# Chemical Science

## Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/chemicalscience

## Chemical Science

### EDGE ARTICLE

## Ultra-Sensitive pH Control of Supramolecular Polymers and Hydrogels: pK<sub>a</sub> Matching of Biomimetic Monomers

B. J. Cafferty, R. R. Avirah, G. B. Schuster and N. V. Hud\*

Achieving self-assembly in water with small molecules that disassemble in response to minimal changes in environmental conditions is important for the development of many promising materials. We report the most sensitive possible pH control of a supramolecular polymer assembly - demonstrated with hydrogel-forming monomers that mimic nucleic acid base pairing. Bidirectional pH-responsive supramolecular polymers are formed upon the self-assembly of three monomers with two pK<sub>a</sub> matched recognition units (one acidic and one basic). These supramolecular assemblies are most stable when the pH equals the  $pK_a$  of the monomers, but disassemble in response to small pH changes near neutrality (exhibiting the theoretical limit for pH-dependent stability). At pH 7 a hydrogel forms in solutions that are 0.7% by weight in monomer, however, the hydrogel dissolves at pH 6 and pH 8. Additionally, we show that hydrogel stability is finely tuned by employing monomer mixtures that frustrate formation of insoluble aggregates. These results illustrate the advantages of using  $pK_a$  matched recognition elements and polymer heterogeneity in the development of responsive materials. Finally, these same recognition elements have recently been proposed as possible ancestors of the current bases of RNA. In this context, the assemblies described here might also provide clues to how the first genetic polymers were driven between states of base pairing and non-base pairing by periodic changes in environmental pH.

Cite this: DOI: 10.1039/x0xx00000x

Received ooth January 2012, Accepted ooth January 2012

DOI: 10.1039/x0xx00000x

www.rsc.org/

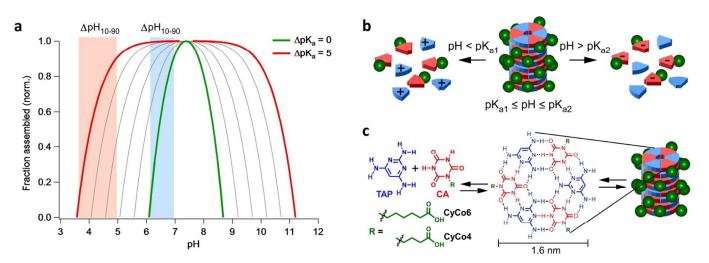
#### Introduction

Precise control over the self-assembly of small molecules in water underlies developmental strategies for many promising materials,<sup>1, 2</sup> including those for use in drug delivery and tissue engineering.<sup>3, 4</sup> Control of supramolecular polymer assembly and hydrogel formation by low molecular weight gelators can be achieved by using monomers with intermolecular interactions that are altered by environmental changes, such as pH, temperature and the addition of other molecules or ions.<sup>5-8</sup> Unfortunately, it is the case with many low molecular weight gelators that the same interactions that enable hydrogelation also lead to the formation of precipitates and, overtime, gel collapse.<sup>9, 10</sup> Therefore, new approaches that allow precise control over the supramolecular assembly process and increased gel lifetime are critical for supramolecular systems to be integrated into functional materials.

An appealing approach to the design of self-assembling structures for biological applications is the creation of supramolecular polymers that respond in a highly non-linear manner to modest pH changes within the physiological range.<sup>11</sup> Bidirection pH control of self-assembly has been demonstrated for monomers that contain acidic and basic groups that promote assembly through charge pairing within the pH range between the pK<sub>a</sub>s of the charged groups.<sup>12, 13</sup> The pH range over which such an assembly is stable and responsive, typically defined as  $\Delta pH_{10.90}$  (where 10-90% of the assemblies are present in solution), is coupled to the pK<sub>a</sub>s of the acidic and basic recognition elements. The response of a system to changes in pH can therefore be enhanced by minimizing the  $\Delta pK_a$  of the

recognition elements (Fig. 1a and ESI). While bidirectional systems commonly employ amines and carboxylic acids to enable charge pairing, these functionalities do not have  $pK_{as}$  near pH 7 and have a  $\Delta pK_{a}$  of at least 5, making these systems mostly insensitive to pH changes near neutrality (Fig. 1a). We hypothesized that maximum pH sensitivity for a supramolecular polymerization process that responds to pH changes that are physiologically relevant could be achieved with assemblies that employ recognition elements with  $pK_{as}$  matched near pH 7 (Fig. 1b).

