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Biosynthesis and characterization of *Ocimum sanctum* green silver nanoparticles and unravelling their enhanced anti-filarial activity through a HRAMS proteomics approach

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The available anti-filarial medications are largely ineffective against adult filarial worms. Also, these drugs have several drawbacks such as toxicity and development of resistance owing to long-term usage. Green nanomedicine may offer better solutions for Lymphatic Filariasis treatment due to its tiny size, biocompatibility, and better penetration at considerably lower costs with higher therapeutic efficacy. In the present study, *Ocimum sanctum* silver nanoparticles (OSAgNPs) were bio-synthesized and their anti-filarial efficacy was evaluated against adult filarial parasites. The green nanoparticles were characterized by UV-VIS spectroscopy, XRD, FTIR, SEM, and TEM analysis. The OSAgNPs significantly affected the motility and viability of adult *Setaria cervi* parasites after 4 h of incubation at concentrations higher than 0.5 $\mu\text{g ml}^{-1}$. Proteomics analysis by high resolution accurate mass spectrometry revealed that 213 proteins were differentially expressed following OSAgNP treatment. Mostly these DEPs belonged to the many biochemical and molecular pathways of parasites such as muscle proteins, antioxidant proteins, heat shock proteins, signal recognition proteins, and energy metabolism-related proteins. Undoubtedly, this study will open new avenues for the development of novel anti-filarial drugs based on green nanoparticles.

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1. Introduction

Lymphatic filariasis (LF) is a mosquito transmitted disease common in tropical and subtropical regions of the world. Around 882 million people in 44 countries are at risk of LF infection. People acquire LF infections usually in childhood and the symptoms like lymphodema, elephantiasis, and hydrocele start developing in later years.¹ The World Health Organization launched a Global Program to Eliminate Lymphatic Filariasis (GPELF) in the year 2000 for the control and elimination of LF. Under this program, anti-filarial drugs ivermectin, diethylcarbamazine and albendazole (IDA) were administered to the populations residing in endemic countries. However, these drugs are only effective against the larval stages of filarial parasites and have almost no effect on adult parasites. Not only this, the drugs are commonly associated with several side effects, and complications in co-infections also their continuous use leads to drug resistance in the parasites.² Therefore, there is an urgent requirement to develop new anti-filarial agents that have adulticidal activity and are effective in lower doses.

A significant subfield of nanoscience and nanotechnology is now bioinspired technology for nanoparticle (NP) production.

In addition to their numerous uses in the synthesis of NPs, plant biomass is primarily targeted as a catalyst for chemical synthesis. According to recent studies, silver nanoparticles (AgNPs) have a variety of medical uses, including cancer diagnosis and treatment as well as anti-bacterial, anti-fungal, anti-diabetic, and anti-inflammatory properties.³ In terms of resource accessibility, security, reaction speed, and the viability of large-scale synthesis, using plant extracts to synthesize AgNPs is preferable. According to reports, the phytochemicals found in plant extracts can reduce metal ions to nanoparticles and eventually eliminate the need for harmful chemicals, high pressure, temperature, and energy.⁴ To create NPs of various sizes and shapes, a variety of plant materials, including leaf extracts, fruit, bark, fruit peels, roots, and calluses, have been investigated.⁵

The genus *Ocimum* has several medicinal properties such as anti-viral, anti-microbial, anti-fungal, anti-helminthic and is also used as a mosquito repellent.⁶ Some prior studies have shown the anthelmintic activities of *Ocimum* and its essential oils against different gastrointestinal nematodes.^{7,8} In a previous study, we investigated the anti-filarial activity of *Ocimum sanctum* (OS) and bioactive compounds against filarial parasites *Setaria cervi* under *in vitro* and *ex vivo* conditions.⁹ The findings from the study provided evidence for the multi-inhibitory potential of *Ocimum sanctum* against several filarial

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proteins/enzymes. The green synthesized silver nanoparticles of *Ocimum* sp. have been reported for their antibacterial and antimicrobial activities.^{10,11} However, no reports are available for the anti-parasitic or anti-filarial activity of green synthesized nanoparticles of *Ocimum sanctum*. Hence, the current work was undertaken to explore the anti-filarial activity of *Ocimum sanctum* silver nanoparticles (OSAgNPs). The study findings may offer fresh perspectives for developing environmentally responsible and long-lasting treatments for filarial infections as well as aid in addressing the issue of drug resistance in present therapies.

2. Experimental section

2.1. Preparation of *Ocimum sanctum* extract

Fresh healthy leaves of *Ocimum sanctum* Linn. (OS) were collected from the botanical gardens of Banaras Hindu University, U.P. India. The OS leaves were identified by a senior taxonomist Prof. Shashi Pandey, Department of Botany, Institute of Science, Banaras Hindu University. Leaves were first rinsed with distilled water to remove the dust and shade dried for a week on the tabletop.⁹ Fine powder of OS leaves was prepared with the help of a mixer grinder and stored at 4 °C for further use. The 5% aqueous leaf extract was prepared from a fine powder of OS leaves. 5 g of leaf powder was boiled in 100 ml sterile distilled water at 60 °C for 40 min. Further, the leaf extract was filtered using Whatman filter paper No. 1 and the light brown solution was stored at 4 °C for further use.¹²

2.2. Biosynthesis of *Ocimum sanctum* silver nanoparticles (OSAgNPs)

5 ml aqueous extract of OS was added to 100 ml of 2 mM AgNO₃ solution. The mixture was left for stirring at 25 °C overnight. The color of the solution changed over from light brown to colloidal dark brown indicating the formation of silver nanoparticles. The OSAgNPs were separated by centrifugation at 12 000 rpm for 30 min followed by three washings with Mili Q water. Then, the OSAgNPs were dried in a lyophilizer and stored at 4 °C for further studies.¹³

2.3. Characterization of OSAgNPs

2.3.1. UV-visible spectroscopy. OSAgNPs were inspected visually as the color changed in solution from light brown to dark colloidal brown *i.e.*, an indication of OSAgNPs formation. Further, the absorption spectra of OSAgNPs solution were recorded from 300 to 700 nm over a period of 24 hours with a UV-Vis Spectrophotometer (Biotek synergy H1 microplate reader).

2.3.2. Fourier-transform infrared spectroscopy (FTIR). For chemical identification, *Ocimum sanctum* L. aqueous leaf extract and synthesized OSAgNPs were subjected to Fourier Transform Infrared Spectrophotometer (FTIR) (PerkinElmer Spectrum Version 10.4.3) by using the potassium bromide (KBr) pellet method. The *Ocimum sanctum* extract and OSAgNPs were first lyophilized, then 1 mg of either sample was mixed with 200 mg of potassium bromide powder and pulverized. The pulverized

mixture was fixed in a pellet-forming die and a pressure of ~8 tons was applied under a vacuum. Further, the characterization was done at a resolution of 4 cm⁻¹ in the range of 4000–500 cm⁻¹.

2.3.3. XRD and EDX studies. The XRD pattern of OSAgNPs was obtained at the central instrument facility, Department of Chemistry, BHU, Varanasi. The X-ray diffraction spectra were recorded in a D8 ECO ADVANCE (Bruker) with the range of 20° to 80° at 2θ angles for the identification of phase variety and grain size of synthesized OSAgNPs. The XRD analysis was done with CuKα radiation at a voltage of 40 kV and 15 mA with a scan rate of 0.083/s. Elemental identification of OSAgNPs was done by energy dispersive spectroscopy (EDAX, Zeiss Germany) at an accelerating voltage of 0–20 keV.

2.3.4. SEM and TEM imaging. The OSAgNPs samples for SEM imaging were mounted on an aluminum stub and sputter coated with colloidal gold and images were taken at 20 kV in a Scanning electron microscope Carl Zeiss EVO18 (Germany) at the Dept. of Geology, BHU, Varanasi, India.¹⁴ The fine powder of OSAgNPs was dispersed in Mili Q water by ultrasonication and coated on a carbon copper grid. Next, the HRTEM images were obtained in a high resolution transmission electron microscope, FEI-Tecnaï G2 20 TWIN at the central instrument facility, IIT, BHU, Varanasi, India. An accelerating voltage of 200 kV was applied¹⁵ and the OSAgNPs were visualized with Resolve RT software. The images of nanoparticles were recorded at scale bar of 200 to 10 nm and the particle sizes were measured in Image J software.

2.4. Parasite collection and maintenance

Adult live female *S. cervi* worms were recovered from the peritoneal folds of Indian water buffaloes from local slaughterhouses. Worms were brought to the laboratory in commercially available Krebs's Ringer bicarbonate buffer (KRB) supplemented with streptomycin (100 µg ml⁻¹), penicillin (100 I.U. per ml), glutamine (5 mM), and 0.5% glucose [KRB Maintenance Medium (KRBMM)]. Worms were washed with KRB and incubated in KRBMM at 37 °C in a CO₂ incubator for an hour before subsequent treatments.⁹

2.5. Exposure of adult female *S. cervi* worms to OSAgNPs

Adult female *S. cervi* worms in equal numbers ($n = 10$) were incubated in 20 ml of KRB maintenance medium with different concentrations of OSAgNPs (0.1 µg ml⁻¹, 0.5 µg ml⁻¹, and 1 µg ml⁻¹) for 4 h at 37 °C, 5% CO₂ and 95% humidity in a CO₂ incubator. *S. cervi* worms maintained in KRB maintenance media only served as the control group. After 4 h of incubation *S. cervi* worms were allowed to recover for 1 h in fresh KRB maintenance medium. Next, worms were thoroughly washed with PBS, homogenized, and stored at –20 °C for further use.¹⁶ The protein estimation was done by Bradford's method¹⁷ and Bovine Serum Albumin (BSA) was used as a standard.

