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Carbonic anhydrase assisted acidogenic fermentation of forest residues for low carbon hydrogen and volatile fatty acid production: enhanced *in situ* CO₂ reduction and microbiological analysis

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Carbonic anhydrase (CA) is considered an efficient enzyme for fermentation systems exhibiting a wide range of applications, enhancing both the efficacy and output of the fermentation process. The present study aimed to evaluate the production of acidogenic biohydrogen (bioH₂) and volatile fatty acids (VFA) using forest residues as a renewable feedstock. Specifically, the study examined the integration of CA derived from *Desulfovibrio vulgaris* into the acidogenic fermentation (AF) process. The experimental procedure involved a cascade design conducted in two distinct phases. In phase I, the concentration of CA in the AF was systematically optimized, with glucose serving as the substrate. In phase II, three influential parameters (pH, pressurization with *in situ* generated gas and organic load) were evaluated on AF in association with optimized CA concentration from phase I. In phase II, glucose was replaced with renewable sugars obtained from forest residues after steam explosion pretreatment followed by enzymatic saccharification. The incorporation of CA in AF was found to be beneficial in steering acidogenic metabolites. Alkaline conditions (pH 8) promoted bioH₂, yielding 210.9 mLH₂ gCOD⁻¹, while introducing CA further increased output to 266.6 mLH₂ gCOD⁻¹. This enzymatic intervention improved the production of bioH₂ conversion efficiency (HCE) from 45.3% to 57.2%. Pressurizing the system accelerated VFA production with complete utilization of *in situ* produced H₂ + CO₂ compared to non-pressurized systems. Particularly, caproic acid production was improved under pressurized conditions which was accomplished by the targeted enrichment of chain-elongating bacteria in the mixed culture. The microbial diversity analysis showed the dominance of *Firmicutes* suggesting a significant degree of adaptation to the experimental contexts, leading to an enhanced production of acidogenic metabolites.

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1. Introduction

Currently, there has been an increasing emphasis on addressing climate change and ensuring energy security, thus the need for renewable sources to contribute to the world's increased energy supply. Among the various renewable sources, forest waste/residues have drawn attention to the rising worldwide demand for energy and the need to cut greenhouse gas (GHG) emissions.¹ Forest residues are considered renewable, unlike fossil fuels, which deplete long-term carbon

storage; second, forest biomass recycles carbon, rather than releasing it into the atmosphere.

Due to its geographical positioning and climatic conditions, Sweden exhibits favorable characteristics for a growing number of forests, which encompass around 60% of the nation's land area. The yearly growth of the forest amounts to around 100 MCM (million cubic meter) of standing volume, and the amount of timber harvested is between 50–80 MCM of standing volume annually during the last decades.^{2,3} The process of forest cutting results in the production of a significant quantity of byproducts referred to as residues, which are typically regarded as waste materials.⁴ Logging residues may constitute a substantial environmental concern owing to their potential to undergo decomposition and subsequently emit methane, a potent greenhouse gas. In addition, they possess the potential to provide a fire risk and may serve as a magnet for various forms of pests and illnesses.²

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Forest residues are considered a valuable resource due to their inherent advantages and characteristics demonstrating the capacity to produce a wide array of products including bio-fuels, renewable chemicals and other value-added.^{5,6} While energy production from forest logging leftovers is an efficient method for mitigating environmental impact.^{7,8} It is pertinent to highlight that the utilization of forest biomass residues, possesses the capacity to function as a feasible and environmentally sustainable energy resource for the country.⁷ Consequently, several governmental initiatives have been implemented in Sweden with the aim of fostering the use of forest residuals. Furthermore, several regional and municipal policies have been implemented to encourage the use of residual forest biomass.^{3,9,10}

Hydrogen (H₂), being a clean-burning fuel, holds immense potential as an energy carrier in the pursuit of industrial decarbonization including steelmaking and cement production.

Acidogenic fermentation (AF), an intermediate stage of anaerobic digestion (AD) and is a highly versatile platform that facilitates the conversion of organic waste/wastewater into a diverse range of value-added products, such as biohydrogen and volatile fatty acids (VFAs).¹¹ AF process holds immense significance in the realm of bioenergy and bio-based chemicals production, as it facilitates the retrieval of valuable resources from waste streams, thereby aiding in waste minimization and fostering environmental sustainability.^{12,13} AF is a complex interaction of microorganisms that facilitates the breakdown of organic matter in anaerobic conditions, thereby producing hydrogen gas along with volatile fatty acids (VFAs).^{13,14} VFAs are a class of organic acids that exhibit a diverse array of industrial applications. Acetic, propionic acid and butyric acid are the major VFAs produced.¹⁵ Harnessing the potential of wood biomass/residues as a feedstock for AF holds immense potential in advancing sustainability and driving the growth of a circular bioeconomy.

The complex structural of wood biomass makes it difficult to use as a substrate for microbes to access and degrade the sugars contained in the cellulose and hemicellulose. Besides its main role, lignin provides an important barrier to microbial decomposition. To render wood biomass amenable to microbial activities, pretreatment is necessary to facilitate the decomposition of its intricate structure and enhance the accessibility of sugars to microbes. Steam explosion (SE) is a physicochemical pretreatment method that is a highly effective for lignocellulosic biomass pretreatment.^{16,17}

The generation of CO₂ is an integral part of the acidogenic/acetogenic/methanogenic process. Metabolic pathways for various products include decarboxylation steps, leading to decreased product yield and quick release of CO₂ into the environment. The acidogenic process results in the production of 2 moles of CO₂ for every two moles of acetate and butyrate produced through the corresponding pathways.^{11,18} Similarly, over 40% of carbon is emitted as CO₂ during bio-methanation process (CH₃COOH → CH₄ + CO₂). Employing carbonic anhydrase in the fermentation system for *in situ* CO₂ capture offers a novel opportunity to transform it into renewable chemicals

and low carbon fuels along with reduction of CO₂ emissions into the environment.^{19,20} Moreover, utilizing biogenic CO₂ is favored to guarantee that the products stay carbon neutral and help in achieving climate neutrality. For instance, Europe produces 69.7 Mt of CO₂ annually from the upgrading and combustion of biogas, bioethanol, and other chemicals produced through fermentation processes.²¹

CO₂ capture has proven to be significant in enhancing the efficacy of various fermentation procedures, including ethanol, lactic, succinic acid, and biogas production.^{22,23} For example, the CO₂ generated during the ethanol fermentation process was effectively captured using a packed column filled with NaOH, KOH, or NH₄OH, facilitating the absorption of CO₂. The resulted carbonate solution serves a dual purpose: firstly, it aids in maintaining the desired pH levels, and secondly, it acts as a valuable source of CO₂ for a succinic acid fermentation process.²² CAs, which are ubiquitous enzymes are known to be highly prevalent among bacteria, archaea, and eukaryotes, wherein they are postulated to fulfill a multitude of roles.^{24,25} CA serve as catalysts for the reversible hydration reaction of carbon dioxide (CO₂) into bicarbonate, accelerating the limiting step of CO₂ solubilization in aqueous media.^{26,27} As such, CA has the potential to mitigate CO₂ levels in bioconversion processes resulting in elevated levels of CO₂ uptake by the microbes or enzymes and providing a viable carbon supply for the production of high-added value products, such as bio-fuels and chemicals.^{28–31} In a study pertaining to ethanol fermentation, the use of CA *in situ* during fermentation was found to enhance the production of ethanol by 20%.²⁸

In the present study, we evaluated the effect of supplementation of CA derived from *Desulfovibrio vulgaris* to the acidogenic fermentation process towards biohydrogen and volatile fatty acids production from forest residue hydrolysates derived through stem explosion pretreatment. Specifically, this study provides an insight into the intricate mechanisms of AF towards biohydrogen and volatile fatty acids in the presence of CA, investigating its potential role in regulating the pH levels, while concurrently utilizing the acidogenic CO₂ within the reactor supporting its mitigation. Furthermore, microbial diversity study was conducted to provide and insight into the alterations occurring during AF of FR derived sugars with respect to experimental variations implemented.

2. Materials and methods

2.1. Acidogenic mixed consortia and substrate

The inoculum used in this study is obtained from a biogas production plant located in Luleå, Sweden. Prior to being used as inoculum, the sludge underwent filtration through nylon mesh to remove stones and other solid objects. Filtered sludge was given heat shock pretreatment at 80–90 °C for 1 hour. The pretreated sludge was then enriched using a nutrient media consisting of glucose: 3 g_{COD} L⁻¹, CaCl₂: 5 mg L⁻¹, CoCl₂: 25 mg L⁻¹, CuCl₂: 10.5 mg L⁻¹, FeCl₃: 25 mg L⁻¹, K₂HPO₄:



0.25 g L⁻¹, KH₂PO₄: 0.25 g L⁻¹, MgCl₂: 0.3 g L⁻¹, MnCl₂: 15 mg L⁻¹, NH₄Cl: 0.5 g L⁻¹, and ZnCl₂: 11.5 mg L⁻¹.

2.2. Enzyme production

Enzyme expression was done according to Sjöblom *et al.* (2020).³² The nucleotide sequence of the thermostable CA variant, DvCA8.0, derived from *Desulfovibrio vulgaris* and combined with a polyhistidine tag (6xHis-tag), was precisely integrated within the NdeI and XhoI restriction sites of the pET22b(+) vector.³³ The gene synthesis and cloning were executed by GenScript, (Piscataway, New Jersey, USA). The vector underwent a subsequent transformation process, resulting in its integration into *Escherichia coli* BL21(DE3). Following this transformation, stock cultures were diligently preserved for future use. To synthesize CA, a volume of 500 µL of a cultured population of genetically modified *E. coli* BL21 (DE3) cells was introduced into a 50 mL solution of Luria-Bertani preculture medium, supplemented with 100 µg mL⁻¹ of the antibiotic ampicillin. This mixture was then subjected to an overnight incubation period at a temperature of 37 °C, with continuous agitation at a rate of 180 rpm. A 1% v/v pre-culture was introduced into an auto-inducing lactose medium (specifically ZYP-5052, devoid of trace elements) supplemented with 100 µg mL⁻¹ of ampicillin. After 24 h of cultivation at 37 °C and 180 rpm, the cultures were harvested. Since no CA activity was observed, the harvested culture was centrifuged at 8000 rpm for 5 minutes, and the obtained supernatant was discarded. The 1 L culture cells were resuspended in 200 mL in a 1 : 1 v/v solution of Tris-HCl 0.1 M pH 8.0 and NaOH 0.2 M. After that, they underwent three cycles of lysis at 700 bar pressure in a laboratory homogenizer (SPX, Crawley, UK). Using a pressurized filtration system (Sterlitech, Kent, WA, USA), the cell debris was removed by centrifugation at 8000 rpm for 5 minutes. The recovered supernatant, which is the entire cell lysate, was filtered to a particle size of 0.2 µm. Lastly, a tangential flow filtering system (Millipore, Burlington, MA, USA; MWCO 10 kDa) was used to concentrate the lysate 30 times. CA expression was confirmed by using a *E. coli* culture having a non-cloned pET22b(+) vector as negative control. The CA activity was determined by the Wilbur-Anderson assay. The activity of the lysate was determined as an equivalent of 143 WAU per µL CA.

