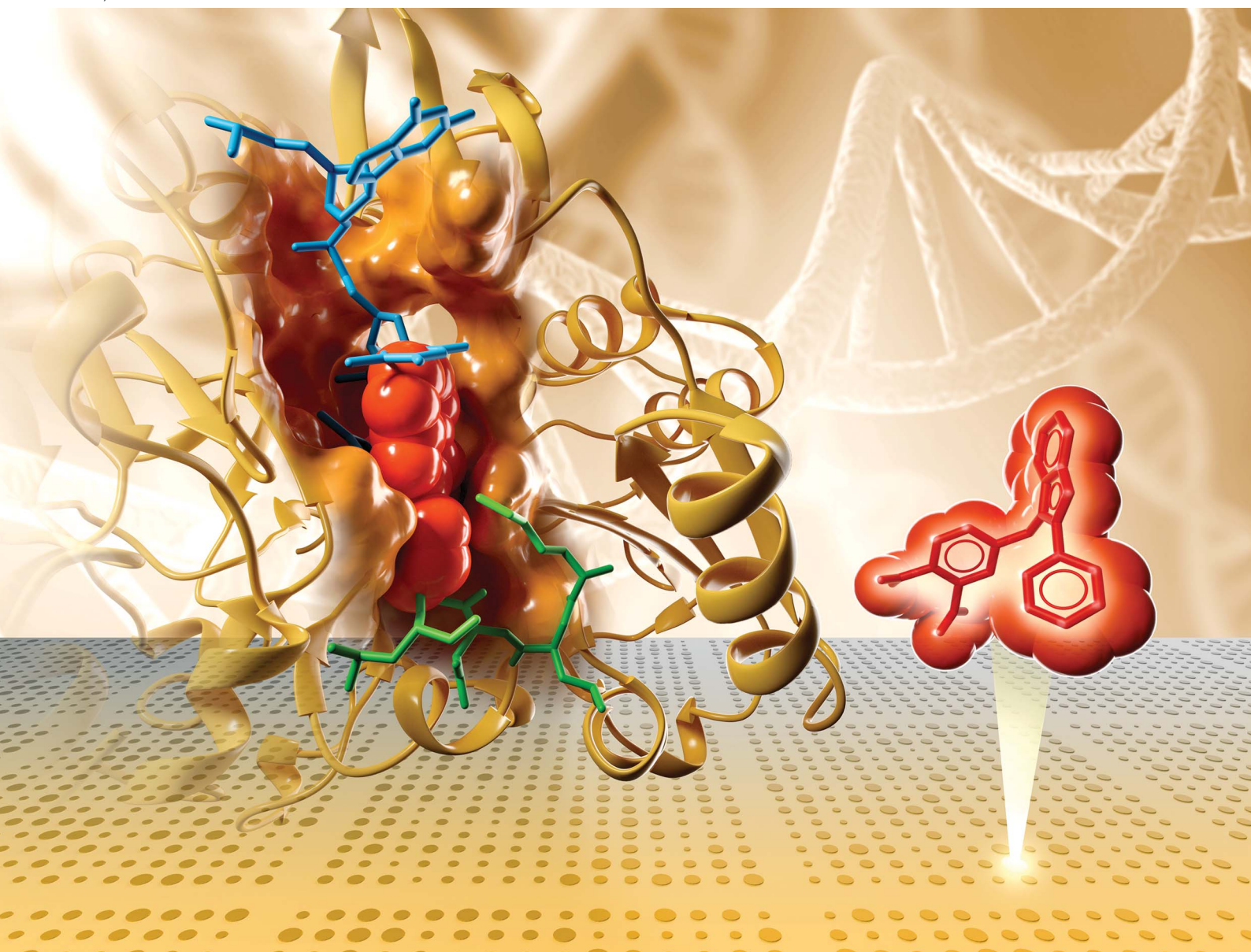


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Small molecule microarray identifies inhibitors of tyrosyl-DNA phosphodiesterase 1 that simultaneously access the catalytic pocket and two substrate binding sites†

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Tyrosyl-DNA phosphodiesterase 1 (TDP1) is a member of the phospholipase D family of enzymes, which catalyzes the removal of both 3'- and 5'-DNA phosphodiester adducts. Importantly, it is capable of reducing the anticancer effects of type I topoisomerase (TOP1) inhibitors by repairing the stalled covalent complexes of TOP1 with DNA. It achieves this by promoting the hydrolysis of the phosphodiester bond between the Y723 residue of human TOP1 and the 3'-phosphate of its DNA substrate. Blocking TDP1 function is an attractive means of enhancing the efficacy of TOP1 inhibitors and overcoming drug resistance. Previously, we reported the use of an X-ray crystallographic screen of more than 600 fragments to identify small molecule variations on phthalic acid and hydroxyquinoline motifs that bind within the TDP1 catalytic pocket. Yet, the majority of these compounds showed limited (millimolar) TDP1 inhibitory potencies. We now report examining a 21 000-member library of drug-like Small Molecules in Microarray (SMM) format for their ability to bind Alexa Fluor 647 (AF647)-labeled TDP1. The screen identified structurally similar *N*,2-diphenylimidazo[1,2-*a*]pyrazin-3-amines as TDP1 binders and catalytic inhibitors. We then explored the core heterocycle skeleton using one-pot Groebke–Blackburn–Bienayme multicomponent reactions and arrived at analogs having higher inhibitory potencies. Solving TDP1 co-crystal structures of a subset of compounds showed their binding at the TDP1 catalytic site, while mimicking substrate interactions. Although our original fragment screen differed significantly from the current microarray protocol, both methods identified ligand–protein interactions containing highly similar elements. Importantly inhibitors identified through the SMM approach show competitive inhibition against TDP1 and access the catalytic phosphate-binding pocket, while simultaneously providing extensions into both the substrate DNA and peptide-binding channels. As such, they represent a platform for further elaboration of trivalent ligands, that could serve as a new genre of potent TDP1 inhibitors.

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Introduction

Tyrosyl-DNA phosphodiesterase 1 (TDP1) is a member of the phospholipase D family of enzymes that catalyses the removal of both 3'- and 5'-DNA phosphodiester adducts.¹ It serves

a variety of physiological roles, including the repair of a broad spectrum of lesions induced by anticancer drugs. Among its important substrates are the stalled topoisomerase type I (TOP1) and DNA covalent complexes produced when a TOP1 tyrosyl residue cleaves one strand of DNA.^{2–4} Functioning in this capacity, TDP1 inhibitors could potentially increase the anticancer activity of TOP1 inhibitors by reducing the repair of TOP1-DNA lesions.^{5,6} Accordingly, such agents may represent a promising new class of therapeutics with potential use for the treatment of cancer in synergistic combination with current TOP1 inhibitors.^{4,7–19}

While significant effort has been devoted to developing TDP1 inhibitors and a variety of inhibitors have been reported, the structural basis for how these inhibitors interact with TDP1 is generally unknown, which may limit the derivation of new

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analogues through structure-based approaches.^{6,20} In addition, many TDP1 inhibitors exhibit physiochemical properties, which could potentially endow them with promiscuous mechanisms of inhibition that would make them of questionable value for further development.^{21–23} Recently, we reported the structures of several small molecules bound to TDP1 that resulted from the crystallographic screening of more than 600 fragments.²⁴ Surprisingly, despite the relatively large size of the enzyme, all molecules were found to bind to the same site of TDP1, specifically within the catalytic phosphate-binding pocket. The uniform topological distribution of bound fragments could be related to the fact that they resulted from soaking preformed crystals, where the conformational mobility of the solid crystalline protein may be more limited than would be found with protein in solution.

