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Study of electrospun nanofibers loaded with Ru(II) phenanthroline complexes as a potential material for use in dye-sensitized solar cells (DSSCs)⁺

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Dye-sensitized solar cells (DSSCs) are an increasingly attractive alternative energy source because of their low cost. Therefore, researchers have intensified efforts over the past decade to increase their energy conversion efficiency by employing new materials in each DSSC component. The present research focuses on synthesizing electrospun nanofibers as a potential new material as a counter electrode in DSSCs. Two Ru(II) half sandwich 1,10 phenanthroline (phen) Ru-1 and 5-amino- phen Ru-2 complexes were prepared for its functionalization. As a deposition medium, poly(caprolactone) (PCL) dissolved in chloroform was used. Different Ru(II) complex concentrations were made at 0.1% wt., 0.5% wt., and 1% wt. Thermal characterization studies using differential scanning calorimetry (DSC) and thermogravimetric analysis (TGA) were conducted to evaluate the behavior and weight loss of the samples with temperature variations. Fourier transform infrared spectroscopy (FTIR) measurements were taken to observe the bond interaction of the ruthenium complexes and the PCL. Finally, scanning electron microscopy (SEM) was used to structurally and morphologically evaluate the fiber distribution and porosity. These fibers have a homogeneous morphology, without bulbs, but with evident solid inlays on the surface, with fibers between ~ 0.58 to 2.47 μ m and percentages of porosity $\sim 45\%$. TGA and DSC thermograms show minor temperature variations that demonstrate the incorporation of the Ru(II) complexes into the fiber. Furthermore, the melting and degradation temperature of the fibers is suitable for use in a DSSC approach. The incorporation of the ruthenium compounds into PCL fibers, along with the addition of the NH₂ group into complex Ru-2, resulted in a higher current density for both anodic and cathodic peaks in Cyclic Voltammetry (CV). It is noteworthy that from I–V curves, PCL-Ru2 1% fibers demonstrated a conductivity of 0.461 µS cm⁻¹, which is comparable to other PCL fibers carrying a higher metal load. Future studies will delve into the mechanical properties of these fibers to highlight their potential for application in this field.

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1 Introduction

Dye-sensitized solar cells (DSSCs) are photovoltaic cells whose main characteristic is to mimic plants in the capture of sunlight, and their objective is to simulate artificial photosynthesis, absorbing solar energy that is later transformed into electrical power.¹

Conventional DSSCs are comprised of four main parts: photo-anode, sensitizing dye layer, electrolyte, and counterelectrode (Fig. 1).¹⁻⁴ Due to their low cost and ability to operate under non-standard conditions, they are ideal for locations where environmental conditions vary widely. DSSCs are efficient in dim and diffuse light (dusk, cloudy days, or even indoors), making them suitable for various applications such as panels, tiles, facades, solar roofs, mobile electronic devices, and intelligent sensors in the automotive and nautical industries.⁵

One of the drawbacks of DSSCs is their low efficiency despite its multiple advantages. Recently, many studies have investigated strategies to increase their efficiency, including developing electrolytic components such as photo-anodes, counterelectrodes, and electrolytes.⁶

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Fig. 1 Assembly of dye-sensitized solar cells (DSSC).9,10



Fig. 2 Assembly of electrospinning device, counter-electrode with electrospinning nanofibers in DSSC. 5,6,9,10

The electrospinning technique is used to fabricate fibers that create three-dimensional fibrous scaffolds with important applications, particularly in medicine and engineering.^{7,8} In this technology, a conductive polymer solution is injected, electrically charged, and directed towards a collector against a ground-connected electrode, closing the circuit. The solution passes through a field of electrostatic forces ranging from 5 to 35 kV, producing a stretching effect that forms fibers from a few nanometers to micrometers.^{6,9,11,12}

Due to their excellent performance, Pt-based counterelectrodes are the most widely used in DSSCs. However, the methods for their manufacture are based on thermal deposition and vacuum bombardment. However, such techniques require high temperatures and complex installations.¹³ Therefore, alternatives to these increased operating costs are required in addition to the cost of Pt compounds.

Several studies have proposed the manufacture of the DSSC's counter-electrode (CE) using the electrospinning technique.¹³⁻²⁵ The CE is typically made of conducting materials that facilitate the regeneration of the photosensitizer, so it plays a crucial role in DSSC because it performs as the catalyst by reducing the oxidized form of the photosensitizer, allowing it to be regenerated and participate in subsequent light absorption and electron transfer processes, thereby completing the DSSC circuit.¹⁶

Transition metals have been used to manufacture electrospun fibers as CE in DSSCs. Bimetallic systems, such as Ni-Co,20,24 Fe-Ni,21 and Co-Ti,23 have been incorporated into carbon nanofibers (CNFs) for this purpose. Additionally, nanofibers with Pt nanoparticles,15 Ni,25 and Zr-O2 (ref. 26) have been reported. Although ruthenium(II) complexes have traditionally been used as photosensitizers in DSSCs,^{27,28} they are also promising for producing electrospun nanofibers as the CE due to their high electrical conductivity and catalytic properties. In a recent study,²⁹ researchers used Ru(II) nanofibers instead of the usual Pt counter-electrode for DSSCs. The nanofibers were created through electrospinning, post-calcination, and hydrogen reduction, resulting in improved photovoltaic performance.29 This is attributed to their high electrical conductivity and unique network structure, which outperforms the traditional Pt CE. In this research, a novel electrospun fibrous scaffold system was created for potential use as a counter electrode in DSSCs (Fig. 2). Poly(caprolactone) (PCL)

nanofibers were utilized and loaded with two different types of Ru(II) complexes ([RuCl (*p*-cymene) (phen)] Cl) **Ru-1** and ([RuCl (*p*-cymene) (5-amino-phen] Cl) **Ru-2**. We predict that introducing the -NH₂ group in the **Ru-2** complex will influence the structure and conductivity of the nanofibers. To the best of our knowledge, these Ru(II) complexes have not been previously used as functionalizing agents of electrospun nanofibers as a material for this potential application in DSSCs.