2.3. Pretreatment of forest residues

Forestry residues (FR; residues gathered from tree thinning or logging, such as tops and branches) was collected from

Swedish forest. The untreated FR composition is presented in Table 1. Steam explosion (SE) pretreatment of FR was performed evaluating the influence of sulphur dioxide (SO₂) and sulfuric acid (H₂SO₄). For treatments employing H₂SO₄, residence time, reaction temperature, and acid quantity were optimized aiming towards maximizing the sugars release and minimizing the inhibitors generated. In detail, different concentrations of H₂SO₄ (1 to 5% w/w), varying temperatures (170 to 215 °C), and different pretreatment times (2 to 15 minutes) were examined. The most promising results were obtained with 1% H₂SO₄ at either 195 °C for 5 minutes or at 215 °C for 10 minutes. Treatment with SO₂ took place at 205 °C, (~18 bar) and 7 min of reaction time. Table 1 displays the composition of the solid fraction resulting from the SE pretreatment of FR (the liquid portion was not used in this study) (Table 1). The solid materials obtained from the SO₂ pretreatment method were used for acidogenic fermentation.

2.4. Acidogenesis: feedstock and fermentation

The feedstock was prepared in two steps. In step I, the solid fraction obtained from the steam explosion process was separated using filter paper (liquid portion was not used in this study). In step II, the separated solid fraction was employed for enzymatic hydrolysis. Enzymatic hydrolysis was carried out using cellulase, enzyme blend (Cellic® CTec2, Sigma-Aldrich; 10 FPU per g solid biomass) in sodium citrate buffer at 50 °C for 30 h.³⁴ Slurry obtained from enzymatic hydrolysis was further centrifuged at 2000 rpm to separate leftover solids particles, the liquid portion containing reducing sugars (composed of glucose, xylose, arabinose, galactose, and mannose) was evaluated as acidogenic fermentation substrate/medium for biohydrogen and volatile fatty acids in serum bottles (200 mL) as reactors. The composition of reducing sugar is provided in section 3.1. The pretreated filtered sludge was used as inoculum with an inoculum-to-substrate ratio of 1 for VS. The study was conducted in two phases (Fig. 1). During phase I, the concentration of CA in the acidogenic fermentation was optimized. Three different concentrations of enzyme were added: 15 µL mL_{feed}⁻¹ (R1), 7.5 µL mL_{feed}⁻¹ (R2), and 3.76 µL mL_{feed}⁻¹ (R3). Furthermore, R4 (only substrate without CA) and R5 (without substrate and CA) were used as control experiments. The substrate used phase I was glucose at an organic load of 12 g_{COD} L⁻¹. In phase II, experiments were conducted in a cascade where initially the best enzyme concentration (from phase I) was selected for its evaluation on acidogenic fermentation using sugars hydrolysate as fermentation

Table 1 Chemical composition of the solids derived from steam explosion of forest residues at varied digestion time and temperature

	Treatment temperature	Hemicellulose (g kg ⁻¹ of dry material)	Cellulose (g kg ⁻¹ of dry material)	Lignin (Klason; g kg ⁻¹ of dry material)	Yield (% w/w)
H ₂ SO ₄	195 °C	35.55	285.3	567	44.9
	215 °C	7.3	249.3	666	43.8
SO ₂	205 °C	23.1	279.9	585	33.4
Untreated forest residue	—	145.1	224.1	384	—



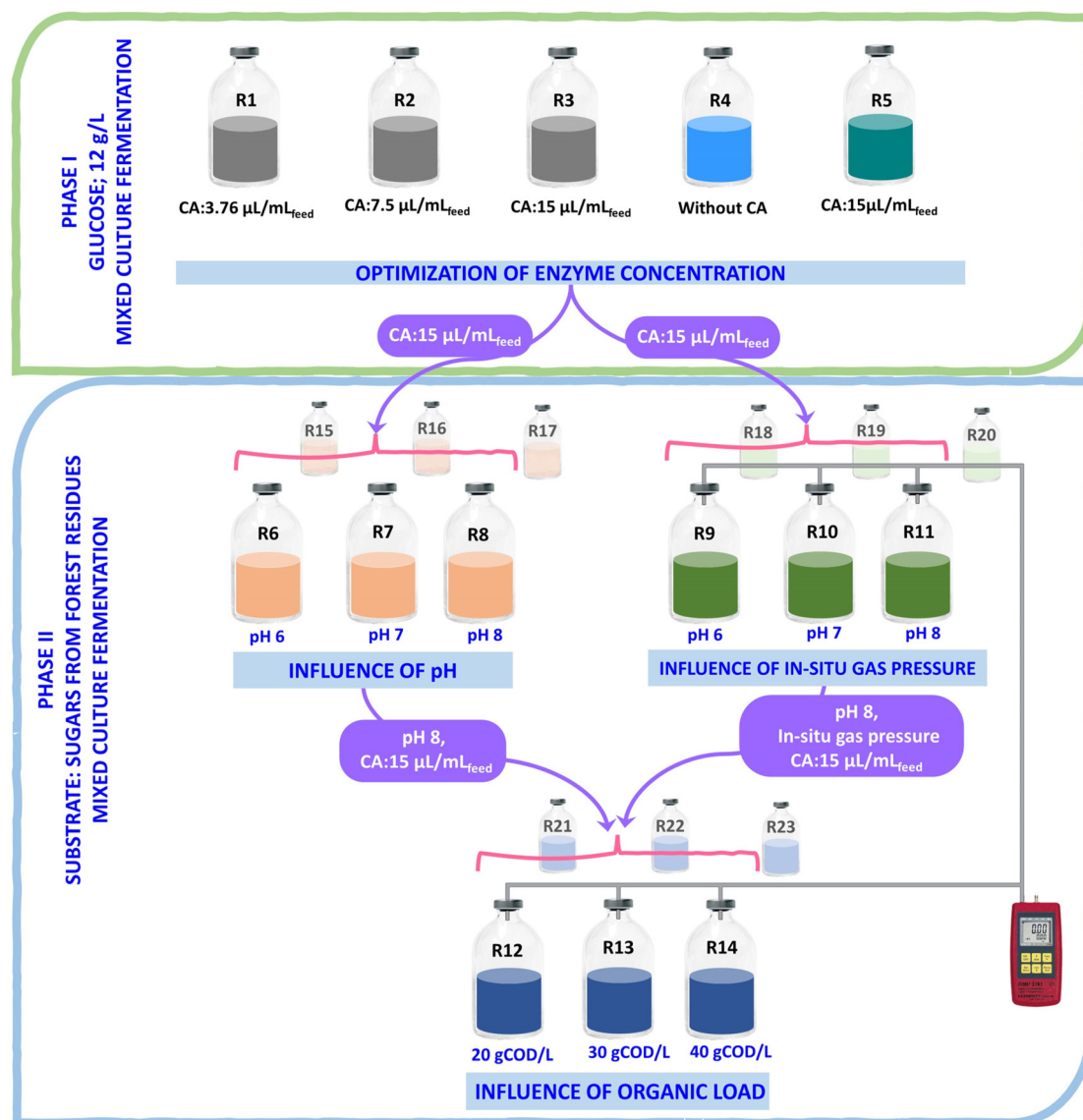


Fig. 1 Schematic presentation of designed experiment to evaluate carbonic anhydrase enzyme addition in acidogenic fermentation evaluating various parameters that influence the process. Reactors 15–23, served as the control configuration for the R6–R14 reactors, as they were operated without carbonic anhydrase enzyme in phase II.

substrate (derived from enzymatic hydrolysis of solids derived from steam explosion of forest residues). Here in phase II, in presence of CA, initially the impact of different pH (R6: pH 6, R7: pH 7 and R8: pH 8) and the influence of *in situ* headspace pressure (*in situ* generated $\text{H}_2 + \text{CO}_2$ from organics fermentation) at pH 6 (R9), pH 7 (R10) and pH 8 (R11) were studied. The optimal identified conditions (best pH + *in situ* pressure/non-pressurization) were then applied at higher organic load (R12: 20 gCOD L^{-1} , R13: 30 gCOD L^{-1} and R14: 40 gCOD L^{-1}). Reactors (R6–14) were operated with CA were compared with its respective control that didn't contain enzyme (R15–R23). In both the phases, nutrient solution (composed of nutrients as indicated in section 2.1 expect glucose) was added along with fermentation media.

2.5. Analytical methods

The change in pH during fermentation was monitored using a pH meter (pHnomenal-pH1100L; VWR, Stockholm, Sweden). VFAs in the fermentation medium were analyzed on a high-performance liquid chromatography system (PerkinElmer, Waltham, MA, USA) equipped with a pump (410 LC), a refractive index detector (RID-6A), and an Aminex HPX-87H column (300 $\times$ 7.8 mm; Bio-Rad, Hercules, CA, USA). The column was maintained at 65 $^{\circ}\text{C}$. The mobile phase was 5 mM H_2SO_4 eluted at a flow rate of 0.6 mL min^{-1} . VFAs were quantified using calibration curves derived from commercially available standards (10 mM, VFA mix; Sigma-Aldrich). The headspace pressure was measured using Greisinger GMH 3111 pressure

gauge equipped with Greisinger GMSD 10 BR Relative pressure gauge GMSD 10 BR. The chemical composition of the biogas was assessed on an Agilent 990 micro-GC instrument, equipped with a COX column (COX UM1MX0.8MMID BF, CP-PORABOND Q, 1MX 0.25 mm $\times$ 3 μ m; Agilent Technologies, Santa Clara, CA, USA). The column was maintained at 110 °C and the injector at 80 °C. Hydrogen energy was calculated considering its LHV value (120 MJ kg⁻¹). An ANOVA was conducted to evaluate the differences between pairs of means for the CA enzyme addition on fermentation process (Table 2). The statistical analysis (Tukey's test) was performed with OriginPro 2021 (Academic edition, version 9.8.0.200).

2.6. DNA extraction

Upon completion of the experiment, on day 3, 1–2 g of biomass was collected from the reactor and preserved at –20 °C. The biomass sample was rinsed with phosphate-buffered saline solution and then subjected to centrifugation at a speed of 8000 rpm for 15 minutes. The DNA was isolated using the FastDNA spin kit for soil (MP Biomedicals, Irvine, CA, USA) according to the manufacturer's protocol. Subsequently, the DNA concentration was determined using the Qubit dsDNA HS/BR assay kit (Thermo Fisher Scientific, Waltham, MA, USA).