2.6. Effect of OSAgNPs on parasite motility and viability

The motility of *S. cervi* parasites exposed to OSAgNPs was monitored by visual inspection at hourly intervals, by an investigator blinded to the experiment. The movement of



worms was scored with either positive or negative signs (+/−).¹⁸ The recovery of filarial worms was checked at the end of the experiment by transferring the worms for 1 h into a fresh KRBMM (without OSAgNPs). The viability of control and treated *S. cervi* parasites was determined by MTT assay according to the method of Mosmann and Coffman.¹⁹ *S. cervi* worms were incubated in PBS medium containing 0.5 mg ml^{−1} MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) for 2 h at 37 °C in dark. The worms were then transferred into 200 μl dimethyl sulphoxide (DMSO) for solubilizing the formazan crystals. After 1 h of incubation, the OD of the solution was measured at 540 nm in a microplate reader (BioRad). All experiments were done in triplicates.

2.7. Estimation of total reactive oxygen species (ROS) production

Total ROS generation was performed on control and OSAgNPs treated *S. cervi* worms by the Nitro blue tetrazolium (NBT) method.²⁰ Firstly, worms were incubated for 1 h in 2% NBT solution at room temperature followed by numerous washings with PBS followed by methanol. Then worms were incubated in 2 M KOH to disrupt the cell membrane and then in DMSO to dissolve the formazan crystals and finally the absorbance was recorded at 620 nm.

2.8. High resolution accurate mass spectrometry (HRAMS) analysis

2.8.1. Sample preparation. 100 μg of protein sample was precipitated in ice-chilled acetone and incubated at −20 °C for overnight. Next, the precipitated protein sample was centrifuged at 10 000 rpm for 10 min at 4 °C. The pellet was dissolved in freshly prepared buffer (40 mM Tris-HCl, pH-7.5, 7 M Urea, 2 M Thiourea, 15 mM DTT) at room temperature, and the dissolved sample was digested with trypsin (Sequencing grade, Promega) at 37 °C for overnight. The extracted peptides were lyophilized and desalted using C18 cartridges. Finally, the peptides were stored at −80 °C for further proteomics analysis.²¹

2.8.2. HRAMS analysis. The sample was subjected to proteomic analysis using high-resolution accurate mass spectrometer facility at Central Discovery Centre, Banaras Hindu University. An Orbitrap Eclipse Tribrid Mass Spectrometer with nano LC and UHPLC was used for peptide analysis. The samples were analyzed using a 120 min linear gradient of buffer B (80% Acetonitrile and 0.1% Formic acid) at a flow rate of 0.300 μl min^{−1}. Full MS scans were done in the range of 200–1600 *m/z* and Thermo Scientific™ Proteome Discoverer™ software was used for peptide identification. For LC-MS/MS analysis control and OSAgNPs treated *S. cervi* protein samples were run in triplicates. The ratio of the mean relative quantitative values for each protein in subsequent samples was used to compute the abundance ratio. The significance of the variation between the samples from the comparison group and the samples from the control group was assessed using *T*-tests. The significance index was calculated using the related *P* value, with *P* < 0.05 being used as the default.

2.9. Gene ontology (GO) annotation and protein–protein interaction (PPI) network analysis

The Gene Ontology (GO) annotation proteome analysis in the UniProt database (<https://www.uniprot.org/>) was done. The accession no. of the respective proteins were searched, in the UniProt database and, the corresponding UniProt IDs were retrieved. Major proteins were characterized into groups by GO annotation, based on molecular function (MF), cellular components (CC), and biological processes (BP). For the classification and visualization of MF, CC, and BP of proteins, histograms were prepared. Further PPI networks for all the differentially expressed proteins using STRING database version 11.5 were constructed.²²

2.10. Statistical analysis

All *in vitro* experiments were performed in triplicates and repeated thrice. The data are expressed as mean ± SD which was calculated using Graph Pad Prism 8.1 software. The statistical significance between control and OSAgNPs treated *S. cervi* worms was calculated with Student's *t*-test. **P* < 0.05 were considered as significant in comparison to control (**P* < 0.05, ***P* < 0.01, ****P* < 0.001 and *****P* < 0.0001).

3. Results and discussion

3.1. Physicochemical characterization of OSAgNPs

The physicochemical characteristics of biosynthesized OSAgNPs were evaluated by using techniques like UV-visible spectroscopy, XRD, FTIR, SEM, and TEM.

3.1.1. Ultraviolet-visible spectroscopy. The reduction of silver ions into nanoparticles with plant extract can be easily monitored by visual inspection of color change due to the surface plasmon resonance (SPR). The color of the OSAgNPs solution changed from light brown or pale yellow to dark colloidal brown within 30 minutes due to the surface plasmon resonance phenomenon. The color change is an indication of the reduction of silver ions into silver particles (Fig. 1A). The formation of silver nanoparticles was further confirmed by UV-VIS spectra recorded as a function of time for 24 h in the range of 300–700 nm. The absorbance maxima of OSAgNPs was observed at 430 nm (Fig. 1B). It was seen that the intensity of the brown color increased with time because of the continuous formation of OSAgNPs in the solution. The excitation of longitudinal plasmon vibrations of silver nanoparticles in the solution phase led to the characteristic absorbance maxima of 430 nm observed in the *Ocimum sanctum* silver nanoparticles (OSAgNPs) solution. The observed absorbance maxima was consistent with previous researches on biogenic silver nanoparticles.^{13,23} Also, the SPR bands were constant at 430 nm for 24 h indicating the absence of aggregation of nanoparticles.

3.1.2. Fourier transform infrared spectroscopy. FTIR analysis was performed to study the possible interactions between the functional groups of biomolecules present in *Ocimum sanctum* leaf extract and silver ions. For FTIR measurements different methods can be employed such as



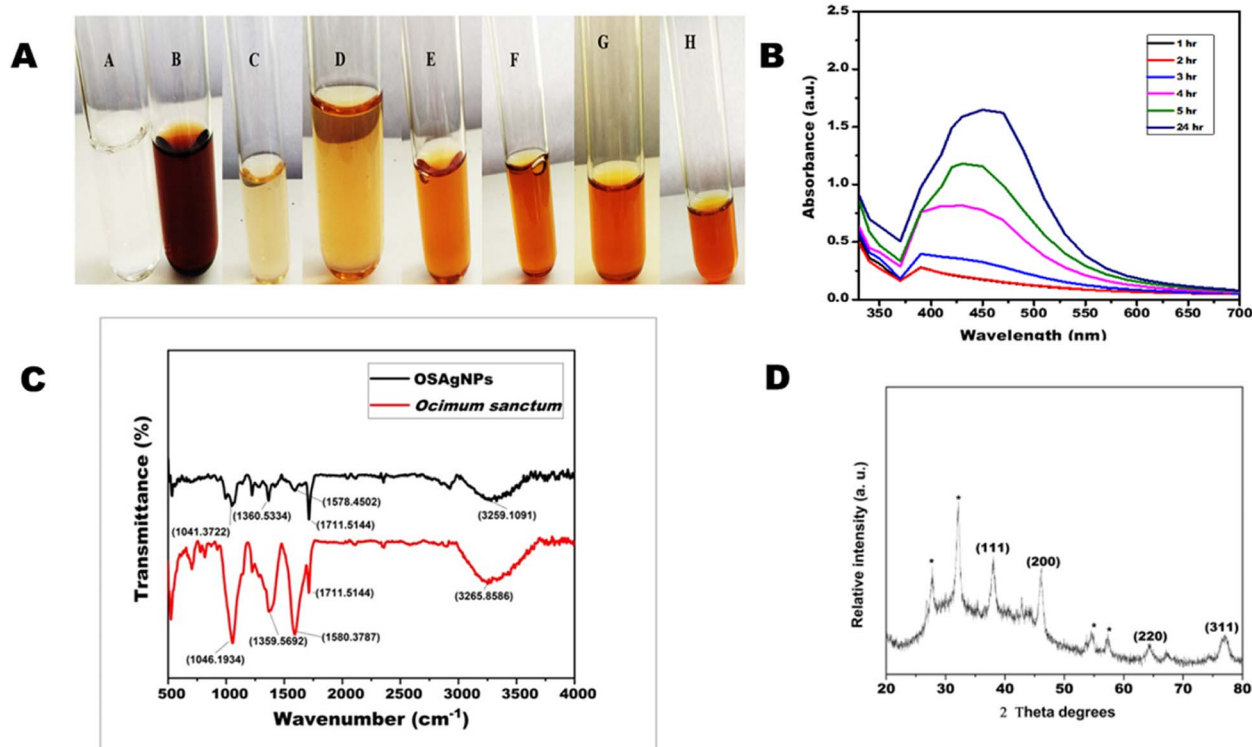


Fig. 1 (A) Change in the color of AgNO_3 solution with time after mixing with *Ocimum sanctum* extract. (A) AgNO_3 solution; (B) *Ocimum* extract; (C) mixture of AgNO_3 solution and *Ocimum sanctum* extract at 0 h; (D) after 1 h (E) after 2 h (F) after 3 h (G) after 5 h; (H) after 24 h. (B) The UV-visible absorption spectrum of silver nanoparticle solution produced from aqueous extract of *Ocimum sanctum* after different time intervals. (C) Fourier-transform infrared absorptive spectra of *Ocimum sanctum* extract (red) and biosynthesized silver nanoparticles (OSAgNPs) (Black). (D) X-ray diffraction (XRD) pattern of green synthesized silver nanoparticles of *Ocimum sanctum* (OSAgNPs).