2 Materials and methods

2.1 Materials

Poly (ɛ-caprolactone) (PCL) (MW: 80 000 Sigma Aldrich), solvents such as chloroform, methanol, and hexane are analytical grade. Commercially available reagents 1,10 phenanthroline and 5-amino-1,10-phenanthroline were obtained from Aldrich Chemical and used as received.

2.2 Experimental

Ruthenium(II) complexes synthesis was carried out in an argonfilled Glove Box or using standard Schlenk techniques. [Ru(pcymene] Cl₂]₂ was prepared using published procedures.³⁰ NMR spectra were recorded at 400 MHz with Bruker Avance III spectrometer at 30 °C unless otherwise specified. ¹H and ¹³C NMR chemical shifts are reported in ppm referenced to residual solvent resonances (¹H NMR: 7.24 for CHCl₃ in CDCl₃, 2.50 for DMSO d_{6} .) ¹³C {¹H} NMR: 77.23, 39.52 respectively). Coupling constants *J* are given in Hertz (Hz). IR spectra were recorded on a PerkinElmer FT-IR 1605 spectrophotometer (ATR mode). Spectra were collected at 20 °C, in the 4000–500 cm⁻¹ range. Four sweeps were performed at a resolution of 16 cm⁻¹. High-Resolution Mass Spectrometry (HRMS) data were obtained in a micrOTOF-Q III MS instrument with electrospray ionization using sodium formate as a calibrant.

2.3 Synthesis of complex Ru-1 [RuCl(p-cymene)(phen)]Cl

To a solution of 1,10-phenanthroline (phen) (0.035 g, 0.1942 mmol) in methanol, $[RuCl_2(p\text{-cymene})]_2$ (0.059 g, 0.0971 mmol) was added and stirred at room temperature for 3 hours. The yellow-red solution was evaporated under vacuum, washed (3 ×

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10 mL) with hexane, and dried under vacuum to afford a yellow solid. Yield: (0.076 g, 80.5%). ¹H NMR (400 MHz, DMSO d_6): δ 0.90 (d, J = 6.8 Hz, 6H –CH(CH₃)₂), 2.17 (s, 3H, CH₃₋*p*-Cymene), 2.62 (sept, J = 6.8 Hz, 1H, –CH(CH₃)₂), 6.14 (d, J = 6.1 Hz, 2H, Ar-*p*-cymene), 6.38 (d, J = 6.2 Hz, 2H, Ar-*p*-cymene), 8.16 (dd, J = 8.16, 5.24 Hz, 2H), 8.3 (s, 2H), 8.94 (d, J = 7.72 Hz, 2H), 9.99 (d, J = 5 Hz, 2H) ppm. ¹³C (¹H} (101 MHz, DMSO d_6) δ : 18.1 (CH₃₋*p*-Cymene), 21.6 (-CH(CH₃)₂), 30.3 (-CH(CH₃)₂), 83.7 (Ar-*p*-cymene), 85.9 (Ar-*p*-cymene), 102.0 (C-iPr, *p*-cymene), 103.9 (C-CH₃, *p*-cymene), 126.3, 127.4, 129.0, 138.7, 145.0, 156.0 (CH= N) ppm. UV-Vis (DMSO, 5 × 10 ⁻⁴ M), λ 302 nm, $\varepsilon = 25$ 150 L cm⁻¹ mol⁻¹, λ 317 nm, $\varepsilon = 17$ 080 L cm⁻¹ mol⁻¹. HRMS (ESI-TOF) *m/z*: [M – Cl]⁺ calculated for C₂₂H₂₂ClN₂Ru 451.0510; found: 451.0502.

2.4 Ru-2 [RuCl (p-cymene)(5-amino-phen)]Cl

To a solution of 5-amino-1,10-phenanthroline (5-amino-phen) (0.060 g, 0.307 mmol) in methanol, $[RuCl_2(p-cymene)]_2$ (0.094)gr, 0.1536 mmol) was added, and stirred for 3 hours; the orange-red solution was evaporated in vacuo, washed (3×10) mL) with hexane, to afford a red solid. Yield: $(0.127 \text{ g}, 82.2\%)^{1}$ H RMN (400 MHz, DMSO d_6): δ 0.88 (dd, J = 4.9 Hz, 6H, - $CH(CH_3)_2$, 2.16 (s, 3H, CH_3 , *p*-Cymene), 2.57 (sept, I = 6.8 Hz, 1H, -CH(CH₃)₂), 6.05 (dd, J = 10.9, 6.6 Hz, 2H, Ar-p-cymene), 6.27 (dd, J = 6.5, 0.8 Hz, 2H, Ar-p-cymene), 6.96 (s, 1H), 7.00 (s, 1H), 7.81 (dd, *J* = 8.3, 5.1 Hz, 1H), 8.09 (dd, *J* = 8.4, 5.2 Hz, 1H), 8.41 (dd, J = 8.3, 0.7 Hz, 1H), 9.12 (d, J = 8 Hz, 1H), 9.43 (dd, J = 5.1, 0.9 Hz, 1H), 9.88 (d, J = 4.5 Hz, 1H) ppm. ¹³C {¹H} (DMSO d_6 , 101 MHz) δ: 18.1 (CH₃.p-Cymene), 21.5 (-CH(CH₃)₂), 30.3(-CH(CH₃)₂), 83.5 (Ar-*p*-cymene), 85.8 (Ar-*p*-cymene), 102.6 (C-iPr, p-cymene), 103.4 (C-CH₃, p-cymene), 123.0, 124.6, 125.8, 132.1, 133.9, 134.7, 138.9, 145.9, 150.2 (CH=N), 155.6 (CH=N) ppm UV-Vis (DMSO, 5×10^{-4} M), λ 270 nm, $\varepsilon = 12690$ L cm⁻¹ mol⁻¹, λ 297 nm, ε = 12 890 L cm⁻¹ mol⁻¹, λ 344 nm, ε = 4840 L cm⁻¹ mol^{-1} , λ 437 nm, $\varepsilon = 1680$ L cm⁻¹ mol⁻¹. HRMS (ESI-TOF) *m/z*: $[M - Cl]^+$. Calculated for $C_{22}H_{23}ClN_3Ru[M]^+$ 466.0621; found: 466.0625.