Work on the coassembly of monomers containing the triazines melamine and cyanuric acid (Cy), or their related pyrimidines triaminopyrimidine (TAP) and barbituric acid, as complementary hydrogen-bonding recognition units has fundamentally advanced our understanding of artificial self-assembly,9-11 particularly in organic solvents where initial investigations were performed. In water, precipitation occurs immediately upon combining any two of these complementary heterocycles, but the addition of a hydrophilic side chain to one or both of the recognition elements promotes the formation of water soluble supramolecular assemblies.<sup>14, 15</sup> Of these heterocycles, TAP and Cy are unique as they are complementary and pK<sub>a</sub> matched near neutrality.<sup>14</sup> Critically, Cy is predominantly negatively charged above this shared  $pK_a$ , while **TAP** is positively charged below it, precisely the properties necessary for a selfassembling system with ultimate possible sensitivity to pH. Additionally, TAP has recently been shown to readily react with ribose to form nucleosides under plausibly prebiotic conditions that assemble with Cy to form supramolecular polymers and hydrogels, which has implications regarding the origin of the nucleobases of the hypothetical 'pre-RNA world'.<sup>16</sup>



**Fig. 1** (a) Theoretical limits for pH-dependent bidirectional assembly systems with  $\Delta pK_as$  ranging from 0 to 5. The sensitivity of the assembly to change in pH,  $\Delta pH_{10,30}$ , is highlighted in blue for  $\Delta pK_a = 0$  (pK<sub>a</sub> matched), and pink for  $\Delta pK_a = 5$  (the minimal  $\Delta pK_a$  for assembly systems that use amines and carboxylic acids for charge pairing). Note that the pH range over which assemblies are present increases with increasing  $\Delta pK_a$ ; the system with  $\Delta pK_a =$ 0 shows the greatest sensitivity to changes in pH. Calculations were performed using idealized model systems that all have a minimum assembly concentration (MAC) of 15 mM in each monomer at pH 7.4, and a total sample concentration of 35 mM in each monomer. See Notes and References<sup>†</sup> for additional model details and ESI for derivation and example calculation. (b) Graphical representation of pH-mediated assembly and disassembly by monomers with  $pK_a$ -matched recognition units. (c) Chemical structures of triaminopyrimidine (**TAP**), cyanuric acid (**CA**, R = H), **CyCo6**, **CyCo4**, and proposed rosette and stacked rosette assemblies.

а

We surmised that reversible supramolecular polymers could be formed from a mixed system of monomeric units having an integrated bidirectional pH-sensitive switch. To test this approach, we prepared water soluble monomers composed of **Cy** substituted with a hexanoic acid (**CyCo6**) or a butyric acid (**CyCo4**) (Fig. 1c).<sup>17</sup> Mimicking nucleic acid base pairing, hydrogen bonding directs the assembly of **TAP** and the **CyCo** monomers into hexameric rosette structures, and hydrophobic interactions cause these rosettes to stack into supramolecular polymers that can then non-covalently crosslink to form a hydrogel matrix. Supramolecular assemblies were found to be most stable when composed of all three monomers and to exhibit the maximal sensitivity to change in pH for a supramolecular polymerization system.

#### **Results and discussion**

A shear-thinning, thermally reversible supramolecular hydrogel forms spontaneously when one equivalent of TAP is added to a 20 mM solution of CyCo6 (0.7% by weight in total monomer), at pH 7 and 20 °C. Gelation is not observed below pH 6 or above pH 8 (Fig. 2a); demonstrating the pH sensitivity of the gel-forming process. At pH 7 and pH 8 TAP and CyCo6 remain in solution for times greater than six months, but precipitation occurs at pH 6 within minutes. Under the same conditions CyCo4 forms a precipitate immediately when mixed with TAP (Fig. S1). Incorporation of a cogelator has been shown to enhance the lifetime of the hydrogel phase.<sup>18</sup> We hypothesized that non-uniform structures formed by the inclusion of closely related CyCo6 as a co-monomer into TAP-CyCo4 assemblies would frustrate regular interactions between linear stacks of monomers. As predicted, mixtures of CyCo4 and CyCo6 form hydrogels with TAP whose gel lifetime at pH 7 increases as the ratio of CyCo6 to CyCo4 increases (Fig. S1). For example, a solution containing TAP, CyCo4 and CyCo6 at a ratio of 1:0.8:0.2, respectively, forms a gel that begins to precipitate within five minutes while a solution at a 1:0.33:0.67 ratio in each molecule, respectively, forms a gel that is stable for greater than 5 hours. At pH 6 the trimolecular assemblies are stable against precipitation for over an hour, while the

