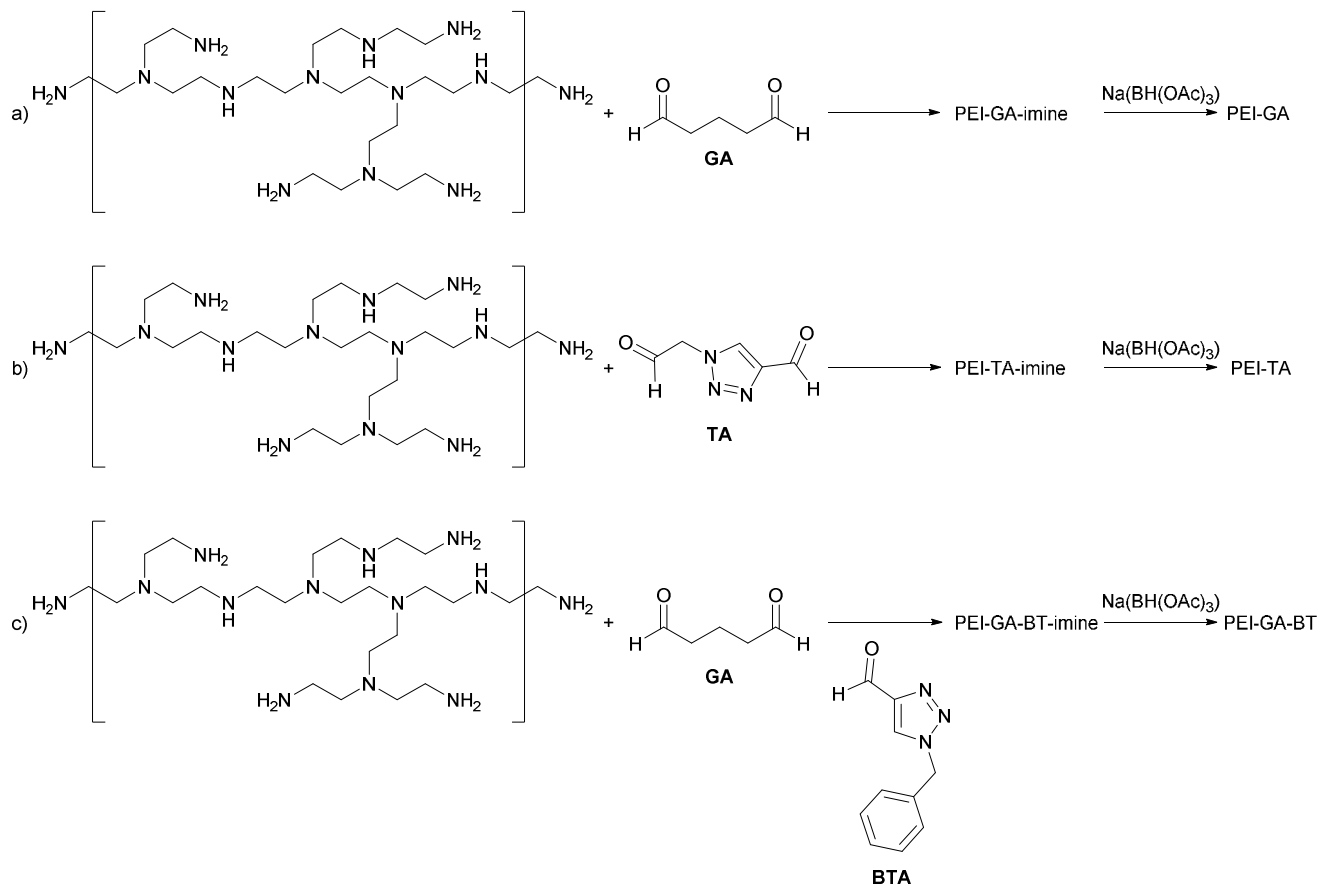


Figure 1. (a) Tris[(benzyl)triazol]methylamine (TBTA); (b) $[\text{Cu}^{\text{II}}_2(\text{TBTA})_2]$; and (c) $[\text{Cu}^{\text{I}}\text{Cl}(\text{TBTA})]$, (b) and (c) are calculated at PBE/DNP(4.4).

Polyethyleneimine (PEI) has a high affinity and selectivity for Cu ions.²⁰⁻²² A common way to increase the stability of PEI on a surface is by chemical cross-linking using glutaraldehyde^{23,24} or epichlorohydrine.²⁵ The nature of the cross-linker can alter the affinity and selectivity of PEI to metal ions. Moreover, its crosslinked form allowing reuse after regeneration,^{25,26} making it an interesting polymer for industrial uses such as waste water treatment.²⁶ Also the selectivity can be changed by modifying the crosslinked structure with other coordinating groups.^{27,28} Considering the ability of PEI to bind to Cu(II), we hypothesise that PEI crosslinked with a triazole group will coordinate both Cu(II) and Cu(I) effectively. The crosslinker employed here consists of a triazole ring with aldehyde functionalities on the 1- and 4-positions. For convenience, we refer to the crosslinker as a triazole dialdehyde (TA). The cross-linked network of PEI-TA is also expected to be able to satisfy the different

coordination geometries required for Cu(I) and Cu(II), i.e. 4-coordinated tetrahedral for Cu(I) and 5-coordinated trigonal bipyramidal for Cu(II). Glutaraldehyde (GA) was used as a reference crosslinker. It has already been shown that a cross-linked PEI-GA film can absorb copper from extremely dilute copper solutions.^{29,30} Our focus here is different since we aim to tune the affinity of the film towards a specific oxidation state of copper. To investigate the versatility of introducing triazole groups, we have also functionalized PEI-GA with a mixture of (benzyl)triazole carbaldehyde (BTA) and GA to achieve a Cu(I) stabilizing structure. To prevent further hydrolysis the imine bonds in the cross-linked polymers were reduced to amine-functionalities.³¹ The effect of this reduction on the copper uptake was also studied. Scheme 1 shows the reagents as well as the cross-linking reactions to prepare PEI-GA, PEI-TA, and PEI-GA-BT.



Scheme 1. (a) Glutaraldehyde-crosslinked PEI (PEI-GA); (b) triazolodialdehyde-crosslinked PEI (PEI-TA); and (c) (benzyl)triazole-functionalized glutaraldehyde-crosslinked PEI (PEI-GA-BT).

