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## Dynamic acoustic field activated cell separation (DAFACS)

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Advances in diagnostics, cell and stem cell technologies drive the development of application specific tools for cell and particle separation. Acoustic micro-particle separation offers a promising avenue for high-throughput, label-free, high recovery, cell and particle separation and isolation in regenerative medicine. Here, we demonstrate a novel approach utilizing on dynamic acoustic field that is capable of separating an arbitrary size range of cells. We first demonstrate the method with separation of different diameter between 6 and 45 µm and different density of particles in a heterogeneous medium. Dynamic acoustic field are then used to separate dorsal root ganglion cells. The shearless, label free and low damage characteristics make this method of manipulation particularly suited for biological applications. Advantages of using dynamic acoustic field for the separation of cells include its inherent safety and biocompatibility, the possibility to operate over large distances (centimetres), high purity (ratio of particle population, up to 100 %), and high efficiency (ratio of separated particles to separate, up to 100 %).

## Introduction

Separating and sorting cells and micro-organisms from a heterogeneous mixture is a fundamental step in basic biological<sup>1</sup>, chemical and clinical studies<sup>2</sup>, enabling regenerative medicine, stem cell research, clinical sample preparation, and improved food safety<sup>3</sup>. Biological cell separation needs to enrich a target cell population whilst minimising the presence of unwanted cells or contaminants. At present the standard systems for cell separation are fluorescently activated cell sorters (FACS), or sedimentation<sup>4</sup>. FACS requires fluorescent labelling, making it complex and expensive, whereas sedimentation has a low ratio of recovery leading to loss of material. As a consequence there is considerable demand for the discovery of alternative methods. Microfluidic systems<sup>5,6,7</sup> e.g. deterministic sorting<sup>8</sup>, inertial microfluidics<sup>9</sup>, dielectrophoretic<sup>10</sup>, and ultrasonic devices<sup>11,12,13,14,15</sup> present viable routes to superior technologies for cell sorting. Techniques utilising ultrasonics are particularly valuable because of the potential for efficient sorting of relatively large microfluidic volumes in a robust, harmless and scalable technology.

Ultrasonic forces are non-invasive and can effectively manipulate cells for applications such as medium exchange<sup>16</sup>, sample concentration<sup>17,18,19</sup>, sorting<sup>20,15</sup>, enhanced bio-detection and immuno-assays. Achieving cell separation<sup>21</sup> utilizing ultrasonic manipulation by frequency sweeping<sup>11,22</sup> has been previously demonstrated however, this method has several critical disadvantages. Firstly unstable forces are generated which leads to differences in the movement of individual trapped particles of the same size or property<sup>23,24</sup>. Secondly, this method exclusively allows small particle displacement, which in turn limits sorting efficiency. Thirdly, frequency sweep sorting inherently lacks flexibility because the frequencies that can be used are limited by the types and dimensions of the transducers. Finally, the dimensional properties of the whole resonator may pose some constraints on the frequency regime.

An alternative to manipulating acoustic frequency is to control the signal phase; it has been demonstrated that shifting the phase of acoustic travelling waves can be used to control the position of micro-particles suspended in an aqueous media<sup>25</sup>. When two opposing transducers are excited, a linear interference pattern of nodes and antinodes is formed in the interstitial media. The micro-particles are trapped at the minima of the potential acoustic energy density<sup>26</sup>. Electronically shifting the excitation phase of one of the transducers, with respect to the other, proportionally translates the linear interference pattern in the direction of the added phase delay. The shift of the node position translates to a change of the position of trapped micro-particles<sup>27,28</sup>.

The new technique devised utilizes a dynamic acoustic field (DAF) with a time-varying phase delay between two opposing travelling waves that results in the separation of particles or cells over large distances (of the order of centimetres). The method shows a high degree of separation selectivity and throughput, making it suitable for applications such as cell sorting. The forces generated by this method are very stable<sup>23</sup> allowing better spatial separation

(improved control and manipulation) of particles and cells to be achieved and maintained compared to results realized using frequency sweeping. This ultimately results in separation of sub-populations in the sample volume with much higher purity.

In this paper, we demonstrate that the flow-less DAF method can be used to separate particles within a sample volume depending on their size or density. We also study the discriminative ability of the method for particles of different size or densities. The application of DAF to primary pig dorsal root ganglion neuron as a contact-less means of separating these neurons from debris and smaller cells, which results from tissue digestion, is demonstrated.