2.7. Library preparation, DNA sequencing, and bioinformatics processing

Amplicon libraries were prepared according to a standard protocol to target the variable region 4 (abV4C) of the 16S rRNA gene in bacteria and archaea.³⁵ In summary, a total of 10 nanograms of extracted DNA was used to perform polymerase chain reaction (PCR) amplification of the abV4C amplicons in both bacterial and archaeal samples. Each PCR reaction had a volume of 25 μ L, consisting of 12.5 μ L of PCR BIO Ultra mix and 400 nM concentrations of both forward and reverse primers. The forward primer [515FB] 5'-GTGYCAGCMGCCGCGGTAA-3' and reverse primer [806RB] 5'-GGACTACNVGGGTWTCTAAT-3' were developed according to the instructions provided in the Illumina Technical Manual.³⁵ Primer tails facilitate the linkage of Illumina Nextera adaptors for sequencing in a following PCR. The amplicon libraries were purified following the standard CleanNGS SPRI beads

protocol (CleanNA, NL) with a bead to sample ratio of 4:5. The DNA was eluted in 25 μ L of nuclease free water from Qiagen in Germany. The DNA concentration was determined using the Qubit dsDNA HS Assay kit (Thermo Fisher Scientific, USA). Conducting gel electrophoresis with the TapeStation 2200 and D1000/High sensitivity D1000 screentapes (Agilent, USA) was essential in verifying the size and purity of a portion of the amplicon libraries. A detailed overview of the methods employed in DNA sequencing, library preparation, and bioinformatics processing has been previously described.³⁶

3. Results and discussion

3.1. Pretreatment of forest residues

Two different catalysts, i.e. H₂SO₄ and SO₂, were tested for the pretreatment of forest residuals (FR) which contained mainly tops and branches from the trees (mixture of softwood and hardwood). SO₂ pretreated solids were used for the subsequent biogas experiments. As shown in Table 1, the SE of forest residues with SO₂ at 205 °C resulted to production of pretreated solids containing 279.9 g kg_{solid}⁻¹ cellulose, 23.1 g kg_{solid}⁻¹ hemicellulose and 585 g kg_{solid}⁻¹ lignin. Further, the steam exploded FR (solid fraction) was subjected to enzyme hydrolysis (10 FPU per g_{solids}). Enzyme hydrolysis resulted in reducing sugars (53.8 g L⁻¹) composed of glucose (48.3 g L⁻¹) followed by xylose (3.1 g L⁻¹), arabinose (0.6 g L⁻¹), galactose (0.4 g L⁻¹) and mannose (1.4 g L⁻¹). This reducing sugar was employed as a renewable substrate in the second phase of the study.

3.2. Biogas production

The experiment consisted of two distinct phases. During phase I, glucose was used as the substrate for the acidogenic fermentation (AF) to determine the optimal concentration of CA. Fig. 2 depicts biogas output with respect to varied concentration of CA (R1: 3.76 μ L mL_{feed}⁻¹, R2: 7.5 μ L mL_{feed}⁻¹, R3: 15 μ L mL_{feed}⁻¹) at initial pH 7. The biogas production was found to be the highest in R3, with a cumulative gas volume of 430 mL g_{COD}⁻¹ followed by R4 (410 mL g_{COD}⁻¹; no CA addition), R2 (390 mL g_{COD}⁻¹) and R1 (385 mL g_{COD}⁻¹). Interestingly, the reactor (R5) that was operated without substrate but supplemented with enzyme (CA: 15 μ L mL_{feed}⁻¹)

Table 2 ANOVA (Tukey test) table showing the effect of carbonic anhydrase addition during fermentation of sugars

Comparison between reactors operated with varied concentration of CA enzyme	Mean difference	<i>q</i> value	Probability	Significance ^{a,b,c}
R2–R1	44.41	56.41	0	1
R3–R1	104.31	132.5	0	1
R3–R2	59.9	76.08	0	1
R4–R1	39.28	49.89	0	1
R4–R2	–5.13	6.52	0.007	1
R4–R3	–65.03	82.6	0	1

^a Significance value equal 1 indicates the difference of the mean is significant at $\alpha = 0.05$. ^b Significance value equal 0 indicates the difference of the mean is not significant at $\alpha = 0.05$. ^c Significance value equal –1 indicates the difference of the mean is not tested.



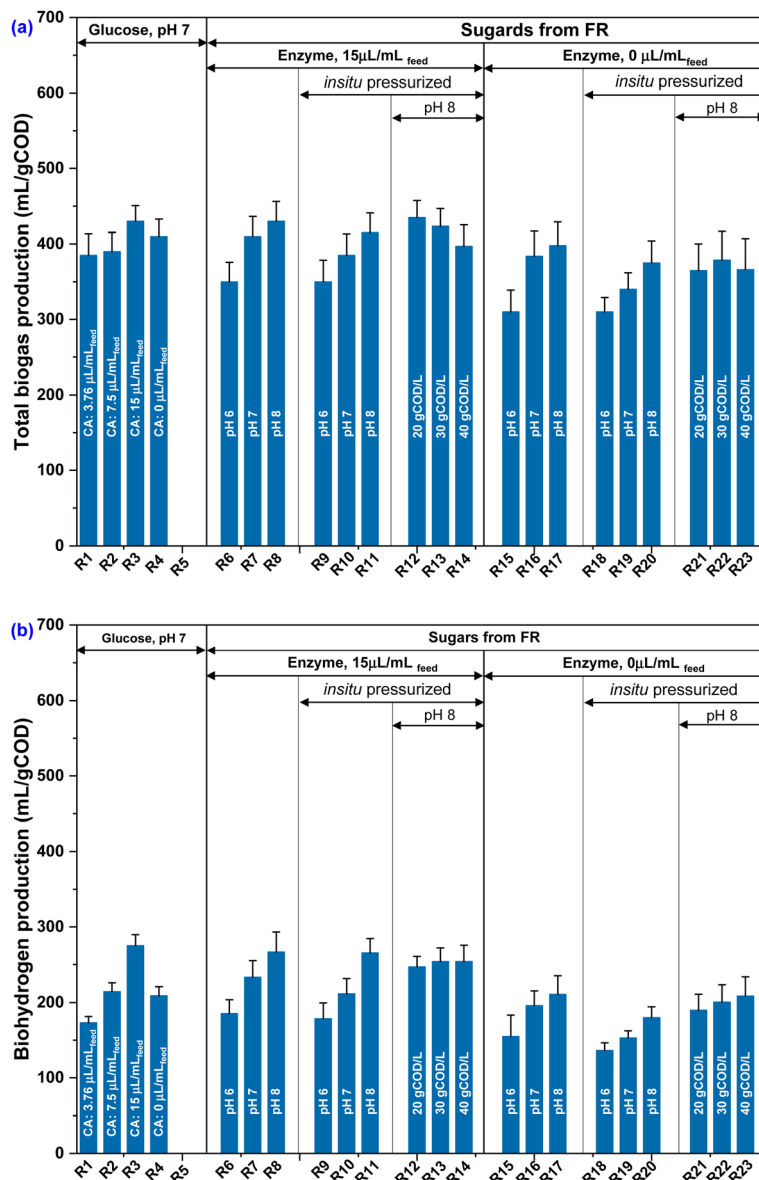


Fig. 2 (a) Biogas production and (b) biohydrogen production recorded during fermentation of glucose and sugars from forest residuals under various experimental conditions.

failed to demonstrate biogas production, thereby suggesting that the enzyme introduced did not serve as a substrate for the mixed culture fermentation. The optimization process was examined using one-way ANOVA. The analysis of variance (ANOVA) showed that the effects of CA on the fermentation process are statistically significant, indicating that the mean difference between the selected CA concentrations was significantly greater with R3 (Table 2). Phase II of the AF experiments included substituting glucose with renewable sugars derived from forest residuals, and the optimal concentration of CA ($15 \text{ L mL}_{\text{feed}}^{-1}$) was selected based on its excellent acidogenic performance towards increased production of gases observed in phase I.

In phase II, during evaluating effect of three different pH, the highest biogas production was notice at pH 8 (R8:

$430 \text{ mL g}_{\text{COD}}^{-1}$) compared to pH 7 (R7: $410 \text{ mL g}_{\text{COD}}^{-1}$) and pH 6 (R6: $350 \text{ mL g}_{\text{COD}}^{-1}$). These findings highlight the importance of alkaline conditions and the efficiency of pre-treatment method adopted for lignocellulosic biomass in delivering sugars for fermentation. The fermentation micro-environment, specifically in an alkaline redox microenvironment, exhibited greater potential for enhanced biohydrogen production when compared to its corresponding neutral and acidic conditions. In contrast to an acidic pH, an alkaline redox microenvironment elicits an enhanced hydrolytic effect on carbohydrates and proteins. This is achieved through the ionization of charged groups, such as carboxylic groups, thereby promoting the accessibility of soluble and readily assimilable substrates for fermentation.^{18,37} Typically,

this phenomenon occurs during the initial hour of the fermentation process. In the context of an elevated alkaline pH, it is noteworthy that a significant proportion of carbohydrates exhibit the capacity for anaerobic biodegradation. Furthermore, alkaline redox conditions serve to impede the proliferation of hydrogenoclastic methanogens, thereby preserving the molecular hydrogen (H_2) in its original form and preventing its conversion into terminal methanogenic byproducts, such as CH_4 . Presence of carbonic anhydrase enzyme along with alkaline pH conditions significantly contributes to the buffering of these biological systems.^{18,37} In contrast, reactors R15 (310 mL g_{COD}^{-1}), R16 (384 mL g_{COD}^{-1}), and R17 (398 mL g_{COD}^{-1}) operated without CA as control setup for R6, R7 and R8 exhibited a reduced biogas production. In addition, headspace *in situ* pressurization (4.8–5.1 bar) did not improve biogas output, as this was declined to 415 mL g_{COD}^{-1} (R11; pH 8), 385 mL g_{COD}^{-1} (R10; pH 7) and 350 mL g_{COD}^{-1} (R9; pH 6). Absence of CA for *in situ* pressurized reactors (R9–11) considerably lowered the biogas values to 310 mL g_{COD}^{-1} (R18; pH 6), 340 mL g_{COD}^{-1} (R19; pH 7) and 375 mL g_{COD}^{-1} (R20; pH 8). The pressure within the headspace has the potential to affect the partial pressure of H_2 in the system.^{38,39} The concept of partial pressure refers to the pressure exerted by an individual gas within a mixture of gases.⁴⁰ Within a reactor, gases produced by microbes including H_2 during fermentation processes. The partial pressure of H_2 is a significant determinant influencing the microbial metabolites and its profile through various biochemical reactions. Under pressurized condition the transfer of H^+ ions and electrons play a crucial role in facilitating the normal progression of the reactions. Furthermore, the decrease in the partial pressure of H_2 leads to a decrease in the Gibbs free energy of reactions involving H_2 as a product.^{40,41} This change in energy favors the production of H_2 , particularly in reactions like homoacetogenesis. However, it is important to note that this reduction in partial pressure can be detrimental to hydrogen consumption when it is lowered.^{39,40,42} Pressurized system may result in the saturation of CO_2 in the medium of the reactors, potentially impacting pH levels and causing changes to microbial activity leading to a shift in the metabolic pathway.⁴³ Nevertheless, the presence of carbonic anhydrase in the reactor can effectively mitigate this issue by utilizing *in situ* accumulated CO_2 within the reactor. This suggest regulation of headspace pressure that could be one of the strategies to control the metabolic pathways towards the target product.^{20,39,44} Combining best pH (8), and pressurized condition along with CA enzyme (15 μL mL $_{feed}^{-1}$), the influence of organic load (OL) was examined which showed an increase in biogas production up to 30 $g_{COD} L^{-1}$, slightly declined at 50 $g_{COD} L^{-1}$. Through the implementation of CA enzyme-enhanced fermentation, the greater biogas production occurred at a concentration of 20 $g_{COD} L^{-1}$ (R12: 435 mL g_{COD}^{-1}), and 30 $g_{COD} L^{-1}$ (R13: 423 mL g_{COD}^{-1}) and declined at 40 $g_{COD} L^{-1}$ (R14: 397 mL g_{COD}^{-1}). Conversely, in the absence of CA enzyme, the biogas production was notably

lower even under identical experimental conditions, with volumes of 365 mL g_{COD}^{-1} (R21: 20 $g_{COD} L^{-1}$), 378 mL g_{COD}^{-1} (R22: 30 $g_{COD} L^{-1}$) and 366 mL g_{COD}^{-1} (R23: 40 $g_{COD} L^{-1}$). This marked difference in biogas production clearly demonstrates the significant role played by CA enzyme during the fermentation process.

