



Cite this: *RSC Adv.*, 2021, 11, 30229

Microwave-assisted synthesis of 4-oxo-2-butenic acids by aldol-condensation of glyoxylic acid†

Mélanie Uguen,^{‡*} Conghao Gai,^{‡,b} Lukas J. Sprenger,^a Hang Liu,^b
Andrew G. Leach^{‡,c} and Michael J. Waring^{‡*,a}

4-Oxobutenic acids are useful as biologically active species and as versatile intermediates for further derivatisation. Currently, routes to their synthesis can be problematic and lack generality. Reaction conditions for the synthesis of 4-oxo-2-butenic acid by microwave-assisted aldol-condensation between methyl ketone derivatives and glyoxylic acid have been developed. They provide the desired products in moderate to excellent yields for a wide range of substrates, by applying a simple procedure to accessible starting materials. The investigation revealed different conditions are required depending on the nature of the methylketone substituent, with aryl derivatives proceeding best using tosic acid and aliphatic substrates reacting best with pyrrolidine and acetic acid. This substituent effect is rationalised by frontier orbital calculations. Overall, this work provides methods for synthesis of 4-oxo-butenic acids across a broad range of substrates.

Received 19th July 2021
Accepted 1st September 2021

DOI: 10.1039/d1ra05539a

rsc.li/rsc-advances

Introduction

4-Oxo-2-butenic acids are interesting building blocks for drug discovery. Several derivatives have shown biological activity in their own right, for example to treat cancer,^{1,2} neurodegenerative,³ metabolic,⁴ and gastric⁵ conditions as well as antimicrobial^{6,7} or antifungal⁶ properties (Fig. 1). In addition, their high reactivity makes them versatile intermediates for further derivatisation.

Preparation of 4-oxo-2-butenic acids has often proven to be scope-limited, with Friedel–Crafts acylations⁴ used for aromatic substrates (Fig. 2) and oxidative furan-opening⁸ for aliphatic ones (Fig. 2). Although we managed to obtain the desired 4-oxo-2-butenic acid product when applying the oxidative furan oxidative conditions to an electron-rich aromatic example, we were unable to identify conditions compatible with electron-deficient aromatic substrates.

Aldol-condensation with glyoxylic acid is compatible with a larger range of starting materials with a few examples available in the literature, mainly using acetophenone derivatives as substrates (Fig. 2). Literature reported conditions are typically

acid-promoted, most commonly by acetic acid,^{7,9–12} sulphuric acid,^{13–17} phosphoric acid,^{13,18} toluene-4-sulfonic acid¹⁹ and formic acid,²⁰ with two acids frequently used together.^{12,19,21–28} The acid promoters are usually used neat or in large excess under reflux for relatively long periods of time. A few base-promoted procedures are also described in the literature, using potassium carbonate^{29,30} or sodium hydroxide at reflux or under reduced pressure.³¹ All these conditions are quite harsh on the reactants and resulting products, thus limiting the scope of the transformation.

Therefore, we decided to investigate the aldol-condensation with glyoxylic acid to identify efficient conditions for the preparation of 4-oxo-2-butenic acid derivatives. We aimed to

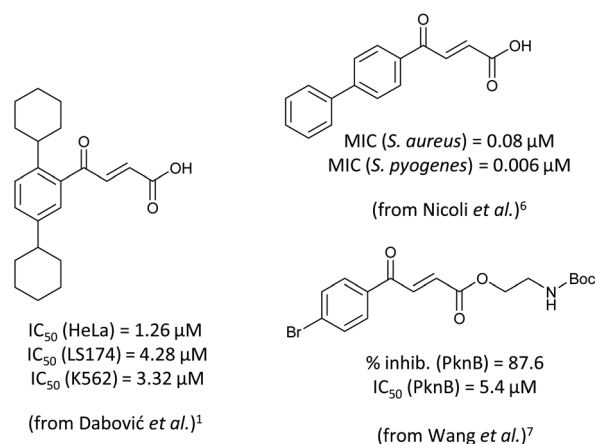


Fig. 1 Examples of biologically active 4-oxo-2-butenic acids and analogues.

^aCancer Research UK Drug Discovery Unit, Newcastle University Centre for Cancer, Chemistry, School of Natural and Environmental Sciences, Newcastle University, Bedson Building, Newcastle upon Tyne NE1 7RU, UK. E-mail: mike.waring@ncl.ac.uk

^bOrganic Chemistry Group, College of Pharmacy, Naval Medical University, Shanghai, 200433, P. R. China

^cDivision of Pharmacy and Optometry, School of Health Sciences, University of Manchester, Manchester M13 9PT, UK

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d1ra05539a

‡ Joint first authors.



Previous routes to synthesise 4-oxo-2-butenic acids:

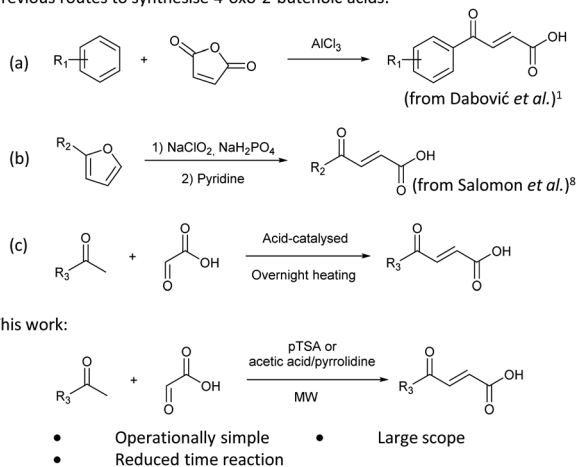


Fig. 2 Synthetic routes for the preparation of 4-oxo-2-butenic acid derivatives.

develop operationally simple conditions compatible with a large range of substrates by using microwave heating, which had the potential to reduce reaction times and increase yields.