## **Experimental results and discussion**

Initial experiments were conducted with synthetic particles of various sizes and density in order to demonstrate the feasibility of the approach. Previous studies have found that these particles are a reasonable surrogate for biological cells<sup>29</sup>. Once DAF had been successfully used to discriminate within heterogeneous mixtures particles by their sizes and density in multiple cycles, we progressed on to show the capacity of DAF to isolate dorsal root ganglion cells from a mixture of cells and debris as develops during tissue digestion.

**Principle of operation**. The time variant primary acoustic force combined with the viscous force enable discriminating particles according to their physico-mechanical properties (size and density). The acoustic force  $F_a$ , equation (1), is dependent on the particle radius and particle density. Smaller or less dense particles and cells experience a lower acoustic force  $F_a$ , with respect to larger or denser particles and cells, therefore particle and cells of different sizes and different densities will separate. This outlines the fundamental concept utilising the DAF method.

The technique relies on a repeated cycling pattern of the phase difference between two excited transducers from 0° to 360°. Within each cycle the phase is swept completely through 360° over a time  $t_{ramp}$  and then allowed to rest for a period  $t_{rest}$  before commencing the next cycle. The interplay between the rate at which the phase is swept, and the length of the rest time, is at the core of the separation technique. During  $t_{ramp}$ , under the correct conditions, the particles of interest experience a strong acoustic force compared to the viscous force.

For a system with particles trapped in one of many nodes, N, the particles of interest closely follow the node that traps them and travel from the initial position of the node  $N_i$  to the initial position of the next node  $N_{i+1}$  (Fig. 2 and 3). This controlled manipulation occurs since a phase shift of 360° moves each node exactly one integer node position, a distance  $\Lambda$  (Fig. 1). If at the end of the  $t_{ramp}$  period, the smaller particles have not travelled more than half way (Fig. 2 and 3) from their initial position of  $N_i$  to the next node position  $N_{i+1}$ , they will relax to their starting position during  $t_{rest}$ , whereas the particles of interest, which have travelled past the midpoint  $N_i$  and  $N_{i+1}$ , will relax towards the next node position  $N_{i+1}$ .

The interplay between acoustic force and viscous drag force is at the heart of the discriminating ability of the DAF method with  $t_{rest}$  serving as an equilibrating interval, during which particles settle at their nearest nodes before the next cycle begins. Assuming the length of  $t_{ramp}$  has been selected correctly, the best choice of  $t_{rest}$  to achieve cell or particle discrimination depends on the balance between the acoustic force,  $F_a$ , equation (1) and the viscous force,  $F_v$ , equation (3).  $t_{rest}$  and  $t_{ramp}$  was measured for different sizes of particles, by applying a step change of 180° in phase to one of the transducers<sup>25</sup>. In water, polystyrene particles of 6, 10 and 45 µm diameter respectively require a time delay of 5, 2 and 0.5 s respectively to reach their equilibrium positions. This serves as the basis to establish the timescale for  $t_{rest}$  and  $t_{ramp}$  required to discriminate between particles during the experiments. The DAF method can then be optimized to achieve the optimum separation performance in

specific applications. These optimum values of  $t_{ramp}$  and  $t_{rest}$  are dependent on the viscosity of the medium, as well as the density and size of the particles.



Figure 1. The effect of the applied phase shift on large and small particles. The large particles move with greater velocity than small particles, thus when the phase shift reaches the rest period at 360° the large particles continue forward and reach the next acoustic node and are trapped. However, the small, light particles relax back to the initial node.

Acoustic separation simulation. The mechanism of particle separation was studied by using a numerical model to predict the particle behaviour under dynamic acoustic field. The code was developed in Visual Basic from scratch. We modelled particle behaviour under different dynamic acoustic field, particle properties (size and density), liquid viscosity and flow. The program sums the forces acting on the particles taking the primary acoustic force  $F_a$ , equation (1), and viscous force  $F_{\nu}$ , equation (3) as a function of time into account.

The equation of motion of a particle labelled by its position r(t) is given by:

$$\frac{\partial^2 r}{\partial t^2} = \frac{\partial v}{\partial t} = \frac{F_{tot}(r, v, t)}{m} = a(r, v, t)$$
(4)

 $F_{tot}$  being the sum of  $F_a$  and  $F_v$ , *m* the mass of the particle, *a* the acceleration as a function of particle position *r* and the relative particle velocity *v* at a time *t*.

These equations of motion are integrated step by step using the Verlet  $algorithm^{30,31}$ , equations (5) and (6) to produce the movement of the particles.

$$r(t + \Delta t) = 2r(t) - r(t - \Delta t) + a(t)\Delta t^{2}$$
(5)

$$v(t) = \frac{r(t + \Delta t) - r(t - \Delta t)}{2\Delta t}$$
(6)

where r is the particle position, v its velocity and  $\Delta t$  the integration time step.