bimolecular assemblies precipitate rapidly, demonstrating a synergistic relationship among the monomers.

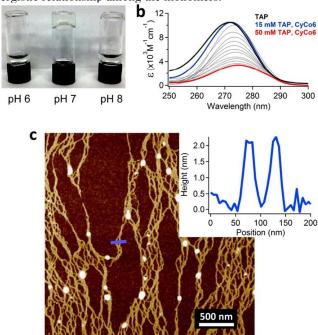


Fig. 2 (a) Inverted bottle test showing gelation of TAP with CyCo6 at pH 7 and no gelation at pH 6 or pH 8. All three solutions are 20 mM in both monomers. (b) UV spectra of solutions of TAP alone and TAP with CyCo6 at equimolar concentrations from 15 to 50 mM. (c) AFM topographic image of TAP-CyCo6 fibers. Insert shows the profile delineated by the blue line in the panel.

The gels' mechanical properties were further studied by oscillatory rheology with solutions that were 1.4% by weight in monomer. Hydrogels consisting of either **TAP-CyCo6** or **TAP-CyCo4-CyCo6** (1:0.33:0.66) exhibited a predominant elastic character over the time scales examined, as evident by a Journal Name

greater storage modulus, G', than loss modulus, G" (Fig. S2). Frequency sweeps were performed at a constant strain value of 1.0%, corresponding to the linear regime as revealed by strain sweeps (Fig. S3). In addition, strain sweeps indicate that G' values are similar at pHs near neutrality (ca. +/- 0.5 pH units) for the biomolecular and trimolecular systems, while G" values are found to vary more markedly with these subtle pH variations and between the systems. More substantial effects were observed when the pH of the solutions was increased to 8.5, as both dynamic moduli decreased by over an order of magnitude. These results indicate that the mechanical properties of the gel phase can be altered with modest pH change.

The nature of **TAP-CyCo** assemblies at the molecular level was investigated using spectroscopic and microscopic techniques. The UV/vis absorption spectrum of TAP in the gel phase is red shifted relative to the spectrum of free TAP, indicative of intermolecular  $\pi$ - $\pi$  electron interactions that are typical of J-type associations by planar molecules (Fig. 2b). The spectrum of samples containing 1:1 TAP-CyCo6 and TAP-CyCo4-CyCo6, at various ratios where the total Cy containing monomer is equal to the concentration of added TAP, revert to that of free TAP upon dilution from 40 mM to 15 mM (in TAP) (Fig. S4), indicating the absence of intermolecular stacking interactions at concentrations below 15 mM. AFM imaging of the TAP-CyCo6 and the trimolecular gel phase shows micron-length linear structures with heights of approximately 2 nm (Fig. 2c and Fig. S5). These structures are consistent with the expected width of the stacked rosette assembly. The observation of extended networks of fibers by AFM is also consistent with the gelation properties of the assemblies.

<sup>1</sup>H NMR spectroscopy indicates the incorporation of these monomers into supramolecular assemblies by exhibiting a concentration-dependent loss of integrated resonance intensity.<sup>19</sup> Specifically, <sup>1</sup>H spectra of **TAP-CyCo6** solutions below 15 mM in each monomer show **CyCo6** resonances with chemical shifts that are identical to **CyCo6** alone in solution and with integrated intensities that are directly proportional to concentration (Fig. 3). In contrast, raising **TAP** and **CyCo6** concentrations above 15 mM results in no further increase of the integrated intensity of **CyCo6** resonances, nor are changes observed in <sup>1</sup>H chemical shifts. This observation indicates that at the higher concentrations, **CyCo6** and **TAP** are slowly exchanging between their free (unassembled) states and incorporation into large assemblies.