2.5 Preparation of polymeric solution (PCL)

A 10% (v/w) solution of PCL in chloroform was used. The preparation was done at room temperature (\sim 25 °C) and humidity of \sim 33%, under constant stirring (200 rpm) for 24 h until a homogeneous state.

2.6 Electrospinning

To a 10% (v/w) chloroform solution of PCL (*vide supra*), the Ru(II) complexes (**Ru-1** and **Ru-2**) were added according to the concentrations in Table 1 and stirred at 850 rpm for 1 hour until a homogeneous mixture was obtained. To electrospun the samples (at room temperature and relative humidity of 32–36%), 1.5 mL of the solutions were poured into a 5 mL syringe at a distance of 10 cm from the needle to the collector. A voltage of 20 kV was applied using a power source, with a constant injection flow of 0.5 mL min⁻¹, using a needle 22 G × 40 mm, and an exposition time of 200 min (Table 1).

Table	1	Concentration	of	Ru(II)	complexes	solutions	used	for
electro	osp	pinning						

Samples	Ru complex concentration (wt%			
PCL/Ru-1 0.1%	0.1			
PCL/Ru-1 1.0%	1.0			
PCL/Ru-2 0.1%	0.1			
PCL/Ru-2 0.5%	0.5			
PCL/Ru-2 1.0%	1.0			

2.7 Scanning electron microscopy (SEM)

The morphology and diameter of the polymer fibers were determined by SEM. A small section of the fibrous material was placed in a JEOL JSM 7600F field emission microscope. The samples were prepared in advance, randomly cutting about 5×5 mm pieces and placing them on an aluminum base with magnetic tape to fix the sample on the surface. This system was coated by sputtering with gold to prevent surface charging effects, and a 20 kV accelerated voltage was used to obtain the images.

The diameter of the fibers was calculated using the program ImageJ in its option MultiMeasure. Thirty measurements were made of at least two different images of each sample; then, the average and standard deviation were calculated using the option summarize. Similarly, the porosity of the samples was calculated with the same program under its threshold option. The data obtained was plotted using Minitab17.

2.8 Fourier transform infrared spectroscopy (FTIR)

ATR-Thermo Scientific Nicolet 6700 was used to determine the infrared spectra by the Fourier transform, measuring the attenuated total reflectance in the 4000–400 cm⁻¹ range. The spectra found were analyzed with OMNIC software.

2.9 Thermal behavior analysis (TGA and DSC)

2.9.1 Thermogravimetric analysis (TGA). TGA was performed by measuring the change in mass by increasing the temperature. The test was carried out using a heating rate of 10 °C min⁻¹ from room temperature (± 20 °C) to 700 °C in an unsealed platinum sampler, using nitrogen as a carrier gas with a flow of 20 mL min⁻¹. The equipment used was a Shimadzu model TGA-Q5500. The mass of the samples analyzed varied between 5 to 10 mg. This technique allowed us to determine the temperature at which thermal degradation begins (T_{onset}) and the change in mass per temperature increase. The curves derived from TGA (DRTG) were generated to identify the maximum degradation temperature ($T_{deg max}$). In addition, TA instruments software was used to determine the temperatures.

2.9.2 Differential scanning calorimetry (DSC). Samples were characterized using DSC equipment Shimadzu model DSC-Q100, programmed to heat from room temperature ($\pm 20 \circ$ C) to 400 °C at a heat rate of 10 °C min⁻¹ (first run), with a carrier gas flow of 20 mL min⁻¹. The mass of the samples analyzed varied between 5 to 10 mg. After the run, the

furnace was cooled with liquid nitrogen to reach a temperature of −20 °C TA instrumentsTM software was used for the identification of the temperatures.

This analysis determined the glass transition temperature $(T_{\rm m})$ and the degradation temperature $(T_{\rm d})$. The samples were evaluated on a TA Instruments DSC Q5000 equipment in unsealed aluminum trays, programmed to heat from room temperature (± 20 °C) to 300 °C at a heat rate of 10 °C min⁻¹ in a nitrogen atmosphere. The thermogram was analyzed using TA Universal Analysis software.

2.10 Cyclic voltammetry (CV)

The CV technique provided information on electrochemical reactions and the redox potential of electroactive species in the fibers, using a reference electrode of silver/silver chloride (Ag/ AgCl) with a potential +0.25 V with respect to the standard hydrogen electrode (ENH) 25 °C. As a counter-electrode, copper was used, and the electrolytic solution was a 0.1 M. acetonitrile HPLC (CH₃CN) and tetrafluoroborate tetrabutylammonium (C₁₆H₃₆BF₄N). Measurements were made by integrating nanofiber samples into a gold working electrode.