Results and discussion

Initially, 4-methoxyacetophenone was used as a test substrate. Reactions with glyoxylic acid under basic conditions were first attempted but showed moderate conversion to the desired product (**1**) (Table S1†). Reaction *via* the pyrrolidine-derived enamine also showed moderate conversion, with the initial aldol adduct obtained in larger amount than the desired aldol-

condensation product. Attempts to force the water elimination by addition of tosyl chloride led to an improved conversion of the starting material. It was proposed that the tosyl chloride was hydrolysed *in situ* and the reaction was catalysed by tosic acid. Accordingly, carrying out the direct tosic acid-promoted aldol-condensation provided the desired product in good yield (70%), confirming the previous hypothesis (Table 1, entry 1). These conditions were applied to the more electron deficient 4-cyanoacetophenone, in which case, even after prolonged heating, some starting ketone remained (Table 1, entry 2). To improve the conversion towards the formation of the product **2** as well as reducing the heating time, microwave-assisted heating was performed using similar conditions, leading to a moderate isolated yield of **2** (32%, Table 1, entry 3). Optimisation of the irradiation time and temperature allowed an increase of the yield to 45% with a reduced reaction time of 1 h (Table 1, entry 4). Shorter reaction time led to incomplete conversion of the starting material whereas increase of the temperature from 160 °C to 180 °C led to partial or total degradation of the desired product (Table 1, entries 5–7), suggesting that heating at 160 °C for 1 hour were giving the best results. These findings suggest that increased temperature and pressure allowed by the microwave reactor are key to drive the

Table 2 Scope evaluation of the aldol-condensation reaction with glyoxylic acid promoted by tosic acid^a

R	Product	Yield	R	Product	Yield
	1	94%		11	78%
	2	45%		3	0%
	4	66%		12	0%
	5	61%		13	0%
	6	93%		14	0%
	7	76%		15	30% ^{b,c}
	8	62%		16	0%
	9	52%		17	0%
	10	55%		18	0%

Table 1 Reaction conditions attempted for aldol-condensation with glyoxylic acid

Entry	R	Cond. ^a	Heat.	Time	Temp. (°C)	Isol. yield
1		A	Conv.	48 h	80	70%
2		A	Conv.	72 h	80	n.c. ^b
3		A	MW	16 h	100	32%
4		A	MW	1 h	160	45%
5		A	MW	10 min	180	<20% ^b
6		A	MW	15 min	180	40%
7		A	MW	5 min	160	0%
8		A	MW	16 h	100	0%
9		B	MW	8 h	80	25%
10		B	MW	10 min	100	14%
11		B	MW	8 h	60	52%

^a Conditions A: 3.0 eq. glyoxylic acid monohydrate, 1.0 eq. TsOH monohydrate, dioxane; conditions B: 3.0 eq. glyoxylic acid monohydrate, 1.0 eq. pyrrolidine, 1.0 eq. acetic acid, MeOH.

^b Products not isolated.

^a Typical scale: 2.4 mmol, 6 mL of solvent. ^b MW temperature and time: 100 °C, 16 h. ^c NMR yield.



reaction while allowing a simple reaction set-up. Therefore, these optimised conditions were also applied to 4-methoxyacetophenone and showed an improved yield of 94% (Table 2).

When applying these conditions to cyclohexylmethyl ketone starting material, no desired product **3** was formed (Table 1, entry 8). The major product was the aldol adduct intermediate. However, treatment of cyclohexylmethyl ketone with glyoxylic acid, in the presence of pyrrolidine and acetic acid (Table 1, entry 9), using microwave-assisted heating, enabled the isolation of the desired product **3** in 25% yield. Increasing the temperature from 80 °C to 100 °C led to product degradation, even with decreased reaction time (Table 1, entry 10), but decreasing the temperature to 60 °C improved the yield to 52% (Table 1, entry 11).

Hence, two sets of microwave-assisted conditions for the formation of 4-oxo-2-butenic acid by aldol-condensation of glyoxylic acid with acetyl derivative have been developed. A scope evaluation of these conditions was performed on electron-rich aromatic (**1**), electron-poor aromatic (**2**, **6–9**), halogen-containing electron-neutral aromatic (**10** and **11**), aliphatic ring (**3**, **12–13**), heteroatom-containing (**14**) and aliphatic chain (**15–18**) 4-oxo-2-butenic acid derivatives (Tables 2 and 3).

TsOH-promoted aldol-condensation provided the desired products in good to excellent yields for aromatic substrates generating products **1**, **2** and **4–11** in 45–94% yield (Table 2).

Table 3 Scope evaluation of the aldol-condensation reaction with glyoxylic acid promoted by acetic acid and pyrrolidine^a

R	Product	Yield	R	Product	Yield
	1	12%		11	<4%
	2	0%		3	52%
	4	5%		12	63%
	5	5%		13	52%
	6	0%		14	43%
	7	5%		15	0%
	8	5%		16	71%
	9	4%		17	92%
	10	4%		18	0%

^a Typical scale: 2.4 mmol, 6 mL of solvent.

Electron donating and withdrawing substituents and *ortho*, *meta* and *para* substitution patterns were well tolerated. No desired product formation was observed when these conditions were applied to aliphatic substrates **3**, **12–14** and **16–18**. Pentan-2-one, however, yielded the desired product **15** under the TsOH-promoted conditions (by NMR) as a minor component relative to the expected internal aldol-condensation product **19** (ratio 1 : 1.5, Fig. S1†). After heating at 100 °C for 16 hours, **19** was the only product formed (99% yield). Hence, this suggests, as expected, that the tosic acid mediated reaction may not be compatible with methyl ketones bearing an additional enolisable centre.

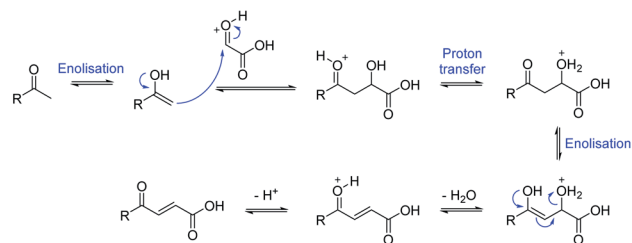
The pyrrolidine-acetic acid conditions were also applied to the selected substrates (Table 3). With aliphatic substrates, these conditions yielded the desired products **3** and **12–17** with yields between 43% and 92%, in which no product was obtained with the TsOH-promoted conditions. In this case, substrates with additional enolisable centres were tolerated (**3**, **12–17**) and the internal aldol adduct was not observed. No desired product was obtained for the *t*-butyl ketone **18**, presumably due to increased steric hindrance. Aromatic products **1**, **4**, **5** and **7–11** were obtained in poor 4 to 12% yields, much lower than with the TsOH-promoted conditions. However, no product was observed for the electron-poor examples **2** and **6** nor for the aliphatic chain **15** (Table 3).

For all the examples above, the obtained 4-oxo-2-butenic acids all had the *E* conformation for the alkene bond confirmed by NMR. No trace of the *Z* alkene was observed, demonstrating the stereoselectivity of this transformation.

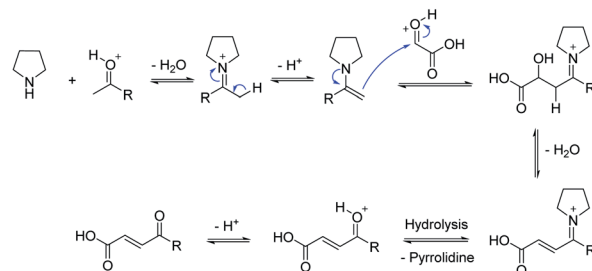
Finally, scale-up of the synthesis of **2** from 0.69 mmol to 6.9 mmol demonstrated the scalability of the TsOH-promoted reaction with no change in yield. Scale up of the synthesis of **3** using the pyrrolidine-acetic acid conditions from 4.0 mmol to 7.2 mmol gave increased yield (from 38% to 52%).

