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A simple electrochemical immunosensor based on a chitosan/reduced graphene oxide nanocomposite for sensitive detection of biomarkers of malignant melanoma†

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The sensitive and specific detection of tumor biomarkers is crucial for early diagnosis and treatment of malignant melanoma. Immunoassay with a simple sensing interface and high sensitivity is highly desirable. In this work, a simple electrochemical immunosensor based on a chitosan/reduced graphene oxide (CS-rGO) nanocomposite was developed for sensitive determination of an S-100B protein, a tumor marker of malignant melanoma. CS-rGO nanocomposites were prepared by chemical reduction of graphene oxide in the presence of chitosan and modified on glassy carbon electrode (GCE) to provide a biofriendly, conductive, and easily chemically modified matrix for further immobilization of antibodies. Anti-S-100B antibodies were grafted onto the chitosan molecules to fabricate the immunorecognition interface by a simple glutaraldehyde cross-linking method. Electrochemical determination of S-100B was achieved by measuring the decreased current signal of solution phase electrochemical probes, which originated from the increased steric hindrance and insulation caused by the formation of antigen-antibody complexes at the electrode interface. Due to the good conductivity, high surface area, excellent biocompatibility, and good film-forming ability of CS-rGO, the constructed immunosensor exhibited good stability, high selectivity and sensitivity, a wide dynamic range from 10 fg mL⁻¹ to 1 ng mL⁻¹ and a low limit of detection of 1.9 pg mL⁻¹ (S/N = 3). Moreover, the sensor was also applicable for the sensitive detection of S-100B protein in real human serum samples.

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1. Introduction

Malignant melanoma is one of the fastest growing malignant tumors.¹ As the most malignant skin tumor, it can occur in different tissues such as skin, mucous membranes (*e.g.* alimentary tract, respiratory tract, and genitourinary tract),² eye glucose membrane, soft tissue, *etc.*³ Malignant melanoma has a lower age of death than other solid tumors.⁴ In addition, malignant melanoma lacks specific treatment other than early surgical resection and has a poor prognosis. Thus, early diagnosis and treatment of malignant melanoma is extremely important to improve survival.⁵ As an acidic calcium-binding protein, S-100B is the first marker to specifically label tumor cells of malignant melanoma and its value in the early diagnosis of malignant melanoma has been fully confirmed.⁶ When the level of S-100B in serum exceeds 0.5 ng mL⁻¹, it is generally

considered to be a pathological concentration. The concentration range of 0.15–0.5 ng mL⁻¹ is between pathological and normal, and the concentration below 0.15 ng mL⁻¹ is normal.⁷ Therefore, sensitive detection of S-100B in serum based on a simple method is of great significance.

Until now, quantification of S-100B levels in serum is mainly based on various immunoassay methods including enzyme-linked immunosorbent assay, fluorescence immunoassay, radioimmunoassay, and electrochemiluminescence immunoassay.^{8–11} However, these methods typically suffer from expensive instruments, specialized operating techniques, high costs, or low sensitivity. In contrast, electrochemical immunoassays combine the advantages of rapid detection, simple instrument, low cost, easy integration, and potential for high throughput.^{12–14} It is highly desirable to construct an efficient immunosensor for fast, sensitive, and low-cost detection of S-100B.

The introduction of functional nanomaterials to improve the performance of electro-chemical immunosensors has attracted much attention in recent years.^{15,16} Graphene based materials have recently attracted great interests due to the unique structure, multidimensional scale (*e.g.* zero dimension-0D graphene quantum dots,^{17–19} two dimensional-2D graphene nanosheets,^{20–22} three-dimensional-3D graphene foam^{23–25}) and rich

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properties, such as good electrical conductivity, high surface area, and good hybridization ability with other functional materials *etc.* Thus, graphene based materials have shown great potential in the fields of (bio)sensor, energy storage, and drug delivery, *etc.*^{26–31} However, the graphene material is prone to agglomeration due to the strong van der Waals forces between the layers. It has been proven that biofunctionalization of graphene material can effectively avoid the agglomeration between layers, and improve its hydrophilicity and biocompatibility.³² Chitosan (poly(1,4)-2-amino-2-deoxy- β -D-glucan, CS) is a biological polysaccharide obtained by partial deacetylation of natural chitin. As an important biopolymer, CS is rich in sources and has good biocompatibility. The abundant hydroxyl and amino groups in its molecule make it easy to be derivatized and functionalized. Although CS has good film-forming properties, excellent biocompatibility and high chemical reactivity, CS film has poor mechanical property and usually exhibits easy hydrolysis/swelling in water. It has been proven that graphene materials can effectively improve the strength, stability, water resistance of polymer.³³