Progress in the development of TDP1 inhibitors could be significantly assisted by expanding the diversity of molecular structures that are capable of accessing and binding to the catalytic machinery. Following up on the reasoning that contacts between neighbouring proteins in the crystal lattice during our initial fragment screens may have limited access to some potential binding sites, we performed a second screen using a small molecule microarray (SMM) approach, where protein–ligand interactions would occur in an aqueous milieu. Toward this end, we screened fluorescently labeled TDP1 in a binding assay against a SMM of 21 000 spatially arrayed drug-like molecules bound to a glass surface and achieved an approximate overall 0.5% hit rate. Based on structural similarities, we examined a subset of compounds for their ability to inhibit TDP1 catalysis *in vitro*. This led us to identify the *N*,2-diphenylimidazo[1,2-*a*]pyrazin-3-amine nucleus as a new TDP1-binding motif exhibiting low-micromolar inhibitory potencies. We further extended this class of compounds to the *N*,2-diphenylimidazo[1,2-*a*]pyridin-3-amine nucleus using one-pot Groebke–Blackburn–Bienayme multicomponent reactions that employ readily available building blocks of aldehyde, isocyanide and pyrazin-2-amine or pyridin-2-amine. By combining structural features of *N*,2-diphenylimidazo[1,2-*a*]pyrazin-3-amine with phthalic acid we were able to confirm by X-ray crystallography that these inhibitors bind within the catalytic site of TDP1, where they replicate aspects of the DNA–TOP1 substrate complex. Remarkably, ligands resulting from the SMM approach bound to the same phosphate-binding site as those obtained from crystallographic screening. Additionally, inhibitors identified through the SMM approach not only accessed the catalytic phosphate-binding pocket, but also simultaneously provided extensions into both the substrate DNA and peptide-binding channels. As such, they represent a potential platform for further elaboration into trivalent ligands, that could serve as a new genre of potent TDP1 inhibitors.

Results

Small molecule microarray (SMM) screening to identify new TDP1-binding motif

SMM can offer a highly efficient and compact platform to interrogate large numbers of compounds.²⁵ Previously, we

utilized the catalytic domain of TDP1 (residues 148–608) to screen for TDP1-binding fragments using X-ray crystallography.²⁴ Within this catalytic domain there are three surface-situated Cys residues (C205, C251 and C413) that are potentially accessible for fluorescence labelling using maleimide–thiol coupling reactions. In our current work, we labeled TDP1 (148–608) with Alexa Fluor 647 (AF647) employing a commercially available AF647 C2 maleimide dye kit (Fig. 1A).²⁶ We then compared the enzymatic activity of the AF647-tagged TDP1 (148–608) construct with unlabelled catalytic domain as well as with the full-length wild-type TDP1 using an *in vitro* gel-based TDP1 assay (Fig. 1B).²⁴ The AF647-labeled TDP1 was found to have catalytic activity intermediate between the more active TDP1 (148–608) and the less active wild-type TDP1 (Fig. 1C), indicating that the AF647-labeled TDP1 (148–608) was suitable for SMM binding screen.

Next, to identify compounds that selectively bind to TDP1, we screened AF647-labeled TDP1 (148–608) against drug-like small molecules using a SMM (Fig. 2A).^{27,28} Briefly, a library of 21 000 compounds was covalently immobilized on glass slides using isocyanate surface chemistry, as described.²⁹ Incubating the SMM with the AF647-labeled TDP1 (148–608) identified 109 hits (0.5% hit rate), which were assigned visually into similar structural groups (Table S1†). Based on this grouping, 37 representative compounds

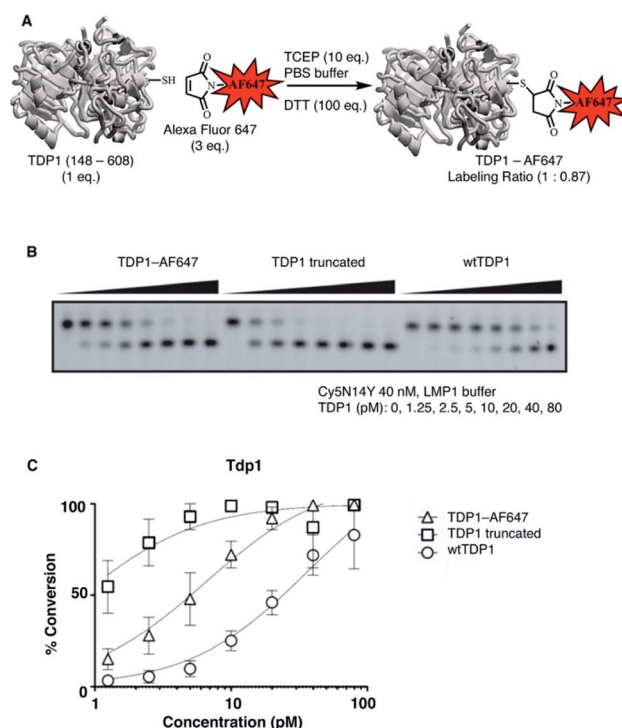


Fig. 1 Labelling TDP1 (148–608) with Alexa Fluor 647. (A) Procedural schematic of reacting the catalytic domain of TDP1 (148–608) with Alexa Fluor 647 (AF647) maleimide; (B) gel-based assay showing the activity of AF647-labeled TDP1 (148–608) (TDP1-AF647) as compared with the catalytic domain of unlabelled TDP1 (148–608) (TDP1 truncated) and wild-type full-length TDP1 (wtTDP1). (C) Plot of enzyme activity data shown in panel (B).



