



 Cite this: *RSC Adv.*, 2023, 13, 9055

# Preparation of a single and reusable biopolymer-based film for the extraction and preconcentration of anti-inflammatory drugs from environmental water samples†

 Maite V. Aguinaga Martínez, Claudia E. Domini\* and Carolina C. Acebal \*

One of the main goals of green chemistry is to reduce the use of toxic materials and the generation of hazardous waste, both during method development and in the synthesis of the materials used. Thus, a biodegradable, single and reusable material composed of agarose and multi-walled carbon nanotubes was proposed. The film preparation was carefully optimized in order to obtain a one-piece sorbent, with high extraction efficiency and the possibility of reuse. The film was tested in the simultaneous extraction and preconcentration of three non-steroidal anti-inflammatory drugs (ketorolac, ketoprofen and piroxicam) from environmental water samples. The optimal extraction parameters were as follows: isopropyl alcohol as the activation solvent, a sample pH value of 3.0, extraction time of 30 min, 2.00 mL of acetonitrile as the eluent, an elution time of 5 minutes, and a sample volume of 250.00 mL. Under these conditions, the film was reusable 50 times without losing its extraction capacity significantly. HPLC with a photodiode array detector was used for the separation and determination. The method presented a linear range between 0.10 and 1.2  $\mu\text{g L}^{-1}$ , good sensitivity with limits of detection between 0.0075 and 0.0089  $\mu\text{g L}^{-1}$ , and quantification between 0.025 and 0.030  $\mu\text{g L}^{-1}$ . In addition, low RSD values (0.46–3.13%) were obtained demonstrating satisfactory precision. Stream water samples were analyzed, and recoveries between 82.0 and 109.0% were obtained.

 Received 9th January 2023  
 Accepted 10th March 2023

DOI: 10.1039/d3ra00166k

[rsc.li/rsc-advances](https://rsc.li/rsc-advances)

## 1. Introduction

Solid-phase extraction (SPE) is probably the most frequently adopted technique used for the extraction and preconcentration of pollutants in environmental samples. In comparison with the conventional liquid–liquid extraction (LLE) technique, it is simpler and consumes lower amounts of organic solvents.<sup>1</sup> In addition, a wide variety of sorbents of different selectivity allows its application for the extraction of several organic compounds.<sup>2</sup> Sorbent materials have an important function in obtaining high extraction yields and high enrichment factors of analytes.<sup>3</sup> In recent years, the development of new and better materials with higher extraction capacity and improved mechanical or physicochemical stability, compared to traditional sorbents, has drawn attention.<sup>4</sup> On the other hand, there is a significant tendency to improve the sustainability of the sample preparation procedure.<sup>5</sup> Particularly, biopolymers that are obtained from biological macromolecules have been used to replace chemically synthesized sorbents. These materials

present the advantage of being highly renewable, biodegradable and non-toxic.<sup>6</sup>

Agarose is a natural polymer obtained from seaweed formed by repeated units of 1,3- $\beta$ -D-galactose and 3,6-anhydro- $\alpha$ -L-galactose.<sup>7,8</sup> It is considered as a green material because of its biodegradable nature<sup>9</sup> and its gelling capacity that facilitates the fabrication of film-based layers. In addition, the gel structure of agarose can be conveniently modified by adding different materials while the gelation process occurs, obtaining interesting hybrid materials that are suitable for extraction processes.<sup>6</sup> In this sense, agarose was mainly combined with well-established sorbents as multi-walled carbon nanotubes (MWCNTs) and C18 to perform the extraction of different pollutants from water samples and some beverages.<sup>9–12</sup>

Non-steroidal anti-inflammatory drugs (NSAIDs) belong to the group of the most widely used pharmaceuticals, showing analgesic and anti-inflammatory effects.<sup>13</sup> Although the Food and Drug Administration (FDA) has a guidance on the safe disposal of pharmaceuticals,<sup>14</sup> this information is unknown for the majority of the population. Thus, NSAIDs are discarded and can enter into the environment through domestic, industrial and hospital wastewaters since the existing water treatment plants are not designed to entirely remove them.<sup>15</sup> They present high solubility in water, relatively low environmental stability

*INQUISUR, Departamento de Química, Universidad Nacional del Sur (UNS)-CONICET, Av. Alem 1253, Bahía Blanca, 8000, Argentina. E-mail: cdomini@criba.edu.ar; cacebal@uns.edu.ar*

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d3ra00166k>



and can be biodegraded.<sup>16</sup> However, this is offset by its continued introduction into surface waters.<sup>17</sup> Different studies have demonstrated that NSAIDs can produce toxic effects on living organism in the aquatic environment, such as fish and invertebrates, even at low concentration levels.<sup>18</sup> Thus, its quantification in different water sources is of great interest. Particularly, some authors were able to determine NSAIDs of massive consumption like ibuprofen and diclofenac found in urbanized areas of Argentina, specially from cities located in the province of Buenos Aires, Córdoba and La Pampa.<sup>19,20</sup>

Several materials have been exploited as sorbents to perform the SPE of NSAIDs from environmental water samples. In most methods, commercial sorbents, such as C18,<sup>21</sup> Strata X cartridges<sup>22,23</sup> and Oasis HLB<sup>24,25</sup> have been used. Among new materials, metal-organic frameworks (MOFs) and MWCNTs have also been proposed to extract NSAIDs in these matrixes.<sup>26-29</sup> In addition, core-shell polyaniline/polyacrylonitrile nanofibers<sup>3</sup> and polyethyleneimine-600 decorated magnetic microporous organic network nanosphere<sup>30</sup> have been synthesized for extracting some of these analytes with good results. Concerning renewable materials, Ibrahim *et al.* proposed a MWCNTs-agarose-chitosan composite to perform the solid-phase microextraction of naproxen, diclofenac and mefenamic acid by using non-reusable pieces of the film inserted in a hypodermic needle.<sup>31</sup>

In this work, a film composed of agarose and MWCNTs was prepared for the simultaneous extraction and determination of ketorolac, ketoprofen and piroxicam. The new film is utilized as a single piece and it can be reused several times, resulting in a high reproducibility, less reagent use (and waste generation) and lower analysis costs. In addition, this hybrid material is potentially biodegradable since both agarose and MWCNTs are materials that can be degraded by bacteria and enzymes.<sup>32,33</sup> The film was successfully used in the extraction of the selected NSAIDs from environmental water samples of Bahia Blanca's region.