The computer program can simulate the movement of *P* particles among several types of particles differing by their radius and density. Figure 2 denotes the predicted particle separation of two classes of particles differing by size. The ratio between  $t_{ramp}$  and  $t_{rest}$  is 1, with a  $t_{ramp} = t_{rest} = 5$  s. As predicted, large and small particles are separated.



Figure 2. Particle behaviour under the dynamic acoustic field predicted by the simulation. Under the influence of the phase shift during  $t_{ramp}$ , the larger (10 µm particle size; in green) and smaller particles (6 µm particle size; in red) separate over time (5 s). The smaller particles are submitted to a smaller acoustic force and thus cannot follow the phase shift, whereas the large particles are moved and slip to the position of the next node ( $N_{i+1}$ ) during the  $t_{ramp}$ . During  $t_{rest}$ , the larger particles stabilize on the next nodes ( $N_{i+1}$ ) whilst the smaller particles move back to their initial position ( $N_i$ ) and the process can be repeated indefinitely, shifting the larger particles continuously further to the right. The fine vertical lines represent the nodes at *t* indicated with the particle position at  $d\Phi$  indicated.

Acoustic separation by particle size. To investigate the potential of this method to discriminate particles by size, sets of polystyrene particles (Polysciences Europe, Germany) with varying diameter, (a) 10 and 45  $\mu$ m and (b) 6 and 10  $\mu$ m were subjected to dynamic acoustic field.

The transducers were excited at a frequency of 4.00 MHz with an amplitude of 8 V<sub>pp</sub>. At this frequency, the wavelength of the sound waves in water is  $\lambda = 370 \ \mu\text{m}$  ( the velocity of sound velocity in water is 1480 m/s)<sup>32</sup>. A schematic of the experimental setup is shown in Fig. 3. The particles agglomerate at the nodes with a separation of  $\lambda/2 = \Lambda = 185 \ \mu\text{m}^{33,34}$ . To estimate the order of magnitude of the acoustic pressure experienced by the particles of different diameters (6, 10, and 45  $\mu$ m), we recorded the time taken for the particles to agglomerate at the nodes starting from a randomly distributed motion state. These experiments were repeated

5 times at 5 different locations within the device using time-lapse microscopy. The viscous force,  $F_{\nu}$ , derived from the dimensions and density of the particles as well as the viscosity of the medium (3), allows the acoustic pressure amplitude acting on the particles to be calculated. The acoustic pressure was found to be 91 ± 7 kPa, 62 ± 4 kPa and 48 ± 2 kPa for 6, 10 and 45 µm diameter particles, respectively.



**Figure 3. Schematic of the experimental setup,** with a signal generator (left) driving two opposing ultrasonic transducers (TDX), with a programmable phase shift. The signal was amplified by 10x by two purpose built amplifiers. The octagon well with the transducers was placed on a light microscope for observation.

The separation experiments were conducted using two mixtures of particles, each at a particle density of 4.99 x 10<sup>5</sup> particles/mL. Mixture A contained 10 and 45  $\mu$ m diameter particles at a ratio of 1:100. Mixture B contained 6 and 10  $\mu$ m diameter particles at a ratio of 1:100. At these concentrations, no aggregation of particles was observed. As predicted, the time variant acoustic field was able to manipulate, and at the same time, separate particles according to their size (Fig. 4). Fig. 4a shows the expected behaviour of the large and small particles over time (black =  $t_0$ , purple  $t_1$ , red  $t_2$ , yellow  $t_3$ ) in relation to the initial position of the nodes (indicated by  $N_i$  and  $N_{i+1}$ ) and antinodes (indicated by  $N_{i+1/2}$ ) of the acoustic landscape. During  $t_{ramp}$ , the large particles, experiencing larger acoustic force, follow the moving node and thus move over a smaller distance. During  $t_{rest}$ , the smaller particles return to their initial position  $N_i$ , while the larger particles settle at the next node  $N_{i+1}$ . Fig. 4b shows particle traces as function of time (71 frames

covering 36 s). In this case, the 45  $\mu$ m diameter particles follow the shifted acoustic field (moving towards the right hand side) while the 10  $\mu$ m diameter particles stay close to the position of the original node. The time between frames was 0.5 s. The average movement of small and large particles was 3 ± 1  $\mu$ m/s and 30 ± 1  $\mu$ m/s respectively during *t<sub>ramp</sub>*.