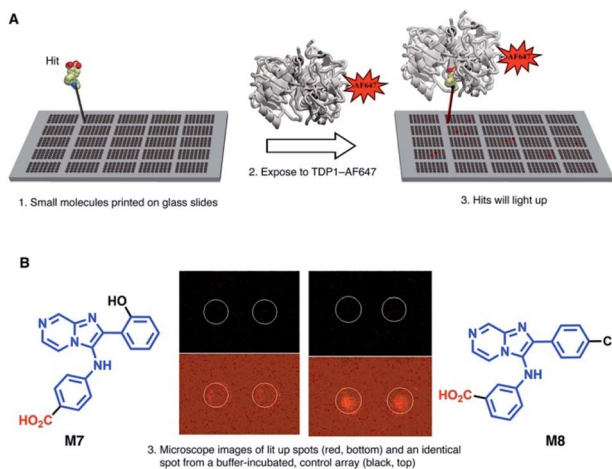


Fig. 2 (A) Schematic representation of the SMM screen using AF647-labeled TDP1 (148–608) to identify hits that bind to TDP1. (B) Chemical structures of **M7** and **M8** and microscope images of their SMM spots. The shared *N*,*N*-diphenylimidazo[1,2-*a*]pyrazin-3-amine core is highlighted in blue.

were evaluated for their ability to inhibit TDP1 catalysis in gel-based TDP1 assays (Fig. S1 and S2†).³⁰ Compounds **M7** and **M8** (Fig. 2B) showed micromolar inhibitory potencies (**M7** $IC_{50} = 3 \mu\text{M}$; **M8** $IC_{50} = 233 \mu\text{M}$). Both **M7** and **M8** share an imidazo[1,2-*a*]pyrazin-3-amine core, with **M7** having a 4-aminobenzoic acid group and **M8** having a 3-aminobenzoic acid moiety. By this process, we identified the 2-phenylimidazo[1,2-*a*]pyrazin-3-phenylamine nucleus as a preferred TDP1-binding motif.

Inhibition of TDP1 by an imidazo[1,2-*a*]pyrazin-3-amine is competitive

We evaluated a series of commercially available analogues having an imidazo[1,2-*a*]pyrazin-3-amine core as well as a small number of synthetic analogues (Table S1†). We identified as being approximately two-fold more potent than **M7**, compound **7b**, which lacks the 2-hydroxyl group on the 2-phenyl ring of the imidazo[1,2-*a*]pyrazin-3-amine nucleus (Fig. 3A and S2†). In addition, we confirmed that **7b** exhibits a competitive mode of inhibition with a K_i value of $0.309 \pm 0.122 \mu\text{M}$ (Fig. 3B).

The imidazo[1,2-*a*]pyrazin-3-amines were initially all obtained commercially (Table S2†). However, these can be prepared by one-pot Groebke–Blackburn–Bienayme multicomponent reactions, in which freshly synthesized arylisocyanides (**3**) are coupled with aromatic aldehydes (**4**) in the presence of pyrazine-2-amines (**5**) (Scheme 1).³¹ Typically, the resulting products can be obtained in high purity by simple precipitation out of the reaction mixtures. We observed that product yields were generally at least 40%. However, yields were often substantially lower when electron withdrawing substituents were present on either the aldehyde or isocyanate components. Similar trends have been previously reported.³² Using this procedure and isolating product by precipitation, we obtained **7b** as a white amorphous solid spectroscopically identical to

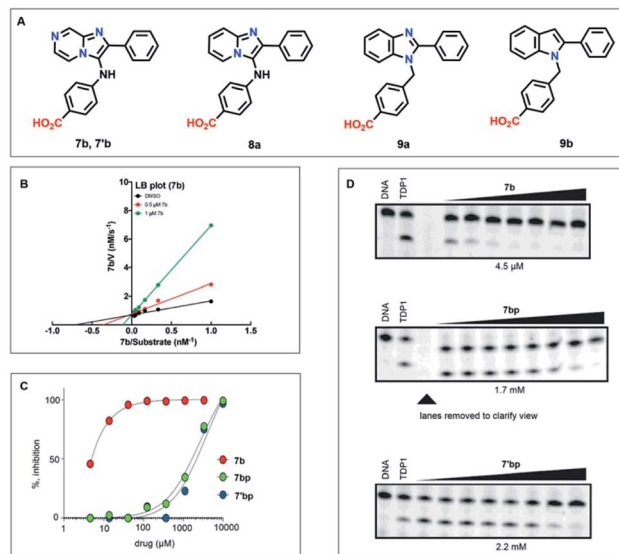
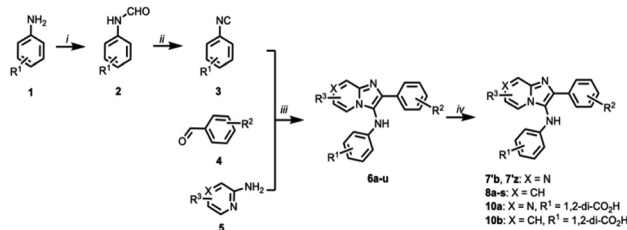


Fig. 3 (A) Structures of TDP1 inhibitors discussed in the text. (B) Lineweaver–Burk plot (LB plot) of TDP1 inhibition by **7b** showing a competitive mode of inhibition (constant of inhibition $K_i = 0.309 \pm 0.122 \mu\text{M}$). (C) TDP1 inhibition curves of **7b** (commercial solid), **7bp** (HPLC-purified **7b**), and **7'bp** (prepared and HPLC-purified sample from our laboratory). (D) Gel-based TDP1 inhibition assay using **7b**, **7bp**, and **7'bp**. Drug concentration for each band: 4.6, 13.7, 41.2, 123.5, 370, 1111, 3333, 10 000 (μM).



Scheme 1 Preparation of TDP1 inhibitors by one-pot Groebke–Blackburn–Bienayme multicomponent reactions. Reagents and Conditions: (i) Ac_2O , HCO_2H , 50°C ; (ii) POCl_3 , Et_3N , 0°C ; (iii) AcOH , MeOH , rt. (X = N: imidazo[1,2-*a*]pyrazin-2-amine or X = CH: imidazo[1,2-*a*]pyridin-2-amine); (iv) NaOH (2N, aq.), MeOH .

the original commercial sample (low resolution mass spectrum, ¹H- and ¹³C-NMR, Fig. 3A). The synthetic **7b** obtained in this fashion behaved in similar fashion to the commercial material **7b** in TDP1 assays.

Examination of structural variation of the imidazo[1,2-*a*]pyrazin-3-amine core

We next examined analogues by removing various nitrogen atoms from the parent nucleus. Deletion of the pyrazine 7-nitrogen of **7b** yielded the corresponding 2-phenyl-3-phenylamine-[1,2-*a*]pyridine nucleus (**8a**, Fig. 3A). Shifting the bridgehead 4-nitrogen of **8a** to the 3-position and conversion of the 3-phenylamine to 3-benzyl moiety provided the 1-benzyl-2-phenyl-1*H*-benzo[*d*]imidazole ring system (**9a**). Finally, removal of the 3-nitrogen from **9a** gave the 1-benzyl-2-phenyl-1*H*-benzo



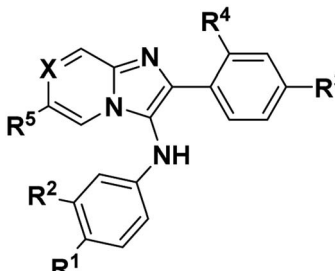
[*d*]imidazole nucleus (**9b**) (Fig. 3A). For comparison purposes, we retained the 4-carboxy group found in the parent **7b**. In the biochemical assays, analogues based on the [1,2-*a*]pyridine and 1*H*-benzo[*d*]imidazole nuclei (**9a** and **9b**, respectively) gave IC₅₀ values > 100 μM. Deletion of the pyrazine 7-nitrogen of **7b** to yield 4-((2-phenylimidazo[1,2-*a*]pyridine-3-yl)amino)benzoic acid (**8a**) resulted in retention of inhibitory potency (IC₅₀ = 9.3 μM) relative to the parent **7b** (IC₅₀ = 4.5 μM). We prepared a variety of analogues based the imidazo[1,2-*a*]pyrazin-3-amine core and subjected them to *in vitro* gel-based TDP1 inhibition assays (Table S2†).