2 Experimental section

2.1 Materials and reagents

1-Benzyl-1H-1,2,3-triazole-4-carbaldehyde (**BTA**, Scheme 1) was prepared according to a previously reported procedure (Scheme 2).³² In this work, we have used a branched polyethyleneimine from Sigma Aldrich (average Mw ~25,000 by LS, average Mn ~10,000 by GPC, branched). Benzyl azide was purchased from Alfa Aesar. Remaining chemicals were purchased from Sigma-Aldrich.

2.2 Characterizations

FTIR-ATR spectra were recorded on a Perkin Elmer system Frontier (at a resolution of 2 cm⁻¹ with 20 scans) The UV-vis measurements were carried out using a Perkin Elmer Lambda

900 UV/VIS/NIR spectrometer. Elemental analysis was carried out by Mikroanalytisches Laboratorium KOLBE (www.mikro-lab.de). Scanning electron microscopy (SEM) was performed with a Leo Ultra 55 FEG system equipped with an Oxford Inca EDS X-ray microanalysis system for energy dispersive X-ray spectroscopy (EDS) analysis.

2.3 Synthesis

Scheme 2 shows the synthetic procedures for the crosslinker **TA** as well as the functional group **BTA**.

2.3.1 2-AZIDO-1,1-DIETHOXYETHANE

Bromoacetaldehyde diethyl acetal (5 mmol, 1.00 g), sodium azide (15 mmol, 1.00 g), and a catalytic amount of sodium iodide were mixed in dimethyl sulfoxide (DMSO, 10 mL) and

stirred at 90 °C for 45 hr. Water (25 mL) was then added to the reaction mixture. Diethyl ether (50 mL) was added and the phases were separated. The organic phase was washed with brine (5 × 30 mL) to remove the DMSO residue and dried over MgSO₄ and concentrated under vacuum to yield a yellow oil (0.67 g, 4.21 mmol, 83.1%). ¹H NMR (400 MHz, CDCl₃) δ 4.59 (t, 1H, O-CH-O), 3.75-3.68 (m, 2H, O-CH₂-CH₃), 3.61-3.53 (m, 2H, O-CH₂-CH₃), 3.22 (d, 2H, N₃-CH₂-CH), 1.23 (t, 6H, CH₂-CH₃), ¹³C NMR (400 MHz, CDCl₃) δ 101.42, 62.98, 52.47, 15.26.

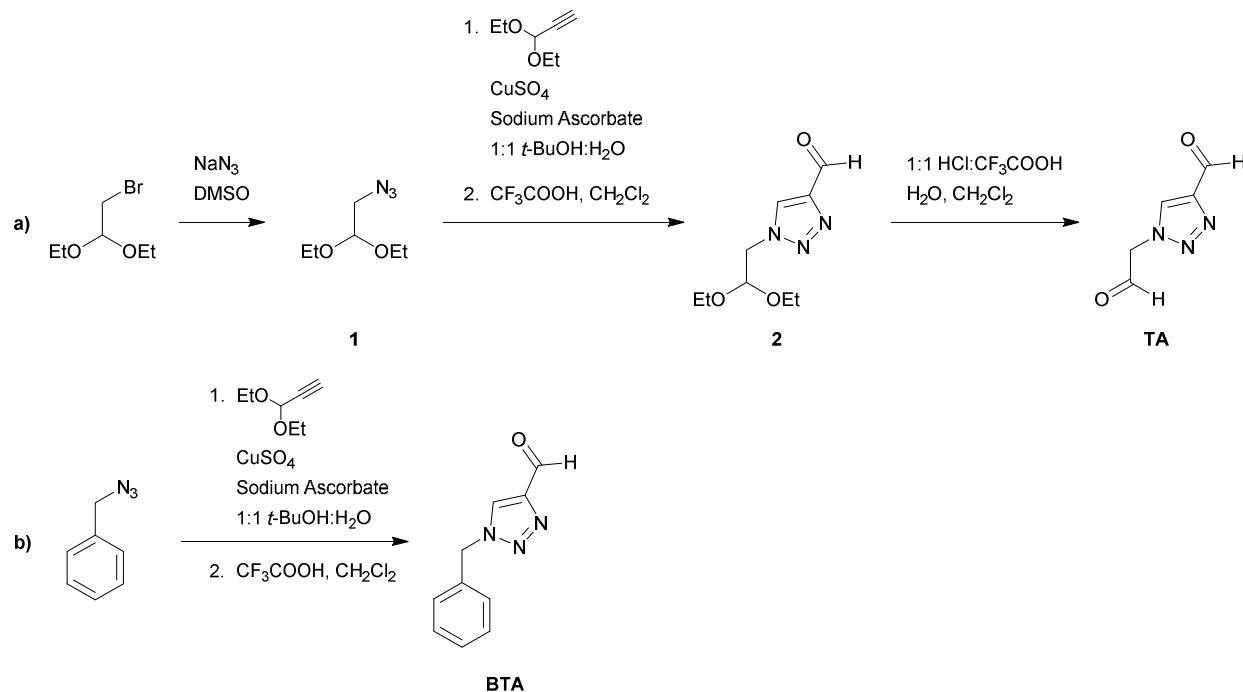
2.3.2 1-(2,2-DIETHOXYETHYL)-1H-1,2,3-TRIAZOLE-4-CARBALDEHYDE (2)

To the solution of **1** (0.67 g, 4.21 mmol) in *t*-BuOH:H₂O (1:1, 15 mL), diethylpropargyl acetal (5.00 mmol, 0.68 g), sodium bicarbonate (0.10 g), CuSO₄ 1 M solution (0.4 mL, 10 mol%), and sodium ascorbate (0.30 g, 30 mol%) were added. The reaction mixture was stirred for 24 hours at 40 °C. Diethyl ether (50 mL) was added and the phases were separated. The organic phase was washed with saturated NaHCO₃ (3 × 30 mL) and brine (2 × 50 mL), dried over MgSO₄ and concentrated under vacuum to afford a yellow oil (0.89 g). This oil was then dissolved in dichloromethane (4 mL), followed by the addition of water (1 mL) and trifluoroacetic acid (TFA) (1 mL). The

mixture was stirred for 12 hours. Diethyl ether (10 mL) was added and the organic phase was separated and washed with saturated NaHCO₃ (3 × 40 mL) and brine (1 × 40 mL), dried over MgSO₄, and concentrated under vacuum to give a yellow oil (0.30 g, 1.40 mmol, 33% over two steps). ¹H NMR (400 MHz, CDCl₃) δ 10.10 (s, 1H, CHO), 8.19 (s, 1H, H-triazole), 4.76 (t, 1H, O-CH-O), 4.49 (d, 2H, N-CH₂-CH), 3.75-3.67 (m, 2H, O-CH₂-CH₃), 3.52-3.44 (m, 2H, O-CH₂-CH₃), 1.15 (t, 6H, -CH₃), ¹³C NMR (400 MHz, CDCl₃) δ 185.0, 147.7, 126.7, 100.1, 63.8, 52.9, 15.1. Elemental analysis: Calc. for C₉H₁₅N₃O₃: C, 50.69; H, 7.09; N, 19.71; O, 22.51. Found: C, 50.48; H, 7.01; N, 19.42; O, 23.09.