## 2. Experimental part

### 2.1 Reagents and materials

The appropriate amount of ketorolac (KEC), ketoprofen (KEN) and piroxicam (PIR) (Sigma-Aldrich, Germany) were diluted in

methanol, MeOH ( $\geq 99\%$ , Sigma-Aldrich, Germany), to prepare 200 mg L<sup>-1</sup> stock solutions that were kept in dark containers (4 °C). Working solutions were prepared daily by diluting a suitable volume of the stock solutions with an acetic acid (Sintorgan, Argentina) solution (pH 3.0). Agarose (BioReagent, for molecular biology, low EEO) was bought from Sigma-Aldrich (Germany). Acetonitrile, ACN ( $\geq 99\%$ , Sigma-Aldrich, Germany) and formic acid (Sintorgan, Argentina) were used for preparation of the mobile phase. Isopropyl alcohol, IPA ( $\geq 99.8\%$ ) was purchased from Dorwil (Argentina). The MWCNTs (13–16 nm, purity >95%) were supplied by Bayer®.

### 2.2 Instrumentation

The HPLC separation was carried out on a HPLC Waters 600 (Waters, USA) combined with a photodiode array detector (PDA). The analytes were separated using a C18 Gemini column (150 mm × 4.6 mm, 5 μm, Phenomenex) under isocratic mode. The mobile phase was composed of ACN and water acidified with 0.07% of formic acid in a 50:50 v/v proportion. The column temperature, the injection volume and the flow rate were 25 °C, 10 μL and 1.0 mL min<sup>-1</sup>, respectively. The separated analytes were detected at 254 nm (KEN), 312 nm (KEC) and 326 nm (PIR).

The MWCNTs solution was dispersed in a Cole-Parmer EW-08895-16 ultrasonic bath. The MWCNTs-agarose film was dried in a laboratory oven (Model SL30S, San Jor, Argentina). A vacuum evaporator system (Speed Vac SPD11V, Thermo Fisher Scientific, Argentina) was used to evaporate the solvent.

The characterization of the agarose and the MWCNTs-agarose films, before and after drying, was performed with a Scanning Electron Microscope, SEM (LEO, EVO 40 XVP) with a secondary electron (SE) detector.

### 2.3 Preparation of the MWCNTs-agarose film

The preparation of the MWCNTs-agarose film was based on the procedure developed by Hong Loh *et al.*<sup>9</sup> with modifications (Fig. 1). Briefly, 0.1 g of agarose and 5.0 mL of water were mixed in a beaker and submerged into a water bath at 80 °C. The solution was magnetically stirred until the agarose was completely dissolved. In another glass beaker, 80 mg of MWCNTs were mixed with 5.0 mL of water and sonicated in an ultrasonic bath for 15 minutes. Then, the dispersed MWCNTs

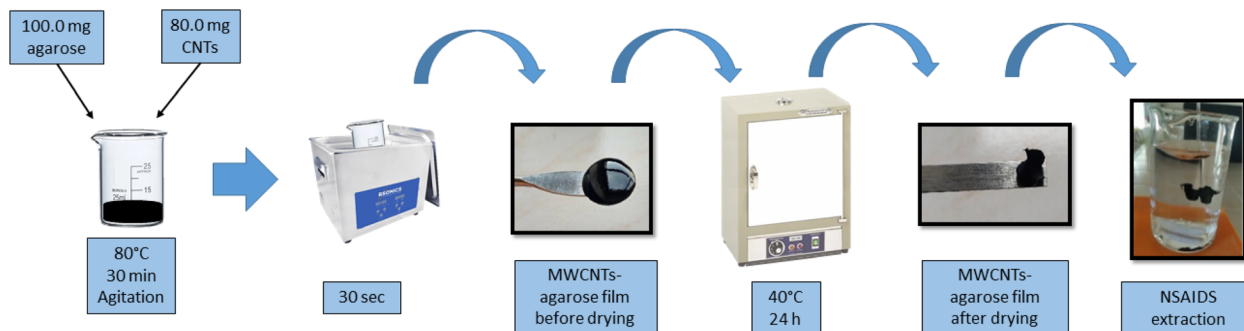


Fig. 1 Schematic representation of the preparation of the MWCNTs-agarose film procedure.



were added to the agarose solution. The final mixture was heated with magnetic stirring at 80 °C for 30 minutes in order to disperse the MWCNTs in the agarose and to achieve a final volume of 5.0 mL. The mixture was placed over a circular glass (5.0 cm diameter) and cooled at 25 °C. The obtained film was dried at 40 °C for 24 h. The images of the obtained MWCNTs-agarose film, before and after drying, are presented in the ESI (Fig. S1†).

#### 2.4 SPE procedure

The MWCNTs-agarose film was first conditioned by submerging it in IPA for 5 minutes, and then washed with distilled water. Next, the film was placed inside 250.00 mL of a standard solution or sample containing the mixture of target analytes. The pH of the standard solution or samples was 3.0. The solution was magnetically stirred and the extraction was performed for 30 minutes. Then, the film was separated from the solution and washed with water. After that, it was placed inside a glass beaker containing 2.00 mL of ACN and sonicated in an ultrasonic bath for 5 minutes for elution. The film was removed from the eluate that was evaporated to dryness using a vacuum evaporator system at 30 °C. Then, the extract was reconstituted in 600 µL of the mobile phase. The extraction blank was performed using distilled water or the sample applying the same procedure.

#### 2.5 Sampling and preparation of samples

The samples were taken from various sampling points of the Naposta stream. Two of them (1: P1 38° 40' 51.822''S 62° 14' 13.707''W; 2: P<sub>2</sub> 38°45'09.0''S 62°14'10.2''W) were situated in the city of Bahía Blanca and the other one (3: P3 38°32'00.1''S 61° 52'26.4''W) was located in the village of Cabildo (Fig. S2, ESI†). The samples were acidified and kept in the fridge (4 °C) until their analysis. Before the extraction procedure, the samples were left at 25 °C, homogenized, and filtered with an ashless filter paper (4–12 µm pore, Microclar, USA). After that, 250.00 mL aliquots were taken, acidified until pH 3.0 and extracted following the procedure mentioned in Section 2.4.

To assess the accuracy of the proposed method, a recovery study was performed. The samples were spiked with KEC, KEN and PIR at three concentration levels within the calibration range in triplicate at the beginning of the analytical process.