Based on the assumption that the reaction proceeds by attack of the protonated glyoxylic acid by the enol form of the

Enol-mediated mechanism:



Enamine-mediated mechanism:



Scheme 1 Mechanisms for the aldol condensation reaction.



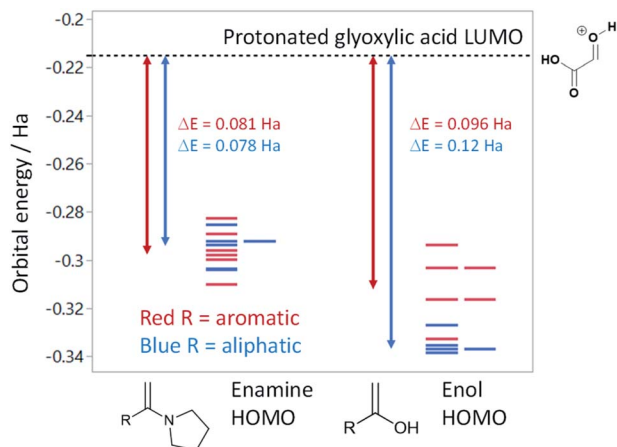


Fig. 3 Calculated HOMO energies at the RHF/6-31+G** level for selected enamines and enols relative to the protonated glyoxylic acid LUMO. Double-headed arrows show the average HOMO–LUMO gap for aromatic (red) and aliphatic (blue) substituents.

methylketone under the tosic acid promoted conditions, and by either the enol or the enamine in the presence of pyrrolidine/acetic acid (Scheme 1) as the rate determining step, the observed differences in reactivity were rationalised from the calculated energy gaps between the protonated glyoxylic acid LUMO and the enol or enamine HOMOs (Fig. 3). Calculations employed the RHF/6-31+G** level of theory in the Gaussian09 suite of programs.³² Geometries were optimised and frequencies computed to verify that they are minima.

HOMOs of the enol form of the aromatic ketones are consistently higher in energy than those of the aliphatic examples. This explains why the TsOH conditions work better on aromatic compounds as the HOMO–LUMO energy gap is significantly reduced (ΔE (aromatic) = 0.096 Ha, ΔE (aliphatic) = 0.12 Ha, $\Delta\Delta E$ = 0.024 Ha). For the aliphatic examples, HOMOs of the enamines are higher in energy compared to their corresponding enols (average HOMO–LUMO ΔE = 0.078 Ha). This provides an explanation as to why the pyrrolidine-acetic acid conditions work better for aliphatic substrates, and that these reactions likely proceed *via* the enamine as the predominant pathway.

Conclusions

We successfully identified two sets of conditions that provide *E*-4-oxo-2-butenic acids using microwave-assisted aldol-condensation with glyoxylic acid. These provide the desired products in good to excellent yields for substrate with various electronic and steric properties (electron-rich, -neutral and -poor aromatic, *ortho*-, *meta*-, and *para*-substituted aromatic, linear and cyclic aliphatic). The reaction can be scaled-up to mmol scale with no effect on the yield. It was observed that tosic acid promoted conditions work better for aromatic substrates whereas acetic acid and pyrrolidine are preferred for aliphatic substrates. These findings can be rationalised by the relative differences in HOMO–LUMO energy gaps for the enol and enamine intermediates. Overall, we describe a convenient and efficient synthesis for 4-oxo-2-butenic acids

which should allow an easier access to these biologically-relevant molecules and their derivatives.

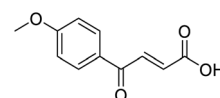
Experimental

General procedure A for the synthesis of 1, 2 and 4–11

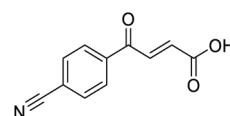
In a Biotage microwave vial, acetyl derivative (1 eq.), glyoxylic acid monohydrate (3 eq.) and TsOH monohydrate (1 eq.) were dissolved in dioxane (2.5 mL mmol^{−1}). The vial was closed with a 20 mm aluminium crimp cap with a PTFE/silicone septum and heated in the microwave for 1 h at 160 °C using the low absorption mode while stirred at 600 rpm with a PTFE stirring bar. 2 M HCl aqueous solution was added to the mixture. This was extracted 3 times with CH₂Cl₂. Combined organic phases were dried over MgSO₄. The solvent was removed under vacuum. A typical scale was 2.4 mmol but the reaction was successfully scaled up and down.

(*E*)-4-(4-Methoxyphenyl)-4-oxobut-2-enoic acid (1).

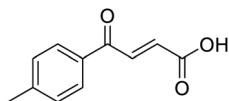
Compound 1 was obtained following General procedure A. Flash chromatography (0 to 10% 0.1 AcOH in MeOH in CH₂Cl₂) provided 1 as a bright yellow solid (2.59 g, 94%). *R*_f = 0.65 (80% EtOAc in PE); mp = 180–182 °C (from MeOH); UV λ_{max} (EtOH/nm) 287.2, 223.6, 200.0; FTIR (cm^{−1}) ν_{max} 2840br (O–H acid), 1699s (C=O acid), 1661s (C=O ketone), 1592s (C=C alkene), 1511s (C=C aromatic), 1420s (O–H acid), 1167 (s, C–O methoxy); ¹H NMR (500 MHz; CDCl₃; Me₄Si) δ_{H} 3.90 (3H, s, CH₃), 6.88 (1H, d, *J* = 15.5 Hz, CH=CH), 6.97–7.02 (2H, m, CH–Ar), 7.97–8.05 (3H, m, CH=CH and CH–Ar); ¹³C NMR (126 MHz; CDCl₃; Me₄Si) δ_{C} 55.77 (CH₃), 114.39 (C–Ar), 129.68 (C–Ar), 130.61 (C=C), 131.55 (C–Ar), 138.77 (C=C), 164.57 (C–Ar), 169.84 (COOH), 187.48 (CO); MS(ES⁺) *m/z* 207.2; HRMS calcd for C₁₁H₁₀O₄ [M + H]⁺ 207.0579, found 207.0555.