2.11 Current-voltage (IV) curves

Current-voltage (IV) curves were obtained inside a Faraday cage at room temperature using a Source Measure Unit Keithley SourceMeter 2450. The sample contacts were prepared with silver paint (PELCO[®] Colloidal Silver Paste Product No. 16032). The current was measured while applying a -10 to 10 V sweep. The following equation was used to obtain the conductivity σ :

$$\sigma = \frac{L}{RA} \tag{1}$$

where R is the ohmic resistance (R = V/I) calculated from IV curves (eqn (1)). A and L are the sample area and thickness, respectively.

3 Results and discussions

Synthesis and characterization of Ru-1 and Ru-2 3.1 complexes

Compounds Ru-1 and Ru-2 were synthesized according to eqn (2) and fully characterized by ¹H NMR, ¹³ C NMR, HRMS, UV-Vis, and FT-IR. Peak assignments were done with 2D NMR Techniques COSY, HSQC, and HMBC (Electronic Supplementary Information, ESI[†]). Compounds are cationic with an octahedral geometry around the metal center, as expected for analogous pcymene Ru(II) complexes.³¹ Compounds are very soluble in most organic solvents.



3.2 Nuclear magnetic resonance (NMR)

As expected, the ¹H NMR spectra of both complexes are very similar (Fig. 3 and 4); the principal difference is the number of signals, owing to the difference in the symmetry of the free ligands, which leads in the case of Ru-2, to a chiral metal center. In general, the signals suffer a downfield shift upon coordination compared with the free phenanthroline ligands (Fig. 3 and 4). The multiplicity of the *p*-cymene ring is also a striking difference; for Ru-1, the signals retain an A2B2 system (as observed in the starting product; Fig. S1[†]), but for Ru-2, an AA'B₂ system can be observed due to non-equivalent-isopropyl protons owing to the chiral environment. Of interest, in Ru-2, the amine peak (NH_2) moves Δ 0.85 ppm to low field, from 6.14 ppm in the free ligand to 6.98 ppm, due to coordination to the metallic center (Fig. 4).

In ¹³C NMR (Fig. S2[†]), for Ru-1, the most characteristic signals are those corresponding to (C=N)- at 156 ppm, the peaks assigned to the $-CH (CH_3)_2$ of the *p*-cymene ring at 21.6 and 30.3 ppm, and -CH₃ at 18.1 ppm. The signals for the aromatic protons at 83.5 ppm and 85.8 ppm and the aromatic ipso carbons of the p-cymene at 103.97 and 102.68 (Fig. S2⁺). The ESI[†] includes the complete assignment of signals for the Ru-1 complex (Fig. S2 and S3[†]), using two-dimensional techniques, COSY (Fig. S4-S6[†]), HSQC (Fig. S7-S9[†]), and HMBC (Fig. S10-S12[†]).

In ¹³C NMR (Fig. S15, ESI[†]), the chiral nature of the complex Ru-2 is corroborated. Two signals corresponding to the inequivalent (C=N) of the 5-amino-1,10-phenanthroline are seen at 155.6 ppm and 150.2 ppm, and four signals corresponding to the aromatic carbons of the *p*-cymene fragment are observed



Fig. 3 Comparison of ¹H RMN of Ru-1 (red) complex and free ligand phen (blue) in DMSO d_{6} .



Fig. 4 Comparison of ¹H NMR of Ru-2 (red) and free ligand (blue) in DMSOd₆.

between 83.5 ppm and 86.11 ppm. The isopropyl groups are diasterotopic, with two signals at 21.5 and 21.54 ppm. The ESI[†] includes the complete assignment of the signals of the Ru-2 complex (Fig. S15 and S16[†]) using two-dimensional techniques, COSY (Fig. S17 and S18[†]), HSQC (Fig. S19 and S20[†]), and HMBC (Fig. S21 and S22[†]).

3.3 UV-vis spectroscopy

The UV-Vis spectra of the Ru(II) complexes can be seen in Fig. 5; for the free phenanthroline ligand, a single peak can be observed at 271 nm (Fig. 5A), attributed to the π - π *-type transitions of the aromatic ring, for complex Ru-1 this band undergoes a bathochromic shift at 302 nm, upon complex formation.³² Also, a new band at 317 nm for Ru-1 can be seen, which can be attributed to the d-d* type transitions for the metal. Moreover, the complex is hyperchromic compared to the free ligand.33,34

The UV-vis spectrum of the 5-amino-phenanthroline ligand (Fig. 5B) shows peaks at 247 nm, 275 nm, and 328 nm, attributed to the n- π^* , π - π^* aromatic transitions of the aromatic ring and to the presence of auxochrome NH₂ in phenanthroline; for Ru-2 these bands suffer a bathochromic shift to 270 nm, 297 nm, and 344 nm when the complex is formed. Fig. 5B shows a new band at 437 nm, which can be attributed to the d-d* type transitions for the metal. As observed earlier, the complex is hyperchromic compared to the free ligand.³²⁻³⁴

3.4 IR spectroscopy

The characteristic v(N=C) bands for phenanthroline (phen) are found at 1691 and 1626 cm⁻¹ (Fig. S25[†]), and for 5-aminephenanthroline (5-phen) at 1634 and 1612 cm^{-1} (Fig. S26†); additional bands between 1592 and 1423 cm⁻¹ can be attributed to the stretching bands of the aromatic rings; for 5-aminephenanthroline (5-phen), bands at 3418 cm⁻¹ and 3315 cm⁻¹