### 3. Results and discussion

#### 3.1 Optimization of the MWCNTs-agarose film preparation procedure

Most of the SPE approaches using biosorbents that can be found in the literature<sup>9–12,31</sup> involved the use of several films or multiple pieces of the same film. This fact can cause difficulties during the extraction procedure, especially during the washing and elution steps. Thus, the main objective of this work was to obtain a single piece sorbent with high extraction efficiency that can be reused as many times as possible. For that purpose, the optimization of the preparation process was performed. The concentration of agarose was the same in all the experiments.

At first, the MWCNTs were dispersed directly in the agarose solution. The accomplishment of a homogeneous dispersion was time consuming since the MWCNTs tended to aggregate. Thus, in order to ensure the homogeneity of the final mixture the dispersion of the MWCNTs was performed in water, assisted by ultrasound, before mixing them with the agarose solution. It was observed that the total dispersion of the MWCNTs required 15 minutes. At lower times, the dispersion was incomplete, resulting in longer times to reach a homogeneous MWCNTs-agarose mixture.

To achieve a satisfactory extraction capacity of the MWCNTs-agarose film, a concentration of MWCNTs higher than that found in the literature was tested.<sup>3,8,11</sup> Therefore, the amount of MWCNTs was evaluated in the range of 30 to 120 mg. The highest recovery values were obtained when the film was prepared with 80 mg of MWCNTs. When the amount of MWCNTs was higher than 80 mg, the film broke in the drying step.

The temperature of the agarose solution was kept in the interval between 80 °C and 85 °C to ensure the complete dissolution of the agarose. It was noticed that when the temperature dropped below 75 °C during the preparation of the film, it broke during drying. This was attributed to a more fragile structure, owing to an incomplete dissolution of the agarose, which hindered the gelation and drying processes.

Also, the MWCNTs film was tested with and without a drying step. When the film was not dry, the extraction of the analytes was not possible. Thus, the temperature and time of the drying step were evaluated. The temperature was assessed between 40 °C and 60 °C, and no difference in the film appearance and recovery values was observed. Therefore, 40 °C was selected as the drying temperature. The drying time was tested between 24 and 48 hours. Satisfactory recovery values were obtained with 24 hours of drying. At lower times, the film was not completely dry, leading to lower recovery values.

A comparison of the different procedures and formats for agarose-based sorbents are shown in Table 1 of the ESI.†

#### 3.2 Characterization of the agarose-MWCNTs films

The composite was examined by SEM to study the morphology of the MWCNTs-agarose film. The surface homogeneity of the MWCNTs in the agarose matrix was observed. Fig. 2 shows the surface of an agarose film after its drying (a), and the morphology of the MWCNTs-agarose film before (b) and after (c) its drying. As it can be seen, while the surface of the agarose film is smooth, the addition of MWCNTs generates the formation of a rough surface which facilitates the interaction between the active sites of the sorbent and the analytes and improves the mass transfer process and its adsorption capacity. Moreover, it can be observed that the surface of the film before drying is rougher than the surface of the film after drying. This fact can be confirmed by observing Fig. 2d and e that corresponds to the cross section of both films. In addition, Fig. 2e shows an adequate blend of the MWCNTs in the agarose matrix, suggesting good dispersion and adhesion.



Table 1 Analytical parameters of the proposed method for the determination of the NSAIDs

Analytes	Linear range ( $\mu\text{g L}^{-1}$ )	$R^2$	LOD <sup>a</sup> ( $\mu\text{g L}^{-1}$ )	LOQ <sup>b</sup> ( $\mu\text{g L}^{-1}$ )	RSD% intra-day <sup>c</sup>	RSD% inter-day <sup>d</sup>
KEC	0.10–1.20	0.997	0.0089	0.030	0.46	3.13
KEN	0.10–1.20	0.998	0.0075	0.025	1.20	3.08
PIR	0.10–1.20	0.998	0.0082	0.027	0.99	2.90

<sup>a</sup> LOD calculated as  $3s A^{-1}$  and. <sup>b</sup> LOQ calculated as  $10s A^{-1}$ , where,  $s$ : standard deviation,  $A$ : slope of the calibration curve. <sup>c</sup>  $n = 5$ ,  $0.6 \mu\text{g L}^{-1}$ . <sup>d</sup>  $0.6 \mu\text{g L}^{-1}$ , measure by triplicate over 3 days.

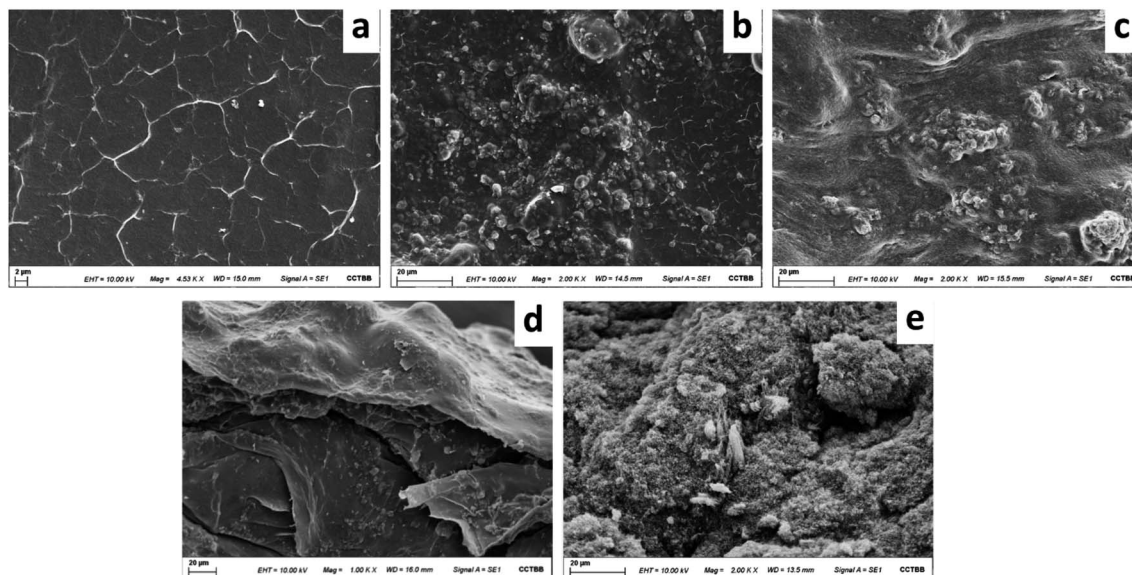


Fig. 2 (a) SEM surface morphology of the agarose film after its drying, (b) SEM surface morphology of the MWCNTs-agarose film before its drying, (c) SEM surface morphology of the MWCNTs-agarose film after its drying, (d) SEM of the cross section of the MWCNTs-agarose film before drying (e) SEM of the cross section of the MWCNTs-agarose film after drying.

