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Rapid Detection of Cocaine, Benzoylecgonine and Methylecgonine in Fingerprints using Surface
Mass Spectrometry

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Fingerprints, drug testing, cocaine, imaging mass spectrometry

Abbreviations

GC- MS Gas Chromatography Mass Spectrometry

BZE Benzoylecgonine

EME Methylecgonine

DESI Desorption Electrospray Ionisation Mass Spectrometry

MALDI-IMS-MS/MS Matrix Assisted Laser Desorption Ion Mobility Tandem Mass Spectrometry

SIMS Secondary Ion Mass Spectrometry

LC-MS Liquid Chromatography Mass Spectrometry

SALDI Surface Assisted Laser Desorption Ionisation

Abstract

Latent fingerprints provide a potential route to the secure, high throughput and non-invasive detection of drugs of abuse. In this study we show for the first time that the excreted metabolites of drugs of abuse can be detected in fingerprints using ambient mass spectrometry. Fingerprints and oral fluid were taken from patients attending a drug and alcohol treatment service. Gas chromatography mass spectrometry (GC-MS) was used to test the oral fluid of patients for the presence of cocaine and benzoylecgonine. The corresponding fingerprints were analysed using Desorption Electrospray Ionization (DESI) which operates under ambient conditions and Ion Mobility Tandem Mass Spectrometry Matrix Assisted Laser Desorption Ionization (MALDI-IMS-MS/MS) and Secondary Ion Mass Spectrometry (SIMS). The detection of cocaine, benzoylecgonine (BZE) and methylecgonine (EME) in latent fingerprints using both DESI and MALDI showed good correlation with oral fluid testing. The sensitivity of SIMS was found to be insufficient for this application. These results provide exciting opportunities for the use of fingerprints as a new sampling medium for secure, non-invasive drug detection. The mass spectrometry techniques used here offer a high level of selectivity and consume only a small area of a single fingerprint, allowing repeat and high throughput analyses of a single sample.

Introduction

The drug testing industry is worth several billion dollars worldwide, and is routinely used by probation services, prisons, courts and other law enforcement agencies. Recently there has been a push towards workplace drug testing as well as new initiatives to test motorists for drug driving [1]. Drug testing is usually carried out by taking a sample of blood or urine from a suspect and using either an antibody assay or chromatographic analysis to detect the relevant drug and its metabolites. However, these methods of sampling have limitations – blood testing can require trained staff, urine testing has associated privacy concerns and in both cases the samples must be treated as a biological hazard, which increases the complexity of sample handling in terms of storage and disposal. Oral fluid and sweat have been proposed as alternative non-invasive collection matrices. These matrices, as well as oral fluid and sweat, frequently require extraction steps from the collection devices or precipitation from the biological fluids prior to analysis and this contributes to the cost of analysis [2].

In contrast, a latent fingerprint can be deposited quickly and transported easily. The identity of the donor is encapsulated within the fingerprint ridge detail, making the test impossible to falsify. It has recently been shown that drugs and their metabolites can be detected in latent fingerprints using antibody reagents [3]. Whilst antibody reagents provide a rapid screening test, non-specific binding can lead to false positive results. Mass spectrometry techniques provide a higher level of specificity, providing confirmation of the identity of the substance detected.

Recent work by Goucher et al [4] has shown that liquid chromatography mass spectrometry (LC-MS) can be used to detect lorazepam, methadone and their metabolites in latent fingerprints. However, a positive detection was only achieved when ten fingerprints were used (and consumed) in combination, making it impractical for use in the field as this precludes repeat analysis. In contrast, recent developments in surface mass spectrometry allow *in-situ* analyses of a small area (less than 100 x 100 μm) of a sample under atmospheric pressure conditions. This provides the potential for rapid sample throughput (often a few seconds per sample, compared with several hours for LC-MS when sample preparation is taken into account), a demonstration that the substance of abuse resides in the fingerprint and not the substrate and the opportunity to take repeat tests. These techniques have been applied previously to fingerprints that have been deliberately contaminated by drugs via contact [5-7] but (to our knowledge) not to unambiguously confirm the presence of drug metabolites in the fingerprints of drug users. In order to confirm ingestion of a drug (as opposed to contact), detection of drug metabolites is essential. Rowell et al [8] used surface assisted laser desorption ionization (SALDI) under vacuum conditions to detect the major metabolite of methadone in natural fingerprints, and was unable to confirm its presence using tandem mass spectrometry.