2.3.3 1-(2-OXOETHYL)-1H-1,2,3-TRIAZOLE-4-CARBALDEHYDE (TA)

To fully hydrolyze **2**, a solution of **2** (2.00 g, 9.4 mmol) in dichloromethane (5 mL) was prepared. To this solution, deionized water (5 mL) was added followed by the addition of trifluoroacetic acid (2.5 mL) and hydrochloric acid (37%, 2.5 mL). After 7 days, the aqueous phase was separated and neutralized by the addition of K₂CO₃ and deionized water. This aqueous solution was used for the crosslinking reaction without further purification.



Scheme 2. Synthetic procedures for (a) the TA crosslinker; (b) benzyl(triazole) aldehyde (BTA).

2.3.4 PEI-GA-IMINE

PEI (2.05 g) was dissolved in deionized water (20 mL) and then added to acetonitrile (150 mL) under mild stirring. Glutaraldehyde (10 mL, 25 wt% aqueous solution) was then added to the mixture. After 60 hours of reaction, the red particles were separated by centrifugation, washed with

acetonitrile (3 × 100 mL) and dried under vacuum to afford a fine red powder (3.64 g) (See section 3 for characterization).

2.3.5 PEI-TA-IMINE

PEI (2.02 g) was dissolved in deionized water (20 mL). The aqueous PEI solution was added to acetonitrile (150 mL) under mild stirring. The aqueous solution of **TA** (from section 2.3.3)

was added to the mixture. After 60 hours of reaction, the yellow particles were separated by centrifugation, washed with acetonitrile (3×100 mL) and dried under vacuum to afford a fine yellow powder (4.50 g) (See section 3 for characterization).

2.3.6 PEI-GA-BT-IMINE

PEI (2.02 g) was dissolved in deionized water (20 mL). The aqueous PEI solution was added to acetonitrile (100 mL) under mild stirring. Glutaraldehyde (10 ml, 25 wt% aqueous solution) and **BTA** (3.00 g in 50 ml acetonitrile) was added to the mixture. After 60 hours of reaction, the red particles were separated by centrifugation, washed with acetonitrile (3×100 mL) and dried under vacuum to afford a fine red powder (6.00 g) (see section 3 for characterization).

2.3.7 IMINE REDUCTION

For successive reduction of the imine bonds in the cross-linked structure of the polymer, the dried polymer (2.50 g) and sodium triacetoxymethylborohydride (12.00 g) were mixed in acetonitrile (100 mL) and left with stirring for 12 hours followed by the addition of water (100 mL) and concentrated sulphuric acid (3 mL) to quench the excess of reducing agent. The mixture was then neutralized by the addition of water and K_2CO_3 . This suspension was centrifuged and the precipitate was washed with deionized water (4×200 mL) and acetonitrile (2×200 mL) and finally dried to give the final product: PEI-GA (2.51 g, red powder); PEI-TA (1.87 g, yellow powder); PEI-GA-BT (2.77 g, red powder).

2.3.8 Cu(II) ABSORPTION

To measure the equilibrium Cu(II) absorption capacity, 50 mg of each polymer was treated with 10 ml of aqueous $CuCl_2$ (100 mM) for 12 hours. The particles were centrifuged and washed with deionized water (3×10 ml) and methanol (3×10 ml) and dried to afford the copper loaded samples.

2.3.9 Cu(I)/Cu(II) EXCHANGE AT THE SURFACE

To observe the changes at the surface, 20 mg of each polymer was placed in a tube (5 mm in diameter). Aqueous $CuSO_4$ (1 mM, 1 mL) was then added carefully and the tube was left for 12 hours to allow the surface to absorb copper. The $CuSO_4$ solution was then replaced with 1 mL of a BCS:EDTA:sodium ascorbate solution (2mM:10mM:1mM) for another 12 hours for the reduction and exchange between the copper loaded interface of polymer and the solution to proceed.

2.3.10 EXCHANGEABLE Cu(I) AND UV-VIS

To investigate the ability of the structure of PEI-GA, PEI-TA, and PEI-GA-BT polymer to stabilize Cu(I), 50 mg from each polymer were first loaded with copper by shaking in an aqueous $CuCl_2$ solution (100 mM, 10 mL) for 12 hours. The copper loaded polymers were then centrifuged and washed with deionized water (3×10 mL), and then treated with 5 mL of a BCS:EDTA:sodium ascorbate solution (2mM:10mM:1mM) for additional 12 hours to allow time for equilibrium. The

resulted suspensions were then centrifuged and 1 mL of the aqueous phase was diluted by adding 4 mL of deionized water just before the UV-vis absorbance measurements (250 nm – 800 nm).

2.3.11 DFT CALCULATIONS

All calculations were carried out using PBE functional,³³ combined with a numerical basis set of valence double zeta quality with polarization d-function on non-hydrogen atoms and p-function on hydrogen, DND³⁴ basis 4.4³⁵ in DMOL3 6.0.^{34,36} All geometries were optimized with a SCF tolerance of 1×10^{-5} Hartree and k-point separation of 0.08 \AA^{-1} . All energies given are pure electronic.

3 Results and discussion

Upon the reaction of primary amines with aldehyde, imine bonds form while water is eliminated. To minimize the negative effect of the presence of initial water in the reaction environment, the initial cross-linking has been carried out in a relatively large volume of acetonitrile to dilute water. The miscibility of acetonitrile and water prevents the phase separation of this solution. Also, this method provides the flexibility to introduce other functional groups which may not be water soluble, but are still soluble in an organic solvent such as acetonitrile as **BTA**. Nevertheless, after initial formation of the crosslinked polymer particles, this latex solution is centrifuged and the particles were dried to remove water. This together with further reduction of the imine bonds to amine stabilizes the particle and prevents from hydrolysis.