Figure 4. Position of the particles as a function of time (represented by colour) (a) A schematic illustration that shows 4 positions of large and small particles. Larger particles, experiencing larger acoustic force, are transported to the next node  $N_{i+1}$  at  $x_2$  whereas smaller particles, experiencing smaller acoustic force, return to  $N_i$  at  $x_0$ . (b) Experimental data of the 45-10 µm particles mixture under the dynamic acoustic field (71 frames covering 36 s). The longer trails show the distance traversed by 45 µm diameter particles, whereas the short trails illustrate how 10 µm diameter particles do not traverse the space. (See supplementary video).

A series of time-lapse images were recorded at regular intervals during separation to produce particle traces. The image stack was then analysed using  $FIJI^{35}$  and the particle positions were extracted. The resulting data (x, y, particle area) was used to calculate efficiency (7) and purity (8) of separation for each experiment by computing the particle

density projected along the node lines and analysing the time variation of this density. Fig. 5a shows the initial position for 10 and 45  $\mu$ m diameter particles for seven consecutive nodes. Fig. 5b shows the average displacement of the particles for five cycles of continuous phase shift including the rest period. It can be seen that the large particles successively travel during  $t_{ramp}$  and stabilise during  $t_{rest}$  at the next node resulting in an overall displacement over time. Simultaneously the small particles move only slightly therefore not crossing the midpoint of the nodes, then returning to their initial node position during  $t_{rest}$ .



Figure 5. Graphs showing (a) the particle density aligned at the nodes (b) the average particle displacement with time. The large particles, experiencing larger acoustic force, move during  $t_{ramp}$  (8 s) and stabilise during  $t_{rest}$  (4 s) at the next node resulting in an overall displacement over time. Simultaneously the small particles, experiencing lower acoustic force, move only slightly, returning to their initial node position during  $t_{rest}$ . The arrows indicate the relaxing movement during  $t_{rest}$ . Experiments were repeated 10 times.

Separation efficiency was simulated using the DAF model outline above with the same parameters as the experiments. Table. I (a) shows the simulation results. The regions coloured red represent the parameters that did not show particle separation, whereas the particles were successfully separated in the blue region. Table. I (b) shows the experimental results conducted replicating the simulation parameters. The blue regions represent those parameters where > 91 % separation was achieved and the red regions represent the parameters where < 80 % separation was achieved. The yellow regions show those parameters where the separation was 80 - 90 % successful. Comparing computational and experimental results, it can be seen that there is a good match between the simulation and the experimental data.

It can be observed that separation performance improves with  $t_{ramp}$  and  $t_{rest}$  until it reaches a maximum. This indicates that the acoustic forces that the shifting nodes exert on the particles of interest need a minimum time to overcome inertia and viscous force in order to move the particles from the node  $N_i$  to the next node  $N_{i+1}$ .  $t_{ramp}$  is the critical parameter, since it has the greatest effect on the separation result. The value of  $t_{rest}$  is less influential on the separation of the entities.



TABLE I. (a) Simulation and (b) experimental results for the separation performance, depending the ramp and rest time. Parameters suitable for separation (> 0.91 purity) are indicated by a blue shade, and those that are unsuitable are shown in red (purity < 80 %). Intermediate values are in yellow (purity between 80 - 90 %).

When suitable parameters are selected the separation process reaches its best experimental performance: with the separation ratio achieving ~100 % purity and efficiency when  $t_{rest} = 4$  s and  $t_{ramp} = 8$  s.

The simulation and experiment was replicated using 6 and 10  $\mu$ m diameter particles. For these particles, it was found that ~97 % of particles separate, with an efficiency and a purity of ~97 %. These results were achieved with a value of  $t_{ramp} = 15$  s and  $t_{rest} = 15$  s. Fig. 6 shows the particle traces as function of time of 6 and 10  $\mu$ m diameter particles mixture (71 frames covering 142 s) under these conditions. The 10  $\mu$ m diameter particles move from node to node (moving towards the right hand side) while the smaller particles (6  $\mu$ m in diameter) remain close to their initial position of the original node. The average movement of the 6 and 10  $\mu$ m diameter particles was calculated as 2 ± 1  $\mu$ m/s and 10 ± 1  $\mu$ m/s respectively during  $t_{ramp}$ .



Figure 6. (a) Particle traces as function of time, (b) Graph showing the average particle displacement with time. The arrow shows the rest period. The 10  $\mu$ m particles, experiencing larger acoustic force, move during  $t_{ramp}$  (15 s) and stabilise during  $t_{rest}$  (15 s) at the next node resulting in an overall displacement over time. Simultaneously the small particles, experiencing lower acoustic force, move only slightly, returning to their initial node position during  $t_{rest}$ . The arrows indicate the relaxing movement during  $t_{rest}$ . Experiments were repeated 5 times.