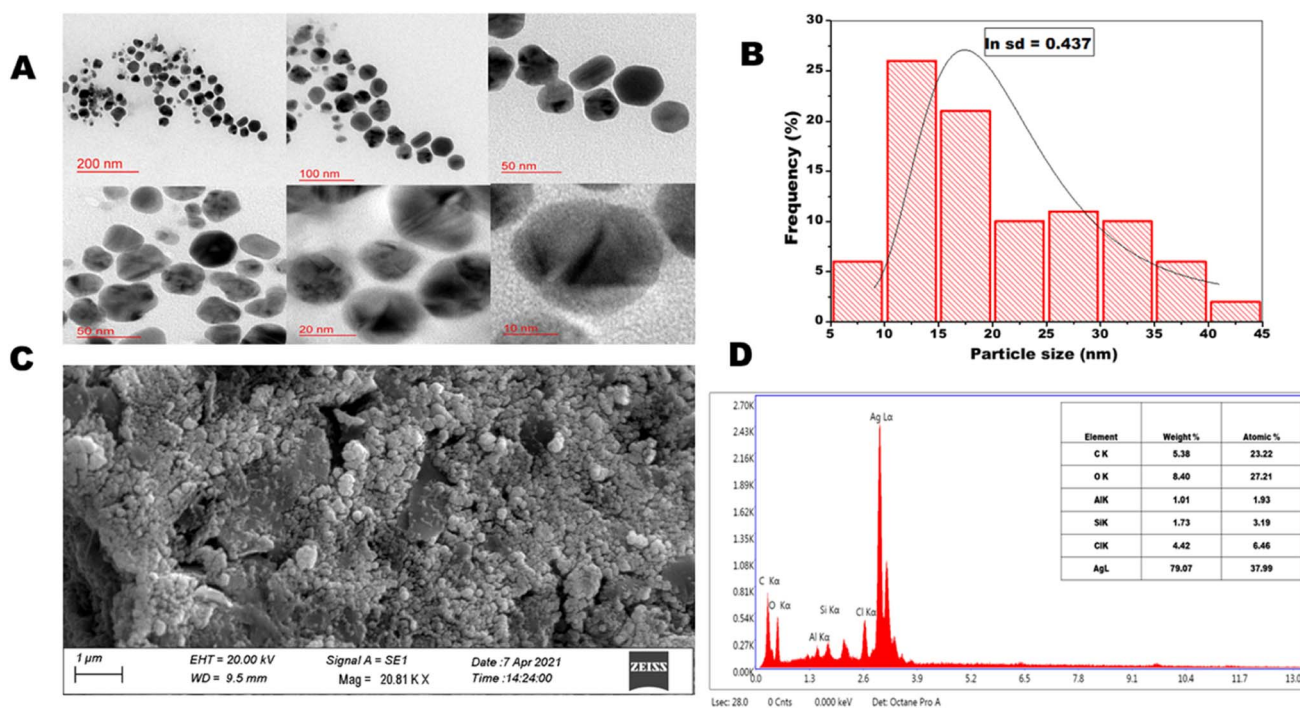


Fig. 2 (A) HRTEM images at scale bar 200, 100, 50, 20, and 10 nm, spherical silver nanoparticles of OSAgNPs. (B) Size distribution histogram of OSAgNPs. (C) SEM image of green synthesized OSAgNPs. (D) Energy dispersive X-ray spectrum and chemical composition of OSAgNPs.



KBr pellet method, Nujol method and diffuse reflection method. In the Nujol method liquid paraffin is used, however to confirm sample absorbance in regions overlapping with liquid paraffin sample, measurements should be repeated with another solvent hexachlorobutadiene. Similarly in the diffuse reflection method some low absorbance bands are also emphasized more than the transmission spectrum, hence we opted for the KBr pellet method. The band values 3285, 2923, 2356, 1710, 1589, 1361, 1212, 1049 cm^{-1} and 3257, 2895, 2348, 1709, 1580, 1368, 1212, 1051 cm^{-1} were obtained for OSAgNPs and OS leaf extract, respectively (Fig. 1C). The FTIR spectra of OSAgNPs and *Ocimum sanctum* leaf extract had several overlapping peaks in the range of 2348 cm^{-1} to 1049 cm^{-1} , confirming the presence of OS extract in the OSAgNPs. The peaks at 3248 cm^{-1} and 1368 cm^{-1} correspond to O–H stretching whereas the 2348 cm^{-1} peak corresponds to O=C=O stretching. The FTIR peaks at 170 cm^{-1} and 1051 cm^{-1} correspond to C=O stretching and the major peak at 1580 cm^{-1} denotes the N–N bending of amines. The peak at 700 cm^{-1} in OS leave extract was absent in OSAgNPs indicating that amine components were oxidized in the process of silver ion reduction. The peak at 3257 cm^{-1} of OS leaf extract shifted to 3285 cm^{-1} in OSAgNPs FTIR spectra and was less intense as the amine groups of proteins may be functioning as reductants for silver ions in nanoparticles. These findings are similar to other earlier studies on biogenic silver nanoparticles synthesized from *C. annum* and *A. paniculata* respectively.^{13,23}

3.1.3. X-ray diffraction. The XRD technique can determine the chemical composition and crystalline structure of nanoparticles. Fig. 1D shows the XRD pattern of green OSAgNPs and the diffraction peaks were obtained at 38°, 45°, 64°, and 77° which represent (111), (200), (220), and (311), respectively. The peaks correlate with the standard miller indices values for powdered diffraction of silver of the Joint Committee on Powdered Diffraction Standards (JCPDS) (File No. 04-0783).²⁴ The broad peaks at 38°, 45°, 64°, and 77° corresponded to face centered cubic lattice arrangement and crystalline structure of silver nanoparticles.¹⁵ In conjunction, some unassigned peaks were also observed in the XRD spectra which could be due to the formation of a bio-organic phase on the top surface of the nanoparticles.²⁵ For determining the size of nanoparticles, the Debye–Scherrer's equation interrelating the average crystalline domain size (D) perpendicular to the reflecting planes, λ the X-ray wavelength, and β the FWHM of diffraction peak and θ which is the diffraction angle as

$$D = 0.9\lambda/\beta \cos \theta$$

was used. The size of the OSAgNPs was calculated from the full width at half maximum (FWHM) of the peak in radians (111) and was found to be ~16.98 nm.

3.1.4. High resolution transmission electron microscopy and scanning electron microscopy. The high resolution transmission electron microscopic images of OSAgNPs are presented in Fig. 2A. The HRTEM analyses showed the particle size to be between 5 to 45 nm (Fig. 2B) which was

estimated by constructing a histogram to deduce the mean particle size. The HRTEM images clearly showed prepared OSAgNPs were nearly spherical but varied in size and the mean particle size of OSAgNPs was 17 nm at pH 7.0., which is very close to the size of the OSAgNPs as estimated with the XRD analysis. It was also observed that the edges of the particles were lighter than the centers, indicating that biomolecules from *Ocimum sanctum* extract would be involved in capping the silver nanoparticles. The surface morphology of synthesized OSAgNPs was checked by scanning electron microscope imaging using a Zeiss SEM system (Fig. 2C). The shape of the nanoparticles can vastly affect their electronic and optical properties and the SEM images clearly showed that the synthesized OSAgNPs were uniform, and smooth in surface morphology.