3.2.1. Biogas composition. The composition of the acidogenic biogas primarily consisted of hydrogen (H_2) and carbon dioxide (CO_2). The biogas composition exhibits variability, dependent upon the specific feedstock and fermentation process employed. However, it is commonly observed that H_2 predominates as the principal constituent, typically constituting a significant proportion of 30–60% of the overall gas volume. During phase I of the experiment, which focused on optimizing enzyme load, it was noted that the introduction of CA had a significant impact on bio H_2 generation. Specifically, the reactor with a highest concentration of the CA enzyme exhibited higher production, delivering 258 mL $_{H_2}$ g_{COD}^{-1} (R3), compared to reactors with lower enzyme loads (R1 and R2) and without enzyme (R4). During phase II of the study, the impact of various pH levels on CA-assisted fermentation of sugars from FR was evaluated. It was observed that the highest bio H_2 production occurred at pH 8 (R8), as opposed to pH 7 (R7) and pH 6 (R6). This trend was consistent even under pressurized conditions, where a greater amount of bio H_2 production was observed at pH 8 (R11) compared to pH 7 (R10) and pH 6 (R9) operations, although *in situ* pressurization did not significantly improved bio H_2 . Continuing with the optimized CA enzyme concentration (15 μL mL $_{feed}^{-1}$) along with pH (8) + pressurized condition, we proceeded to assess these parameters across three distinct organic loads (OL: 20, 30 and 40 $g_{COD} L^{-1}$). An initial enhancement in the production of bio H_2 was observed as the OL was gradually increased while above 30 $g L^{-1}$ COD it remained stable. On the contrary, these configurations were subjected to comparative analysis of fermentation in the absence of CA enzyme setups (R21–R23), revealing a lower bio H_2 yield in each reactor as compared to their respective enzyme-loaded reactors (R12–R14). For example, during pH variations study, the reactors with CA exhibited a significant increase in bio H_2 production compared to the reactors without CA. Here the bio H_2 production was found to be 1.26 (pH 8), 1.2 (pH 6) and 1.19 (pH 7) times higher in the CA supplemented reactors. In a similar manner, the production of bio H_2 showed an increase of around 1.31–1.48 times when using a pressured system in conjunction with CA. Additionally, during studying the effect of OL, the bio H_2 production demonstrated a 1.22–1.30 times higher yield when CA was used. Particularly the volumetric bio H_2 significantly increased with R12 (246.8 mL $_{H_2}$ g_{COD}^{-1}), R13 (254 mL $_{H_2}$ g_{COD}^{-1}), R14 (254.08 mL $_{H_2}$ g_{COD}^{-1}) with an increase in organic load to 20 $g L^{-1}$ COD, 30 $g L^{-1}$ COD and 40 $g L^{-1}$ COD respectively. However, the yield remained low compared to the operation at 12 $g_{COD} L^{-1}$ (265.6 mL $_{H_2}$ g_{COD}^{-1}) despite of possessing symmetrical configurations as with R11 (CA + pH 8 + pressurization). This lower yield might be due to substrate load. Substrate load may affect the stability and yield of bio H_2 gene-



ration process. Under such conditions microbes can redirect their metabolic energy away from bioH₂ synthesis and instead focus on other activities, such as biomass growth and cell maintenance. This could result to a decrease in the total hydrogen output and cause instability of the process.^{45,46} These results indicate the significant role of CA enzyme addition in the reactor during the acidogenic fermentation process. Additionally, an enhanced bioH₂ production seen during the optimization (phase I) may be ascribed to the use of a simpler substrate in comparison to lignocellulosic based sugars (in phase II). Whereas lignocellulosic based sugars are a complex blend of various sugars, organic acids, and other molecules that have the potential to inhibit the proliferation of H₂ producing microflora, hence impacting the overall acidogenic process.

The documented bioH₂ production in phase II is impressive and can be compared with glucose fermentation in phase I, emphasizing the effective adaptation of mixed culture to the lignocellulosic hydrolysate under implemented experimental conditions. Table 3 presents the bioH₂ and other acidogenic metabolites derived from various lignocellulosic based feedstocks (Table 3). Using lignocellulosic feedstocks to produce bioH₂ without costly and energy-intensive purification and neutralization appears promising. The addition of CA was beneficial in promoting the acidogenic process. Enhanced bioH₂ production with CA addition can be ascribed to its ability to regulate pH and improve redox equilibrium. Acidogenic fermentation is often associated with the redox equilibrium within the microbial population, and CA was found to potentially regulate the pH and CO₂ levels. No significant increase in bioH₂ generation was found when the OL was raised to 30 g L⁻¹ and 40 g L⁻¹, in comparison to the levels of 10 g L⁻¹ and 20 g L⁻¹. This might be due to the volumetric nature of the CA enzyme addition (15 μL mL⁻¹ feed⁻¹), as the introduction of more organic load led to an elevated generation of CO₂ in the reactor. However, this rise in CO₂ production was inadequate to compensate for the lower enzyme quantity.

3.3. Influence of CA on volatile fatty acids production

Volatile fatty acids (VFAs) are the byproduct of the metabolic activity of acidogenic/acetogenic bacteria, which occurs

through the oxidative transformation of sugars and various organic compounds during the fermentation process (eqn (1) & (2)).⁴⁷



VFAs that cogenerate during hydrogen fermentation serve as an indicator to determine the progress of the process. Organic waste/wastewater fermentation based VFAs have the potential to fuel sustainability, maximize resources, and used as green platform chemicals in different industries. In this study, the presence of CA in the fermentation medium had an influence on both the concentration and composition of the VFAs. In the phase I, it was found that the net VFA production from glucose was greater in the reactor dosed with a higher enzyme load (0.95 g_{COD} L⁻¹: R3) compared to the control (0.76 g_{COD} L⁻¹: R5) and other two enzyme loads (0.9 g_{COD}: R2 and 0.88 g_{COD} L⁻¹: R1). In phase II, to develop more knowledge of the CA role in the AF process, the results of all the reactors operated with CA (R6–R14) were compared against the efficiency of all the reactors run without CA (R15–R23). In the first experiments evaluating redox conditions, it was seen that a pH 8 enhanced VFA production. Specifically, R8, supplemented with CA yielded 0.9 g_{COD} L⁻¹, while R17, which lacked enzyme, only achieved 0.78 g_{COD} L⁻¹. In the next step, the reactors were subjected to pressurization using *in situ* generated H₂ + CO₂, which along with the addition of CA, the production level raised to 0.95 g_{COD} L⁻¹ (R10, pH 7), which was 1.28 times more than that of the control reactor (R19, pH 7).

Finally, the combined effect of optimized parameters including *in situ* pressurization, alkaline pH 8 and CA were employed to assess the impact of organic load on acidogenic metabolites. The results indicated that the reactor fermenting 20 g_{COD} L⁻¹ (R12) was superior producing VFA of 0.94 g_{COD} L⁻¹, in comparison to the reactors operating even at higher loadings of 30 g_{COD} L⁻¹ (R13) and 40 g_{COD} L⁻¹ (R14). In contrast, in absence of CA, the production value exhibited a significant decrease ranging 0.67–0.84 g_{COD} L⁻¹ in R21–R23 (control to R12–R13). Overall, the net VFA production values with the reactors supported by CA (R6–R14) exhibited

Table 3 Acidogenic metabolites production from various lignocellulosic based biomass

Feedstock/ biomass	Reactor type	Pretreatment/hydrolysis	Acidogenic metabolites (VFA/hydrogen yield)	Ref.
Birch sawdust	2 L glass bottle reactor; AMPTS-II	Organosolv fractionation	121.4 mL g _{VS} ⁻¹ VFAs 20.44 g L ⁻¹	75
Duckweed	125 mL serum bottle	Acid, alkaline, and thermal hydrolyses.	169.3 mL _{H₂} g _{biomass} ⁻¹	76
Rice straw	100 mL serum bottles	Fungus <i>Gymnopus contrarius</i> J2	5.71 mmol g _{biomass} ⁻¹	77
Rice crop wastes	20 L anaerobic bioreactor	Untreated	40 mL H ₂ g _{VS} ⁻¹	78
Corn stover	500 mL glass reactor	Enzymatic hydrolysis: Ctec2	36.1 mL _{H₂} g _{biomass} ⁻¹	79
Pine tree wood	100 mL serum bottles	Dilute acid pretreatment	1.28–1.36 mol _{H₂} mol _{sugar} ⁻¹	80
Waste paper	250 mL serum bottle	Acid hydrolyzed	15.31–139.97 mL _{H₂} g _{sugar} ⁻¹	81
Grass waste	150 mL glass bottles	Ionizing radiation + acid pretreatment	68 mL g _{grass added} ⁻¹	82
Forest residues	200 mL glass bottles	Organosolv fractionation + enzyme hydrolysis	266.6 mL _{H₂} g _{COD} ⁻¹ 0.8 g _{VFA} g _{COD} ⁻¹	This study



1.28–1.12 higher compared to its control reactors (R15–R23) (Table 4).

3.3.1. Influence of CA on distribution of VFAs. The total VFA consisted of a range of fatty acids, including short chain (SCFA) such as acetic, propionic, butyric acids and medium chain (MCFA), such as caproic acid (Fig. 3). The experimental conditions had a significant impact on the concentration and distribution of individual acids in the reactors. The principal formation of acetic acid and butyric acid in a reactor represents a substantial portion (ranging 58.76–91.54%) of the net VFAs suggesting that the production of bioH₂ in the system is mostly linked to the acetate and butyrate metabolic pathways (eqn (1) & (2)). Acetic acid production is the acidogens preferred metabolic route since it offers the highest energy output. But when the H₂ partial pressure rises, the pyruvate-to-acetate conversion pathway becomes energetically unfavorable.^{39,48} This causes a switch in the metabolic pathways to make propionic and butyric acid that are more reduced than acetic acid. Presence of CA in the fermenting media can potentially balance the H₂ partial pressure due to its involvement in the CO₂ reduction. Moreover, other than acetate, other metabolites including propionate, butyrate, lactate, propanol, butanol, H₂, and CO₂, can be biosynthesized from pyruvate.