3.1 Elemental analysis of crosslinked polymers

The elemental analysis of the three crosslinked polymers in Table 1 shows that PEI-GA has 12.6 wt% of nitrogen while the value is 20.3 wt% for PEI-TA which has the triazole group as a part of its crosslinker. For the PEI-GA-BT structure, both the nitrogen and carbon content are higher than for PEI-GA due to the additional benzyl and triazole groups.

Table 1. The elemental analysis of the crosslinked polymers

	C (wt%)	H (wt%)	N (wt%)
PEI-GA	47.1	8.5	12.6
PEI-TA	34.0	7.3	20.3
PEI-GA-BT	49.8	7.2	15.5

3.2 IR spectroscopy

As expected, the IR spectra for PEI-GA, PEI-TA, and PEI-GA-BT show almost the same pattern, mainly due to the similar vibrations of the C-N and C=N bonds within the crosslinked structure of these polymers. This is especially true for PEI-GA and the functionalized PEI-GA-BT, where the main difference is found at 717 cm^{-1} , that is the C=C vibrations of the pendant benzyl group in compound **BT**. The remaining major signals originate from amine vibrations.

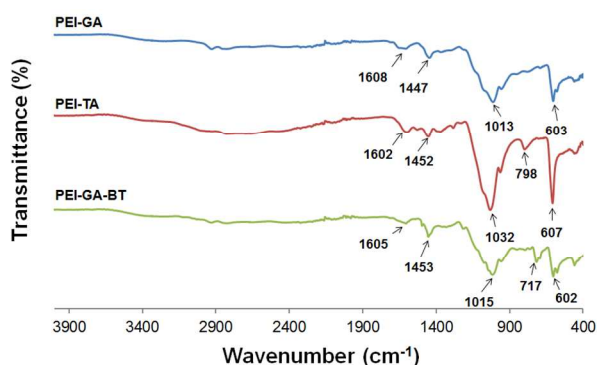


Figure 2. IR spectra for PEI-GA, PEI-GA-BT, and PEI-TA.

3.3 Particle morphology for crosslinked polymers

Figure 3 shows the scanning electron microscopy images of PEI-GA, PEI-TA, and PEI-GA-BT. The effect of the crosslinker on the surface morphology has been already shown for other crosslinking reactions and polymeric systems.³⁷⁻³⁹ During the crosslinking reaction and upon the formation of the cross-linked solid phase, a phase separation occurs.³⁸ The dynamics of the phase separation, which has a determinative role on the morphology of the particles, can be influenced by the crosslinker. In Figure 3, one can observe this difference in the morphology of PEI-GA and PEI-TA. While the PEI-GA structure is more compact, the structure of PEI-TA seems to be more porous. The structure of PEI-GA-BT more resembles PEI-GA, showing that the presence of the BT functional group does not have any strong effect on the final morphology of the particles. For the PEI-GA, there are a small number of spherical particles with varying sizes present. However, these microspheres are less frequent for PEI-GA-BT and absent in the case of PEI-TA.

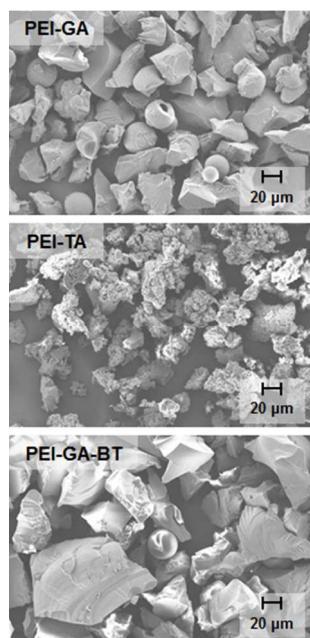


Figure 3. The morphology of the crosslinked polymers PEI-GA, PEI-TA, and PEI-GA-BT.

3.4 Cu(II) absorption

For all the samples, the maximum copper uptake was measured by soaking the polymer in a 100 mM CuCl_2 aqueous solution for 12 hours under vigorous shaking to assure that the equilibrium copper concentration was attained. Elemental analysis of the Cu(II) loaded polymers in Table 2 shows that reduction of the imine bonds reduces the capacity for copper uptake; PEI-GA absorbs 8.1 wt% Cu(II) and 4.5 wt% before and after reduction while for PEI-TA the corresponding values are 12.2 wt% and 5.8 wt% before and after the reduction step. PEI-GA-BT absorbs 3.5 wt% of Cu(II) before and 3.2 wt% after the reductive amination modification. The decrease in Cu(II) absorption upon reduction is probably correlated to the protonation of amine groups in the crosslinked structure. Although this effect can be minimized by optimizing the reduction process, it is ultimately a choice between the durability of the final structure and its copper uptake performance.

Table 2. The absorption capacity for Cu(II) for the cross-linked polymer systems

	Absorption capacity for Cu(II) (wt%) Before imine reduction	Absorption capacity for Cu(II) (wt%) After imine reduction
PEI-GA	8.1	4.5
PEI-TA	12.2	5.8
PEI-GA-BT	3.5	3.2

As it has been already pointed out, the higher capacity for the TA cross-linker is due to the presence of additional triazole functionalities, providing more binding sites for the copper ions. Also the more porous structure of PEI-TA, as compared to PEI-GA and PEI-GA-BT, provides an additional contribution to the higher Cu(II) absorption of this polymer as the Cu(II) absorption occurs mostly at the surface of the particle. Also, it can be seen from Figure 3 that the PEI-GA-BT particles are on average larger than the PEI-GA particles, which explains the lower Cu(II) uptake of this polymer.

3.5 Absorption selectivity

The selectivity of the cross-linked polymers toward copper is important when a flux of copper at the interface is aimed for, otherwise the coordinating sites in the structure may be occupied by other competing metals ions. It has already been shown however, that a thin film of crosslinked PEI-GA can absorb copper from a mixture of metal ions with high selectivity.²⁹ To investigate the selectivity of the crosslinked polymers PEI-GA, PEI-TA, and PEI-GA-BT upon metal coordination, the particles were soaked in 100 mM aqueous solutions of CuCl_2 , ZnCl_2 , and $\text{CuCl}_2/\text{ZnCl}_2$ and the metal content of these samples were measured by elemental analysis. The results are presented in Table 3. Both PEI-GA and PEI-TA

were found to have a very low zinc absorption capacity after reaching equilibrium. PEI-GA and PEI-TA absorb less than 0.2 wt% when there is just ZnCl₂ present in the solution. For the mixed solution of CuCl₂/ZnCl₂, almost no zinc was present in the samples, indicating the strong preference toward Cu(II) versus Zn(II) for both PEI-GA and PEI-TA. The ‘harder’ nature of Zn(II) compared to Cu(II) on one side, and the lack of hard oxygen donor groups in these systems may be a reason for the preference of copper over zinc.