3.1.5. Energy-dispersive X-ray analysis. The energy dispersive X-ray (EDAX) analysis of OSAgNPs was performed at an accelerating voltage of 20 kV and take-off angle of 74.9°. EDAX results showed a strong signal in the Ag (silver) region and weak

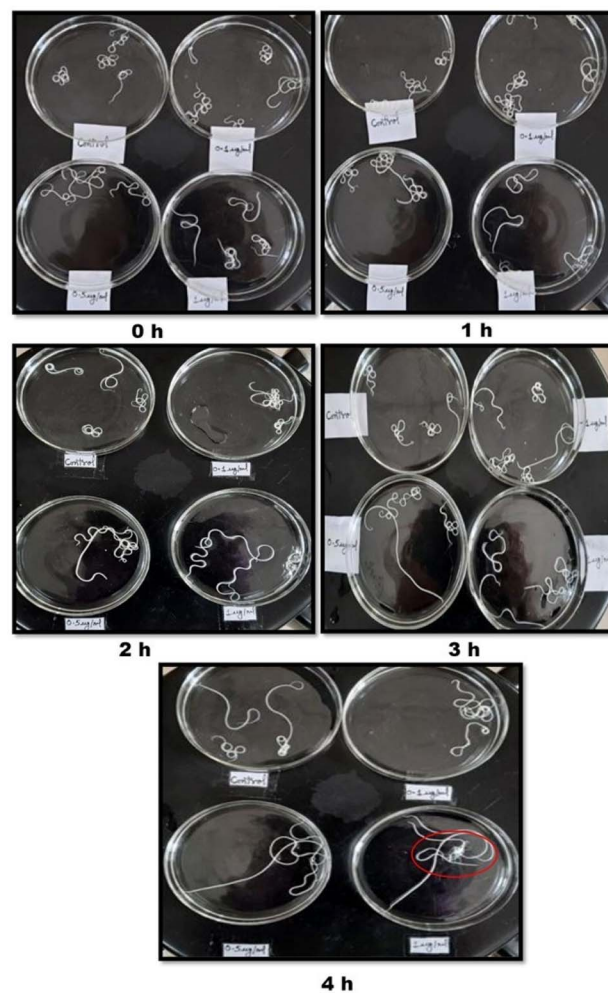


Fig. 3 Adult worms ($n = 10$) of equal size were exposed at 0.1 $\mu\text{g ml}^{-1}$, 0.5 $\mu\text{g ml}^{-1}$ and 1 $\mu\text{g ml}^{-1}$ of OSAgNPs in 20 ml of KRBBM at 37 °C. Images of control and OSAgNPs treated female *S. cervi* worms, after 0, 1, 2, 3 and 4 h of treatment.



signals in the O, C, and Cl regions (Fig. 2D). The quantitative results of EDAX spectra of OSAgNPs confirmed silver (Ag) as the major constituent element (79.07.12%) of the green synthesized OSAgNPs. The other elements present in OSAgNPs were carbon (5.38%), oxygen (8.40%), and chlorine (4.42%). The minor components carbon and oxygen are the main constituents of the OS leaf extract. The capping effect of organic compounds found in the plant extracts often leads to an increase in the stability of the nanoparticles.²⁶

3.2. Effect of OSAgNPs on the motility and viability of *S. cervi*

The motility of *S. cervi* exposed to OSAgNPs reduced in a time and concentration-dependent manner (Fig. 3). The exposure of *S. cervi* to OSAgNPs at 1 $\mu\text{g ml}^{-1}$ concentration resulted in the death of parasites after 3 h of incubation which was confirmed by transferring them to a flask of fresh KRB maintenance medium. These parasites did not show any sign of recovery even after incubation for an hour in the maintenance medium (Table 1). On the other hand, AgNO₃ solution was much less effective in reducing the motility of *S. cervi* at 1 $\mu\text{g ml}^{-1}$ concentration, the parasites easily recovered and became motile after they were transferred to fresh KRBMM.¹³ The viability of adult female *S.*

cervi after treatment with OSAgNPs was estimated by MTT assay. A positive correlation between viability and motility was observed in OSAgNPs treated female *S. cervi* worms. The viability after exposure to 0.1 $\mu\text{g ml}^{-1}$, 0.5 $\mu\text{g ml}^{-1}$ and 1 $\mu\text{g ml}^{-1}$ nanoparticles was reduced to 65%, 40% (P value ≤ 0.001) and 19% (P value ≤ 0.001), after 4 h of incubation, when compared with the control parasites (100% viability) (Fig. 4A). The LC₅₀ value of OSAgNPs treatment was observed at a concentration of 0.36 $\mu\text{g ml}^{-1}$. In earlier studies, the LC₅₀ value of *Andrographis paniculata* silver nanoparticles and *Acacia auriculiformis* silver nanoparticles were reported to be 11.6 $\mu\text{g ml}^{-1}$ and 5.61 $\mu\text{g ml}^{-1}$, respectively.^{13,27} The LC₅₀ value of OSAgNPs is much lower in comparison to other plant based silver nanoparticles. Previously, the effect of *Ocimum sanctum* leaf extract was also tested on the motility of *S. cervi* parasites.⁹ The OS leaf extract at the concentration of 250 $\mu\text{g ml}^{-1}$ and above could significantly inhibit *S. cervi* motility after 5 h of incubation, while, OSAgNPs treated adult parasites failed to revive after 4 h of incubation. The LC₅₀ value of OS leaf extract was 197.24 $\mu\text{g ml}^{-1}$ which reveals that the anti-filarial efficacy of OSAgNPs has increased by 547.88 folds. The motility and viability studies noticeably demonstrated that OSAgNPs are potent anti-filarial adulticides even at very low concentrations.

Table 1 Effect of OSAgNPs on the motility of adult female *Setaria cervi* at different time intervals

Experimental sample (concentration)	Motility of adult <i>S. cervi</i> at different time intervals					
	0 h	1 h	2 h	3 h	4 h	Recovery ^a
Control	++++	++++	++++	++++	++++	++++
OSAgNPs (0.1 $\mu\text{g ml}^{-1}$)	++++	++++	++++	++++	++++	++++
OSAgNPs (0.5 $\mu\text{g ml}^{-1}$)	++++	++++	++++	++	+	++
OSAgNPs (1 $\mu\text{g ml}^{-1}$)	++++	++++	++	–	–	–

^a Motility was checked by visual observation at different time intervals. Adult female *S. cervi* ($N = 5$) of equal size were treated with different concentrations of OSAgNPs. Incubation was done for 4 h in a 20 ml maintenance medium. Motility scored was denoted as '–' no movement; '+' least active; '++' less active; '+++', moderate active; '++++' highly active. Worms of each flask were transferred into a fresh medium free of OSAgNPs and compared with the control group to observe recovery.

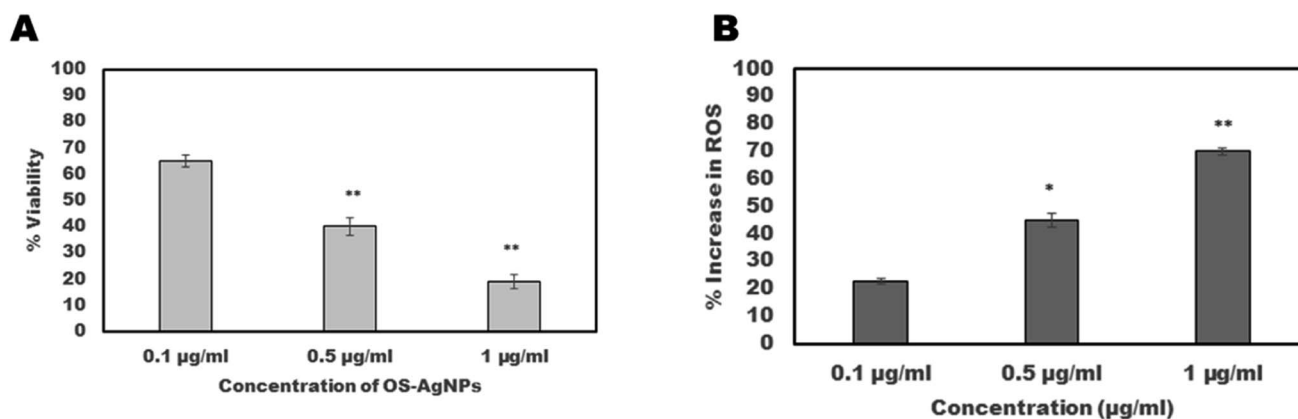


Fig. 4 Adult worms of equal size were exposed at 0.1 $\mu\text{g ml}^{-1}$, 0.5 $\mu\text{g ml}^{-1}$ and 1 $\mu\text{g ml}^{-1}$ of OSAgNPs in 20 ml of KRB maintenance medium at 37 °C. (A) The viability of parasites was determined by MTT assay after 4 h of incubation, percent reduction in viability was calculated with respect to the control group of *S. cervi* worms. (B) Total reactive oxygen species production assessed in percentage after exposure of adult female *S. cervi* worms to OSAgNPs. The data expressed are mean \pm SD of three independent experiments. ** $P \leq 0.001$, * $P \leq 0.05$. Values with $P \leq 0.05$ were considered significant.