It is interesting that in phase I, the synthesis of acetic acid (0.21 g_{COD}) + butyric acid (0.43 g_{COD}) represented 84.55% of the net volatile fatty acids (VFAs) in R4. The concentrations of these specific fatty acids with R4 were somewhat higher compared to the values observed in the reactors supplemented with CA (R1–R3). In contrast, earlier observations indicated that the reactor with CA (R1, R2, and R3) had a higher net VFA production in comparison to R4. This striking difference in the distribution of individual fatty acids can be ascribed to the transformation of SCFAs into MCFAs such as caproic acid within the reactor. Caproic acids is a chain elongated fatty acids which formed through reverse β -oxidation (RBO) pathway where SCFA (acetic and butyric acid) bio-convert to longer carbon chain (caproic acid) in presence of an electron donor. The production of caproic acid through glucose fermentation, aided by CA, resulted in a significant increase in production. Specifically, the production values of 0.15 g_{COD} (R3) followed by 0.13 g_{COD} (R2) and 0.12 g_{COD} (R1) represented a 2.21, 1.93 and 1.83-fold increase, respectively, compared to the R4 (0.068 g_{COD}) operated without CA. This increase in caproic acid production may be ascribed to the utilization of *in situ* CO₂ along with organic substrate for the synthesis of SCFA, which are then transformed into MCFA. Other than RBO, MCFA yield is determined by chain elongation cyclic reactions is fatty acid synthesis. Both cyclic routes need reduced compounds as electron donors such as sugar alcohols, sugars, lactate, ethanol, methanol, and H₂, with ethanol being the most popular for high-rate chain elongation.⁴⁹ Caproic acid, exhibits significant potential in various domains, including the production of biofuels and biochemicals. To enhance the productivity of caproic acid, the microbial process needs proper investigations and regulation considering the influential factors.^{36,50}

Table 4 Acidogenic metabolites production with respect to various experimental conditions

Reactors	R1	R2	R3	R4	R5	R6	R7	R8	R9	R10	R11	R12	R13	R14	R15	R16	R17	R18	R19	R20	R21	R22	R23
H ₂ (%)	45	55	64	51	0	53	57	62	51	55	64	56.76	60	64	50	51	53	44	45	48	52	53	57
CO ₂ (%)	55	45	36	49	0	47	43	38	49	45	36	43.24	40	51.72	50	49	47	56	55	52	45.95	45.62	52.90
Lactic acid (g _{COD} h ⁻¹)	0.006	0.006	0.008	0.009	0	0.013	0.007	0.005	0.031	0.026	0.020	0.039	0.006	0.007	0.012	0.003	0.001	0.001	0.002	0.001	0.010	0.015	0.016
Ethanol (g _{COD} h ⁻¹)	0.001	0.001	0.002	0.001	0	0.023	0.022	0.020	0.023	0.016	0.018	0.027	0.015	0.006	0.019	0.014	0.012	0.031	0.023	0.006	0.001	0.022	0.013
H ₂ energy (kJ g _{COD} ⁻¹)	1.73	2.15	2.75	2.09	0.00	1.86	2.34	2.67	1.79	2.12	2.66	2.47	2.54	2.54	1.55	1.96	2.11	1.36	1.53	1.80	1.90	2.01	2.09
pH	5.13	5.62	5.8	5.1	6.89	5.26	6.57	6.14	5.11	6.21	6.34	6.35	6.21	6.23	4.87	5.26	5.47	4.79	5.19	6.45	6.25	6.24	6.37



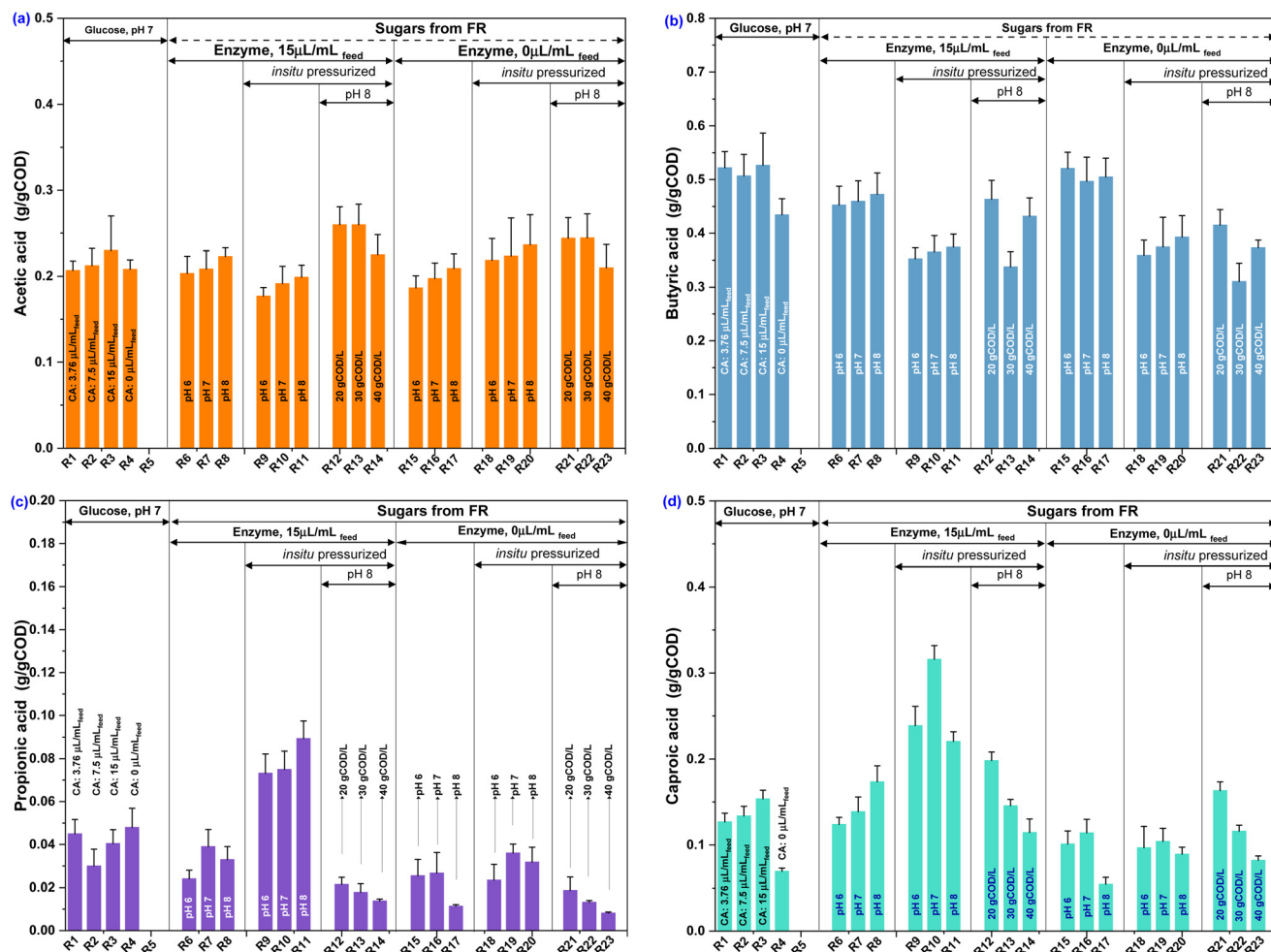


Fig. 3 Composition of (a) acetic acid, (b) butyric acid, (c) propionic acid, and (d) caproic acid with respect to experimental conditions observed during fermentation of glucose and sugars from forest residuals in phase I and phase II respectively.

In phase II, the metabolic route was altered with respect to substrate and fermentation pH (6, 7 and 8) impacting the pattern and distribution of SCFA and MCFA. While acetic and butyric acid production was minimal across all reactors, caproic acid production was significantly influenced. Upon examining the distribution pattern, it was found that SCFA ($H_{Ac} + H_{Bu}$) biosynthesis was favored at acidic (pH 6: 81.59%, R6) and neutral (pH 7: 78.98%, R7) compared to alkaline environment (pH 8: 77.1%, R8), on the other hand, caproic acid biosynthesis was favored by initial pH 8 (0.17 g_{COD} , R8) and pH 7 (0.14 g_{COD} , R7) compared to acidic pH 6 (0.12 g_{COD} , R6).

Further, the alkaline pH was found to be favorable in enhancing acetate and butyrate production when the reactors R9, R10, and R11 were pressurized. The recorded acetate production was 0.20 g_{COD} (R11, pH 8) > 0.19 g_{COD} (R10, pH 7) > 0.18 g_{COD} (R9, pH 6) whereas butyrate production was 0.57 g_{COD} (pH 8, R11) > 0.56 g_{COD} (pH 7, R10) > 0.53 g_{COD} (pH 6, R9). Alkaline pH promotes hydrolysis of substrate and provide excellent buffering capabilities.^{51,52} On the other side,

the catalytic activity of the CA is found to be more efficient in alkaline microenvironment.⁵³ Additionally, CA regulates pH and buffering in biological processes. Specifically, at alkaline pH, this enzyme expedites the swift conversion between CO_2 and bicarbonate ions, contributing to the stabilization of pH.^{54,55} Additionally, the reactors R18, R19, and R20, employed as control equivalents to R9, 10, and R11, exhibited a comparable pattern marked by elevated levels of acetate and butyrate production under alkaline conditions. However, it is important to highlight that the fatty acids in the control reactors remained accumulated, whereas in R9–R11 with CA, the accumulated fatty acids were slightly consumed indicating its transformation.

The synergistic application of pH 7, CA and pressurized system have been observed to exhibit a promising approach in steering the biosynthesis of caproic acid. The observed outcome from this combination delivered a yield of 0.31 g_{COD} , in contrast to the pH 6 (0.23 g_{COD}) and pH 8 (0.22 g_{COD}). Pressurized systems are advantageous since H_2 in the head-space serve as an electron donor along with ethanol and lactic



acid for chain elongation which was further supported by its complete consumption in the reactor.^{39,56} Although pressurized, the yields noticed from the reactors that were operated without CA was 0.097 g_{COD} (pH 6), 0.104 g_{COD} (pH 7), and 0.089 g_{COD} (pH 8) which was 59.5%, 67% and 59.6% lower, respectively compared to the reactors dosed with CA. Furthermore, the partial pressure can influence the formation of H₂ in the reactor. The presence of CA can potentially influence the process by reducing H₂ partial pressure that hinder the process, promoting its production through shifting the reaction. The preferential accumulation of acetate and butyrate at elevated concentrations in the fermentation medium during H₂ fermentation can be ascribed to the pathways that have been and discussed earlier. Moreover, various other metabolites, including propionate was found at lower levels which is an indication towards sustained H₂ production because propionic acid pathway can lead with significant consumption of H₂. Increasing the headspace pressure with *in situ* produced H₂ + CO₂ has been found a way to potentially change the acid product spectrum.