NMR and UV/vis experiments both reveal a cooperative assembly process,<sup>20, 21</sup> with a minimal assembly concentration (MAC) for TAP and CyCo6 of 15 mM in each monomer at 20 °C, pH 7. In agreement with the proposed cooperative assembly, NMR and AFM analysis also indicate an absence of intermediate assemblies, i.e., only free monomers and long noncovalent polymers are present. The driving force of this highly cooperative assembly, and thus the formation of long noncovalent polymers, is apparently the hydrophobic effect. The exposure of the two faces of a rosette to water is predicted to have a large positive free energy (ca. 27 kcal/mol).<sup>15, 7</sup> Moreover, the favourable assembly between **TAP** and **Cy** may be enhanced by pK<sub>a</sub> matching of these recognition elements, as both experimental and theoretical studies have indicated that smaller pK<sub>a</sub> differences between H-bond donor and acceptor groups leads to stronger H-bonding.<sup>23-26</sup>

Trimolecular systems were next investigated by <sup>1</sup>H NMR spectroscopy. The analysis shown in Fig. 3 reveals that the MAC for solutions that contain **TAP:CyCo4:CyCo6** in ratios

of 1:0.5:0.5 is identical to the MAC measured for the bimolecular TAP-CyCo6 system when the sum of the integrated intensity of CyCo4 and CyCo6 resonances in the trimolecular system are compared to the integrated intensities of CyCo6 resonances in the bimolecular solutions (Fig. 3b). The same is true for solutions in which the ratio of TAP:CyCo4:CyCo6 is 1:0.33:0.66 (Fig. 3c). The amount of CyCo4 and CyCo6 incorporated into large assemblies with **TAP** is simply proportional to the ratio of these monomers present in solution; equal concentrations of CyCo4 and CyCo6 are incorporated in the 1:0.5:0.5 samples (Fig. 3a and 3b), and a 1:2 ratio of CyCo4 and CyCo6 are incorporated in the 1:0.33:0.66 samples (Fig. 3c). Additionally, the MAC determined for the mixed systems is the same as that measured for the **TAP-CyCo6** system at pH 7 for temperatures ranging from at least 5 to 35 °C (Fig. S6). These observations confirm that the mixed trimolecular system assembles with the same propensity as the TAP-CyCo6 system and demonstrates that formation of soluble co-assemblies with CyCo4 and CyCo6 are insensitive to the differences in the lengths of their carboxylic acid tails.

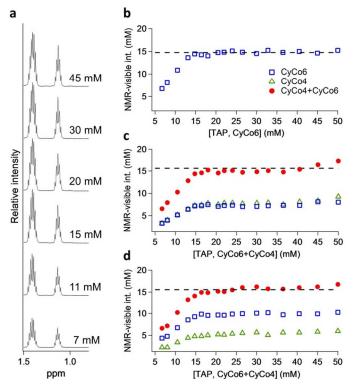
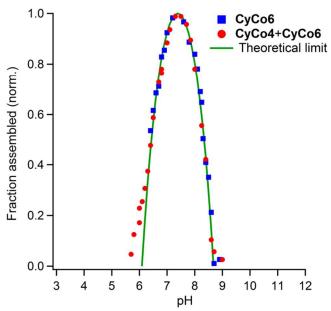


Fig. 3 (a) Representative <sup>1</sup>H NMR spectra of the CyCo6 methylene protons for samples containing various concentrations of TAP and CyCo6 (in equimolar ratios). (b-d) Plots of NMR-visible resonance intensity of unassembled CyCo4 and/or CyCo6 vs actual monomer concentrations in (b) 1:1 TAP and CyCo6 solutions, (c) 1:0.5:0.5 solutions of TAP:CyCo4:CyCo6, and (d) 1:0.33:0.66 solutions of TAP:CyCo4:CyCo6. Dashed lines in plots indicate minimal assembly concentrations of the total CA-containing species.