Table 3. The comparative absorption capacity for Cu(II) and Zn(II) from a 100 mM aqueous solution of MCl₂ (M = Cu(II), Zn(II))

	Metal salt present in the solution	Absorption capacity for Cu(II) (wt%)	Absorption capacity for Zn(II) (wt%)
PEI-GA	CuCl ₂	4.5	-
	ZnCl ₂	-	0.2
	CuCl ₂ /ZnCl ₂	4.5	0.0
PEI-TA	CuCl ₂	5.8	-
	ZnCl ₂	-	0.1
	CuCl ₂ /ZnCl ₂	6.3	0.0
PEI-GA-BT	CuCl ₂	3.2	-
	ZnCl ₂	-	0.7
	CuCl ₂ /ZnCl ₂	1.1	1.0

For PEI-GA-BT however the selectivity for Cu(II) versus Zn(II) is not as good as the other two crosslinked system. PEI-GA-BT absorbs 0.7 wt% Zn(II) when there is only ZnCl₂ is present and 1.0 wt% when both CuCl₂ and ZnCl₂ are present in the solution. The presence of ZnCl₂ also affects the Cu(II) absorption capacity for the PEI-GA-BT crosslinked solution, reducing the Cu(II) content to 1.1 wt% which is less than half

of the copper content of this polymer when there is just CuCl₂ is present in the solution. It has already been shown that for a thin film of glutaraldehyde-crosslinked PEI, even though the dynamic of Zn(II) absorption is much faster at the initial absorption stage, the Cu(II) ions will eventually occupy the coordination sites in the crosslinked network.^{29,30} Our results from Table 3 indicates that although such a dominance by Cu(II) ions at equilibrium is the case for both PEI-GA and PEI-TA, the coordination sites will be distributed almost evenly between Zn(II) and Cu(II) for PEI-GA-BT at equilibrium.

3.6 Computational models of the triazolealdehyde (TA) and glutaraldehyde (GA) crosslinkers

The triazole monomer unit in PEI-TA can make a maximum of two coordinations to one Cu ion, but more than one unit can complex to one Cu ion to make a full coordination sphere. Figure 4a shows a selection of probable binding geometries together with the corresponding complexation energies of the complexes of the model ‘monomers’ for PEI-GA and PEI-TA. It can be seen that the complexation energy for PEI-TA is higher than PEI-GA, i.e. Cu(I) reaches lower energy upon coordination to PEI-TA in comparison with PEI-GA. This is also in consistent with the experimental observations for the crosslinked system. Also, PEI-TA offers more possible coordination sites for Cu ions as opposed to PEI-GA (see Figure 4b). Furthermore, and as it is shown in Figure 4c, a lower energy is achieved when the second ‘PEI-TA monomer’ participates in the coordination sphere. This is a more realistic situation that one may also find in a crosslinked PEI-TA network.

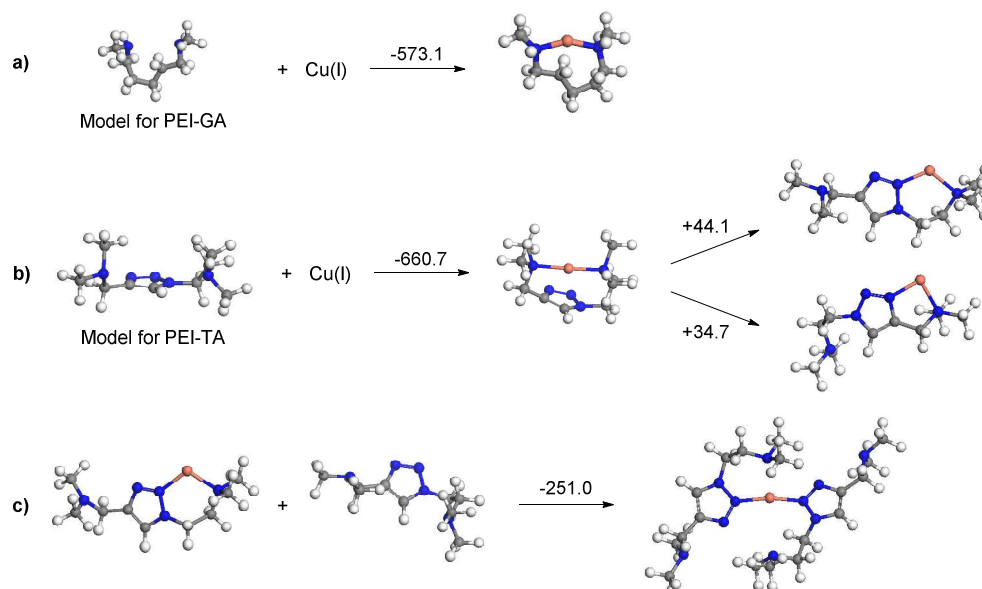


Figure 4. Complexation energies of Cu(I) in kJ·mol⁻¹ with (a) monomers of PEI-GA; (b) monomer of PEI-TA; and (c) dimer of PEI-TA.

3.7 Cu(I) stabilization and exchange

In principle, Cu(I) can be produced *in-situ* by the addition of a mild reducing agent such as sodium ascorbate. This procedure comes with a classic disappearance of the blue colour upon

Cu(II) reduction. However, we have employed a titrating solution that not only contains sodium ascorbate, but also two more ligands that can coordinate strongly and specifically to Cu(I) or Cu(II). In this manner, we can also determine the ability of the polymer structures to retain Cu(I) upon competition with another strongly Cu(I) binding ligand bathocuproine (BC) (Figure 5a). (In this work, we have used the water-soluble bathocuproinedisulfonic sodium salt (BCS) (Figure 5b)). BCS shows a distinct red colour upon coordination to Cu(I) (absorbance wavelength at 488 nm). This has made BCS a useful compound for spectrophotometric determination of Cu(I).⁴⁰ So, even though the quantitative measurement of Cu(I) is not feasible with this spectrophotometric method when a strong Cu(I)-coordinating ligand is present in the environment, the method can still provide valuable information for comparing different Cu-coordinating systems which has been prepared in the course of this work. Also, the strong Cu(I) preference of BCS can shift the $\text{Cu(II)} \rightleftharpoons \text{Cu(I)}$ equilibrium in solution. To minimize this effect, one can use ethylenediaminetetraacetic (EDTA) (Figure 5c) which is able to coordinate specifically and rapidly to Cu(II).⁴⁰