These above experiments were performed using the smallest gap in size of commercially available particles ( $10 \pm 1 \mu m$  and  $6 \pm 0.6 \mu m$ ; Polysciences Europe, Germany). Therefore an experimentally accessible discrimination capability of the acoustic separation device is  $\pm 2 \mu m$  difference of particle diameter.

Acoustic separation by particle density. In this section, particles of same size in diameter but of different density are separated. The acoustic forces are not solely based on particle size

but also depend on density (or particle mass), equation 2. We tested the separation performance against particle density by using 10  $\mu$ m diameter polystyrene particles and iron-oxide filled particles with a density of  $\rho = 1.05$  g/cm<sup>3</sup> and  $\rho = 1.41$  g/cm<sup>3</sup>, respectively. The ratio of the concentration of iron-oxide filled and polystyrene particles was 1:100.

As for previous experiments, Fig. 7 shows particle traces as function of time for 10  $\mu$ m diameter iron-oxide filled and polystyrene particles mixture using DAF (140 frames covering 140 s). It can be seen that the denser particles (iron-oxide filled) are significantly affected by the DAF and continuously travel from left to right, whilst the less dense particles (polystyrene) have a more restricted range of movement close to their starting positions. With these particles a separation performance of ~99 % has been recorded, with an efficiency of ~100 % and a purity of ~98 % for  $t_{ramp} = 14$  s and  $t_{rest} = 14$  s. The average movement of the polystyrene and iron-oxide filled particles was calculated as  $3 \pm 1 \mu$ m/s and  $15 \pm 1 \mu$ m/s during  $t_{ramp}$ .



Figure. 7 (a) Particle traces as function of time, (b) Graph showing the average particle displacement with time. The iron-oxide filled particles, denser, experiencing larger acoustic force, move during  $t_{ramp}$  (14 s) and stabilise during  $t_{rest}$  (14 s) at the next node resulting in an overall displacement over time. Simultaneously the polystyrene particles, less dense, experiencing lower acoustic force, move only slightly, returning to their initial node position during  $t_{rest}$ . The arrows indicate the relaxing movement during  $t_{rest}$ . Experiments were repeated 5 times.

The less dense particles (polystyrene) experience smaller acoustic force (equation (1) and (2)), and thus cannot follow the DAF while the denser particles (iron-oxide filled) track the movement of the dynamic acoustic field. Thus, particles of same diameter but different density can be sorted with the DAF method.

#### Acoustic separation of dorsal root ganglion (DRG) cells from a heterogeneous medium.

As mentioned, particles used in our previous experiments are a reasonable surrogate for biological cells<sup>29</sup>. Since the acoustic force is proportional to the particle volume the

polystyrene particles are thus likely to experience forces of the same order of magnitude as the cells. At higher intensities ultrasound can create physiological effects, however several studies have investigated viability and gene expression at intensities sufficient for manipulation in the MHz range and found no observable detrimental effect on cell viability over extend periods of exposure<sup>36,37,38,39</sup>.

To assess a potential practical application, we applied the dynamic acoustic field to separate porcine dorsal root ganglion (DRG) neurons from a freshly isolated mixture containing myelin debris and other non-neuronal cells. The neurons would normally be separated based on their hydrodynamic state using centrifugation across a Ficoll gradient<sup>40</sup>. The DRG neurons have an average size from 17 to 145  $\mu$ m while the myelin debris, has a size of approximately from 10 to 15  $\mu$ m.

Fig. 8 shows debris (~ 26  $\mu$ m) and a single DRG neuron (~ 85  $\mu$ m) in the presence of an acoustic standing wave. These entities aligned themselves in vertical lines, agglomerating at the nodes of the acoustic field<sup>41</sup>. Dynamic acoustic field were then applied (using a  $t_{ramp}$ and  $t_{rest}$  of 5 s), and the resulting time-lapse overlay is represented in Fig. 9a. Static material does not produce a trace whereas material that has been displaced, shows a trace that moves from left to right. The DRG cell follows the shifted acoustic field (moving towards the right hand side) while the debris exhibits minimal displacement of the original node. The time between frames was 0.5 s. The average movement of the DRG cell was calculated to be 18.5 ± 1  $\mu$ m/s during  $t_{ramp}$ . Fig. 9b shows the displacement of the DRG over time.



Figure 8 Debris (arrows) and a single DRG neuron (arrowhead) in the presence of acoustic standing waves. The cells and debris agglomerate in the nodal line of the acoustic landscape. Scale bar =  $185 \mu m$ .