Of the many surface mass spectrometry techniques available, matrix assisted laser desorption ionization (MALDI) [9-11], desorption electrospray ionization (DESI) [5] and secondary ion mass spectrometry (SIMS) [12-14] have been previously shown to provide chemical images of endogenous and exogenous compounds in a latent fingerprint. In this context, the techniques offer complementary features, these being high sensitivity (MALDI), quantitative precision (SIMS) and high throughput (DESI). All three of the techniques can operate under atmospheric pressure conditions, allowing high throughput analysis. In this paper, we will show for the first time that

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2 cocaine and its metabolites benzoylecgonine (BZE) and methylecgonine (EME) can be detected by
3 these surface mass spectrometry techniques in latent fingerprints, and in particular under
4 atmospheric pressure conditions, offering an exciting new opportunity for drug testing.
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8 **Materials and Methods**

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10 Cocaine, EME, acetone and acetonitrile (ACN) were obtained from Sigma Aldrich (St. Louis,
11 USA). BZE was obtained either from Sigma Aldrich (St. Louis, USA) or from Sigma Aldrich
12 (Poole, UK). Methanol was obtained either from Rathburn (Walkerburn, UK) or from Sigma
13 Aldrich (Poole, UK). Trifluoroacetic acid (TFA), and α -cyano-4-hydroxycinnamic acid (α -CHCA)
14 were purchased from Sigma Aldrich (Poole, UK). Double-side conductive carbon tape was obtained
15 from TAAB (Aldermaston, UK).
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19 **Sample preparation**

20 *Latent Fingerprints*

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22 Latent fingerprints were obtained from five individuals who were attending a drug and alcohol
23 treatment service to receive treatment for drug dependence. "Natural" fingerprints were deposited
24 onto clean glass slides at a pressure between 400-1000 g and barcoded, before being shipped to the
25 two analyzing laboratories. The fingerprints were then stored in a freezer at each lab at -80°C. A
26 corresponding oral fluid sample was collected from each patient using a Quantisal[™] collection kit,
27 Alere Toxicology, UK. The oral fluid samples were screened for cocaine and benzoylecgonine
28 BZE using a standard drug screening procedure utilizing GC-MS/MS at LGC Forensics,
29 Teddington, Middlesex. A favourable ethical opinion for sample collection and analysis was
30 granted from the National Research Ethics Service. In line with the ethics protocol, the patients
31 were not asked whether they had used drugs.
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37 *MALDI*

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39 For MALDI analyses, the fingerprints were taken out of storage and a 0.5 μ L BZE standard solution
40 at a concentration of 100 μ g/mL in 1:9 methanol: water mixed 1:1 with a solution of 5 mg/mL α -
41 CHCA in 70:30 ACN: 0.5% TFA was then spotted onto an area of the glass slide away from the
42 mark. Both the fingerprint and the standard on the glass slide were allowed to dry in a vacuum
43 desiccator for 20 minutes prior to analysis. The entire fingermark was spotted at random points with
44 the MALDI matrix before spectral acquisition. Fingerprints were analysed approximately 3 months
45 after deposition.
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49 *DESI and SIMS*

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51 Gels containing standards of cocaine, BZE and EME at concentrations of 5, 50, 500 and 5000 ng/ml
52 were pipetted onto glass slides and smeared across the slides to create a uniform layer. The gels,
53 which were a few mm in thickness, were made of synthetic gelatine (Sigma Aldrich, UK) and
54 analysed using DESI and SIMS. No sample preparation was required for the fingerprint samples.
55 Fingerprints were analysed approximately 1 month after deposition.
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59 **Instrumentation**

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MALDI ion mobility tandem mass spectrometry (MALDI-IMS-MS/MS) analysis was conducted
using a Synapt[™] G2 HDMS mass spectrometer (Waters Corporations, Manchester, UK)