### 3.3 Optimization of the extraction procedure

The extraction mechanism of the film is mainly due to the interactions between MWCNTs and the analytes through hydrophobic and  $\pi$ - $\pi$  interactions.<sup>34</sup> Particularly, the  $\pi$ - $\pi$  interactions are favored by the structure of the analytes that possess aromatic rings and cyclic molecules with conjugated double carbon-carbon bonds. To confirm that the agarose did not intervene in the extraction, the extraction procedure was performed with a film made only of this material. The results can be seen in Fig. S30 (ESI<sup>†</sup>).

Thus, in order to achieve a high extraction efficiency, different factors that can affect the interactions between the MWCNTs and the analytes were optimized, including activation solvent, extraction time, sample pH value, eluent and eluent volume, sample volume and elution time. All variables were tested for the three analytes and experiments were performed in triplicate using 50.00 mL of the standard solution with a concentration of  $0.10 \text{ mg L}^{-1}$  for the three compounds.

**3.3.1 Sample pH value.** The sample pH is a parameter that strongly influences the efficiency of the extraction, mainly for acidic and basic compounds. NSAIDs are acidic compounds

that mainly exist in their molecular form at pH values lower than their  $pK_a$  values (KEC: 3.5; KEN: 4.0; PIR: 6.3).<sup>35–37</sup> Thus, pH values in the range of 2.0–6.0 were studied by adjusting the pH with acetic acid. It was observed that when the solution pH decreased to 3.0, the extraction efficiency increased significantly for all the analytes, and kept constant at a lower pH. Therefore, pH 3.0 was selected for the next experiments.

**3.3.2 Activation solvent and extraction time.** First, the extraction procedure was performed without activation of the sorbent. As it was observed that the analytes were not extracted by the MWCNTs-agarose film, MeOH and IPA were tested as activation solvents.<sup>11</sup> Since IPA is a less polar solvent than MeOH,<sup>38</sup> the hydrophobic interactions between MWCNTs and the analytes were favored by the use of IPA, leading to higher extraction efficiency. This can be observed in Fig. S4 (ESI<sup>†</sup>) which shows the difference in the spectra between the use of both solvents as conditioning solvents. Thus, IPA was selected to perform the activation of the MWCNTs-agarose film.

On the other hand, the extraction time was studied in the interval between 10 and 40 minutes (Fig. 3a). The highest recoveries were obtained after 30 minutes of extraction. At longer extraction times, recoveries remained constant.



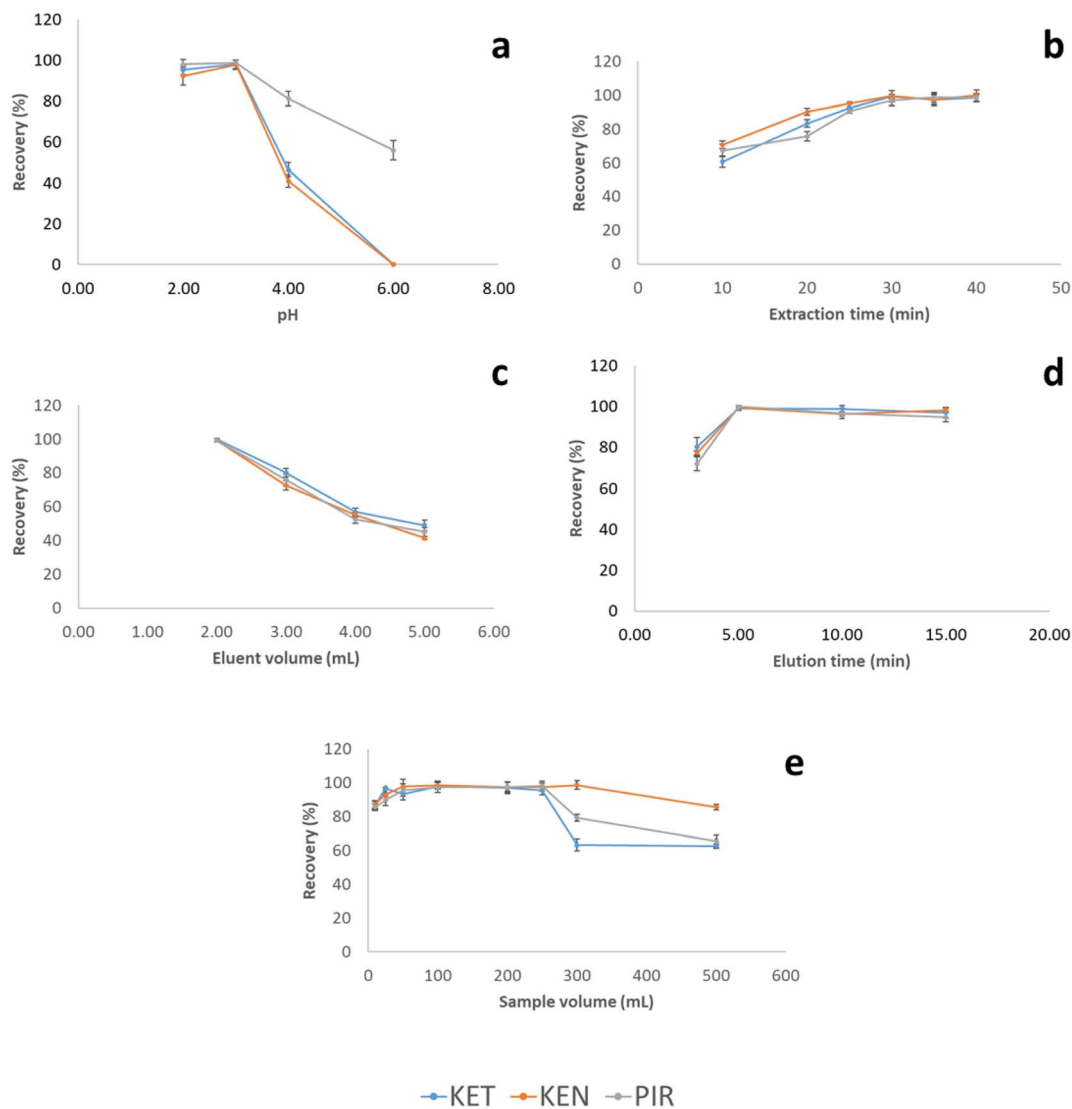


Fig. 3 Effect of the sample pH (a), extraction time (b), eluent volume (c), elution time (d) and sample volume (e) on the NSAIDs recovery.