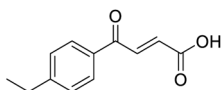
(*E*)-4-(4-Cyanophenyl)-4-oxobut-2-enoic acid (2). Compound 2 was obtained following General procedure A. Normal phase flash chromatography (0 to 10% 0.1% AcOH in MeOH in CH₂Cl₂) yielded compounds 2 as a pale yellow solid (443 mg, 32%). *R*_f = 0.12 (10% MeOH in CH₂Cl₂); mp = 134–140 °C (from MeOH); UV λ_{max} (EtOH/nm) 256.4; FTIR (cm^{−1}) ν 3063br (O–H acid), 2231w (CN); ¹H NMR (500 MHz; MeOD; Me₄Si) δ_{H} 6.83 (1H, d, *J* = 15.6 Hz, CH=CH), 7.91 (1H, d, *J* = 15.6 Hz, CH=CH), 7.93 (2H, d, *J* = 8.4 Hz, CH–Ar), 8.17 (2H, d, *J* = 8.4 Hz, CH–Ar); ¹³C NMR (methanol-*d*₄, 126 MHz) δ_{C} 117.86 (CN), 118.87 (C–Ar), 130.43 (C–Ar), 133.93 (C–Ar), 135.05 (C=C), 136.84 (C=C), 141.19 (C–Ar), 168.23 (COOH), 190.30 (CO); MS (ES⁺) *m/z* = 201.1 [M–H][−]; HRMS calcd for C₁₁H₇NO₃ 200.0348 [M + H]⁺ found 200.0363.



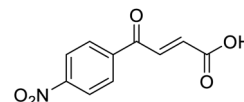
(*E*)-4-Oxo-4-(*p*-tolyl)but-2-enoic acid (4). Compound **4** was obtained following General procedure A. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **4** as a yellow solid (300 mg, 66%). $R_f = 0.39$ (10% MeOH in CH_2Cl_2); mp = 134–136 °C (from MeOH); UV λ_{max} (EtOH/nm) 280.0, 231.1; FTIR (cm^{-1}) ν_{max} 3031br (OH acid), 2923s (CH), 1664s (C=O acid), 1410s (O–H acid); ^1H NMR (600 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{H} 2.40 (3H, s, CH_3), 6.67 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.38 (2H, d, $J = 7.8$ Hz, CH-Ar), 7.87 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.94 (2H, d, $J = 7.8$ Hz, CH-Ar), 13.14 (1H, br s, COOH); ^{13}C NMR (150 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{C} 21.67 (CH_3), 129.34 (C-Ar), 130.04 (C-Ar), 133.09 (C=C), 134.18 (C-Ar), 136.65 (C=C), 145.10 (C-Ar), 166.78 (COOH), 189.30 (CO); MS(ES^+) m/z 191.1 [$\text{M} + \text{H}$] $^+$; HRMS calcd for $\text{C}_{11}\text{H}_{11}\text{O}_3$ [$\text{M} + \text{H}$] $^+$ 191.0629, found 191.0697.



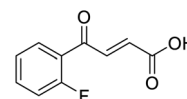
(*E*)-4-(4-Ethylphenyl)-4-oxobut-2-enoic acid (5). Compound **5** was obtained following General procedure A. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **5** as a yellow solid (300 mg, 61%). $R_f = 0.42$ (10% MeOH in CH_2Cl_2); mp = 91–93 °C (from MeOH); UV λ_{max} (EtOH/nm) 282.0, 232.1; FTIR (cm^{-1}) ν_{max} 3052br (OH acid), 2968s (CH), 2934s (CH), 1666s (C=O acid), 1415s (O–H acid); ^1H NMR (600 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{H} 1.20 (3H, t, $J = 7.2$ Hz, CH_3), 2.70 (2H, q, $J = 7.2$ Hz, CH_2), 6.68 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.41 (2H, d, $J = 7.8$ Hz, CH-Ar), 7.87 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.96 (2H, d, $J = 7.8$ Hz, CH-Ar), 13.15 (1H, br s, COOH); ^{13}C NMR (150 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{C} 15.48 (CH_3), 28.68 (CH_2), 128.87 (C-Ar), 129.45 (C-Ar), 133.18 (C=C), 134.43 (C-Ar), 136.62 (C=C), 151.07 (C-Ar), 166.80 (COOH), 189.34 (CO); MS(ES^+) m/z 205.1 [$\text{M} + \text{H}$] $^+$; HRMS calcd for $\text{C}_{12}\text{H}_{13}\text{O}_3$ [$\text{M} + \text{H}$] $^+$ 205.0791, found 205.0865.



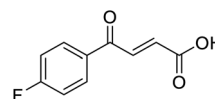
(*E*)-4-(4-Nitrophenyl)-4-oxobut-2-enoic acid (6). Compound **6** was obtained following General procedure A. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **6** as a yellow solid (493 mg, 93%). $R_f = 0.40$ (10% MeOH in CH_2Cl_2); mp = 173–175 °C (from MeOH); UV λ_{max} (EtOH/nm) 269.0; FTIR (cm^{-1}) ν_{max} 2989br (OH acid), 1604s (C=O acid), 1530s (NO_2), 1419s (O–H acid), 1350s (NO_2); ^1H NMR (600 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{H} 6.71 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.87 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 8.25 (2H, d, $J = 9.0$ Hz, CH-Ar), 8.36 (2H, d, $J = 9.0$ Hz, CH-Ar), 13.25 (1H, br s, COOH); ^{13}C NMR (150 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{C} 124.42 (C-Ar), 130.65 (C-Ar), 134.36 (C=C), 136.23 (C=C), 141.23 (C-Ar), 150.63 (C-Ar), 166.56 (COOH), 189.40 (CO); MS(ES^+) m/z 222.1 [$\text{M} + \text{H}$] $^+$; HRMS calcd for $\text{C}_{10}\text{H}_6\text{NO}_5$ [$\text{M}-\text{H}$] $^-$ 220.0329, found 220.0257.



(*E*)-4-(2-Fluorophenyl)-4-oxobut-2-enoic acid (7). Compound **7** was obtained following General procedure A. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **7** as a yellow solid (352 mg, 76%). $R_f = 0.41$ (10% MeOH in CH_2Cl_2); mp = 105–106 °C (from MeOH); UV λ_{max} (EtOH/nm) 246.0; FTIR (cm^{-1}) ν_{max} 3001br (OH acid), 1665s (C=O acid), 1421s (O–H acid); ^1H NMR (600 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{H} 6.62 (1H, dd, $J = 15.6$ Hz, $J_{\text{HF}} = 0.6$ Hz, $\text{CH}=\text{CH}$), 7.37–7.41 (2H, m, CH-Ar), 7.57 (1H, dd, $J = 15.6$ Hz, $J_{\text{HF}} = 3.0$ Hz, $\text{CH}=\text{CH}$), 7.70–7.74 (1H, m, CH-Ar), 7.80–7.83 (1H, m, CH-Ar), 13.21 (1H, br s, COOH); ^{13}C NMR (150 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{C} 117.30 (d, $J_{\text{CF}} = 22.2$ Hz, C-Ar), 125.50 (d, $J_{\text{CF}} = 2.8$ Hz, C-Ar), 125.54 (d, $J_{\text{CF}} = 16.6$ Hz, C-Ar), 131.22 (C=C), 133.33 (C=C), 136.06 (d, $J_{\text{CF}} = 9.0$ Hz, C-Ar), 139.14 (d, $J_{\text{CF}} = 5.1$ Hz, C-Ar), 161.40 (d, $J_{\text{CF}} = 252.0$ Hz, CF-Ar), 166.63 (COOH), 188.54 (CO); ^{19}F NMR (282 MHz; $\text{DMSO}-d_6$) δ_{F} –111.29; MS(ES^+) m/z 195.1 [$\text{M} + \text{H}$] $^+$; HRMS calcd for $\text{C}_{10}\text{H}_8\text{FO}_3$ [$\text{M} + \text{H}$] $^+$ 195.0379, found 195.0452.