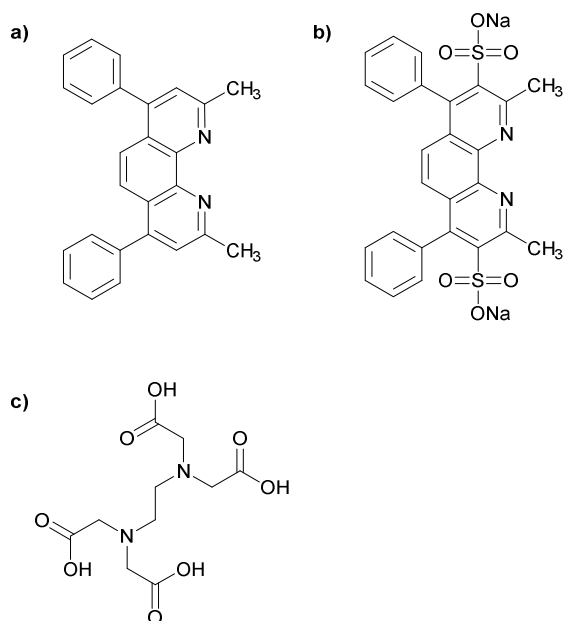
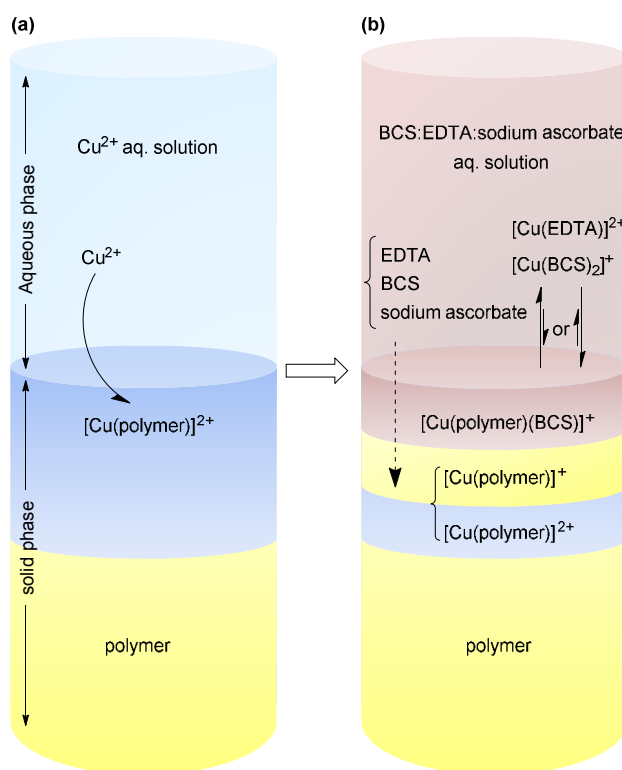


Figure 5. (a) Bathocuproine (BC); (b) bathocuproinedisulfonic sodium salt (BCS); (c) ethylenediaminetetraacetic acid (EDTA).

A schematic representation of a system in which a Cu(II) loaded surface is treated with a solution of BCS:EDTA:sodium ascorbate is shown in Scheme 3. In such a system, there is a competition between different ligands in the solution and at the polymer-solution interface to coordinate to Cu ions. At equilibrium, a typical system is expected to contain a mixture of Cu(I) and Cu(II) coordinated to the different ligands both in solution and in the solid phase. For such a system, the conditional stability constants of the $[\text{Cu}(\text{ligand})]$ complexes as

well as the $[\text{Cu}(\text{ligand } 1)(\text{ligand } 2)]$ ternary complex system are important for the final equilibrium state of the system.



Scheme 3. Schematic representation of (a) the Cu(II) absorption from an aqueous solution of Cu(II); (b) some of the important interactions between the titrating solution of BCS:EDTA:sodium ascorbate and a Cu-loaded surface. The structure-property of the polymer has a determinative influence on the $[\text{Cu}^{\text{I}}(\text{BCS})(\text{polymer})] = [\text{Cu}^{\text{I}}(\text{BCS})_2]$ at equilibrium state.

Figure 6 shows copper soaked PEI-GA, PEI-TA, and PEI-GA-BT polymers before and after exposure to BCS:EDTA:sodium ascorbate aqueous solution. To mimic the conditions as in a typical coating, this experiment was carried out without shaking. The diffusion of sodium ascorbate causes a gradient of Cu(I) and Cu(II) across the polymer. For all these three polymers, the system follows Scheme 3 to different extents. However, the Cu(II) oxidation state gradient is mostly visible for PEI-TA where three different colour waves appear upon the diffusion of sodium ascorbate and the coordinating ligands, i.e. the red top layer which is the ternary complex of $[\text{Cu}^{\text{I}}(\text{BCS})(\text{PEI-TA})]$, the yellow middle layer which has lost the blue colour due to the diffusion of sodium ascorbate, and finally the blue bottom layer which has not yet been influenced by the diffusion layer. For PEI-GA and PEI-GA-BT, however, the dark colour of the original polymers makes such an observation more difficult.

To exclude the diffusion effect and to make some comparisons between these crosslinked structures, the same experiment as in Figure 6 was repeated with shaking. In this case, 50 mg of each polymer was loaded with Cu(II) ions. After washing with deionized water, the polymer was shaken for 12 hours in BCS:EDTA:sodium ascorbate (2mM:10mM:1mM) and the

UV-vis absorption spectra of the solution phase was measured. As shown in Figure 7, even though this experiment is also qualitative in nature, while keeping all other parameters constant, the red colour evolution in the solution together with analysis of the sediment is an indication of the ability of the crosslinked structure to retain Cu(I) upon reduction of Cu(II) ions with sodium ascorbate in competition with a water soluble ligands such as BCS and EDTA. The UV-vis spectra of the titrating solutions in Figure 7d clearly shows that PEI-GA has a

much lower capacity to retain Cu(I) in comparison to PEI-TA which in turn retains almost all the Cu(I) within the structure. PEI-GA-BT stands somewhere in between PEI-GA and PEI-TA. This also suggests that the partitioning of Cu(I) between the crosslinked polymer phase and the solution that contains a strong Cu(I)-coordinating compound can be tuned by introducing a third functionality into the crosslinked PEI-GA system.