1
2 incorporating a Nd:YAG laser operating at 1 kHz. Spectra were acquired in positive and resolution
3 ('Res') mode in the mass range between m/z 100-1000 with a laser power of 200 (a.u.). Transfer
4 fragmentation was used, allowing for ions to be separated based on their mobility and then
5 subjected to collision induced dissociation in the Transfer T-Wave™. This maintained that product
6 ions retained the same ion drift time as their corresponding precursor ion. Collision energy was
7 applied at 26 eV for the dissociation of BZE. Prior to analysis, calibration was performed in the
8 specified mass range using Glu-1-fibrinopeptide. Data processing of all spectra was performed
9 using MassLynx 4.1 (Waters Corporation, Manchester, UK).
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12 DESI analyses were carried out at the National Physical Laboratory, Teddington using a DESI 2D
13 Omni Spray Ion Source (Prosolia, Indianapolis, IN, USA) and an LTQ Orbitrap Velos mass
14 spectrometer (Thermo Scientific, Bremen, Germany), optimised as outlined in Reference 15. Data
15 were acquired using the Orbitrap mass analyser with the highest resolution setting of 100,000 (at
16 m/z 400), except where stated when the linear ion trap analyser was used. The solvent composition
17 was 90:10 MeOH:H₂O supplied at a flow rate of 1.5 μ L/min using a 5 kV bias. The spray was
18 directed through a 50 micron capillary, giving a spot of approximately 300 x 300 μ m on the sample
19 [16]. Approximately 1 mm² of sample was consumed during the analysis. Spectra were collected in
20 positive ion mode using both full scan mode and MS/MS mode.
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24 SIMS analyses were carried out using an ION-TOF GmbH (Münster, Germany) TOF SIMS 5
25 instrument at the University of Surrey, UK. A 25 keV Bi₃⁺ primary ion beam delivering 0.18 pA of
26 current was employed. Images were acquired at 128 x 128 resolution in the MacroRaster mode of
27 operation. Scans of 500 x 500 μ m were acquired. Image data were acquired using 512 cycles per
28 pixel point with 1 scan per pixel. A cycle time of 100 μ s was employed. Mass calibration was
29 performed by assigning peaks with a known mass using IonSpec software (version 4.1). Calibration
30 was carried out using the following ions: CH₃, Na, C₂H₃, CHO, C₂H₅, C₃H₅, C₂H₃O, C₃H₇, C₂H₅O,
31 C₄H₅, C₃H₃O, C₄H₇, C₃H₅O, C₄H₉, C₆H₅, C₇H₇, and Si₂C₅H₁₅O.
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38 Results

39 MALDI IMS-MS/MS

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42 The ion mobility function can provide superior separation of isobaric ions and sensitivity, as ions
43 are separated, in addition to their m/z , also on the basis of a specific 'drift time' (DT) that is unique
44 to that particular ion as it is linked to their collisional cross section[17]. This allows for the reliable
45 detection of biologically relevant concentrations. In this instance, MALDI-IMS-MS/MS has been
46 employed to confirm the presence of BZE based on the matching of the product ions and their DT's
47 in the fingerprint and in the BZE standard. MALDI IMS-MS/MS analysis of both the standard and
48 the mark allowed for Drift Scope Plots (a graphical representation of the ion mobility separation) to
49 be generated and select DT regions of interest containing both the parent and the corresponding
50 product ions (Figure 1A). Spectra from these acquisitions were interrogated for a product ion
51 derived from the collision induced dissociation of BZE which directly corresponded to those
52 observed in the standard (m/z 168.1033) (Figure 1B). Drift time chromatograms could then be
53 produced based on the DT of this product ion, showing that the BZE product ion in the standard and
54 the mark directly coincided (Figure 1C). These data prove, unequivocally, that it is possible to
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2 detect excreted BZE contained within the fingerprint residue with corresponding oral fluid above
3 the 16 ng/ml cut-off determined by the service provider's GC-MS protocol [18] (although it is
4 unknown at the present what is the lowest limit of detection and limit of quantification for
5 fingerprint residue) and highlight the selectivity and sensitivity of modern analytical techniques
6 such as ion mobility mass spectrometry.
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9 10 11 DESI