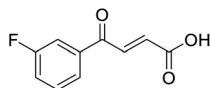
(*E*)-4-(4-Fluorophenyl)-4-oxobut-2-enoic acid (8). Compound **8** was obtained following General procedure A. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **8** as a yellow solid (289 mg, 62%). $R_f = 0.41$ (10% MeOH in CH_2Cl_2); mp = 108–109 °C (from MeOH); UV λ_{max} (EtOH/nm) 243.1; FTIR (cm^{-1}) ν_{max} 2978br (OH acid), 1666s (C=O acid), 1411s (O–H acid); ^1H NMR (600 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{H} 6.68 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.36–7.40 (2H, m, CH-Ar), 7.87 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 8.10–8.14 (2H, m, CH-Ar), 13.07 (1H, br s, COOH); ^{13}C NMR (150 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{C} 116.51 (d, $J_{\text{CF}} = 21.9$ Hz, C-Ar), 132.33 (d, $J_{\text{CF}} = 9.6$ Hz, C-Ar), 133.37 (d, $J_{\text{CF}} = 2.0$ Hz, C-Ar), 133.49 (C=C), 136.40 (C=C), 165.90 (d, $J_{\text{CF}} = 251.6$ Hz, CF-Ar), 166.70 (COOH), 188.54 (CO); ^{19}F NMR (282 MHz; $\text{DMSO}-d_6$) δ_{F} –104.52; MS(ES^+) m/z 195.1 [$\text{M} + \text{H}$] $^+$; HRMS calcd for $\text{C}_{10}\text{H}_8\text{FO}_3$ [$\text{M} + \text{H}$] $^+$ 195.0379, found 195.0468.



(*E*)-4-(3-Fluorophenyl)-4-oxobut-2-enoic acid (9). Compound **9** was obtained following General procedure A. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **9** as a white solid (239 mg, 52%). $R_f = 0.41$ (10% MeOH in CH_2Cl_2); mp = 109–110 °C (from MeOH); UV λ_{max} (EtOH/nm) 246.0; FTIR (cm^{-1}) ν_{max} 3079br (OH acid), 1668s (C=O acid), 1416s (O–H acid); ^1H NMR (600 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{H} 6.70 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.54–7.58 (1H, m, CH-Ar), 7.62–7.65 (1H, m, CH-Ar), 7.79–7.81 (1H, m, CH-Ar), 7.85 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.90 (1H, dd, $J = 7.8$ Hz, $J_{\text{HF}} = 0.6$ Hz, CH-Ar), 13.18 (1H, br s, COOH); ^{13}C NMR (150 MHz; $\text{DMSO}-d_6$; Me_4Si) δ_{C} 115.56 (d,

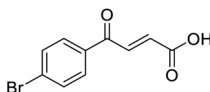


$J_{\text{CF}} = 22.4$ Hz, C-Ar), 121.35 (d, $J_{\text{CF}} = 21.3$ Hz, C-Ar), 125.55 (d, $J_{\text{CF}} = 1.4$ Hz, C-Ar), 131.68 (d, $J_{\text{CF}} = 7.8$ Hz, C-Ar), 133.94 (C=C), 136.23 (C=C), 138.80 (d, $J_{\text{CF}} = 6.2$ Hz, C-Ar), 162.72 (d, $J_{\text{CF}} = 244.5$ Hz, CF-Ar), 166.64 (COOH), 189.02 (CO); ^{19}F -NMR (282 MHz, DMSO- d_6) δ_{F} -111.78; MS(ES^+) m/z 195.1 $[\text{M} + \text{H}]^+$; HRMS calcd for $\text{C}_{10}\text{H}_6\text{FO}_3$ $[\text{M}-\text{H}]^-$ 193.0379, found 193.0306.



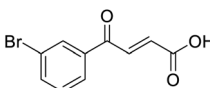
(E)-4-(4-Bromophenyl)-4-oxobut-2-enoic acid (10).

Compound **10** was obtained following General procedure A. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **10** as a white solid (334 mg, 55%). $R_{\text{f}} = 0.41$ (10% MeOH in CH_2Cl_2); mp = 149–152 °C (from MeOH); UV λ_{max} (EtOH/nm) 265.0; FTIR (cm^{-1}) ν_{max} 3020br (OH acid), 1666s (C=O acid), 1418s (O-H acid); ^1H NMR (600 MHz; DMSO- d_6 ; Me_4Si) δ_{H} 6.69 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.78–7.81 (2H, m, CH-Ar), 7.85 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.96–7.98 (2H, m, CH-Ar), 13.09 (1H, br s, COOH); ^{13}C NMR (150 MHz; DMSO- d_6 ; Me_4Si) δ_{C} 128.65 (CBr-Ar), 131.21 (C-Ar), 132.55 (C-Ar), 133.78 (C=C), 135.64 (C-Ar), 136.26 (C=C), 166.69 (COOH), 189.28 (CO); MS(ES^+) m/z 254.1 $[\text{M}(^{79}\text{Br}) + \text{H}]^+$ and 256.1 $[\text{M}(^{81}\text{Br}) + \text{H}]^+$; HRMS calcd for $\text{C}_{10}\text{H}_8^{79}\text{BrO}_3$ $[\text{M} + \text{H}]^+$ 254.9585, found 254.9652.



(E)-4-(3-Bromophenyl)-4-oxobut-2-enoic acid (11).