Table 2 List of upregulated proteins in control vs. OSAGNPs treated group

Accession	Description	Abundance ratio: (treated)/(control)	Abundance ratio <i>P</i> -value: (treated)/(control)	Score sequent HT
A0A4E9FJB5	Myosin heavy chain B (MHC B), putative	2.876	2.58089×10^{-7}	428.86
A0A4E9EPZ8	Troponin family protein	2.067	0.000279915	162.66
A0A3P7GKE0	Protein-serine/threonine phosphatase	2.464	8.99351×10^{-6}	4.08
J9EZR8	T-complex protein 1 subunit gamma	2.962	1.24132×10^{-7}	174.99
J9FCK5	Peptide-methionine (S)-S-oxide reductase	3.486	1.5685×10^{-9}	28.57
A0A0H5SFI5	Cystathionine gamma-lyase	2.066	0.00028413	3.42
J9F677	Proteasome subunit beta	1.498	0.03145365	44.44
A8Q167	Fibronectin type III domain containing protein	1.53	0.024579067	27.11
J9DVN6	Ig-like domain-containing protein	1.782	0.003176492	2.28
A0A4E9EW72	Hypothetical 24.1 kDa protein C01C10.2 in chromosome X, putative	1.744	0.004360374	0
A0A1I8EA17	Translation machinery-associated protein 7 homolog	2.012	0.000452006	18.63
A0A1I8EQR7	Uncharacterized protein	2.12	0.000177617	6.31
J9EZA7	RRM domain-containing protein	1.708	0.00587942	18.04
A0A4E9F572	Uncharacterized protein	1.909	0.001093537	17.55
A0A1I8EDU1	Uncharacterized protein	1.614	0.012592256	46.54
A0A0K0IZG5	V-type proton ATPase subunit G	10.228	1×10^{-17}	2.55
A0A4E9FEI0	Eukaryotic translation initiation factor 3 subunit L	1.945	0.00080233	38.28
A0A4E9FAN7	Troponin T, putative	2.33	2.88402×10^{-5}	100.89
J9B7U2	Uncharacterized protein (Fragment)	1.856	0.001701183	245.07
J9FGC0	RNA recognition domain-containing protein	1.797	0.002817386	12.1
A0A1I8EZZ0	Uncharacterized protein	2.767	6.52958×10^{-7}	565.66
A0A5S6PD06	Uncharacterized protein	2.856	3.0594×10^{-7}	7.75
A0A4E9F302	Uncharacterized protein	3.566	8.16814×10^{-10}	1.94
Q04010	Major body wall myosin	1.596	0.014566794	376.23
A0A3P7GK84	Myosin motor domain-containing protein	6.101	1×10^{-17}	141.91
A0A1P6BIP7	BMA-MYO-3	1.904	0.001131896	1079.53
A0A0K0J5T8	DNA-directed RNA polymerase subunit beta	1.54	0.022774414	5.47
A0A4E9F0V9	Succinate-CoA ligase [ADP-forming] subunit beta, mitochondrial	2.732	8.84871×10^{-7}	115.1
A0A0J9XRM8	Bm14768	1.57	0.017937114	19.38
J9E141	Cystathionine gamma-lyase	2.066	0.00028413	46.78
A0A3P7DV80	Superoxide dismutase [Cu-Zn]	1.563	0.01891622	35.86
A0A0J9XPV1	NADH dehydrogenase [ubiquinone] 1 alpha subcomplex subunit 12	2.593	2.9264×10^{-6}	2.08
A0A1I8ENX1	PITH domain-containing protein	2.57	3.58715×10^{-6}	4.32
A0A0I9N6H9	BMA-MLC-3	2.003	0.000487386	365.3
A0A1I8EDQ8	Myosin tail family protein	2.289	4.116×10^{-5}	1162.25
A0A4E9FKG6	Tropomyosin family protein	1.809	0.002544887	172.22
E3UV59	Glutathione S-transferase	1.729	0.004939501	5.48
A0A4E9F8C5	Ribokinase	3.63	4.87338×10^{-10}	10.3
A0A1I8EJF9	Phosphoglycerate mutase (2,3-diphosphoglycerate-independent)	19.675	1×10^{-17}	267.39
A0A1I8EW76	60S ribosomal protein L18	1.669	0.008113791	4.47
J9F341	Ubiquitin carboxyl-terminal hydrolase	1.509	0.028919869	15.5
A0A1I8EA31	ANK_REP_REGION domain-containing protein	1.764	0.003696815	0
A0A0K0J4J0	NADH dehydrogenase [ubiquinone] flavoprotein 1, mitochondrial	1.607	0.013372543	11.74
J9EMX1	Eukaryotic translation initiation factor 3 subunit K	1.817	0.002374081	9.89
A0A3P7DI98	Uncharacterized protein	1.502	0.030415891	2.16
A0A4E9FDF7	NADAR domain-containing protein	1.643	0.009969112	1.61
J9EFL6	Uncharacterized protein (Fragment)	2.515	5.76792×10^{-6}	82.5
A0A3P7DJC8	Uncharacterized protein	2.247	5.88975×10^{-5}	50.08
A0A1P6CFB8	60S ribosomal protein L7a	2.172	0.000113527	52.15
A0A7I4NMH9	Uncharacterized protein	1.935	0.000869051	330.94
A0A0K0JJA5	Non-specific serine/threonine protein kinase	3.057	5.53284×10^{-8}	1.93
J9EM29	Uncharacterized protein (Fragment)	2.247	5.88587×10^{-5}	7.03
J9E9B2	Uncharacterized protein	1.533	0.023965505	35.65
A0A0J9XP99	BMA-PRPF-4, isoform b	1.896	0.001220539	2.32
A0A4E9F8S5	WD-repeat protein WDC146, putative	1.757	0.003911261	12.73
A0A1I8ERN7	Uncharacterized protein	1.572	0.017604686	28.99
A0A0K0J064	BMA-TIN-10	2.074	0.00026445	6.43



Table 2 (Contd.)

Accession	Description	Abundance ratio: (treated)/(control)	Abundance ratio <i>P</i> -value: (treated)/(control)	Score sequest HT
J9EZX6	Uncharacterized protein	1.984	0.000573261	2.04
J9EY11	Phosphatidylinositol-4,5-bisphosphate 4-phosphatase	1.685	0.007090721	1.77
A0A4E9EX33	Rasputin, putative	1.632	0.010902888	5.34
P90689	Actin	2.138	0.000152517	6421.99
A0A0J9XXG2	Bm4971, isoform c	2.515	5.77475×10^{-6}	916.83
A0A0K0JXX0	Bm9047	1.749	0.004203347	7.77
A0A1I8EG76	Sorting nexin	1.604	0.013650175	4.77
A0A1I8ESS3	Apple domain-containing protein	1.677	0.007605868	5.17
J9EF28	Uncharacterized protein (Fragment)	1.915	0.001035958	132.21
A0A4E9FQW3	Protein-tyrosine-phosphatase	1.814	0.002432581	4.07
A0A4E9FQE3	Importin-beta N-terminal domain containing protein	2.184	0.000102445	17.52
J9B374	Sorting nexin-12	1.596	0.014622216	46.04

3.3. Effect of OSAgNPs on reactive oxygen species production in *S. cervi*

A colorimetric assay employing nitro-blue tetrazolium (NBT) substrate was used for estimating reactive oxygen species production following exposure of parasites to OSAgNPs. The increase in ROS production by female *S. cervi* was directly proportional to the concentration of OSAgNPs (Fig. 4B). The increase in reactive oxygen species was significantly higher in parasites treated with $0.5 \mu\text{g ml}^{-1}$ (P value ≤ 0.05) and $1 \mu\text{g ml}^{-1}$ (P value ≤ 0.001) of OSAgNPs. The generation of reactive oxygen species (ROS) due to cellular metabolic activities is a regular phenomenon in *S. cervi* parasites. However, an increase in ROS levels, can cause oxidative stress which may lead to apoptosis and ultimately culminate in death. It was observed that the exposure of filarial parasites to much lower concentrations of OSAgNPs resulted in significantly higher ROS production, in comparison to other biogenic silver nanoparticles.¹³

3.4. Effect of OSAgNPs on the *S. cervi* proteome by high resolution accurate mass spectrometry

The application of proteomic profiling analysis through HRAMS was applied to investigate differentially expressed proteins in OSAgNPs treated filarial parasites. The proteome profiling, data was analyzed through Thermo Scientific™ Proteome Discoverer™ software and the comparison of control worms with OSAgNPs treated *S. cervi* was done. The alteration in the protein expression was analyzed based on the abundance ratio. An abundance ratio value of ≤ 0.67 was set for downregulated proteins and for upregulated proteins the cut-off value was set to ≥ 1.5 . It was found that 213 proteins were differentially expressed (DEPs) after exposure of filarial parasites to OSAgNPs (Tables 2 and 3), out of which 144 proteins were significantly downregulated whereas 69 were significantly upregulated. Heat shock proteins HSP-70, small heat shock protein, p27 and signaling proteins such as Signal Recognition protein (SRP)-14, SRP-54, and SRP- receptor were highly downregulated after the treatment. The energy metabolism proteins/enzymes such as Guanylate kinase which catalyzes ATP-dependent phosphorylation of GMP into GDP, mannose-6 phosphate isomerase catalyzing conversion of mannose-6 phosphate to

fructose-6 phosphate and, NADH-ubiquinone oxidoreductase 24 kDa subunit (complex-I) of electron transport chain were also found to be downregulated. The crucial enzymes Guanylate kinase and Adenylate kinase, play a role in salvaging nucleotides GMP and AMP, were significantly downregulated thus showing the profound effect of OSAgNPs on nucleotide phosphorylation and energy metabolism. The signal recognition particle (SRP) 14 kDa, SRP-54 kDa, and SRP receptor α subunit were downregulated to an abundance ratio of 0.01 which shows that the targeting of secretory proteins becomes highly compromised after treatment with OSAgNPs. The heat shock protein 70, a stress-responsive molecular chaperone, protects the cell and is implicated as a dominant antigen in parasitic helminths such as *Schistosoma mansoni*, *Onchocerca volvulus*, *Brugia malayi* and *Wuchereria bancrofti*²⁸ was also under-expressed in OSAgNPs treated parasites. The highly antigenic filarial protein BMA-RAL which has been tested as a lymphatic filarial vaccine candidate was significantly downregulated too.²⁹ Moreover, cAMP-dependent protein kinase, an essential enzyme that controls multiple cellular processes by regulating the rate of phosphorylation of serine–threonine in the target proteins was much reduced too. These findings establish that the signal transduction cascades in the filarial worms are severely affected after OSAgNPs treatment.