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14 Analysis of the standards showed that detectable signals for cocaine and BZE were present above 5
15 ng/ml, and above 50 ng/ml for EME. In Figure 2, spectra are presented which show positive
16 identification of cocaine, BZE and EME in a fingerprint sample. Figure 2A shows the MS/MS
17 spectrum from a fingerprint for ions with m/z 304, with the $(M+H)^+$ peak for cocaine detected at a
18 mass accuracy of 4.8 ppm (in full scan mode this was improved to 0.807 ppm) and the detection of
19 the fragment ion at m/z 182. Similarly, Figure 2B shows the MS/MS spectrum from a fingerprint for
20 ions with m/z 290, with the $(M+H)^+$ peak for BZE detected at a mass accuracy of 5.0 ppm
21 (0.016ppm in full scan mode), and detection of the fragment ion at m/z 168, confirming the
22 presence of BZE in the fingerprint. No signal was detected when the spray was moved away from
23 the fingerprint. Finally, Figure 2C shows the MS/MS spectrum, using the linear ion trap analyser,
24 from a fingerprint for ions with m/z 200, with the $(M+H)^+$ peak for EME and detection of the
25 fragment ion at m/z 182, confirming the presence of EME in the fingerprint. Using the Orbitrap
26 mass analyser in full scan mode, EME was detected with a mass accuracy of 0.200 ppm. Table 1
27 compares the results of oral fluid testing and fingerprint testing from DESI of four donors'
28 fingerprints. Donors 1a and 1b represent different fingerprints from the same donor, and there is
29 direct correspondence between the oral fluid test and the two fingerprints analysed by DESI. A
30 similar result is seen for Donor 3. Donor 2's oral fluid tested negative, and the absence of a
31 detectable signal in the DESI spectra from Donor 2's fingerprints are in direct agreement with this.
32 The oral fluid test for Donor 4 tested positive for cocaine and this is also detected in Donor 4's
33 fingerprint. However, EME and BZE were not detected in the fingerprints from this donor.
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39 Table 2 presents data relating to the qualitative repeatability of the DESI method. Three
40 fingerprints were taken from each of a further two donors who tested positive for cocaine in their
41 oral fluid. Three separate points were analysed on each fingerprint. Table 2 shows that cocaine and
42 EME were detected at every point analysed. BZE was detected in 17 out of 18 of the analysis
43 points.
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48 SIMS

49 SIMS analysis of the standards revealed peaks at m/z 168, 124, 105 and 82, representing fragments
50 of cocaine, BZE and EME. Molecular ion peaks could be seen only with the highest concentration
51 of the analytes. In the latent fingerprint samples, only the peak at m/z 82 was detected.
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54 Discussion

55 MALDI IMS-MS/MS and DESI analysis of a "natural" latent fingerprint allowed for the successful
56 detection of the excreted primary metabolite, BZE. DESI was also able to detect EME and, at the
57 time of the MALDI analysis, researchers were specifically asked to look for BZE only, therefore no
58 comparative conclusions can be made on the feasibility of MALDI to detect this secondary
59 metabolite.
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1
2 The ability to detect excreted substances in latent fingerprints proves that surface mass spectrometry
3 techniques could provide the drug testing industry with an exciting new and complementary tool,
4 potentially allowing differentiation between drug consumption and contact solely based on the
5 presence of metabolites in the residue. The significance of detecting cocaine metabolites in a
6 fingerprint is currently unknown, because it is not known whether cocaine in fingerprint residue
7 from contact could metabolise on the skin's surface. Furthermore, the possibility of absorption and
8 metabolism of contact residues and the potential for secondary transfer has never been investigated.
9 Although an image of the distribution of these excreted illicit substances in the fingerprint samples
10 has not yet been attempted by either technique, the use of these techniques in profiling mode
11 enables a rapid confirmatory screening for drugs of interest in less than 2 minutes.
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15 The correspondence between the DESI and MALDI results and the oral fluid measurements is
16 promising, but also shows the importance of developing a quantitative test with a cut-off level. The
17 DESI result for Donor 4 shows that whilst this donor's oral fluid was above the oral fluid screening
18 test cut-off level, only cocaine (and not BZE or EME) was detected in the fingerprint. This could
19 be due to a difference in the (currently unknown) detection window for these metabolites in
20 fingerprints compared with oral fluid or it may be a consequence of insufficient sensitivity of the
21 DESI technique.
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24 No quantification of the DESI data was attempted at this stage since a matrix-matched standard
25 does not exist. Additionally, since the fingerprints are not spatially uniform, an appropriate
26 sampling regime should be developed. The difficulty with quantification of a substance within
27 fingerprint residue is that there is currently no accepted method for determining how much material
28 was deposited in the finger deposition. In contrast, in blood, urine or oral fluid, the sample volume
29 is known. It has been shown previously using an LC-MS method to detect caffeine in fingerprints
30 that different fingers gave different results, presumably due to different contact pressure and/or
31 deposition area from different fingers [19]. We are currently investigating the validity of using
32 different endogenous compounds within a fingerprint for normalisation purposes. Further studies
33 are necessary in this area to establish a quantitative procedure.
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37 Whilst SIMS has been previously shown to be a useful technique for imaging endogenous
38 compounds in latent fingerprints [13,14], its suitability for the detection of drug metabolites in
39 fingerprints is questionable, under the analytical conditions used here. The high level of
40 fragmentation results in only the detection of the fragment ion at m/z 82 in fingerprint samples and
41 standards of concentrations below 5000 ng/ml. This fragment is common to cocaine, BZE and
42 EME as well as other compounds, and therefore the selectivity is insufficient for toxicological
43 application. It should be noted however, that Ar cluster SIMS sources can provide significantly
44 higher molecular ion yields than Bi SIMS for organic compounds, and this may yield the desired
45 selectivity in fingerprint samples. If this were the case, SIMS may provide superior opportunities
46 for quantification compared with DESI and MALDI.
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50 **Conclusion**