When we subjected either the commercial sample **7b** or the synthetic sample **7'b** to reverse-phase HPLC purification (acetonitrile and H₂O with 0.1% TFA as eluents) and then lyophilized the collected eluent, we obtained the corresponding solids, **7bp** and **7'bp**, respectively ("p" indicating purification by HPLC). Both compounds gave identical ¹H NMR spectra, which showed slight differences relative to **7bp** and **7'bp** that were readily attributed to their different salt forms (HCl prior to HPLC purification). We also observed that both **7bp** and **7'bp** showed reduced inhibitory potencies in the TDP1 gel-based assay (Fig. 3D). The reasons for the loss of inhibitory potency are not clear.

Structural variation of the *N*,2-diphenylimidazo[1,2-*a*]pyridine-3-amine core

Using the one-pot multicomponent Groebke–Blackburn–Bienayme protocol, we prepared a set of analogues based on the *N*,2-diphenylimidazo[1,2-*a*]pyridine-3-amine core (Scheme 1, wherein X = CH). We evaluated these analogues using the *in vitro* gel-based TDP1 assay (Table 1). Removal of the carboxylic acid group at the R¹ position (**8a**) resulted in significant loss of inhibitory potency (**6t**, IC₅₀ > 100 μM). While keeping a carboxylic acid at the R¹ position, a variety of substituents were examined at the R³ position. Those substituents that were not proton donors retained inhibitory IC₅₀ values of from 3 μM to 15 μM (*i.e.*, R³ = CH₃, OBn, CF₃ and NO₂; **8b**, **c**, **d**, **e**, respectively). However, inhibitory potencies were severely diminished (IC₅₀ > 100 μM) when proton donors were introduced (*i.e.*, R³ = OH, SO₃H, CO₂H; **8f**, **g**, **h**, respectively, Table 1). Good inhibitory potency was regained by moving the carboxyl group in **8h** from the R³ position to the R⁵ position (**8m**, IC₅₀ = 0.9 μM) or to the X position (**8s**, IC₅₀ = 4.0 μM). Shifting the carboxyl group to the R² position was also well tolerated (**10b**, IC₅₀ = 11.0 μM). Removing the R¹ carboxyl from **8m** resulted in good retention of inhibitory potency (**8k**, IC₅₀ = 2.4 μM), while shifting that carboxyl to the R³ position gave a modest loss in potency (**8i**,

Table 1 Evaluation of imidazo[1,2-*a*]pyridin-3-amines using a gel-based TDP1 assay *in vitro*



Compound	8a	6t	8b	8c	8d	8e	8f	8g	8h	8i	8j
R ¹	CO ₂ H	H	CO ₂ H	CO ₂ H	CO ₂ H	CO ₂ H	CO ₂ H	CO ₂ H	CO ₂ H	H	CO ₂ H
R ²	H	H	H	H	H	H	H	H	H	H	H
R ³	H	H	CH ₃	OBn	CF ₃	NO ₂	OH	SO ₃ H	CO ₂ H	CO ₂ H	H
R ⁴	H	H	H	H	H	H	H	H	H	H	CO ₂ H
R ⁵	H	H	H	H	H	H	H	H	H	H	CO ₂ H
X	CH	CH	CH	CH	CH	CH	CH	CH	CH	CH	CH
IC ₅₀ ^a (μM)	9.3	>100	14.8	12.7	4.2	3.2	>100	>100	>100	40.1	>100

Compound	8k	8l	8m	8n	8o	8p	8q	8r	6u	8s	10a	10b
R ¹	H	NO ₂	CO ₂ H	CO ₂ H	CO ₂ H	CO ₂ H	CO ₂ H	CO ₂ H	NO ₂	CO ₂ H	CO ₂ H	CO ₂ H
R ²	H	H	H	H	H	H	H	H	H	H	CO ₂ H	CO ₂ H
R ³	H	H	H	Ph	H	H	H	Ph	H	H	H	H
R ⁴	H	H	H	H	H	H	H	H	H	H	H	H
R ⁵	CO ₂ H	CO ₂ H	CO ₂ H	CO ₂ H	Br	H	SO ₂ Me	SO ₂ Me	SO ₂ Me	H	H	H
X	CH	CH	CH	CH	CH	CPh	CH	CH	CH	CCO ₂ H	N	CH
IC ₅₀ ^a (μM)	2.4	22.0	0.9	1.5	1.6	1.8	1.8	9.95	16.1	4.0	45.5	11.0

^a Half maximal inhibitory concentrations (IC₅₀ values) obtained from *in vitro* TDP1 gel-based assays.



$IC_{50} = 40.1 \mu M$). Adding a carboxyl group to the R^4 position of **8m** was highly deleterious (**8j**, $IC_{50} > 100 \mu M$). Replacing the R^5 carboxyl in **8m** with a bromine (**8o**, $IC_{50} = 1.6 \mu M$) or methyl-sulfone (**8q**, $IC_{50} = 1.8 \mu M$) was well tolerated, causing little effect on the good inhibitory potency of **8m**. Replacing the R^1 carboxyl in **8m** with a nitro group (**8l**) lowered potency approximately 20-fold (from $IC_{50} = 0.9 \mu M$ to $22.0 \mu M$). Doing a similar replacement in **8q** caused an approximate 10-fold loss of potency (**6u**, $IC_{50} = 16.1 \mu M$). Finally, adding a phenyl ring to the R^3 position of **8m** was well tolerated (**8n**, $IC_{50} = 1.5 \mu M$), but when a similar change was done to **8q**, a modest loss of potency was observed (**8r**, $IC_{50} = 9.9 \mu M$). In summary, a carboxyl group at the R^1 position was generally important for inhibitory potency. While retaining a carboxyl group at the R^1 position, substitution at the R^3 position was well tolerated (**8b**, **c**, **d**, **e**), except when the R^3 substituent was a proton donor and, in these cases, there was a significant loss of inhibitory potency (**8f**, **g**, **h**). Introduction of a carboxyl group at the R^5 or X positions was well tolerated (**8m**, **n**, **s**), but was deleterious at the R^4 position (**8j**). A carboxyl group at the R^5 position gave good inhibitory potency both in the presence (**8m**) and absence (**8k**) of a carboxyl group at the R^1 position. The presence of carboxyl groups simultaneously at the R^1 and R^2 positions was tolerated (**10b**). These compounds showed TDP1 selectivity over TDP2 in gel-based *in vitro* assay (Table S4†). Several of the compounds showed micromolar inhibition in assays employing whole cell extracts (WCE).³⁰ While having little cytotoxicity, some of the compounds were shown to act synergistically with the TOP1 inhibitor camptothecin (CPT) in human colon cancer cell assays (see below). Taken together, the data are consistent with the compounds selectively targeting TDP1.