Figure 9(a) Cell traces as function of time, (b) Graph showing the average cells displacement with time. The DRG cell experiencing larger acoustic force, moves during  $t_{ramp}$  (5 s) and stabilise during  $t_{rest}$  (5 s) at the next node resulting in an overall displacement over time. Simultaneously the debris exhibit minimal displacement of the original node. The arrows indicate the relaxing movement during  $t_{rest}$ . The distance between nodes was 185 µm. Experiments were repeated 4 times.

In these experiments, the DRG cells exhibit similar behaviour to the particles in the previous experiments. Only the large DRG neurons are differentially shifted to the right, while smaller cells and debris remain in their original node.

## Conclusion

We have demonstrated a new technique using dynamic acoustic field to separate particulate materials on the basis of size and density. Separation occurs as long as the balance of the acoustic forces and the viscous force is differentiated between particles types. Since the acoustic forces are a function of size and density of the particles, many types of particles or cell type can be separated. A detailed evaluation has been carried out using polystyrene and iron-oxide filled particles, prior to applying the technique to the separation and purification of recovered dorsal root ganglion cells from the myelin cells. The advantages of DAF for the separation of particles include its inherent safety and biocompatibility, the possibility of operating over large distances (centimetres), the high purity (ratio of particle populations) that can be achieved that is up to 100 % and high efficiency (ratio of separated particles over total number of particles of same type in sample) that can be up to 100 %.

## Methods

Acoustic radiation force. The primary acoustic force  $F_a$ , equation (1), and viscous force  $F_v$ , equation (3), to which the particles are submitted at time *t* are given by equation (1) to equation (3):

$$F_{a} = -\left(\frac{\pi p_{0}^{2} V_{c} \beta_{w}}{2\lambda}\right) \times \phi(\beta, \rho) \times \sin(2kx)$$
(1)

$$\phi(\beta,\rho) = \frac{5\rho_c - 2\rho_w}{2\rho_c + \rho_w} - \frac{\beta_c}{\beta_w}$$
(2)

$$F_{\nu} = -6\pi\eta R\nu \tag{3}$$

where  $p_0$  is the acoustic pressure amplitude,  $V_c$  the volume of the particle,  $\lambda$  the wavelength, k the wave number equals to  $2\pi/\lambda$ , x the distance from a pressure node,  $\rho_c$  and  $\rho_w$  the density of the particle and fluid,  $\beta_c$  and  $\beta_w$  the compressibility of the particle and fluid,  $\eta$  the medium viscosity, R the particle radius, v the relative velocity.

The acoustic contrast factor, equation (2), represented by ( $\phi$ ) in equation (1), depends on both the particle density ( $\rho_c$ ) and its compressibility ( $\beta_c$ ) in relation to the corresponding properties of the surrounding medium ( $\rho_w$ ,  $\beta_w$ ). The equation of the primary radiation force,  $F_a$ , equation (1) states that the acoustic force applied on the particles is proportional to the acoustic pressure amplitude ( $\rho_0$ ) squared and to the volume of the particles ( $V_c$ ). The acoustic dynamic field takes advantages of the size dependency of the mechano-physical properties of the micro-entities being sorting, that scales with particle volume, inducing a primary force which is strongly dependant on particle size ( $r^3$ ) and the medium viscosity.

Acoustic device. The acoustic device, described elsewhere<sup>25</sup>, has been used to demonstrate how dynamic acoustic field technology can be applied to particle sorting. In this paper, only two opposite transducers were used. The pair of transducer was synchronised using an arbitrary waveform generator (TGA12104, Thurlby Thandar Instruments, UK) allowing independent control of the amplitude, phase and frequency of each channel. The waveform generator was controlled by a general purpose interface bus (GPIB) utilizing a script written in Labview (National Instruments, USA). The signals, created by the waveform generator, were amplified and matched by high-speed buffers (AD811 Analog Devices USA; BUF634T, Texas Instruments, USA), before being fed to the transducers via length matched coax cables. Furthermore, an agar layer was introduced into the device to minimize the streaming<sup>42</sup> and maximize the precision in control of the particle movement.