**3.3.3 Type of eluent, volume and elution time.** MeOH and ACN were tested as possible eluents. In order to perform the optimization, the extraction protocol was followed for each analyte and elution was done with 2.0 mL of the eluent. Also,

extraction blanks were performed. Using MeOH as extractant resulted in more unstable baseline for blank than the use of ACN. Mixtures of ACN and water were also tested, observing a decrease of the recoveries of the analytes as the percentage of

Table 2 Recovery percentage values of the NSAIDs determined in surface water samples

Samples <sup>a</sup>	Added concentration ( $\mu\text{g L}^{-1}$ )	KEN	Recovery (%) KEC	PIR
Napostá river (1) <sup>b</sup>	0.30	90.0 $\pm$ 3.3	106.7 $\pm$ 6.7	97.0 $\pm$ 6.7
	0.60	85.0 $\pm$ 6.7	108.3 $\pm$ 6.7	98.3 $\pm$ 1.7
	1.10	82.7 $\pm$ 1.8	109.0 $\pm$ 1.8	99.1 $\pm$ 3.6
Napostá river (2) <sup>b</sup>	0.30	100.0 $\pm$ 13.3	106.7 $\pm$ 6.7	90.0 $\pm$ 16.7
	0.60	91.7 $\pm$ 8.3	100.0 $\pm$ 3.3	96.0 $\pm$ 8.3
	1.10	96.4 $\pm$ 5.4	99.1 $\pm$ 4.5	99.1 $\pm$ 2.7
Napostá Chico stream	0.30	103.3 $\pm$ 3.3	100.0 $\pm$ 3.3	100.0 $\pm$ 6.7
	0.60	83.3 $\pm$ 1.7	99.1 $\pm$ 3.3	82.0 $\pm$ 15.0
	1.10	102.7 $\pm$ 2.7	99.7 $\pm$ 3.6	99.1 $\pm$ 4.5

<sup>a</sup> The recoveries were calculated in triplicate. <sup>b</sup> (1) and (2) were different sampling points.



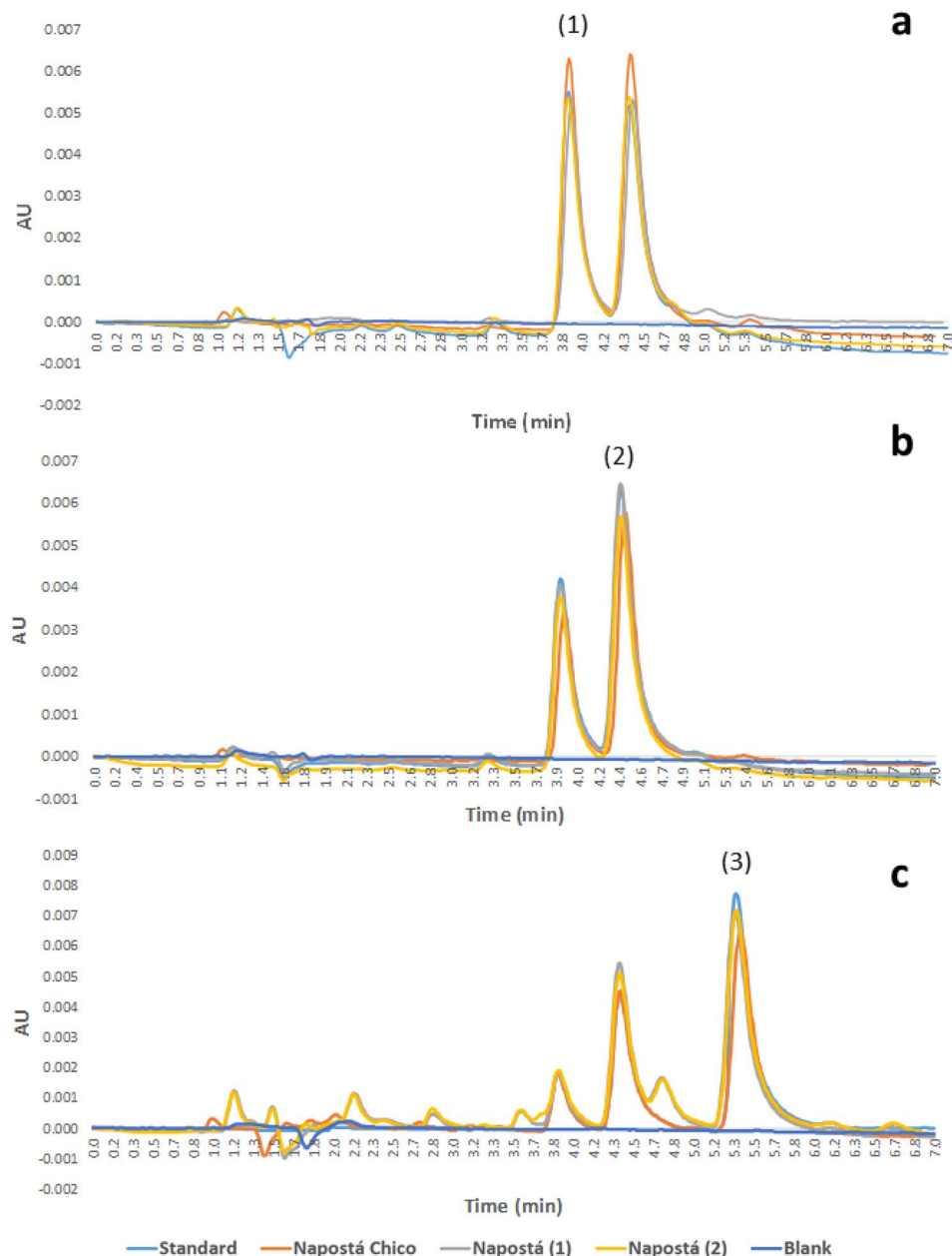


Fig. 4 Chromatograms corresponding to the standard solution and spiked water samples measured at  $\lambda = 310$  nm for KEC (a),  $\lambda = 326$  nm for PIR (b) and  $\lambda = 254$  nm for KEN (c). Peak identification: (1) KEC, (2) PIR and (3) KEN. Extraction conditions: concentration of NSAIDs,  $0.30 \mu\text{g L}^{-1}$ ; sample volume, 250.00 mL; mass of MWCNTs, 80 mg; volume of desorption solvent, 2.00 mL; sample pH, 3.0. Chromatographic conditions: 150 mm  $\times$  4.6 mm, 5  $\mu\text{m}$  particle size C18 Gemini Phenomenex column; mobile phase: ACN-water 0.07% formic acid (50 : 50 v/v); injection volume: 10  $\mu\text{L}$ ; flow rate: 1.0 mL  $\text{min}^{-1}$ .

water increased. Therefore, ACN was selected as the elution solvent.