Determination of Lineweaver–Burk plots for **8a**, **10a** and **10b**

We had initially prepared a Lineweaver–Burk analysis of the mode of TDP1 inhibition by **7b** and found that it was competitive in nature (Fig. 3B). Additionally, we now subjected **8a** (the corresponding *N*,2-diphenylimidazo[1,2-*a*]pyridine-3-amine version of **7b**), to Lineweaver–Burk analysis and found that it also inhibited TDP1 in a competitive fashion (Fig. S3†). Similarly, we subjected **10a** (the 3,4-dicarboxy variant of 4-carboxy-containing **7b**) and **10b** (the 3,4-dicarboxy variant of 4-carboxy-containing **8a**) to Lineweaver–Burk analysis and found that these also exhibited competitive modes of inhibition with micromolar K_i values (Fig. S3†).

Demonstration that the TDP1 inhibitors act in synergistic fashion with the TOP1 inhibitor camptothecin (CPT)

To establish whether our TDP1 inhibitors could act in cells as TDP1 inhibitors, we performed combination experiments with the classical TOP1 inhibitor camptothecin (CPT)³ in assays using the human colon cancer cell line HCT116 (Table S4 and Fig. S4–S7†). Synergy scores were calculated based on Synergy-Finder 2.0 (Table S4 and Fig. S6†).³³ Synergy scores were larger than 10 for **M7** (15.6), **6u** (10.8), **8m** (13.4), **8n** (11.3), **8o** (16.3), **8s** (20.7) and **10b** (17.9), indicating that the our TDP1 inhibitors act synergistically with CPT in human cancer cells.

Determination of protein–ligand interactions by X-ray crystallography

Recently, we disclosed the first X-ray crystal structures of low molecular weight molecules bound within the TDP1 catalytic pocket.²⁴ To determine the ligand–protein interactions for our current SMM-derived inhibitors, we solved the X-ray crystal structures of **10a** and **10b** bound to the TDP1 catalytic domain at 1.86 and 1.70 Å, respectively (Fig. 4 and Table S3†). Compound **10a** is anchored in the active site pocket of TDP1 by direct hydrogen bonds *via* its carboxylate head group. Hydrogen bonds are formed between the carboxylate oxygen atoms and the side chain O γ atom of S399 (2.8 Å hydrogen bond distance), K495 N ζ atom (2.7 Å), H493 N ϵ 2 atom (2.9 Å), N283 N δ 2 atom (3.0 Å), K265 N ζ atom (2.6 Å), and H263 N δ 1 atom (2.8 Å). A water molecule (A823), forms a hydrogen bond bridge to the carboxylate O27 atom of **10a** (2.6 Å) and the backbone carbonyl oxygen of S514 (2.7 Å). Stabilizing hydrophobic interactions with **10a** are provided by the side chain atoms of Y204, P461, and W590 (Fig. 4A). Compound **10b** binds to the active site in essentially the same manner with the carboxylate headgroups bound to the catalytic residues. Hydrogen bonds are formed between the carboxylate oxygen atoms of **10b** and the side chain O γ atom of S399 (2.8 Å), the N ζ atom of K495 (2.6 Å), N ϵ 2 atom of H493 (2.4 Å), N δ 2 atom of N283 (2.7 Å), and N ζ atom of K265 (2.7 Å), and N ϵ 2 atom of H263 (2.7 Å) (Fig. 4B).

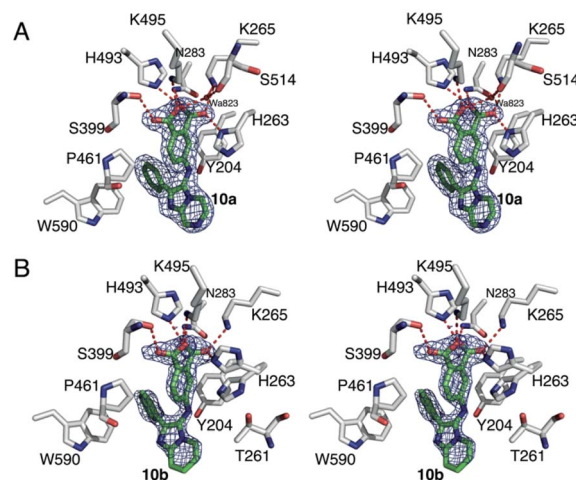


Fig. 4 Crystal structures of TDP1 bound **10a** and **10b**. (A) Stereoview of the active site of TDP1 (carbon atoms in gray, oxygen atoms in red, nitrogen atoms in blue) from the crystal structure of TDP1 bound to compound **10a** (PDB code: 6W7L, carbon atoms in green). The fit of compound **10a** is shown to the final $2F_o - F_c$ electron density map (blue) at 1.86 Å resolution and contoured at 1.0σ level. (B) Stereoview of the active site of TDP1 (carbon atoms in gray, oxygen atoms in red, nitrogen atoms in blue) from the crystal structure of TDP1 bound to compound **10b** (PDB code: 6W7K, carbon atoms in green). The fit of compound **10b** is shown to the final $2F_o - F_c$ electron density map (blue) at 1.7 Å resolution and contoured at 1.0σ level (atoms in green, nitrogen atoms in blue, and oxygen atoms in red). The binding site of a TOP1 derived peptide (cyan sticks) and a DNA tetranucleotide AGTT (yellow sticks) and bound vanadate (VO_4 , magenta sphere) are shown.



Discussion

As with many other phospholipase D members, the active site of TDP1 contains two conserved signature “HKN” motifs (H263, K265, N283 and H493, K495, N516) in close proximity to a substrate-binding channel.^{5,34–36} The enzymatic reaction occurs through two stages. In the first step, the N ϵ 2 nitrogen of H263 executes an in-line S $_N$ 2 attack onto the DNA 3'-phosphate ester group (Fig. 5A). This proceeds through a bipyramidal transition state, which is thought to be stabilized by the amine side chains of flanking K265 and K495 residues. A phosphoryl-mediated covalent bond is formed between the H263 imidazole ring and the 3'-phosphate group of the DNA substrate, while the incipient negative charge on the departing alcohol is neutralized by protonation from the H493 residue, which acts a general acid (Fig. 5A and B).³⁵ When the substrate consists of a DNA-TOP1 cleavage complex, the alcohol component represents the sidechain hydroxyl of the Y723 residue of TOP1. However, because a complex between native TOP1 and DNA is poorly cleaved, it has been shown that TOP1 must be processed prior to serving as a TDP1 substrate.^{35,37,38} The reaction cycle is completed by hydrolysis of the DNA – H263 phosphonamide bond through nucleophilic attack of a water molecule under general base-catalysed activation by the H493 N ϵ 2 nitrogen (Fig. 5C).