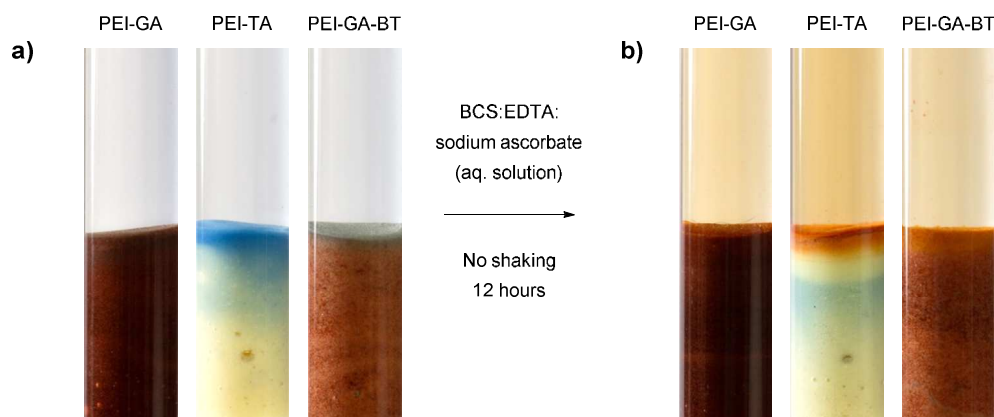


Figure 6. (a) Cu(II) absorption from a 1 mM CuSO_4 aqueous solution under stagnant condition for 12 hours; (b) reduction by sodium ascorbate for 12 hours by a BCS:EDTA:sodium ascorbate (2mM:10mM:1mM).

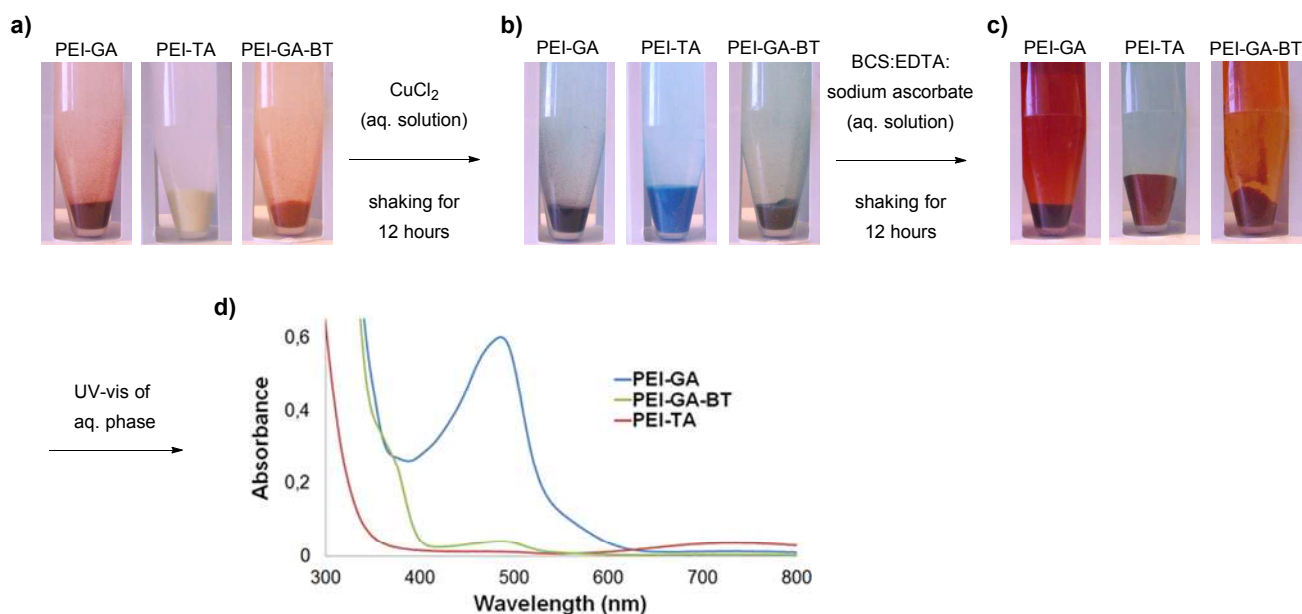


Figure 7. (a) PEI-GA, PEI-TA, and PEI-GA-BT; (b) Cu(II) loaded PEI-GA, PEI-TA, and PEI-GA-BT; (c) titration of Cu-loaded PEI-GA, PEI-TA, and PEI-GA-BT crosslinked polymers with BCS:EDTA:sodium ascorbate solution; (d) UV-vis spectroscopy of the aq. phase from the titration step.

The Cu content of the sediment in Figure 7c has been also measured using EDS. The results are presented in Table 4. As can be seen, the Cu(I) content in the case of PEI-TA is the highest (3.6 wt%) while it was found to be lower for both PEI-GA and PEI-GA-BT (2.2 wt% and 1.4 wt% respectively).

Table 4 The copper content and weight increase of polymer sediments after titration of Cu(II)-coordinated polymers with BCS:EDTA:sodium ascorbate solution.

	Cu(I) content in the polymer sediment after titration (wt%)*	Weight increase of the sediment after titration (%)
PEI-GA	2.2	4
PEI-TA	3.6	20
PEI-GA-BT	1.4	4

(*measured by EDS)

The weight-percent of Cu(I) in all of the polymer networks are lower after the titration. One reason for this, as stated earlier, is that the copper ions leave the polymer structure upon formation of water-soluble $[\text{Cu}(\text{BCS})_2]^+$ and/or $[\text{Cu}(\text{EDTA})]^{2+}$ complexes, which is mainly the case for PEI-GA and PEI-GA-BT. However, for PEI-TA, the coordination of BCS to the copper ions in the polymer structure is the main reason that the total Cu(I) weight percent in the final sediment is lower. This happens to a much lesser extent for PEI-GA and PEI-GA-BT. Measuring the total weight increase of the polymer sediments after titration also demonstrates that although the total weight increase is 4% for PEI-GA and PEI-GA-BT, the weight of PEI-TA increases by 20% after titration, mainly due to the coordination of BCS to the Cu ions in the polymer network.