FTIR spectroscopy of PCL and PCL/Ru-1 fibers (wave no. = cm⁻¹). Fig. 6

corresponding to the symmetrical and asymmetric stretching frequencies of the amino group NH₂ can be observed. These vibrations move 4 cm⁻¹ for phen (Fig. S27†) and 9 cm⁻¹ for 5phen (Fig. S28[†]) upon coordination to the metal center, as seen in similar phenanthroline complexes.35-37

Fig. 6 compares the different FTIR spectra of the PCL and PCL/Ru-1 fibers. An intense signal between 1722 and 1723 cm⁻¹ can be found in all the samples, characteristic of the carbonyl group of PCL,³⁸⁻⁴² ν (CH) bands between 2935 to 2940 cm⁻¹, which corresponds to the alkyl groups, can also be observed. In addition, multiple signals can be observed between 1168 and 1172 cm⁻¹, corresponding to the alkyl ester group with symmetrical and asymmetric stretching. Finally, the small signal of 728 cm⁻¹ corresponds to methyl chains (CH₂), part of the basic polymer skeleton. However, the latter does not provide evidence of the presence of the **Ru-1** complex since the ν (N=C) bands at 1632 and 1511 cm⁻¹ (Fig. S27[†]), corresponding to the complex, are not observed. The absence of these signals may be due to the low proportion of the complex in the fibers (Table 1).

In the case of the PCL/Ru-2 fibers (Fig. 7), the signal for the amine group (NH₂) corresponding to the Ru-2 complex is absent



Fig. 5 (A) Comparison of the UV-vis spectrum for Ru-1 complex (blue) and free ligand (red) in DMSO. (B) UV-vis spectrum comparison for complex Ru-2 (blue) and free ligand (red) in DMSO.



Fig. 7 FTIR spectroscopy of PCL and PCL/Ru-2 fibers (wave no. = cm⁻¹).

in the polymer fibers (Fig. S28†). As observed (*vide supra*), the signals for the PCL/Ru-2 samples mainly correspond to the PCL control. The signals between 2939 to 2941 cm⁻¹ corresponding to the vibration of the alkyl groups are a little more pronounced in the fibers with a higher concentration of Ru(II) complex (0.5 and 1.0%); compared to the PCL control spectrum, these bands generally move slightly to higher wave numbers (2940 2941 cm⁻¹ *vs.* 2935–2939 cm⁻¹).

These signals, as before, do not provide evidence of the presence of the **Ru-2** complex, possibly due to the low proportion of the complex in the fibers (Table 1).

3.4.1 Electrospinning of PCL. Poly(caprolactone) is a highly versatile material widely used in medical applications due to its biocompatibility and gradual biodegradation. As electrospun nanofibers, these materials possess unique structural features that make them an excellent choice for solar cells, such as a large surface area, small pore diameter, and high porosity.⁹

Various factors influence the formation of PCL fibers in electrospinning, including solvent composition, electrolyte parameters, and polymer concentration. Preventing bulb formation is crucial for consistent, high-quality fiber production. Chloroform is the favoured solvent for electrospinning PCL because it produces uniform fibers without bulbs. Other solvents, such as formic acid and acetic acid, were also studied, including binary solvents, such as formic acid/chloroform. Still, they resulted in the formation of bulbs.⁴³

A study conducted by the Enis group⁴⁴ in 2016 revealed that a combination of formic acid and chloroform resulted in fibers of around 500 nm with bulbs. However, further research discovered that a binary formic acid and acetic acid solvent produced fibers at the nanometric scale more effectively. Our previous study found that a concentration of 10% chloroform was optimal for creating polymeric fibers.⁴⁵ We also examined various electrospinning parameters and concluded that a fixed power of 20 kV and a constant flow rate of 0.450 mL h⁻¹ using a 22 G × 40 mm needle at a distance of 10 cm from the collector led to successful fiber formation.⁴⁶

3.4.2 Scanning electron microscopy (SEM). PCL has been widely used as a base polymer to manufacture electrospun fibers.⁴³ For example, when mixed with chitosan, the resulting fibers diameters are \sim 400 nm;⁴⁷ with zinc oxide (ZnO), fibers were between 511–1019 nm,⁴⁸ and with reduced graphene oxide fibers diameters range from 380–410 nm,⁴⁹ amongst others.

Fig. 8 shows the morphology and fiber diameter distribution of the PCLc, PCL/Ru-1, and PCL-Ru-2 fibers from this study. PCL control fibers (PCLc) are randomly distributed in the collector (Fig. 8A). While not uniformly sized, there are no prominent bulbous formations; the fibers generally display varying thicknesses, with some areas thicker than others. Some areas may show signs of swelling; although bulbs exist, they aren't numerous. Additionally, very thin and extensive fibers are present, resulting in a diversity of diameters. Overall, the structure forms a three-dimensional scaffold with fibers arranged haphazardly. Upon closer examination using SEM (Fig. 8A), the surface of the PCL fibers appears smooth and continuous, displaying some irregularities but lacking noticeable porosity. The average fiber diameter for the PCLc sample is $2.47 \pm 0.68 \ \mu m$ with a higher distribution between 2.35–3.33μm, as shown in Table 2 and Fig. 8A. A random arrangement in the collector is observed for all ruthenium fibers (Fig. 8B-F).

Table 2 summarizes the average diameters and porosity percentages of each sample. For most of the PCL/Ru-1 0.1%,



Fig. 8 SEM micrographies of the nanofibers. (A) PCL control (1500 x), (B) PCL/Ru-1 0.1% (1500 x), (C) PCL/Ru-1 1.0% (1500 x), (D) PCL/Ru-2 0.1% (1500 x), (E) PCL/Ru-2 0.5% (1500 x), (F) PCL/Ru-2 1.0% (1500 x).