Insights into the mechanisms of substrate binding and phosphate ester cleavage have been informed by X-ray crystal structures of fully active N-terminally truncated TDP1 (149–608) without³⁴ and with vanadate or tungstate bound within the catalytic site both without³⁵ and with single-strand DNA and

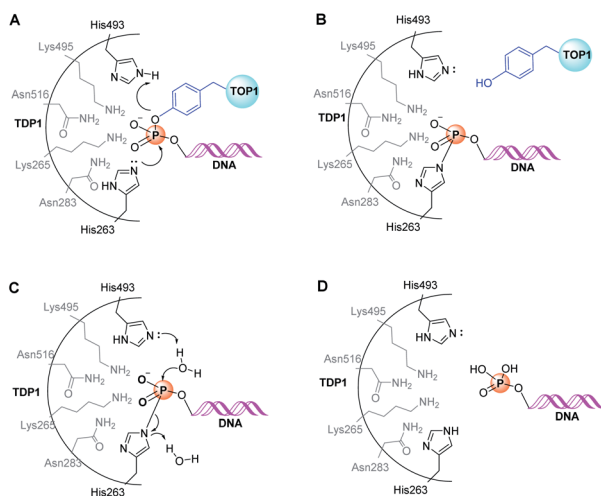


Fig. 5 Schematic representation of TDP1 catalysis. (A) Nucleophilic attacks on the phosphate group of the TOP1-DNA complex by the imidazole N atom of H263 (B) Formation of a phosphoryl-histidine covalent intermediate with displacement of the TOP1 tyrosyl residue. (C) Nucleophilic attack on the phosphoryl-histidine intermediate by a H493-activated water molecule. (D) Generation and release of a final 3'-phosphate DNA product. Residue numbers refer to human TDP1. TOP1-DNA complex are color-coded (phosphate group is tan; TOP1-derived peptide is cyan with tyrosyl residue in blue; DNA is magenta). Curved arrows denote the transfer of electron pairs.

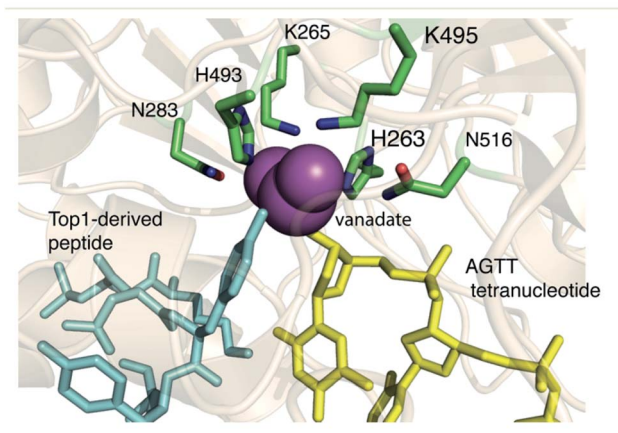


Fig. 6 The crystal structure of TDP1 (wheat ribbons, PDB code: 1RFF) illustrating the overall 3D-structure of the enzyme and highlighting features of the active site region. The residues forming the HKN motifs are depicted in stick format (carbon atoms in green, nitrogen atoms in blue, and oxygen atoms in red). The binding site of a TOP1 derived peptide (cyan sticks) and a DNA tetranucleotide AGTT (yellow sticks) and bound vanadate (VO $_4$, magenta sphere) are shown.

TOP1-derived peptide substrates (Fig. 6).^{36,39} Vanadate and tungstate serve as phosphoryl hydrolysis transition state intermediate mimetics that clarify the catalytic functions of residues within the TDP1 active site. More recently, crystal structures of TDP1 in complex with double-strand DNA have shed further light on binding interactions of DNA.⁴⁰

X-ray crystal structures of 10a and 10b bound to TDP1

Although the HKN-containing phosphoryl-binding pocket is well defined, to date structure-based inhibitor development has been challenging. A potentially important advance was recently made by our report of several low molecular weight compounds bound within the TDP1 phosphate binding pocket identified by soaking TDP1 crystals with fragment cocktails. These ligands bound deeply within the phosphate-binding pocket where they recapitulated features of substrate interactions with key residues of the catalytic machinery.²⁴ Many of the compounds were simple phthalic acids (Fig. 7).

The current crystal structures of **10a** (PDB code: 6W7L) and **10b** (PDB code: 6W7K) show that they include the same important interactions with the catalytic phosphate-binding

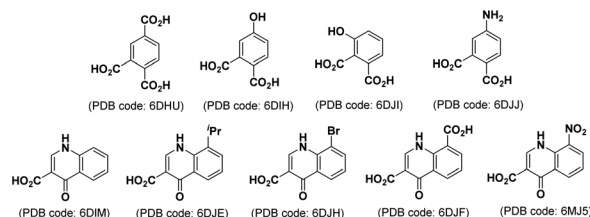


Fig. 7 Structures of low molecular weight TDP1-binding compounds identified by soaking TDP1 crystals with fragment cocktails.²⁴



pocket as our previously identified structurally more simple phthalic acid-containing inhibitors.²⁴ An important distinction of our SMM-derived inhibitors, is that they contain additional 2-phenylimidazo[1,2-*a*]pyrazine (for **10a**) or 2-phenylimidazo[1,2-

a]pyridine (for **10b**) nuclei that are held above the phthalic acid moiety, with the latter serving to anchor the molecules to the phosphate-binding pocket. This is exemplified by overlaying **10b** with TDP1-bound 4-aminophthalic acid (Fig. 8A).²⁴

Our new TDP1-binding platforms represent a potentially important advance beyond our previously identified phthalic acid inhibitors, due to the possibilities they afford for exploring substrate binding regions outside the catalytic pocket. A three-component picture emerges relating TDP1 structure to enzymatic function. First, there is a well-formed catalytic pocket (red surface, Fig. 8C), whose bottom is formed by the two HKN motifs. Second, extending from one side of the active site is a long narrow positively charged cleft, where the single-stranded DNA binds (magenta surface, Fig. 8C). Third, projecting outward from the catalytic center in the direction opposite to the DNA-binding cleft is a more open and less positively charged channel, which is occupied by the TOP1-derived peptide (tan surface, Fig. 8C) (Fig. 8B and C). While low molecular weight molecules, such as 4-aminophthalic acid efficiently access elements of the phosphate-binding pocket, they are largely confined within this region.²⁴ The more structurally complex molecular architectures of agents discovered as a result of the current SMM screen, such as **10a** and **10b**, extend above the phosphate-binding pocket and overlap with both the peptide and DNA components of the bi-substrate complex (Fig. 8B). This provides projections directed toward both the DNA and peptide-binding channels that potentially afford points from which to elaborate extensions into these channels (Fig. 8C). Therefore, inhibitors identified in our current work represent a platform for rationale-based construction of trivalent ligands capable of simultaneously accessing the catalytic phosphate-binding pocket and the neighbouring DNA and peptide-binding channels.