**Separation performance.** We studied the separation performance in purity and efficiency depending on the ramping time and the resting time of the phase. Separation purity and separation efficiency are two figures of merit that can used to assess separation performance. The separation purity and efficiency can be expressed as:

$$Efficiency = \frac{Target \ cells \ in \ desired \ area}{Total \ target \ cells \ in \ sample} \cdot 100\%$$
(7)

$$Purity = \frac{Target \ cells \ in \ desired \ area}{Total \ cells \ in \ desired \ area} \cdot 100\%$$
(8)

The demonstrated results in table I were calculated using the following formula which is the average of purity and efficiency as described above.

$$Final \ grade = \frac{Efficiency + Purity}{2} \tag{9}$$

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## References

- <sup>1</sup> Diethard Mattanovich and Nicole Borth, Microbial Cell Factories **5** (1), 12 (2006).
- <sup>2</sup> Dimitri Pappas and Kelong Wang, Analytica Chimica Acta **601** (1), 26 (2007).
- <sup>3</sup> S. Neethirajan, I. Kobayashi, M. Nakajima, D. Wu, S. Nandagopal, and F. Lin, Lab Chip **11** (9), 1574 (2011).
- <sup>4</sup> Richard G. Miller and R. A. Phillips, Journal of Cellular Physiology **73** (3), 191 (1969).
- <sup>5</sup> Yan Gao, Wenjie Li, and Dimitri Pappas, Analyst **138** (17), 4714 (2013).
- <sup>6</sup> Andreas Lenshof and Thomas Laurell, Chemical Society Reviews **39** (3), 1203 (2010).
- <sup>7</sup> Yuchao Chen, Peng Li, Po-Hsun Huang, Yuliang Xie, John D. Mai, Lin Wang, Nam-Trung Nguyen, and Tony Jun Huang, Lab on a Chip **14** (4), 626 (2014).
- <sup>8</sup> John A. Davis, David W. Inglis, Keith J. Morton, David A. Lawrence, Lotien R. Huang, Stephen Y. Chou, James C. Sturm, and Robert H. Austin, Proceedings of the National Academy of Sciences **103** (40), 14779 (2006).
- <sup>9</sup> AliAsgarS Bhagat, SathyakumarS Kuntaegowdanahalli, and Ian Papautsky, Microfluid Nanofluid 7 (2), 217 (2009).
- <sup>10</sup> Ana Valero, Thomas Braschler, Nicolas Demierre, and Philippe Renaud, Biomicrofluidics **4** (2) (2010).
- <sup>11</sup> Nick Harris, Rosemary Boltryk, Peter Glynne-Jones, and Martyn Hill, Physics Procedia **3** (1), 277 (2010).
- <sup>12</sup> Lothar Schmid, David A. Weitz, and Thomas Franke, Lab on a Chip **14** (19), 3710 (2014).
- <sup>13</sup> Ola Jakobsson, Carl Grenvall, Maria Nordin, Mikael Evander, and Thomas Laurell, Lab on a Chip **14** (11), 1943 (2014).
- <sup>14</sup> Xiaoyun Ding, Sz-Chin Steven Lin, Michael Ian Lapsley, Sixing Li, Xiang Guo, Chung Yu Chan, I. Kao Chiang, Lin Wang, J. Philip McCoy, and Tony Jun Huang, Lab on a Chip **12** (21), 4228 (2012).
- <sup>15</sup> X. Ding, Z. Peng, S. C. Lin, M. Geri, S. Li, P. Li, Y. Chen, M. Dao, S. Suresh, and T. J. Huang, Proceedings of the National Academy of Sciences of the United States of America **111** (36), 12992 (2014).
- <sup>16</sup> Jeremy J. Hawkes, Robert W. Barber, David R. Emerson, and W. Terence Coakley, Lab on a Chip 4 (5), 446 (2004).
- <sup>17</sup> Martyn Hill, Yijun Shen, and Jeremy J. Hawkes, Ultrasonics **40** (1-8), 385 (2002).
- <sup>18</sup> D. Carugo, T. Octon, W. Messaoudi, A. L. Fisher, M. Carboni, N. R. Harris, M. Hill, and P. Glynne-Jones, Lab Chip 14 (19), 3830 (2014).
- <sup>19</sup> M. Nordin and T. Laurell, Lab Chip **12** (22), 4610 (2012).
- <sup>20</sup> F. Petersson, A. Nilsson, C. Holm, H. Jonsson, and T. Laurell, Analyst **129** (10), 938 (2004).
- <sup>21</sup> Allen H. J. Yang and H. Tom Soh, Analytical Chemistry **84** (24), 10756 (2012).
- <sup>22</sup> Yang Liu and Kian-Meng Lim, Lab on a Chip **11** (18), 3167 (2011).
- <sup>23</sup> T. Kozuka, T. Tuziuti, H. Mitome, F. Arai, and T. Fukuda, in *Micromechatronics and Human Science, 2000. MHS 2000. Proceedings of 2000 International Symposium on* (IEEE, 2000), pp. 201.
- <sup>24</sup> T. Kozuka, T. Tuziuti, H. Mitome, and T. Fukuda, in *Micro Machine and Human Science, 1995. MHS '95., Proceedings of the Sixth International Symposium on* (IEEE, 1995), pp. 179.
- <sup>25</sup> A. L. Bernassau, C. R. P. Courtney, J. Beeley, B. W. Drinkwater, and D. R. S. Cumming, Applied Physics Letters **102** (16), 164101 (2013).
- <sup>26</sup> A. Bernassau, P. MacPherson, J. Beeley, B. W. Drinkwater, and D. Cumming, Biomedical Microdevices 15 (2), 289 (2013).
- A. L. Bernassau, Ong Chun-Kiat, Ma Yong, P. G. A. Macpherson, C. R. P. Courtney, M. Riehle, B. W. Drinkwater, and D. R. S. Cumming, Ultrasonics, Ferroelectrics and Frequency Control, IEEE Transactions on 58 (10), 2132 (2011).
- <sup>28</sup> C. R. P. Courtney, C. K. Ong, B. W. Drinkwater, A. L. Bernassau, P. D. Wilcox, and D. R. S. Cumming, Proceedings of the Royal Society A: Mathematical, Physical and Engineering Science 468, 337 (2012).
- <sup>29</sup> Jinjie Shi, Daniel Ahmed, Xiaole Mao, Sz-Chin Steven Lin, Aitan Lawit, and Tony Jun Huang, Lab on a Chip 9 (20), 2890 (2009).
- <sup>30</sup> Loup Verlet, Physical Review **165** (1), 201 (1968).
- <sup>31</sup> Loup Verlet, Physical Review **159** (1), 98 (1967).
- <sup>32</sup> Michael Baumann, Vincent Daanen, Antoine Leroy, and Jocelyne Troccaz, in *Computer Vision Approaches to Medical Image Analysis*, edited by ReinhardR Beichel and Milan Sonka (Springer Berlin Heidelberg, 2006), Vol. 4241, pp. 248.