Here, a simple electrochemical immunosensor was developed based on CS/reduced graphene oxide (CS-rGO) nanocomposite for sensitive detection of the biomarker of malignant melanoma, S-100B. The reduction of graphene oxide (GO) in presence of CS avoided the agglomerate of graphene materials and the obtained CS-rGO nanocomposite exhibited good dispersibility in water and monolayer structure. When CS-rGO was drop-coated on glassy carbon electrode (GCE), the biocompatible surface with high stability and excellent electron transfer was beneficial for covalent immobilization of recognition antibodies. Sensitive electrochemical detection of S-100B could be realized by utilizing the reduction of the signal of the redox probe ($\text{Fe}[\text{CN}]_6^{3-/4-}$) after the formation of the antigen-antibody complex. The sensor construction was simple and easy operated. The introduction of rGO not only improved the electron transfer rate, but also significantly enhanced the stability of the modified film. Since CS-rGO can provide a biocompatible environment for the immobilized antibodies, the fabricated immunosensor exhibited high stability and achieved sensitive electrochemical detection of S-100B in human serum. This work demonstrates an efficient strategy for easy construction of electrochemical immunosensors with simple fabrication, high sensitivity, good stability, providing a new strategy for electrochemical detection of tumor markers.

2. Experimental

2.1 Materials and reagents

Mouse anti-human S-100B monoclonal antibody (Ab), S-100B antigen, carcinoembryonic antigen (CEA), prostate specific antigen (PSA) and carcinoma antigen 125 (CA125) and carcinoma antigen 199 (CA199) were purchased from Beijing KEY-BIO Biotech Co., Ltd. (Beijing, China). Bone gamma-carboxyglutamate protein (BGP) was purchased from NanJing OkayBio Co., Ltd. (Nanjing, China). Bovine serum albumin (BSA), sodium phosphate dibasic dodecahydrate ($\text{Na}_2\text{HPO}_4 \cdot 12\text{H}_2\text{O}$), potassium ferricyanide ($\text{K}_3[\text{Fe}(\text{CN})_6]$), tetrapotassium

hexacyanoferrate trihydrate ($\text{K}_4[\text{Fe}(\text{CN})_6]$) were obtained from Aladdin Biochemical Technology Co., Ltd (Shanghai, China). Sodium dihydrogen phosphate dehydrate ($\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$) was obtained from Shanghai Macklin Biochemical Technology Co., Ltd. (Shanghai, China). Phosphate buffer solution (PBS) was prepared using Na_2HPO_4 and NaH_2PO_4 . Human blood serum (healthy man) was provided by Shanxi Bethune Hospital (Taiyuan, China). All other chemicals were of analytical grade and used without further treatment. Ultrapure water ($18.2 \text{ M}\Omega \text{ cm}$) was prepared by the Milli-Q system and used throughout the work.

2.2 Characterizations

Scanning electron microscopy (SEM) investigation was performed on a SU8100 micro-scope (Hitachi, Japan). The applied acceleration voltage was 10 kV. UV-visible absorption spectroscopy (UV-vis) was obtained by UV-2450 spectrophotometer (Shimadzu, Japan). Transmission electron microscope (TEM) measurement was performed at an acceleration voltage of 200 kV (JEM-2100, JEOL, Japan). Fourier transform infrared spectroscopy (FT-IR) was measured by a Vertex 70 spectrometer (Bruker, Germany) at room temperature on power-pressed KBr pellets. X-ray photoelectron spectroscopy (XPS) analysis was carried out using Mg K α radiation (250 W, 14 kV) on a PHI5300 electron spectrometer (PE Ltd, USA). Atomic force microscopic (AFM) image was obtained on a Bruker Multimode 8 (Bruker, Inc, USA). All electrochemical experiments included cyclic voltammetry (CV) and differential pulse voltammetry (DPV) were performed on the electrochemical workstation (Autolab PGSTAT302N, Metrohm, Switzerland) using the traditional three electrode system. Briefly, the working electrode adopted bare or modified glassy carbon electrode (GCE). The reference electrode was a silver/silver chloride (Ag/AgCl) electrode saturated with potassium chloride, and the counter electrode was a platinum wire electrode. Raman spectra was measured using a 960FT-Raman spectrometer (Thermo Nicolet, USA).