In the final step of phase II, examining the impact of organic load in conjunction with CA and pressurization, it was found that 20 g_{COD} L⁻¹ was superior to 30 g_{COD} L⁻¹ and 40 g_{COD} L⁻¹, resulting in the production of 0.94 g_{COD} (R12), 0.76 g_{COD} (R13), and 0.79 g_{COD} (R14) respectively. In contrast, the formation of MCFA without CA was extremely poor and restricted to 0.19 g_{COD} (20 g_{COD} L⁻¹), 0.14 g_{COD} (30 g_{COD} L⁻¹), and 0.11 g_{COD} (40 g_{COD} L⁻¹) in R21, R22 and R23 respectively which was 1.46, 2.72, and 2.68 times lower than its respective CA dosed reactors (R12, R13 and R14). It was expected that the yields of SCFA and MCFA would be greater at higher organic loads. The same pattern appeared in the case of bioH₂, where no significant rise was noticed. This suggests that the dosage of CA in the fermenting medium was inadequate, resulting in a limited solubilization of *in situ* CO₂ in the reactor. Consequently, this disruption of the redox equilibrium within the system occurred.

3.4. Inorganic (IC) and organic carbon (OC) utilization

An efficient utilization of organic and inorganic carbon plays a crucial role in determining the efficiency of acidogenic/aceto-genic towards biohydrogen and VFA production signifying the importance in the context of biomass hydrogen fermentation. The addition of CA significantly influenced the consumption of both the organic substrate (sugars from forest residuals) and the inorganic carbon (CO₂). Among all the 23 reactors operated in phase I and II, the highest carbon utilization efficiency above 85% was showed by the reactors dosed with CA (Fig. 4). In phase I, the complete utilization of the substrate in R3 can be attributed to the simpler substrate fermentation by mixed culture accompanied by CA addition converting 1.23 g_{COD} of organic along with 154.8 mL of CO₂ to microbial metabolites. In contrast, the reactor without CA (R4) achieved only 74.54% of substrate utilization, transforming only organic part (0.97 g_{COD}) to SCFA/MCFA leaving 200.9 mL of CO₂ unused. In the second part of the experiment, the reactors

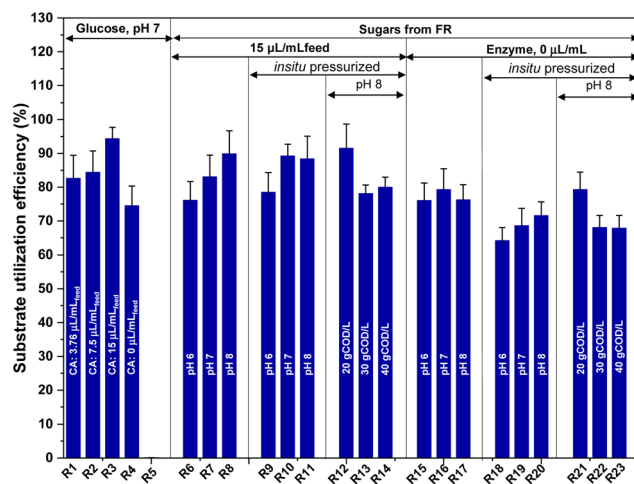


Fig. 4 Substrate utilization efficiency exhibited by individual reactors fermenting glucose and sugars from forest residuals during phase I and phase II studies.

with CA operated under alkaline conditions and pressurization (R8, R10, R11 and R12) exhibited >85% substrate utilization. R12 exhibited the highest substrate utilization efficiency of 91.54%, followed by R8 (89.90%), R10 (89.22%), and R11 (88.39%) with simultaneous utilization of CO₂. These reactors successfully transformed substrate of 1.19 g_{COD}, 1.17 g_{COD}, 1.16 g_{COD}, and 1.15 g_{COD}, respectively, into metabolites. The control reactors (R15–R23), which were run in parallel with the specified configuration without enzymes, demonstrated that the absence of CA may have an influence on the formation of metabolites inside the reactor, leading to a decrease in the efficiency of product generation. Reactors supplemented with CA used almost all the *in situ* CO₂ they produced, whereas lack of CA in R13–R23 used just 12–37%. These results suggest that presence of CA not only helped to enhance organic substrate transformation to metabolites but also assisted in the *in situ* CO₂ utilization. The utilization of CO₂ through the addition of CA within the reactor presents a promising avenue for effectively managing and harnessing its potential while mitigating its environmental impact. This approach holds the potential to not only minimize the adverse effects of CO₂ but also create additional value in the process. The utilization of CA in fermentation system is anticipated to result in a favorable response in terms of promoting the proliferation and metabolic functionality of bacteria such as homoacetogens that exhibit the capability utilizing CO₂ and H₂.²⁷ CA can potentially impact a specific group of microorganisms, steering them towards the efficient utilization of CO₂ by providing a conveniently accessible carbon substrate. This substrate is particularly advantageous for bacteria that rely on CO₂ as their primary source of carbon. Additionally, the use of CA in the mitigation of CO₂ particularly during fermentation process exhibits considerable potential as a technology to enhance the efficacy of the processes along with controlling the release of GHG into the atmosphere.



3.4.1. Distribution of substrate. A COD mass balance was performed on the fermentation system to evaluate the reactor performance in the presence and absence of CA, focusing on influent COD, metabolite transformation, and residual substrate in the liquid phase. The mass balance demonstrated a significant impact of CA on gaseous and liquid metabolites within the reactors. Carbon transformation (in terms of COD) to metabolites was significantly higher during the pH variation study in the presence of CA (R6: 0.804 g_{COD}; R7: 0.846 g_{COD}; R8: 0.902 g_{COD}) compared to control reactors without the enzyme (R15: 0.834 g_{COD}; R16: 0.835 g_{COD}; R17: 0.780 g_{COD}) (Fig. 5). Most of the carbon transformation occurred in the creation of butyric acid rather than acetic acid. It was determined that most of the bioH₂ was produced *via* the routes involving acetic and butyric acid fractions in the medium. When the systems were closed, the carbon conversion to metabolites was higher in reactors dosed with CA (R9: 0.842 g_{COD}, R10: 0.948 g_{COD}, R11: 0.883 g_{COD}) compared to those operating without CA (R18: 0.688 g_{COD}, R19: 0.738 g_{COD}, R20: 0.75 g_{COD}). Moreover, the *in situ* produced CO₂ was completely used in the fermentation process, specifically with R9–R11. During organic load testing, the conversion of organic carbon into metabolites was greater in reactors with CA (R12: 0.94 g_{COD}; R13: 0.76 g_{COD}; R14: 0.78 g_{COD}) compared to reactors without CA (R21: 0.841 g_{COD}; R22: 0.684 g_{COD}; R23: 0.674 g_{COD}). CO₂ produced and utilized in the reactor also play a role in the production of organic acids, influenced by the bacteria and fermentation conditions. Determining the exact transformation of organic carbon and subsequent CO₂ generation into organic acids during mixed culture fermentation poses challenges.

3.5. Microbiological analysis

Microbiome investigations were conducted to assess the microbial composition with respect to the progression of the acidogenic metabolites by a mixed culture, both in the presence and absence of CA. Fig. 6 and 7 presents a comprehensive depiction of the relative abundance of various microbial species that constitute the acidogens enrichment at the end of the experiment. The microbial community structure of all 18 reactors (R6–R23) operated during phase II was assessed by comparing the reactors with (R6–R14) and without CA (R15–R23). The Venn diagram illustrates greater levels of genetic variation and difference among the reactors operated at different experimental conditions. Fig. 6a shows the comparative analysis utilizing the 6 samples obtained from the reactors, which were subjected to varying pH conditions, demonstrated that among the 475 operational taxonomic units (OTUs), a total of 5 OTUs (1.05%) were found to be present in all six groups, regardless of the presence or absence of the enzyme in the reactor. This observation suggests that these OTUs might exhibit a species-specific nature. The R6, followed by R7, R8, R15, R16, and R17, exhibited a total of 71, 70, 67, 69, 66, and 74 unique OTUs respectively. The reactors with (R9–R11) and those without CA (R18–R20) operated under pressurized conditions shared 12 OTUs, however CA-dosed reactors possessed 65 (R9), 68 (R10), and 63 (R11) unique

OTUs, while control to these reactors displayed 73 (R18), 100 (R19), and 68 (R20) (Fig. 6b). During OL operation, all the six reactors exhibited a shared set of 9 OTUs among them. The reactors, with CA, displayed unique OTUs of 60 (R12), 67 (R13), and 60 (R14). In contrast, without CA operation exhibited unique OTUs of 65 (R21), 72 (R22), and 69 (R23) respectively (Fig. 6).

3.5.1. The shift on bacterial community. The community structures at phylum and genus level can be observed in Fig. 7. A major shift in the microbial community structure was observed with respect to experimental conditions applied. All the reactors (R6–R23) were enriched with Firmicutes (99.49–58.31%), followed by *Bacteroidota* (5.13–2.8%) and *Actinobacteriota* (1.31–1.03%). Relative abundance of phylum Firmicutes dominated the whole process, particularly *Clostridiaceae* families. The introduction of CA profoundly impacted the composition of the bacterial community within the reactors, resulting in significant differences from the control reactors. Upon individual examination, it was observed that the reactors with CA (R6–R14) had more Firmicutes than those without CA (R15–R23). Specifically, the abundance of Firmicutes was $93.31 \pm 2.44\%$ (R6–R8) during pH optimization, $84.08 \pm 2.76\%$ (R9–R11) during *in situ* pressurization, and 95.12% during OL variation operation (R12–R14). In contrast, Firmicutes dominance declined without CA, with $80.87 \pm 3.58\%$ (R16–17), $71.12 \pm 4.74\%$ (R18–20), and $77.07 \pm 4.07\%$ (R21–23). In terms of the genera that produce hydrogen, reactors with CA were dominated by *Clostridium sensu stricto*, *Paraclostridium*, and *Romboutsia*, but the growth of these genera was less in reactors operated without CA. *Clostridium sensu stricto* 1, can utilize a diverse range of carbon sources including glucose, xylose, sucrose, hemicellulose, cellulose, and starch. This metabolic versatility has been found to contribute to the efficient production of hydrogen through a dark fermentation/acidogenesis.^{36,57} The enrichment of the hydrogen and volatile fatty acid (VFA) producing bacterium *Clostridium sensu stricto* 1 was observed to be significantly higher in reactors R6 (57.7%), R7 (52.7%) and R8 (53.6%) that were operated with the addition of CA, as compared to reactors R15 (19.5%), R16 (18.1%), and R17 (18.5%) that were operated without CA. This effect was observed at pH 6, pH 7, and pH 8, respectively. A relatively poor bioH₂ production from the control reactors can be attributed to the low number of hydrogen production genera. This finding is corroborated by the previous studies that demonstrated *Clostridium sensu stricto* as the primary hydrogen producer and have observed a correlation between its abundance and hydrogen yield.^{36,57,58} Yang and Wang examined microbial community structure changes in dark fermentative hydrogen generation and found that *Clostridium sensu stricto*1 contributed more to hydrogen production.⁵⁹ Kumar *et al.* used experimental design to determine optimum hydrogen generation settings and discovered that *Clostridium* sp. dominated.⁶⁰ During *in situ* pressurization, propagation of *Clostridium sensu stricto* was slightly higher with R9, R10 and R11 (21–44%) compared to the reactors operated without CA (R18, R19 and R20). Similarly, the existence of