We note that the precipitate of a 1:0.5:0.5 ratio **TAP:CyCo4:CyCo6** sample is enriched in **CyCo4** (Fig. S7), which is likely the result of monomer rearrangement producing regions enriched in **TAP-CyCo4** assemblies that precipitate due to the greater propensity of this particular assembly to aggregate. This proposed mechanism of precipitation and observed precipitate enrichment in **CyCo4** is consistent with a recent report by Meijer and coworkers, which showed that rearrangement of monomers within and between noncovalent polymers in water can require on the order of hours.<sup>27</sup>

Precipitation of TAP-CyCo4 and TAP-CyCo6 assemblies below pH 7 likely results from intermolecular hydrogen bonding of the neutral carboxylic acid groups that bundle the linear assemblies into large aggregates, as has been seen with other supramolecular polymerization systems that contain monomers with carboxylic acid groups.<sup>9</sup> Cooperative interactions of the mixed monomers are highly synergistic, enabling both the CyCo4 monomer to form long-lived gels and extending the lifetime of CyCo6 assemblies under unfavorable conditions (i.e., low pH). It has been previously shown that substantial modifications in the peripheral groups of recognition elements that also form rosette assemblies can result in the adoption of alternative assembly modes.<sup>28, 29</sup> However, less substantial changes are known to lead to soluble rosette nanotubes or insoluble tightly packed rosette assemblies.<sup>30-32</sup> As demonstrated here, the addition of a structurally similar co-monomer allows the rosette assembly unit to be maintained while simultaneously leading to large differences in assembly solubility due to the frustration of regular packing (and thus crystallization) between the assemblies.

The pH dependent assembly of both bimolecular (**TAP-CyCo6**) and trimolecular (**TAP-CyCo6-CyCo4**) supramolecular polymers were investigated by <sup>1</sup>H NMR spectroscopy. Any deviation in pH away from 7.4 corresponded to disassembly of the supramolecular polymer (Fig. 4 and S8). Complete disassembly occurs when the pH is below 5.7 or above pH 9.0. As anticipated, precipitation was evident when the bimolecular system was evaluated at any pH below 6.5 (Fig. S8). However, for the three-component system, no precipitation was observed over this entire pH range.



**Fig. 4** Plot of the normalized fraction of monomer assembled in solutions that were 35 mM in **TAP** and **CyCo** monomer based on the <sup>1</sup>H NMR visible concentration of unassembled **CyCo6** (in 1:1 **TAP:CyCo6** solutions) (blue) and **CyCo4+CyCo6** (in 1:0.33:0.66 **TAP:CyCo4:CyCo6** solutions) (red) as a function of pH. The theoretical limit governing supramolecular assembly for a  $pK_a$ -matched system as a function of pH is shown as a green line. See Notes and References<sup>†</sup> for additional model details and ESI for derivation. Note that below pH 6.3 the solution

containing only **TAP** and **CyCo6** immediately formed a precipitate, so data points for mixtures below this pH could not be obtained.

The behaviour of the trimolecular system quantitatively illustrates the high level of structural control that can be achieved with pK<sub>a</sub>-matched monomers. Specifically, а comparison of the experimental data to the theoretical limit for a multimolecular assembly as a function of pH demonstrates that the TAP-CyCo system has the theoretically maximum sensitivity to change in pH that can be obtained for a supramolecular polymerization system (Fig. 4). An observation that is consistent with the TAP-CyCo assemblies being the most pH responsive supramolecular polymers ever reported. We note that our observation of the TAP-CyCo system being maximally assembled at pH 7.4 indicates pKa-matched monomers at this pH, which is about 0.5 pH units above the reported pK<sub>a</sub>s of **TAP** and **Cy**. Consistent with this observation, the apparent pKas of TAP, CyCo6 and CyCo4 were individually determined to each be around pH 7.4 in the buffer conditions of our experiments (Fig. S9).