2.3 Preparation of CS-rGO nanocomposite

Graphene oxide (GO) was prepared by Hummers' method. CS-rGO was prepared as previously reported.³⁴ Briefly, GO solution (1 mg mL^{-1} , 4 mL) was added into CS solution (0.25%, pH = 3.0) with stirring until the solution became clear and transparent. Subsequently, the solution was then sonicated for 30 min before adding hydrazine hydrate (20 μL). Afterwards, the mixture reacted at 80 $^\circ\text{C}$ for 3 h with stirring to reduce GO. Then, the product solution was centrifuged at 3000 rpm for 10 min to remove large particles. The supernatant was collected and further centrifuged at 15 000 rpm for 10 min to obtain CS-rGO nanocomposite. The resulting solid was washed thoroughly with 0.1 mM HCl and then dried by lyophilization. Then, CS-rGO power was obtained for the fabrication of immunosensor based on CS-rGO and the electrochemical determination of S-100B.

2.4 Fabrication of immunosensor

Glassy carbon electrode (GCE, 3 mm in diameter) was used as the supporting electrode for constructing the immunosensor.



Before use, GCE was sequentially polished with 0.3 and 0.05 μm Al_2O_3 power slurry to obtain a mirror-like surface. Then, the electrode was thoroughly cleaned with acetone and ultrapure water for 3 min under ultrasonic treatment. The construction of the immunosensor included three steps. Firstly, GCE was modified with CS-rGO nanocomposites. Briefly, 10 μL of CS-rGO solution (0.1 mg mL^{-1}) was drop-coated on GCE followed with drying at room temperature. Secondly, S-100B antibody (Ab) was covalently immobilized on CS-rGO/GCE using glutaraldehyde (GA) as the linker. To activate the amino groups in CS molecules, 20 μL GA (0.01 wt%) was drop-coated on the surface of CS-rGO/GCE and incubated at 25 $^\circ\text{C}$ for 15 min in dark. After thorough washing with ultrapure water to remove residual GA, the resulting electrode was immersed in the Ab solution (50 $\mu\text{g mL}^{-1}$ in 0.01 M PBS, pH = 7.4). After incubating for 90 min at 25 $^\circ\text{C}$, the unbound Ab on electrode surface was adequately rinsed with PBS (0.01 M PBS, pH = 7.4). Finally, the non-specific sites on the obtained Ab/CS-rGO/GCE were blocked by incubation with BSA (1%) at room temperature for 1 h. After carefully rinsed with PBS (0.01 M, pH = 7.4) to remove residual BSA, the immunosensor was finally obtained and denoted as BSA/Ab/CS-rGO/GCE.

2.5 Fabrication of immunosensor

$\text{Fe}(\text{CN})_6^{3-}/\text{Fe}(\text{CN})_6^{4-}$ was used as the electrochemical probe to detect S-100B. When Ab on the surface of the immunoelectrode selectively recognized S-100B, the formation of antigen-antibody complex reduced the signal of the electrochemical probe. 0.1 M KCl containing 2.5 mM $\text{Fe}(\text{CN})_6^{3-}/\text{Fe}(\text{CN})_6^{4-}$ was used as the electrolyte for the detection of S-100B. The DPV curves before and after incubation the immunosensors with different concentrations of S-100B at 37 $^\circ\text{C}$ for 30 min were recorded. For real sample analysis, different concentrations of S-100B were added to fresh human serum to simulate the different levels of S-100B in cancer patients. Then, the obtained serum was diluted by a factor of 50 and S-100B was detected using the immunosensor for the fabrication of immunosensor based on CS-rGO and the electrochemical determination of S-100B.