51 We have demonstrated the use of established and emerging analytical techniques for the analysis of
52 cocaine and its metabolites in natural fingerprints. DESI was carried out under ambient conditions,
53 and atmospheric pressure MALDI systems are also available. This, combined with the ability to
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2 analyse the sample in-situ and with minimal sample preparation allows for very rapid sample
3 throughput compared with chromatographic methods. The high sensitivity of the methods and
4 selectivity to the analytes described has enabled us to detect metabolites of cocaine drugs from
5 those who use illicit substances. Secondly, although the fingerprints are not spatially uniform, we
6 are optimistic that these approaches can be developed for quantitative analysis of fingerprint
7 residues.
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11
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16 sample collection and the National Institute for Health Research for sponsoring this research.
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Donor	Oral fluid Screening Result	DESI fingerprint analysis		
		Cocaine	BZE	EME
DESI				
1a	Positive	y	y	y
1b	Positive	y	y	y
2	Negative	n	n	n
3	Positive	y	y	y
4	Positive	y	n	n
MALDI				
5	Positive	y	y	N/A

Table 1 : Oral fluid test results and corresponding DESI and MALDI signals in the fingerprints of five donors

	Position	Cocaine	BZE	EME
Donor 6				
Lthumb	1	✓	✓	✓
	2	✓	✓	✓
	3	✓	☒	✓
Rindex	1	✓	✓	✓
	2	✓	✓	✓
	3	✓	✓	✓
Rring	1	✓	✓	✓
	2	✓	✓	✓
	3	✓	✓	✓
Donor 7				
Rindex	1	✓	✓	✓
	2	✓	✓	✓
	3	✓	✓	✓
Rmiddle	1	✓	✓	✓
	2	✓	✓	✓
	3	✓	✓	✓
Rthumb	1	✓	✓	✓
	2	✓	✓	✓
	3	✓	✓	✓

Table 2 : DESI signals for cocaine, BZE and EME across three positions on three different fingerprints of two donors