4 Conclusions and outlook

We have shown that chemical modification of polyethyleneimine with a triazole-based crosslinker results in a material with a very strong affinity to Cu(I) and Cu(II) which are the most common and biologically most important oxidation states of copper. In particular, the crosslinker increased the affinity to Cu(I). Furthermore, we have prepared a polyethyleneimine-glutaraldehyde-(benzyl)triazole polymer that allows effective release of Cu(I). Finally, we have shown that a combination of the polymers can be used to fine tune the uptake and release of copper across an interface, which opens up for a new approach to antibiofouling. Our proposed antibiofouling mechanism has the potential to lead to biocide-free antibiofouling coatings for a wide range of freshwater applications as well as static marine constructions and vessels, with the added benefit of remediating copper from copper-contaminated harbours once a vessel is exposed to the open sea. Yet there are also three significant environmental dimensions of this science. First, the advance should enable us to supplant the use of existing coatings, which release highly toxic substances into the ocean, causing untold environmental damage. Second, the new generation of coatings will minimise the spread of invasive organisms on the hulls of ocean-going vessels. The third environmental benefit is related to the deleterious effects of copper on marine and aquatic biodiversity: in the longer term, the technology will significantly decrease copper concentrations in polluted water, resulting in increased biodiversity.

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1. M. R. Bleackley and R. T. A. MacGillivray, *Biometals*, 2011, **24**, 785-809.
2. K. Balamurugan and W. Schaffner, *Biochim. Biophys. Acta-Mol. Cell Res.*, 2006, **1763**, 737-746.
3. M. M. O. Pena, J. Lee and D. J. Thiele, *J. Nutr.*, 1999, **129**, 1251-1260.
4. E. Almeida, T. C. Diamantino and O. de Sousa, *Prog. Org. Coat.*, 2007, **59**, 2-20.
5. L. D. Chambers, K. R. Stokes, F. C. Walsh and R. J. K. Wood, *Surf. Coat. Technol.*, 2006, **201**, 3642-3652.
6. N. Voulvoulis, M. D. Scrimshaw and J. N. Lester, *Appl. Organomet. Chem.*, 1999, **13**, 135-143.
7. E. Ytreberg, J. Karlsson and B. Eklund, *Sci. Total Environ.*, 2010, **408**, 2459-2466.
8. W. G. Sunda and R. R. L. Guillard, *J. Mar. Res.*, 1976, **34**, 511-529.
9. V. F. Vetere, M. C. Perez, R. Romagnoli, M. E. Stupak and B. del Amo, *J. Coat. Technol.*, 1997, **69**, 39-45.
10. L. Ciavatta, D. Ferri and R. Palombi, *J. Inorg. Nucl. Chem.*, 1980, **42**, 593-598.
11. J. W. Moffett and R. G. Zika, *Mar. Chem.*, 1983, **13**, 239-251.
12. J. F. Jiang, I. A. Nadas, M. A. Kim and K. J. Franz, *Inorg. Chem.*, 2005, **44**, 9787-9794.
13. Y. Nose, E. M. Rees and D. J. Thiele, *Trends Biochem. Sci.*, 2006, **31**, 604-607.
14. K. D. Karlin, J. C. Hayes, S. Juen, J. P. Hutchinson and J. Zubieta, *Inorg. Chem.*, 1982, **21**, 4106-4108.
15. M. A. Trojer, A. Movahedi, H. Blanck and M. Nyden, *J. Chem.*, 2013.
16. J. T. Rubino and K. J. Franz, *J. Inorg. Biochem.*, 2012, **107**, 129-143.
17. T. R. Chan, R. Hilgraf, K. B. Sharpless and V. V. Fokin, *Org. Lett.*, 2004, **6**, 2853-2855.
18. P. S. Donnelly, S. D. Zanatta, S. C. Zammit, J. M. White and S. J. Williams, *Chem. Commun.*, 2008, 2459-2461.
19. A. Movahedi, K. Moth-Poulsen, J. Eklöf, M. Nyden and N. Kann, *React. Funct. Polym.*, 2014, **82**, 1-8.
20. J. Jia, A. H. Wu and S. J. Luan, *Colloid Surf. A-Physicochem. Eng. Asp.*, 2014, **449**, 1-7.
21. B. L. Rivas, E. D. Pereira and I. Moreno-Villoslada, *Prog. Polym. Sci.*, 2003, **28**, 173-208.

22. F. Sabermahani and M. A. Taher, *Anal. Chim. Acta*, 2006, **565**, 152-156.
23. Y. L. Chen, B. C. Pan, S. J. Zhang, H. Y. Li, L. Lv and W. M. Zhang, *J. Hazard. Mater.*, 2011, **190**, 1037-1044.
24. W. J. Tong, C. Y. Gao and H. Möhwald, *Polym. Adv. Technol.*, 2008, **19**, 817-823.
25. J. Jia, A. H. Wu and S. J. Luan, *Phys. Chem. Chem. Phys.*, 2014, **16**, 16158-16165.
26. D. M. Saad, E. M. Cukrowska and H. Tutu, *Toxicol Environ. Chem.*, 2011, **93**, 914-924.
27. D. M. G. Saad, E. M. Cukrowska and H. Tutu, *Toxicol Environ. Chem.*, 2012, **94**, 1916-1929.
28. D. M. G. Saad, E. M. Cukrowska and H. Tutu, *Toxicol Environ. Chem.*, 2013, **95**, 409-421.
29. J. B. Linden, M. Larsson, B. R. Coad, W. M. Skinner and M. Nyden, *RSC Advances*, 2014, **4**, 25063-25066.
30. S. J. Miklavcic, M. Nyden, J. B. Linden and J. Schulz, *RSC Advances*, 2014, **4**, 60349-60362.
31. A. F. AbdelMagid, K. G. Carson, B. D. Harris, C. A. Maryanoff and R. D. Shah, *J. Org. Chem.*, 1996, **61**, 3849-3862.
32. T. R. Chan and V. V. Fokin, *QSAR Comb. Sci.*, 2007, **26**, 1274-1279.
33. J. P. Perdew, K. Burke and M. Ernzerhof, *Phys. Rev. Lett.*, 1996, **77**, 3865-3868.
34. B. Delley, *J. Chem. Phys.*, 1990, **92**, 508-517.
35. B. Delley, *J. Phys. Chem. A*, 2006, **110**, 13632-13639.
36. B. Delley, *J. Chem. Phys.*, 2000, **113**, 7756-7764.
37. D. Kim, D. Y. Lee, K. Lee and S. Choe, *Macromol. Res.*, 2009, **17**, 250-258.
38. K. Kim, Y. Kim, N. R. Ko and S. Choe, *Polymer*, 2011, **52**, 5439-5444.
39. B. Peng and A. Imhof, *Soft Matter*, 2015, **11**, 3589-3598.
40. J. W. Moffett, R. G. Zika and R. G. Petasne, *Anal. Chim. Acta*, 1985, **175**, 171-179.