3. Results and discussion

3.1 Preparation and characterization of CS-rGO nanocomposite

Although graphene has excellent electron transfer rate and high specific surface area, it has highly hydrophobic surface and strong van der Waals force between layers. Thus, graphene is commonly difficult to be dispersed in water and the agglomeration caused by interlayer stacking will significantly reduce its specific surface area. However, the precursor for the preparation of rGO, GO, is hydrophilic and has good dispersibility in water.³⁴ As illustrated in Fig. 1, CS-rGO with good dispersibility in aqueous solution was prepared in this work. Briefly, GO was reduced by hydrazine in the presence of CS. The CS-rGO nanocomposite drop-coated on GCE provided abundant amine groups to react with GA to introduce aldehyde groups for

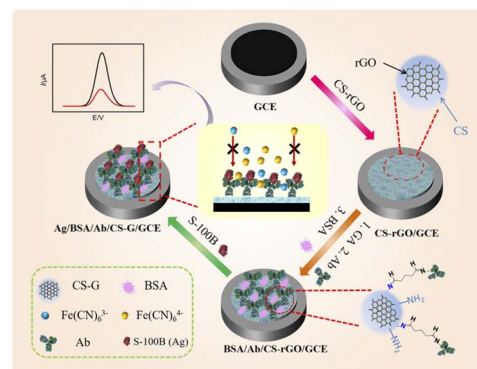


Fig. 1 Schematic illustration for the fabrication of immunosensor based on CS-rGO and the electrochemical determination of S-100B.

covalent immobilization of recognitive Ab. After blocking the nonspecific site with BSA, the immunosensor was prepared.

The morphology of the synthesized CS-rGO was characterized by SEM. As shown in Fig. 2A, CS-rGO has a unique folded structure of graphene. TEM images of GO and CS-rGO were also given in Fig. S1.[†] As shown, GO exhibited a thin-layered structure with some wrinkles. After reduction by hydrazine and binding to CS, the as-prepared CS-rGO nanocomposite retained the characteristic sheet-like structure. In addition, no agglomerated structure was observed, indicating that the reduction of GO in the presence of CS can avoid agglomeration of graphene material. The morphology and thickness of CS-rGO were further verified by AFM. As shown in Fig. S2,[†] CS-rGO was single-layered sheet with a thickness of about 3.5 nm. This thickness is higher than that of monolayer rGO (commonly around 1.0 nm). As CS bound on the surface of rGO through electrostatic or hydrogen bonding or van der Waals forces, the thickness of CS-rGO increased. Thus, the composite of chitosan with rGO did not change the monolayer structure of rGO, but resulted in an increase in its sheet thickness.

The successful composite of CS and rGO was characterized by ultraviolet-visible absorption spectroscopy (UV-vis, Fig. 2B). In case of GO, the $\pi-\pi^*$ transitions of C=C and the $n-\pi^*$ transitions of C=O were revealed corresponding to 230 nm and 300 nm, respectively. When GO reacted with hydrazine, on the one hand, the $\pi-\pi^*$ transitions of C=C red-shifted to 265 nm, proving that the increase of the conjugated structure through the reduction of GO. In addition, the absorption peak corresponding to the $n-\pi^*$ transition became indistinct,³⁵ which was

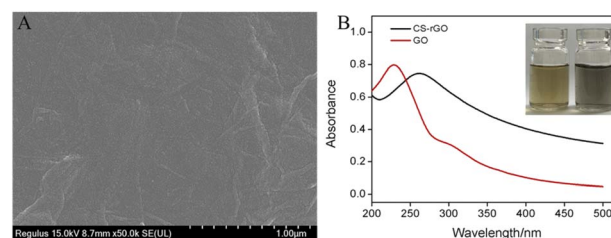


Fig. 2 (A) SEM image of CS-rGO film modified on GCE. (B) UV-vis spectra of GO and CS-rGO.



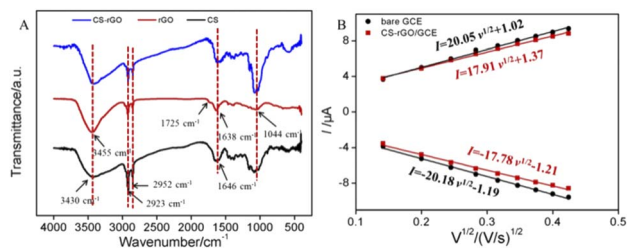


Fig. 3 (A) FR-IR spectra of CS, CS-rGO, and rGO synthesized without CS. (B) Relationship between $I/v^{1/2}$ derived from the CVs of bare GCE and CS-rGO/GCE obtained in 0.5 mM $K_3[Fe(CN)_6]$ containing 0.1 M KCl.

attributed to the reduction of surface oxygen-containing functional groups after GO is reduced to rGO. These phenomena demonstrated the successful preparation of CS-rGO.