The major detoxifying enzymes Glutathione S-transferase (GST), Superoxide dismutase (SOD) and Methionine sulfoxide reductase (MSR) that help in scavenging oxidants and act as parasites' first line of defence were upregulated after treatment with OSAgNPs. The enzyme GST and SOD are also involved in xenobiotics metabolism and their upregulation signifies an increased need for detoxification following treatment with nanoparticles. The enzyme Methionine Sulfoxide Reductase (MSR) protects against oxidative damage by reversing methionine sulphoxides to methionine. Similarly, the PITH (C-terminal proteasome interacting domain of Thioredoxin-like) domain containing protein which is involved in transcription, signal transduction and chromatin organization was also upregulated. PITH is present in exosomes/vesicles and, tends to coregulate with mitochondrial protein components. The homologs of the protein are being studied in Alzheimer, Parkinson and certain type of cancer,^{30,31} however, its role in



Table 3 List of downregulated proteins in control vs. OSAGNPs treated group

Accession	Description	Abundance ratio: (treated)/(control)	Abundance ratio <i>P</i> -value: (treated)/(control)	Score sequent HT
A0A4E9F713	Collagen IV NC1 domain-containing protein	0.39	5.31376×10^{-5}	116.96
J9EH11	CULLIN_2 domain-containing protein	0.01	1×10^{-17}	0
A0A4E9EXI3	Uncharacterized protein	0.568	0.020732457	22.32
A0A4E9ERK7	p27, putative	0.668	0.117288598	217.31
A0A0K0JST2	Bm7534	0.556	0.01613457	1.65
A0A0J9XSV8	Calreticulin	0.01	1×10^{-17}	161.07
A0A0J9Y695	Eukaryotic translation initiation factor 3 subunit H	0.498	0.003584272	28.44
A0A4E9FD64	GOLGA2L5 domain-containing protein	0.01	1×10^{-17}	11.44
A0A3P7EM85	RRM domain-containing protein (Fragment)	0.469	0.001388476	12.59
A0A4E9F3S9	Mitochondrial glycoprotein	0.511	0.005198066	15.25
A0A3P7EGS9	PDZ domain-containing protein	0.01	1×10^{-17}	2.23
A0A0I9R2V8	Bm3963, isoform a	0.76	0.331949126	22.57
A0A4E9F9W9	Uncharacterized protein	0.01	1×10^{-17}	2.18
A0A1I8EN40	Guanylate kinase	0.01	1×10^{-17}	9.09
J9EUV2	DNA/pantothenate metabolism flavoprotein	0.652	0.093188782	7.86
J9BCB6	Uncharacterized protein	0.438	0.000452327	72.54
A0A4E9ETN8	Tryparedoxin, putative	0.633	0.069411568	15.31
A0A1I8ERA9	Sulfhydryl oxidase	0.642	0.081051968	8.66
J9F6A2	SH3 domain-containing protein	0.366	1.49581×10^{-5}	2.53
A0A3P7G2R2	H15 domain-containing protein	0.616	0.052815285	15.77
A0A4E9FDH4	Uncharacterized protein	0.568	0.020720135	9.37
A0A4E9EU80	Uncharacterized protein	0.473	0.001627636	0
A0A4E9EXG9	RWD domain-containing protein	0.01	1×10^{-17}	12.11
A0A1I8EQ15	Serine/threonine-protein phosphatase	0.642	0.080969648	32.49
A0A0K0IZH4	U6 snRNA-associated Sm-like protein LSM1	0.269	9.05566×10^{-9}	7.59
A0A4E9ERH0	MHD domain-containing protein	0.01	1×10^{-17}	0
A0A183XC56	Uncharacterized protein	0.66	0.104647575	20.31
A0A1I8ESY8	Uncharacterized protein	0.292	8.06802×10^{-8}	9.87
A0A0K0JCT4	Bm4012, isoform c	0.604	0.042325562	128.7
A0A1I9GCQ7	BMA-CBN-1	0.571	0.02201965	2.6
A0A7I4KFL2	Histone H2B	0.479	0.001972824	76.96
A0A3P7DEX3	Eukaryotic translation initiation factor 3 subunit C	0.649	0.089448353	30.51
A0A1I8EDE4	Uncharacterized protein	0.662	0.108420961	101.17
A0A1I8EVX5	Mannose-6-phosphate isomerase	0.01	1×10^{-17}	13.31
A0A4E9EZZ3	Uncharacterized protein	0.497	0.003400259	5.59
A0A0I9N5X2	BMA-HINT-3	0.01	1×10^{-17}	10.98
A8Q043	cAMP-dependent protein kinase regulatory chain, putative	0.01	1×10^{-17}	10.84
J9FG14	Heat shock 70 protein (Fragment)	0.458	0.000947605	1581.93
J9B0S3	Peptide-methionine (R)-S-oxide reductase	0.613	0.050003528	12.97
A0A4E9F7P5	Intermediate filament protein Ov1, putative	0.498	0.003551624	148.68
A0A4E9EQU4	Uncharacterized protein	0.654	0.096035911	43.53
A0A0I9N461	ADP/ATP translocase	0.664	0.111721821	100.19
A0A5S6PI69	SPARC, putative	0.584	0.028931832	28.78
A0A4E9FJK0	Thioredoxin domain-containing protein	0.666	0.114569485	646.62
A0A3P7FL15	Oxidored_FMN domain-containing protein	0.541	0.011170915	64.16
J9BGY5	Uncharacterized protein	0.01	1×10^{-17}	5.07
A0A4E9FPI2	Protein-L-isoaspartate O-methyltransferase	0.523	0.007061644	19.29
A0A0K0IYI4	Proteasome subunit beta	0.384	3.90786×10^{-5}	12.38
A0A1I8ENA1	ATP-dependent RNA helicase DDX1	0.415	0.000169853	84.71
A0A4E9FGF9	Multifunctional methyltransferase subunit TRM112-like protein	0.01	1×10^{-17}	4.78
J9F2G5	Signal recognition particle 14 kDa protein	0.01	1×10^{-17}	2.35
J9EP49	Papain family cysteine protease containing protein (Fragment)	0.574	0.023624401	4.97
J9F1F7	Splicing factor 45	0.01	1×10^{-17}	0
A0A4E9FRB2	Serine/threonine-protein phosphatase 2A activator	0.319	6.85623×10^{-7}	36.1
A0A3P7GMD3	Uncharacterized protein	0.548	0.0132847	194.96
A0A0H5S5D4	Bm2017	0.654	0.096546488	0
J9ELW9	Chaperonin GroL	0.432	0.000356664	651.97
A0A3P7E0J8	Uncharacterized protein	0.611	0.048638726	18.78
A0A183XS33	PPM-type phosphatase domain-containing protein	0.659	0.104228142	7.46



Table 3 (Contd.)