- <sup>33</sup> Thomas Laurell, Filip Petersson, and Andreas Nilsson, Chemical Society Reviews **36** (3), 492 (2007).
- <sup>34</sup> T. Kozuka, T. Tuziuti, H. Mitome, and T. Fukuda, in *Micro Machine and Human Science, 1994. Proceedings., 1994 5th International Symposium on* (IEEE, 1994), p. 83.
- <sup>35</sup> J. Schindelin, I. Arganda-Carreras, E. Frise, V. Kaynig, M. Longair, T. Pietzsch, S. Preibisch, C. Rueden, S. Saalfeld, B. Schmid, J. Y. Tinevez, D. J. White, V. Hartenstein, K. Eliceiri, P. Tomancak, and A. Cardona, Nature methods **9** (7), 676 (2012).
- <sup>36</sup> H. Bohm, P. Anthony, M. R. Davey, L. G. Briarty, J. B. Power, K. C. Lowe, E. Benes, and M. Groschl, Ultrasonics **38** (1), 629 (2000).
- <sup>37</sup> Despina Bazou, Roisin Kearney, Fiona Mansergh, Celine Bourdon, Jane Farrar, and Michael Wride, Ultrasound in Medicine & Biology **37** (2), 321 (2011).
- <sup>38</sup> Bruno Vanherberghen, Otto Manneberg, Athanasia Christakou, Thomas Frisk, Mathias Ohlin, Hans M. Hertz, Bjorn Onfelt, and Martin Wiklund, Lab on a Chip **10** (20), 2727 (2010).
- <sup>39</sup> J. Hultstrom, O. Manneberg, K. Dopf, H. M. Hertz, H. Brismar, and M. Wiklund, Ultrasound Med Biol. 2007 Jan;33(1):145-51. **33**, 175 (2006).
- <sup>40</sup> R. Gilabert and P. McNaughton, Journal of neuroscience methods **71** (2), 191 (1997).
- <sup>41</sup> F. Gesellchen, A. L. Bernassau, T. Dejardin, D. R. S. Cumming, and M. O. Riehle, Lab on a Chip **14** (13), 2266 (2014).
- <sup>42</sup> A. L. Bernassau, P. Glynne-Jones, F. Gesellchen, M. Riehle, M. Hill, and D. R. S. Cumming, Ultrasonics 54 (1), 268 (2014).

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#### Contributions

G.S. did the experiments and analysis of the data. Acoustic technologies were devised by A.L.B. with input from D.R.S.C. M.O.R and J.N.R. conceived the idea of applying the dynamic acoustic field to dorsal root ganglion cells and extracted the cells from pig tissue. D.R.S.C, M.O.R and A.L.B drafted the paper. All authors commented on the final manuscript.

#### **Competing financial interests**

The authors declare no competing financial interests.

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