Fig. 3A showed the FR-IR spectra of three materials including CS, rGO, and CS-rGO. Amongst, rGO was obtained by reduction of GO under the same conditions without CS. For chitosan, the peaks at 3430 cm^{-1} corresponded to the stretching vibration absorption peak of $-OH$, peaks at 2923 cm^{-1} and 2852 cm^{-1} were the stretching vibration absorption peaks of $C-H$. The presence of the characteristic peak of $N-H$ at 1646 cm^{-1} indicated that there had chemically reactive amino groups in chitosan.^{36,37} For rGO, the vibrational absorption peak of $-OH$ at 3455 cm^{-1} , the stretching vibration of $C=O$ at 1725 cm^{-1} , and the stretching vibration peak of $C=C$ at 1638 cm^{-1} , and the stretching vibration peak of $C-O-C$ at 1044 cm^{-1} revealed both the sp^2 carbon structure and the surface oxygen-containing groups on rGO.³⁸ In case of CS-rGO, the FT-IR spectrum contained characteristic absorption peaks of rGO and CS, indicating successful preparation of the nanocomposite material.

The changes of chemical groups during the preparation of CS-rGO from GO were further investigated by XPS. As shown in Fig. S3A and B,[†] the high-resolution C 1s spectra of both GO and CS-rGO displayed four types of carbon atoms, including $C-C/C=C$ (sp^2 carbon atoms, 284.6 eV), $C-O$ (epoxy or alkoxy group, 286.7 eV), $C=O$ (aldehyde or ketone group, 287.2 eV) and $O-C=O$ (carboxyl group, 288.5 eV). However, the $C-C/C=C$ peak in CS-rGO became larger, and the peak intensities of $C=O$ and $COOH$ greatly reduced, indicating that GO was successfully reduced. In the N 1s spectrum of CS-rGO in Fig. S3C,[†] characteristic peaks of $-NH_2/-NH-$ and $-NH_3^+$ appeared, proving that chitosan successfully composited on rGO.

Fig. S4[†] displayed the Raman spectra of GO and CS-rGO. As shown, two prominent peaks at $\sim 1350\text{ cm}^{-1}$ (D band) and $\sim 1590\text{ cm}^{-1}$ (G band) were observed, which corresponded to a breathing mode of κ -point photons of A_{1g} symmetry and the first order scattering of the E_{2g} phonon of sp^2 C atoms, respectively. When GO was reduced in presence of CS, the obtained CS-rGO nanocomposite had a higher intensity ratio ($r = I_D/I_G$, 1.32) in comparison with that of GO ($r = 0.85$), suggesting the increase of sp^3 defects within the sp^2 carbon network upon the reduction.³⁹

3.2 Electrochemical characterization of CS-rGO/ITO

Fig. S5A[†] showed the cyclic voltammetry (CV) curves of CS/GCE and CS-rGO/GCE in $Fe(CN)_6^{3-/4-}$ solution. As seen, CS-rGO/

GCE had a significantly improved electrochemical signal compared with that of CS/GCE, which was attributed to the high electron transfer rate of rGO. It was noteworthy that the peak current obtained on CS-rGO/GCE exhibited good stability during the continuous 20 scanning cycles (Fig. S5B[†]). The peak current remained 98% of the initial signal after 20 consecutive scans. In contrast, the electrochemical signal of redox probe remarkably increased when CS/GCE was continuously scanned (Fig. S6[†]), indicating the instability of the modified CS layer resulting from the swelling or shedding of the CS. Thus, the introduced rGO can significantly improve the stability of CS film, which might be ascribed to the cross-linking between CS and rGO through electrostatic interaction or hydrogen bonding or van der Waals force.

The effect of scan rate on the electrochemical response was also investigated. Fig. S7[†] showed the cyclic voltammetry curves obtained on CS-rGO/GCE or bare GCE at different scan rates. As seen from Fig. 3B, the peak current had a good linear relationship with the square root of the scan rate, proving a diffusion-controlled process on both electrodes. The electrochemical active surface area (ECSA) of bare GCE can be calculated to be 0.058 cm^2 by the Randles-Sevcik equation,^{40–42} while that of CS-rGO/GCE was 0.052 cm^2 .