Accession	Description	Abundance ratio: (treated)/(control)	Abundance ratio <i>P</i> -value: (treated)/(control)	Score sequent HT
J9ELM0	Protein disulfide-isomerase	0.665	0.112101409	157.49
A0A1P6C045	Histone H4	0.322	8.78598×10^{-7}	82.76
A0A4E9FES7	UNC-52/perlecan, putative	0.564	0.019072398	103.28
A0A0H5S4P3	Adenylate kinase isoenzyme 1	0.606	0.0440707	376.45
A0A1I8E9A9	Uncharacterized protein	0.635	0.071944787	104.72
A0A1I8EIH1	Splicing factor 3A subunit 3	0.653	0.095344137	14.46
A0A4E9F8W1	UBC core domain-containing protein	0.643	0.081190023	120.48
J9EZX2	Uncharacterized protein	0.593	0.034436113	15.15
A0A1I8EM83	Leucine-tRNA ligase	0.01	1×10^{-17}	12.29
A0A7I4KM80	Proteasome activator pa28 beta subunit family protein	0.405	0.000107715	19.71
A0A3P7FGQ8	Tyrosine-tRNA ligase	0.629	0.066026078	9
J9FBW7	Small heat shock protein	0.54	0.011032497	15.88
A0A0H5S2P4	60S ribosomal protein L29	0.46	0.001015711	6.39
J9F0A4	TBC domain-containing protein	0.454	0.000821992	4.52
A0A1I8EIF1	UAS domain-containing protein	0.411	0.000146767	11.4
A0A0K0JCV4	Bm4025	0.654	0.095908276	11.41
A0A4E9EUK8	Uncharacterized protein	0.392	5.82324×10^{-5}	5.94
A0A3P7G9V9	Dolichol-phosphate mannosyltransferase subunit 1	0.01	1×10^{-17}	0
J9F457	26S proteasome non-ATPase regulatory subunit Nin1/mts3 family protein	0.64	0.078214319	4.73
A0A3P7EFR0	Uncharacterized protein	0.551	0.014245454	11.33
A0A3P7DS02	Uncharacterized protein	0.558	0.016577961	4.82
A0A0K0JKQ9	Bm5876	0.01	1×10^{-17}	12.57
A8QH13	Transforming growth factor b homolog, putative	0.667	0.116229835	13.78
A0A7I4KGB0	NADH-ubiquinone oxidoreductase 24 kDa subunit, mitochondrial, putative	0.01	1×10^{-17}	5.19
A0A3P7EDP1	Striatin domain-containing protein	0.587	0.030641171	2.92
A0A4E9FC72	Histone H3, putative	0.635	0.071845133	82.83
P90699	Putative RNA binding protein	0.575	0.024181177	15.49
A0A7I4KHH8	BMA-ZYX-1	0.408	0.00012468	2.47
A0A4E9FZZ7	RRM domain-containing protein	0.478	0.00192784	85.38
J9BDI3	60S ribosomal protein L32	0.485	0.002361855	62.91
A0A0K0JD86	BMA-CUL-4	0.01	1×10^{-17}	4.53
A0A0H5S845	Coatomeer subunit zeta	0.461	0.001082192	15.05
A0A0H5S6P0	Bm4536	0.282	3.24203×10^{-8}	32.26
A0A3P7DW72	Uncharacterized protein (Fragment)	0.67	0.121165365	12.46
A0A4E9EYB0	Cleavage stimulation factor, putative	0.01	1×10^{-17}	0
J9EYL6	Uncharacterized protein	0.592	0.034162971	11.31
A0A1I9GCK8	Bm15494	0.01	1×10^{-17}	2.16
A0A4E9FT71	Signal recognition particle 54 kDa protein	0.01	1×10^{-17}	4.94
A0A4E9EYE7	RRM domain-containing protein	0.01	1×10^{-17}	4.58
A0A3P7F7C6	C2H2-type domain-containing protein	0.01	1×10^{-17}	2.27
A0A1I8EI32	ATP-binding cassette	0.639	0.076728778	7.42
A0A1I8EZH2	Phosphorylase b kinase regulatory subunit	0.501	0.003888383	4.1
A0A1I8EC76	T6PP_N domain-containing protein	0.618	0.05412924	6.13
J9F6W8	Uncharacterized protein (Fragment)	0.61	0.047472365	6.33
A0A0K0JR82	BMA-UBC-14	0.01	1×10^{-17}	2.05
A0A1I9G4M2	Bm7621, isoform c	0.41	0.000138793	2.39
A0A1I9G3L0	BMA-MTCH-1, isoform d	0.01	1×10^{-17}	12.43
A0A1I8EMJ6	Fatty acid oxidation complex	0.639	0.076995583	10.78
A0A3P7EML4	26S proteasome non-ATPase regulatory subunit 6	0.673	0.126627232	35.64
A0A1I9G7E2	BMA-ACIN-1, isoform a	0.541	0.011148325	7.98
A0A1I9G795	BMA-ATX-2, isoform j	0.01	1×10^{-17}	10.65
A0A1I8EHC6	WD_REPEATS_REGION domain-containing protein	0.01	1×10^{-17}	7.77
J9DX04	RRM domain-containing protein (Fragment)	0.525	0.007567659	2.29
A0A1P6CD63	Histone H2A	0.596	0.036774833	95.88
J9BHL4	Uncharacterized protein	0.65	0.090764387	6.45
A0A1I8EHZ9	Glutamate-tRNA ligase	0.508	0.004665651	21.48
A0A4E9FA17	Uncharacterized protein	0.655	0.097713949	26.48
A0A3P7DJ83	Transmembrane protein 258	0.42	0.000213686	6.25
J9B6T8	U4/U6 small nuclear ribonucleoprotein Prp31	0.516	0.005939834	3.51
A0A0I9NAH1	Bm12984	0.01	1×10^{-17}	0



Table 3 (Contd.)

Accession	Description	Abundance ratio: (treated)/(control)	Abundance ratio <i>P</i> -value: (treated)/(control)	Score sequent HT
A0A0J9Y4U4	BMA-EGL-30	0.569	0.021392617	2.43
A0A1I8EQI0	Uncharacterized protein	0.342	3.48676×10^{-6}	17.46
J9E5G9	Myosin_tail_1 domain-containing protein (Fragment)	0.56	0.017638063	58.44
J9FF86	ATP-dependent Clp protease proteolytic subunit	0.01	1×10^{-17}	11.74
A0A4E9FJB8	Phosphoglycerate mutase family protein	0.327	1.18996×10^{-6}	25.98
A0A0H5S5U2	BMA-CUL-3	0.01	1×10^{-17}	6.22
A0A1I8EF89	Phenylalanine-tRNA ligase	0.561	0.01791942	7.23
J9EZY7	Alanine-tRNA ligase	0.538	0.010388903	11.52
A0A0J9Y3N8	RNA helicase	0.01	1×10^{-17}	3.2
A0A0J9Y0Q8	BMA-HIP-1	0.607	0.045063353	73.05
A0A1I8ESJ2	PHB domain-containing protein	0.449	0.000683132	36.93
A0A1I8EPC5	Ge1_WD40 domain-containing protein	0.598	0.03795328	11.08
A0A4E9FGA6	Signal recognition particle receptor alpha subunit, putative	0.01	1×10^{-17}	2.03
A0A1I8EHS3	Uncharacterized protein	0.01	1×10^{-17}	7.6
A0A183XKF1	PIPK domain-containing protein	0.01	1×10^{-17}	2.11
A0A5S6PNY5	Spectrin beta chain	0.564	0.018973378	23.71
A0A4E9F4P0	Vacuolar ATP synthase subunit D, putative	0.674	0.128267858	2.06
A0A0H5S1U2	NADH dehydrogenase [ubiquinone] 1 beta subcomplex subunit 10	0.539	0.010693356	7.81
A0A1I9G4A9	BMA-ALH-11, isoform b (Fragment)	0.519	0.006464945	42
A0A0I9NA30	BMA-LSM-6	0.604	0.042599403	35.12
A0A4E9FA01	Glyceraldehyde-3-phosphate dehydrogenase	0.635	0.072353055	2650.13
A0A7I4NJV0	ATP-dependent (S)-NAD(P)H-hydrate dehydratase	0.651	0.092093112	15
A0A1I9GB23	Bm12220 (Fragment)	0.01	1×10^{-17}	6.67
A0A4E9F6R1	SHSP domain-containing protein	0.66	0.105509104	20.58
A0A7I4KLU1	BMA-RAL-1, isoform b	0.01	1×10^{-17}	6.77

filarial parasites and LF progression is not well characterized to date.

The major structural elements actin, myosin, and Troponin T were upregulated in the OSAgNPs treated filarial worms. Myosin and actin form the molecular machines for the contraction of sarcomeres and convert chemical energy to mechanical energy. It is quite possible that treatment with OSAgNPs resulted in extensive muscular contractions, ultimately leading to the rupture of the body wall of the parasites (Fig. 3) which could be visually seen after 4 h of treatment. The enzyme ribokinase catalyzing the conversion of ATP to ADP was upregulated, corroborating well with the downregulation of the Guanylate kinase and Adenylate kinase. The TCA cycle enzyme succinate-CoA ligase (ADP forming) was highly expressed too, confirming an increase in the production of ATP following OSAgNPs treatment. Furthermore, the glycolytic enzyme Phosphoglycerate mutase was also highly upregulated. The 2,3-diphosphoglycerate independent (iPGM) form of the enzyme has already been well studied in *O. volvulus*, *B. malayi* and *W. bancrofti* and implicated as a potential drug target.³²

3.5. Gene ontology and functional classification of DEPs

Gene Ontology annotation analysis of most significant DEPs based on molecular function, biological process, and cellular components is shown in Fig. 5. In terms of molecular function, major DEPs participated in ATP binding, kinase activity, metal

ion binding, nucleic acid binding, oxidoreductase activity, dimerization activity, and as structural constituents of chromatin. Major DEPs in the category of biological processes belonged to mRNA processing, biosynthetic process, catabolic process, tRNA aminoacylation, metabolic process, transport process, development process and stress response. Furthermore, the myosin complex, nucleus, plasma membrane, cytoplasm, mitochondria, spliceosome complex, nucleosome, and translation initiation factor were significantly enriched in the cellular components. Differential expression patterns of proteins revealed that treatment of filarial worms with OSAgNPs resulted in the downregulation of several proteins that were involved in energy metabolism, signal transduction, stress response, chaperone proteins as well as highly antigenic proteins.