Compound **11** was obtained following General procedure A. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **11** as a white solid (476 mg, 78%). $R_{\text{f}} = 0.41$ (10% MeOH in CH_2Cl_2); mp = 148–151 °C (from MeOH); UV λ_{max} (EtOH/nm) 244.8; FTIR (cm^{-1}) ν_{max} 2992br (OH acid), 1672s (C=O acid), 1416s (O-H acid); ^1H NMR (600 MHz; DMSO- d_6 ; Me_4Si) δ_{H} 6.70 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.55 (1H, t, $J = 7.8$ Hz, CH-Ar), 7.85 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 7.91–7.92 (1H, m, CH-Ar), 8.04–8.05 (1H, m, CH-Ar), 8.15–8.16 (1H, m, CH-Ar), 13.08 (1H, br s, COOH); ^{13}C NMR (150 MHz; DMSO- d_6 ; Me_4Si) δ_{C} 122.81 (CBr-Ar), 128.32 (C-Ar), 131.57 (C-Ar), 131.68 (C-Ar), 134.03 (C=C), 136.24 (C=C), 136.95 (C-Ar), 138.66 (C-Ar), 166.66 (COOH), 189.05 (CO); MS(ES^+) m/z 254.1 $[\text{M}(^{79}\text{Br}) + \text{H}]^+$ and 256.1 $[\text{M}(^{81}\text{Br}) + \text{H}]^+$; HRMS calcd for $\text{C}_{10}\text{H}_6^{79}\text{BrO}_3$ $[\text{M}-\text{H}]^-$ 252.9585, found 252.9503.

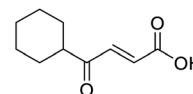


General procedure B for the synthesis of 3, 12–14 and 16–17

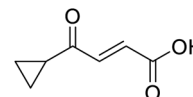
In a Biotage microwave vial, methyl ketone-containing compound (1 eq.), glyoxylic acid monohydrate (3 eq.) and acetic acid (1 eq.) were dissolved in MeOH (2.5 mL mmol $^{-1}$) then pyrrolidine (1 eq.) was added. The vial was sealed with a 20 mm

aluminium crimp cap with PTFE/silicon septa and the mixture stirred for 5 min using a PTFE stirring bar. The mixture was irradiated using the MW at 60 °C for 8 h while stirred at 600 rpm. The solvent was removed under vacuum. A typical scale was 2.4 mmol but the reaction was successfully scaled up and down.

(E)-4-Cyclohexyl-4-oxobut-2-enoic acid (3). Compound **3** was obtained following General procedure B. Flash chromatography (0 to 15% 0.1% AcOH in MeOH in CH_2Cl_2) yielded **3** as a beige solid (277 mg, 38%). $R_{\text{f}} = 0.32$ (5% MeOH in CH_2Cl_2); mp = 113–115 °C (from MeOH); UV λ_{max} (EtOH/nm) 330.8, 219.8; FTIR (cm^{-1}) ν_{max} 3062br (OH acid), 2922s (CH), 2851s (CH), 1660s (C=O acid), 1427s (O-H acid); ^1H NMR (500 MHz; MeOD; Me_4Si) δ_{H} 1.05–1.21 (5H, m, Cy), 1.51–1.67 (5H, m, Cy), 2.50 (1H, tt, $J = 10.7, 3.4$ Hz, Cy), 6.42 (1H, d, $J = 15.9$ Hz, C=C), 6.91 (1H, d, $J = 15.9$ Hz, C=C); ^{13}C NMR (126 MHz; MeOD; Me_4Si) δ_{C} 26.49 (Cyc), 29.31 (Cyc), 50.35 (Cyc), 132.21 (C=C), 139.56 (C=C), 168.50 (COOH), 204.38 (CO); MS(ES^+) m/z 183.1 $[\text{M} + \text{H}]^+$; HRMS calcd for $\text{C}_{10}\text{H}_{14}\text{O}_3$ $[\text{M}-\text{H}]^-$ 181.0870, found 181.0870.



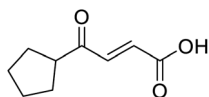
(E)-4-Cyclopropyl-4-oxobut-2-enoic acid (12). Compound **12** was obtained following General procedure B. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **12** as a yellow oil (213 mg, 63%). $R_{\text{f}} = 0.40$ (10% MeOH in CH_2Cl_2); UV λ_{max} (EtOH/nm) 223.0; FTIR (cm^{-1}) ν_{max} 3012br (OH acid), 1715s (C=O acid), 1392s (O-H acid); ^1H NMR (600 MHz; DMSO- d_6 ; Me_4Si) δ_{H} 0.96–0.98 (2H, m, CHH-cyclopropane), 0.99–1.03 (2H, m, CHH-cyclopropane), 2.47–2.51 (1H, m, CH-cyclopropane), 6.70 (1H, d, $J = 16.0$ Hz, $\text{CH}=\text{CH}$), 7.03 (1H, d, $J = 16.0$ Hz, $\text{CH}=\text{CH}$), 13.06 (1H, br s, COOH); ^{13}C NMR (150 MHz; DMSO- d_6 ; Me_4Si) δ_{C} 12.07 (CH_2), 19.82 (CH), 131.92 (C=C), 139.56 (C=C), 167.00 (COOH), 200.20 (CO); MS(ES^+) m/z 141.1 $[\text{M} + \text{H}]^+$; HRMS calcd for $\text{C}_7\text{H}_8\text{O}_3$ $[\text{M}-\text{H}]^-$ 139.0473, found 139.0400.



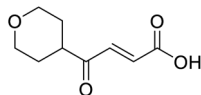
(E)-4-Cyclopentyl-4-oxobut-2-enoic acid (13). Compound **13** was obtained following General procedure B. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **13** as a yellow oil (210 mg, 52%). $R_{\text{f}} = 0.38$ (10% MeOH in CH_2Cl_2); UV λ_{max} (EtOH/nm) 221.0; FTIR (cm^{-1}) ν_{max} 3065br (OH acid), 2951s (CH), 2869s (CH), 1687s (C=O acid), 1428s (O-H acid); ^1H NMR (600 MHz; DMSO- d_6 ; Me_4Si) δ_{H} 1.54–1.58 (4H, m, CH_2CH_2 -cyclopentane), 1.64–1.85 (4H, m, CH_2CH_2 -cyclopentane), 3.31 (1H, m, CH-cyclopentane), 6.62 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 6.99 (1H, d, $J = 15.6$ Hz, $\text{CH}=\text{CH}$), 13.09 (1H, br s, COOH); ^{13}C NMR (150 MHz; DMSO- d_6 ; Me_4Si) δ_{C} 26.09 (CH_2CH_3 -cyclopentane), 28.63 (CH_2CH_3 -cyclopentane), 49.34 (CH), 132.00 (C=C), 139.26 (C=C), 166.97 (COOH), 202.25 (CO); MS(ES^+) m/z



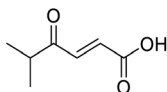
169.1 $[M + H]^+$; HRMS calcd for $C_9H_{12}O_3$ $[M-H]^-$ 167.0798, found 167.0725.