#### Conclusions

We present here a new design strategy for the formation of supramolecular structures that are optimally responsive to changes in pH by utilizing pK<sub>a</sub> matched complementary monomeric recognition elements, acidic and basic monomers. Systems that are pKa matched enable dual, highly non-linear pH control of the assembly, which is most favoured in a solution in which the pH equals the pK<sub>a</sub> of the monomers. The TAP-Cy assembly motif is exceptional because both recognition elements have pKas near neutrality, making systems that employ these complimentary heterocycles attractive for the development of materials useful for biological applications, as small pH changes near physiological pH will elicit an appreciable and predicable response.<sup>11, 33, 34</sup> For example, materials that can undergo a structural transition upon encountering slightly acidic conditions, as found in the extracellular microenvironment of some tumours (pH 6.3-7.2), could find applications in drug delivery and diagnostics.<sup>11</sup>, A significant observation of this study is that heterogeneity within the supramolecular polymers stabilizes the hydrogel phase against precipitation. The ability of mixed assemblies formed by multiple monomers with the same recognition elements but different side chains that suppress the interactions that cause precipitation further supports this approach as a general principle for the design of wellbehaved, self-assembling noncovalent polymer systems. In addition to their importance for materials development, the self-assembly properties of these pKa-matched recognition elements, and their resemblance to RNA nucleobases, have implications regarding the search for the original nucleobases of the pre-RNA world; which we have previously discussed elsewhere.16, 36

#### Acknowledgements

This work was jointly supported by the NSF and the NASA Astrobiology Program, under the NSF Center for Chemical Evolution, CHE-1004570 (NVH, BJC), and the NASA Exobiology Program NNX13AI02G (NVH, GBS, RA). We thank L.A. Bottomley for use of AFM, A. Fernandez-Nieves for use of rheometer, M. Pelaez-Fernandez, I. Gállego, D.M. Fialho, L.A. Lyon and R. Krishnamurthy for discussions.

#### Notes and references

26.

27.

29.

30.

35.

\* Department of Chemistry and Biochemistry, Georgia Institute of Technology, Atlanta, Georgia 30332 (USA). Email: hud@gatech.edu.

† Theoretical limits for the assembly of monomers with various  $\Delta p K_a s$  (0-5) were determined using idealized model systems with 15 mM MACs (at pH 7.4) and solution concentrations (free and assembled) of 35 mM in each monomer. Curves of fraction assembled as a function of pH were generated by taking the product of the pairing monomers at pH 7.4 (the  $\Delta p K_a$  midpoint) as the solubility constant of each system. This constant and the calculated monomer ionization equilibrium were then used to calculate the maximum expected assembly as a function of pH (normalized to fraction assembled at pH 7.4). See ESI for derivation and example.

Electronic Supplementary Information (ESI) available: Materials, methods and supplemental images. See DOI: 10.1039/b000000x/

- R. J. Wojtecki, M. A. Meador and S. J. Rowan, *Nat Mater.*, 2011, 10, 14-27.
- 2. T. Aida, E. W. Meijer and S. I. Stupp, *Science*, 2012, **335**, 813-817.
- J. D. Hartgerink, E. Beniash and S. I. Stupp, Science, 2001, 294, 1684-1688.
- 4. A. R. Hirst, B. Escuder, J. F. Miravet and D. K. Smith, *Angew. Chem. Int. Ed. Engl.*, 2008, **47**, 8002-8018.
- P. Besenius, G. Portale, P. H. H. Bomans, H. M. Janssen, A. R. A. Palmans and E. W. Meijer, *Proc. Natl. Acad. Sci. USA*, 2010, 107, 17888-17893.
- T. Fenske, H. G. Korth, A. Mohr and C. Schmuck, *Chem.-Eur. J.*, 2012, 18, 738-755.
- L. E. Buerkle and S. J. Rowan, *Chem. Soc. Rev.*, 2012, 41, 6089-6102.
- M. D. Segarra-Maset, V. J. Nebot, J. F. Miravet and B. Escuder, *Chem. Soc. Rev.*, 2013, **42**, 7086-7098.
- C. Tang, A. M. Smith, R. F. Collins, R. V. Ulijn and A. Saiani, Langmuir, 2009, 25, 9447-9453.
- D. J. Adams, K. Morris, L. Chen, L. C. Serpell, J. Bacsa and G. M. Day, *Soft Matter*, 2010, 6, 4144.
- A. Ghosh, M. Haverick, K. Stump, X. Yang, M. F. Tweedle and J. E. Goldberger, *J. Am. Chem. Soc.*, 2012, **134**, 3647-3650.
- H. Frisch, J. P. Unsleber, D. Ludeker, M. Peterlechner, G. Brunklaus, M. Waller and P. Besenius, *Angew. Chem. Int. Ed. Engl.*, 2013, **52**, 10097-10101.
- T. Fenske, W. Meyer-Zaika, H. G. Korth, H. Vieker, A. Turchanin and C. Schmuck, J. Am. Chem. Soc., 2013, 8342–8349.
- 14. M. Ma and D. Bong, *Langmuir*, 2011, **27**, 8841-8853.
- B. J. Cafferty, I. Gallego, M. C. Chen, K. I. Farley, R. Eritja and N. V. Hud, J. Am. Chem. Soc., 2013, 135, 2447-2450.
- M. C. Chen, B. J. Cafferty, I. Mamajanov, I. Gállego, J. Khanam, R. Krishnamurthy and N. V. Hud, J. Am. Chem. Soc., 2014, DOI: 10.1021/ja410124v.
- 17. K. Hager, A. Franz and A. Hirsch, *Chem.-Eur. J.*, 2006, **12**, 2663-2679.
- L. E. Buerkle, Z. Li, A. M. Jamieson and S. J. Rowan, *Langmuir*, 2009, 25, 8833-8840.
- 19. D. C. Duncan and D. G. Whitten, *Langmuir*, 2000, 16, 6445-6452.
- A. R. Hirst, I. A. Coates, T. R. Boucheteau, J. F. Miravet, B. Escuder, V. Castelletto, I. W. Hamley and D. K. Smith, *J. Am. Chem. Soc.*, 2008, 130, 9113-9121.
- T. F. De Greef, M. M. Smulders, M. Wolffs, A. P. Schenning, R. P. Sijbesma and E. W. Meijer, *Chem. Rev.*, 2009, **109**, 5687-5754.
- 22. D. Chandler, *Nature*, 2005, **437**, 640-647.
- 23. C. L. Perrin and J. B. Nielson, *Annu. Rev. Phys. Chem.*, 1997, **48**, 511-544.
- J. Chen, M. A. McAllister, J. K. Lee and K. N. Houk, J. Org. Chem., 1998, 63, 4611-4619.
- 25. P. Acharya, P. Cheruku, S. Chatterjee, S. Acharya and J. Chattopadhyaya, J. Am. Chem. Soc., 2004, **126**, 2862-2869.