Table 2 Average fiber diameter and percentage of porosity⁴

Sample	Average fiber diameter $(\bar{x} \pm \sigma) (\mu m)$	Percentage of porosity (%)	Higher fiber diameter distribution (μm)	
PCLc	2.47 ± 0.68	57.12	2.35-3.33	
PCL/Ru-1 0.1%	0.92 ± 0.27	26.47	0.57-0.87	
PCL/Ru-1 1.0%	0.58 ± 0.24	26.01	0.56-0.84	
PCL/Ru-2 0.1%	1.05 ± 0.17	39.45	0.92-1.15	
PCL/Ru-2 0.5%	2.14 ± 0.44	40.96	1.77-2.27	
PCL/Ru-2 1.0%	0.87 ± 0.23	48.43	0.63-0.88	

PCL/Ru-1 1.0%, PCL/Ru-2 0.1%, and PCL/Ru-2 1.0%, a decrease in thickness can be observed so fibers no longer appear swollen. A three-dimensional structure was obtained, which integrates homogeneous thin fibers. There is no presence of bulbs or artifacts in the scaffolding. The average diameter of the PCL/Ru-1 0.1% sample fibers is 0.92 \pm 0.27 μm with a higher distribution between 0.57–0.87 µm, which is thinner than the PCL control fibers. In general, the average diameter of all the fibers is thinner than the PCL control, except for PCL/Ru-2 0.5% fibers with a diameter of 2.14 \pm 0.44 μ m, mainly concentrated between 1.77-2.27 µm. Nevertheless, the fiber diameter is thicker for Ru-2 than for Ru-1 complex fibers, potentially attributed to the presence of the amine group within the Ru-2 complex. Despite this difference, the overall structure maintains a three-dimensional arrangement that combines slender and broader fibers without any observable presence of protrusions or abnormalities within the scaffold. The observed residues (Fig. 8E) are most likely the Ru-2 complex not embedded in the fiber, thus remaining on the surface of the electrospun fibers, giving the thickest diameter of all the fibers (Table 2). The latter is a convenient feature for this work, whose objective is that the complex can be exposed to photons and thus capture them and promote the transfer of electrons in said 3D network.

As mentioned earlier, the thickest fibers were PCL/Ru-2 0.5%, with an average diameter of 2.14 \pm 0.44 µm (higher distribution 1.77–2.27 µm), and the thinnest fibers were PCL/Ru-1 0.1%, with a diameter of 0.58 \pm 0.24 µm (higher distribution 0.56–0.84 µm) (Table 2). The PCLc samples have the highest percentage of porosity with 57.12%, while the lowest percentage corresponds to PCL/Ru-1 1% and PCL/Ru-1 0.1% fibers with 26.01% and 26.47%, respectively. The rest of the samples are in a range of ~45%. The fiber diameter and % of porosity does not have a linear relationship. Albeit, in general, for **Ru-2** fibers, the % of porosity increases as the concentration of **Ru-2** complex increases in the fiber. Furthermore, the % of porosity for **Ru-2** > **Ru-1** nanofibers. The presence of the amine group is possibly responsible for the increase in fiber diameter (thickness) and the percentage of porosity.

For DSSC applications, various fiber diameters have been described. For instance, a study reported carbon fibers with a diameter of 230 μ m, which are much thicker than the fibers discussed in this research and were suggested to form part of the photoanode.⁵⁰ Conversely, thinner poly (methyl methacrylate) (PMMA) fibers have also been reported for CSST solar cells, which have an average diameter of 350 nm.⁵¹

3.4.3 Thermogravimetric analysis (TGA) and differential scanning calorimetry (DSC). The thermogravimetric analysis (Table 3) provides information on the thermal stability of PCL and PCL/Ru-1 fibers. PCL fibers lose 10% of their mass at 374 ° C, and PCL/Ru-1 0.1% fibers lose 6% at 347 °C, while PCL/Ru-1, 1.0% fibers, lose 10% of their mass at 344 °C (Fig. 9A). For PCL/Ru-2 samples, PCL fibers lose 8% of their mass at 373 °C, and PCL/Ru-2 0.1% fibers lose 15% at 365 °C, compared to PCL/Ru-2 1% fibers which lose 19% of their mass at 361 °C (Fig. 10A). Finally, PCL/Ru-2 1.0% fibers lose 17% of their mass at 353 °C.

Table 3 TGA analysis of PLC and PLC/Ru-1 and R-2 nanofibers Sample Initial degradation Critical degradation 374 °C, 90% PCL 435 °C, 11% 344 °C, 90% 390 °C, 14% Ru-1 1% 347 °C, 94% 416 °C, 9% Ru-1 0.1% 373 °C. 92% 435 °C. 11% PCL Ru-2 1% 361 °C, 81% 416 °C, 17% 353 °C, 83% 422 °C, 2% Ru-2 0.5% Ru-2 0.1% 365 °C, 85% 424 °C, 0.01%



Fig. 9 (A) TGA thermograph of PCL and PCL/Ru-1 fibers, (B) DSC thermograph of PCL and PCL/Ru-1 fibers.



Fig. 10 (A) TGA thermograph of PCL and PCL/Ru-2 fibers, (B) DSC thermograph of PCL and PCL/Ru-2 fibers.

Concerning the initial mass loss, which corresponds to the evaporation of the solvent, the higher the % of the Ru(II) complex, the lower the temperature needed to start with the initial decomposition temperature (TDI). PCL's TDI is reported to be 378.3 °C, similar to our work ⁴⁷ (Table 3).