(E)-4-Oxo-4-(tetrahydro-2H-pyran-4-yl)but-2-enoic acid (14). Compound **14** was obtained following General procedure B. Flash chromatography (0 to 10% 0.1% AcOH in MeOH in CH_2Cl_2) yielded **14** as an orange solid (321 mg, 38%). R_f = 0.41 (5% MeOH in CH_2Cl_2); mp = 90–92 °C (from MeOH); UV λ_{max} (EtOH/nm) 243.0, 201.7; FTIR (cm^{-1}) ν_{max} 3067br (O–H acid), 2963 (CH), 2841 (CH), 1669s (C=O acid), 1424s (O–H acid), 1117s (C–O ether); 1H NMR (500 MHz; MeOD; Me_4Si) δ_H 1.53 (2H, dtd, J = 13.4, 11.4, 4.2 Hz, Pyr), 1.71 (2H, ddd, J = 13.4, 4.2, 2.1 Hz, Pyr), 2.93 (1H, tt, J = 11.4, 3.8 Hz, Pyr), 3.42 (2H, td, J = 11.4, 2.1 Hz, Pyr), 3.87 (2H, ddd, J = 11.4, 4.2, 2.1 Hz, Pyr), 6.62 (1H, d, J = 15.9 Hz, C=C), 7.07 (1H, d, J = 15.9 Hz, C=C); ^{13}C NMR (126 MHz; MeOD; Me_4Si) δ_C 29.06 (Pyr), 47.01 (Pyr), 68.07 (Pyr), 132.75 (C=C), 139.07 (C=C), 168.44 (COOH), 202.59 (CO); MS(ES+) m/z 185.1 $[M + H]^+$; HRMS calcd for $C_9H_{12}O_4$ $[M + H]^+$ 183.0663, found 183.0644.

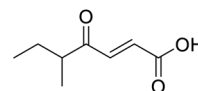


(E)-5-Methyl-4-oxohex-2-enoic acid (16). Compound **16** was obtained following General procedure B. Normal phase flash chromatography (0 to 10% 0.1% AcOH in MeOH in CH_2Cl_2) yielded compound **16** as a pale yellow oil (53 mg, 37%). R_f = 0.57 (5% MeOH in CH_2Cl_2); UV λ_{max} (EtOH/nm) 221.8; FTIR (cm^{-1}) ν_{max} 3376br (O–H acid), 1686s (C=O acid), 1638s (C=O ketone), 1466 (O–H acid); 1H NMR (500 MHz; MeOD; Me_4Si) δ_H 0.99 (6H, d, J = 6.8 Hz, Me), 2.83 (1H, hept, J = 6.8 Hz, CH), 6.54 (1H, d, J = 15.9 Hz, C=C), 7.01 (1H, d, J = 15.9 Hz, C=C); ^{13}C NMR (126 MHz; MeOD; Me_4Si) δ_C 18.24 (Me), 40.57 (CMe₂), 132.44 (C=C), 139.36 (C=C), 168.56 (COOH), 205.15 (CO); MS(ES+) m/z 143.1 $[M + H]^+$; HRMS calcd for $C_7H_{10}O_3$ $[M-H]^-$ 141.0557, found 141.0536.

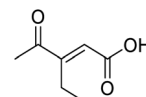


(E)-5-Methyl-4-oxohept-2-enoic acid (17). Compound **17** was obtained following General procedure B. Flash chromatography (0 to 50% EtOAc in petroleum ether) yielded **17** as a yellow oil (343 mg, 92%). R_f = 0.39 (10% MeOH in CH_2Cl_2); UV λ_{max} (EtOH/nm) 272.4; FTIR (cm^{-1}) ν_{max} 2968br (OH acid), 1631s (C=O acid), 1454s (O–H acid); 1H NMR (600 MHz; DMSO- d_6 ; Me_4Si) δ_H 0.82 (3H, t, J = 7.2 Hz, CH_2CH_3), 1.02 (3H, d, J = 6.6 Hz, CH_3), 1.37 (1H, dq, J = 6.6 Hz and 7.2 Hz, CHH), 1.37 (1H, dq, J = 6.6 Hz and 7.2 Hz, CHH), 2.89 (1H, sext, J = 6.6 Hz, CH), 6.62 (1H, d, J = 15.6 Hz, $CH=CH$), 7.05 (1H, d, J = 15.6 Hz, $CH=CH$), 13.10 (1H, br s, COOH); ^{13}C NMR (150 MHz; DMSO- d_6 ; Me_4Si) δ_C 11.61 (CH_2CH_3), 15.59 (CH_3), 25.53 (CH_2CH_3), 45.52 (CH), 131.93 (C=C), 138.73 (C=C), 166.92 (COOH),

203.66 (CO); MS(ES+) m/z 157.1 $[M + H]^+$; HRMS calcd for $C_8H_{12}O_3$ $[M-H]^-$ 155.0786, found 155.0713.



(E)-3-Ethyl-4-oxopent-2-enoic acid (19). Compound **19** was obtained following General procedure A. Normal phase flash chromatography (0 to 10% 0.1% AcOH in MeOH in CH_2Cl_2) yielded compound **19** as a yellow oil (132 mg, 99%). R_f = 0.63 (5% MeOH in CH_2Cl_2); UV λ_{max} (EtOH/nm) 220.9; FTIR (cm^{-1}) ν 3144br (O–H acid), 2974 (CH), 1681s (C=O acid), 1362 (O–H acid); 1H NMR (500 MHz; MeOD; Me_4Si) δ_H 0.99 (3H, t, J = 7.5 Hz, Me), 2.37 (3H, s, COMe), 2.71 (2H, q, J = 7.5 Hz, CH_2), 6.64 (1H, s, C=C); ^{13}C NMR (126 MHz; MeOD; Me_4Si) δ_C 14.06 (Me), 20.87 (CH_2), 26.56 (COMe), 128.17 (C=C), 156.25 (C=C), 169.07 (COOH), 201.87 (CO); MS(ES+) m/z 143.1 $[M + H]^+$; HRMS calcd for $C_7H_{10}O_3$ $[M-H]^-$ 141.0557, found 141.0560.



Author contributions

The investigation was performed by M. U., C. G., L. J. S. and H. L. under the supervision of M. J. W. A. G. L. carried out the molecular orbital calculations. The manuscript was written by M. U. and reviewed by M. J. W.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We thank Newcastle University (SagE doctoral training programme studentship award to MU), Cancer Research UK (C2115/A21421, Newcastle Drug Discovery Unit Programme Grant) and the Erasmus+ programme of the European Union (studentship award to LJS) for funding.