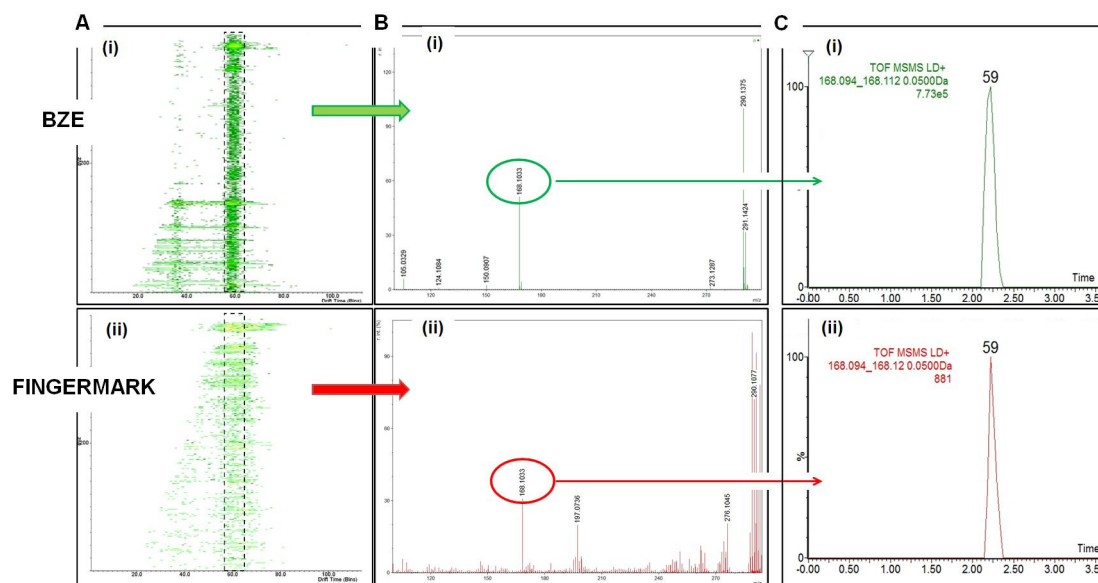


Figure 1 Detection of BZE from a fingerprint from a drug user in a rehabilitation centre by MALDI IMS-MS/MS. (Figure 1A) Drift scope plots of BZE standard and BZE in the fingerprint (generated through transfer fragmentation experiment). (Figure 1B) MS/MS spectra of BZE standard and BZE in the fingerprint after the selection of the BZE parent ion and products ion same drift time plume in the fingerprint and in the standard. Selection of the ion product at m/z 168.1033 in common to the BZE standard and BZE in fingerprint showing superimposable drift time chromatograms.

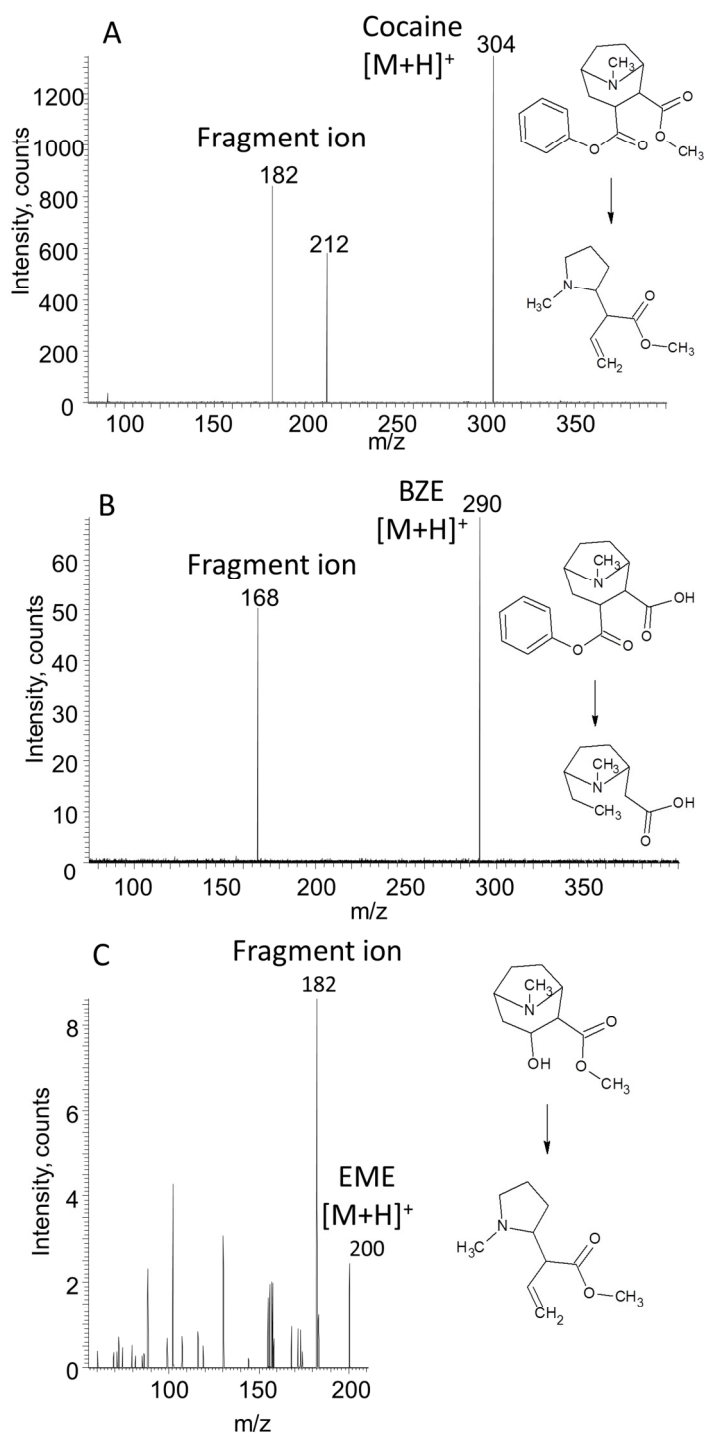


Figure 2 Detection of cocaine, BZE and EME in a fingerprint from a drug user attending a rehabilitation centre using DESI MS/MS. Spectra from a fingerprint with ions of m/z (Figure 1A) 304 (cocaine) (Figure 1B) 290 (BZE) (Figure 1C) 200 (EME)