Furthermore, PCL fibers lost most of their mass at 435 °C, while PCL/Ru-1 0.1% fibers lost mass at 416 °C, and PCL/Ru-1 1.0% sample lost the total mass at 390 °C (Fig. 9A). PCL/Ru-2 0.1% fibers lost most of their mass at 424 °C, PCL/Ru-2 0.5% scaffolding at 422 °C, and PCL/Ru-2, 1.0% samples lost their total mass at 416 °C (Fig. 10A). These temperatures correspond to the maximum temperature (T_{max}). T_{max} for PCL is reported to be 412.2 °C, ⁵² which is close to the results obtained in this study (Fig. 9A and 10A).

In the case of the results obtained in differential scanning calorimetry (DSC), it should be considered that poly (caprolactone) is a semi-crystalline polymer with a regular structure, with a reported melting temperature (T_m) between 59 °C and 64 °C. Its glass transition temperature (T_g) is at -60 °C, which gives it high hardness due to its amorphous domains when they are in the elastic state.⁵³

In Fig. 9B and 10B, the DSC thermograms of the fibers of PCLc, PCL/Ru-1, and PCL/Ru-2 are observed, and all the samples have presented a change of temperature flux between \sim 57 to 61 °C; this temperature corresponds to $T_{\rm m}$, reported in the literature.⁵³ (Table 4). However, there are some variations in temperature between samples, with no significant difference.

The PCL and PCL/Ru-1 fibers have a decomposition or degradation temperature (T_d) of approximately 375–415 °C (Fig. 9B), while for PCL/Ru-2, it is between 407–410 °C (Fig. 10B).

Sample	Melting temperature	Decomposition temperature		
PCL	61 °C	412 °C		
Ru-1 1%	62 °C	375 °C		
Ru-1 0.1%	62 °C	385 °C		
PCL	61 °C	412 °C		
Ru-2 1%	59 °C	408 °C		
Ru-2 0.5%	58 °C	411 °C		
Ru-2 0.1%	61 °C	409 °C		

These temperatures correspond to the maximum temperature (T_{max}) of the TGA thermograms for the same samples (Table 4). The reported degradation temperature for PCL is ~380 °C.⁵⁴

It has been reported that the operating temperatures of DSSC range from -20 °C to 70 °C,^{2,55,56} so the fusion and decomposition temperatures of the samples will not affect the optimal functioning of the solar cell.

3.4.4 Cyclic voltammetry (CV). The redox pair in the voltamperogram (Fig. 11) shows a series of peaks. The cathodic peak (left) represents the reduction phenomena, and the anodic peak (right) represents the oxidation phenomena. The gray line indicates the potential of the gold electrode without fibers. Notably, PCL fibers display a more significant catalytic activity compared to the fibers with the Ru(II) complexes. The current of the anodic peak ($I_{p, a}$) increases by adding **Ru-1** 0.1%; however, it decreases by 0.16 mA by increasing the concentration to 1.0%. Likewise, the cathode peak current ($I_{p, c}$) increases when **Ru-1** 0.1% is incorporated and decreases by 0.36 mA when the concentration increases (Table 5). Although the PCL fibers display a more significant catalytic activity than the **Ru-2** sample



Fig. 11 (A) Cyclic voltamperogram of **Ru-1**, and (B) **Ru-2** nanofibers (C) voltamperogram) assembly of the reference electrolytic cell and counter-electrode, (D) equidistant positioning of electrodes, (E) working electrode configuration, (F). The gold electrode is used as the working electrode in CV tests.

Table 5 Electrochemical parameters obtained from cyclic voltammetry⁴

Data	Ru-1 1.0%	Ru-1 0.1%	Ru-2 1.0%	Ru-2 0.5%	Ru-2 0.1%	PCL _{Ru-1}	PCL _{Ru-2}
$I_{\rm p, a}$ (mA)	9.34	9.50	10.60	6.70	5.38	6.14	6.17
$I_{\rm p, c}$ (mA)	8.38	8.74	8.47	7.15	5.63	3.29	2.90
$E_{\rm p,a}(V)$	0.870	0.870	1.012	0.818	0.916	0.928	0.930
$E_{\rm p, c}(V)$	-0.753	-0.849	-0.940	-0.812	-0.958	-0.955	-0.956
$E_{p/2,a}(V)$	0.348	0.390	0.437	0.384	0.377	0.928	0.031

^{*a*} $I_{p, a}$: Current of anodic peak. $I_{p, c}$: Cathode peak current. $E_{p, a}$: Anodic peak potential. $E_{p, c}$: Cathodic peak potential.

at 0.1 wt, increasing the **Ru-2** concentration beyond 0.5 wt% leads to an elevation in the anodic peak current, promoting oxidation processes on the surface of the **Ru-2** fibers (Fig. 11).

Catalytic activity is observed from the binding of the PCL sample, even without the presence of the **Ru-1** compound. The current density of the anodic peak ($I_{p, a}$) increases by adding **Ru-1** 0.1%; however, it decreases by 0.16 mA by increasing the concentration to 1.0%. Likewise, the cathode peak current density ($I_{p, c}$) increases when **Ru-1** 0.1% is incorporated and decreases by 0.36 mA when the concentration increases (Table 5).