- A. E. Engelhart, T. H. Morton and N. V. Hud, *Chem. Commun.*, 2009, 647-649.
- L. Albertazzi, F. J. Martinez-Veracoechea, C. M. A. Leenders, I. K. Voets, D. Frenkel and E. W. Meijer, *Proc. Natl. Acad. Sci. USA*, 2013, **110**, 12203-12208.
- G. M. Whitesides, E. E. Simanek, J. P. Mathias, C. T. Seto, D. Chin, M. Mammen and D. M. Gordon, *Acc. Chem. Res.*, 1995, 28, 37-44.
  - A. G. Bielejewska, C. E. Marjo, L. J. Prins, P. Timmerman, F. de Jong and D. N. Reinhoudt, *J. Am. Chem. Soc.*, 2001, **123**, 7518-7533.
  - A. Marsh, M. Silvestri and J.-M. Lehn, Chem. Commun., 1996, 1527.
- M. Mascal, N. M. Hext, R. Warmuth, J. R. Arnall-Culliford, M. H. Moore and J. P. Turkenburg, J. Org. Chem., 1999, 64, 8479-8484.
- A. Durmus, G. Gunbas, S. C. Farmer, M. M. Olmstead, M. Mascal, B. Legese, J. Y. Cho, R. L. Beingessner, T. Yamazaki and H. Fenniri, *J. Org. Chem.*, 2013, **78**, 11421-11426.
- D. Schmaljohann, *Adv. Drug Deliv. Rev.*, 2006, **58**, 1655-1670.
  M. A. Stuart, W. T. Huck, J. Genzer, M. Muller, C. Ober, M.
  - M. A. Stuart, W. T. Huck, J. Genzer, M. Muller, C. Ober, M. Stamm, G. B. Sukhorukov, I. Szleifer, V. V. Tsukruk, M. Urban, F. Winnik, S. Zauscher, I. Luzinov and S. Minko, *Nat Mater.*, 2010, **9**, 101-113.
  - S. Mura, J. Nicolas and P. Couvreur, *Nat Mater.*, 2013, **12**, 991-1003.
- N. V. Hud, B. J. Cafferty, R. Krishnamurthy and L. D. Williams, *Chem. Biol.*, 2013, 20, 466-474.