The volume of ACN needed to elute quantitatively the analytes was studied in the range of 2.00 to 5.00 mL. 2.00 mL of ACN was the minimum volume needed to completely cover the MWCNTs-agarose film. Within this range, the volume of eluent to obtain the highest recovery value was 2.00 mL (Fig. 3b). The decrease in the recovery values above this volume can be attributed to a dilution effect. Due to the low elution volume, the elution step was assisted by ultrasound since the stirring of

the solution led to the breaking of the film. The elution time in the ultrasonic bath was tested in the range of 2 to 15 minutes. The highest recovery was achieved at 5 minutes of elution, and there was no significant improvement in the recoveries for all NSAIDs at higher times.

**3.3.4 Breakthrough volume.** Breakthrough volume data is important in order to assure that the analytes are not purged off the sorbent bed during sample loading. This value is also important to calculate the enrichment factor. Thus, the suitable volume to be loaded into the MWCNTs-agarose film was studied



Table 3 Comparison of SPE and SPE-based methods for NSAIDs' determination<sup>a</sup>

Analytes	SPE and SPE-based extraction technique	SPE sorbent	Amount of sorbent (mg)	Eluent volume (mL)	Sample volume (mL)	Sorbent reused	Detection technique	LOD ( $\mu\text{g L}^{-1}$ )	RSD (%)	Ref.
Tolfenamic acid, naproxen, clofibrac acid, diclofenac, indomethacin, ibuprofen, aspirin, ketoprofen	SPE	PANI/Pan NFsM	3	0.5	10	Yes (20 times)	UPLC-MS/MS	0.2–5.0 $\text{ng L}^{-1}$	0.4–11.1 <sup>c</sup>	3
Ketorolac	SPE	C18	1000	5.0	250	n. i.	HPLC-UV	2.83	1.8	21
Ketoprofen, naproxen, piroxicam, diclofenac, indometacin, carprofen, meclufenamic acid	SPE	Strata-X	200	6.0	200	n. i.	HPLC-Q-Orbitrap-HRMS	3.7–11.7 $\text{ng L}^{-1}$	6.2–32.7 <sup>b</sup>	22
Ketoprofen, naproxen, diclofenac, ibuprofen	SPE	Strata-X	200	6.0	200	n. i.	HPLC-Q-Orbitrap-HRMS	n. i.	4.2–56 <sup>c</sup>	23
Ketoprofen	SPE	Oasis@HLB	n. i.	n. i.	2.5	n. i.	HPLC-MS/MS	n. i.	2–20	24
Ketoprofen, naproxen, flurbiprofen, diclofenac sodium and ibuprofen	SPE	CF@UiO-66-NH <sub>2</sub>	50	0.5	20	Yes (5 times)	UPLC-PDA	0.12–3.50	2.36–8.49	27
Tolfenamic acid, diclofenac, flunixin, ibuprofen, carprofen, ketoprofen, mefenamic acid, meloxicam, naproxen, niflumic acid, vedapropfen, flufenamic acid	SPE	CNTs	20	10.0	100.0	n. i.	LC-QqQ-MS/MS	0.01–1.3 $\text{ng L}^{-1}$	2–14 <sup>d</sup>	28
1-Naphthylacetic acid, naproxen, ibuprofen, ketoprofen	MSPE	Fe <sub>3</sub> O <sub>4</sub> @MON-PEI <sub>600</sub>	4.0	0.4	10.0	Yes (8 times)	HPLC-UV	0.042–0.149	0.3–4.5 <sup>d</sup>	30
Naproxen, diclofenac sodium salt, mefenamic acid	SPME	Agarose-chitosan-MWCNTs composite film	0.6 (four discs per extraction)	0.1	10	No	HPLC-UV	0.89–8.05	1.03–4.59 <sup>c</sup>	31
Carprofen, mefenamic acid naproxen, diclofenac, ketoprofen	SB-SPE	MIL-101(Cr)/PVA	60 $\text{mg mL}^{-1}$	1.0	10.0	Yes	HPLC-MS/MS	0.011–0.093	1.67–9.52	41
Ketorolac, piroxicam, ketoprofen		Agarose-MWCNTs composite film	80	2.0	250.0	Yes (50 times)	HPLC-PDA	0.0075–0.0089	2.90–3.13	Our

<sup>a</sup> MSPE: magnetic solid phase extraction; SB-SPE: stir bar solid phase extraction. <sup>b</sup> Calculated from the samples. <sup>c</sup> Calculated from the samples at different concentration levels. <sup>d</sup> Calculated at different concentration levels.

by drowning the film into increasing volumes of the sample, in the range of 10.00 to 500.00 mL, and keeping the analytes mass constant. The experimental data is shown in Fig. 3c. As it can be observed, no significant variations of the recoveries occurred (95.5–100.0%) when the sample volume increased from 10.00 to 250.00 mL for the three analytes. After that volume, the recoveries of KEC and KEN decreased with the increase of the sample volume. Thus, 250.00 mL was selected as the optimal sample volume.

### 3.4 Reusability of the MWCNTs-agarose film

The reuse and stability of the sorbent are important factors to assess its performance. To evaluate this, the MWCNTs-agarose film was subjected to consecutive cycles of adsorption/elution under the optimized conditions. At the end of the working day, the film was washed repeatedly with ACN and distilled water, and then it was dried until reuse. The obtained recovery percentage values were satisfactory and remained approximately constant up to 50 times, demonstrating that the film was stable and reusable, without losing the extraction capacity for the target analytes.