3.6. PPI network analysis

The protein-protein interaction (PPI) analysis of differentially expressed proteins was predicted using the STRING tool. Out of the 69 upregulated proteins, 25 were interrelated to each other, whereas out of the 144 downregulated proteins, 64 were found to interact with each other (Fig. 6A and B). The PPI network of upregulated proteins revealed that troponin, actin, and tropomyosin were highly connected to each other and also with many other upregulated proteins. The downregulated proteins like signal recognition particles, HSP60, PIP kinase protein 2, Matrin-type domain containing protein exhibited the most number of



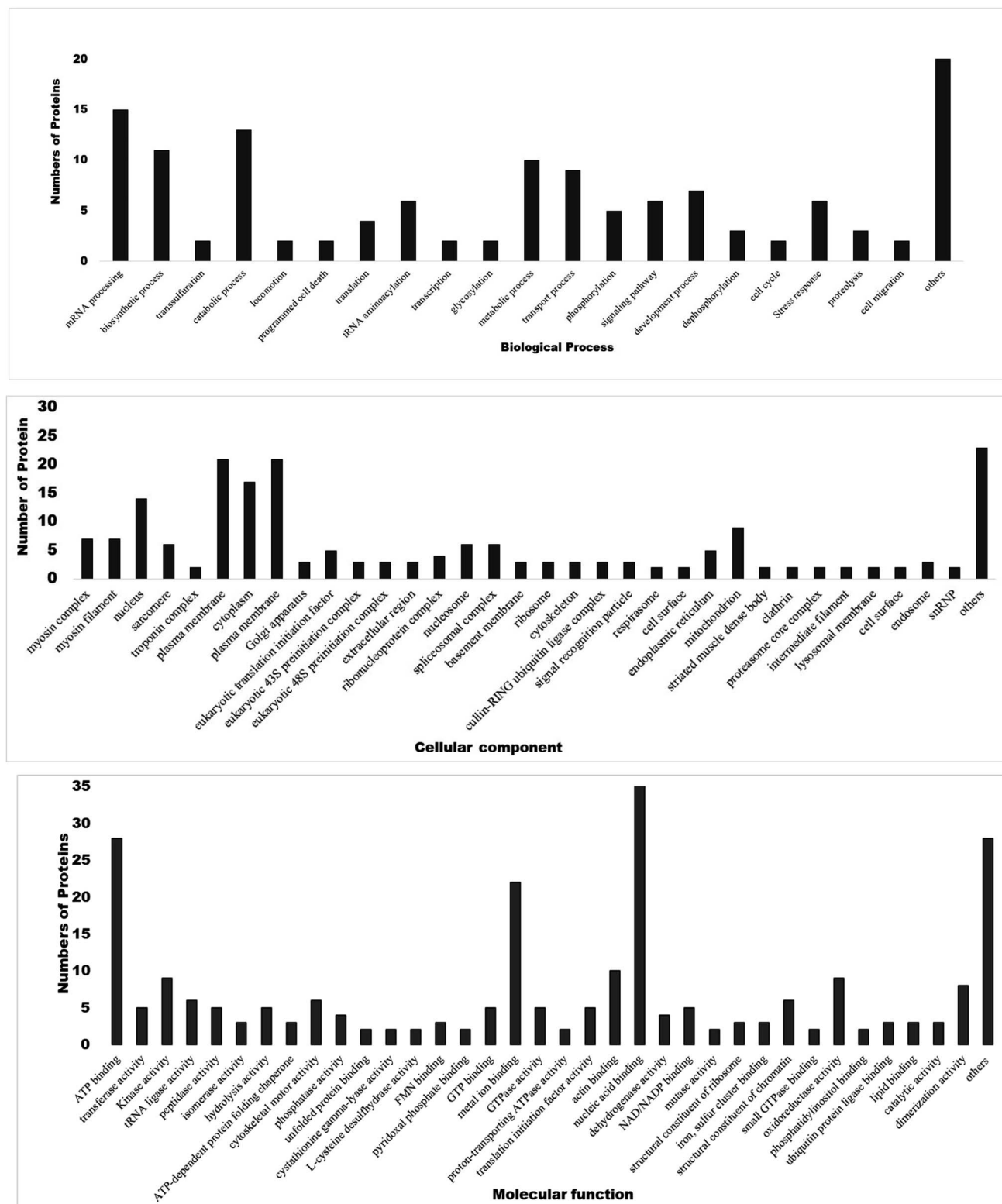


Fig. 5 Gene ontology annotation analysis of major DEPs classified into three main categories *i.e.*, molecular functions, biological processes and cellular components.

connections in the PPI network. The *in silico* analysis indicates that the highly connected differentially expressed proteins play crucial roles in filarial metabolism *via* different signalling

pathways and are also sensitive to anti-filarial molecules. Thus, it is anticipated that some of these key proteins may be developed and utilized as therapeutic markers in filarial treatments.



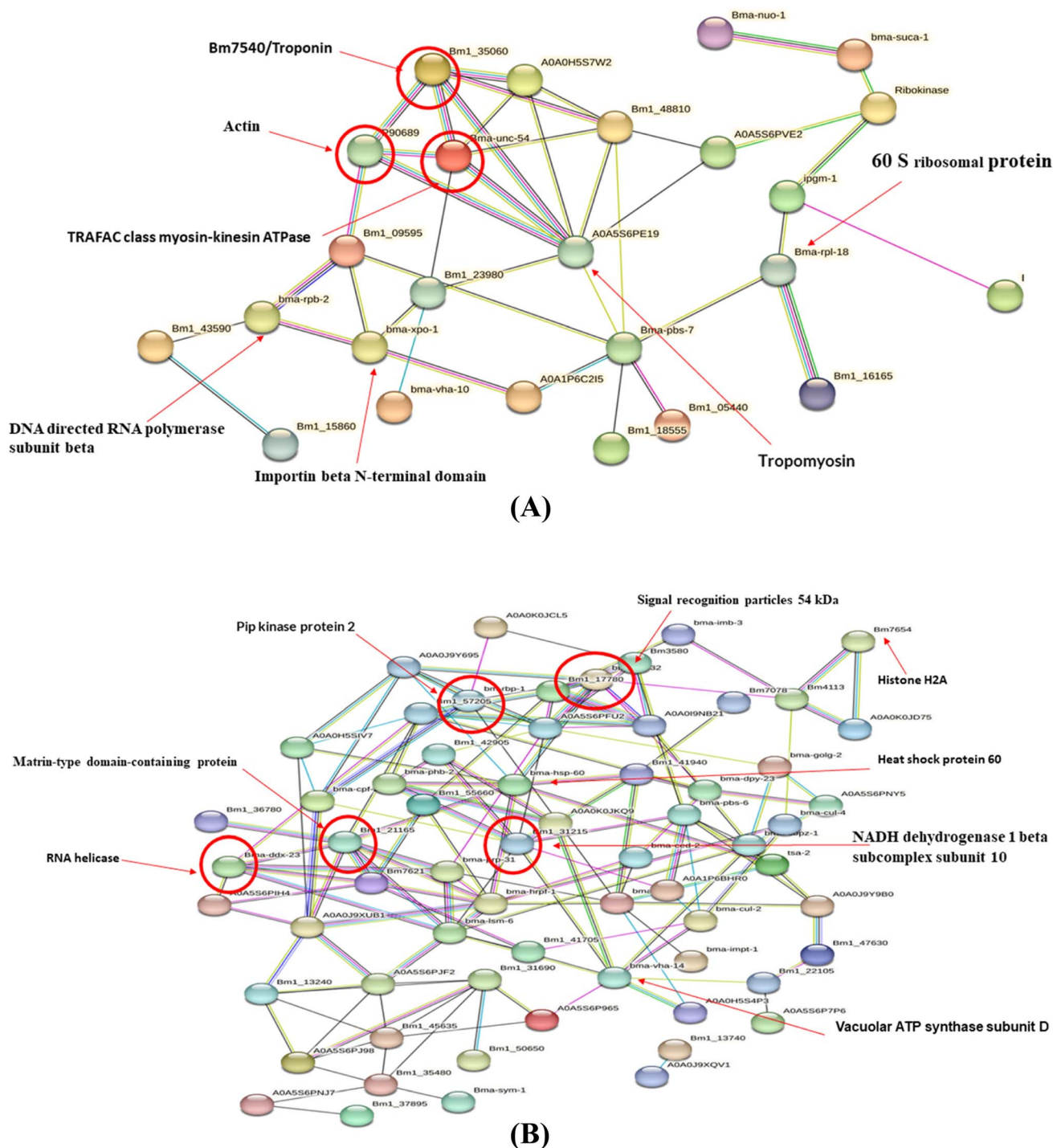


Fig. 6 Protein-protein interaction of major DEPs. (A) Upregulated proteins (B) downregulated proteins.

In summary, the objective of this work was to evaluate the adulticidal activity of naturally synthesized *Ocimum sanctum* silver nanoparticles and to develop an understanding of its molecular mechanism of action on the filarial parasites. Additionally, the study is opening up new avenues for future research on the effect of OSAGNPs on other filarial parasites, thus broadening the horizons for advanced research in filariasis treatment and management.

4. Conclusion

In this study, *Ocimum sanctum* silver nanoparticles were synthesized using an eco-friendly, rapid, easy and reproducible method. The green OSAGNPs had profound effect on the metabolism and survival of *S. cervi* filarial parasites and were also highly effective at very low concentrations. The HRMS proteomics results proved that the exposure of parasites to



OSAgNPs resulted in disruption of important signalling and metabolic pathways. To the best of our knowledge, this is the first report of OSAgNPs exhibiting anti-filarial effect against mature filarial parasites under *ex vivo* conditions. Undoubtedly, this study will open new avenues for designing and developing new nano-herbal anti-filarial drugs.

Ethical statement

Indian water buffaloes are slaughtered for food as part of non-vegetarian diets. The *Setaria cervi* worms were procured from the peritoneal lavage of freshly butchered buffaloes. Worms were purchased and brought to the laboratory in KRBMM for further experimental work.

Author contributions

Ayushi Mishra: conceptualization, methodology, data curation, formal analysis, investigation, writing – original draft, review & editing. Sunil Kumar: methodology, writing and editing. Anchal Singh: supervision, project administration, validation, writing – original draft, review & editing. All authors have read and agreed to the published version of the manuscript.

Conflicts of interest

The authors declare no conflict of interest.

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