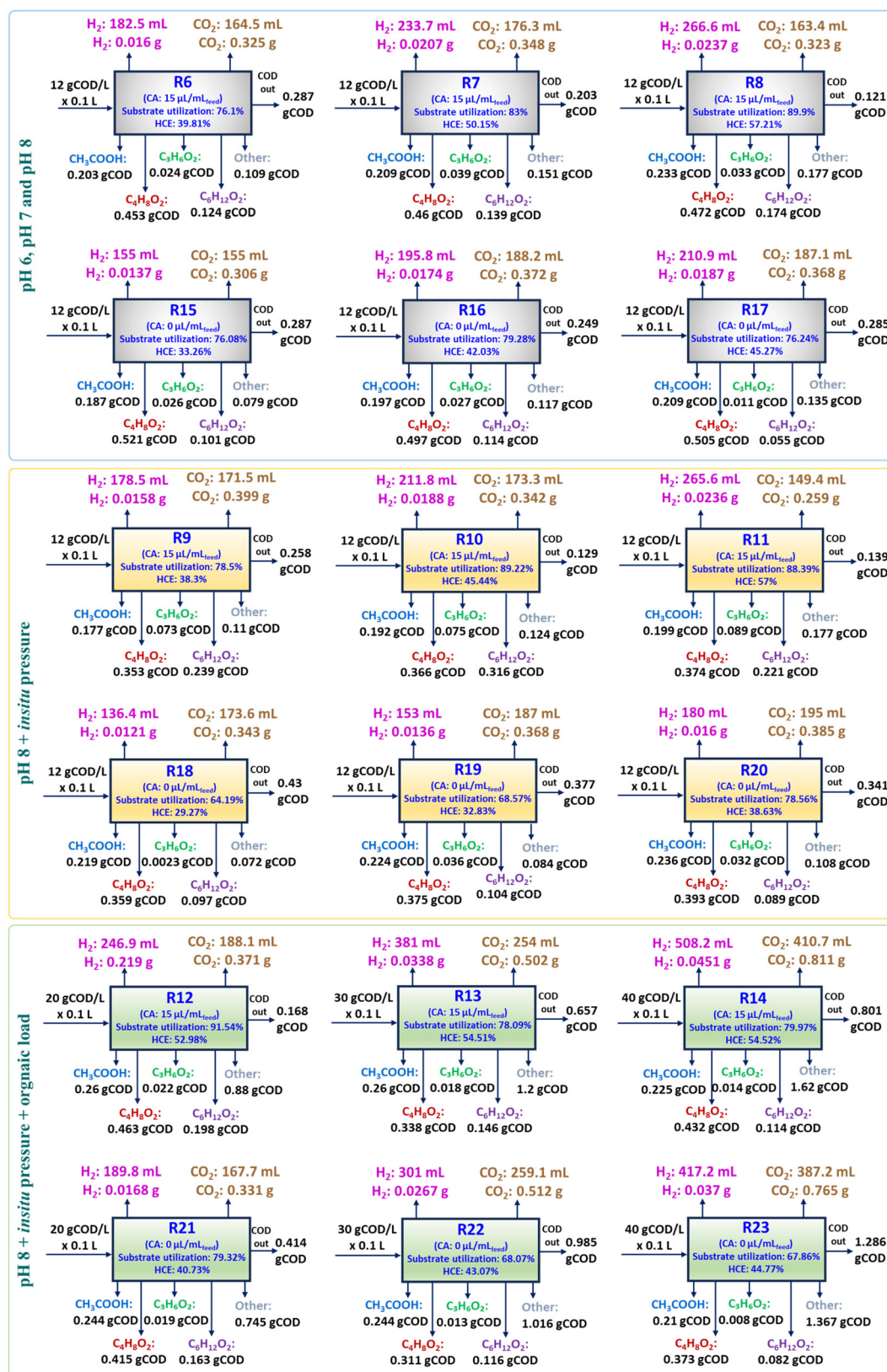


Fig. 5 The impact of carbonic anhydrase on the distribution of substrate in the reactors during mixed culture fermentation.



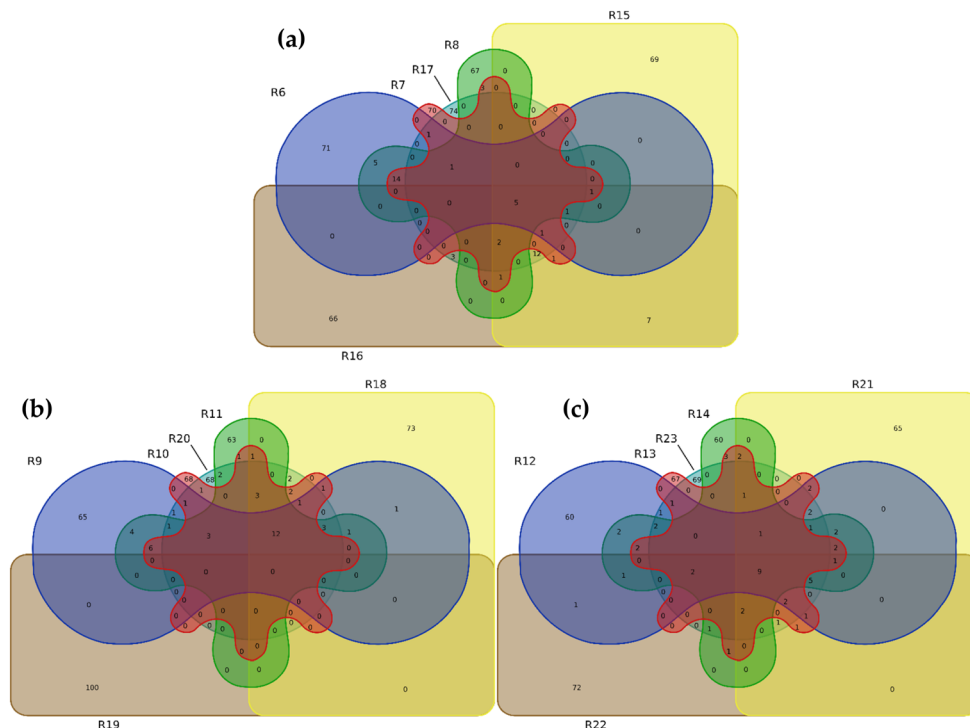


Fig. 6 Venn diagrams showing the number of bacterial OTUs shared among reactors fermenting sugars from FR in phase II under different experimental conditions. Reactors with CA (R6, R7 and R8) compared with its respective control reactors (R15, R16 and R17) (a). Similarly, "R9, R10, R11" compared with "R18, R19, R20" (b) and "R12, R13, R14" compared with "R21, R22, R23" (c).

Clostridium sensu stricto 1 was significantly reduced in the reactors operated without CA (R21, R22 and R23), as compared to the reactors supplemented with CA (R12, R13 and R14).

Another hydrogen-producing genus that dominated reactors was *Paraclostridium* which is an obligate anaerobe that can efficiently produce hydrogen from wastewater sludge and glucose by dark fermentation.^{59,61} The documented significant production of hydrogen particularly with R6, R7, and R8 can be connected to the concurrent proliferation of *Paraclostridium* in conjunction with *Clostridium sensu stricto*. The prevalence of *Paraclostridium*, was seen in the reactors operated with CA was higher at pH 6 (23.8%, R6) in comparison to pH 7 (20.9%, R7) and pH 8 (19.4%, R8). Conversely, in the absence of enzymes, these percentages significantly decreased to a range of 3–8% in R15, R16 and R17. *Paraclostridium*, an obligate anaerobe, exhibits the ability to undergo sporulation. The utilization of organic carbon by *Paraclostridium* sp. result with an efficient hydrogen production. The addition of CA did not stimulate a growth of *Paraclostridium* in pressurized reactors, as its proliferation was restricted to a range of 4–10%. In contrast, the operation without CA under pressurized conditions resulted in the total elimination of *Paraclostridium*. The suppressed development growth of *Paraclostridium* under pressurization condition indicated that the pressure could have impacts on its growth. However, the observed developed headspace pressure (3–4 bar) was found to be beneficial for other genera, including *Clostridium sensu stricto 1*, *Clostridium sensu stricto 12*,

Romboutsia, and *Caproiciproducens*. These pressure-adapted genera are especially known for their ability to produce biohydrogen and volatile fatty acids.

Clostridium sensu stricto 1, *Clostridium sensu stricto 12*, *Romboutsia*, *Paraclostridium*, and *Caproiciproducens* were the genera associated with biohydrogen and VFAs production. The *Clostridium butyricum*, a bacterium belonging to the genera *Clostridium sensu stricto 1* (Firmicutes), exhibited a strong association with the implemented experimental conditions. The Firmicutes bacterial populations exhibited dominance in terms of hydrogen production, with the related species of the *Clostridium* genus occupying the prominent population position in the reactor, thereby resulting in enhanced efficiency of hydrogen production. The primary metabolic pathways associated with hydrogen production encompass the acetate and butyrate pathway. The experimental findings show a strong positive connection between acetate and butyrate biosynthesis with hydrogen cogeneration. *Clostridium* has garnered interest for hydrogen fermentation. Mixed culture inoculum investigations related most *Clostridium* genus to high hydrogen production from a variety of organic substrates.⁶² This study contained more than four genera of *Clostridium*. One of the most common fermentative H₂ producer is *Clostridium butyricum*, which makes up to 17–27.65% of all bacteria in the reactor dosed with CA operated under *in situ* pressure.

Romboutsia, a member of the family *Peptostreptococcaceae* within the phylum *Firmicutes* has the capacity to metabolize