The use of a biopolymers as supports for the sorbents is important not only because the material obtained is potentially non-toxic and biodegradable but also to avoid other types of immobilization that require organic reagents that can contaminate the environment. However, in the works that can be found in the literature, the proposed sorbents are not reusable or used few times and discarded after each sample extraction (Table S1, ESI†). This represents a great disadvantage since, although they are biodegradable materials, the preparation of different films for each extraction reduces the reproducibility of the method, increases the quantity of reagents needed and it is time-consuming.

### 3.5 Analytical performance

Table 1 shows the analytical figures of merit that were calculated to evaluate the proposed method. Calibration plots for the target compounds were built in the range of 0.10 to 1.20  $\mu\text{g L}^{-1}$  by performing the extraction of a mixture of the analytes' standard solutions. Correlation coefficients higher than 0.99 were obtained for all analytes. LOD values were 0.0089, 0.0075 and 0.0082  $\mu\text{g L}^{-1}$  for KEC, KEN and PIR, respectively, and LOQ values were 0.030, 0.025 and 0.027  $\mu\text{g L}^{-1}$  for KEC, KEN and PIR, respectively. The intra-day precision was assessed by calculating the relative standard deviation (RSD%) with the values obtained when the analysis was performed with a 0.6  $\mu\text{g L}^{-1}$  standard solution of each compound five times within a day. In order to study the inter-day precision, the solutions with the same concentration were measured by triplicate on three consecutive days. The obtained RSD values demonstrated a high precision of the extraction method.

### 3.6 Application to environmental water samples

The applicability of the MWCNTs-agarose film in the extraction of the analytes from surface water samples was evaluated. The samples were taken from different points of the Napostá stream

(sampling points 1 and 2), which provides the city of Bahía Blanca with drinking water. The sampling point 3 was collected from the Napostá chico stream. Sampling sites were taken at strategic locations in and around the city. Point 1 and point 2 were selected before entering the city and before the mouth of the stream enters the estuary, respectively. These points were chosen due to the fact that there may be clandestine sewage discharges that arrive through freshwater courses'.<sup>39</sup> On the other hand, the sampling point of the Naposta Chico stream was selected after the last town (Cabildo) that crosses the stream.

Even though the sampling points were selected taking into account a potential contamination of the water, residues of the target compounds were not found at concentrations above the quantitation limit of the method.

To evaluate the accuracy of the proposed method, a recovery study was performed. For that, the samples were spiked at the beginning of the analytical process and treated as described in Section 2.4. Table 2 shows the added concentrations (0.30, 0.60 and 1.10  $\mu\text{g L}^{-1}$ ) and the recoveries obtained for KEN, KEC and PIR, which ranged from 82.0% to 109.0%, with most of the results between 90.0 and 103.3%.

Fig. 4 shows the chromatograms of the 0.30  $\mu\text{g L}^{-1}$  standard solutions of the analytes and spiked samples at the same concentration level for comparison.

The slope of the calibration plot and the slope of the calibration plot built using a sample spiked at the same concentration levels were compared to evaluate the matrix effect.<sup>40</sup> At the calculated probability level ( $p > 0.1$ ), there was no significant difference between the slopes. Thus, it can be concluded that no matrix effect was present.

Different SPE methods that have been found in the literature for the extraction of the studied analytes are shown in Table 3. They have been compared in terms of SPE-based extraction technique, type of sorbent, SPE parameters, reusability of the sorbent, detection technique, LOD and RSD values. The use of less amount of sorbent and lower volume of organic solvent were the main advantages, together with the reusability of the MWCNTs-agarose film several times. As it can be observed, it was possible to obtain preconcentration factors comparable with most of the methods that were found in the literature. In addition, the obtained LOD value was significantly lower than the LODs of the methods that use the same detection technique (molecular absorption spectrometry). Furthermore, it was comparable with some of the methods that use mass spectrometry as detection technique. Concerning RSD, the obtained values were similar or even lower than the published methods.

## 4. Conclusions

Agarose, which is a non-toxic, renewable and biodegradable material, was used for the immobilization of MWCNTs. The MWCNTs-agarose film presented excellent sorbent properties and allowed the extraction and preconcentration of KEC, KEN and PIR, obtaining satisfactory analytical parameters. In addition, the film was reusable up to 50 times without losing its extraction capacity. Furthermore, it was possible to perform the





extraction with a single film, which notably simplified the experimental procedure. The proposed method was successfully applied for the quantification of the selected analytes in natural water samples of Bahia Blanca's region with satisfactory results. As far as we know, it is the first time that these compounds were determined in this region, which is of great importance to obtain the state of the art of their presence/absence. In this case, the analytes were not found in the selected sampling points at the detection limits that were achieved by the proposed method.

## Conflicts of interest

The authors declare no competing interests.

## Acknowledgements

Authors gratefully acknowledge to Universidad Nacional del Sur (PGI 24/Q119, PGI 24/Q099) and Comisión Nacional de Investigaciones Científicas y Técnicas (CONICET), PIP 11220200102603CO. Also, the authors would like to thank to Agencia Nacional de Promoción Científica y Tecnológica (PICT-2019-04458).