Notes and references

- 1 B. J. Drakulić, T. P. Stanojković, Ž. S. Žižak and M. M. Dabović, *Eur. J. Med. Chem.*, 2011, **46**, 3265–3273.
- 2 M. D. Vitorović-Todorović, A. Erić-Nikolić, B. Kolundžija, E. Hamel, S. Ristić, I. O. Juranić and B. J. Drakulić, *Eur. J. Med. Chem.*, 2013, **62**, 40–50.
- 3 A. Giordani, P. Pevarello, M. Cini, R. Bormetti, F. Greco, S. Toma, C. Speciale and M. Varasi, *Bioorg. Med. Chem. Lett.*, 1998, **8**, 2907–2912.
- 4 L. A. Arnold, A. Kosinski, E. Estébanez-Perpiñá and R. K. Guy, *J. Med. Chem.*, 2007, **50**, 5269–5280.



- 5 M. Bianchi, A. Butti, Y. Christidis, J. Perronnet, F. Barzaghi, R. Cesana and A. Nencioni, *Eur. J. Med. Chem.*, 1988, **23**, 45–52.
- 6 A. Dal Pozzo, M. Acquasaliente, G. Donzelli, P. De Maria and M. C. Nicoli, *J. Med. Chem.*, 1987, **30**, 1674–1677.
- 7 C. Xu, X. Bai, J. Xu, J. Ren, Y. Xing, Z. Li, J. Wang, J. Shi, L. Yu and Y. Wang, *RSC Adv.*, 2017, **7**, 4763–4775.
- 8 S. P. Annangudi, M. Sun and R. G. Salomon, *Synlett*, 2005, **2005**, 1468–1470.
- 9 B. G. Sundar, T. Bailey, E. Bacon, L. Aimone, Z. Huang, J. Lyons, R. Raddatz and R. Hudkins, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 5543–5546.
- 10 X. Li, N. Liu, H. Zhang, S. E. Knudson, H. J. Li, C. T. Lai, C. Simmerling, R. A. Slayden and P. J. Tonge, *ACS Med. Chem. Lett.*, 2011, **2**, 818–823.
- 11 J. Sun, L. Chen, C. Liu, Z. Wang, D. Zuo, J. Pan, H. Qi, K. Bao, Y. Wu and W. Zhang, *Chem. Biol. Drug Des.*, 2015, **86**, 1541–1547.
- 12 J. Ren, J. Xu, G. Zhang, C. Xu, L. L. Zhao, X. F. You, Y. Wang, Y. Lu, L. Yu and J. Wang, *Bioorg. Med. Chem. Lett.*, 2019, **29**, 539–543.
- 13 P. Jakubec, D. Berkeš, A. Kolarović and F. Považanec, *Synthesis*, 2006, 4032–4040.
- 14 N. Chakor, S. Dallavalle, L. Musso and M. Moretti, *Tetrahedron Lett.*, 2008, **49**, 5056–5058.
- 15 W. L. Wan, J. B. Wu, F. Lei, X. L. Li, L. Hai and Y. Wu, *Chin. Chem. Lett.*, 2012, **23**, 1343–1346.
- 16 X. Wang, Y. Yan, B. Gong, X. Tang, Q. Li, Y. Meng, H. E. Xu and W. Yi, *Bull. Korean Chem. Soc.*, 2013, **34**, 3143–3146.
- 17 A. S. Amarasekara and U. Ha, *Synth. Commun.*, 2018, **48**, 2533–2538.
- 18 R. Scheffold and P. Dubs, *Helv. Chim. Acta*, 1967, **50**, 798–808.
- 19 R. Messer, A. Schmitz, L. Moesch and R. Häner, *J. Org. Chem.*, 2004, **69**, 8558–8560.
- 20 V. P. Ghidu, M. A. Ilies, T. Cullen, R. Pollet and M. Abou-Gharbia, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 259–261.
- 21 A. Bolognese and G. Scherillo, *J. Org. Chem.*, 1977, **42**, 3867–3869.
- 22 K. Kaczanowska, H. Eickhoff, K. Albert, K. H. Wiesmüller and A. P. Schaffner, *J. Heterocycl. Chem.*, 2011, **48**, 792–798.
- 23 C. Desel, K. Werninghaus, M. Ritter, K. Jozefowski, J. Wenzel, N. Russkamp, U. Schleicher, D. Christensen, S. Wirtz, C. Kirschning, E. M. Agger, C. P. da Costa and R. Lang, *PLoS One*, 2013, **8**, 1–9.
- 24 S. Zhao, J. B. Lin, Y. Y. Zhao, Y. M. Liang and P. F. Xu, *Org. Lett.*, 2014, **16**, 1802–1805.
- 25 M. A. I. Salem, M. I. Marzouk and A. M. El-Kazak, *Molecules*, 2016, **21**, 249.
- 26 S. K. Wang, M. T. Chen, D. Y. Zhao, X. You and Q. L. Luo, *Adv. Synth. Catal.*, 2016, **358**, 4093–4099.
- 27 H. Chen, S. Zhao, S. Cheng, X. Dai, X. Xu, W. Yuan and X. Zhang, *J. Heterocycl. Chem.*, 2019, **56**, 1672–1683.
- 28 M. Lemurell, J. Ulander, H. Emtenäs, S. Winiwarter, J. Broddefalk, M. Swanson, M. A. Hayes, L. Prieto Garcia, A. Westin Eriksson, J. Meuller, J. Cassel, G. Saarinen, Z. Q. Yuan, C. Löfberg, S. Karlsson, M. Sundqvist and C. Whatling, *J. Med. Chem.*, 2019, **62**, 4325–4349.
- 29 M. Yamaguchi, N. Maruyama, T. Koga, T. Kuroki, K. Kamei, M. Hamana, M. Akima and N. Ohi, *Chem. Pharm. Bull.*, 1994, **42**, 1850–1853.
- 30 M. Krasavin, A. Shetnev, S. Baykov, S. Kalinin, A. Nocentini, V. Sharoyko, G. Poli, T. Tuccinardi, M. Korsakov, T. B. Tennikova and C. T. Supuran, *Eur. J. Med. Chem.*, 2019, **168**, 301–314.
- 31 J. Krall, C. H. Jensen, F. Bavo, C. B. Falk-Petersen, A. S. Haugaard, S. B. Vogensen, Y. Tian, M. Nittegaard-Nielsen, S. B. Sigurdardóttir, J. Kehler, K. T. Kongstad, D. E. Gloriam, R. P. Clausen, K. Harpsøe, P. Wellendorph and B. Frølund, *J. Med. Chem.*, 2017, **60**, 9022–9039.
- 32 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, J. J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman and D. J. Fox, *Gaussian09, Revision D.02*, Gaussian, Inc., Wallingford CT, 2016.