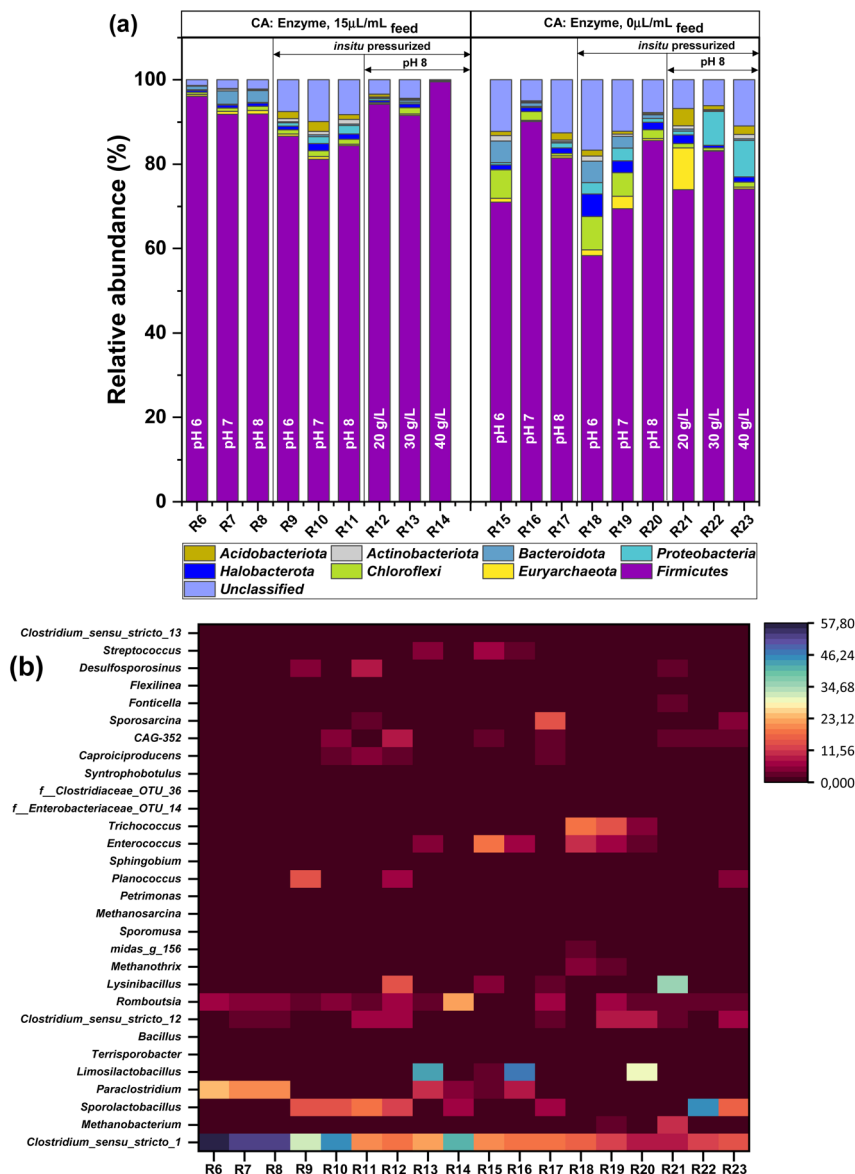


Fig. 7 A heatmap was generated to illustrate the relative abundance of genera (%) in the specific reactor during phase II operation, with a focus on comparing enzyme-supplemented versus enzyme-less fermentation under various experimental conditions (a–b).

carbohydrates, resulting in the production of acetate, formate, ethanol, and lactic acid.^{58,63} The observed increased prevalence of *Romboutsia*, in conjunction with other genera, exhibited a consistent pattern with the elevated VFA and bioH₂ in the reactors. The abundance of *Romboutsia* in during pH optimization was $5.2 \pm 0.84\%$ (R6–R8), while without CA it was a slightly lower fraction ($2.72 \pm 1.72\%$), highlighting the significance of CA in *in situ* buffering and promoting favorable condition for microbial growth. Further, growth of *Romboutsia* was marginally decreased particularly under pressurization indicating that developed *in situ* pressure was unfavorable on its propagation regardless of the presence or absence of CA. However, moving towards increase in organic load proved to be beneficial as it notably facilitated the proliferation of the

bioH₂ and VFA producer *Romboutsia*, especially in the reactors with CA (10.23%, R12–R14) over without CA (2.86%, R21–R23).

The biosynthesis of caproic acid can be attributed to the enrichment of chain elongator bacteria in the mixed culture. *Caproiciproducens* belongs to the family *Ruminococcaceae* under firmicutes known for caproic acid along with molecular hydrogen and other VFAs production.^{64,65} Chain elongation driven by lactate occurs in a few species of *Caproiciproducens*. During lactate oxidation, the yielded pyruvate undergoes subsequent oxidation, resulting in the formation of acetyl-CoA and CO₂ with simultaneous electrons in the form of reduced ferredoxin. The acetyl-CoA, which is obtained through a process, serves as substrate for the biosynthesis of butyrate and caproate *via* a reverse β -oxidation pathway.⁶⁶ The reactors



R11 and R12 exhibited a presence of *Caproiciproducens* (>3%), particularly when dosed with CA and subjected to *in situ* pressurization. These reactors showed a lactic acid consumption at the rate of 0.02 g_{COD} h⁻¹ (R11), and 0.039 g_{COD} h⁻¹ (R12) leading to its complete utilization within 48-hour. On the other side, the proliferation of *Caproiciproducens* was much lower (<3%) in the other reactors. Despite being below 3% in these reactors, the production of caproic acid showed significant efficacy, potentially attributable to the other chain elongators and their synergistic interactions within the reactors. The batch experiments demonstrated the production of lactic acid and ethanol, as well as the substantial consumption and transformation of these molecules into short-chain fatty acids (SCFAs) and medium-chain fatty acids (MCFAs). These findings indicate the presence of a beneficial synergy in the mixed culture, despite the presence of lactic acid bacteria (LAB), which ultimately enhanced the yield of the desired products.

Few setups in this study (particularly when the reactors were pressurized; R9, R10, R11, R12, R22 and R23) showed the dominance of LAB. The contribution of LAB, specifically *Sporolactobacillus*, in bioH₂ production remains an area that requires further elucidation. Previous studies showed that LAB can have a negative impact on bioH₂ production due to substrate competition or with release of bacteriocins. For example, Cieciura-Włoch *et al.* (2020) observed instability in the bioH₂ production attributing towards the prevalence of the Lactobacillaceae family accounting for 59% of the total microbial population in the reactor.⁶⁷ As per the findings of Park *et al.*, there was a sudden termination in bioH₂ production due to accumulation of lactic acid.⁶⁸ In the study conducted by Chatellard *et al.* (2016), it was observed that the presence of various carbohydrates had distinct effects on the growth of mixed bacterial communities. Specifically, hexoses were found to promote the proliferation of LAB, while pentoses were observed to stimulate the growth of BioH₂ producing bacteria.⁶⁹ Forest residues hydrolysate used as substrate in this study, containing of good fraction of glucose which theoretically provides equal conditions for the proliferation of both lactic acid and hydrogen producing bacterial species. Nevertheless, other research has shown that LAB have a superior growth rate and enhanced substrate absorption capabilities compared to *Clostridia*, hence granting them a competitive advantage over other fermentative bacterial species.^{70,71}

In this study, lactic acid synthesis was observed on day one, but its complete utilization was demonstrated as well by day 3, indicating its transformation to other metabolites such as chain elongated butyric or caproic acid. This finding aligns with previous studies investigating microbial diversity derived from hydrogen-producing bioreactors where Esquivel-Elizondo *et al.* (2017) postulated that *Lactobacillaceae* are the primary producers of butyrate in the reactor.⁷² Likewise, other studies have made similar assumptions on volatile fatty acids production.^{36,58,71} In a study conducted by De Leeuw (2019), it was observed that the presence of *Caproiciproducens* ASV2 in the reactor, alongside *Lactobacillus rhamnosus* ASV4, during

the initial high level of total VFAs production resulted in a higher production of butyric and caproic acids, while the production of acetic acid was lower. This outcome is likely attributed to a chain elongation process facilitated by *Caproiciproducens*, utilising lactic acid generated by *Lactobacilli* as an electron donor.⁷³ External electron donors, such as lactic acid or ethanol, in the media promote the development of chain elongators. However, lactic acid and ethanol are also byproducts of fermentation. The findings of our study clearly show that the complete utilization of these electron donors within the reactor serves as the primary source of electrons for the chain elongation process. On contrary, studies that have documented the presence of LAB with H₂ production indicating the potential role of LAB in providing supplementary substrates to hydrogen-producing bacteria, either in the form of lactate or through the direct conversion of lactate into hydrogen.

The typical chain elongator *Clostridium kluyveri*, is known for caproic acid production from acetic and butyric acid and is a member of the genus *Clostridium sensu stricto* 12, which also included homoacetogens. Genera *Clostridium sensu stricto* 12 had the second-highest growth after *Clostridium sensu stricto* 1. Reactors supplemented with enzyme showed the good association between *Clostridium sensu stricto* 12 and consumption of ethanol, H₂ and *in situ* CO₂. Particularly the abundance of *Clostridium sensu stricto* 12 was higher with the reactor operated under *in situ* pressurization. The application of pressurization potentially elicited an augmented uptake of carbon dioxide in the presence of an enzyme, thereby promoting the proliferation of the microorganism *Clostridium sensu stricto* 12. Furthermore, the application of pressurization conditions promotes the utilization of H₂ and CO₂ by homoacetogens.⁷⁴ Similar to the pattern of lactic acid consumption, ethanol was found to be consumed at a rate ranging from 0.001 to 0.031 g_{COD} h⁻¹ until it was completely consumed. The production and consumption of ethanol was slightly higher with the reactors operated at pH 6 (0.023–0.031 g_{COD} h⁻¹) compared to pH 7 (0.022–0.023 g_{COD} h⁻¹) and pH 8 (0.013–0.014 g_{COD} h⁻¹). The increased fraction of two genera *Clostridium sensu stricto* 12 and *Caproiciproducens* in the mixed culture was responsible for the production of hexanoic acid through chain elongation process. The observed shifts in microbial communities and the selective enrichment of certain bacterial species during mixed culture fermentation highlight the importance of experimental conditions shaping the structure and adaptability of the community towards biohydrogen and VFAs from wood biomass hydrolysate.

Exploring bio-based chemicals and fuels as a sustainable alternative to fossil-based production is a widely explored method for ensuring fuel and chemical supply while also reducing CO₂ emissions. Even though the CO₂ produced from fermentation processes is classified as biogenic since the carbon is sourced from sustainable resources and is part of a natural biogenic carbon cycle. It is not recommended to overlook CO₂ emissions from fermentation processes because of their substantial impact on greenhouse gas emissions. Implementing



the traditional fermentation process in conjunction with the CA enzyme-based method can be a viable option contributing to reducing carbon emissions in a roundabout yet efficient manner, transforming CO₂ into biobased fuels and chemicals. Employing an appropriate treatment such as stem explosion fractionization, forest residues can serve as an excellent feedstock for biobased products. This study particularly focused on examining the effects of CA on mixed culture fermentation of stem exploded forest residues optimizing various influential factors. However further research in carbonic anhydrase-based fermentation at different scales are necessary to gain greater insight of the process and regulate the system to enhance the recovery of metabolites.

4. Conclusion

Forest residues exhibit great potential as a renewable feedstock for the production of biobased fuels and chemicals. In this study, we have successfully demonstrated the potential of carbonic anhydrase derived from *Desulfovibrio vulgaris* on the acidogenic process across various experimental settings. The addition of CA not only enhanced the production of acidogenic metabolites such as biohydrogen and volatile fatty acids, but also contributed to the mitigation of *in situ* CO₂. The individual and intricate interactions of the chosen factors (pH, organic load, and *in situ* gas pressurization) were observed to greatly enhance the acidogenic product outcome compared to their individual effects in the presence of CA. The abundance of biohydrogen and VFA-producing *clostridium* genera and its synergistic interaction with other bacterial community in the mixed culture, indicated the crucial role of CA in regulating the process efficiency, leading to the development of an environment conducive to the efficient synthesis of acidogenic/acetogenic metabolites. Overall, this strategy of utilizing produced acidogenic CO₂ within a system is in line with the principles of circular economy, waste reduction, and sustainable resource management. It helps maximize the benefits of bioprocesses, such as anaerobic digestion, while minimizing environmental impacts and contributing to a more sustainable and environmentally friendly approach to huge quantities of forest residuals utilization for bioenergy and other value-added products biosynthesis.

Conflicts of interest

The authors declare no conflict of interest.

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