Electrofiber voltammetry tests lead to understanding and visualizing the potential of these materials in redox processes.55 These electron exchange processes are of great importance in manufacturing power generation devices. The reference voltammogram for both the pure Au electrode and the electrode covered with the PCL electrospun fiber allows us to evaluate the effect and role of the two Ru species in oxidation and reduction.⁵⁶ The voltammograms show us that the fibers based on Ru-1 and Ru-2 complexes give rise to a similar oxidation and reduction process. The incorporation of Ru-1 at 0.1% favours the anodic processes in the different electrocatalytic sites arranged in the PCL scaffolds. It compares to the performance of the electrospun fibers with the Ru-2 compound, specifically at 1%; it reaches a higher value of anode current intensity, indicating that the arrangements between the active centers of Ru and the PCL scaffolds encourage electronic transfer. The behaviour observed in the CV of PCL is indicative that the fibers undergo a non-reversible oxidation-reduction process, considering the anodic and cathodic current intensity. While the incorporation of Ru-1 and Ru-2 complexes favours reversibility, considering the values of the current intensities of the anodic and cathodic peaks, they are almost comparable. All these phenomena are associated with the Ru compounds' structure; the fact that the Ru-2 complex contains an amino group generates an electronic delocalization effect that improves charge transfer in redox processes.57-59

3.4.5 Nanofiber conductivity. The term conductive nanofiber was adopted due to the wide variety of applications in which nanofibers are used to support conductive coatings or nanostructures. In principle, by functionalizing the nanofibers, their conductivity is improved.⁵⁵ However, some polymeric structures with great industrial potential, such as PCL, are electrical insulators, and enhancing their conductivity remains one of their main challenges.

Fig. 12 shows the I–V curves taken for **PCL-Ru1** and **PCL-Ru2** nanofibers. The nanofibers were electrically contacted with silver paint, and measurements were made on one-inch-long samples. The curves were taken from -10 to 10 V, and an ohmic contact was observed. Therefore, it was possible to extract the conductivity through the geometric relationship:

$$\sigma = \frac{L}{RA}$$

where *L* is the sample's length, *A* is the cross-sectional area, and *R* is the ohmic resistance (R = V/I).

Fig. 12a shows the *I–V* curves taken for **PCL-Ru1** and **PCL-Ru2** nanofibers, where the slope is the inverse of the resistance. The steeper the slope, the higher the conductivity. The *I–V* curves show that the 1% PCL-Ru2 nanofiber has higher conductivity with the same load percentages as the **PCL-Ru1** nanofiber.

Based on the *I–V* curves shown in Fig. 12a, the behaviour appears similar at a low concentration of 0.1% of the Ru(II) complexes. However, as the amount of Ru(II) complex increases, so does the conductivity. While Ru-1 1% and Ru-2 0.5% show similar curves, the **Ru-2** 1% nanofiber demonstrates the highest conductivity. Fig. 12b displays the calculated conductivity, which reveals an increase in conductivity with an increase in the concentration of **Ru-1** and **Ru-2** complexes in the PCL nanofiber, regardless of structure. Notably, PCL with **Ru-2** complex exhibits higher conductivities than **Ru-1**; for instance, the conductivity value of 0.5% in **Ru-2** is slightly higher than 1% in **Ru-1**.

The obtained range of conductivity values falls between 0.102 and 0.461 μ S cm⁻¹. Previous studies have reported similar conductivities for PCL with 12% LiBF₄ (0.10 μ S cm⁻¹)⁶⁰ and PCL with 20% LiSCN (0.443 μ S cm⁻¹).⁶¹ Our results suggest that it is possible to increase the conductivity of PCL with lower concentrations using the discussed Ru(π) complexes.



Fig. 12 (a) I-V Curves and (b) conductivity.



Fig. 13 Possible interaction between the PCL polymer with (A) Ru(n) complexes, (B) Ru-2 complex.

The increase in the nanofibers' conductivity by increasing the concentration of the **Ru-1** and **Ru-2** complexes is why the cathodic and anodic currents increase in the cyclic voltammetry curves. Ruthenium(π) complexes enhanced the charge transfer with the electrolyte.

3.4.6 Interaction of Ru(π) complexes (Ru-1 and Ru-2) with PCL fibers. A possible interaction between the Ru(π) complexes with PCL is depicted in Fig. 13. In this interaction, the coordinated chloride ligand (Cl⁻) is displaced by the carbonyl group of the poly (caprolactone) polymer, which coordinates through the lone pair on the carbonyl moiety. The complex remains cationic, with a + 2 charge on the metallic center and an 18-electron count. In Fig. 13b, the interaction occurs through hydrogen bonding between the amino group and the carbonyl of PCL, but this is only possible for **Ru-2**, which may explain the observed differences in behavior.

4 Conclusions

In this work, we successfully utilized the electrospinning technique to create poly (caprolactone) microfibers that were functionalized with Ru(II) complexes. The electrochemical properties of these fibers were analyzed to assess their potential as a counter electrode in DSSCs. These fibers have a uniform morphology, with no bulbs but visible solid inlays on the surface. They range in size from approximately 10 to 25 µm and with porosity levels of \sim 45%. While FTIR spectroscopy did not provide evidence of Ru(II) compound incorporation into the polymer fibers, TGA and DSC thermograms indicated minor temperature variations, demonstrating their incorporation. Moreover, the fibers' melting and degradation temperature are suitable for use in DSSC. The incorporation of ruthenium complex into PCL fibers, along with the addition of the NH₂ group in Ru-2, resulted in a higher current density for both anodic and cathodic peaks. Notably, PCL-Ru2 1% fibers demonstrated a conductivity of 0.461 μ S cm⁻¹, comparable to other PCL fibers with a higher metal load. The latter are the desired characteristics of a counter electrode material. Therefore, PCL-Ru2 fibers appear suitable for evaluation as a counter electrode in DSSC. No direct relation between the morphology of the nanofibers and the increase in the material's conductivity is observed. The effect appears to be caused by the presence of the ruthenium and the amino group that generates a charge delocalization effect. More experiments are underway to assess the latter and increase the concentration of the Ru-2 complex in the nanofibers.

Conflicts of interest

There are no conflicts to declare.

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