3.3 Fabrication of immunosensor

Fig. 4A displayed the CV curves of $Fe(CN)_6^{3-/4-}$ on different electrodes obtained during the construction of the immunosensor. When CS-rGO was modified on the surface of GCE, the CV peak signal of the electrochemical probe on the CS-rGO/GCE electrode was weakly reduced accompanying a slightly larger peak-to-peak difference in comparison with GCE since CS was a non-conductive biopolymer.⁴³ After further activation by GA, the peak signal on GA/CS-rGO/GCE slightly reduced due to the possible cross-linking between CS molecules. The following covalent immobilization of Ab (Ab/GA/CS-rGO/GCE) and BSA blocking (BSA/Ab/GA/CS-rGO/GCE) resulted in a significant decrease in the current signal and a further increase in the peak-to-peak difference, which was attributed to the hindered electron transfer by the non-conductive protein layer. When the fabricated immunosensor (BSA/Ab/GA/CS-rGO/GCE) was incubated with S-100B, only extremely weak probe signal was detected. Thus, the formation of antigen-antibody complexes significantly reduced electron transfer on the electrode surface. The DPV curves in Fig. 4B demonstrated the consistent results,

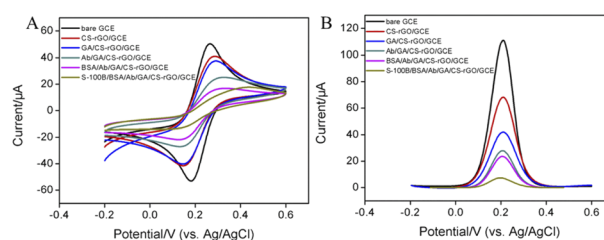


Fig. 4 CV (A) and DPV (B) curves obtained on different electrodes obtained in the fabrication of the immunosensor or after incubated with S-100B in $Fe(CN)_6^{3-/4-}$ solution.



indicating efficient fabrication of the immunosensor and the specific recognition of S-100B on the immuno-recognitive surface.

3.4 Electrochemical determination of S-100B using the developed immunosensor

The formation of antigen–antibody complex on the surface of immunosensor significantly reduced the signal of the redox probe ($\text{Fe}(\text{CN})_6^{3-}/\text{Fe}(\text{CN})_6^{4-}$), realizing electrochemical determination of S-100B. The performance of the developed immunosensor for the electrochemical determination of S-100B was investigated. Fig. 5A displayed DPV curves obtained on the fabricated immunosensor, BSA/Ab/GA/CS-rGO/GCE, in presence of different concentration of S-100B. As shown, the peak current gradually decreased when the concentration of S-100B increased. A linear correlation was revealed between the peak current (I , μA) and the logarithm value of the concentration of S-100B ($\log C$) in the range from 10 fg mL^{-1} to 1 ng mL^{-1} ($I = -2.62 \log C_{\text{S-100B}} + 23.4$, $R^2 = 0.990$, Fig. 5B). The limit of detection (LOD) was 1.9 fg mL^{-1} at a signal-to-noise ratio of 3 ($S/N = 3$).

Comparison between determination of S-100B using different methods was demonstrated in Table S1.^{†44–49} The LOD is lower than that obtained using fluorescent determination based on antibody-conjugated magnetic beads and quantum dots (MB-Ab/S100B/Ab-QD),⁴⁴ or electrochemical impedance spectroscopy (EIS) determination based on antibody binding to cysteamine modified on interdigitate-zigzag biochip using glutaraldehyde as cross-linking agent (IDZB/Cys/GA/anti-S100B),⁴⁵ electrochemical determination based on Au-coated magnetic NPs/thiol-ended Ab,⁴⁶ but higher than photo-electrochemical determination based on immobilization of Ab on green reduced graphene oxide and decorated with gold nanoparticles (Ab/rGO-Au),⁴⁷ or electrochemical determination by immobilization of Ab on 4-nitrobenzenediazonium/GA modified graphene screen printed electrode (GSPE/4-NBD/GA/Ab),⁴⁸ or poly(ethyleneimine) modified poly(methyl methacrylate) (PEI-PMMA) modified electrode.⁴⁹