## References

- R. E. Majors, in *Handbook of Sample Preparation*, ed. Pawliszyn J. and Lord H. L., Wiley & Sons, Inc, 2010, pp. 53–79.
- A. Azzouz, S. K. Kailasa, S. S. Lee, A. J. Rascón, E. Ballesteros, M. Zhang and K. H. Kim, *Trends Anal. Chem.*, 2018, **108**, 347–369.
- N. Jian, L. Qian, C. Wang, R. Li, Q. Xu and J. Li, *J. Hazard. Mater.*, 2019, **363**, 81–89.
- J. Plotka-Wasyłka, N. Szczepańska and J. Namieśnik, *Trends Anal. Chem.*, 2016, **77**, 23–43.
- I. Pacheco-Fernández, D. W. Allgaier-Díaz, G. Mastellone, C. Cagliero, D. D. Díaz and V. Pino, *Trends Anal. Chem.*, 2020, **125**, 115839.
- H. Tabani, M. Alexović, J. Sabo and M. Ramos Payán, *Talanta*, 2021, **224**, 121892.
- M. N. H. Rozaini, N. F. Semail, B. Saad, S. Kamaruzaman, W. N. Abdullah, N. A. Rahim and N. Yahaya, *Talanta*, 2019, **199**, 522–531.
- M. K. Yazdi, A. Taghizadeh, M. Taghizadeh, F. J. Stadler, M. Farokhi, F. Mottaghitalab, P. Zarrantaj, J. D. Ramsey, F. Seidi, M. R. Saeb and M. Mozafari, *J. Controlled Release*, 2020, **326**, 523–543.
- S. Hong Loh, M. Marsin Sanagi, W. Aini Wan Ibrahim and M. Noor Hasan, *Talanta*, 2013, **106**, 200–205.
- S. Hong Loh, P. Ee Neoh, C. Tung Tai and S. Kamaruzaman, *Malaysian J. Anal. Sci.*, 2018, **22**, 1–7.
- M. Marsin Sanagi, M. Nazirul Muzakkir Mat Jais, S. Kamaruzaman, W. Aini Wan Ibrahim and U. Baig, *Anal. Methods*, 2015, **7**, 2862–2868.
- N. T. Ng, M. M. Sanagi, W. N. W. Ibrahim and W. A. W. Ibrahim, *Food Chem.*, 2017, **222**, 28–34.
- Y. Zhou, J. Xu, N. Lu, X. Wu and X. Hou, *Talanta*, 2021, **225**, 121846.
- Food and Drug Administration, *Information for Consumers and Patients, Safe disposal of medicines*, <https://www.fda.gov/drugs/ensuring-safe-use-medicine/safe-disposal-medicines>, Accessed March 7, 2023.
- A. Gogoi, P. Mazumder, V. K. Tyagi, G. T. Chaminda, A. K. An and M. Kumar, *Groundw. Sustain. Dev.*, 2018, **6**, 169–180.
- K. Świacka, J. Maculewicz, D. Kowalska, M. Caban, K. Smolarz and J. Świeżak, *J. Hazard. Mater.*, 2022, **424**, 127350.
- K. Świacka, A. Michnowska, J. Maculewicz, M. Caban and K. Smolarz, *Environ. Pollut.*, 2021, **273**, 115891.
- H. Wang, H. Xi, L. Xu, M. Jin, W. Zhao and H. Liu, *Sci. Total Environ.*, 2021, **788**, 147819.
- Y. Elorriaga, D. J. Marino, P. Carriquiriborde and A. E. Ronco, *Bull. Environ. Contam. Toxicol.*, 2013, **90**, 397–400.
- Y. Elorriaga, D. J. Marino, P. Carriquiriborde and A. E. Ronco, *Int. J. Environ. Health*, 2013, **6**(4), 330–339.
- H. H. Ortega Soto, J. J. Ramírez García, P. Gamboa Suárez and A. M. Dávila Estrada, *Int. J. Photoenergy*, 2017, 6781310.
- C. L. Chitescu, G. Kaklamanos, A. I. Nicolau and A. A. M. L. Stolker, *Sci. Total Environ.*, 2015, **532**, 501–511.
- I. Pugajeva, J. Rusko, I. Perkons and V. Bartkevics, *J. Pharm. Biomed. Anal.*, 2017, **133**, 64–74.
- J. Camilleria, R. Baudota, L. Wiesta, E. Vullieta, C. Cren-Olivéa and G. Daniele, *Int. J. Environ. Anal. Chem.*, 2015, **95**, 67–81.
- A. Kot-Wasik, A. Jakimska and M. Śliwka-Kaszyńska, *Environ. Monit. Assess.*, 2016, **188**, 661.
- A. Amiri and F. Ghaemi, *J. Chromatogr. A*, 2021, **1648**, 462168.
- Y. Gao, S. Wang, N. Zhang, X. Xu and T. Bao, *J. Chromatogr. A*, 2021, **1652**, 462349.
- I. Reinholds, I. Pugajeva, Dz. Zacs, E. Lundanes, J. Rusko, I. Perkons and V. Bartkevics, *Environ. Monit. Assess.*, 2017, **189**, 568.
- T. Wang, S. Liu, G. Gao, P. Zhao, N. Lu, X. Lun and X. Hou, *Microchim. Acta*, 2017, **184**, 2981–2990.
- X. Q. He, Y. Y. Cui and C. X. Yang, *Talanta*, 2021, **233**, 122471.
- W. N. Wan Ibrahim, M. Marsin Sanagi, N. S. Mohamad Hanapi, S. Kamaruzaman, N. Yahaya and W. A. Wan Ibrahim, *J. Sep. Sci.*, 2018, **41**, 2942–2951.
- M. Chen, X. Qin and G. Zeng, *Trends Biotechnol.*, 2017, **35**(9), 836–846.
- C. Jiang, Z. Liu, D. Cheng and X. Mao, *Biotechnol. Adv.*, 2020, **45**, 107641.
- J. Plotka-Wasyłka, N. Szczepańska, M. de La Guardia and J. Namieśnik, *TrAC Trend. Anal. Chem.*, 2016, **77**, 23–43.
- National Center for Biotechnology Information, *PubChem Compound Summary for CID 3826, Ketorolac*, 2022, <https://pubchem.ncbi.nlm.nih.gov/compound/Ketorolac>, Accessed March 7, 2023.
- National Center for Biotechnology Information, *PubChem Compound Summary for CID 3825, Ketoprofen*, <https://pubchem.ncbi.nlm.nih.gov/compound/Ketoprofen>, Accessed March 7, 2023.



- [pubchem.ncbi.nlm.nih.gov/compound/Ketoprofen](https://pubchem.ncbi.nlm.nih.gov/compound/Ketoprofen), 2022, Accessed March 7, 2023.
- 37 National Center for Biotechnology Information, *PubChem Compound Summary for CID 54676228, Piroxicam*, <https://pubchem.ncbi.nlm.nih.gov/compound/Piroxicam>, 2022, Accessed March 7, 2023.
- 38 M. Razali, C. Didaskalou, J. F. Kim, M. Babaei, E. Drioli, Y. M. Lee and G. Szekely, *ACS Appl. Mater. Interfaces*, 2017, **9**(12), 11279–11289.
- 39 M. E. Streitenberger and M. D. Baldini, *Rev. Int. Contam. Ambiental*, 2016, **32**(2), 243–248.
- 40 D. L. Massart, *Handbook of Chemometrics and Qualimetrics*, 1st edn, Elsevier Science, 1997.
- 41 Y. Wang, M. Ji, X. Wu, T. Wang, J. Wang and X. Hou, *Microchem. J.*, 2019, **146**, 214–219.