Conclusions

A number of TDP1 inhibitors have previously been reported with some exerting synergy with TOP1 inhibitors.^{10,12,14,16,18,19} However, the precise manner in which these ligands bind to the TDP1 protein are unknown, which limits further development using structure-based approaches. Our current work represents a qualitative departure from previous efforts in the way that it first interrogates the ability of ligands to bind to TDP1 using a microarray including more than 20 000 ligands. It then secondarily examines the ability of binders to inhibit the catalytic activity of TDP1 using highly specific biochemical assays. The SMM screen identified structurally similar *N*,2-diphenylimidazo[1,2-*a*]pyrazin-3-amines as TDP1 binders and catalytic inhibitors, which we then explored using one-pot Groebke–Blackburn–Bienayme multicomponent reactions. In so doing, we have achieved analogues having high inhibitory potencies. We have also shown that members of the series exhibit competitive modes of inhibition and that certain compounds can function in synergistic fashion with a TOP1 inhibitor. Finally, we solved the crystal structures of a subset of compounds bound within the TDP1 catalytic site. Importantly, inhibitors identified through the SMM approach are anchored

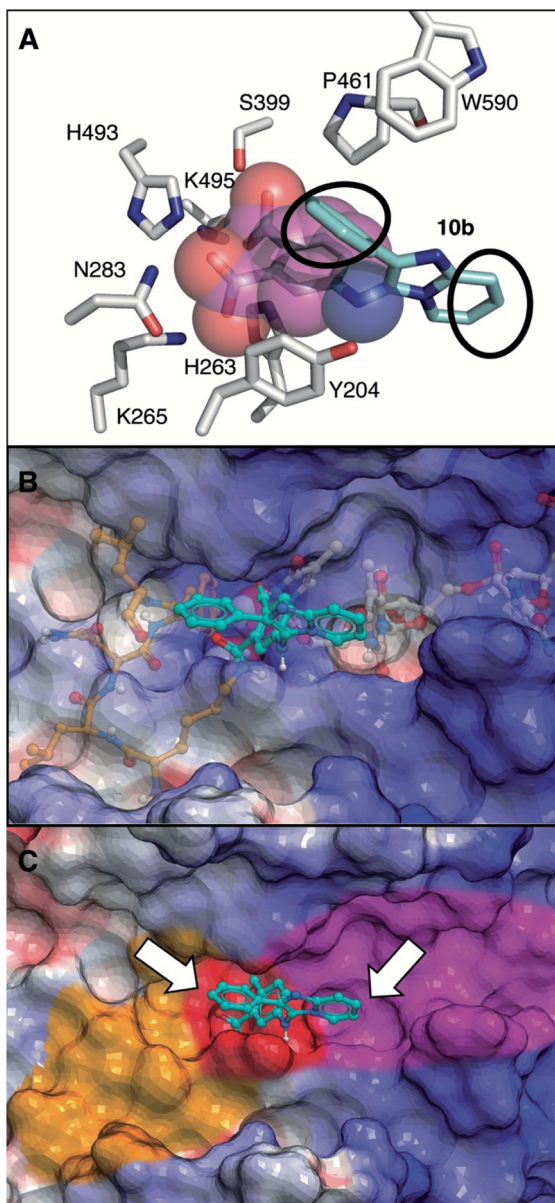


Fig. 8 View of the TDP1 active site (carbon atoms in gray, nitrogen in blue, and oxygen in red). (A) Comparison of the binding modes of 4-aminophthalic acid (depicted as space-filling with carbons in magenta; PDB code: 6DJJ)²⁴ and **10b** (carbons in cyan; PDB code: 6W7K) with black circles showing sites of potential extension above and beyond the phosphate-binding pocket. (B) Overlay of **10b** onto with previously reported semi-transparent structures of single-strand DNA (carbons in grey) and TOP1-derived substrate peptide (carbons in tan) with vanadate as a phosphate mimetic (bound space filling) (PDB code: 1NOP).³⁵ Electrostatic surface colored with positive charge blue, negative charge red and neutral white. (C) Bound **10b** with phosphate-binding catalytic pocket colored red; DNA-binding pocket colored magenta and peptide-binding pocket colored tan. Arrows indicate positions on **10b** extending into substrate binding channels.



within the catalytic phosphate-binding pocket, while simultaneously providing extensions into both the substrate DNA and peptide-binding channels.

In summary, using a SMM approach, our current work has permitted the elaboration of our previously discovered monovalent fragments into platforms that may form a foundation upon which a new genre of potent multivalent TDP1 inhibitors can be built. Multivalency affords a well-known means of achieving significant enhancement in binding affinity, where it is not uncommon for increases in affinity to be orders-of-magnitude relative to mono-valent ligands.^{41,42} Our compounds present a new class of ligands that have been established by X-ray crystallography to bind within the catalytic pocket and extend out of the pocket with projections toward the DNA and peptide substrate binding channels. Given the current state of TDP1 inhibitor development, this achievement is significant in its implications for future development of trivalent ligands with potentially high TDP1 affinity and selectivity. Our findings make a particularly important contribution to the field, given the historic challenges faced by developing TDP1 inhibitors.

Conflicts of interest

Aspects of the work presented are contained within one or more patent applications.