3.5 Selectivity of the immunosensor

To further explore the selectivity of the immunosensor, the response obtained when the immunosensor was incubated with several common tumor biomarkers including BGP, PSA, CEA, CA125, CA199, were investigated. As shown in Fig. 6, the current obtained on the immunosensor almost did not change in

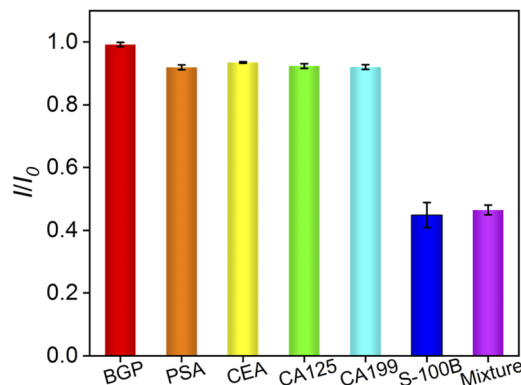


Fig. 6 The currents obtained on the developed immunosensor in absence (control) or presence of the indicated tumor biomarker.

Table 1 Determination of S-100B in human serum samples

Sample	Added (pg mL ⁻¹)	Found (pg mL ⁻¹)	RSD (% n = 3)	Recovery (%)
Human serum	5.00	4.87	3.4	97.4
	50.0	52.6	1.4	105
	500	488	3.5	97.6

absence (control) or presence of one of the possible interferences. On the contrary, the peak current decreased greatly when S-100B or the mixture of all proteins was added.

3.6 Reproducibility and stability of the immunosensor

The reproducibility and stability of the constructed immunosensor were further investigated. Five immunosensors were prepared in parallel. After the fabricated immunosensors were incubated with S-100B (0.1 ng mL^{-1}), the current signal of the electrochemical probe on different electrodes were similar with a low relative standard deviation (RSD, 1.8%), indicating that the electrode fabrication had good reproducibility. The storage stability of the developed immunosensor was also investigated. When the immunosensor was stored at 4°C for different times, the RSD for the determination of S-100B (10 pg mL^{-1}) was 2.1%, demonstrating high stability.

3.7 Determination of S-100B in human serum

Electrochemical determination of S-100B in human serum was investigated using standard addition method. Briefly, different concentrations of S-100B were artificially added into the human serum. Then, the human serum was diluted by a factor of 50. As shown, the recovery of the concentration of S-100B ranged from 97.4% to 105% with an RSD no more than 3.5% (Table 1), indicating good reliability for the determination of S-100B.

4. Conclusions

In summary, a simple immunosensor sensor was developed for sensitive determination of S-100B based on the modification of electrode using chitosan-reduced graphene oxide

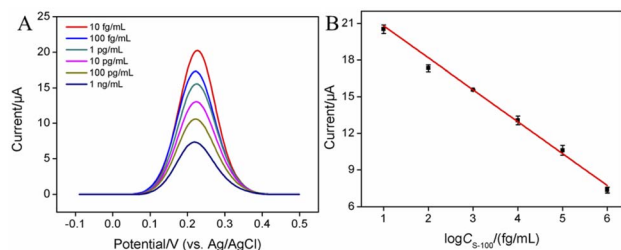


Fig. 5 (A) DPV curves of the immunosensor in the presence of different concentration (10 fg mL^{-1} to 1 ng mL^{-1}) of S-100B. (B) Calibration curve for the determination of S-100B.



nanocomposite. CS-rGO nanocomposites integrate the functional properties of graphene and chitosan, exhibiting two-dimensional monolayer structure, good electrical conductivity, biocompatibility, and modifiability. In comparison with pure CS film, the interaction between CS and rGO improve the stability of the film. The amino groups in CS molecules were activated by the functional linker, glutaraldehyde, for the further covalent immobilization of the recognitive antibody. As the formation of the antigen-antibody complex decreases the signal of the redox probe, the electrochemical determination of S-100B was realized with wide dynamic range from 10 fg mL⁻¹ to 1 ng mL⁻¹ and a low limit of detection of 1.9 fg mL⁻¹. Owing to the biocompatible environment for the immobilization of Ab and the good electron transfer of rGO, the developed sensor has advantages of high sensitivity and good stability, indicating great potential for the determination of tumor biomarkers.

Ethical statement

Human serum was used in this work. Informed consent was obtained from all human subjects.

Conflicts of interest

There are no conflicts to declare.

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