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References

- 1 A. S. Kawale and L. F. Povirk, *Nucleic Acids Res.*, 2018, **46**, 520–537.
- 2 H. U. Barthelmes, M. Habermeyer, M. O. Christensen, C. Mielke, H. Interthal, J. J. Pouliot, F. Boege and D. Marko, *J. Biol. Chem.*, 2004, **279**, 55618–55625.
- 3 Y. Pommier, *Chem. Rev.*, 2009, **109**, 2894–2902.
- 4 G. L. Beretta, G. Cossa, L. Gatti, F. Zunino and P. Perego, *Curr. Med. Chem.*, 2010, **17**, 1500–1508.
- 5 Y. Pommier, S.-y. N. Huang, R. Gao, B. B. Das, J. Murai and C. Marchand, *DNA Repair*, 2014, **19**, 114–129.
- 6 S. S. Laev, N. F. Salakhutdinov and O. I. Lavrik, *Bioorg. Med. Chem.*, 2016, **24**, 5017–5027.
- 7 H. Interthal, H. J. Chen, T. E. Kehl-Fie, J. Zotzmann, J. B. Leppard and J. J. Champoux, *EMBO J.*, 2005, **24**, 2224–2233.
- 8 E. Q. Comeaux and R. C. A. M. van Waardenburg, *Drug Metab. Rev.*, 2014, **46**, 494–507.
- 9 R. Gao, B. B. Das, R. Chatterjee, O. D. Abaan, K. Agama, R. Matuo, C. Vinson, P. S. Meltzer and Y. Pommier, *DNA Repair*, 2014, **13**, 1–9.
- 10 A. Zakharenko, N. Dyrkheeva and O. Lavrik, *Med. Res. Rev.*, 2019, **39**, 1427–1441.
- 11 E. J. Brettrager and C. A. M. van Waardenburg, *Cancer Drug Resist.*, 2019, **2**, 1153–1163.
- 12 E. V. Koldysheva, A. P. Men'shchikova, E. L. Lushnikova, N. A. Popova, V. I. Kaledin, V. P. Nikolin, A. L. Zakharenko, O. A. Luzina, N. F. Salakhutdinov and O. I. Lavrik, *Bull. Exp. Biol. Med.*, 2019, **166**, 661–666.
- 13 A. Thomas and Y. Pommier, *Clin. Cancer Res.*, 2019, **25**, 6581.
- 14 A. L. Zakharenko, O. A. Luzina, D. N. Sokolov, V. I. Kaledin, V. P. Nikolin, N. A. Popova, J. Patel, O. D. Zakharova, A. A. Chepanova, A. Zafar, J. Reynisson, E. Leung, I. K. H. Leung, K. P. Volcho, N. F. Salakhutdinov and O. I. Lavrik, *Eur. J. Med. Chem.*, 2019, **161**, 581–593.
- 15 O. Luzina, A. Filimonov, A. Zakharenko, A. Chepanova, O. Zakharova, E. Ilina, N. Dyrkheeva, G. Likhatskaya, N. Salakhutdinov and O. Lavrik, *J. Nat. Prod.*, 2020, **83**, 2320–2329.
- 16 A. L. Zakharenko, M. S. Drenichev, N. S. Dyrkheeva, G. A. Ivanov, V. E. Oslovsky, E. S. Ilina, I. A. Chernyshova, O. I. Lavrik and S. N. Mikhailov, *Molecules*, 2020, **25**, 3694.
- 17 E. V. Koldysheva, A. P. Men'shchikova, E. L. Lushnikova, N. A. Popova, V. I. Kaledin, V. P. Nikolin, A. L. Zakharenko, O. A. Luzina, N. F. Salakhutdinov and O. I. Lavrik, *Bull. Exp. Biol. Med.*, 2019, **166**, 661–666.
- 18 T. M. Khomenko, A. L. Zakharenko, A. A. Chepanova, E. S. Ilina, O. D. Zakharova, V. I. Kaledin, V. P. Nikolin, N. A. Popova, D. V. Korchagina, J. Reynisson, R. Chand, D. M. Ayine-Tora, J. Patel, I. K. H. Leung, K. P. Volcho, N. F. Salakhutdinov and O. I. Lavrik, *Int. J. Mol. Sci.*, 2020, **21**, 126.
- 19 E. D. Gladkova, I. V. Nechepurenko, R. A. Bredikhin, A. A. Chepanova, A. L. Zakharenko, O. A. Luzina, E. S. Ilina, N. S. Dyrkheeva, E. M. Mamontova, R. O. Anarbaev, J. Reynisson, K. P. Volcho, N. F. Salakhutdinov and O. I. Lavrik, *Int. J. Mol. Sci.*, 2020, **21**, 7162.
- 20 T. S. Dexheimer, S.-y. N. Huang, S. Antony, C. Marchand and Y. Pommier, *Adv. Anticancer Agents Med. Chem.*, 2013, **2**, 444–471.
- 21 J. B. Baell, L. Ferrins, H. Falk and G. Nikolakopoulos, *Aust. J. Chem.*, 2013, **66**, 1483–1494.



- 22 D. A. Erlanson, *J. Med. Chem.*, 2015, **58**, 2088–2090.
- 23 J. B. Baell and J. W. M. Nissink, *ACS Chem. Biol.*, 2018, **13**, 36–44.
- 24 G. T. Lountos, X. Z. Zhao, T. R. Burke, E. Kiselev, Y. Pommier, J. E. Tropea, D. Needle and D. S. Waugh, *Nucleic Acids Res.*, 2019, **47**, 10134–10150.
- 25 M. Uttamchandani and S. Q. Yao, *Methods Mol. Biol.*, 2010, **669**, 1–15.
- 26 K. Zlotkowski, W. M. Hewitt, R. S. Sinniah, J. E. Tropea, D. Needle, G. T. Lountos, J. J. Barchi, D. S. Waugh and J. S. Schneekloth, *SLAS Discovery*, 2017, **22**, 760–766.
- 27 J. E. Bradner, O. M. McPherson and A. N. Koehler, *Nat. Protoc.*, 2006, **1**, 2344–2352.
- 28 J. L. Duffner, P. A. Clemons and A. N. Koehler, *Curr. Opin. Chem. Biol.*, 2007, **11**, 74–82.
- 29 J. Sztuba-Solinska, S. R. Shenoy, P. Gareiss, L. R. H. Krumpke, S. F. J. Le Grice, B. R. O'Keefe and J. S. Schneekloth, *J. Am. Chem. Soc.*, 2014, **136**, 8402–8410.
- 30 C. Marchand, S.-y. N. Huang, T. S. Dexheimer, W. A. Lea, B. T. Mott, A. Chergui, A. Naumova, A. G. Stephen, A. S. Rosenthal, G. Rai, J. Murai, R. Gao, D. J. Maloney, A. Jadhav, W. L. Jorgensen, A. Simeonov and Y. Pommier, *Mol. Cancer Ther.*, 2014, **13**, 2116–2126.
- 31 S. Shaaban and B. F. Abdel-Wahab, *Mol. Diversity*, 2016, **20**, 233–254.
- 32 A. Boltjes and A. Doemling, *Eur. J. Org. Chem.*, 2019, **2019**, 7007–7049.
- 33 A. Ianevski, A. K. Giri and T. Aittokallio, *Nucleic Acids Res.*, 2020, **48**, W488–W493.
- 34 D. R. Davies, H. Interthal, J. J. Champoux and W. G. J. Hol, *Structure*, 2002, **10**, 237–248.
- 35 D. R. Davies, H. Interthal, J. J. Champoux and W. G. J. Hol, *J. Mol. Biol.*, 2002, **324**, 917–932.
- 36 D. R. Davies, H. Interthal, J. J. Champoux and W. G. J. Hol, *Chem. Biol.*, 2003, **10**, 139–147.
- 37 Y. Sun, L. K. Saha, S. Saha, U. Jo and Y. Pommier, *DNA Repair*, 2020, **94**, 102926.
- 38 Y. Sun, L. M. Miller Jenkins, Y. P. Su, K. C. Nitiss, J. L. Nitiss and Y. Pommier, *Sci. Adv.*, 2020, **6**, eaba6290.
- 39 D. R. Davies, H. Interthal, J. J. Champoux and W. G. J. Hol, *J. Med. Chem.*, 2004, **47**, 829–837.
- 40 F. J. Flett, E. Ruksenaite, L. A. Armstrong, S. Bharati, R. Carloni, E. R. Morris, C. L. Mackay, H. Interthal and J. M. Richardson, *Nat. Commun.*, 2018, **9**, 24.
- 41 V. M. Krishnamurthy, L. A. Estrofi and G. M. Whitesides, in *Fragment-based Approaches in Drug Discovery*, ed. W. J. a. D. A. Erlanson, Wiley-VCH GmbH & Co., Weinheim, DE, 2006, pp. 11–53.
- 42 C. Chittasupho, *Ther. Delivery*, 2012, **3**, 1171–1187.

